



Hodson, AJ., B, B., Pearce, D., Laybourn-Parry, J., & Tranter, M. (2015). Cryospheric ecosystems: a synthesis of snowpack and glacial research. *Environmental Research Letters*, *10*(11), [110201]. https://doi.org/10.1088/1748-9326/10/11/110201

Publisher's PDF, also known as Version of record License (if available): CC BY Link to published version (if available):

10.1088/1748-9326/10/11/110201

Link to publication record in Explore Bristol Research PDF-document

This is the final published version of the article (version of record). It first appeared online via IOP Publishing at http://dx.doi.org/10.1088/1748-9326/10/11/110201. Please refer to any applicable terms of use of the publisher.

University of Bristol - Explore Bristol Research General rights

This document is made available in accordance with publisher policies. Please cite only the published version using the reference above. Full terms of use are available: http://www.bristol.ac.uk/red/research-policy/pure/user-guides/ebr-terms/



Home Search Collections Journals About Contact us My IOPscience

Cryospheric ecosystems: a synthesis of snowpack and glacial research

This content has been downloaded from IOPscience. Please scroll down to see the full text.

2015 Environ. Res. Lett. 10 110201

(http://iopscience.iop.org/1748-9326/10/11/110201)

View the table of contents for this issue, or go to the journal homepage for more

Download details:

IP Address: 137.222.139.41

This content was downloaded on 13/11/2015 at 14:51

Please note that terms and conditions apply.

Environmental Research Letters



OPEN ACCESS

EDITORIAL

Cryospheric ecosystems: a synthesis of snowpack and glacial research

Andy Hodson ^{1,2}, Ben Brock ³, David Pearce ^{4,5}, Johanna Laybourn-Parry ⁶ and Martyn Tranter ⁶

- ¹ Department of Geography, University of Sheffield, S10 2TN, UK
- ² Arctic Geology, University Centre on Svalbard, N-9171 Longyearbyen, Norway
- Department of Geography, University of Northumbria, NE1 8ST, UK
- Department of Applied Sciences, University of Northumbria NE1 8ST, UK
- Arctic Biology, University Centre on Svalbard, N-9171 Longyearbyen, Norway
- ⁶ School of Geographical Sciences, University of Bristol, BS8 1s, UK

28 October 2015

PUBLISHED

Content from this work may be used under the terms of the Creative Commons Attribution 3.0 licence.

Any further distribution of this work must maintain attribution to the author(s) and the title of the work, journal citation and DOI.



Abstract

The fourteen letters that contributed to this focus issue on cryospheric ecosytems provide an excellent basis for considering the state of the science following a marked increase in research attention since the new millennium. Research letters from the focus issue provide significant insights into the biogeochemical and biological processes associated with snow, glacier ice and glacial sediments. This has been achieved via a significant, empirical effort that has given particular emphasis to glacier surface habitats. However, far less is known about aerobiology, glacial snow covers, supraglacial lakes and sub-ice sedimentary habitats, whose access for sampling and *in-situ* monitoring remains a great challenge to scientists. Furthermore, the use of models to explore key fluxes, processes and impacts of a changing glacial cryosphere are conspicuous by their absence. As a result, a range of process investigations and modelling studies are required to address the increasing urgency and uncertainty that is associated with understanding the response of cryospheric ecosystems to global change.

Introduction

Since the turn of the last millennium, our perception of glaciers and ice sheets has shifted from one of a lifeless, abiotic realm to a distinct biome with a microbial biomass broadly similar to all of Earth's unfrozen freshwater (Priscu et al 2008, Hodson et al 2008, Anesio and Laybourn-Parry 2012). This paradigm shift was a long time coming, because the very earliest explorers crossing the Greenland Ice Sheet in the 1890s had already drawn our attention to the discolouring of snow and ice surfaces by microorganisms, and investigations of snow micro-biota were conducted during expeditions to Antarctica in the 1900s by groups under the leadership of Scott, Mawson and others (e.g. McLean 1918). The meiofauna, algae, Protozoa and bacteria were then studied from the 1930s onwards, including during the 1957 International Geophysical Year (Sisler 1961). However, the data were qualitative rather than quantitative. We had an idea of what organisms were present but no information on their role. It was not until the 1960s

that snow algal photosynthesis was quantified (Fogg 1967): a challenging measurement that has been replicated by few studies since (e.g. Mosser et al 1977). Investigations of bacterial and viral production only started at the beginning of this century (Laybourn-Parry et al 2012). However, in spite of this protracted development history, we are now in a position where a profoundly important role for glaciers and ice sheets in global biogeochemical cycles is being realised (e.g. Wadham et al 2013, Hood et al 2015;) and demands for a better understanding of the implications of global cryospheric change are being made by many new research groups. This synthesis therefore places the research letters in this Cryospheric Ecosystems Focus Issue in the context of the recent literature and reviews our present understanding of the cascade of biogeochemical and ecological processes through ice-bound and ice-marginal terrestrial habitats. The intention is to identify the multidisciplinary approaches needed to understand global cryospheric change, and to identify where future research effort should be directed.

