

Corrosion Inhibition and Adsorption Properties of Ethanol Extract of *Vernonia Amygdalina* for the Corrosion of Mild Steel in H₂SO₄

A.O. Odiongenyi,¹ S.A. Odoemelam,¹ N.O. Eddy^{2,*}

¹Department of Chemistry, Michael Okpara University of Agriculture,
Umudike, Abia State, Nigeria

²Department of Chemistry, Ahmadu Bello University, Zaria, Kaduna State, Nigeria

Received 19 September 2008; accepted 4 December 2008

Abstract

Inhibitive and adsorption properties of ethanol extract of *vernonia amygdalina* for the corrosion of mild steel were studied using weight loss, thermometric, gasometric and IR methods of monitoring corrosion. The results revealed that ethanol extract of *Vernonia amygdalina* inhibited the corrosion of mild steel. The inhibition efficiency of the extract increased as the concentration of the extract increases. The inhibitor was found to function by being adsorbed on the surface of mild steel. The adsorption of the inhibitor followed the Langmuir adsorption isotherm. IR spectra of the corrosion product (without inhibitor), the extract and the corrosion product (with the inhibitor) confirmed that ethanol extract of *vernonia amygdalina* is an adsorption inhibitor. Phytochemical studies also revealed that ethanol *vernonia amygdalina* contains tannin, saponnins, flavanoid and anthraquinone, all of them contributing to the corrosion inhibition. Physical adsorption mechanism has been proposed from the values of some of the thermodynamic parameters obtained.

Keywords: corrosion inhibition, mild steel, adsorption, *vernonia amygdalina*.

Introduction

The use of inhibitors is one of the best methods of protecting metals against corrosion [1-2]. Most corrosion inhibitors are organic compounds having hetero atoms in their aromatic or long carbon chain [1-3]. However, there is increasing concern about the toxicity of most corrosion inhibitors. The toxic effect does not only affect living organisms but also poison the environment [2].

* Corresponding author. E-mail address: nabukeddy@yahoo.com

Due to the toxicity of some corrosion inhibitors, there has been increasing search for green corrosion inhibitors [3]. Inhibitors in this class are those that are environmentally friendly and are gotten from natural products such as plant extracts [4]. Several studies have been carried out on the inhibition of corrosion of metals by plant extract [5-11]. In most of these and other studies, nothing has been reported on the use of ethanol extract of *vernonia amygdalina* for the inhibition of the corrosion of mild steel in H₂SO₄. The present study is aimed at investigating the inhibitive properties of ethanol extract of *vernonia amygdalina* on the corrosion of mild steel in H₂SO₄. *Vernonia amygdalina* is an annual, erect, branched and hairy herb, having a height of 30 -120 cm. The leaves of the plant are simple and alternate. The plant is geographically distributed in West African countries including Nigeria [30]. According to Gill [31], the extract of *vernonia amygdalina* is medicinal and is used in curing headache and fever. Notable chemical constituents of ethanol extract of *vernonia amygdalina* are glycoside-vernoniside, vernonium and 15 sesquiterpenes lactones [31].

Experimental details

Materials preparation

Mild steel of composition (wt%) Mn (0.6), P (0.36), C (0.15), S (0.07) and Fe (98.79) was used for the study. The sheet was mechanically press-cut to form different coupons, each of dimension, 5x4x0.11 cm. Each coupon was degreased by washing with ethanol. The washed sample was dipped in acetone, removed and allowed to dry in air before use. All reagents used for the study were analar grade and double distilled water was used for their preparation.

Extraction of plants

Samples of *vernonia amygdalina* obtained from the Akwa Ibom State garden were dried, ground and soaked in a solution of ethanol. After 48 hours, the sample was cooled and filtered. The filtrate was subjected to evaporation (in order to leave the sample free of the ethanol) using a rotary evaporator. The stock solution of the extract so obtained was used in preparing 0.1, 0.2, 0.3, 0.4 and 0.5 g/L of 2.5 M H₂SO₄ and 0.1 M for used in gasometric/thermometric and gravimetric analysis, respectively.

Gasometric method

Hydrogen evolution measurements were carried out at 303 and 333 K as described in literature [12]. From the volume of hydrogen gas evolved per minute, corrosion rate (CR), inhibition efficiency (%I) and degree of surface coverage, were calculated using Equations 1, 2 and 3, respectively.

$$\%I = \left(1 - \frac{V_{Ht}^1}{V_{Ht}^0}\right) \times 100 \quad (1)$$

$$CR = (V_{Ht}^0 - V_{Ht}^1)/t \quad (2)$$

$$\theta = \%I / 100 = \left(1 - \frac{V_{Ht}^I}{V_{Ht}^o} \right) \quad (3)$$

where V_{Ht}^I is the volume of hydrogen gas at time t for inhibited solution and V_{Ht}^o is the volume of hydrogen gas evolved at time t for uninhibited solution.

