

# **HHS Public Access**

Author manuscript *Mol Nutr Food Res.* Author manuscript; available in PMC 2019 September 01.

Published in final edited form as:

Mol Nutr Food Res. 2018 September ; 62(18): e1800079. doi:10.1002/mnfr.201800079.

## **Cruciferous Vegetables, Isothiocyanates and Bladder Cancer Prevention**

Besma Abbaoui<sup>1,2,3</sup>, Christopher R. Lucas<sup>3,4,5</sup>, Ken M. Riedl<sup>2,5</sup>, Steven K. Clinton<sup>5,6</sup>, and Amir Mortazavi<sup>5,6,\*</sup>

<sup>1</sup>Foods for Health Discovery Theme, The College of Food, Agricultural and Environmental Sciences; The Ohio State University, Columbus, OH 43210

<sup>2</sup>Department of Food Science and Technology, The College of Food, Agricultural, and Environmental Sciences; The Ohio State University, Columbus, OH, 43210

<sup>3</sup>Integrated Biomedical Science Graduate Program, College of Medicine; The Ohio State University, Columbus, OH, 43210

<sup>4</sup>Department of Mechanical and Aerospace Engineering, The College of Engineering; The Ohio State University, Columbus, OH, 43210

<sup>5</sup>Comprehensive Cancer Center, The Ohio State University, Columbus, OH, 43210

<sup>6</sup>Division of Medical Oncology, Department of Internal Medicine, College of Medicine; The Ohio State University, Columbus, OH, 43210

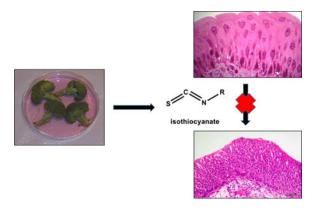
## Abstract

Bladder cancer is a significant health burden due to its high prevalence, risk of mortality, morbidity, and high cost of medical care. Epidemiologic evidence suggests that diets rich in cruciferous vegetables, particularly broccoli, are associated with lower bladder cancer risk. Phytochemicals in cruciferous vegetables, such as glucosinolates, which are enzymatically hydrolyzed to bioactive isothiocyanates, are possible mediators of an anticancer effect. In vitro studies have shown inhibition of bladder cancer cell lines, cell cycle arrest and induction of apoptosis by these isothiocyanates, in particular sulforaphane and erucin. Although, not yet completely understood, many mechanisms of anti-cancer activity at the steps of cancer initiation, promotion and progression have been attributed to these isothiocyanates. They target multiple pathways including the adaptive stress response, phase I/II enzyme modulation, pro-growth, survival, -inflammatory signaling, angiogenesis, and even epigenetic modulation. Multiple in vivo studies have shown the bioavailability of isothiocyanates and their anti-tumoral effects. Although human studies are limited, they support oral bioavailability with reasonable plasma and urine concentrations achieved. Overall, both cell and animal studies support a potential role for isothiocyanates in bladder cancer prevention and treatment. Future studies are necessary to examine clinically relevant outcomes and define guidelines on ameliorating the bladder cancer burden.

<sup>&</sup>lt;sup>\*</sup>Correspondence to: Amir Mortazavi, Division of Medical Oncology, Department of Internal Medicine, College of Medicine, The Ohio State University and The Comprehensive Cancer Center, A454 Starling-Loving Hall, 320 West 10th Ave, Columbus, OH 43210, USA.

## Graphical Abstract.

There is mounting evidence that isothiocyanates, derived from cruciferous vegetables, show strong promise to prevent bladder cancer, specifically transitional cell carcinoma (TCC) of the urinary bladder.



#### **Keywords**

isothiocyanates; bladder cancer; cruciferous vegetables; glucosinolates; chemoprevention

## I. Introduction

Our interest in bladder cancer risk and prevention was motivated by collaboration on a series of large epidemiological studies that elucidated and quantified the potential roles of tobacco, geographic location, fluids, and dietary components on the risk of bladder cancer [1-5]. Interestingly, among the vast array of fruits and vegetables available in North America, only cruciferous vegetables emerged with significant associations with reduced bladder cancer risk. The objective of this review is to critically evaluate the accumulated literature regarding the potential role of cruciferous vegetables in bladder cancer prevention and as a possible adjunct to treatment. Cruciferous vegetables contain a complex mix of phytochemicals, however, this review will focus on isothiocyanates (ITC)s. We begin by defining the burden of bladd*e*r cancer and its pathophysiology. We then explore the cruciferous vegetable family, with a focus upon bioactive isothiocyanates, and summarize the present epidemiologic evidence for employing this class of vegetables in bladder cancer prevention and/or treatment. Next, we will define the chemistry and bioavailability of cruciferous vegetable isothiocyanates. We then proceed to review in vitro and in vivo experimental studies evaluating the effects of cruciferous vegetables and isothiocyanates in bladder cancer, followed by a synopsis of the diverse potential mechanisms of actions suggested to be at play in the bioactivity of these compounds. Finally, we summarize present findings from human interventions to assess the potential of these compounds to prevent bladder cancer and then discuss strategies for future larger scale human clinical trials and how we should proceed as a field to help ameliorate the bladder cancer burden.

## II. The Bladder Cancer Burden

Carcinoma of the urinary bladder presents a significant health care burden in the United States and around the globe. In the United States, bladder cancer is the sixth most common cancer and the eighth most common cause of cancer death, in men, with an estimated 79,000 new cases and 17,000 deaths in 2017 [6, 7]. It is the second most prevalent neoplasm in men 60 years of age or older [8–11]. Worldwide, almost 400,000 new cases are diagnosed annually with approximately 150,000 deaths [12, 13]. Strikingly, bladder cancer recurrence rate is the highest among all malignancies [14] and it has the highest lifetime treatment costs per patient, of all cancers, due to its high recurrence rate and ongoing invasive monitoring requirements [15].

Bladder cancers are derived from the urothelium, also referred to as the transitional epithelial lining of the bladder. More than 90% of bladder cancers are transitional cell carcinoma. The less common bladder cancers are squamous cell carcinoma, adenocarcinoma, and small cell carcinoma, perhaps representing different cells of origin [16, 17].

Tobacco smoking is the most important risk factor of bladder cancer, estimated to cause about half of all cases, increasing relative risk by 2- to 4-fold, and is a growing international concern due to expansion in tobacco use in developing nations [18]. Dramatically, the risk of bladder cancer is directly related to the intensity and duration of smoking, and quitting visibly reduces risk [14]. The increased risk of developing bladder cancer due to smoking is attributed to exposure to a family of known bladder carcinogens present in tobacco smoke, called aromatic amines. The mechanism of action of aromatic amines in the genesis of bladder cancer is not completely currently clear, however there has been an associated link between aromatic amine exposure and a genotoxic effect, specifically DNA adduction and mutagenicity [14]. One can also be exposed to carcinogenic aromatic amines through occupational exposure. Interestingly, bladder cancer was one of the first cancers associated with an industrial process. Epidemiological studies of urinary bladder cancers began in 1985 with a study of excessive occurrence of bladder cancer among workers in the aniline dye industry, with aniline being an aromatic amine, with then accumulating evidence clearly pointing to the relationship between bladder cancer and certain industrial chemicals with known carcinogenic effects [11]. Occupational exposure to different chemical carcinogens accounts for 10–20% of bladder cancers [19]. Some of the occupations that are linked to bladder cancer include painters, miners; and metal, rubber, leather and cement industry workers. Over 40 agents, from contemporary exposure, with a reported role in urothelial carcinogenesis have been identified, including 2-naphthylamine found in the dye and rubber industry as well as tobacco smoke and 4,4'-methylenebis[2-chloroanline] or [MBOCA] for short, which is used in polyurethane production [20]. Bladder infection with Schistosoma haematobium is a very important risk factor for bladder cancer in some parts of the world. Schistosomiasis is most prevalent in East Africa and the Middle East, and thus the most common cancer in men and the second in women [11, 21]. The chlorine [22] and arsenic in drinking water are also potential risk factors [10, 23, 24]. However, some studies suggest that greater total daily fluid intake may reduce the risk of bladder cancer, perhaps by dilution of carcinogens and promoting chemicals [3]. Acquired genetic alterations in tumor

suppressor genes and oncogenes play a critical role in bladder cancer initiation and progression [16]. However, there is no clear hereditary pattern and there appears to be only a small increased risk in relatives of bladder cancer patients [25]. There remains a critical need for effective screening as well as primary and tertiary preventive strategies, in addition to continued smoking cessation efforts.

Transitional cell carcinoma is generally classified into two pathways as summarized in Figure 1. The majority present with non-invasive cancers, which encompasses approximately 80% of urothelial carcinomas, with the remainder presenting with invasive disease. The non-invasive disease is usually managed by cystoscopic resection of the tumor, often followed by intravesical cytotoxic and/or immune therapy. Those with superficial disease have a 5-year survival rate of almost 90%, when optimally monitored and treated. However, non-invasive cancer has an approximately 70% chance of recurring, and a 10–20% chance of progressing into invasive disease [26, 27]. Due to the very high probability of recurrence, and risk of ultimately converting to invasive disease, the current standard-of-care mandates frequent urine tests, cystoscopies and resections, which is challenging and costly, yet this may present a unique and attractive opportunity for possible preventive strategies.

The *de novo* invasive form, in contrast, has a far grimmer prognosis with greater than 50% of patients developing metastatic disease and a 5-yr survival of 6% in the metastatic setting [28–30]. Interestingly, these two clinically different scenarios have distinct mechanisms of molecular carcinogenesis. The non-invasive variant is typically a result of gain-of-function mutations of the *ras* or similar growth regulating pathways, including *ras* itself and fibroblast growth factor receptor 3 (FGFR3) [31–33]. In contrast, the invasive variant is most commonly associated with inactivation of tumor suppressor genes, such as p53 and RB [16, 34].

There remains a strong need for the development of effective treatments and preventative strategies to help ameliorate the bladder cancer burden. Furthermore, the development of novel biomarkers of bladder cancer initiation and progression, and monitoring of treatment are also greatly needed.

## III. History of Cruciferous Vegetables

The mustard family of plants (Brassicaceae or Cruciferae) have a long history of human cultivation and consumption. It is documented that the Ancient Greeks, Romans, Indians and Chinese all utilized and greatly valued cruciferous vegetables. Interestingly, Brassicaceae crops provide the greatest diversity of crops used by man derived from a single species [35]. The diversity of cauliflower and broccoli-like vegetables from *Brassica oleracea* (wild cabbage) occurred in Europe, probably evolving from germplasm introduced in Roman times from the Eastern Mediterranean. Broccoli (*Brassica oleracea italica*) is a name of Italian origin, coming from the Latin 'brachium', meaning an arm or branch and refers to the edible floral shoots on brassica plants, which include cabbages and turnips. It became popular in Northern Europe in the 18<sup>th</sup> century. Broccoli, as a single main green head (called calabrese and named from the Calabria region of Italy), was introduced in the United States by Italian immigrants in the early 20<sup>th</sup> century. It is considered a 'convenience' vegetable

and has spread back to Europe from the United States into Japan and other countries in the Pacific Rim over the past 50 years [35]. Today, broccoli is one of the most consumed cruciferous vegetables in the United States, however it pales in comparison to the consumption of the top five fruits and vegetables: lettuce, tomatoes, potatoes, bananas and orange products. For example, the Continuing Survey of Food Intakes estimated 39–42% consume iceberg lettuce and tomatoes, while only 3% consumed broccoli [36]. Worldwide, cruciferous vegetables are commonly consumed and include cabbage, Brussels sprouts, cauliflower, kale, collard greens, arugula (rocket), bok choy, watercress, canola, mustard seeds, radish, daikon, wasabi and horseradish.

Plants from this family are readily distinguished by a cruciform (cross-shaped) corolla, six stamens (the outer two shorter than the inner four), a capsule often with a septum and a pungent watery sap [37]. However, taxonomic classification of Brassicaceae (338 genera and 3709 species) is one of great complexity and controversy. This is because boundaries between species have been poorly delineated, and largely artificially circumscribed until 2006. The use of molecular biology techniques have helped to more clearly define the taxonomy of this important family of plants [37].

## IV. Epidemiology of Cruciferous Vegetables, Isothiocyanates and Bladder

## Cancer

A landmark series of reports from the Health Professional's Follow-Up Study, a prospective cohort epidemiologic study, involving over 47,000 men and published in the late 1990's provided new insight into risk factors for bladder cancer, particularly diet and nutrition [1, 3–5]. Included in these, was a thorough examination of estimated fruit and vegetable intake and the risk of bladder cancer, reporting that intake of cruciferous vegetables, particularly broccoli, had a strong inverse association with bladder cancer risk, with those consuming <1 serving of broccoli/week vs. >1 serving/week associated with a 29% lower risk, while 2 servings/week experienced a 39% lower risk (p=.009) [5]. Supporting these findings, a retrospective case-control study of over 1400 participants, from MD Anderson Cancer Center, showed that high ITC intake was associated with 29% decreased risk of bladder cancer and the protective effect was most evident in older individuals and ever- and heavysmokers. The study also correlated N-acetyltransferase (NAT2) slow acetylators, an enzymatic process involved in carcinogen metabolism, with increased bladder cancer in Caucasians [38]. A recent meta-analysis found that increasing intake of fruits, vegetables, cruciferous vegetables, citrus fruits and fruits and vegetables combined were associated with a statistically significant reduction in bladder cancer risk with similar results observed in a linear dose-response analysis [39]. Furthermore, another meta-analysis including cohort and case-control studies, evaluated the relationship between cruciferous vegetables intake and risk of bladder cancer, finding a significantly decreased risk of bladder cancer in overall cruciferous vegetables intake group and subgroup of case-control studies but not detected in cohort studies [40]. A hospital-based case-controlled study, involving 275 individuals with incident, primary bladder cancer, observed a strong and statistically significant inverse association between bladder cancer risk and raw cruciferous vegetables intake, with the inverse association remaining consistent among current and heavy smokers with three or

more servings of cruciferous vegetables a month [41]. Furthermore, with an average 8-year follow-up, a strong and significant inverse association was observed between bladder cancer mortality and broccoli intake, particularly raw broccoli intake [42]. Interestingly, in a Multiethnic Cohort Study, it was found that in women, total fruits and vegetables, total vegetables, yellow-orange vegetables and total fruits and citrus fruits were all inversely associated with the risk of invasive bladder cancer in risk factor-adjusted models. However, for men, no association for fruits, vegetables, or nutrients were found, overall; although inverse associations were observed for vegetable intake among current smokers, and in ethnic specific analyses, specifically in Latino men [43].

