

Coupling the modules of EMT and stemness: A tunable 'stemness window' model

Mohit Kumar Jolly^{1,3,*}, Dongya Jia^{1,2,*}, Marcelo Boareto^{1,7}, Sendurai A. Mani⁸, Kenneth J. Pienta^{9,10,11,12}, Eshel Ben-Jacob^{1,5,6,†}, Herbert Levine^{1,3,4,5}

¹Center for Theoretical Biological Physics, Rice University, Houston, TX 77005-1827, USA

²Graduate Program in Systems, Synthetic and Physical Biology, Rice University, Houston, TX 77005-1827, USA

³Department of Bioengineering, Rice University, Houston, TX 77005-1827, USA

⁴Department of Physics and Astronomy, Rice University, Houston, TX 77005-1827, USA

⁵Department of Biosciences, Rice University, Houston, TX 77005-1827, USA

⁶School of Physics and Astronomy and The Sagol School of Neuroscience, Tel-Aviv University, Tel-Aviv 69978, Israel

⁷Institute of Physics, University of Sao Paulo, Sao Paulo 05508, Brazil

⁸Department of Translational Molecular Pathology, and Metastasis Research Center, The University of Texas MD Anderson Cancer Center, Houston, TX 77030, USA

⁹The James Brady Urological Institute, Johns Hopkins School of Medicine, Baltimore, MD 21287, USA

¹⁰Departments of Urology, Johns Hopkins School of Medicine, Baltimore, MD 21287, USA

¹¹Department of Oncology, Johns Hopkins School of Medicine, Baltimore, MD 21287, USA

¹²Department of Pharmacology and Molecular Sciences, Johns Hopkins School of Medicine, Baltimore, MD 21287, USA

*These authors have contributed equally to this work

†Deceased on June 5, 2015.

Correspondence to:

Herbert Levine, **e-mail:** herbert.levine@rice.edu

Keywords: *partial EMT, stemness window, cancer stem cells, OVOL, multistability*

Received: June 02, 2015

Accepted: July 10, 2015

Published: July 23, 2015

ABSTRACT

Metastasis of carcinoma involves migration of tumor cells to distant organs and initiate secondary tumors. Migration requires a complete or partial Epithelial-to-Mesenchymal Transition (EMT), and tumor-initiation requires cells possessing stemness. Epithelial cells (E) undergoing a complete EMT to become mesenchymal (M) have been suggested to be more likely to possess stemness. However, recent studies suggest that stemness can also be associated with cells undergoing a partial EMT (hybrid E/M phenotype). Therefore, the correlation between EMT and stemness remains elusive. Here, using a theoretical framework that couples the core EMT and stemness modules (miR-200/ZEB and LIN28/let-7), we demonstrate that the positioning of 'stemness window' on the 'EMT axis' need not be universal; rather it can be fine-tuned. Particularly, we present OVOL as an example of a modulating factor that, due to its coupling with miR-200/ZEB/LIN28/let-7 circuit, fine-tunes the EMT-stemness interplay. Coupling OVOL can inhibit the stemness likelihood of M and elevate that of the hybrid E/M (partial EMT) phenotype, thereby pulling the 'stemness window' away from the M end of 'EMT axis'. Our results unify various apparently contradictory experimental findings regarding the interconnection between EMT and stemness, corroborate the emerging notion that partial EMT associates with stemness, and offer new testable predictions.

INTRODUCTION

Metastasis and tumor relapse remain clinically insurmountable and claim more than 90% of cancer-related deaths [1]. It is believed that metastasis begins when some cancer cells of the primary tumor undergo an Epithelial-to-Mesenchymal Transition (EMT) and migrate towards blood vessels. These metastatic cells transit in the bloodstream as Circulating Tumor Cells (CTCs) and then exit at distant organs. There they may undergo the reverse of EMT, a Mesenchymal-to-Epithelial Transition (MET) and grow into secondary tumors [2]. On the other hand, tumor relapse is thought to be caused by therapy-resistant Cancer Stem Cells (CSCs) that can repopulate a tumor. Previous experimental studies have shown that these cell-fate decisions of phenotypic transition (EMT/MET) and gaining stem cell properties (stemness) are interconnected via underlying gene regulatory networks [3]. However, the basic principles of this interconnection remain enigmatic, hence limiting major therapeutic advances.

The decisions of EMT/MET and attaining stemness are much more flexible than imagined earlier—neither is EMT/MET a binary process nor is stemness a fixed inherent trait of a few cells. Cells undergoing EMT/MET can attain a hybrid epithelial/mesenchymal (E/M) phenotype that has combined epithelial (cell-cell adhesion) and mesenchymal (motility) traits. These combined traits enable them to migrate collectively, as observed in cluster migration of CTCs in the lung, prostate and breast cancer patients [4–7]. Also, CSCs and non-CSCs can interconvert and maintain a dynamic equilibrium among themselves [8–11]. Such plasticity blurs the direct one-to-one correlation between a complete EMT and an increased likelihood of being a CSC as postulated in earlier work [12–14]. As we will discuss below, this blurring can allow hybrid E/M (partial EMT) as well as epithelial cells to gain stemness, as observed in recent experimental studies [15–20].

Deciphering the EMT-stemness interplay requires a rigorous analysis of the decision-making modules of EMT and stemness, and the coupling between them. EMT decision-making is governed by a mutually inhibitory feedback loop of miR-200/ZEB [21, 22], such that epithelial cells have (high miR-200, low ZEB), mesenchymal cells have (low miR-200, high ZEB) and hybrid E/M cells have (medium miR-200, medium ZEB) [23, 24] (Figure 1A, Figure S1). On the other hand, stemness is regulated by a mutually inhibitory loop of LIN28/let-7 [25] that can allow three states - D (Down - low LIN28, high let-7), U (Up - high LIN28, low let-7) and D/U (Down/Up - medium LIN28, medium let-7) [18]. LIN28 activates the pluripotency marker OCT4 [26]. Both very high and very low levels of OCT4 lead to loss of stem cell properties (stemness), therefore OCT4 levels must be within a range of intermediate levels to

acquire or maintain stemness [27–30]. Such intermediate levels of OCT4 are generally attained by medium levels of LIN28 or equivalently D/U state, therefore, the D/U state usually associates with stemness [18]. Consequently, the phenotype(s)—E (no EMT), M (complete EMT), and E/M (partial EMT)—that can have medium OCT4 (or corresponding LIN28) levels can gain stemness, or in other words, lie in a ‘stemness window’.

A key player that has been reported independently both in regulating EMT and stemness, and is therefore, well-positioned to tune the EMT-stemness interplay is the transcription factor OVOL. OVOL is a well-studied regulator of embryogenesis that is involved in the differentiation of epidermal progenitor cells [31, 32] and is self-inhibitory [33]. In a developmental example of EMT—the case of mammary morphogenesis—OVOL is expressed in terminal end bud (TEB) cells that migrate collectively forming finger-like projections [34]. It can maintain TEB cells in a hybrid E/M phenotype and operate as a ‘critical molecular brake on EMT’ by preventing the ‘TEB cells that have gained partial plasticity’ from undergoing complete EMT [34]. Also, in a pathological EMT context, OVOL can drive MET by forming a double negative feedback loop with ZEB [35]. Collectively, coupling miR-200/ZEB with OVOL significantly expands the range of parameters or physiological conditions for the existence of the hybrid E/M phenotype—such that without OVOL, E/M phenotype can only exist in combination with the mesenchymal phenotype (the possible phases are {E, E/M, M} and {E/M, M}), however, with OVOL, the E/M phenotype can either exist alone or co-exist with the epithelial phenotype (phases {E/M} and E, E/M}) (Figure 1B-1D, ref. [36]). However, the mechanism by which OVOL influences the EMT-stemness interplay remains elusive.

Here, we present a theoretical approach to investigate how OVOL modulates EMT-stemness interplay. First, we investigate the coupled dynamics of EMT and stemness circuits—(miR-200/ZEB) and (LIN28/let-7) respectively—to elucidate how different relative strengths of the two links coupling these two circuits—miR-200 inhibiting LIN28 (referred to as ‘feed-forward coupling’ hereafter) and let-7 inhibiting ZEB (referred to as ‘feed-backward coupling’ hereafter)—affect EMT-stemness interplay. Further, we couple OVOL to this combined network, and compare the EMT-stemness correlation obtained with that obtained for the combined network without OVOL. We find that OVOL can enable cells in the epithelial and hybrid E/M states, but not the mesenchymal one, to gain stemness, i.e. it prevents the ‘stemness window’ from sliding completely towards the M end of the axis. Consequently, inhibition or loss of OVOL can allow the cells that undergo a complete EMT (M phenotype) to gain stemness. Our study serves two purposes—it presents an example of how various coupling factors may influence the EMT-stemness correlation, and it highlights the specific effects of OVOL as an example of a modulating factor.