Thermometric method

Measurements of temperature were carried out according to the method described by Eddy and Ebenso [13]. From the rise in temperature per minute, the reaction number (RN) and inhibition efficiency were calculated using Equations 4 and 5.

$$RN \left(^\circ C \text{ min}^{-1} \right) = \frac{T_m - T_i}{t} \quad (4)$$

$$\%I = \frac{RN_{aq} - RN_{wi}}{RN_{aq}} \times 100 \quad (5)$$

where RN_{aq} is the reaction number in the absence of inhibitors (blank solution), and RN_{wi} is the reaction number of 2.5 M H_2SO_4 containing studied inhibitors.

Gravimetric analysis

In gravimetric experiment, a previously weighed metal (mild steel) coupon was completely immersed in 250 mL of the test solution in an open beaker. The beaker was inserted into a water bath maintained at a temperature of 30 °C. After every 24 hours, each sample was withdrawn from the test solution, washed in a solution containing 50% NaOH and 100 g/L of zinc dust. The washed sample was dried with acetone before re-weighing. The difference in weight for a period of 168 h was taken as total weight loss. From the weight loss results, the inhibition efficiency (%I) of the inhibitor and degree of surface coverage were calculated using Equations 6 and 7, respectively,

$$\%I = (1 - W_1/W_2) \times 100 \quad (6)$$

$$\theta = 1 - W_1/W_2 \quad (7)$$

where W_1 and W_2 are the weight losses (g/dm^3) for mild steel in the presence and absence of the inhibitor in H_2SO_4 solution, respectively, and θ is the degree of surface coverage of the inhibitor.

Chemical analysis

IR analysis was carried out using a Buck model 500M infra red spectrophotometer. The samples were prepared using nujol oil. Photochemical analysis of the extract was carried out according to the method reported by Onyeka and Nwabekwe [14].

Results and discussions

Fig. 1 shows the variation of weight loss with time during the corrosion of mild steel in 0.02 – 0.5 M H_2SO_4 at 303 K. From the figure, it can be seen that weight loss of mild steel increases linearly with time, indicating that the rate of corrosion of mild steel in H_2SO_4 increases with time. Values of weight loss were also found to vary with concentration of H_2SO_4 in such a way that the trend of increase in weight loss with time was $0.5 > 0.1 > 0.02 > 0.01$ M H_2SO_4 . Fig. 2 shows the plot of variation of weight loss with time during the corrosion of mild steel in 0.1 M H_2SO_4 in the presence of various concentrations of ethanol extract of *vernonia amygdalina* as an inhibitor. It was also observed that the addition of ethanol extract of *Vernonia amagdalina* to the corrodent led to a reduction in weight loss compared to that of the blank. This indicates that ethanol extract of *vernonia amagdalina* inhibited the corrosion of mild steel in H_2SO_4 .

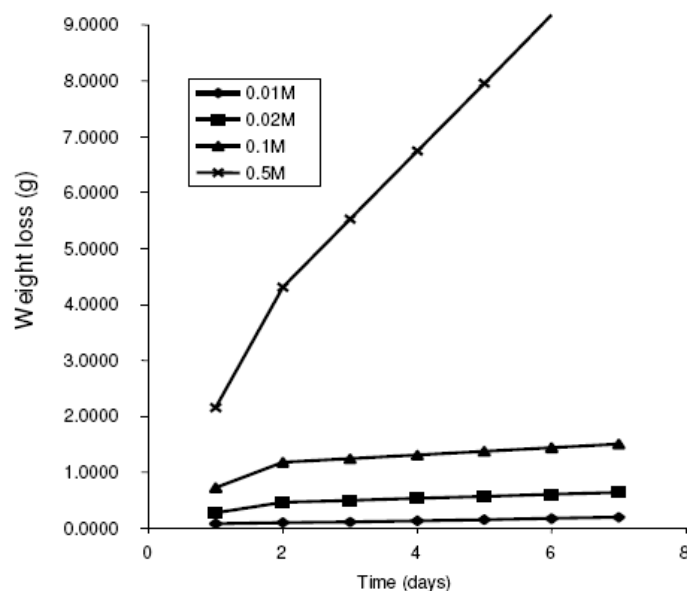


Figure 1. Variation of weight loss with time for the corrosion of mild steel in various concentrations of H_2SO_4 at 303 K.