There is emerging evidence that bladder cancer risk increases significantly in individuals who carry genetic variants of phase II enzymes such as glutathione S-transferase (GST) and NAD(P)H:quinone oxidoreductase 1 (NQO1) [44–47]. Phase II enzymes act as cell protectants by detoxifying against potential carcinogens and oxidants. Interestingly, isothiocyanates found in cruciferous vegetables have been shown to induce phase II enzyme activity and this may in part explain their epidemiologic association with decreased bladder cancer risk [33]. This may explain why high consumption of fruits and vegetables, particularly cruciferous vegetables, is associated with reduced risk of bladder cancer [48–51]. Furthermore, genetic variations in DNA repair genes, including ERCC2, D312N and XPC, have also been shown to be associated with increased bladder cancer risk [52].

Conversely, some epidemiologic studies examining cruciferous vegetables intake and bladder cancer risk, have shown no association of decreased risk [53, 54]. For example, a prospective study from Alpha-Tocopherol Beta-Carotene Cancer Prevention Study did not find an association between total fruits and vegetables intake or cruciferous vegetables intake and bladder cancer risk in smokers [2]. In addition, a Swedish prospective population-based cohort study of 82,002 women and men found no association between total fruits and vegetables or cruciferous vegetables intake [55]. A Meta- Analysis on fruits and vegetables intake and risk of bladder cancer risk also did not see a correlation between cruciferous vegetables and bladder cancer risk, however, an inverse relationship between bladder cancer and green leafy vegetables was seen [56].

Inconsistencies in epidemiologic findings may be related to multiple reasons [57]. Firstly, intakes from diet assessment tools cannot capture many variables that may significantly contribute to the potentially anticancer activity of bioactive in fruits and vegetables. The many types of cruciferous vegetables and their multitude of genetic strains will show a diverse array of bioactives. In addition, cooking methods may impact stability, absorption and bioavailability. The context of the meal where cruciferous vegetables are consumed may also impact host exposure to bioactives [58, 59]. In recent years, many studies are showing exceptionally diverse pharmacokinetics of phytochemicals in humans as host genetics impact the metabolism and degradation of phytochemicals [60–62]. Furthermore, epidemiologic studies must consider the exposures to known bladder carcinogens as critical variables and examine key interactions with diet and host genetic polymorphisms associated with carcinogen metabolism. Overall, in spite of the recognized caveats, the epidemiologic findings support a hypothesis that cruciferous vegetable intake is related to decreased risk of bladder cancer. Further well-designed prospective studies addressing the interactive risk

factors are needed to explore the potential protective impact of cruciferous vegetables over the life cycle of bladder carcinogenesis [40]. In addition to human studies, one logical and necessary step is to test these hypotheses in relevant pre-clinical bladder cancer models of superficial and invasive disease.

## V. Chemistry, Bioavailability and Metabolism of Cruciferous Vegetable Isothiocyanates

A remarkable characteristic of cruciferous plants is their high content of glucosinolates, which often approaches greater than 1% of their dry weight [63]. These compounds serve an important evolutionary protective role as they are a component of the myrosinase system where myrosinase enzymes are sequestered in the intact plant separately from glucosinolate substrates. When the plant is damaged, by an insect or by human chewing, chopping or digestion for example, myrosinase catalyzes the conversion of inactive glucosinolate precursors to an unstable intermediate, which rearranges to produce a nitrile, thiocyanate, or isothiocyanate that protect the plant as illustrated in Figure 2. Isothiocyanates have been shown to have broad antibiotic properties including antimicrobial, nematocidal, antifungal and antiprotozoal [64]. They also cause specific positive and negative feeding cues for some insects, can exhibit insecticidal properties as well as allelopathy, where they can suppress the growth of neighboring plants [63]. Diverse glucosinolates and their corresponding isothiocyanates are common among cruciferous vegetables, with the glucosinolate backbone demonstrating over 100 different R groups [65] and classified as alkyl, indolyl, or benzyl derivatives. Isothiocyanates are often volatile adding spiciness to food products such as horse radish, wasabi, radishes, and many cruciferous sprouts such as those of broccoli. An emerging body of literature involves the bioactivity and fate of indolyl glucosinolates in humans. Downstream metabolites, indole-3-carbinol and di-indolyl methane, are wellstudied in this realm however, this review's focus is on the alkyl isothiocyanates sulforaphane and erucin which have established efficacy in many experimental models of cancer including of the bladder.

The glucosinolate content of cruciferous plants varies greatly, influenced by a multitude of factors including the region, season, climactic conditions of the specific year grown, length and mode of storage and soil conditions among many other factors. However, genetics have been shown to have a greater effect on glucosinolate content over environment [66–68]. Previous damage to a plant primes for higher production of glucosinolates as a protective mechanism, watering also increases overall levels [69]. Contrastingly, lower growing temperatures, with plants harvested in the winter or autumn versus the spring or summer as well as selenium-enriched soil, causes decreased plant glucosinolate levels [67, 68]. Young broccoli plants, especially seeds and sprouts, have 20–50 times the levels of glucosinolates than more mature market-stage plants [70]. Since glucosinolates serve in plant defense, the myrosinase system has evolved to be most potent in immature organs to protect a plant until it reaches its reproductive stage.

Plant tissue damage usually induces glucosinolate hydrolysis to isothiocyanates and subsequent isothiocyanate losses if vegetables are not immediately consumed. Many food

handling practices employed for storage and preparation are disruptive. When vegetables are cooked, glucosinolates can be reduced by 30-60% via thermal degradation but primarily due to leaching into the cooking water when boiled [71–73]. Thawing of frozen cruciferous vegetables, without previous inactivation of myrosinase, leads to almost complete loss of glucosinolate as the cell structure is broken and water becomes available for reactions to occur [74]. Commercial transport, distribution and storage during the retail sale period also lead to significant losses, with a 40% loss of aliphatic glucosinolates after 7 days of storage and a 66% loss after 10 days [74]. To minimize these losses, rapid cooking at sub-boiling temperatures and with scant cooking water – e.g. sautéing, steaming, or stir-frying thermally inactivates myrosinase without leaching and thereby stabilizes glucosinolate levels. Greater isothiocyanate yields at consumption can be realized by pre-treating vegetables at 60  $^{\circ}$ C to selectively inactivate epithiospecifier protein (ESP) such that a greater proportion of intermediates rearrange to isothiocyanates upon processing or chewing [74– 76]. It has also been shown that isothiocyanates in broccoli juice are relatively thermolabile and pressure stable [77]. In addition, High Pressure Processing (HPP) caused no negative effects on the glucosinolate-myrosinase system in broccoli sprouts [78].

The bioavailability and metabolism of vegetables isothiocyanates is a key issue when considering the potential impact these compounds may have on human health. Glucosinolates have limited bioavailability, yet when converted to isothiocyanates, metabolites are recovered in biological samples. Collectively, published data in rats and humans suggests that isothiocyanates from fresh cruciferous vegetables can reach µM concentrations in the blood, accumulate in tissues and persist with a half-life of approximately 2 hours [79]. After conversion by plant myrosinase, isothiocyanates can be transported from the gastrointestinal tract by passive, facilitated or active transport [73]. The high activity of myrosinase in fresh cruciferous vegetables rapidly produces isothiocyanates during chewing and isothiocyanate metabolites can reach their maximum concentration in blood within 30 minutes [80, 81]. During uptake by enterocytes or once they reach the blood and liver, isothiocyanates which are reactive electrophiles and usually trapped and further metabolized. A major first step of metabolism is by conjugation to glutathione (GSH) by Glutathione S-Transferase (GST) enzymes or by direct reaction with GSH or cysteine since thiol groups are strong nucleophiles. Consequently human genetic polymorphisms in GSTs were expected to be one contributing factor impacting isothiocyanate metabolite concentrations *in vivo* yet were not terribly predictive of recoveries [80, 82, 83]. Isothiocyanates conjugated to GSH are processed by the mercapturic acid pathway leading eventually to N-acetylcysteine (NAC) conjugates which are the major form present in urine, as depicted in Figure 3. Acetylated isothiocyanates have demonstrated effects on histone acetylation as histone deacetylases inhibitors. Although NAC-isothiocyanate conjugates are more stable than isothiocyanates themselves, NAC conjugates exist in an equilibrium with free isothiocyanates and the reversible nature of these bonds [84] might explain in part how conjugated isothiocyanates continue to influence biological systems. Free isothiocyanates can also react with protein thiols on the cysteine-rich protein, KEAP-1 thus modulating the KEAP1-Nrf2-antioxidant response element (ARE) signaling pathway [85] where several cysteine residues were reversibly modified through conjugation to sulforaphane.

If myrosinase in foods is inactivated, glucosinolates reach the large intestine where they can be degraded by the resident microflora if they express the requisite myrosinase activity [86]. In a study by Clarke et al. [87] isothiocyanate metabolite levels in humans were measured after delivering a glucosinolate supplement devoid of myrosinase activity and compared to matched amounts of glucosinolates with high levels of endogenous plant myrosinase. Isothiocyanate bioavailability was ~40–80 fold higher in urine of volunteers when consumed as fresh broccoli sprouts [87]. More recently, researchers have aimed to encourage the growth of bacteria in the lower gut with myrosinase activity and thereby enhance conversion to isothiocyanates when humans ingest cooked cruciferous vegetables [88]. This is a promising strategy to maximize cruciferous vegetables health benefits since cruciferous vegetables are often preserved through cooking making isothiocyanate exposure reliant on the gut microbiome.

Two of the key isothiocyanates showing potent bioactivity in a variety of chemoprevention studies are sulforaphane (SFN) and erucin (ECN) which both reach the bladder via the urine after consumption of cruciferous vegetables. This has been shown in vivo where rats were fed purified glucosinolates. SFN and ECN conjugates as well as free SFN and ECN were found in the urine [89]. In addition, a study performed in over 18,000 men with colorectal cancer showed that cruciferous vegetable consumption, in single-void urine samples, led to urinary isothiocyanates concentrations that averaged 2.75 µmol/g creatinine, where urinary ITC concentrations were expressed in units of urinary creatinine to account for varying water contents of urine samples [90]. Cumulative excretion of isothiocyanates in urine have been shown to be about 50 times higher than the maximum concentration in the plasma, making the feasibility of isothiocyanate chemoprevention particularly powerful in the bladder [91]. Interestingly, SFN and ECN represent redox partners of one another (sulfinyl and thio-ether, respectively) and both are present in broccoli sprouts. When SFN and ECN were administered individually to mice, metabolites of both SFN and ECN were detected in biological samples [76, 92] demonstrating interconversion between the two forms. In studies by Clarke et al. [87, 93] it was found that between individuals there was great variance in the ratio of urinary ECN/SFN metabolites and these differences were reproducible over time. In the mouse studies this ratio was similar whether SFN or ECN was incorporated into chow, suggesting a steady state ratio. The varied human ECN/SFN ratios are provocative and may be predictive of individual biological impacts of isothiocyanate exposures. If so, elucidating the biochemical basis would be of great value to steer ratios towards optimal benefits.

More recently an analogous type of alkyl isothiocyanate with similar phase II enzymeinducing activity and chemopreventive promise has come to the forefront. Glucoraphenin and glucoraphasatin occur in radish roots, sprouts, and seeds and upon hydrolysis are converted to sulforaphane and raphasatin. They share the same structure with sulforaphane and erucin except contain a double bond at the 3-position of the alkyl chain. The two isothiocyanates occur with similar abundance to one another and in comparisons with broccoli sprouts for bioactivity, radish sprouts are similarly potent. Also of interest is that radishes are mostly devoid of ESP and thus generate higher yields of isothiocyanate without special pre-treatments [94–96].

#### Human Studies in Isothiocyanate Metabolism

Several small-scale clinical studies have been reported examining the safety, tolerance and metabolism of isothiocyanates. A randomized, placebo-controlled, double-blind Phase I clinical trial of healthy volunteers used 25 µmol or 100 µmol of glucosinolates or 25 µmol of pure isothiocyanate (which is equivalent to about 10 g of dried broccoli or 1 g of dried broccoli sprouts) for 7 days showed no significant toxicities and much higher and more consistent excretion of isothiocyanates in the cohort fed isothiocyanates directly compared to those given glucosinolates [97]. A study comparing administration of a broccoli sprout derived sulforaphane-rich drink (SFR) vs. a glucoraphanin rich drink (GRR) found that the SFR led to fastest and highest peak concentration of serum SFN, while the GRR had considerable slower elimination rates of SFN, therefore implying that the optimal formulation would be a combination of SFR and GRR to achieve peak concentration for activation of targets as well as prolonged inhibition of other protective actions of SFN [98]. Healthy human volunteers given 200 µmol of broccoli sprout isothiocyanates exhibited peak plasma concentrations of 0.94-2.27 µmol/L at 1 hour and declined with first order kinetics with a half-life =  $1.77 \pm 0.13h$  [81]. Furthermore, raw broccoli was shown to result in faster absorption, higher bioavailability and higher peak plasma concentrations in human volunteers over cooked broccoli [99].