Snowpacks and snowmelt

Seasonal snow cover exerts a fundamental control upon biological production in underlying habitats: a problem that is best understood in the context of soil and tundra ecology in boreal and Arctic landscapes (e.g. Luus et al 2013). Interestingly, in the context of glacial snowpacks, changes in the distribution and persistence of snow cover are reasonably well known through mass balance studies (e.g. Mernild et al 2013), yet its influence upon the ecological and biogeochemical processes in underlying glacial habitats remains largely unexplored. However, the importance of glacial snow cover as a control upon subglacial biogeochemical processes is at least established in the literature. For example, several studies have demonstrated how the seasonal recession of glacial snowpacks greatly influences the evolution of subglacial drainage networks: a process that is linked to the aeration of sediments at the glacier bed (and thus the redox environment that characterises their microbial ecology) and also the delivery of nutrients and organic matter by surface melt (Hodson et al 2008, Tranter et al 2005). As snowlines retreat up-glacier, large fluxes of surface meltwater enter the subglacial environment from the glacier surface, and transform long residence time, poorly oxygenated meltwater flowpaths into well-aerated arteries or conduits at the glacier bed. Sediments in close proximity to these conduits will favour oxic or sub-oxic biogeochemical processes, while those that remain at distance are most likely to remain anoxic, and thus conducive to processes such as sulphate reduction and even methanogenesis, especially beneath ice sheets, where inputs of surface meltwater are negligible (e.g. Wadham et al 2012). However, our characterisation of the redox changes that occur beneath melting ice masses has been largely restricted to outflow measurements of stable isotopes and dissolved oxygen at the ice margin (e.g. Wynn et al 2006, Irvine-Fynn and Hodson 2010), or in-situ measurements via invasive boreholes (e.g. Christner et al 2014). Understanding the spatio-temporal variation of redox conditions beneath ice masses will therefore remain a major research priority for the future, because it will reveal new insights into subglacial ecology and show how the redox-sensitive acquisition of certain key nutrients (such as N, Fe and S) by meltwaters will change with increasing melt.

Our understanding of the snowpack as an ecosystem in its own right (rather than as a mere mediator of the abiotic conditions that lie beneath it) is also beginning to improve, but again, this focus is not sufficiently directed towards glacial snow covers. For example, Larose *et al* (2013) show how microbial community structure and nitrogen cycling within coastal tundra snows may be identified using molecular techniques during spring melt and in spite of the low temperatures that limit water availability for metabolic processes. This work follows a number of other important

studies of 'dry snow' biology and biogeochemistry, some of which have emphasised the role of clay particles or dust in biogeochemical transformation of nitrogen (Amoroso *et al* 2009). These reduce the likelihood of its photolytic loss to the atmosphere (e.g. Beine *et al* 2003, Björkman *et al* 2014), and therefore enhance the opportunities for assimilation into cryospheric ecosystems. The nitrogen cycle therefore remains one of the best examples of how closely the biological, chemical and physical processes can be integrated in cryospheric ecosystems. In spite of this, there still persists a tendency for studies to depend heavily upon just one technique or approach.

Takeuchi et al (2013) compared the microbial dynamics in seasonal (ablation area) and perennial (accumulation area) glacial snowpacks, revealing distinct differences in their algal taxa (e.g. Cd. Nivalis on snow and A. nordenskioldii, M. berggrenii and the Oscillatoracea on the ice). The duration and intensity of algal 'blooms' within these different habitats is an important research problem, because they can increase surface melting by reducing the snow/ice reflectance (albedo). This particular issue is attracting most research attention on the Greenland Ice Sheet, because there are strong reasons to expect autotrophic processes to expand rapidly across the low gradient upper ablation area of this ice sheet in future (e.g. Uetake et al 2010, Wientjes et al 2010, Yallop et al 2012), Cook et al 2012, Lutz et al 2014.