Fig. 3 shows the variation of volume of hydrogen gas evolved with time during the corrosion of mild steel in 1.0 – 2.5 M of H_2SO_4 . From the figure, it can be seen that the volume of hydrogen gas evolved increased with increase in the period of contact and with the concentration of H_2SO_4 , indicating that the rate of corrosion of mild steel in H_2SO_4 increased as the period of contact and concentration of H_2SO_4 increase. Figs. 4 and 5 show the variation of volumes of hydrogen gas evolved with time for the corrosion of mild steel in 2.5 M H_2SO_4 containing various concentrations of ethanol extract of *vernonia amygdalina* at 303 and 333 K, respectively. From the figures, it is seen that the volume of hydrogen gas evolved, hence the rate of corrosion of mild steel, also increased with increase in the concentration of ethanol extract of *vernonia amagdalina* and with the period of contact. It was also observed that the volume of hydrogen gas

evolved by the blank (2.5 M H₂SO₄) was greater than the volumes evolved by solutions containing various concentrations of ethanol extract of *vernonia amygdalina*, indicating that the extract inhibited the corrosion of mild steel in H₂SO₄.

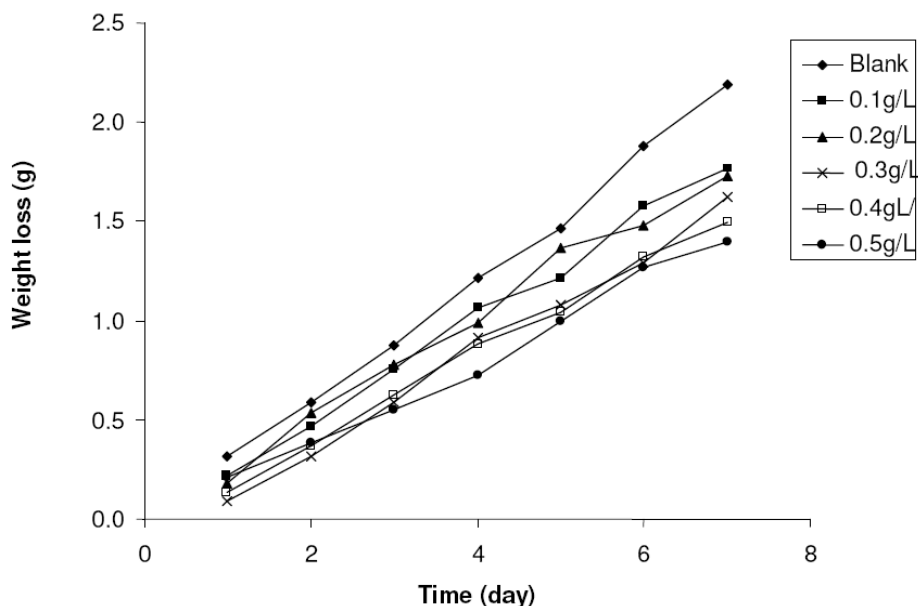


Figure 2. Variation of weight loss with time for the corrosion of mild steel in H₂SO₄ containing various concentrations of ethanol extract of *vernonia amygdalina*.

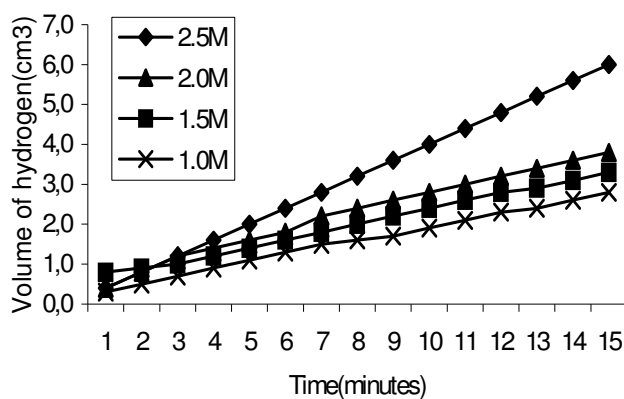


Figure 3. Variation of the volume of hydrogen gas evolved with time for the corrosion of mild steel in various concentrations of H₂SO₄.

The reaction number (RN) obtained from thermometric measurement, for the corrosion of mild steel in 2.5 M H₂SO₄ was 0.095. In the presence of various concentrations of ethanol extract of *vernonia amygdalina*, values of reaction number decreased to the range of 0.05 to 0.075 °C/min. The decrease in RN in the presence of ethanol extract of *vernonia amygdalina* also indicated that ethanol extract of *vernonia amygdalina* retarded the corrosion of mild steel in H₂SO₄.