Interestingly, smokers consuming 250 g/day of steamed broccoli showed 41% decreased levels of oxidized DNA lesions and 23% increased resistance to  $H_2O_2$  induced DNA strand breaks in peripheral blood mononuclear cells [100]. Another study looked at eight healthy women undergoing reduction mammoplasty who were given a single dose of broccoli sprout preparation containing 200 µmol of SFN. Following oral dosing, SFN metabolites were readily measurable in human breast tissue and were enriched in the epithelium [101]. One study conducted in China, reported striking inter- individual differences in the bioavailability of broccoli sprout isothiocyanates, an important consideration in the design of future human clinical trials [102]. Overall, the current literature supports the oral bioavailability of isothiocyanates, producing biologically active plasma and urine concentrations after consumption of broccoli, sprouts and pure isothiocyanates. Yet, there is clearly a need for further carefully controlled and powered clinical studies examining metabolism, bioavailability and safety of isothiocyanates. In addition, studies documenting the bioactivity of these compounds in relation to modulation of bladder carcinogenesis are also imperative.

## VI. In Vitro and In Vivo Studies of Isothiocyanates and Bladder Cancer

There are limited *in vitro* and *in vivo* experimental studies examining at the potential inhibitory effects of cruciferous vegetables or their components on bladder carcinogenesis. In addition, there is a need for more *in vivo* bioavailability and metabolism studies of isothiocyanates to determine if they can reach appreciable plasma concentrations and reach important target organs and tissues progressing through the carcinogenesis cascade. Furthermore, addition studies focusing upon cellular targets mediating critical steps in bladder carcinogenesis are needed.

The ability of phytochemicals from food to inhibit carcinogenesis began over 5 decades ago, where the observation that animals fed a complex diet were at a lesser risk of developing

cancer than those consuming a semi-purified diet [103]. The two diets were nutritionally comparable in regard to major nutrients but differed significantly in the amount of nonnutrients they contained. Dr. Lee Wattenberg subsequently fed a semi-purified diet and incorporated single food components one at a time to determine which of these exhibits an anticancer effect. He found that cruciferous vegetables are partly responsible for the observed anti-cancer effect of a chow diet [104]. In 1992, Paul Talalay reported the isothiocyanate sulforaphane to be a potent inducer of phase II enzyme activity and suggested that it may be responsible for the anti-cancer effects seen with broccoli consumption [105, 106]. Subsequently, several *in vitro* and *in vivo* studies utilizing broccoli and broccoli sprout extracts as well as pure isothiocyanates have been performed looking at their effects in multiple cancers including lung, esophageal, prostate, breast and colorectal among several others [106–111]. However, the literature on bladder cancer and cruciferous vegetables is much more limited. In vitro, human bladder carcinoma cells (UMUC3 and T24) were significantly inhibited by isothiocyanates at doses of  $7.5-30 \mu$ M, leading to induction of apoptosis and arrested cell cycle progression in the G2/M and S phases [112]. Our data also shows induction of apoptosis and cell cycle arrest in cell lines, ranging from superficial to invasive cell lines, treated with sulforaphane or erucin, with less toxic effects on normal human urothelial cells [92].

There are few in vivo bladder cancer studies utilizing isothiocyanates. In a long term (36 week) rat study, animals fed 160 µmol/kg bw/day broccoli sprout extract and given 0.05% N-butyl-N-(4-hydroxybutyl) nitrosamine (BBN), a specific bladder carcinogen, lead to a significant decrease in incidence, multiplicity, size and progression of bladder cancer. This was attributed to the broccoli extract's ability to significantly induce glutathione-Stransferase and NAD(P)H:quinine oxidoreductase 1, enzymes that detoxify oxidants and carcinogens [113]. Another study showed that sulforaphane can inhibit 4-aminobiphenylinduced DNA damage in RT4 cells and in bladder tissue [114]. It has also been shown that allyl isothiocyanate rich mustard seed powder (71.5 mg/kg giving from a singirin dose of 9 µnmol/kg) can inhibit bladder cancer growth and block muscle invasion by 34.5% in an orthotopic rat bladder cancer model [115]. Also, in a study of n-butyl-(4hydroxybutyl)nitrosamine (BHBN) induced bladder tumors in rats, both 2-mercaptoethane sulfonate (MESNA) and 6-phenylhexyl isothiocyanate inhibited tumorigenesis, but phenethyl isothiocyanate did not [116]. We have also shown the broccoli isothiocyanates have the ability to inhibit established bladder cancer, utilizing a UMUC3 subcutaneous xenograft tumor model [92]. Additional cellular, animal and preclinical studies are described in a recent review on cruciferous vegetables, isothiocyanates and the prevention of bladder cancer [117].

#### Potential Toxicity of Isothiocyanates

Although isothiocyanates have been shown to exhibit anti-bladder cancer effects, they have also been shown to potential toxic effects. One of the major toxic effects of isothiocyanates involves their effects on thyroid function, interfering with iodine uptake and thyroid hormone synthesis and leading to hypothyroidism and goiter (enlargement of the thyroid gland) [97, 118, 119]. An inhibition of proper thyroid function may impact metabolism in almost all tissues, including effects on reproductive organs. In studies looking at animal feed

rich in cruciferous vegetables, a reduction in fertility of male and female animals, as well as growth reduction, and reduction in milk and egg production have been observed [120]. Furthermore, glucosinolate hydrolysis may cause irritation of gastro-intestinal mucosa, which may lead to local necrosis and hepatotoxicity [121].

There have even been studies that have linked isothiocyanates, in low concentrations, with compromising the function of immune cells and impairing genome stability [122]. Isothiocyanates are extremely electrophilic and have the ability to bind sulphydryl groups of biologically important molecules, contributing to their potential toxicity. In fact, some isothiocyanates have also been shown to have bladder cancer inducing properties, particularly phenethyl isothiocyanate (PEITC) and benzyl isothiocyanate (BITC). In rats fed a diet including 0.1% PEITC for 45 weeks, 92% of the mice formed bladder carcinomas, which remained even when the phenethyl isothiocyanate diet was discontinued [123]. Both pre- and post- initiation events were studied. Pre-initiation, 0.1% PIETC or BITC for 14 days lead to increased inflammatory cell infiltration and hyperplasia of the bladder epithelium [124]. In addition, post-initiation effects of 0.1 % PEITC and BITC were examined in urinary bladder carcinogenesis with or without pre-treatment with diethylnitrosamine (DEN) and BBN, and both isothiocyanates showed strong promoter and some complete carcinogenic potential of bladder cancer [125]. One possible reason for this observation could be the accumulated dose of urinary isothiocyanates with this dosing regimen. It is estimated that NAC-ITCs could have been constantly maintained at levels far greater than 1 mmol/L and that although isothiocyanates can be beneficial at preventing or inhibiting bladder cancer growth at lower doses, at excessively higher doses, they may be harmful [112]. In addition, it is possible that the benzyl or phenethyl isothiocyanates may have some harmful effects, but this has not been reported for other isothiocyanates such as sulforaphane or erucin. Broccoli and broccoli sprouts do not have appreciable concentrations of these isothiocyanates and may be a better option over cruciferous vegetables containing higher levels of PIETC or BITC such as cauliflower, cabbage or Brussels sprouts, especially if these compounds are used in high doses for prevention or treatment of bladder cancer [126].

Toxicity can be explained by isothiocyanates ability to readily accumulate in bladder tissue. Given a rat that would consume an average of 16 g of food/day with a diet containing 0.1% isothiocyanate and produces 30 ml of urine in 24 hours, the rat will consume ~100  $\mu$ mol isothiocyanate daily [127]. This would lead to a urinary concentration of ~1900  $\mu$ M in the bladder which is much higher than can be achieved by human consumption and may explain the toxicity observed. Furthermore, cell culture studies have shown that only low concentrations of 5–20  $\mu$ M are necessary to inhibit the growth of cancer cells further supporting the idea that large doses of isothiocyanates are not necessary [112, 128]. However, because other organs do not accumulate isothiocyanates and cannot reach such high concentrations for chemopreventive effects in these organs. Overall cruciferous plants at usual intake appear to be safe in humans [97] with the exception of allergies and with special precaution taken by those taking blood-thinning medications such as Warfarin [129].

## VII. Mechanisms of Action of Isothiocyanates and Bladder Cancer

A wide body of evidence suggests that isothiocyanates possess potent anti-cancer activity via induction of apoptosis through several distinct molecular mechanisms of action that include targeting the Keap-1- and Nrf2-dependent adaptive stress response, inhibition of phase I enzymes (i.e. cytochrome p450), interfering with inflammatory and pro-growth/-survival intracellular signaling pathways, and through disruption of mitochondrial function, reviewed extensively in [130, 131].

#### Modulation of Carcinogen Metabolizing Enzymes

A well-defined isothiocyanate anti-cancer mechanism of action, which may modulate bladder cancer initiation, involves inhibition of phase I enzymes (i.e. cytochrome p450) and the induction of phase II enzymes, such as glutathione-S transferase (GST), quinone reductase (QR), and NAD(P)H:quinone oxidoreductase-1 (NOQ1), reviewed in [131]. Phase I enzymes cause carcinogenic compounds to become more hydrophilic through numerous oxidation, reduction, and hydrolysis reactions, causing increased reactivity and DNA damage [132, 133]. Furthermore, certain cytochrome p450 enzymes are associated with an increased risk of bladder cancer [127]. In contrast, phase II enzymes cause conjugation of the reactive intermediates, rendering them more water-soluble and therefore excreted in the urine through the mercapturic acid pathway [132, 133]. GSH is regulated by phase II enzymes, and is a principle ligand that binds with electrophiles and reactive oxygen species (ROS) [132].

Isothiocyanates have also been demonstrated to interact with the bladder epithelium and induce GST and NQO1, well-known cytoprotective enzymes capable of detoxifying carcinogens [134]. Interestingly, NQO1 has also been shown to stabilize the tumor suppressor p53 [135]. The bladder was also shown to be one of the most responsive tissues for the induction of these enzymes by broccoli sprout extracts which could especially be effective in protecting the bladder against cancer initiation [33]. Isothiocyanates appear to induce phase II enzymes at the transcription level, through the antioxidant response element (ARE) in the 5'-upstream region of mRNA [132, 136, 137]. This response element is activated through the binding of nuclear factor E2-related factor 2 (Nrf2), which can be regulated through mitogen activated protein kinase (MAPK), phosphatidylinositol 3-kinase (PI3K), and other kinases and phase II enzyme inducers [132, 136]. When cytosolic Nrf2 is released from its chaperone, it translocates to the nucleus to induce the transcription of phase II enzymes [132]. Since induction occurs at the level of transcription, chemoprevention through this mechanism of action is thought to be more effective prior to the presence of the carcinogen.

#### Modulation of Cell Cycle and Apoptosis

Evidence indicates isothiocyanates may enhance the deletion of initiated cells from damaged tissue through inducing programmed cell death. One way in which isothiocyanates have been shown to do this is through modulation of the cell cycle. Invasive bladder cancer cells, treated with sulforaphane, have been shown to undergo cell cycle arrest at the G2-M and S phases [138]. In addition, isothiocyanates have been shown to enhance sensitivity to

apoptosis. Sulforaphane (SFN) has also been shown to induce caspase-mediated (caspase 8 and 9) apoptosis in both bladder and prostate cancer cells, as well as cause the overexpression of Bax and down-regulation of Bcl-2 [139]. We have shown that broccoli isothiocyanates, sulforaphane and erucin, can significantly induce apoptosis in superficial (RT4) and invasive (J82, UMUC3) human bladder cancer cell lines, by induction of caspase 3/7 activity and PARP cleavage. We have also shown this phenomenon *in vivo* in a UMUC3 subcutaneous xenograft model, where apoptosis was induced, revealed by an increase in PARP cleavage when mice were treated with broccoli sprout diet. Furthermore, we revealed RT4, J82 and UMUC3 cells to accumulate in the G2/M phase of the cell cycle when treated with either SFN or ECN that was regulated via suppressed levels of survivin, epidermal growth factor receptor (EGFR), and human epidermal growth factor receptor 2 (HER2/neu) [92].

A previous study by Savio, et. al. revealed that allyl isothiocyanate (AITC) (mustard cell essential oil) casued cell cycle arrest, increased apoptosis rates, and varying genotoxicity, which was dependent on the mutational status of TP53 [140]. Savio and colleagues also showed that while AITC affected the BAX/BCL2 pathway in RT4 cells (wt TP53), AITC affected the cytokinesis related ANLN and S100P in T24 (mutant TP53) cells. This suggests AITC regulates bladder cancer cell gene expression in a TP53 genotype-dependent manner [141].

Isothiocyanates have been shown to bind to microtubules to ultimately induce cell cycle arrest and apoptosis. A study by Overby, et. al. showed that isothiocyanates disrupted microtubules in *Arabodopsis thalinia* and in AY-27 rat bladder cancer cells suggesting that plant and mammalian cells share isothiocyante-mediated anti-growth mechanisms [142].