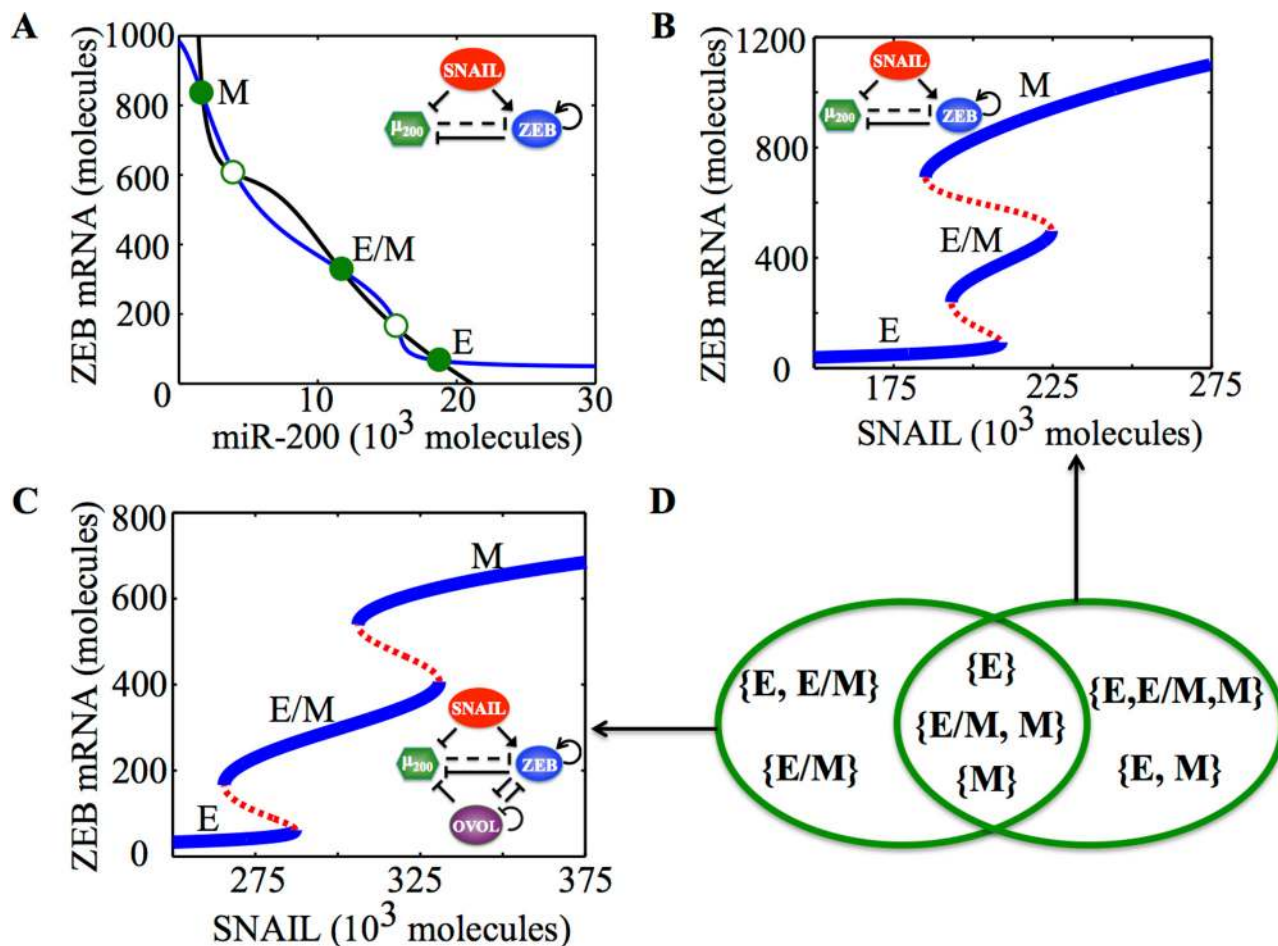


Figure 1: Dynamical system characteristics of the EMT decision-making circuit - (miR-200/ZEB). **A.** Nullclines of (miR-200/ZEB) circuit for SNAIL = 200×10^3 molecules representing three steady states: Epithelial (E—high miR-200, low ZEB), hybrid Epithelial/ Mesenchymal (E/M—medium miR-200, medium ZEB) and Mesenchymal (low miR-200, high ZEB). Solid green dots represent stable states; hollow green dots represent unstable states. **B.** Bifurcation levels of mRNA levels of ZEB in response to SNAIL for miR-200/ZEB circuit. **C.** Bifurcation levels of mRNA levels of ZEB in response to SNAIL for miR-200/ZEB/OVOL circuit. Different phenotypes obtained at different levels of SNAIL are labeled alongside. **D.** Comparing the different phases (co-existence of different phenotypes) obtained for miR-200/ZEB in (B) and miR-200/ZEB/OVOL circuits in (C).

RESULTS

Coupling the decision-making modules of EMT and stemness

The EMT and stemness decision-making modules—(miR-200/ZEB) and (LIN28/let-7)—are driven by SNAIL and NF- κ B respectively (Figure 2A, 2B). These inputs govern which phenotypes or states a cell can adopt, even in the absence of any coupling. These modules are connected via two links: miR-200 inhibiting LIN28 ('feed-forward coupling') and let-7 inhibiting ZEB ('feed-backward coupling'). The strengths of these couplings are represented by variables α_1 and α_2 . Previously, we demonstrated that in the absence of any 'feed-backward coupling' (inhibition of ZEB by let-7), both the hybrid E/M and M phenotypes can gain stemness, with the hybrid E/M being more likely to do so [18]. As the

first step to decipher the full EMT-stemness interplay, we include here the 'feed-backward coupling' from let-7 to ZEB and calculate the total number of stable steady states of the combined network (miR-200/ZEB/ LIN28/let-7) at different values of (α_1, α_2).

We start with the case when both (miR-200/ZEB) and (LIN28/let-7) are stand-alone (i.e. without any coupling, $\alpha_1 = \alpha_2 = 0$) tristable circuits, i.e. the input parameters are set such that cells can attain any of the three phenotypes—E, E/M and M—and each of them independently can be associated with any of the three states of the LIN28/let-7 circuit—D, D/U, and U—thus leading to a total of $3 \times 3 = 9$ stable states. Each of these stable states must be defined by a set of two variables, for example, the levels of LIN28 and levels of miR-200, as shown in the phenotypic map (Figure 2C). As either the 'feed-forward coupling' or 'feed-backward coupling' gets stronger, or in other words, as either α_1 (strength

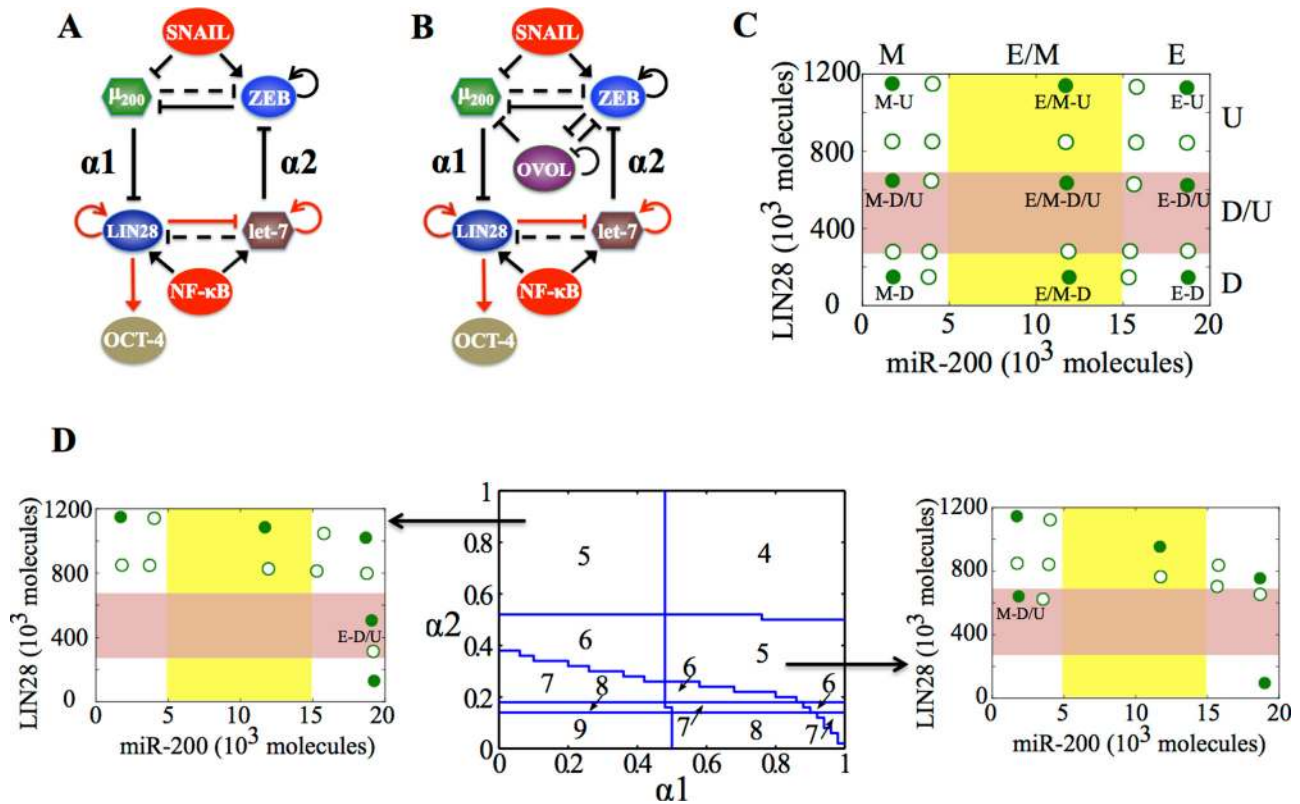


Figure 2: Coupling the decision-making circuits for EMT and stemness. **A.** Regulatory network showing coupling of miR-200/ZEB circuit with LIN28/let-7 circuit. **B.** Regulatory network showing coupling of miR-200/ZEB/LIN28/let-7 circuit with the transcription factor OVOL. A black solid arrow denotes transcriptional activation, and a black solid bar denotes transcriptional inhibition. Black dashed line indicates microRNA-mediated regulation, and red solid arrows imply non-transcriptional activation. α_1 and α_2 are coupling parameters between the EMT circuit (miR-200/ZEB) and the stemness circuit (LIN28/let-7) that vary from 0 to 1. The larger the value of α_1 , the stronger the inhibition of LIN28 by miR-200; and the larger the value of α_2 , the stronger the inhibition of ZEB by let-7. **C.** Phenotypic map representing all possible steady states of the coupled circuit (miR-200/ZEB/LIN28/let-7) for SNAIL = 200×10^3 molecules and NF- κ B = 25×10^3 molecules, at $\alpha_1 = \alpha_2 = 0.05$ (very weak ‘feed-forward’ and ‘feed-backward’ coupling). Red shaded region defines the ‘stemness window’ based on relative LIN28 levels or equivalently relative OCT4 levels, and yellow shaded region represents range of miR-200 levels for the existence of the hybrid E/M phenotype, as noted in [23] for (miR-200/ZEB) circuit. **D.** (center) Phase diagram representing the number of stable steady states of the coupled circuit (miR-200/ZEB/LIN28/let-7) for varying values of (α_1 , α_2) at SNAIL = 200×10^3 , and NF- κ B = 25×10^3 molecules. (left) Stable states of the coupled circuit at $\alpha_1 = 0.3$ and $\alpha_2 = 0.9$. (right) Stable states of the coupled circuit at $\alpha_1 = 0.9$ and $\alpha_2 = 0.3$. Green solid dots represent stable states, and green hollow dots are for unstable states. The labels ‘E-D/U’ (left) and ‘M-D/U’ (right) denote that epithelial and mesenchymal phenotypes respectively lie in stemness window.

of inhibition of LIN28 by miR-200) or α_2 (strength of inhibition of ZEB by let-7) increases, the number of total stable states decreases, indicating that some stable states can coalesce at different coupling strengths (α_1 , α_2) (Figure 2D, middle).