Ice surface habitats

Glacier surfaces are characterised by inorganic and organic debris layers that support significant rates of biological activity when liquid water becomes available. The bioflocculation of dust and other impurities such as black carbon upon this layer produce aggregate particles called cryoconite (e.g. Takeuchi et al 2010, Hodson 2014). The structure and persistence of cryoconite is becoming increasingly well researched, with conceptual models of cell-mineral attachment akin to those employed in biofilm research being used to explain the role of exudates in the cementation of the aggregate (Hodson et al 2010, Takeuchi et al 2010). Zarsky et al (2013) and Langford et al (2011) studied perhaps the largest aggregate clasts yet described in the literature, whose formation was enhanced by ammonium deposition from nearby sea birds, and reactive clay particles from cliffs overlooking the Aldegondabreen glacier (Svalbard). Interestingly, the size of these aggregates and the presence of labile carbon enabled low redox environments to develop, and thus form anoxia in an environment that is otherwise conducive to oxygenation (Hodson et al 2008). The proximity of Aldegondabreen to local sources of guano and fine sediment supply also gave the large aggregates studied by Zarsky et al (2013) the capacity to support ammonia-oxidising archaea. Local environmental factors therefore influence the functional activity of

microorganisms on glacier surfaces. However, while the influence of bird colonies is now established (Mindl et al 2007, Grzesiak et al 2015), other environmental factors and gradients are less understood (e.g. Stibal et al 2011). Recent insights have been provided by diatom biogeography in cryoconite-filled surface melt pools ('cryoconite holes') of the arid McMurdo Dry Valleys, Antarctica (Stanish et al 2013), which showed a clear increase in diatom species richness with distance from the coast, further influence by nearby ephemeral streams and/or perennial (ice covered) lakes, and a sediment size effect. The sediment size effect suggests that a greater understanding of sediment provenance and transport history should be sought. For example, Nagatsuka et al (2014) showed how the long-range transport of fine aeolian particles may be established using their rare earth element composition and then used to understand bioflocculation into Asian cryoconite. Future work should therefore integrate the techniques described above to understand better the linkages between sediment provenance and microbial diversity. Studies of the long-term changes in the storage of dust and associated microorganisms in large ice sheets should also be included, because ice core analyses from the Greenland Ice Sheet have revealed clear evidence for longerterm changes forced by dust deposition over glacial cycles (Miteva et al 2009).

The above synthesis shows that there exists a clear bias towards cryoconite research within the literature which needs to be addressed before we understand the cascade of nutrients, energy and life that is initiated by seasonal melting on glacier surfaces. However, the use of cryoconite as a model ecosystem for the exploration of tight linkages between physical, chemical and biological drivers should not be overlooked. For example, Bellas et al (2013) show that since there is a food web within cryoconite dominated by microorganisms, viruses can play an important role in bacterial mortality and the subsequent release of nutrients and organic carbon for recycling. The same processes are invoked in streams upon McMurdo Dry Valley glaciers by Foreman et al (2013). Quantifying this biological driver of DOC cycling and considering it alongside climate forcing is a research priority for the future, because it is increasingly clear that glacial meltwaters are distinctive on account of the high proportion of labile, proteinaceous organic matter within their DOC pool (e.g. Spencer et al 2014). Modern molecular techniques should also be employed to pinpoint the precise microorganisms and compounds involved, because much of the recent evidence seems to suggest that the labile DOC is produced in-situ (Barker et al 2006).

Despite conceptual advances in our understanding of snow and ice surface microbial communities and their potential feedbacks to melting and runoff nutrient transfer, the direction and magnitude of net ecosystem exchange (NEE) of CO₂ with the atmosphere

remains unclear. Great emphasis is currently placed upon the importance of autotrophic processes on account of their albedo-reducing potential (e.g. Yallop et al 2012) and likely importance for the generation of labile DOC. However, Edwards et al (2013a) used molecular techniques to reveal how the phylogenetic composition of microbial communities in cryoconite debris can be dominated by heterotrophic bacteria in the cryoconite of a north-facing Alpine glacier. This work also showed how functional genes were most likely adapted to the exploitation of allochthonous C, N and P. Similarly, subsidy of allochthonous organic matter and a likely net heterotrophic ecosystem have been emphasised in the study of Svalbard glaciers (Stibal et al 2008). Furthermore, debris-covered glaciers, where the thickness of surficial debris is sufficient to suppress ablation (rather than promote it) are even more likely to promote heterotrophic processes (e.g. Franzetti et al 2013). However, although these glacier types have been subject to limited investigation, recent CO₂ flux measurements have revealed that the debris layer can in fact be a significant sink of CO₂, perhaps due to weathering processes (Wang et al 2014).