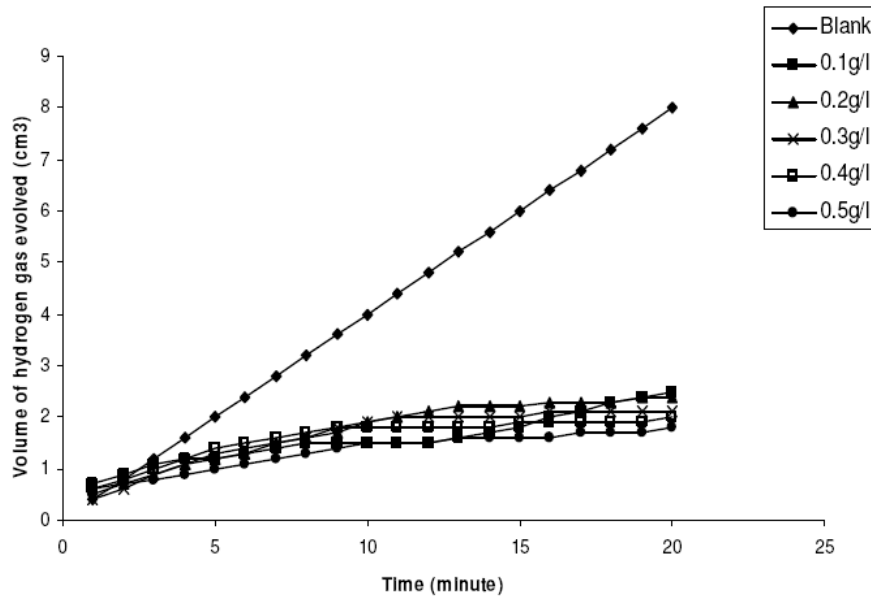


Figure 4. Variation of the volume of hydrogen gas evolved with time for the corrosion of mild steel in H_2SO_4 containing various concentrations of ethanol extract of *vernonia amygdalina* at 303 K.

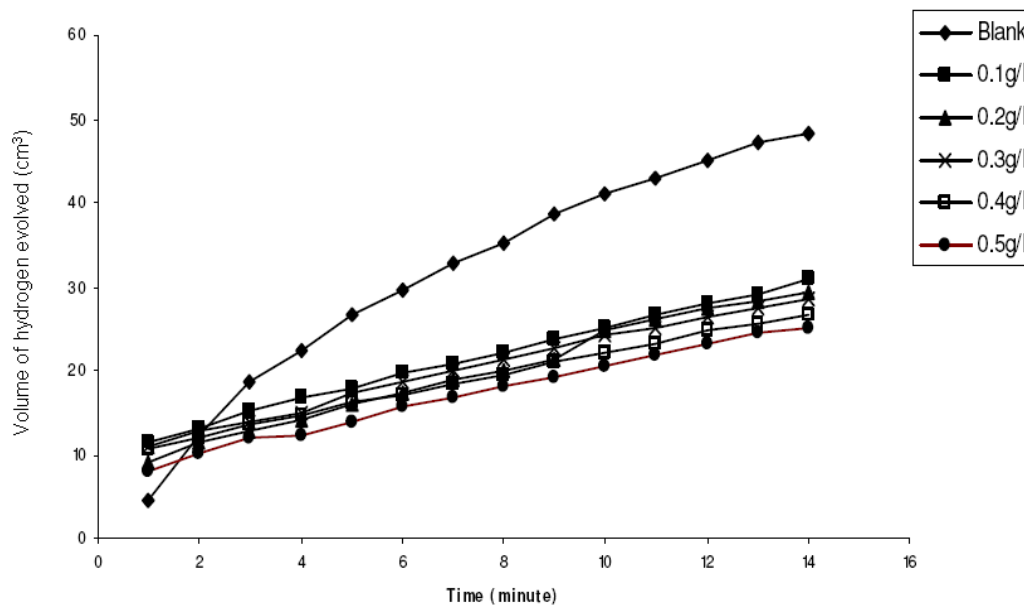


Figure 5. Variation of the volume of hydrogen gas evolved with time for the corrosion of mild steel in H_2SO_4 containing various concentrations of ethanol extract of *vernonia amygdalina* at 333 K.

Effect of ethanol extract of vernonia amygdalina on the corrosion of mild steel

Ethanol extract of *vernonia amygdalina* decreased the rate of weight loss, the rate of evolution of hydrogen gas and the rate of change of temperature. This confirms that ethanol extract of *vernonia amygdalina* inhibits the corrosion of mild steel in H_2SO_4 . Values of inhibition efficiency (%I) and corrosion rates obtained from weight loss, gasometric and thermometric methods (for the inhibition of the corrosion of mild steel in H_2SO_4 by various concentrations of ethanol extract of *vernonia amygdalina*) are recorded in Table 1. The results

indicate that inhibition efficiencies obtained from weight loss measurement are higher than those obtained from gasometric and thermometric methods, indicating that the average inhibition efficiency of the inhibitor is better than its instantaneous inhibition efficiencies. However, data obtained from gasometric and thermometric methods strongly correlated with those obtained from weight loss measurements ($r = 0.9803$ and 0.9151 for gasometry and thermometry data, respectively), confirming that ethanol extract of *vernonia amygdalina* is a good inhibitor for the corrosion of mild steel in H_2SO_4 .