We found that even when the total number of stable states of the coupled circuit (miR-200/ZEB/ LIN28/let-7) is the same, different phenotype(s) can gain stemness, depending on exact values of strengths of ‘feed-forward coupling’ and ‘feed-backward coupling’, or equivalently (α_1 , α_2) (Figure 2D, left, right). The likelihood of gaining stemness of a phenotype is defined on the basis of relative OCT4 levels (relative to the saturation level of OCT4 when it is activated by a threshold level of LIN28). If that relative OCT4 level is between 0.25–0.65, the corresponding phenotype is highly likely to gain stemness or in other words, it lies in the ‘stemness window’.

This definition of a ‘stemness window’ is based on experimental observations that both too high and too low levels of OCT4 can lead the cell to differentiate, hence medium OCT4 levels (usually corresponding to D/U [18]) correspond to pluripotency or stemness [27–30]. This range of OCT4 levels defining the ‘stemness window’ can be tumor-specific. The results presented here are for this hypothesized range and serve to demonstrate the overall concept.

To investigate further how ‘feed-forward coupling’ (miR-200 inhibiting LIN28) and ‘feed-backward coupling’ (let-7 inhibiting ZEB) affect which phenotypes—E, M and E/M—gain stemness, we calculated the stable states of the system at different values of (α_1 , α_2) and plotted the ‘stemness region’ for each phenotype, i.e. range of values of (α_1 , α_2) for which each phenotype can gain stemness. For some range of the values of (α_1 , α_2),

more than one phenotype can gain stemness, or in other words, the ‘stemness regions’ of the three phenotypes (E, M and E/M) can overlap among themselves. For instance, at low strengths of both ‘feed-forward coupling’ and ‘feed-backward coupling’ (low α_1 , low α_2), all three phenotypes can gain stemness (black shaded region in Figure 3A), indicating that the ‘stemness window’ can cover the ‘EMT axis’, thereby enabling a rich phenotypic plasticity for cells that gain stemness. However, as either of the coupling strengths increase, this plasticity is restricted. Specifically, the epithelial (E) phenotype can be associated with stemness only at weak ‘feed-forward coupling’ (low α_1), irrespective of the strength of ‘feed-backward coupling’ (α_2) (green shaded region in Figure 3A). Conversely, the mesenchymal (M) phenotype can be associated with stemness only at weak ‘feed-backward coupling’ (low α_2), irrespective of the strength of ‘feed-forward coupling’ (α_1) (red and violet shaded regions in Figure 3A).

Therefore, the relative strength of these two coupling links govern exactly where the ‘stemness window’ lies on the ‘EMT axis’; strong ‘feed-forward coupling’ (high α_1 ; strong inhibition of miR-200 on LIN28) push it towards the M end of the axis, while strong ‘feed-backward coupling’ (high α_2 ; strong inhibition of let-7 on ZEB) push it towards the E end (Figure 3B, 3C). Our results reflect that the position of ‘stemness window’ on EMT axis is not universal, but rather coupling-dependent; hence cells in all three phenotypes—E, M and E/M—can possibly gain stemness depending on the relative levels of coupling strengths (α_1 , α_2) between the EMT and stemness decision-making modules - (miR-200/ZEB) and (LIN28/let-7).

Next, we choose a different value of SNAIL such that at no coupling ($\alpha_1 = \alpha_2 = 0$), cells are all in the {M} phase, but can associate with any of the three states of (LIN28/let-7) circuit, i.e. hence a total $3 * 1 = 3$ of stable states. Here, as α_1 and α_2 increase, total number of

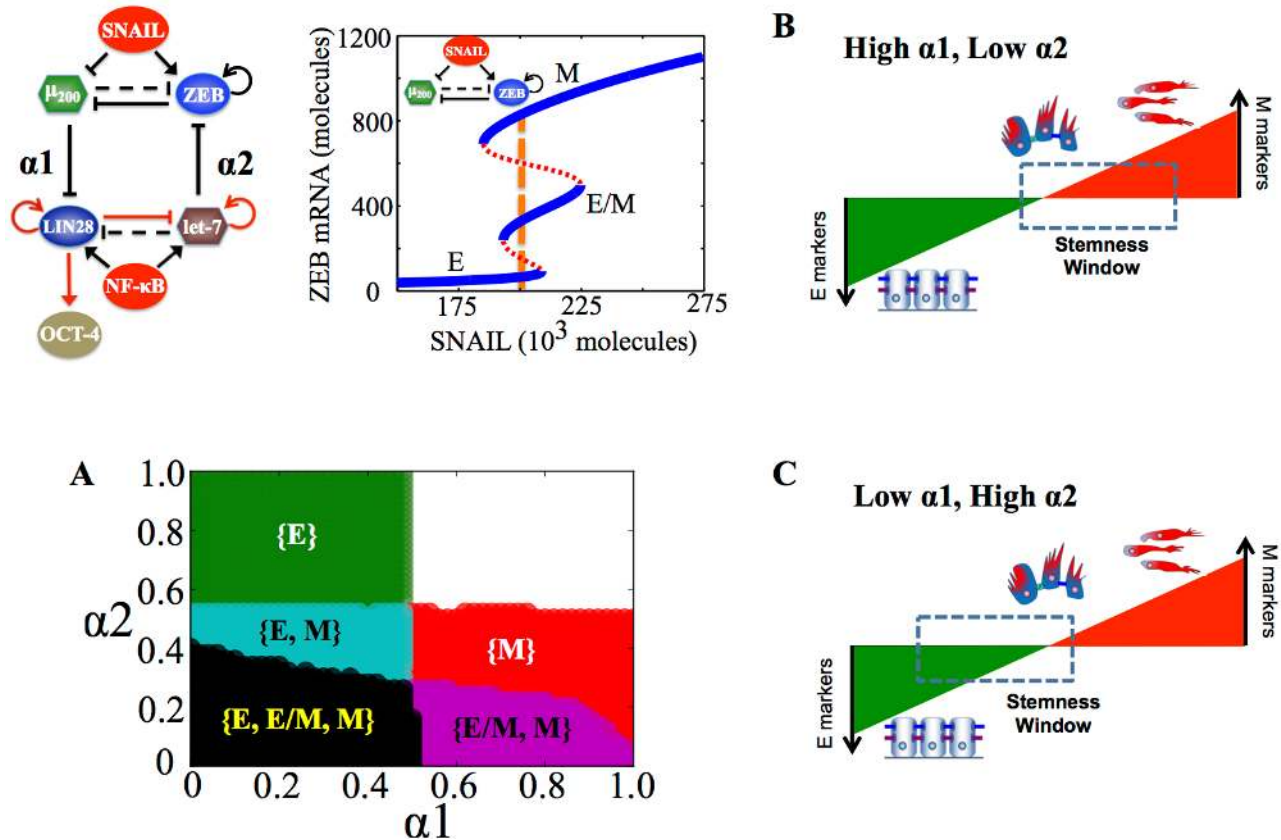


Figure 3: Stemness regions for different phenotypes under various (α_1 , α_2), when cells are in {E, E/M, M} phase at $\alpha_1 = \alpha_2 = 0$. A. Phase diagram of the circuit representing the values of (α_1 , α_2) for which the different phenotypes—E, M and E/M—can lie in stemness window respectively, for SNAIL = $200 * 10^3$ molecules and NF-kB = $25 * 10^3$ molecules. Areas with different colors represent different combinations of phenotypes than can gain stemness for that range of (α_1 , α_2). B, C. Model for tunable ‘stemness window’ on the ‘EMT axis’, such that (high α_1 , low α_2) allows for stemness to be associated with {E/M, M} and {M}, pushing the ‘stemness window’ towards the M end of the EMT axis, and (low α_1 , high α_2) allows for stemness to be associated with {E}, pushing the ‘stemness window’ towards the E end of the EMT axis. The results presented here are for the circuit presented here, and SNAIL levels driving this circuit are marked by a dashed line in bifurcation diagram presented here.

stable states increases, and again, this total number is not sufficient to determine which phenotype(s) correlate with stemness; as before, this correlation depends on the exact values of (α_1 , α_2) (Figure S2, S3).

Generally, an increase or decrease in number of states shows that in general, increasing coupling between the two decision-making circuits; (miR-200/ZEB) and (LIN28/let-7) creates non-trivial associations between the states of EMT circuit and those of stemness circuit, therefore regulating which combination of phenotype(s) gain stemness. These results demonstrate that the set of phenotypes that can gain stemness, or in other words, the positioning of the ‘stemness window’ on the ‘EMT axis’, depends at least on two factors: (a) the relative strength of coupling links between EMT and stemness decision-making modules, and (b) the external input signals on EMT circuit, such as SNAIL that governs the different set of phenotypes a cell can attain—E, M, and E/M.

OVOL precludes the mesenchymal phenotype from gaining stemness

Aside from the coupling strengths, it is expected that other factors may help determine the ‘context’ for the coupled EMT-stemness circuits. To show the effect of one such factor, we focus on the transcription factor OVOL and investigate how OVOL affects the correlation between EMT and stemness. To proceed, we analyze the coupled circuits without OVOL (miR-200/ZEB/LIN28/let-7) and with OVOL (miR-200 /ZEB/LIN28/let-7/OVOL) for varying values of SNAIL, such that without any coupling ($\alpha_1 = \alpha_2 = 0$), cells can attain either only the mesenchymal phenotype ($\{M\}$) or one of the two phenotypes: mesenchymal or partial EMT ($\{E/M, M\}$).

We start with the case of high levels of SNAIL such that at $\alpha_1 = \alpha_2 = 0$, all cells are in a mesenchymal (M) phenotype and can gain stemness (monostable phase $\{M\}$ at $\alpha_1 = \alpha_2 = 0$ in Figure 4A). We investigate the behavior at different values of α_1 and α_2 , and plot ‘stemness regions’ for the phenotypes E, M, and E/M. In absence of OVOL, at a strong inhibition of ZEB by let-7 or ‘feed-backward coupling’ ($\alpha_2 > 0.7$), ZEB is suppressed strongly so as to allow the existence of the hybrid E/M phenotype in addition to the M phenotype and both of them lie in the ‘stemness window’ (Figure 4A, 4B). In the presence of OVOL, both these phenotypes—hybrid E/M and M—can gain stemness at a relatively weaker inhibition of ZEB by let-7 ($\alpha_2 > 0.2$), probably because OVOL is also inhibiting ZEB (Figure 4C). Further, at a strong ‘feed-backward coupling’ ($\alpha_2 > 0.6$), the M phenotype can no longer gain stemness, and only the hybrid E/M and E phenotypes lie in the ‘stemness window’ (Figure 4C, 4D). Also, at slightly higher levels of SNAIL, in the absence of OVOL, the only phenotype that lies in the stemness window is mesenchymal (M), but in presence of OVOL, the only phenotype that can gain stemness is the

hybrid E/M phenotype (Figure S4). These results suggest that at strong ‘feed-backward coupling’, OVOL can drive the M phenotype out of the ‘stemness window’.