The most recent NEE model that included both heterotrophic and autotrophic production on the Greenland Ice Sheet indicated a net autotrophic system dominated by micro-algae rather than cryoconite (Cook et al 2012). Therefore we must challenge the view that biological production on glacier surfaces is dominated by cryoconite before we can deduce whether glaciers are net sources or sinks of atmospheric CO₂. The causes and likely distribution of net heterotrophy also deserves equal attention. One of the greater uncertainties in this context is the so-called bacterial growth efficiency, which Foreman et al (2013) suggest is likely to be as low as 1—2% in a range of ice surface habitats and thus capable of explaining a large proportion of the CO₂ transfer to the atmosphere that is inferred from incubation studies (e.g. Telling et al 2012). However, the empirical approaches used to calculate this crucial parameter might not be appropriate for supraglacial ecosystems.

Supraglacial streams and lakes

Supraglacial streams are crucial vectors in glacial ecosystems, yet their role in moderating (rather than merely advecting) the flux of nutrients, organic matter and cells downstream during the melt season remains unclear. A study on Mendenhall Glacier in Alaska was perhaps the first to directly assess nutrient conservation in these streams by using a tracer addition approach, finding evidence for DOC retention and NO₃ production (Scott *et al* 2010). Work by Foreman *et al* (2013) considered the ecology and biogeochemistry of supraglacial streams of Antarctica, and emphasised how they are subject to high UV irradiance levels that are harmful for aquatic life. In many aquatic ecosystems, humic substances play an important role

in the absorption of UV radiation. However, intense photo-degradation, low rates of production and flushing by meltwater contributed to their absence in the streams of Foreman et al's study. As a result, Foreman et al (2013) describe how other adaptations, including pigmentation and ice nucleation, were important for success, and enabled an autotrophic community to support a heterotrophic bacterial community through the production of young, labile (proteinaceous) DOM. By contrast, Cawley et al (2013) found that stable hypersaline coastal ponds in Antarctica develop humic substances to a far greater degree than freshwater supraglacial lakes and streams described above. This occurs without any plant cover within the watershed and enabled the researchers to describe a muchneeded reference microbial fulvic acid from this environment. The results therefore suggest that photo-degradation is not the most important factor governing the lack of humic substances upon glaciers, because the process also occurs on ice shelf and coastal ponds where humic substances accumulate. Ice shelf ponds perhaps make better cases for comparison to glacier surface lakes than coastal ponds, although their abiotic conditions can be surprisingly different. For example, Sorrel et al (2013) showed that primary production was greatest in basal brines of the stratified ponds: conditions that are clearly unlikely on glacier surfaces. They also found N limitation and a tight coupling of primary and secondary production there, with the former dominating by far. Surprisingly, the N limitation existed in spite of a large and potentially available organic C and N pool. The recalcitrance of the older organic nutrients accumulating in the ice shelf lakes clearly deserves further attention, because it is at odds with the emerging paradigm of labile autochthonous DOM production in ice surface ecosystems.

The excess of primary production over bacterial production in some icy habitats and the reverse situation (net heterotrophy) in others suggest that the lability and composition of DOM exported by meltwaters might vary according to ambient light conditions and nutrient availability. Therefore, unless viral lysis is a dominant control upon DOM acquisition by meltwater, there should be detectable temporal and spatial changes in the lability of DOM draining glaciers and ice sheets. To date, however, there are an insufficient number of studies with which to assess this. Furthermore, any changes in DOM characteristics due to variations in *in-situ* production and processing need to be isolated from a strong signal associated with DOM elution throughout snow cover depletion (Meyer and Wania 2011) and seasonal inputs from ice-marginal ecosystems (Spencer et al 2014). Until experimental work has isolated these various controls, the impact of labile DOM export from glaciers and ice sheets will remain unclear. Further, the fluorometric techniques that dominate the study of DOM composition need to

augmented with other techniques, because not all DOM is detectable by fluorescence, and the molecular composition of the DOM needs to be better understood (Singer *et al* 2012). Recent research suggests Fourier Transform Ion Cyclotron Resonance Mass Spectrometry (Singer *et al* 2012) and Nuclear Magnetic Resonance (Paulter *et al* 2012) show promise.