Table 1. Values of inhibition efficiency and degree of surface coverage for the inhibition of the corrosion of mild steel at various concentrations of ethanol extract of *vernonia amygdalina*.

Con. (g/L)	Gasometric		Weight loss		Thermometric			
	%I (303 K)	CR (cm^3/min) (303 K)	%I (333 K)	CR (cm^3/min) (333 K)	%I (303 K)	CR (g/cm^3h^{-1}) (303 K)	%I (303 K)	CR ($^{\circ}C/min$) (303 K)
0.1	76.32	0.090	37.00	1.44	89.94	0.313	21.05	0.075
0.2	76.32	0.090	40.53	1.39	90.35	0.261	21.05	0.075
0.3	77.63	0.085	44.27	1.26	91.59	0.254	21.05	0.075
0.4	81.58	0.070	44.93	1.23	93.59	0.250	47.37	0.050
0.5	82.89	0.065	46.92	1.15	95.75	0.206	47.37	0.050

Effect of temperature

The effect of temperature on the corrosion reaction of mild steel in the absence and presence of ethanol extract of *vernonia amygdalina* was investigated using Arrhenius equation,

$$\log \frac{CR_2}{CR_1} = \frac{E_a}{2.303R} \left(\frac{1}{T_1} - \frac{1}{T_2} \right) \quad (8)$$

From hydrogen evolution measurements, values of CR (cm^3/min) were calculated and substituted into equation 6 to obtain E_a values. These values (Table 2) ranged from 74.00 to 77.44 kJ/mol, indicating that *vernonia amygdalina* is adsorbed on the surface of mild steel by physical adsorption [3]. Also, the average value of E_a obtained for the blank (36.00 kJ/mol) was lower than values obtained for systems containing various concentrations of ethanol extract of *vernonia amygdalina*, indicating that this extract retards the corrosion of mild steel in H_2SO_4 [19-21].

Table 2. Some thermodynamic parameters for the adsorption of ethanol extract of *vernonia amygdalina* on the surface of mild steel.

Concentration (g/L)	E _a (J/mol)	Q _{ads} (kJ/mol)
Blank	36.00	-
0.1	74.24	-38.25
0.2	72.86	-52.34
0.3	77.44	-47.62
0.4	74.00	-39.17
0.5	72.78	-49.82

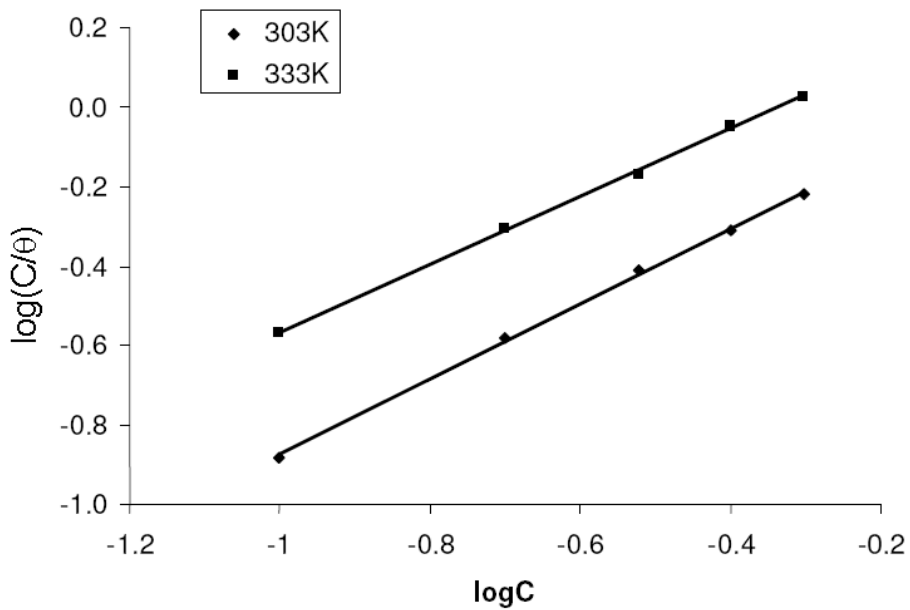


Figure 6. Langmuir isotherm for the adsorption of ethanol extract of *vernonia amygdalina* on the surface of mild steel.

Thermodynamics/adsorption consideration

The heat adsorbed during the inhibition of the corrosion of mild steel by ethanol extract of *vernonia amygdalina* was calculated using equation 9 [1-2]

$$Q_{ads} = 2.303R \left[\log \left(\frac{\theta_2}{1-\theta_2} \right) - \log \left(\frac{\theta_1}{1-\theta_1} \right) \right] \times \left(\frac{T_1 T_2}{T_2 - T_1} \right) \text{kJmol}^{-1} \quad (9)$$

Values of Q_{ads} are recorded in Table 2. These values ranged from -38.2481 to 52.3433 kJ/mol, indicating that the adsorption of ethanol extract of *vernonia*

amygdalina on mild steel surface is exothermic [17-19]. Also, since the reactions were carried out under constant pressure, the heat adsorbed is also equal to the enthalpy change [22-23].