In the absence of OVOL, for a given range of α_1 and α_2 , all the three phenotypes (E, M, and E/M) can gain stemness, representing a very flexible positioning of ‘stemness window’ all over the ‘EMT axis’. In other words, there can exist a rich phenotypic plasticity for the stem cells, as cellular stochastic fluctuations can cause motility transitions without necessarily interfering with a cell’s stemness. In the presence of OVOL, however, this plasticity is significantly curtailed as there is no tristable $\{E, E/M, M\}$ phase overlapping with the stemness region (compare the black area in Figure 4C vs that in Figure 4A).

A different value of SNAIL can be chosen, such that without coupling, cells can attain either a mesenchymal or partial EMT phenotype (the $\{E/M, M\}$ phase at $\alpha_1 = \alpha_2 = 0$ in Figure 5A). In the absence of OVOL, a large range of values of (α_1 , α_2) allows any of the three phenotypes (E, M, and E/M) to gain stemness, thus representing an enriched plasticity for stem cells as observed earlier too. However, in the presence of OVOL, this tristable phase $\{E, M, E/M\}$ disappears (compare the black area in Figure 5C vs that in Figure 5A). Further, in absence of OVOL, none of the three phenotypes can lie in the stemness window at strong coupling ($\alpha_1 = \alpha_2 = 1$) (white area in Figure 5A), however, at the same values of α_1 and α_2 , both hybrid E/M and E phenotypes can gain stemness after including OVOL (Figure 5A-5D).

Consistently, in stemness region diagrams, the presence of OVOL allows for a broad range of (α_1 , α_2) values that enables an exclusive and exhaustive association of the hybrid E/M phenotype with stemness, a feature not observed for the circuit without OVOL (compare the blue area in Figure 5C vs that in Figure 5A). A significant stemness region also exists for the cells in the $\{E, E/M\}$ phase, i.e. cells both in epithelial and hybrid E/M phenotype can gain stemness (Figure 5B). Collectively, these results suggest that OVOL restricts the sliding of ‘stemness window’ towards the M end of the ‘EMT axis’, or in other words, it typically precludes the mesenchymal phenotype from gaining stemness.

Inhibition of OVOL allows mesenchymal phenotype to gain stemness

Next, we investigate the effect of OVOL in modulating EMT-stemness interplay, such that at no coupling ($\alpha_1 = \alpha_2 = 0$), all cells are in the hybrid E/M phenotype. Importantly, this $\{E/M\}$ phase is allowed only when OVOL is coupled to miR-200/ZEB and is not present in the absence of OVOL (Figure 1D, ref. [36]). Therefore, instead of comparing our results for the combined circuit without OVOL, here we consider the effects of an external signal controlling OVOL so as to better understand its role.

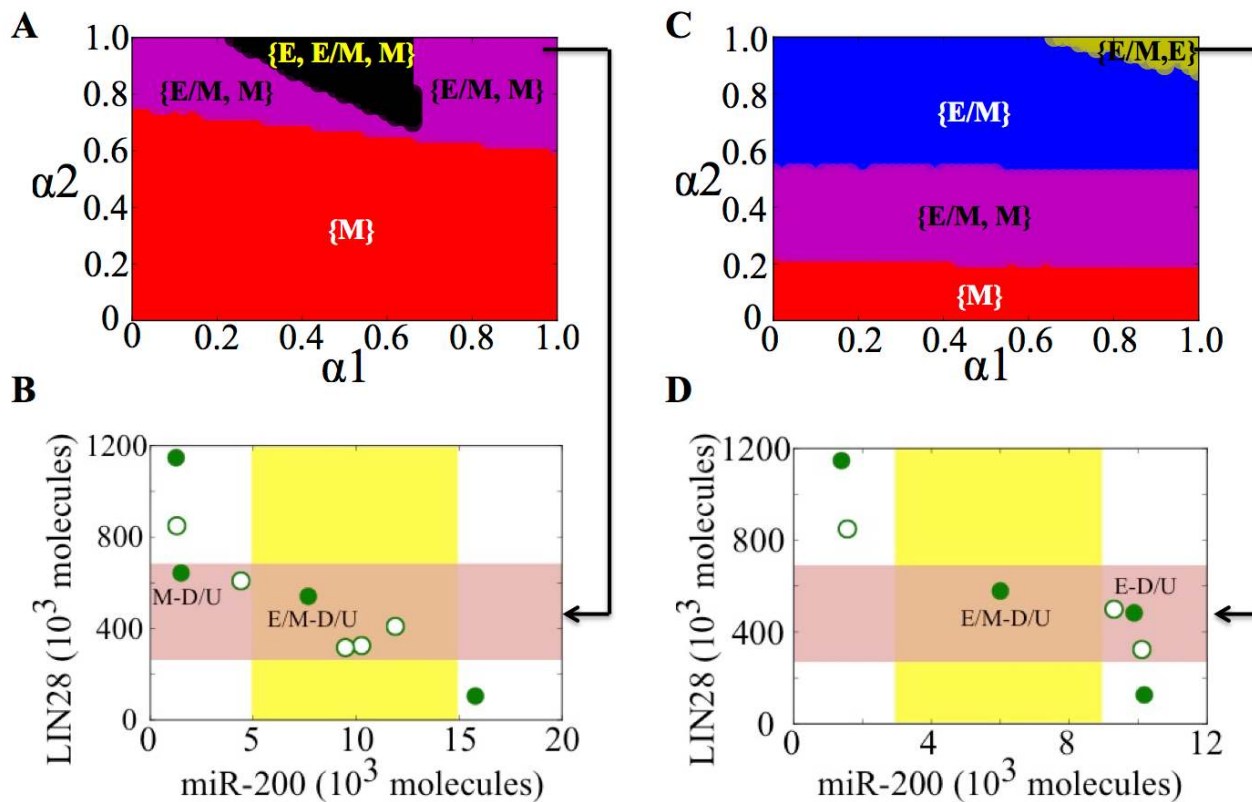
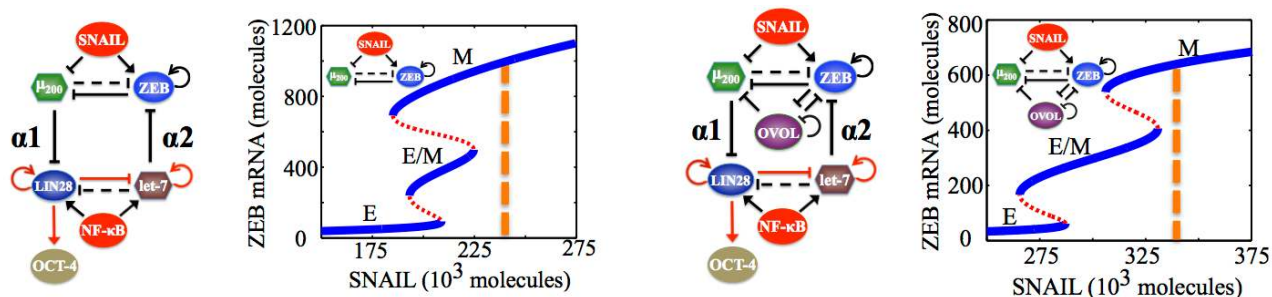


Figure 4: State-space characteristics of the coupled networks miR-200/ZEB/LIN28/let-7 and miR-200/ZEB/LIN28/let-7/OVOL, when cells are in {M} phase at $\alpha_1 = \alpha_2 = 0$. **A.** Phase diagram of the circuit miR-200/ZEB/LIN28/let-7 representing the values of (α_1, α_2) for which the different phenotypes can lie in stemness window, for SNAIL = 240×10^3 molecules and NF- κ B = 25×10^3 molecules. **B.** Phenotypic map of the coupled circuit at $\alpha_1 = \alpha_2 = 0.95$ and at external signals SNAIL = 240×10^3 molecules and NF- κ B = 25×10^3 molecules. Red shaded area shows the ‘stemness window’ based on relative OCT4 levels, and yellow shaded area represents the range of miR-200 levels for the existence of the hybrid E/M phenotype, as noted in [23] for (miR-200/ZEB) circuit and in [36] for (miR-200/ZEB/OVOL) circuit. **C, D.** represent a similar case for (A), (B) respectively but for the circuit with OVOL, therefore SNAIL = 340×10^3 molecules. Different colors represent different combinations of phenotypes that can gain stemness. Steady state diagram and phase diagram in every column are for the circuit drawn in the topmost row of that column, such that at $\alpha_1 = \alpha_2 = 0$ (no coupling between the EMT and stemness circuits), cells are in the phase marked by dashed lines in the bifurcation diagram drawn next to the circuit.