The use of space-time substitution in the study of

Beyond the ice limit

glacier forefields has made them valuable observatories for studying ecosystem succession. This research has begun to give increasing emphasis to the role of microbial processes in biogeochemical cycles and soil development (Schütte et al 2009, Zumsteg et al 2012, Bernasconi et al 2011, Bradley et al 2014), and therefore complements the studies of higher trophic levels that have dominated in the past (e.g. Chappin et al 1994). For example, Brunner et al (2014) examined pioneering fungi in the Damma Glacier forefield—and showed that a heterogeneous range of carbohydrates typified early ecosystem development and influenced which particular organic acids are used to mine nutrients like phosphorus from glacial till. The efficacy of these processes, along with inorganic acid production by other heterotrophs and the extended opportunities for rock-water interaction that young glacial till affords (Engstrom et al 2000), are most likely the key reasons why nitrogen and carbon are usually reported as the limiting nutrient following deglaciation (Bernasconi et al 2011). The immediate question to arise from this work is how influential fungi and yeasts are in the other habitats where debris is present, such as in cryoconite and subglacial till (e.g. Butinar et al 2007, Turchetti et al 2008, Edwards et al 2013b). Frey et al (2013) gave emphasis to the microbial autotrophic community in the Damma Glacier forefield and also found that heterogeneity within different successional stages was greater than expected. In this case it was shown how cyanobacteria and green algae communities along the successional gradient clearly shifted and that each soil environment selected for its own photoautotrophic community. These studies therefore show how autotrophic and heterotrophic microbial communities deserve more research attention before the links between ecological succession and soil development can be fully understood. Further, they are important for developing a better understanding of how expanding glacier forefields influence aquatic biogeochemistry (Engstrom et al 2000, Milner and Robertson 2010) and the composition of nutrients exported to downstream ecosystems (e.g. Nowak et al 2014).

Conclusions and recommendations for future research

Over the last few decades, the traditional view that glaciers and ice sheets are passive, abiotic cryospheric environments has been justifiably overturned in favour of a paradigm that treats them as one of Earth's most vulnerable biomes. It is now very clear that cryospheric environments support relatively simple yet important ecosystems, whose tight integration between physical, chemical and biological processes offer ideal opportunities for conducting research with a broad appeal to ecologists. This research is now being given a sense of great urgency on account of the vulnerability of icy habitats to climate change (Hood et al 2014, Yde et al 2011) and the compelling cases being made for developing a better understand the consequences of nutrient and organic matter losses from ice to downstream ecosystems. The following future research priorities deserve attention in these contexts:

- We must continue working across the 'ice divide' and integrate expertise from glacial and non-glacial research. It is very clear that the wider importance of glaciers in global biogeochemical cycles needs transdisciplinary interaction among researchers at dedicated conferences and workshops, because there is still too much fragmentation to address the key uncertainties.
- Quantitative, not qualitative, insights into the sensitivity of cryospheric ecosystems to climate change must be sought. Particular attention needs to be given to how small temperature increases might give rise to very significant changes in the biological production upon the flatter parts of polar ice sheets and ice shelves, where the expanse of water-soaked snow could increase markedly in the near future.
- The representation of the above processes in conceptual or physically-based models must improve as we move away from empirical understanding and adopt a more mechanistic approach. For example, models of carbon, water and nutrient fluxes are urgently needed to improve understanding of how polar ice sheets fertilise adjacent seas. Critical feedbacks such as the influence of biological production upon ice albedo and thus melting also require modelling. At present, models remain conspicuous by their absence in glacial cryospheric ecology, yet there are several being used to explore sea ice biogeochemistry and biological production from which we can draw inspiration (e.g. Pogson et al 2011).
- Particular emphasis has been given to the surface processes in this synthesis. This was no coincidence:

surface meltwater is the key vector for understanding how the transfer of nutrients, cells and organic matter will respond to climate change. However, atmospheric scientists and aerobiologists must help us understand the inoculation of such habitats. Further, subglacial environments will also play a crucial role in regulating surface-derived meltwater outputs. The community therefore needs to continue to address the great technological challenges associated with sampling and monitoring the environments above and below the ice surface.

With these research drivers it is hoped that we can begin to understand the biogeochemical and ecological consequences of global mass balance decline in the cryosphere.

Acknowledgments

Hodson acknowledges an award from the Natural Environment Research Council (NE/H014446/1).

References

Amoroso A, Domine F, Esposito G, Morin S, Savarino J,
Nardino M and Beine H J 2009 Microorganisms in dry
polar snow are involved in the exchanges of reactive nitrogen
species with the atmosphere *Environ. Sci. Technol.* 44
714–9

Anesio A M and Laybourn-Parry J 2012 Glaciers and ice sheets as a biome *Trends in Ecology and Evolution* 27 219–25

Bardgett R D *et al* 2007 Heterotrophic microbial communities use ancient carbon following glacial retreat *Biol. Lett.* 3 487–90

Barker J D, Sharp M J, Fitzsimons S J and Turner R J 2006 Abundance and dynamics of dissolved organic carbon in glacier systems *Arctic, Antarctic, and Alpine Research* 38 163–72

Beine H J, Domine F, Ianniello A, Nardino M, Allegrini I, Teinilä K and Hillamo R 2003 Fluxes of nitrates between snow surfaces and the atmosphere in the European high Arctic *Atmos. Chem. Phys.* 3 335–46