Additionally, SFN was also shown to induce mitotic arrest, however, not in the G2 phase of the cell cycle. Park, et. al. revealed that 5637 human bladder cancer cells treated with SFN underwent caspase-dependent apoptosis via accumulation of reactive oxygen species (ROS) [143]. Mechanistically, Jo and colleagues showed that SFN treated T24 human bladder cancer cells underwent apoptosis via the mitochondria-mediated intrinsic pathway with concomitant elevation of ROS, and activation of endoplasmic reticulum stress (ER) and the Nrf2 signaling pathway suggesting that ER and Nrf2 may represent targets for SFN-dependent apoptosis [144]. Another report revealed that SFN not only affected the epithelial-to-mesenchymal transition but also targeted both the OX-2/MMP2,9/ZEB1, Snail and miR-200c/ZEB signaling pathways in T24 bladder cancer cells [145].

In addition to SFN acting as a single therapeutic, Islam and colleagues showed that SFN in combination with the carbonic anhydrase inhibitor acetazolamide (AZ) targeted both the pH homeostasis pathway and the PI3K/Akt intracellular signaling pro-survival pathway allowing for increased anti-tumor efficacy *in vitro* and *in vivo* [146]. SFN+AZ combination treatment allowed for down-regulation of components of pH homeostasis related carbonic anhydrase 9 (CA9), E-cadherin, N-cadherin, and vimentin proteins while diminishing the epithelial-to-mesenchymal transition and decreased PI3k/Akt survival signaling suggesting a potential link between pH homeostasis and pro-survival signaling [146]. Another previous report showed that SFN inhibited proliferation of BIU87 bladder cancer cells via

upregulation of IGFBP-3 expression levels and negatively regulated nuclear factor- $\kappa$ B (NF- $\kappa$ B) signaling [147]. Collectively, these findings suggest that isothiocyanates, specifically SFN targets multiple pathways in bladder cancer cells including the intrinsic apoptotic and pro-growth/-survival intracellular signaling pathways to promote apoptotic cell death.

In addition to apoptotic induction in bladder cancer cells through modulation of the adaptive stress response, phase I/II enzymes, pro-growth/-survival signaling, and the intrinsic apoptotic pathway, SFN has also been shown to be effective in inhibiting all essential steps of cancer vessel formation from proangiogenic signaling to endothelial cell proliferation, migration and tube formation [148, 149]. Furthermore, SFN has been shown to inhibit cancer cell metastasis. Utilizing B16F-10 melanoma cells, which are highly metastatic to the lungs, when injected through the mouse tail vein, SFN was shown to reduce metastasis, by inhibiting matrix metalloproteinases (MMPs). MMPs are proteases capable of degrading the extracellular matrix, and thereby promoting metastasis [150]. Together, these findings suggest that isothiocyanates are compounds with potent anti-tumor effects with several distinct mechanisms of action that inhibit proliferation, and induce apoptosis and cell cycle arrest.

#### **Epigenetic Modulation**

There is emerging evidence that broccoli isothiocyanates modulate epigenetic activity, partly through microRNA (miR) regulation and act as potent inhibitors of histone deacetylases (HDACs) and DNA methyltransferases (DNMTs), reviewed in [151]. The classical central causes of cancer revolve around changes in DNA structure leading to cell growth deregulation. However, the emerging field of epigenetics reveals that histone changes can also lead to cancer progression, through opening or closing regions of oncogenes or tumor suppressor genes, respectively [152–155]. HDAC activity has been shown to occur in many cancers, leading to deregulation of differentiation, cell cycle, and apoptosis. The tumor suppressor p21 appears to be a target of HDACs and is 'silenced', however, when HDAC inhibitors are utilized, the p21 gene remains open and inhibits cancer progression [156]. Sulforaphane has been shown to exhibit HDAC inhibitory activity in prostate cancer cells in vitro [157]. In vivo, a xenograft model with human prostate cancer cells also exhibited HDAC inhibition, when mice were fed SFN [158]. HDAC inhibition by SFN has also been shown in humans fed broccoli sprouts, where peripheral blood mononuclear cells were isolated and HDAC inhibitory activity was assessed [159]. Our laboratory has recently shown that both isothiocyantes SFN and ECN have the ability to significantly inhibit HDAC activity as well as histone acetyltransferase (HAT) activity, leading to small increased levels in acetylated histone H3 [160]. Furthermore, we have shown a potential novel epigenetic mechanism of broccoli isothiocyanate action, whereby histone phosphorylation decreased with concomitant increases in PP1B and PP2A phosphatase activity in the presence of SFN and ECN treatment [160], where we have previously identified increased histone phosphorylation as a biomarker of bladder cancer progression [161]. Taken together with evidence in the literature, our findings suggest that isothiocyanates act as effective epigenetic modifiers both as HDAC inhibitors and now as HAT inhibitors and phosphatase enhancers to ultimately affect the novel biomarker, histone H1 phosphorylation, to induce anti-tumor activity. To summarize, isothiocyanates have tremendous potential both as therapeutic and

preventative compounds due to their ability to target multiple pathways with numerous mechanisms of action in bladder cancer including the adaptive stress response, phase I/II enzyme modulation, pro-growth/-survival/-inflammatory signaling, the intrinsic apoptotic pathway, and epigenetic modulation.

## VIII. Future Approaches

One potential challenge in the field of isothiocyanates as therapeutic and preventative agents for bladder cancer is targeting the compounds specifically to tumor cells, while leaving healthy tissue unharmed. One novel therapeutic approach is via the emerging fields of nanotechnology and nanomedicine for targeted and/or enhanced drug delivery. A recent report demonstrated the ability to construct rhodamine B isothiocyanate-labelled polyacrylic acid-coated cobalt ferrite nanoparticles (NPs) that were endocytosed with significantly greater efficacy by cancer urothelial cells compared to normal urothelial cells suggesting a selective uptake mechanism by cancerous urothelial cells [162]. In addition, Zhang, et. al. built several versions of  $\beta$ -cyclodextrin functionalized mesoporous silica NPs with hydroxyl, amino, and thiol groups and tested mucoadhesitivity properties on urothelium and found that thiol-modified NPs [163]. Furthermore, doxorubicin-loaded thiol-modified NPs were shown to undergo sustained drug release upon acidification [163]. Collectively, these findings represent a promising and exciting novel area of bladder cancer research that merges tumor biology and nanotechnology.

A deeper understanding of potential mechanisms of action may help us define better strategies for prevention and therapy by cruciferous vegetables or their components. The future integration of genomics, transcriptomics, proteomics and metabolomics, called systems biology, will offer deeper insights into the study of both bladder carcinogenesis as well as the cruciferous vegetable family. For example, these tools may identify new biomarkers in urine or other samples to effectively diagnose bladder cancer earlier and further define subtypes for study. A recent publication shows a pattern of fourteen metabolites including lactic acid, leucine, valine, phenylalanine, glutamate, histidine, aspartic acid, tyrosine, serine, uracil, hypoxanthine, carnitine, pyruvic acid and citric acid linked to early bladder carcinogenesis [164]. Furthermore, metabolomics has been used to study how growing conditions, such as light exposure, can ultimately change the nutritional and phytochemical value, and ultimately the cancer fighting abilities, of broccoli sprouts [165]. Metabolomics has additionally been used to help identify metabolite and transcript biomarkers, which could be useful in the cultivation of broccoli and other Brassica vegetables for increased insect resistance [166]. Fascinatingly, underlying mechanisms and key molecular targets involved in the ability of cruciferous vegetables to improve human health have been characterized through plasma metabolite profiles before and after human consumption of broccoli sprouts. The investigation identified several potential molecular targets of crucifers, including fatty acids, glutathione, glutamine, cysteine, dehydroepiandrosterone and deoxyuridine monophosphate, aiding in the study of established and emerging health benefits that the consumption of cruciferous vegetables may possess [167]. It is apparent that future research utilizing metabolomics and its integration with

additional –omics technologies has strong potential of helping both the fields of bladder cancer and isothiocyanates significantly advance.

## IX. Concluding Thoughts

The natural history and pathophysiology of bladder cancer and the current standard-of-care surveillance programs allow for several opportunities to utilize preventive and treatment strategies. The anatomy of the bladder allows cancers at this site to be monitored easily through outpatient cystoscopy, urine cytology, and novel biomarker-based urine tests. Principally, the bladder is a storage compartment for urine, which provides a route of exposure for excreted carcinogens, as suggested by the strong association between bladder cancer risk and exposure to environmental carcinogens, such as tobacco [168]. However, this can also be exploited by enhancing concentrations of anti-carcinogenic compounds, for example from the diet, for prevention. As we reflect upon several decades of cancer prevention efforts, we have seen major accomplishments [169–176]. Yet, we face enormous challenges as a discipline/field with several large high profile human chemoprevention studies showing no benefit [177–179]. We firmly believe that successful human prevention studies are those with the strongest portfolio of supportive preclinical data, particularly with in vivo experimental models. Thus, additional preclinical studies examining multiple formulations of cruciferous vegetables relevant to human application are needed in a variety of modern models of bladder carcinogenesis [180]. Furthermore, critical human clinical trials utilizing isothiocyanate rich food products, extracts, concentrates or chemically pure phytochemicals are needed to examine bioavailability, safety, and biomarkers of impact in bladder carcinogenesis and other cancers.

Through the performance of well-designed rodent and human investigations, we have the ability to determine the stages of bladder carcinogenesis where cruciferous vegetables may impact risk. If these foods or compounds do indeed prove to demonstrate clear and potent bioactivity, they can be utilized to improve upon the bladder cancer burden in several ways. One strategy would be "primary" chemoprevention by targeting high-risk populations such as heavy smokers and those with environmental/occupational exposures [181]. Another strategy is a "tertiary" chemopreventive approach, in patients who had a successful complete cystoscopic resection of their superficial bladder cancer. In this case, the goal is to reduce or eliminate the well-known high risk of recurrent tumors and the progression to invasive disease. In addition, patients with diagnosed invasive cancers and scheduled for cystectomy could be fed well-characterized isothiocyanate-rich food products in the period between diagnosis and surgery in order to examine the dietary impact on molecular and cellular events in an established cancer. Furthermore, data for additive or synergistic effects are necessary.

Finally, it is also conceivable to combine dietary or phytochemical interventions with established and emerging treatments for bladder cancer, such as immunotherapy, chemotherapy, and radiation, although additional preclinical evidence and supportive mechanistic studies are needed. While foods alone are unlikely to have therapeutic effects on high grade and aggressive cancers, particularly those in the metastatic state, it is conceivable that foods may enhance the activity of other therapeutic agents when provided in novel

combinations or sequence. Based upon our work and others', we believe that the use of cruciferous vegetable consumption, or the development of specific food products rich in cruciferous vegetable components, warrants additional studies in bladder cancer prevention as well as part of future effective treatment strategies. The bladder offers multiple opportunities for intervention with foods or phytochemicals in cancer prevention and it is up to us to best harness these opportunities.

## Acknowledgements

C.R.L is a recipient of a National Institutes of Health T32 Award in Oncology Training Fellowship at The Ohio State University Comprehensive Cancer Center, T32 CA009338. This publication was made possible through support from Foods for Health, a focus area of the Discovery Themes initiative at The Ohio State University.

## List of Abbreviations:

ITC	Isothiocyanate
GLU	Glucosinolate
SFN	Sulforaphane
ECN, ERN	Erucin
GSH	Glutathione
GST	Glutathione S-Transferase
NAC	N-acetylcysteine
PEITC	phenethyl isothiocyanate
BITC	benzyl isothiocyanate
Nrf2	nuclear factor E2-related factor 2
РІЗК	phosphatidylinositol 3-kinase
AITC	allyl isothiocyanate
HDAC	histone deacetylase
HAT	histone acetyltransferase

## References

- Michaud DS, Clinton SK, Rimm EB, Willett WC, Giovannucci E, Risk of bladder cancer by geographic region in a U.S. cohort of male health professionals. Epidemiology 2001, 12, 719– 726. [PubMed: 11679802]
- [2]. Michaud DS, Pietinen P, Taylor PR, Virtanen M, Virtamo J, Albanes D, Intakes of fruits and vegetables, carotenoids and vitamins A, E, C in relation to the risk of bladder cancer in the ATBC cohort study. Br J Cancer 2002, 87, 960–965. [PubMed: 12434284]
- [3]. Michaud DS, Spiegelman D, Clinton SK, Rimm EB, Curhan GC, Willett WC, Giovannucci EL, Fluid intake and the risk of bladder cancer in men. N Engl J Med 1999, 340, 1390–1397. [PubMed: 10228189]