As before, we explore the effect of different values of coupling strengths (α_1, α_2) on the EMT-stemness coupling, by calculating ‘stemness regions’ for all three phenotypes over the entire range of values of the coupling parameters (α_1, α_2). At weak ‘feed-backward coupling’ (low α_2), only the hybrid E/M phenotype can gain stemness, however, as the strength of this coupling increases, ZEB is further

repressed so as to allow the existence of the epithelial phenotype in addition to hybrid E/M phenotype, and both these phenotypes lie in the ‘stemness window’ (Figure 6A). At $\alpha_1 = \alpha_2 = 1$, only the E phenotype can gain stemness, i.e. the ‘stemness window’ moves completely to E end of the ‘EMT axis’ as the levels of ZEB are suppressed strongly by let-7 (very strong ‘feed-backward coupling’)

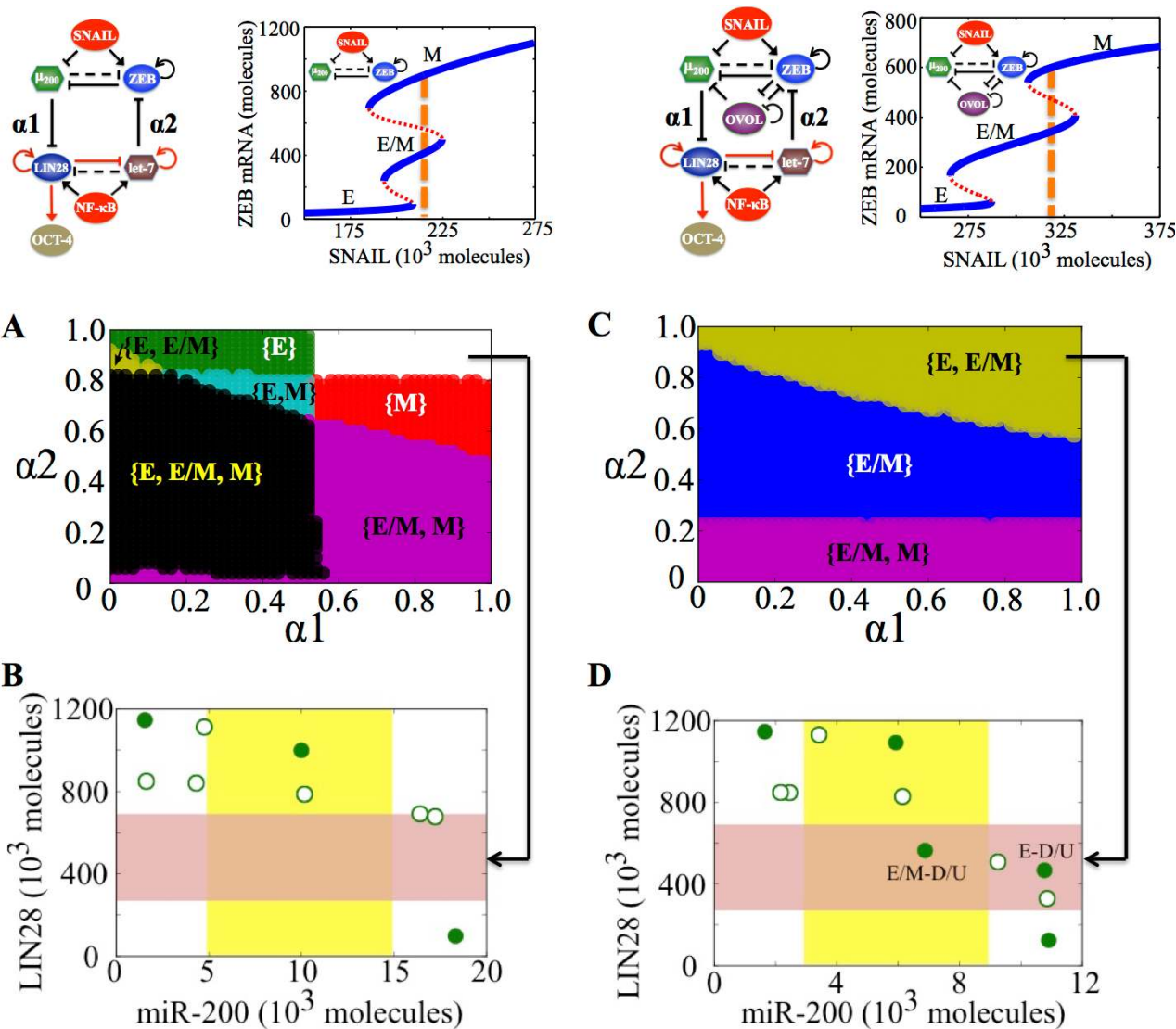


Figure 5: State-space characteristics of the coupled networks miR-200/ZEB/LIN28/let-7 and miR-200/ZEB/LIN28/let-7/OVOL, when cells are in {E/M, M} phase at $\alpha_1 = \alpha_2 = 0$. **A.** Phase diagram of the circuit miR-200/ZEB/LIN28/let-7 representing the values of (α_1, α_2) for which the different phenotypes can lie in stemness window, for SNAIL = 210×10^3 molecules and NF- κ B = 25×10^3 molecules. **B.** Phenotypic map of the coupled circuit at $\alpha_1 = \alpha_2 = 0.9$ and at external signals SNAIL = 210×10^3 molecules and NF- κ B = 25×10^3 molecules. Red shaded area shows the ‘stemness window’ based on relative OCT4 levels, and yellow shaded area represents the range of miR-200 levels for the existence of the hybrid E/M phenotype, as noted in [23] for (miR-200/ZEB) circuit and in [36] for (miR-200/ZEB/ OVOL) circuit. **C, D.** represent a similar case for (A), (B) respectively but for the circuit with OVOL, therefore SNAIL = 320×10^3 molecules. Different colors represent different combinations of phenotypes that can gain stemness. Steady state diagram and phase diagram in every column are for the circuit drawn in the topmost row of that column, such that at $\alpha_1 = \alpha_2 = 0$ (no coupling between the EMT and stemness circuits), cells are in the phase marked by dashed lines in the bifurcation diagram drawn next to the circuit.

(Figure 6A, 6C). Consistently, upon overexpression of OVOL (through an external activation signal), the E phenotype can lie in ‘stemness window’ for a broad range of coupling strengths (α_1, α_2) (Figure S5). Next, an external inhibition signal on OVOL is applied and the stemness regions for E, E/M, and M phenotypes are calculated. Inhibiting OVOL allows both M and E/M phenotypes to gain stemness without any coupling ($\{E/M, M\}$ phase at $\alpha_1 = \alpha_2 = 0$ in Figure 6B) as well as for

weak ‘feed-backward coupling’ ($\alpha_2 < 0.2$) irrespective of the strength of ‘feed-forward coupling’. Further, it reduces the range of (α_1, α_2) over which $\{E/M\}$ and $\{E, E/M\}$ phases can be associated with stemness, and enables the association of $\{E/M, M\}$ phase with stemness (Figure 6B), i.e. inhibition of OVOL allows the ‘stemness window’ to slide towards the M end of the ‘EMT axis’, especially with the ‘feed-forward coupling’ being stronger than the ‘feed-backward coupling’ (low α_2) (Figure 6C, 6D).

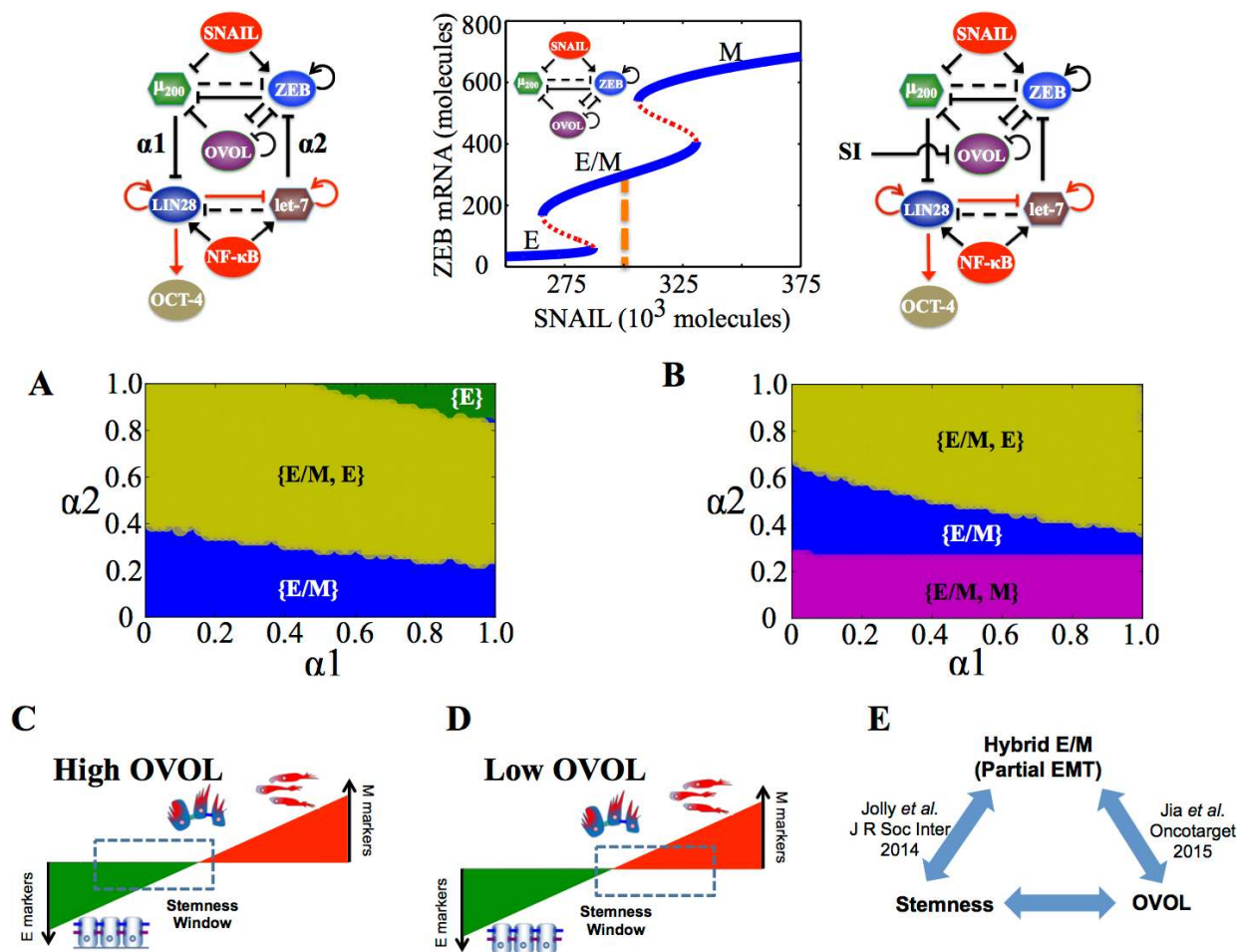


Figure 6: State-space characteristics of the miR-200/ZEB/let-7/OVOL circuit when the cells are in {E/M} phase at $\alpha_1 = \alpha_2 = 0$. A, B. Phase diagram of the circuit representing the values of (α_1, α_2) for which different phenotypes can attain stemness or lie in the ‘stemness window’, for SNAIL = 300×10^3 molecules and NF- κ B = 25×10^3 molecules. (A) denotes the case without inhibition signal on OVOL (as shown in the circuit above (A)) and (B) is for the circuit with inhibition signal on OVOL (as shown in the circuit above (B)). Both (A, B) represent the case such that cells are in {E/M} phase at $\alpha_1 = \alpha_2 = 0$ (as shown by dashed line in the bifurcation figure). C–D. Flexible ‘stemness window’ model showing that (C) high levels of OVOL can pull the ‘stemness window’ towards the E end of the axis, and (D) low levels can let it slide towards the M end of the axis. E. Cartoon showing the interconnections between partial EMT or hybrid E/M phenotype, stemness and the effect of OVOL (Jolly et al. [18] suggest that hybrid E/M phenotype is enriched in stemness, Jia et al. [36] propose that endogenous OVOL levels can expand the range of existence of the hybrid E/M phenotype, and results here show that endogenous levels of OVOL can enable an association of stemness with the hybrid E/M phenotype).