Table 3. Langmuir adsorption parameters for the adsorption of ethanol extract of *vernonia amygdalina* on the surface of mild steel.

Langmuir	Temperature (K)	logK	slope	ΔG_{ads} (kJ/g)	R^2
	303	0.0723	0.9475	-11.56	0.9989
	333	0.2843	0.8518	-11.74	0.9996

Values of degree of surface coverage calculated from hydrogen evolution measurements have been used to evaluate the adsorption characteristics of ethanol extract of *vernonia amygdalina* on mild steel surface. The data obtained were found to fit Langmuir adsorption isotherm. The assumptions of Langmuir adsorption isotherm can be represented by equation below [19 – 23]:

$$C/\theta = 1/k + C \quad (10)$$

which can also be written as follows,

$$\log(C/\theta) = \log C - \log K \quad (11)$$

By plotting values of $\log(C/\theta)$ versus $\log C$, linear plots were generated (Fig. 6), confirming that the experimental data fitted the Langmuir adsorption isotherm for the adsorption of ethanol extract of *vernonia amygdalina* on mild steel, meaning that there is no interaction between the adsorbed species..

Values of free energy of adsorption, ΔG_{ads} , of ethanol extract of *vernonia amygdalina* on mild steel surface were calculated using equation 12 [23-27]

$$\Delta G_{\text{ads}} = -2.303RT \log(55.5K) \quad (12)$$

where R is the gas constant, T is the temperature and K is the equilibrium constant of adsorption of ethanol extract of *vernonia amygdalina* on the surface of mild steel, and 55.5 is the concentration of water in the solution. Values of ΔG_{ads} calculated from equation 12 are recorded in Table 3. These values are negative, indicating spontaneous adsorption of the inhibitor on the surface of mild steel [28-29].

In order to further support the adsorption behaviour of the inhibitor on the surface of mild steel, IR spectroscopy was employed. Fig. 7 shows the IR spectrum of ethanol extract of *vernonia amygdalina* alone. Fig. 8 shows the IR spectrum of the corrosion product (without the inhibitor), while Fig. 9 shows the IR spectrum of the corrosion product when ethanol extract of *vernonia amygdalina* was used as an inhibitor. From Fig. 7, it is seen that the extract exhibited broad adsorption band at 3402.06 cm^{-1} (peak height = 56.880 cm), indicating the presence of alcohol or phenol functional group (i.e –OH). An

adsorption band was also found at 1046.01 cm^{-1} (peak height = 87.459 cm), suggesting the presence of $-\text{CO}$ stretch.

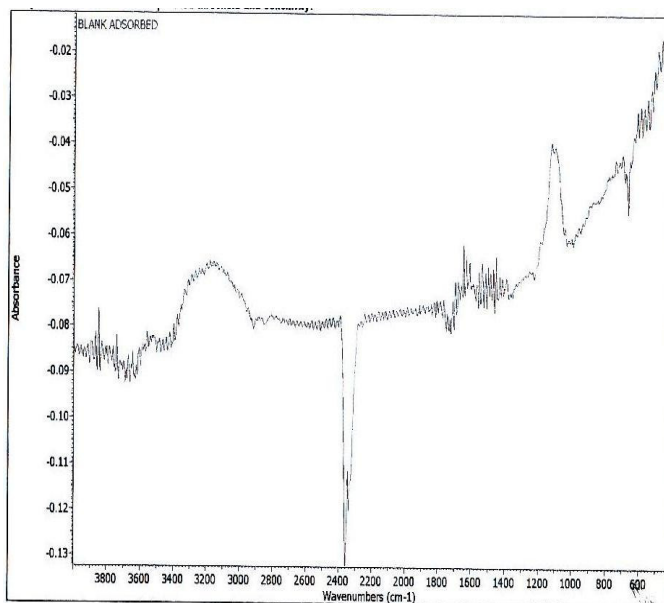


Figure 7. IR spectrum of the corrosion product (without the inhibitor).