- [4]. Michaud DS, Spiegelman D, Clinton SK, Rimm EB, Willett WC, Giovannucci E, Prospective study of dietary supplements, macronutrients, micronutrients, and risk of bladder cancer in US men. Am J Epidemiol 2000, 152, 1145–1153. [PubMed: 11130620]
- [5]. Michaud DS, Spiegelman D, Clinton SK, Rimm EB, Willett WC, Giovannucci EL, Fruit and vegetable intake and incidence of bladder cancer in a male prospective cohort. J Natl Cancer Inst 1999, 91, 605–613. [PubMed: 10203279]
- [6]. Siegel RL, Miller KD, Jemal A, Cancer statistics, 2016. CA Cancer J Clin 2016, 66, 7–30. [PubMed: 26742998]
- [7]. Siegel RL, Miller KD, Jemal A, Cancer Statistics, 2017. CA Cancer J Clin 2017, 67, 7–30. [PubMed: 28055103]
- [8]. Vercelli M, Quaglia A, Parodi S, Crosignani P, Cancer prevalence in the elderly. ITAPREVAL Working Group. *Tumori* 1999, 85, 391–399.
- [9]. Jemal A, Thomas A, Murray T, Thun M, Cancer statistics, 2002. CA Cancer J Clin 2002, 52, 23– 47. [PubMed: 11814064]
- [10]. Mitra AP, Cote RJ, Molecular pathogenesis and diagnostics of bladder cancer. Annu Rev Pathol 2009, 4, 251–285. [PubMed: 18840072]
- [11]. Mostafa MH, Sheweita SA, O'Connor PJ, Relationship between schistosomiasis and bladder cancer. Clin Microbiol Rev 1999, 12, 97–111. [PubMed: 9880476]
- [12]. Antoni S, Ferlay J, Soerjomataram I, Znaor A, Jemal A, Bray F, Bladder Cancer Incidence and Mortality: A Global Overview and Recent Trends. Eur Urol 2017, 71, 96–108. [PubMed: 27370177]
- [13]. Torre LA, Bray F, Siegel RL, Ferlay J, Lortet-Tieulent J, Jemal A, Global cancer statistics, 2012. CA Cancer J Clin 2015, 65, 87–108. [PubMed: 25651787]
- [14]. Besaratinia A, Tommasi S, Genotoxicity of tobacco smoke-derived aromatic amines and bladder cancer: current state of knowledge and future research directions. FASEB J 2013, 27, 2090–2100.
  [PubMed: 23449930]
- [15]. Sievert KD, Amend B, Nagele U, Schilling D, Bedke J, Horstmann M, Hennenlotter J, Kruck S, Stenzl A, Economic aspects of bladder cancer: what are the benefits and costs? World J Urol 2009, 27, 295–300. [PubMed: 19271220]
- [16]. Wu XR, Urothelial tumorigenesis: a tale of divergent pathways. Nat Rev Cancer 2005, 5, 713– 725. [PubMed: 16110317]
- [17]. Crawford JM, The origins of bladder cancer. Lab Invest 2008, 88, 686–693. [PubMed: 18475256]
- [18]. Freedman ND, Silverman DT, Hollenbeck AR, Schatzkin A, Abnet CC, Association between smoking and risk of bladder cancer among men and women. JAMA 2011, 306, 737–745.[PubMed: 21846855]
- [19]. Jung I, Messing E, Molecular mechanisms and pathways in bladder cancer development and progression. Cancer Control 2000, 7, 325–334. [PubMed: 10895126]
- [20]. Cumberbatch MG, Cox A, Teare D, Catto JW, Contemporary Occupational Carcinogen Exposure and Bladder Cancer: A Systematic Review and Meta-analysis. JAMA Oncol 2015, 1, 1282–1290. [PubMed: 26448641]
- [21]. el-Mawla NG, el-Bolkainy MN, Khaled HM, Bladder cancer in Africa: update. Semin Oncol 2001, 28, 174–178. [PubMed: 11301380]
- [22]. Villanueva CM, Fernandez F, Malats N, Grimalt JO, Kogevinas M, Meta-analysis of studies on individual consumption of chlorinated drinking water and bladder cancer. J Epidemiol Community Health 2003, 57, 166–173. [PubMed: 12594192]
- [23]. Augustine A, Hebert JR, Kabat GC, Wynder EL, Bladder cancer in relation to cigarette smoking. Cancer Res 1988, 48, 4405–4408. [PubMed: 3390836]
- [24]. Marshall G, Ferreccio C, Yuan Y, Bates MN, Steinmaus C, Selvin S, Liaw J, Smith AH, Fiftyyear study of lung and bladder cancer mortality in Chile related to arsenic in drinking water. J Natl Cancer Inst 2007, 99, 920–928. [PubMed: 17565158]
- [25]. Goldgar DE, Easton DF, Cannon-Albright LA, Skolnick MH, Systematic population-based assessment of cancer risk in first-degree relatives of cancer probands. J Natl Cancer Inst 1994, 86, 1600–1608. [PubMed: 7932824]

- [26]. Parekh DJ, Bochner BH, Dalbagni G, Superficial and muscle-invasive bladder cancer: principles of management for outcomes assessments. J Clin Oncol 2006, 24, 5519–5527. [PubMed: 17158537]
- [27]. Sexton WJ, Wiegand LR, Correa JJ, Politis C, Dickinson SI, Kang LC, Bladder cancer: a review of non-muscle invasive disease. Cancer Control 2010, 17, 256–268. [PubMed: 20861813]
- [28]. Stein JP, Lieskovsky G, Cote R, Groshen S, Feng AC, Boyd S, Skinner E, Bochner B, Thangathurai D, Mikhail M, Raghavan D, Skinner DG, Radical cystectomy in the treatment of invasive bladder cancer: long-term results in 1,054 patients. J Clin Oncol 2001, 19, 666–675. [PubMed: 11157016]
- [29]. Shariat SF, Karakiewicz PI, Palapattu GS, Lotan Y, Rogers CG, Amiel GE, Vazina A, Gupta A, Bastian PJ, Sagalowsky AI, Schoenberg MP, Lerner SP, Outcomes of radical cystectomy for transitional cell carcinoma of the bladder: a contemporary series from the Bladder Cancer Research Consortium. J Urol 2006, 176, 2414–2422; discussion 2422. [PubMed: 17085118]
- [30]. von der Maase H, Sengelov L, Roberts JT, Ricci S, Dogliotti L, Oliver T, Moore MJ, Zimmermann A, Arning M, Long-term survival results of a randomized trial comparing gemcitabine plus cisplatin, with methotrexate, vinblastine, doxorubicin, plus cisplatin in patients with bladder cancer. J Clin Oncol 2005, 23, 4602–4608. [PubMed: 16034041]
- [31]. van Rhijn BWG, Lurkin I, Radvanyi F, Kirkels WJ, van der Kwast TH, Zwarthoff EC, 2001, pp. 1265–1268.
- [32]. Jebar AH, Hurst CD, Tomlinson DC, Johnston C, Taylor CF, Knowles MA, FGFR3 and Ras gene mutations are mutually exclusive genetic events in urothelial cell carcinoma. Oncogene 2005, 24, 5218–5225. [PubMed: 15897885]
- [33]. Zhang Y, Munday R, Jobson HE, Munday CM, Lister C, Wilson P, Fahey JW, Mhawech-Fauceglia P, Induction of GST and NQO1 in cultured bladder cells and in the urinary bladders of rats by an extract of broccoli (Brassica oleracea italica) sprouts. J Agric Food Chem 2006, 54, 9370–9376. [PubMed: 17147420]
- [34]. Wu XR, Biology of urothelial tumorigenesis: insights from genetically engineered mice. Cancer Metastasis Rev 2009, 28, 281–290. [PubMed: 20012171]
- [35]. Dixon GR, Vegetable Brassicas and Related Crucifers, CAB International, London 2007.
- [36]. Johnston CS, Taylor CA, Hampl JS, More Americans are eating "5 a day" but intakes of dark green and cruciferous vegetables remain low. J Nutr 2000, 130, 3063–3067. [PubMed: 11110870]
- [37]. Franzke A, Lysak MA, Al-Shehbaz IA, Koch MA, Mummenhoff K, Cabbage family affairs: the evolutionary history of Brassicaceae. Trends Plant Sci, 16, 108–116. [PubMed: 21177137]
- [38]. Zhao H, Lin J, Grossman HB, Hernandez LM, Dinney CP, Wu X, Dietary isothiocyanates, GSTM1, GSTT1, NAT2 polymorphisms and bladder cancer risk. Int J Cancer 2007, 120, 2208– 2213. [PubMed: 17290402]
- [39]. Yao B, Yan Y, Ye X, Fang H, Xu H, Liu Y, Li S, Zhao Y, Intake of fruit and vegetables and risk of bladder cancer: a dose-response meta-analysis of observational studies. Cancer Causes Control 2014, 25, 1645–1658. [PubMed: 25248495]
- [40]. Liu B, Mao Q, Lin Y, Zhou F, Xie L, The association of cruciferous vegetables intake and risk of bladder cancer: a meta-analysis. World J Urol 2013, 31, 127–133. [PubMed: 22391648]
- [41]. Tang L, Zirpoli GR, Guru K, Moysich KB, Zhang Y, Ambrosone CB, McCann SE, Consumption of raw cruciferous vegetables is inversely associated with bladder cancer risk. Cancer Epidemiol Biomarkers Prev 2008, 17, 938–944. [PubMed: 18398034]
- [42]. Tang L, Zirpoli GR, Guru K, Moysich KB, Zhang Y, Ambrosone CB, McCann SE, Intake of cruciferous vegetables modifies bladder cancer survival. Cancer Epidemiol Biomarkers Prev 2010, 19, 1806–1811. [PubMed: 20551305]
- [43]. Park SY, Ollberding NJ, Woolcott CG, Wilkens LR, Henderson BE, Kolonel LN, Fruit and vegetable intakes are associated with lower risk of bladder cancer among women in the Multiethnic Cohort Study. J Nutr 2013, 143, 1283–1292. [PubMed: 23739308]
- [44]. Salagovic J, Kalina I, Habalova V, Hrivnak M, Valansky L, Biros E, The role of human glutathione S-transferases M1 and T1 in individual susceptibility to bladder cancer. Physiol Res 1999, 48, 465–471. [PubMed: 10783912]

- [45]. Schulz WA, Krummeck A, Rosinger I, Eickelmann P, Neuhaus C, Ebert T, Schmitz-Drager BJ, Sies H, Increased frequency of a null-allele for NAD(P)H: quinone oxidoreductase in patients with urological malignancies. Pharmacogenetics 1997, 7, 235–239. [PubMed: 9241663]
- [46]. Park SJ, Zhao H, Spitz MR, Grossman HB, Wu X, An association between NQO1 genetic polymorphism and risk of bladder cancer. Mutat Res 2003, 536, 131–137. [PubMed: 12694753]
- [47]. Toruner GA, Akyerli C, Ucar A, Aki T, Atsu N, Ozen H, Tez M, Cetinkaya M, Ozcelik T, Polymorphisms of glutathione S-transferase genes (GSTM1, GSTP1 and GSTT1) and bladder cancer susceptibility in the Turkish population. Arch Toxicol 2001, 75, 459–464. [PubMed: 11757669]
- [48]. Kim MK, Park JH, Conference on "Multidisciplinary approaches to nutritional problems". Symposium on "Nutrition and health". Cruciferous vegetable intake and the risk of human cancer: epidemiological evidence. Proc Nutr Soc 2009, 68, 103–110. [PubMed: 19061536]
- [49]. Lin J, Kamat A, Gu J, Chen M, Dinney CP, Forman MR, Wu X, Dietary intake of vegetables and fruits and the modification effects of GSTM1 and NAT2 genotypes on bladder cancer risk. Cancer Epidemiol Biomarkers Prev 2009, 18, 2090–2097. [PubMed: 19549811]
- [50]. To K, Zhao Y, Jiang H, Hu K, Wang M, Wu J, Lee C, Yokom DW, Stratford AL, Klinge U, Mertens PR, Chen CS, Bally M, Yapp D, Dunn SE, The phosphoinositide- dependent kinase-1 inhibitor 2-amino-N-[4-[5-(2-phenanthrenyl)-3-(trifluoromethyl)-1H-pyrazol-1-yl]phenyl]-ace tamide (OSU-03012) prevents Y-box binding protein-1 from inducing epidermal growth factor receptor. Mol Pharmacol 2007, 72, 641–652. [PubMed: 17595327]
- [51]. Nagano J, Kono S, Preston DL, Moriwaki H, Sharp GB, Koyama K, Mabuchi K, Bladder-cancer incidence in relation to vegetable and fruit consumption: a prospective study of atomic-bomb survivors. Int J Cancer 2000, 86, 132–138. [PubMed: 10728607]
- [52]. Stern MC, Lin J, Figueroa JD, Kelsey KT, Kiltie AE, Yuan JM, Matullo G, Fletcher T, Benhamou S, Taylor JA, Placidi D, Zhang ZF, Steineck G, Rothman N, Kogevinas M, Silverman D, Malats N, Chanock S, Wu X, Karagas MR, Andrew AS, Nelson HH, Bishop DT, Sak SC, Choudhury A, Barrett JH, Elliot F, Corral R, Joshi AD, Gago-Dominguez M, Cortessis VK, Xiang YB, Gao YT, Vineis P, Sacerdote C, Guarrera S, Polidoro S, Allione A, Gurzau E, Koppova K, Kumar R, Rudnai P, Porru S, Carta A, Campagna M, Arici C, Park SS, Garcia-Closas M, Polymorphisms in DNA repair genes, smoking, and bladder cancer risk: findings from the international consortium of bladder cancer. Cancer Res 2009, 69, 6857–6864. [PubMed: 19706757]
- [53]. Vieira AR, Vingeliene S, Chan DS, Aune D, Abar L, Navarro Rosenblatt D, Greenwood DC, Norat T, Fruits, vegetables, and bladder cancer risk: a systematic review and meta-analysis. Cancer Med 2015, 4, 136–146. [PubMed: 25461441]
- [54]. Vrieling A, Lifestyle and bladder cancer prevention: no consistent evidence from cohort studies. Eur J Epidemiol 2017, 32, 1033–1035. [PubMed: 28871470]
- [55]. Larsson SC, Andersson SO, Johansson JE, Wolk A, Fruit and vegetable consumption and risk of bladder cancer: a prospective cohort study. Cancer Epidemiol Biomarkers Prev 2008, 17, 2519– 2522. [PubMed: 18768526]
- [56]. Xu C, Zeng XT, Liu TZ, Zhang C, Yang ZH, Li S, Chen XY, Fruits and vegetables intake and risk of bladder cancer: a PRISMA-compliant systematic review and dose-response meta-analysis of prospective cohort studies. Medicine (Baltimore) 2015, 94, e759. [PubMed: 25929912]
- [57]. World Cancer Research Fund, Food, Nutrition and the Prevention of Cancer: A Global Perspective, American Institute for Cancer Research, Washington, DC 2007.
- [58]. Kopec RE, Cooperstone JL, Schweiggert RM, Young GS, Harrison EH, Francis DM, Clinton SK, Schwartz SJ, Avocado consumption enhances human postprandial provitamin A absorption and conversion from a novel high-beta-carotene tomato sauce and from carrots. J Nutr 2014, 144, 1158–1166. [PubMed: 24899156]
- [59]. Unlu NZ, Bohn T, Clinton SK, Schwartz SJ, Carotenoid absorption from salad and salsa by humans is enhanced by the addition of avocado or avocado oil. J Nutr 2005, 135, 431–436. [PubMed: 15735074]
- [60]. Moran NE, Novotny JA, Cichon MJ, Riedl KM, Rogers RB, Grainger EM, Schwartz SJ, Erdman JW, Jr., Clinton SK, Absorption and Distribution Kinetics of the 13C-Labeled Tomato Carotenoid Phytoene in Healthy Adults. J Nutr 2016, 146, 368–376. [PubMed: 26674763]