In summary, at a strong ‘feed-backward coupling’ and in presence of endogenous levels of OVOL, the M phenotype is precluded from gaining stemness. Consistently, if either OVOL is inhibited or the ‘feed-backward coupling’ (inhibition of ZEB by let-7) is weakened, ZEB levels can be maintained at high levels due to its self-activation, hence enabling the M phenotype as well as increasing its likelihood to gain stemness. Therefore, OVOL and the inhibition of ZEB by let-7 can act synergistically to preclude the complete EMT (or mesenchymal) phenotype from gaining stemness, therefore indicating a possible decoupling between complete EMT and stemness.

DISCUSSION

Coupling between EMT and stemness (or tumor-initiating properties) directly connects two of the most fatal aspects of cancer—metastasis and tumor relapse. The emergence of stemness when cells undergo a complete EMT has been studied at a phenomenological level [13, 14]; however, an understanding of how the underlying decision-making modules couple these traits together has largely remained underexplored. Here, using a theoretical framework modeling the connections between the decision-making modules of EMT (miR-200/ZEB) and stemness (LIN28/let-7), we reveal how the strengths of

the two links between these modules—inhibition of LIN28 by miR-200 (‘feed-forward coupling’) and inhibition of ZEB by let-7 (‘feed-backward coupling’)—determine which phenotype(s) gain stemness, or in other words, where the ‘stemness window’ lies on the ‘EMT axis’. We also investigate the role of OVOL, a transcription factor noted for its roles in regulating stemness and shepherding EMT/MET [31–39].

We find that relative strengths of ‘feed-forward coupling’ and ‘feed-backward coupling’ can affect the position of ‘stemness window’ on the ‘EMT axis’—strong inhibition of LIN28 by miR-200 pushes it towards mesenchymal (M) end of the axis, but strong inhibition of ZEB by let-7 pushes it towards the E end. miR-200 family that is highly expressed in epithelial phenotype [21, 22], has been experimentally shown to inhibit the gain of stemness [40]. However, the predicted role of let-7 needs further investigation.

We further show that OVOL modulates the positioning of the ‘stemness window’ on the ‘EMT axis’ in multiple ways: (a) it precludes the association of an entirely mesenchymal phenotype with stemness, (b) it significantly increases the likelihood for the hybrid E/M phenotype to gain stemness, and (c) it restricts the range of physiological conditions (or parameters) under which minor cellular stochastic fluctuations can cause motility transitions in stem cells, or in other words, it prevents the ‘stemness window’ from sliding all over the ‘EMT axis’. These results explain why the loss of OVOL may result in unchecked or aberrant plasticity of stem or progenitor cells [34, 41]. In developmental contexts, this unrestricted plasticity can impair the differentiation potential of stem or progenitor cells [41]; however, higher plasticity might be advantageous for tumor progression by potentially maximizing the range of conditions under which cells can gain stemness. Therefore, such enriched plasticity is more likely to be a hallmark of cancer but not necessarily that of homeostasis or development, thus offering some insight into possible differences in EMT during cancer metastasis (type III EMT) vs. EMT during embryonic development and tissue regeneration (type I, II EMT). This observation also offers a possible explanation of why the loss of OVOL correlates with poor survival [35].

Further, our results reveal that OVOL can not only ‘maintain’ the hybrid E/M phenotype [34, 36], but also largely associate it with stemness (Figure 6E). Based on its proposed effect in enabling collective migration of tumor cell clusters and conferring them with rich tumor-initiating properties, OVOL can be an important therapeutic target. Reducing OVOL levels could break apart clusters of CTCs in the circulation. These clusters (made of cells with the E/M phenotype) may be the key drivers of metastasis and ‘breaking’ them could deprive them of many advantages of cluster migration—resistance to anoikis, more tumor-initiating potential, ease of extravasation, and finally ‘priming’ for subsequent dissemination

[18, 42–44]. Recent diagnostic attempts have mostly focused on isolating individual CTCs [44], however, these results suggest that the most effective diagnostic approach would be to isolate and characterize clusters of CTCs.

Importantly, our study unifies many apparently contradictory experiments such as those indicating that a complete EMT leads to increased stemness [13, 14], MET associates with enhanced stemness [19, 20, 45], and partial EMT phenotype has the maximum stemness [17, 46–48]. We demonstrate that the ‘stemness window’ [46] is quite flexible or tunable on the ‘EMT axis’. Consequently, different phenotypes can gain stemness, either alone or in different combinations among themselves, depending on (a) strengths of ‘feed-forward’ and ‘feed-backward’ coupling between the EMT and stemness circuits, (b) levels of external signals to the circuits such as SNAIL, and (c) ‘phenotypic stability factors’ such as OVOL [49]. Because different cell lines operate in different regimes of coupling strengths and levels of SNAIL and OVOL, each of them is likely to have its own EMT-stemness coupling, slightly different from the other. In an actual tumor, different microenvironments could lead to spatially varying correlations between stemness and motility phenotypes, as recently indicated [15].

Aside from unifying many apparently contradictory studies concerning EMT-stemness interplay, our study also offers new testable predictions. For instance, we predict that the loss or inhibition of OVOL can promote the ability of mesenchymal (M) cells to gain stemness. Loss of OVOL has been shown to enable the transition of hybrid E/M cells to being completely mesenchymal [34], however, the expression of OVOL needs to be measured in mesenchymal stem cells for verifying its predicted role in enabling M cells to be stem-like. Importantly, we present here OVOL as one example of a modulating factor that can fine-tune the E/M state as well as acquisition of stemness. Such a role can also be potentially played by other pro-epithelial genes such as E-cadherin and P-cadherin [50]; the need for additional factors could arise, for instance, because the inhibition of ZEB by OVOL might be relevant in some biological contexts where EMT is operative [34, 35, 41], but not necessarily all. Given the emerging notion that most tumors *in vivo* undergo only a partial EMT [51], it would be important to identify other modulating factors that can fine-tune the hybrid E/M phenotype. The E/M phenotype, sometimes referred to as an ‘EMT-like’ state [48], also needs to be characterized further functionally in the contexts of both physiological and pathological EMT. Here, we characterize the motility phenotypes discretely (E, M and E/M) based on the tristable behavior of the core EMT circuit miR-200/ZEB, however, including other components in the model such as E-cadherin and P-cadherin that are regulated during EMT might fine-tune this characterization to appear more close to being a continuum of phenotypes [52, 53].

Similar to EMT, many other cellular decisions are not necessarily binary but rather ternary and governed by underlying three-way gene circuits. Dynamical characteristics of such three-way gene circuits have been theoretically investigated individually [54–57]. However, to the best of our knowledge, ours is the first computational study that couples two three-way genetic switches to elucidate the operating principles of coupled cellular decision-making. It is not yet clear that how many of our results depend on the specific details of the two circuits considered here. Hence, future studies using more generic models of tristable circuits are required. Further, to investigate the ‘underlying organizing principles’ [58] of coupled cellular decision-making in cancer, the decision-making circuits of EMT and stemness should be coupled with circuits regulating other hallmarks of cancer such as deregulated cellular metabolism [58–60].

With an increasing interest in decoding the signaling pathways [36, 61–68] underlying many hallmarks of cancer to elucidate their ‘underlying organizing principles’ [58], the theoretical approach presented here can serve as a basis for incorporating other intracellular and extracellular signals, and also aid in investigating the efficacy of different therapeutic strategies that target EMT and/or CSCs.

MATERIALS AND METHODS

Model formulation

Our coupled circuit model of EMT and stemness (miR-200/ZEB/LIN28/let-7) has six components - microRNA miR-200 (M), ZEB mRNA (Z), ZEB protein (Z_p), microRNA let-7 (L), LIN28 mRNA (L), and LIN28 protein (L_p)—and includes multiple modes of regulation—transcriptional (ZEB inhibiting miR-200, and self-activation of ZEB), microRNA-mediated (miR-200 inhibiting ZEB and LIN28, and let-7 inhibiting both LIN28 and ZEB), microRNA processing (LIN28’s inhibition of let-7 and self-activation of let-7) and translational (self-activation of LIN28). We generalized and extended the theoretical framework devised by Lu *et al.* [23] for the EMT circuit, and Jolly *et al.* [18] for the stemness circuit, and include (a) two coupling links, ‘feed-forward coupling’ (inhibition of LIN28 by miR-200), and ‘feed-backward coupling’ (inhibition of ZEB by let-7), and (b) coupling of miR-200/ZEB with OVOL. Except for miRNA-mediated regulation with a large number of binding sites of miRNA on its target mRNA (such as for miR-200 inhibiting ZEB with 6 binding sites), all other effects have been modeled via shifted Hill functions ($H^{S^+}(X, \lambda)$ for activation and $H^{S^-}(X, \lambda)$ for inhibition). Shifted Hill functions are defined as the weighted sum of positive Hill function $H^+(X)$ and negative Hill function $H^-(X)$

$$H^{\pm}(X) = \frac{\lambda H^{\pm}(X) + 1}{\lambda H^{\pm}(X) + 1 + \alpha}$$

represents the fold-change in the production rate of Y due

to regulation by X. For activation, $\lambda > 1$; for inhibition, $\lambda < 1$; and for no effect, $\lambda = 1$ [23]. For the two links coupling the circuits (miR-200 inhibiting LIN28 and let-7 inhibiting ZEB), the strength of inhibition is denoted by $\alpha = 1 - \lambda$, $0 < \alpha < 1$; larger the value of α , stronger the inhibition, and thus shifted Hill function is denoted by .

Effects of miR-200 on ZEB include both degradation of mRNA and translational inhibition by miRNAs that can themselves be degraded after binding and forming complexes with the mRNAs [23].