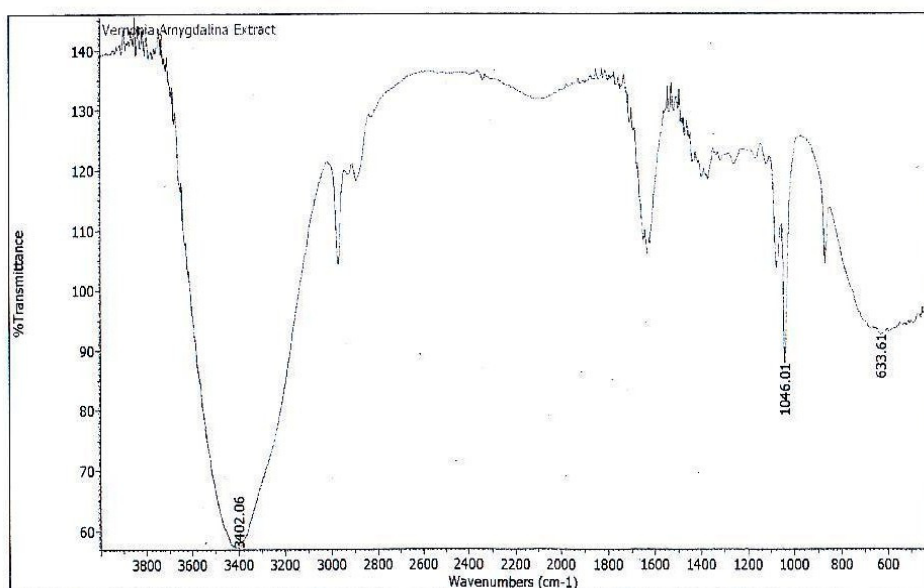


Figure 8. IR spectrum of ethanol extract of *vernonia amygdalina*.

The adsorption band at 633.61 cm^{-1} (peak height = 92.300 cm) suggests the presence of carbon-carbon triple bond (i.e., alkyne type of compound). However, in the presence of the inhibitor, IR spectrum of the corrosion product revealed that the $-\text{OH}$ stretch (3407.06 cm^{-1}) was shifted to 3435.22 cm^{-1} and the $\text{C}=\text{O}$ stretch (1046.01 cm^{-1}) was shifted to 1632.74 cm^{-1} , while $\text{C}-\text{H}$ bend (633.61 cm^{-1})

¹) was missing, indicating that there is interaction between the ethanol extract of *vernonia amygdalina* and the surface of mild steel.

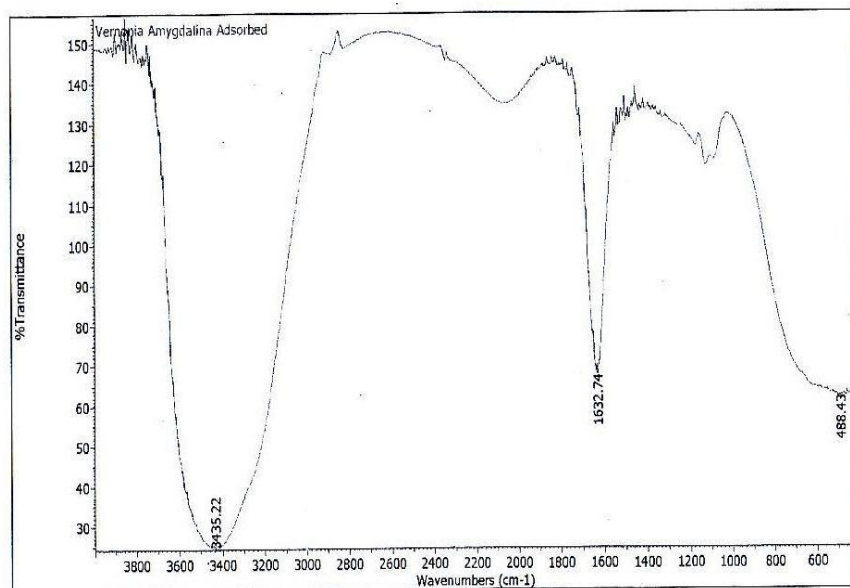


Figure 9. IR spectrum of the corrosion product showing adsorption in the presence of ethanol extract of *vernonia amygdalina*.

Phytochemical composition of aqueous and ethanol extract of vernonia amygdalina

The phytochemical composition of ethanol extract of *vernonia amygdalina* is shown in Table 4. The results indicated that the aqueous extract does not contain saponins, terpenes, tannins, flavanoids, phlobatannins, anthraquinones, cardiac glycoside and alkaloid, while the ethanol extract contains all of them, except flavanoid and phlobatannins. This indicates that the inhibition efficiency of the extract is due to the presence of some or all of the above listed phytochemical constituents. Eddy and Ebenso [1] also stated that saponins, tannins and alkaloids are active constituents of most green inhibitors.

Table 4. Phytochemical constituents of aqueous and ethanol extract of *vernonia amygdalina*.

Phytochemicals	Aqueous extract	Ethanol extract
Saponins	-	++
Terpenes	-	++
Tannins	-	++
Flavonoid	-	-
Phlobatannins	-	-
Anthraquinones	-	+++
Cardiac glycoside	-	++
Alkaloids	-	++

+++ = highly present, ++ = moderately present, - = absent or present in negligible quantity.