Bellas C M, Anesio A M, Telling J, Stibal M, Tranter M and Davis S 2013 Viral impacts on bacterial communities in Arctic cryoconite *Environ. Res. Lett.* 8 045021

Bernasconi S M 39 others 2011 Chemical and biological gradients along the Damma glacier soil chronosequence, Switzerland *Vadose Zone J.* **10** 867–83

Björkman M P et al 2014 Nitrate postdeposition processes in Svalbard surface snow J. of Geophysical Research: Atmospheres 119 12–953

Bradley J A, Singaraye J S and Anesio A M 2014 Microbial community dynamics in the forefield of glaciers *Proc. R. Soc.* B 281 20140882

Brunner I, Goren A and Schlumpf A 2014 Patterns of organic acids exuded by pioneering fungi from a glacier forefield are affected by carbohydrate sources *Environ. Res. Lett.* 9 025002

Butinar L, Spencer-Martins I and Gunde-Cimerman N 2007 Yeasts in high Arctic glaciers: the discovery of a new habitat for eukaryotic microorganisms *Antonie van Leeuwenhoek* 91 277–89

Cawley K M, McKnight D M, Miller P, Cory R, Fimmen R L, Guerard J, Dieser M, Jaros C, Chin Y-P and Foreman C 2013 Characterization of fulvic acid fractions of dissolved organic matter during ice-out in a hyper-eutrophic, coastal pond in Antarctica *Environ. Res. Lett.* 8 045015

- Chapin F S, Walker L R, Fastie C L and Sharman L C 1994 Mechanisms of primary succession following deglaciation at Glacier Bay, Alaska *Ecological Monographs* 64 149–75
- Christner B C et al 2014 A microbial ecosystem beneath the West Antarctic ice sheet Nature 512 310–3
- Cook J M, Hodson A J, Anesio A M, Hanna E, Yallop M, Stibal M, Telling J and Huybrechts P 2012 An improved estimate of microbially mediated carbon fluxes from the Greenland ice sheet J. Glaciol. 58 1098–108
- Edwards A, Pachebat J A, Swain M, Hegarty M, Hodson A J, Irvine-Fynn T D and Sattler B 2013a A metagenomic snapshot of taxonomic and functional diversity in an alpine glacier cryoconite ecosystem *Environ. Res. Lett.* **8** 035003
- Edwards A, Douglas B, Anesio A M, Rassner S M, Irvine-Fynn T D, Sattler B and Griffith G W 2013b A distinctive fungal community inhabiting cryoconite holes on glaciers in Svalbard *Fungal Ecol.* 6 168–76
- Engstrom D R, Fritz S C, Almendinger J E and Juggins S 2000 Chemical and biological trends during lake evolution in recently deglaciated terrain *Nature* 408 161–6
- Fogg G E 1967 Observations on the snow algae of the South Orkney Islands *Proc. R. Soc.* B **252** 279–87
- Foreman C M, Cory R M, Morris C E, San Clements M D, Smith H J, Lisle J T, Miller P L, Chin Y-P and McKnight D M 2013 Microbial growth under humic-free conditions in a supraglacial stream system on the Cotton Glacier Antarctica Environ. Res. Lett. 8 035022
- Franzetti A, Tatangelo V, Gandolfi I, Bertolini V, Bestetti G, Diolaiuti G, D'Agata C, Mihalcea C, Smiraglia C and Ambrosini R 2013 Bacterial community structure on two alpine debris-covered glaciers and biogeography of Polaromonas phylotypes *ISME J.* 7 1483–92
- Frey B, Bühler L, Schmutz S, Zumsteg A and Furrer G 2013 Molecular characterization of phototrophic microorganisms in the forefield of a receding glacier in the Swiss Alps *Environ*. *Res. Lett.* 8 015033
- Gerringa L J, Alderkamp A C, Laan P, Thuroczy C E, De Baar H J, Mills M M, van Dijken GL, van Haren H and Arrigo K R 2012 Iron from melting glaciers fuels the phytoplankton blooms in amundsen sea Southern Ocean: iron biogeochemistry *Deep* Sea Research: II. Topical Studies in Oceanography 71 16–31
- Grzesiak J, Górniak D, Świątecki A, Aleksandrzak-Piekarczyk T, Szatraj K and Zdanowski M K 2015 Microbial community development on the surface of Hans and Werenskiold Glaciers Svalbard, Arctic: a comparison *Extremophiles* 19 885–97
- Hodson A J 2014 Understanding the dynamics of black carbon and associated contaminants in glacial systems Wiley