SNAIL and NF- κ B are treated as external signals for EMT and stemness circuits respectively with their effects captured by shifted Hill functions. Similarly, including OVOL in our analysis entails including two more components: OVOL mRNA (O) and OVOL protein (O_p). The regulations involving OVOL and miR-200/ZEB loop as well as the external activation or inhibition signal on OVOL is also captured by shifted Hill functions. Details of the model construction for the miR-200/ ZEB/LIN28/let-7/OVOL and parameter values used in the model can be found in SI sections 1, 2, and 3 and Tables S1-5. The model is quite robust with respect to changes in parameter values as discussed in SI section 4 (Figure S1). Codes were implemented in Python using PyDSTool [69]. Figures 2 and S2 were drawn in MATLAB.

ACKNOWLEDGMENTS

We have benefitted from useful discussions with Mary C. Farach-Carson and Jose’ N. Onuchic. H.L. and E.B.-J. were supported by NSF Center for Theoretical Biological Physics (NSF grant no. PHY-1427654) and by the Cancer Prevention and Research Institute of Texas (CPRIT) at Rice University. E.B.-J. was also supported by a grant from the Tauber Family Funds and the Maguy-Glass Chair in Physics of Complex Systems at Tel Aviv University. S.A.M. was supported by a grant from the National Institutes of Health (5R01CA155243). K.J.P. was supported by grants from National Cancer Institute (U54CA143803, CA163124, CA093900 and CA143055). M.B. was also supported by FAPESP (Grant 2013/14388-8).

FUNDING

National Science Foundation (NSF grant no. PHY-1427654) Cancer Prevention and Research Institute of Texas (CPRIT) at Rice University Tauber Family Funds and the Maguy-Glass Chair in Physics of Complex Systems at Tel Aviv University National Institutes of Health (5R01CA155243) National Cancer Institute (U54CA143803, CA163124, CA093900 and CA143055) FAPESP (Grant 2013/14388-8)

CONFLICTS OF INTERESTS

The authors declare no conflict of interest.

REFERENCES

1. Gupta GP, Massagué J. Cancer metastasis: building a framework. *Cell*. 2006; 127:679–95.
2. Nieto MA. Epithelial plasticity: a common theme in embryonic and cancer cells. *Science*. 2013; 342:1234850.
3. May CD, Sphyris N, Evans KW, Werden SJ, Guo W, Mani SA. Epithelial-mesenchymal transition and cancer stem cells: a dangerously dynamic duo in breast cancer progression. *Breast Cancer Res*. 2011; 13:202.
4. Lecharpentier A, Vielh P, Perez-Moreno P, Planchard D, Soria JC, Farace F. Detection of circulating tumour cells with a hybrid (epithelial/mesenchymal) phenotype in patients with metastatic non-small cell lung cancer. *Br J Cancer*. 2011; 105:1338–41.
5. Hou J-M, Krebs M, Ward T, Sloane R, Priest L, Hughes A, Clack G, Ranson M, Blackhall F, Dive C. Circulating Tumor Cells as a Window on Metastasis Biology in Lung Cancer. *Am J Pathol*. 2011; 178:989–96.
6. Armstrong AJ, Marengo MS, Oltean S, Kemeny G, Bitting RL, Turnbull JD, Herold CI, Marcom PK, George DJ, Garcia-Blanco MA. Circulating tumor cells from patients with advanced prostate and breast cancer display both epithelial and mesenchymal markers. *Mol Cancer Res*. 2011; 9:997–1007.
7. Yu M, Bardia A, Wittner BS, Stott SL, Smas ME, Ting DT, Isakoff SJ, Ciciliano JC, Wells MN, Shah AM, Conchannon KF, Donaldson MC, Sequist L V, et al. Circulating breast tumor cells exhibit dynamic changes in epithelial and mesenchymal composition. *Science*. 2013; 339:580–4.
8. Wang W, Quan Y, Fu Q, Liu Y, Liang Y, Wu J, Yang G, Luo C, Ouyang Q, Wang Y. Dynamics between cancer cell subpopulations reveals a model coordinating with both hierarchical and stochastic concepts. *PLoS One*. 2014; 9:1–9.
9. Yang G, Quan Y, Wang W, Fu Q, Wu J, Mei T, Li J, Tang Y, Luo C, Ouyang Q, Chen S, Wu L, Hei TK, et al. Dynamic equilibrium between cancer stem cells and non-stem cancer cells in human SW620 and MCF-7 cancer cell populations. *Br J Cancer*. 2012; 106:1512–9.
10. Iliopoulos D, Hirsch HA, Wang G, Struhl K. Inducible formation of breast cancer stem cells and their dynamic equilibrium with non-stem cancer cells via IL6 secretion. *Proc Natl Acad Sci U S A*. 2011; 108:1397–402.
11. Gupta PB, Fillmore CM, Jiang G, Shapira SD, Tao K, Kuperwasser C, Lander ES. Stochastic state transitions give rise to phenotypic equilibrium in populations of cancer cells. *Cell*. Elsevier Inc. 2011; 146:633–44.
12. Turner C, Kohandel M. Investigating the link between epithelial-mesenchymal transition and the cancer stem cell phenotype: A mathematical approach. *J Theor Biol*. 2010; 265:329–35.
13. Mani SA, Guo W, Liao M-J, Eaton EN, Ayyanan A, Zhou AY, Brooks M, Reinhard F, Zhang CC, Shipitsin M, Campbell LL, Polyak K, Briskin C, et al. The epithelial-mesenchymal transition generates cells with properties of stem cells. *Cell*. 2008; 133:704–15.
14. Morel A-P, Lièvre M, Thomas C, Hinkal G, Ansieau S, Puisieux A. Generation of breast cancer stem cells through epithelial-mesenchymal transition. *PLoS One*. 2008; 3:e2888.
15. Liu S, Cong Y, Wang D, Sun Y, Deng L, Liu Y, Martin-Trevino R, Shang L, McDermott SP, Landis MD, Hong S, Adams A, D'Angelo R, et al. Breast cancer stem cells transition between epithelial and mesenchymal states reflective of their normal counterparts. *Stem Cell Reports*. 2014; 2:78–91.
16. Celià-Terrassa T, Meca-Cortés Ó, Mateo F, De Paz AM, Rubio N, Arnal-Estapé A, Ell BJ, Bermudo R, Díaz A, Guerra-Rebollo M, Lozano JJ, Estarás C, Ulloa C, et al. Epithelial-mesenchymal transition can suppress major attributes of human epithelial tumor-initiating cells. *J Clin Invest*. 2012; 122:1849–68.
17. Grosse-Wilde A, Fouquier d' Herouei A, McIntosh E, Ertaylan G, Skupin A, Kuestner RE, del Sol A, Walters K-A, Huang S. Stemness of the hybrid epithelial/mesenchymal state in breast cancer and its association with poor survival. *PLoS One*. 2015; 10:e0126522.
18. Jolly MK, Huang B, Lu M, Mani SA, Levine H, Ben-Jacob E. Towards elucidating the connection between epithelial - mesenchymal transitions and stemness. *J R Soc Interface*. 2014; 11:20140962.
19. Tsai JH, Donaher JL, Murphy DA, Chau S, Yang J. Spatiotemporal regulation of epithelial-mesenchymal transition is essential for squamous cell carcinoma metastasis. *Cancer Cell*. 2012; 22:725–36.
20. Ocaña OH, Córcoles R, Fabra A, Moreno-Bueno G, Acloque H, Vega S, Barrallo-Gimeno A, Cano A, Nieto MA. Metastatic colonization requires the repression of the epithelial-mesenchymal transition inducer Prrx1. *Cancer Cell*. 2012; 22:709–24.
21. Burk U, Schubert J, Wellner U, Schmalhofer O, Vincan E, Spaderna S, Brabletz T. A reciprocal repression between ZEB1 and members of the miR-200 family promotes EMT and invasion in cancer cells. *EMBO Rep*. 2008; 9:582–9.
22. Bracken CP, Gregory PA, Kolesnikoff N, Bert AG, Wang J, Shannon MF, Goodall GJ. A double-negative feedback loop between ZEB1-SIP1 and the microRNA-200 family regulates epithelial-mesenchymal transition. *Cancer Res*. 2008; 68:7846–54.
23. Lu M, Jolly MK, Levine H, Onuchic JN, Ben-Jacob E. MicroRNA-based regulation of epithelial-hybrid-mesenchymal fate determination. *Proc Natl Acad Sci U S A*. 2013; 110:18174–9.
24. Lu M, Jolly MK, Onuchic J, Ben-Jacob E. Toward Decoding the Principles of Cancer Metastasis Circuits. *Cancer Res*. 2014; 74:4574–87.