Conclusion

From the study, the following conclusions are made:

1. ethanol extract of *vernonia amygdalina* is a good inhibitor for the corrosion of mild steel in H₂SO₄;
2. the inhibitor acts by being adsorbed on the surface of mild steel according to classical Langmuir adsorption isotherm;
3. adsorption of ethanol extract of *vernonia amygdalina* on the surface of mild steel is spontaneous and occurs by physical adsorption.

Acknowledgement

The authors are grateful to Ndifreke Nde, Isangedihi Ating and S.A. Umoren for supporting the research. The authors are also grateful to Mrs. Edikan Nnabuk Eddy for typesetting the work.

References

1. N.O. Eddy and E.E. Ebenso, *Afri. J. Pure Appl. Chem.* 2 (2008) 046.
2. N.O. Eddy and S.A. Odoemelam, *Mat. Sci. (India)* 4 (2008) 9.
3. A. Al-Sehaibani, *Mater. Wissen. Werkst. Tech.* 31 (2000) 1060.
4. G.O. Avwiri and F.O. Igho, *Materials Letter* 57 (2001) 3705.
5. E. H. El Ashry, A. El Nemir, S. A. Esawy and S. Ragab, *Electrochimica Acta* 51 (2006) 3957.
6. A.Y. El-Etre and M. Abdallah, *Corros. Sci.* 42 (2000) 731.
7. P.C. Okafor, M.I. Ikpi, I.E. Uwah, E.E. Ebenso, U.J. Ekpe and S.A. Umoren, *Corros. Sci.* 50 (2008) 2310.
8. M.I. Awad, *J. Appl. Electrochem.* 36 (2006) 1163.
9. S.A. Odoemelam and N.O. Eddy, *J. Surf. Sci. Technol.* 24 (2008) 1.
10. S. Rajendran, S.V. Ganga, J. Arockiaselvi, and A.J. Amalraj, *Bull. Electrochem.* 21 (2005) 367.
11. M.G. Sethuran and P.B. Raja, *Pigment & Resin Technol.* 34 (2006) 327.
12. E.E. Ebenso, *Bull. Electrochem.* 19 (2003) 209.
13. S.A. Odoemelam and N.O. Eddy, *J. Surf. Sci. & Technol.* 24 (2008) 1.
14. E.U. Onyeka and I.O. Nwabekwe, *Nigerian Food J.* 25 (2007) 67.
15. O.K. Abiola, N.C. Oforka and E.E. Ebenso, *JCSE.* 5 (2004) 1.
16. N.O. Eddy and A.S. Ekop, *Mat. Sci.* 4 (2008) 10.
17. P. Atkins, *Physical Chemistry*, 7th edition, Oxford: Oxford press (2002).
18. K.K. Sharma and L.K. Sharma, *A textbook of physical chemistry*, 4th revised ed., India: Vikas Pub. House. PVT Ltd. (2004).
19. M. Abdallah, *Corrosion Sci.* 46 (2004) 1981.
20. S. Acharya and S.N. Upadhyay, *Trans. Indian Inst. Met.* 57 (2004) 297.
21. Y.K. Agrawal, J.D. Talati, M.D. Shah, M.N. Desai and N.K. Shah, *Corrosion Sci.* 46 (2003) 633.

22. H. Ashassi-Sorkhabi, B. Shaabani and D. Seifzadeh, *Appl. Surf. Sci.* 239 (2005) 154.
23. A. Yurt, G. Bereket, A. Rivrak, A. Balaban and B. Erk, *J. Appl. Electrochem.* 35 (2005) 1025.
24. S.A. Umoren, E.E. Ebenso, P.C. Okafor and O. Ogbode, *Pigment & Resin Technol.* 35 (2006a) 346.
25. S.A. Umoren, I.B. Obot, E.E. Ebenso, P.C. Okafor, O. Ogbobe and E.E. Oguzie, *Anti-Corrosion Methods and Materials* 53 (2006b) 277.
26. S.A. Umoren, O. Ogbobe, E.E. Ebenso and U.J. Ekpe, *Pigment & Resin Technol.* 35 (2006c) 284.
27. E.E. Oguzie, *Pigment and Resin Technol.* 34 (2005) 321.
28. E.E. Ebenso, U.J. Ekpe, E. Jackson, O.K. Abiola and N.C. Oforika, *J. Appl. Polym. Sci.* 100 (2004) 2889.
29. N.O. Eddy, A.O. Odiongenyi and S.A. Odoemelam, *Adv. in Nat. & Appl. Sci.* 2 (2008) 35.
30. A. Mann, M. Gbate and A.N. Umar, *Medicinal and Economic Plants of Nupeland*. Jube-Evans Books Publication. Bida, Nigeria.
31. L.S. Gill, *Ethnomedical uses of plants in Nigeria*. UNIBEN press, Benin city, Nigeria.