 Interdisciplinary Reviews: Water 1 141–9
- Hodson AJ, Anesio A M, Tranter M, Fountain A G, Osborn M, Priscu J, Laybourn-Parry J and Sattler B 2008 Glacial Ecosystems *Ecological Monographs* **78** 41–67
- Hodson A J, Cameron K A, Bøggild C E, Irvine-Fynn T D, Langford H, Pearce D A and Banwart S A 2010 The structure, biological activity and biogeochemistry of cryoconite aggregates upon an Arctic valley glacier: Longyearbreen, Svalbard J. Glaciol. 56 349–62
- Hood E and Scott D T 2008 Riverine organic matter and nutrients in southeast Alaska affected by glacial coverage *Nature Geosci.* 1
- Hood E, Battin T J, Fellman J, O'Neel S and Spencer R G 2015 Storage and release of organic carbon from glaciers and ice sheets *Nature Geosci.* 8 91–6
- Irvine-Fynn T D L and Hodson A J 2010 Biogeochemistry and dissolved oxygen dynamics at a subglacial upwelling, Midtre Lovénbreen, Svalbard *Ann. Glaciol.* 51 41–6
- Langford H, Hodson A and Banwart S 2011 Using FTIR spectroscopy to characterise the soil mineralogy and geochemistry of cryoconite from Aldegondabreen glacier, Svalbard *Appl. Geochem.* 26 S206–9
- Larose C, Dommergue A and Vogel T M 2013 Microbial nitrogen cycling in Arctic snowpacks Environ. Res. Lett. 8 035004

- Laybourn-Parry J, Tranter M and Hodson A J 2012 *The Ecology of*Snow and Ice Environments (Oxford: Oxford University Press)
- Luus K A, Kelly R E J, Lin J C, Humphreys E R, Lafleur P M and Oechel W C 2013 Modeling the influence of snow cover on low Arctic net ecosystem exchange *Environ. Res. Lett.* 8 035045
- Lutz S, Anesio A M, Villar S E J and Benning L G 2014 Variations of algal communities cause darkening of a Greenland glacier FEMS Microbiol. Ecol. 89 402–14
- McLean A L 1918 Bacteria of ice and snow in Antarctica *Nature* 102 35–9
- Mernild S H, Pelto M, Malmros J K, Yde J C, Knudsen N T and Hanna E 2013 Identification of snow ablation rate, ELA, AAR and net mass balance using transient snowline variations on two Arctic glaciers *J. Glaciol.* 59 649–59
- Meyer T and Wania F 2011 Modeling the elution of organic chemicals from a melting homogeneous snow pack *Water Res.* 45 3627–37
- Milner A M and Robertson A L 2010 Colonization and successional theory? River Res. Appl. 26 26–35
- Mindl B, Anesio A M, Meirer K, Hodson A J, Laybourn-Parry J, Sommaruga R and Sattler B 2007 Factors influencing bacterial dynamics along a transect from supraglacial runoff to proglacial lakes of a high Arctic glacier *FEMS Microbial Ecol.* 59 307–17
- Miteva V, Teacher C, Sowers T and Brenchley J 2009 Comparison of the microbial diversity at different depths of the GISP2 Greenland ice core in relationship to deposition climates *Environ. Microbiol.* 11 640–56
- Mosser J L, Mosser A G and Brock T D 1977 Photosynthesis in the snow: the alga *Chlamydomonas nivalis* Chlorophyceae *J. Phycol.* **13** 22–7
- Nagatsuka N, Takeuchi N, Nakano T, Shin K and Kokado E 2014 Geographical variations in Sr and Nd isotopic ratios of cryoconite on Asian glaciers *Environ. Res. Lett.* **9** 045007
- Nowak A and Hodson A 2014 Changes in meltwater chemistry over a 20-year period following a thermal regime switch from polythermal to cold-based glaciation at Austre Brøggerbreen, Svalbard *Polar Res.* 33 22779
- Nowak A and Hodson A 2015 On the biogeochemical response of a glacierized High Arctic watershed to climate change: revealing patterns, processes and heterogeneity among micro-catchments *Hydrol. Process.* 29 1588–603
- Pautler B G, Woods G C, Dubnick A, Simpson A J, Sharp M J, Fitzsimons S J and Simpson M J 2012 Molecular characterization of dissolved organic matter in glacial ice: coupling natural abundance 1H NMR and fluorescence spectroscopy *Environ. Sci. Technol.* 46 3753–61
- Pogson L, Tremblay B, Lavoie D, Michel C and Vancoppenolle M 2011 Development and validation of a one-dimensional snow-ice algae model against observations in Resolute Passage, Canadian Arctic Archipelago *J. Geophys. Res.* 116 C4
- Priscu J C, Tulczyk S, Studinger M, Kennicutt M C, Christner B C and Foreman C M 2008 Antarctic subglacial water: origin, evolution and ecology *Polar Lakes and Rivers* ed W F Vincent and J Laybourn-Parry (New York: OUP) ch 7
- Roberts T J, Hodson A J, Evans C D and Holmen K 2010 Modelling the impacts of a nitrogen pollution event on the biogeochemistry of an Arctic glacier *Ann. Glaciol.* 51 163–70
- Schroth A W, Crusius J, Chever F, Bostick B C and Rouxel O J 2011 Glacial influence on the geochemistry of riverine iron fluxes to the Gulf of Alaska and effects of deglaciation *Geophys. Res. Lett.* 38 16
- Schütte U M, Abdo Z, Bent S J, Williams C J, Schneider G M, Solheim B and Forney L J 2009 Bacterial succession in a glacier foreland of the High Arctic *ISME J.* 3 1258–68
- Scott D, Hood E and Nassry M 2010 In-stream uptake and retention of C, N and P in a supraglacial stream *Ann. Glaciol.* 51 80–6
- Singer G A, Fasching C, Wilhelm L, Niggemann J, Steier P,
 Dittmar T and Battin T J 2012 Biogeochemically diverse
 organic matter in Alpine glaciers and its downstream fate *Nat. Geosci.* 5 710–4