25. Yang X, Lin X, Zhong X, Kaur S, Li N, Liang S, Lassus H, Wang L, Katsaros D, Montone K, Zhao X, Zhang Y, Bützow R, et al. Double-negative feedback loop between reprogramming factor LIN28 and microRNA let-7 regulates aldehyde dehydrogenase 1-positive cancer stem cells. *Cancer Res.* 2010; 70:9463–72.
26. Qiu C, Ma Y, Wang J, Peng S, Huang Y. Lin28-mediated post-transcriptional regulation of Oct4 expression in human embryonic stem cells. *Nucleic Acids Res.* 2010; 38:1240–8.
27. Niwa H, Miyazaki J, Smith AG. Quantitative expression of Oct-3/4 defines differentiation, dedifferentiation or self-renewal of ES cells. *Nat Genet.* 2000; 24:372–6.
28. Theunissen TW, Van Oosten AL, Castelo-Branco G, Hall J, Smith A, Silva JCR. Nanog overcomes reprogramming barriers and induces pluripotency in minimal conditions. *Curr Biol.* 2011; 21:65–71.
29. Karwacki-Neisius V, Göke J, Osorno R, Halbritter F, Ng JH, Weiße AY, Wong FCK, Gagliardi A, Mullin NP, Festuccia N, Colby D, Tomlinson SR, Ng HH, et al. Reduced Oct4 expression directs a robust pluripotent state with distinct signaling activity and increased enhancer occupancy by Oct4 and Nanog. *Cell Stem Cell.* 2013; 12:531–45.
30. Shu J, Wu C, Wu Y, Li Z, Shao S, Zhao W, Tang X, Yang H, Shen L, Zuo X, Yang W, Shi Y, Chi X, et al. Induction of pluripotency in mouse somatic cells with lineage specifiers. *Cell.* Elsevier Inc 2013; 153:963–75.
31. Dai X, Schonbaum C, Degenstein L, Bai W, Mahowald A, Fuchs E. The ovo gene required for cuticle formation and oogenesis in flies is involved in hair formation and spermatogenesis in mice. *Genes Dev.* 1998; 12:3452–63.
32. Nair M, Teng A, Bilanchone V, Agrawal A, Li B, Dai X. *Ovol1* regulates the growth arrest of embryonic epidermal progenitor cells and represses *c-myc* transcription. *J Cell Biol.* 2006; 173:253–64.
33. Nair M, Bilanchone V, Ortt K, Sinha S, Dai X. *Ovol1* represses its own transcription by competing with transcription activator *c-Myb* and by recruiting histone deacetylase activity. *Nucleic Acids Res.* 2007; 35:1687–97.
34. Watanabe K, Villarreal-Ponce A, Sun P, Salmans ML, Fallahi M, Andersen B, Dai X. Mammary morphogenesis and regeneration require the inhibition of EMT at terminal end buds by *Ovol2* transcriptional repressor. *Dev Cell.* 2014; 29:59–74.
35. Roca H, Hernandez J, Weidner S, McEachin RC, Fuller D, Sud S, Schumann T, Wilkinson JE, Zaslavsky A, Li H, Maher CA, Daignault-Newton S, Healy PN, et al. Transcription Factors *OVOL1* and *OVOL2* Induce the Mesenchymal to Epithelial Transition in Human Cancer. *PLoS One.* 2013; 8:e76773.
36. Jia D, Jolly MK, Boareto M, Parsana P, Mooney SM, Pienta KJ, Levine H, Ben-Jacob E. *OVOL* guides the epithelial-hybrid-mesenchymal transition. *Oncotarget.* 2015; 6:15436–48.
37. Li S, Yang J. *Ovol* Proteins: Guardians against EMT during Epithelial Differentiation. *Dev Cell.* 2014; 29:1–2.
38. Zhang T, Zhu Q, Xie Z, Chen Y, Qiao Y, Li L, Jing N. The zinc finger transcription factor *Ovol2* acts downstream of the bone morphogenetic protein pathway to regulate the cell fate decision between neuroectoderm and mesendoderm. *J Biol Chem.* 2013; 288:6166–77.
39. Roca H, Pande M, Huo JS, Hernandez J, Cavalcoli JD, Pienta KJ, McEachin RC. A bioinformatics approach reveals novel interactions of the *OVOL* transcription factors in the regulation of epithelial - mesenchymal cell reprogramming and cancer progression. *BMC Syst Biol.* 2014; 8:29.
40. Lim Y-Y, Wright JA, Attema JL, Gregory PA, Bert AG, Smith E, Thomas D, Lopez AF, Drew PA, Khew-Goodall Y, Goodall GJ. Epigenetic modulation of the miR-200 family is associated with transition to a breast cancer stem-cell-like state. *J Cell Sci.* 2013; 126:2256–66.
41. Lee B, Villarreal-Ponce A, Fallahi M, Ovadia J, Sun P, Yu Q-C, Ito S, Sinha S, Nie Q, Dai X. Transcriptional mechanisms link epithelial plasticity to adhesion and differentiation of epidermal progenitor cells. *Dev Cell.* 2014; 29:47–58.
42. Bednarz-Knoll N, Alix-Panabières C, Pantel K. Plasticity of disseminating cancer cells in patients with epithelial malignancies. *Cancer Metastasis Rev.* 2012; 31:673–87.
43. Aceto N, Bardia A, Miyamoto DT, Donaldson MC, Wittner BS, Spencer JA, Yu M, Pely A, Engstrom A, Zhu H, Brannigan BW, Kapur R, Stott SL, et al. Circulating tumor cell clusters are oligoclonal precursors of breast cancer metastasis. *Cell.* 2014; 158:1110–22.
44. Joosse SA, Gorges TM, Pantel K. Biology, detection, and clinical implications of circulating tumor cells. *EMBO Mol Med.* 2015; 7:1–11.
45. Samavarchi-Tehrani P, Golipour A, David L, Sung H-KK, Beyer TA, Datti A, Woltjen K, Nagy A, Wrana JL. Functional genomics reveals a BMP-driven mesenchymal-to-epithelial transition in the initiation of somatic cell reprogramming. *Cell Stem Cell.* Elsevier Inc. 2010; 7:64–77.
46. Ombrato L, Malanchi I. The EMT universe: space between cancer cell dissemination and metastasis initiation. *Crit Rev Oncog* 2014; 19:349–61.
47. Strauss R, Li Z-Y, Liu Y, Beyer I, Persson J, Sova P, Möller T, Pesonen S, Hemminki A, Hamerlik P, Drescher C, Urban N, Bartek J, et al. Analysis of epithelial and mesenchymal markers in ovarian cancer reveals phenotypic heterogeneity and plasticity. *PLoS One.* 2011; 6:e16186.
48. Ruscetti M, Quach B, Dadashian EL, Mulholland DJ, Hong W. Tracking and Functional Characterization of Epithelial-Mesenchymal Transition and Mesenchymal Tumor Cells During Prostate Cancer Metastasis. *Cancer Res.* 2015; 75:2749–59.
49. Yaswen P. Reinforcing targeted therapeutics with phenotypic stability factors. *Cell Cycle.* 2015; 13:3818–22.
50. Ribeiro AS, Paredes J. P-Cadherin Linking Breast Cancer Stem Cells and Invasion: A Promising Marker to Identify

- an “Intermediate/Metastable” EMT State. *Front Oncol.* 2015; 4.
51. De Herreros AG. Epithelial to mesenchymal transition in tumor cells as consequence of phenotypic instability. *Front Cell Dev Biol.* 2014; 2.
 52. Schliekelman MJ, Taguchi A, Zhu J, Dai X, Rodriguez J, Celiktas M, Zhang Q, Chin A, Wong C, Wang H, McFerrin L, Selemat S, Yang C, et al. Molecular portraits of epithelial, mesenchymal and hybrid states in lung adenocarcinoma and their relevance to survival. *Cancer Res.* 2015; 75:1789–800.
 53. Tan TZ, Miow QH, Miki Y, Noda T, Mori S, Huang RY, Thiery JP. Epithelial-mesenchymal transition spectrum quantification and its efficacy in deciphering survival and drug responses of cancer patients. *EMBO Mol Med.* 2014; 6:1279–93.
 54. Andreucot M, Halley JD, Winkler DA, Huang S. A general model for binary cell fate decision gene circuits with degeneracy: indeterminacy and switch behavior in the absence of cooperativity. *PLoS One.* 2011; 6:e19358.
 55. Guantes R, Poyatos JF. Multistable decision switches for flexible control of epigenetic differentiation. *PLoS Comput Biol.* 2008; 4:e1000235.
 56. Macía J, Widder S, Solé R. Why are cellular switches Boolean? General conditions for multistable genetic circuits. *J Theor Biol.* 2009; 261:126–35.
 57. Chang HH, Hemberg M, Barahona M, Ingber DE, Huang S. Transcriptome-wide noise controls lineage choice in mammalian progenitor cells. *Nature.* 2008; 453:544–7.
 58. Hanahan D, Weinberg RA. Hallmarks of cancer: The next generation. *Cell.* 2011; 144:646–74.
 59. Yang L, Moss T, Mangala L, Marini J, Zhao H, Wahling S, Armaiz-Pena G, Jiang D, Achreja A, Win J, Roopaimoole R, Rodriguez-Aguayo C, Mercado-Urbe I, et al. Metabolic shifts toward glutamine regulator tumor growth, invasion and bioenergetics in ovarian cancer. *Mol Syst Biol.* 2014; 10:1–23.
 60. LeBleu VS, O’Connell JT, Gonzalez Herrera KN, Wikman H, Pantel K, Haigis MC, de Carvalho FM, Damascena A, Domingos Chinen LT, Rocha RM, Asara JM, Kalluri R. PGC-1 α mediates mitochondrial biogenesis and oxidative phosphorylation in cancer cells to promote metastasis. *Nat Cell Biol.* 2014; 16:992–1003.
 61. Steinway SN, Gomez Tejeda Zañudo J, Ding W, Rountree CB, Feith DJ, Loughran TP, Albert R. Network modeling of TGF β signaling in hepatocellular carcinoma epithelial-to-mesenchymal transition reveals joint Sonic hedgehog and Wnt pathway activation. *Cancer Res.* 2014; 74:5963–77.
 62. Huang B, Lu M, Jolly MK, Tsarfaty I, Onuchic J, Ben-Jacob E. The three-way switch operation of Rac1/RhoA GTPase-based circuit controlling amoeboid-hybrid-mesenchymal transition. *Sci Rep.* 2014; 4.
 63. Chung S, Cooper CR, Farach-Carsron MC, Ogunnaike BA. A control engineering approach to understanding the TGF- β paradox in cancer. *J R Soc Interface.* 2012; 9:1389–97.
 64. Lee J, Lee J, Farquhar KS, Yun J, Frankenberger CA, Bevilacqua E, Yeung K, Kim E-J, Balázsi G, Rosner MR. Network of mutually repressive metastasis regulators can promote cell heterogeneity and metastatic transitions. *Proc Natl Acad Sci U S A.* 2014; 111:E364–73.
 65. Aguda BD, Kim Y, Piper-Hunter MG, Friedman A, Marsh CB. MicroRNA regulation of a cancer network: consequences of the feedback loops involving miR-17–92, E2F, and Myc. *Proc Natl Acad Sci U S A.* 2008; 105:19678–83.
 66. Jolly MK, Boareto M, Lu M, Onuchic JN, Clementi C, Ben-Jacob E. Operating Principles of Notch-Delta-Jagged module of cell-cell communication. *New J Phys.* 2015; 17:055021.
 67. Boareto M, Jolly MK, Lu M, Onuchic JN, Clementi C, Ben-Jacob E. Jagged-Delta asymmetry in Notch signaling can give rise to a Sender/Receiver hybrid phenotype. *Proc Natl Acad Sci.* 2015; 112:E402–9.
 68. Lee J, Tiwari A, Shum V, Mills GB, Mancini MA, Igoshin OA, Balázsi G. Unraveling the regulatory connections between two controllers of breast cancer cell fate. *Nucleic Acids Res.* 2014; 42:6839–49.
 69. Clewley R. Hybrid models and biological model reduction with PyDSTool. *PLoS Comput Biol.* 2012; 8:e1002628.