- [61]. Moran NE, Cichon MJ, Riedl KM, Grainger EM, Schwartz SJ, Novotny JA, Erdman JW, Jr., Clinton SK, Compartmental and noncompartmental modeling of (1)(3)Clycopene absorption, isomerization, and distribution kinetics in healthy adults. Am J Clin Nutr 2015, 102, 1436–1449. [PubMed: 26561629]
- [62]. Moran NE, Rogers RB, Lu CH, Conlon LE, Lila MA, Clinton SK, Erdman JW, Jr., Biosynthesis of highly enriched 13C-lycopene for human metabolic studies using repeated batch tomato cell culturing with 13C-glucose. Food Chem 2013, 139, 631–639. [PubMed: 23561155]
- [63]. Assayed ME, Abd El-Aty AM, Cruciferous plants: phytochemical toxicity versus cancer chemoprotection. Mini Rev Med Chem 2009, 9, 1470–1478. [PubMed: 20205629]
- [64]. Hopkins RJ, van Dam NM, van Loon JJ, Role of glucosinolates in insect-plant relationships and multitrophic interactions. Annu Rev Entomol 2009, 54, 57–83. [PubMed: 18811249]
- [65]. Fahey JW, Zalcmann AT, Talalay P, The chemical diversity and distribution of glucosinolates and isothiocyanates among plants. Phytochemistry 2001, 56, 5–51. [PubMed: 11198818]
- [66]. Schonhof I, Krumbein A, Brueckner B, Genotypic effects on glucosinolates and sensory properties of broccoli and cauliflower. Nahrung 2004, 48, 25–33. [PubMed: 15053347]
- [67]. Kushad MM, Brown AF, Kurilich AC, Juvik JA, Klein BP, Wallig MA, Jeffery EH, Variation of glucosinolates in vegetable crops of *Brassica oleracea*. J Agric Food Chem 1999, 47, 1541–1548. [PubMed: 10564014]
- [68]. Farnham MW, Wilson PE, Stephenson KK, Fahey JW, Genetic and environmental effects on glucosinolate content and chemoprotective potency of broccoli. Plant Breeding 2004, 123, 60–65.
- [69]. Shelton AL, Within-plant variation in glucosinolate concentrations of Raphanus sativus across multiple scales. J Chem Ecology 2005, 31, 1711–1731.
- [70]. Fahey JW, Zhang Y, Talalay P, Broccoli sprouts: an exceptionally rich source of inducers of enzymes that protect against chemical carcinogens. Proc Natl Acad Sci U S A 1997, 94, 10367– 10372. [PubMed: 9294217]
- [71]. Mithen RF, Dekker M, Verkerk R, Rabot S, Johnson IT, The nutritional significance, biosynthesis and bioavailability of glucosinolates in human foods. J Sci Food Agric 2000, 967–984.
- [72]. McNaughton SA, Marks GC, Development of a food composition database for the estimation of dietary intakes of glucosinolates, the biologically active constituents of cruciferous vegetables. British J Nutr 2003, 90, 687–697.
- [73]. Holst B, Williamson G, A critical review of the bioavailability of glucosinolates and related compounds. Nat Prod Rep 2004, 21, 425–447. [PubMed: 15162227]
- [74]. Vallejo F, Tomas-Barberan F, Garcia-Viguera C, Health-promoting compounds in broccoli as influenced by refrigerated transport and retail sale period. J Agric Food Chem 2003, 51, 3029– 3034. [PubMed: 12720387]
- [75]. Nho CW, Jeffery E, The synergistic upregulation of phase II detoxification enzymes by glucosinolate breakdown products in cruciferous vegetables. Toxicol Appl Pharmacol 2001, 174, 146–152. [PubMed: 11446830]
- [76]. Bricker GV, Riedl KM, Ralston RA, Tober KL, Oberyszyn TM, Schwartz SJ, Isothiocyanate metabolism, distribution, and interconversion in mice following consumption of thermally processed broccoli sprouts or purified sulforaphane. Mol Nutr Food Res 2014, 58, 1991–2000. [PubMed: 24975513]
- [77]. Van Eylen D, Oey I, Hendrickx M, Van Loey A, Kinetics of the stability of broccoli (Brassica oleracea Cv. Italica) myrosinase and isothiocyanates in broccoli juice during pressure/ temperature treatments. J Agric Food Chem 2007, 55, 2163–2170. [PubMed: 17305356]
- [78]. Westphal A, Riedl KM, Cooperstone JL, Kamat S, Balasubramaniam VM, Schwartz SJ, Bohm V, High-Pressure Processing of Broccoli Sprouts: Influence on Bioactivation of Glucosinolates to Isothiocyanates. J Agric Food Chem 2017, 65, 8578–8585. [PubMed: 28929757]
- [79]. Clarke JD, Dashwood RH, Ho E, Multi-targeted prevention of cancer by sulforaphane. Cancer Lett 2008, 269, 291–304. [PubMed: 18504070]
- [80]. Gasper AV, Al-Janobi A, Smith JA, Bacon JR, Fortun P, Atherton C, Taylor MA, Hawkey CJ, Barrett DA, Mithen RF, Glutathione S-transferase M1 polymorphism and metabolism of sulforaphane from standard and high-glucosinolate broccoli. Am J Clin Nutr 2005, 82, 1283– 1291. [PubMed: 16332662]

- [81]. Ye L, Dinkova-Kostova AT, Wade KL, Zhang Y, Shapiro TA, Talalay P, Quantitative determination of dithiocarbamates in human plasma, serum, erythrocytes and urine: pharmacokinetics of broccoli sprout isothiocyanates in humans. Clinica Chimica Acta 2002, 316, 43–53.
- [82]. Steck SE, Gammon MD, Hebert JR, Wall DE, Zeisel SH, GSTM1, GSTT1, GSTP1, and GSTA1 polymorphisms and urinary isothiocyanate metabolites following broccoli consumption in humans. J Nutr 2007, 137, 904–909. [PubMed: 17374652]
- [83]. Steck SE, Gaudet MM, Britton JA, Teitelbaum SL, Terry MB, Neugut AI, Santella RM, Gammon MD, Interactions among GSTM1, GSTT1 and GSTP1 polymorphisms, cruciferous vegetable intake and breast cancer risk. Carcinogenesis 2007, 28, 1954–1959. [PubMed: 17693660]
- [84]. Conaway CC, Jiao D, Chung FL, Inhibition of rat liver cytochrome P450 isozymes by isothiocyanates and their conjugates: a structure-activity relationship study. Carcinogenesis 1996, 17, 2423–2427. [PubMed: 8968058]
- [85]. O'Mealey GB, Berry WL, Plafker SM, Sulforaphane is a Nrf2-independent inhibitor of mitochondrial fission. Redox Biol 2017, 11, 103–110. [PubMed: 27889639]
- [86]. Krul C, Humblot C, Philippe C, Vermeulen M, van Nuenen M, Havenaar R, Rabot S, Metabolism of sinigrin (2-propenyl glucosinolate) by the human colonic microflora in a dynamic in vitro large-intestinal model. Carcinogenesis 2002, 23, 1009–1016. [PubMed: 12082023]
- [87]. Clarke JD, Hsu A, Riedl K, Bella D, Schwartz SJ, Stevens JF, Ho E, Bioavailability and interconversion of sulforaphane and erucin in human subjects consuming broccoli sprouts or broccoli supplement in a cross-over study design. Pharmacol Res 2011, 64, 456–463. [PubMed: 21816223]
- [88]. Angelino D, Dosz EB, Sun J, Hoeflinger JL, Van Tassell ML, Chen P, Harnly JM, Miller MJ, Jeffery EH, Myrosinase-dependent and -independent formation and control of isothiocyanate products of glucosinolate hydrolysis. Front Plant Sci 2015, 6, 831. [PubMed: 26500669]
- [89]. Bheemreddy RM, Jeffery EH, The metabolic fate of purified glucoraphanin in F344 rats. J Agric Food Chem 2007, 55, 2861–2866. [PubMed: 17367161]
- [90]. Moy KA, Yuan JM, Chung FL, Van Den Berg D, Wang R, Gao YT, Yu MC, Urinary total isothiocyanates and colorectal cancer: a prospective study of men in Shanghai, China. Cancer Epidemiol Biomarkers Prev 2008, 17, 1354–1359. [PubMed: 18559550]
- [91]. Ye L, Dinkova-Kostova AT, Wade KL, Zhang Y, Shapiro TA, Talalay P, Quantitative determination of dithiocarbamates in human plasma, serum, erythrocytes and urine: pharmacokinetics of broccoli sprout isothiocyanates in humans. Clin Chim Acta 2002, 316, 43– 53. [PubMed: 11750273]
- [92]. Abbaoui B, Riedl KM, Ralston RA, Thomas-Ahner JM, Schwartz SJ, Clinton SK, Mortazavi A, Inhibition of bladder cancer by broccoli isothiocyanates sulforaphane and erucin: characterization, metabolism, and interconversion. Mol Nutr Food Res 2012, 56, 1675–1687. [PubMed: 23038615]
- [93]. Clarke JD, Riedl K, Bella D, Schwartz SJ, Stevens JF, Ho E, Comparison of isothiocyanate metabolite levels and histone deacetylase activity in human subjects consuming broccoli sprouts or broccoli supplement. J Agric Food Chem 2011, 59, 10955–10963. [PubMed: 21928849]
- [94]. Abdull Razis AF, De Nicola GR, Pagnotta E, Iori R, Ioannides C, 4-Methylsulfanyl-3-butenyl isothiocyanate derived from glucoraphasatin is a potent inducer of rat hepatic phase II enzymes and a potential chemopreventive agent. Arch Toxicol 2012, 86, 183–194. [PubMed: 21960141]
- [95]. Suzuki I, Cho YM, Hirata T, Toyoda T, Akagi JI, Nakamura Y, Sasaki A, Nakamura T, Okamoto S, Shirota K, Suetome N, Nishikawa A, Ogawa K, Toxic effects of 4-methylthio-3-butenyl isothiocyanate (Raphasatin) in the rat urinary bladder without genotoxicity. J Appl Toxicol 2017, 37, 485–494. [PubMed: 27633481]
- [96]. Pawlik A, Wala M, Hac A, Felczykowska A, Herman-Antosiewicz A, Sulforaphene, an isothiocyanate present in radish plants, inhibits proliferation of human breast cancer cells. Phytomedicine 2017, 29, 1–10. [PubMed: 28515021]
- [97]. Shapiro TA, Fahey JW, Dinkova-Kostova AT, Holtzclaw WD, Stephenson KK, Wade KL, Ye L, Talalay P, Safety, tolerance, and metabolism of broccoli sprout glucosinolates and isothiocyanates: a clinical phase I study. Nutr Cancer 2006, 55, 53–62. [PubMed: 16965241]

- [98]. Egner PA, Chen JG, Wang JB, Wu Y, Sun Y, Lu JH, Zhu J, Zhang YH, Chen YS, Friesen MD, Jacobson LP, Munoz A, Ng D, Qian GS, Zhu YR, Chen TY, Botting NP, Zhang Q, Fahey JW, Talalay P, Groopman JD, Kensler TW, Bioavailability of Sulforaphane from two broccoli sprout beverages: results of a short-term, cross-over clinical trial in Qidong, China. Cancer Prev Res (Phila), 4, 384–395. [PubMed: 21372038]
- [99]. Vermeulen M, Klopping-Ketelaars IW, van den Berg R, Vaes WH, Bioavailability and kinetics of sulforaphane in humans after consumption of cooked versus raw broccoli. J Agric Food Chem 2008, 56, 10505–10509. [PubMed: 18950181]
- [100]. Riso P, Martini D, Moller P, Loft S, Bonacina G, Moro M, Porrini M, DNA damage and repair activity after broccoli intake in young healthy smokers. Mutagenesis, 25, 595–602. [PubMed: 20713433]
- [101]. Cornblatt BS, Ye L, Dinkova-Kostova AT, Erb M, Fahey JW, Singh NK, Chen MS, Stierer T, Garrett-Mayer E, Argani P, Davidson NE, Talalay P, Kensler TW, Visvanathan K, Preclinical and clinical evaluation of sulforaphane for chemoprevention in the breast. Carcinogenesis 2007, 28, 1485–1490. [PubMed: 17347138]
- [102]. Kensler TW, Chen JG, Egner PA, Fahey JW, Jacobson LP, Stephenson KK, Ye L, Coady JL, Wang JB, Wu Y, Sun Y, Zhang QN, Zhang BC, Zhu YR, Qian GS, Carmella SG, Hecht SS, Benning L, Gange SJ, Groopman JD, Talalay P, Effects of glucosinolate-rich broccoli sprouts on urinary levels of aflatoxin-DNA adducts and phenanthrene tetraols in a randomized clinical trial in He Zuo township, Qidong, People's Republic of China. Cancer Epidemiol Biomarkers Prev 2005, 14, 2605–2613. [PubMed: 16284385]
- [103]. Silverstone H, Solomon RD, Tannenbaum A, Relative influences of natural and semipurified diets on tumor formation in mice. Cancer Res 1952, 12, 750–756. [PubMed: 12988206]
- [104]. Wattenberg LW, Inhibition of carcinogenesis by naturally-occurring and synthetic compounds. Basic Life Sci 1990, 52, 155–166. [PubMed: 2183767]
- [105]. Zhang Y, Talalay P, Cho CG, Posner GH, A major inducer of anticarcinogenic protective enzymes from broccoli: isolation and elucidation of structure. Proc Natl Acad Sci U S A 1992, 89, 2399–2403. [PubMed: 1549603]
- [106]. Zhang Y, Talalay P, Anticarcinogenic activities of organic isothiocyanates: chemistry and mechanisms. Cancer Res 1994, 54, 1976s–1981s. [PubMed: 8137323]
- [107]. Aragon-Ching JB, Implications for chemoprevention of prostate cancer with intake of cruciferous vegetables. Asian J Androl, 13, 357–358. [PubMed: 21499283]
- [108]. Stoner GD, Morrissey DT, Heur YH, Daniel EM, Galati AJ, Wagner SA, Inhibitory effects of phenethyl isothiocyanate on N-nitrosobenzylmethylamine carcinogenesis in the rat esophagus. Cancer Res 1991, 51, 2063–2068. [PubMed: 2009525]
- [109]. Conaway CC, Yang YM, Chung FL, Isothiocyanates as cancer chemopreventive agents: their biological activities and metabolism in rodents and humans. Curr Drug Metab 2002, 3, 233–255. [PubMed: 12083319]
- [110]. Hecht SS, Chemoprevention by isothiocyanates. J Cell Biochem Suppl 1995, 22, 195–209.[PubMed: 8538199]
- [111]. Lynn A, Collins A, Fuller Z, Hillman K, Ratcliffe B, Cruciferous vegetables and colo-rectal cancer. Proc Nutr Soc 2006, 65, 135–144. [PubMed: 16441953]
- [112]. Tang L, Zhang Y, Dietary isothiocyanates inhibit the growth of human bladder carcinoma cells. J Nutr 2004, 134, 2004–2010. [PubMed: 15284390]
- [113]. Munday R, Mhawech-Fauceglia P, Munday CM, Paonessa JD, Tang L, Munday JS, Lister C, Wilson P, Fahey JW, Davis W, Zhang Y, Inhibition of urinary bladder carcinogenesis by broccoli sprouts. Cancer Res 2008, 68, 1593–1600. [PubMed: 18310317]
- [114]. Ding Y, Paonessa JD, Randall KL, Argoti D, Chen L, Vouros P, Zhang Y, Sulforaphane inhibits 4-aminobiphenyl-induced DNA damage in bladder cells and tissues. Carcinogenesis, 31, 1999– 2003.
- [115]. Bhattacharya A, Tang L, Li Y, Geng F, Paonessa JD, Chen SC, Wong MK, Zhang Y, Inhibition of bladder cancer development by allyl isothiocyanate. Carcinogenesis, 31, 281–286. [PubMed: 19955395]

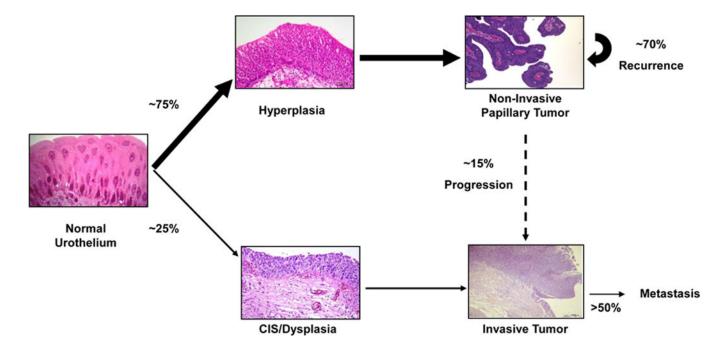
- [116]. Nishikawa A, Morse MA, Chung FL, Inhibitory effects of 2-mercaptoethane sulfonate and 6phenylhexyl isothiocyanate on urinary bladder tumorigenesis in rats induced by N-butyl-N-(4hydroxybutyl)nitrosamine. Cancer Lett 2003, 193, 11–16. [PubMed: 12691818]
- [117]. Veeranki OL, Bhattacharya A, Tang L, Marshall JR, Zhang Y, Cruciferous vegetables, isothiocyanates, and prevention of bladder cancer. Curr Pharmacol Rep 2015, 1, 272–282. [PubMed: 26273545]
- [118]. Sarne D, Effects of Environment, Chemicals and Drugs on Thyroid Function, Endotext.org, South Dartmouth (MA) 2016.
- [119]. MK Tripathi AM, Glucosinolates in animal nutrition: A review. Animal Feed Science and Technology 2007, 132, 1–27.
- [120]. Manoj K,Triphathi M, AS, Prospects and Problems of Dietary Glucosinolates in Animal Feeding. J Adv Dairy Res 2017, 5.
- [121]. King G, Molecular genetics and breeding of vegetable brassicas Euphytica 1990, 50, 97–112.
- [122]. Grundemann C, Huber R, Chemoprevention with isothiocyanates From bench to bedside. Cancer Lett 2017, 414, 26–33. [PubMed: 29111351]
- [123]. Sugiura S, Ogawa K, Hirose M, Takeshita F, Asamoto M, Shirai T, Reversibility of proliferative lesions and induction of non-papillary tumors in rat urinary bladder treated with phenylethyl isothiocyanate. Carcinogenesis 2003, 24, 547–553. [PubMed: 12663517]
- [124]. Akagi K, Sano M, Ogawa K, Hirose M, Goshima H, Shirai T, Involvement of toxicity as an early event in urinary bladder carcinogenesis induced by phenethyl isothiocyanate, benzyl isothiocyanate, and analogues in F344 rats. Toxicol Pathol 2003, 31, 388–396. [PubMed: 12851104]
- [125]. Hirose M, Yamaguchi T, Kimoto N, Ogawa K, Futakuchi M, Sano M, Shirai T, Strong promoting activity of phenylethyl isothiocyanate and benzyl isothiocyanate on urinary bladder carcinogenesis in F344 male rats. Int J Cancer 1998, 77, 773–777. [PubMed: 9688312]
- [126]. VanEtten CH, Daxenbichler ME, Williams PH, Kwolek WF, Glucosinolates and derived products in cruciferous vegetables. Analysis of the edible part from twenty-two varieties of cabbage. J Agric Food Chem 1976, 24, 452–455. [PubMed: 1270657]
- [127]. Tang L, Zhang Y, Isothiocyanates in the chemoprevention of bladder cancer. Current Drug Metabolism 2004, 5, 193–201. [PubMed: 15078196]
- [128]. Gamet-Payrastre L, Li P, Lumeau S, Cassar G, Dupont M, Chevolleau S, Gasc N, Tulliez J, Terce F, Sulforaphane, a naturally occurring isothiocyanate, induces cell cycle arrest and apoptosis in HT29 human colon cancer cells. Cancer Research 2000, 60, 1426–1433. [PubMed: 10728709]
- [129]. Scott O, Galicia-Connolly E, Adams D, Surette S, Vohra S, Yager JY, The safety of cruciferous plants in humans: a systematic review. J Biomed Biotechnol 2012, 2012, 503241. [PubMed: 22500092]
- [130]. Yang L, Palliyaguru DL, Kensler TW, Frugal chemoprevention: targeting Nrf2 with foods rich in sulforaphane. Semin Oncol 2016, 43, 146–153. [PubMed: 26970133]
- [131]. Leone A, Diorio G, Sexton W, Schell M, Alexandrow M, Fahey JW, Kumar NB, Sulforaphane for the chemoprevention of bladder cancer: molecular mechanism targeted approach. Oncotarget 2017, 8, 35412–35424. [PubMed: 28423681]
- [132]. Talalay P, Fahey JW, Phytochemicals from cruciferous plants protect against cancer by modulating carcinogen metabolism. J Nutrition 2001, 131, 3027S–3033S. [PubMed: 11694642]
- [133]. Keum YS, Khor TO, Lin W, Shen G, Kwon KH, Barve A, Li W, Kong AN, Pharmacokinetics and pharmacodynamics of broccoli sprouts on the suppression of prostate cancer in transgenic adenocarcinoma of mouse prostate (TRAMP) mice: implication of induction of Nrf2, HO-1 and apoptosis and the suppression of Akt-dependent kinase pathway. Pharm Res 2009, 26, 2324– 2331. [PubMed: 19669099]
- [134]. Hayes JD, Pulford DJ, The glutathione S-transferase supergene family: regulation of GST and the contribution of the isoenzymes to cancer chemoprotection and drug resistance. Crit Rev Biochem Mol Biol 1995, 30, 445–600. [PubMed: 8770536]
- [135]. Nioi P, Hayes JD, Contribution of NAD(P)H:quinone oxidoreductase 1 to protection against carcinogenesis, and regulation of its gene by the Nrf2 basic-region leucine zipper and the

arylhydrocarbon receptor basic helix-loop-helix transcription factors. Mutat Res 2004, 555, 149–171. [PubMed: 15476858]

- [136]. Zhang Y, Cancer-preventive isothiocyanates: measurement of human exposure and mechanism of action. Mutation Research 2004, 555, 173–190. [PubMed: 15476859]
- [137]. Steinkellner H, Rabot S, Freywald C, Nobis E, Scharf G, Chabicovsky M, Knasmuller S, Kassie F, Effects of cruciferous vegetables and their constituents on drug metabolizing enzymes involved in the bioactivation of DNA-reactive dietary carcinogens. Mutation Research 2001, 480–481, 285–297.
- [138]. Tang L, Zhang Y, Jobson HE, Li J, Stephenson KK, Wade KL, Fahey JW, Potent activation of mitochondria-mediated apoptosis and arrest in S and M phases of cancer cells by a broccoli sprout extract. Mol Cancer Ther 2006, 5, 935–944. [PubMed: 16648564]
- [139]. Singh AV, Xiao D, Lew KL, Dhir R, Singh SV, Sulforaphane induces caspase- mediated apoptosis in cultured PC-3 human prostate cancer cells and retards growth of PC-3 xenografts in vivo. Carcinogenesis 2004, 25, 83–90. [PubMed: 14514658]
- [140]. Savio AL, da Silva GN, de Camargo EA, Salvadori DM, Cell cycle kinetics, apoptosis rates, DNA damage and TP53 gene expression in bladder cancer cells treated with allyl isothiocyanate (mustard essential oil). Mutat Res 2014, 762, 40–46. [PubMed: 24625788]
- [141]. Savio AL, da Silva GN, Salvadori DM, Inhibition of bladder cancer cell proliferation by allyl isothiocyanate (mustard essential oil). Mutat Res 2015, 771, 29–35. [PubMed: 25771977]
- [142]. Overby A, Baevre MS, Thangstad OP, Bones AM, Disintegration of microtubules in Arabidopsis thaliana and bladder cancer cells by isothiocyanates. Front Plant Sci 2015, 6, 6. [PubMed: 25657654]
- [143]. Park HS, Han MH, Kim GY, Moon SK, Kim WJ, Hwang HJ, Park KY, Choi YH, Sulforaphane induces reactive oxygen species-mediated mitotic arrest and subsequent apoptosis in human bladder cancer 5637 cells. Food Chem Toxicol 2014, 64, 157–165. [PubMed: 24296129]
- [144]. Jo GH, Kim GY, Kim WJ, Park KY, Choi YH, Sulforaphane induces apoptosis in T24 human urinary bladder cancer cells through a reactive oxygen species-mediated mitochondrial pathway: the involvement of endoplasmic reticulum stress and the Nrf2 signaling pathway. Int J Oncol 2014, 45, 1497–1506. [PubMed: 24993616]
- [145]. Shan Y, Zhang L, Bao Y, Li B, He C, Gao M, Feng X, Xu W, Zhang X, Wang S, Epithelialmesenchymal transition, a novel target of sulforaphane via COX-2/MMP2, 9/Snail, ZEB1 and miR-200c/ZEB1 pathways in human bladder cancer cells. J Nutr Biochem 2013, 24, 1062–1069. [PubMed: 23159064]
- [146]. Islam SS, Mokhtari RB, Akbari P, Hatina J, Yeger H, Farhat WA, Simultaneous Targeting of Bladder Tumor Growth, Survival, and Epithelial-to-Mesenchymal Transition with a Novel Therapeutic Combination of Acetazolamide (AZ) and Sulforaphane (SFN). Target Oncol 2016, 11, 209–227. [PubMed: 26453055]
- [147]. Dang YM, Huang G, Chen YR, Dang ZF, Chen C, Liu FL, Guo YF, Xie XD, Sulforaphane inhibits the proliferation of the BIU87 bladder cancer cell line via IGFBP-3 elevation. Asian Pac J Cancer Prev 2014, 15, 1517–1520. [PubMed: 24641360]
- [148]. Bertl E, Bartsch H, Gerhauser C, Inhibition of angiogenesis and endothelial cell functions are novel sulforaphane-mediated mechanisms in chemoprevention. Mol Cancer Ther 2006, 5, 575– 585. [PubMed: 16546971]
- [149]. Asakage M, Tsuno NH, Kitayama J, Tsuchiya T, Yoneyama S, Yamada J, Okaji Y, Kaisaki S, Osada T, Takahashi K, Nagawa H, Sulforaphane induces inhibition of human umbilical vein endothelial cells proliferation by apoptosis. Angiogenesis 2006, 9, 83–91. [PubMed: 16821112]
- [150]. Thejass P, Kuttan G, Antimetastatic activity of Sulforaphane. Life Sci 2006, 78, 3043–3050.[PubMed: 16600309]
- [151]. Royston KJ, Tollefsbol TO, The Epigenetic Impact of Cruciferous Vegetables on Cancer Prevention. Curr Pharmacol Rep 2015, 1, 46–51. [PubMed: 25774338]
- [152]. Marks P, Rifkind RA, Richon VM, Breslow R, Miller T, Kelly WK, Histone deacetylases and cancer: causes and therapies. Nat Rev Cancer 2001, 1, 194–202. [PubMed: 11902574]
- [153]. Johnstone RW, Licht JD, Histone deacetylase inhibitors in cancer therapy: is transcription the primary target? Cancer Cell 2003, 4, 13–18. [PubMed: 12892709]

- [154]. Dashwood RH, Myzak MC, Ho E, Dietary HDAC inhibitors: time to rethink weak ligands in cancer chemoprevention? Carcinogenesis 2006, 27, 344–349. [PubMed: 16267097]
- [155]. Dashwood RH, Ho E, Dietary histone deacetylase inhibitors: from cells to mice to man. Semin Cancer Biol 2007, 17, 363–369. [PubMed: 17555985]
- [156]. Halkidou K, Gaughan L, Cook S, Leung HY, Neal DE, Robson CN, Upregulation and nuclear recruitment of HDAC1 in hormone refractory prostate cancer. Prostate 2004, 59, 177–189. [PubMed: 15042618]
- [157]. Myzak MC, Hardin K, Wang R, Dashwood RH, Ho E, Sulforaphane inhibits histone deacetylase activity in BPH-1, LnCaP and PC-3 prostate epithelial cells. Carcinogenesis 2006, 27, 811–819. [PubMed: 16280330]
- [158]. Myzak MC, Dashwood WM, Orner GA, Ho E, Dashwood RH, Sulforaphane inhibits histone deacetylase in vivo and suppresses tumorigenesis in Apc-minus mice. Faseb J 2006, 20, 506–508. [PubMed: 16407454]
- [159]. Myzak MC, Tong P, Dashwood WM, Dashwood RH, Ho E, Sulforaphane retards the growth of human PC-3 xenografts and inhibits HDAC activity in human subjects. Exp Biol Med (Maywood) 2007, 232, 227–234. [PubMed: 17259330]
- [160]. Abbaoui B, Telu KH, Lucas CR, Thomas-Ahner JM, Schwartz SJ, Clinton SK, Freitas MA, Mortazavi A, The impact of cruciferous vegetable isothiocyanates on histone acetylation and histone phosphorylation in bladder cancer. J Proteomics 2017, 156, 94–103. [PubMed: 28132875]
- [161]. Telu KH, Abbaoui B, Thomas-Ahner JM, Zynger DL, Clinton SK, Freitas MA, Mortazavi A, Alterations of histone H1 phosphorylation during bladder carcinogenesis. J Proteome Res 2013, 12, 3317–3326. [PubMed: 23675690]
- [162]. Lojk J, Bregar VB, Strojan K, Hudoklin S, Veranic P, Pavlin M, Kreft ME, Increased endocytosis of magnetic nanoparticles into cancerous urothelial cells versus normal urothelial cells. Histochem Cell Biol 2017.
- [163]. Zhang Q, Neoh KG, Xu L, Lu S, Kang ET, Mahendran R, Chiong E, Functionalized mesoporous silica nanoparticles with mucoadhesive and sustained drug release properties for potential bladder cancer therapy. Langmuir 2014, 30, 6151–6161. [PubMed: 24824061]
- [164]. Cheng Y, Yang X, Deng X, Zhang X, Li P, Tao J, Qin C, Wei J, Lu Q, Metabolomics in bladder cancer: a systematic review. Int J Clin Exp Med 2015, 8, 11052–11063. [PubMed: 26379905]
- [165]. Maldini M, Natella F, Baima S, Morelli G, Scaccini C, Langridge J, Astarita G, Untargeted Metabolomics Reveals Predominant Alterations in Lipid Metabolism Following Light Exposure in Broccoli Sprouts. Int J Mol Sci 2015, 16, 13678–13691. [PubMed: 26084047]
- [166]. Ku KM, Becker TM, Juvik JA, Transcriptome and Metabolome Analyses of Glucosinolates in Two Broccoli Cultivars Following Jasmonate Treatment for the Induction of Glucosinolate Defense to Trichoplusia ni (Hubner). Int J Mol Sci 2016, 17.
- [167]. Housley L, Magana AA, Hsu A, Beaver LM, Wong CP, Stevens JF, Choi J, Jiang Y, Bella D, Williams DE, Maier CS, Shannon J, Dashwood RH, Ho E, Untargeted Metabolomic Screen Reveals Changes in Human Plasma Metabolite Profiles Following Consumption of Fresh Broccoli Sprouts. Mol Nutr Food Res 2018.
- [168]. Wynder EL, Goldsmith R, The epidemiology of bladder cancer: a second look. Cancer 1977, 40, 1246–1268. [PubMed: 332323]
- [169]. Fisher B, Costantino JP, Wickerham DL, Cecchini RS, Cronin WM, Robidoux A, Bevers TB, Kavanah MT, Atkins JN, Margolese RG, Runowicz CD, James JM, Ford LG, Wolmark N, Tamoxifen for the prevention of breast cancer: current status of the National Surgical Adjuvant Breast and Bowel Project P-1 study. J Natl Cancer Inst 2005, 97, 1652–1662. [PubMed: 16288118]
- [170]. Vogel VG, Costantino JP, Wickerham DL, Cronin WM, Cecchini RS, Atkins JN, Bevers TB, Fehrenbacher L, Pajon ER, Jr., Wade JL, 3rd, Robidoux A, Margolese RG, James J, Lippman SM, Runowicz CD, Ganz PA, Reis SE, McCaskill-Stevens W, Ford LG, Jordan VC, Wolmark N, Effects of tamoxifen vs raloxifene on the risk of developing invasive breast cancer and other disease outcomes: the NSABP Study of Tamoxifen and Raloxifene (STAR) P-2 trial. Jama 2006, 295, 2727–2741. [PubMed: 16754727]

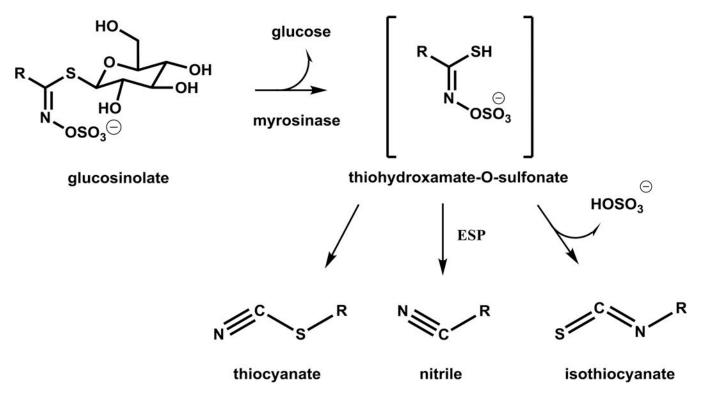
- [171]. Wickerham DL, Costantino JP, Vogel VG, Cronin WM, Cecchini RS, Ford LG, Wolmark N, The use of tamoxifen and raloxifene for the prevention of breast cancer. Recent Results Cancer Res 2009, 181, 113–119. [PubMed: 19213563]
- [172]. Andriole GL, Bostwick DG, Brawley OW, Gomella LG, Marberger M, Montorsi F, Pettaway CA, Tammela TL, Teloken C, Tindall DJ, Somerville MC, Wilson TH, Fowler IL, Rittmaster RS, Effect of dutasteride on the risk of prostate cancer. N Engl J Med, 362, 1192–1202.
- [173]. Reed AB, Parekh DJ, The utility of 5-alpha reductase inhibitors in the prevention and diagnosis of prostate cancer. Curr Opin Urol 2009, 19, 238–242. [PubMed: 19318950]
- [174]. Asano TK, McLeod RS, Non steroidal anti-inflammatory drugs (NSAID) and Aspirin for preventing colorectal adenomas and carcinomas. Cochrane Database Syst Rev 2004, CD004079.
- [175]. Peek RM, Jr., Prevention of colorectal cancer through the use of COX-2 selective inhibitors. Cancer Chemother Pharmacol 2004, 54 Suppl 1, S50–56. [PubMed: 15309515]
- [176]. Sandler RS, Aspirin and other nonsteroidal anti-inflammatory agents in the prevention of colorectal cancer. Important Adv Oncol 1996, 123–137. [PubMed: 8791132]
- [177]. Albanes D, Heinonen OP, Huttunen JK, Taylor PR, Virtamo J, Edwards BK, Haapakoski J, Rautalahti M, Hartman AM, Palmgren J, et al., Effects of alpha-tocopherol and beta-carotene supplements on cancer incidence in the Alpha-Tocopherol Beta-Carotene Cancer Prevention Study. Am J Clin Nutr 1995, 62, 1427S–1430S. [PubMed: 7495243]
- [178]. Goralczyk R, Beta-carotene and lung cancer in smokers: review of hypotheses and status of research. Nutr Cancer 2009, 61, 767–774. [PubMed: 20155614]
- [179]. Lippman SM, Klein EA, Goodman PJ, Lucia MS, Thompson IM, Ford LG, Parnes HL, Minasian LM, Gaziano JM, Hartline JA, Parsons JK, Bearden JD, 3rd, Crawford ED, Goodman GE, Claudio J, Winquist E, Cook ED, Karp DD, Walther P, Lieber MM, Kristal AR, Darke AK, Arnold KB, Ganz PA, Santella RM, Albanes D, Taylor PR, Probstfield JL, Jagpal TJ, Crowley JJ, Meyskens FL, Jr., Baker LH, Coltman CA, Jr., Effect of selenium and vitamin E on risk of prostate cancer and other cancers: the Selenium and Vitamin E Cancer Prevention Trial (SELECT). Jama 2009, 301, 39–51. [PubMed: 19066370]
- [180]. Wu XR, Lin JH, Walz T, Haner M, Yu J, Aebi U, Sun TT, Mammalian uroplakins. A group of highly conserved urothelial differentiation-related membrane proteins. J Biol Chem 1994, 269, 13716–13724. [PubMed: 8175808]
- [181]. La Rochelle J, Kamat A, Grossman HB, Pantuck A, Chemoprevention of bladder cancer. BJU Int 2008, 102, 1274–1278. [PubMed: 19035892]



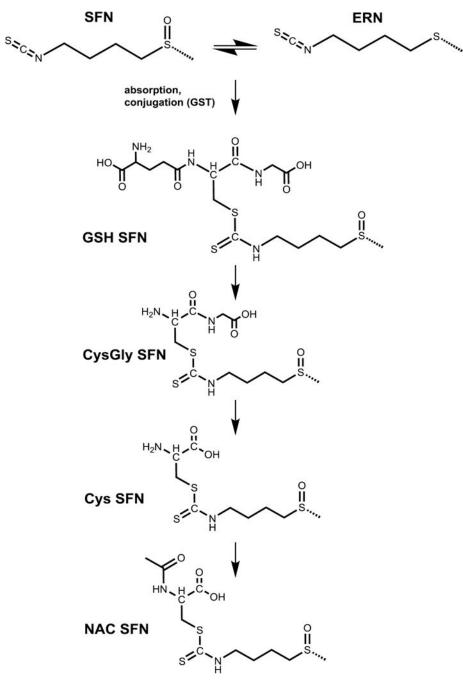
#### Figure 1. Transitional Cell Carcinoma Pathogenesis.

Transitional Cell Carcinoma of the urinary bladder tends to follow two pathways of pathogenesis. The more common non-invasive papillary tumor pathway has a high survival rate, but a high chance of recurrence, and a chance of progression. The invasive tumor pathway has a very high likelihood of developing into metastatic disease and a much lower rate of survival than the non-invasive tumor. *Images, 'unpublished findings'*.

Page 30



**Figure 2.** Glucosinolate to isothiocyanate conversion. Glucosinolate (GLU) hydrolysis mediated by myrosinase (MYR) (plant or gut) produces an unstable intermediate which spontaneously rearranges to thiocyanate, isothiocyanate (ITC), or to nitrile facilitated by epithiospecifier protein (ESP).



#### Figure 3. Mercapturic Acid Pathway.

Alkyl isothiocyanates such as sulforaphane (SFN) and erucin (ERN), produced by myrosinase (MYR) action on glucosinolates after decompartmentalization of broccoli and broccoli sprouts (chewing or processing), can be interconverted post-absorption. Either can be scavenged by glutathione (spontaneous or mediated by glutathione S-transferase (GST)) to form a glutathione (GSH) conjugate which is rapidly processed by the mercapturic acid pathway to N-acetyl cysteine isothiocyanate (ITC) which predominates in urine. SFN is used as the example here although ERN is presumably metabolized in the same manner. It is not

known whether interconversion occurs between free ITC forms, after conjugation, or from either form.