- Sisler F D 1961 Geomicrobiolgy of Antarctica, In: National Academy of Sciences—National Research Council Committee on Polar Research Publication No 839 Science in Antarctica: I. The Life Sciences in Antarctica Washington, DC pp 147–50
- Sorrell B K, Hawes I and Safi K 2013 Nitrogen and carbon limitation of planktonic primary production and phytoplankton—bacterioplankton coupling in ponds on the McMurdo Ice Shelf, Antarctica *Environ. Res. Lett.* 8 035043
- Spencer R G, Vermilyea A, Fellman J, Raymond P, Stubbins A, Scott D and Hood E 2014 Seasonal variability of organic matter composition in an Alaskan glacier outflow: insights into glacier carbon sources Environ. Res. Lett. 9 055005
- Stanish L F, Bagshaw E A, McKnight D M, Fountain A G and Tranter M 2013 Environmental factors influencing diatom communities in Antarctic cryoconite holes *Environ. Res. Lett.* 8 045006
- Stibal M, Telling J, Cook J, Mak K M, Hodson A and Anesio A M 2011 Environmental Controls on microbial abundance and activity on the Greenland Ice Sheet: a multivariate analysis approach *Microb. Ecol.* 63 74–84
- Stibal M, Tranter M, Benning L G and Řehák J 2008 Microbial primary production on an Arctic glacier is insignificant in comparison with allochthonous organic carbon input Environ. Microbiol. 10 2172–8
- Takeuchi N 2013 Seasonal and altitudinal variations in snow algal communities on an Alaskan glacier Gulkana glacier in the Alaska range *Environ. Res. Lett.* **8** 035002
- Takeuchi N, Nishiyama H and Li Z 2010 Structure and formation process of cryoconite granules on Ürümqi glacier No 1, Tien Shan, China *Ann. Glaciol.* 51 9–14
- Telling J, Anesio A M, Tranter M, Stibal M, Hawkings J, Irvine-Fynn T, Hodson A J, Butler C, Yallop M and Wadham J 2012 Controls on the autochthonous production and respiration of organic matter in cryoconite holes on high Arctic glaciers J. Geophys. Res. 117 G01017
- Tranter M, Skidmore M and Wadham J 2005 Hydrological controls on microbial communities in subglacial environments *Hydrol. Process.* **19** 995–8

- Turchetti B, Buzzini P, Goretti M, Branda E, Diolaiuti G, D'Agata C, Smiraglia C and Vaughan-Martini A 2008 Psychrophilic yeasts in glacial environments of Alpine glaciers *FEMS Microbiol. Ecol.* **63** 73–83
- Uetake J, Naganuma T, Hebsgaard MB, Kanda H and Kohshima S 2010 Communities of algae and cyanobacteria on glaciers in west Greenland *Polar Science* $4\,71-80$
- Wadham J L et al 2012 Potential methane reservoirs beneath Antarctica Nature 488 633–7
- Wadham J L, De'ath R, Monteiro F M, Tranter M, Ridgwell A, Raiswell R and Tulaczyk S 2013 The potential role of the Antarctic Ice Sheet in global biogeochemical cycles Earth Environ. Sci. Trans. R. Soc. Edinburgh 104 55–67
- Wang J, Haidong H and Zhang S 2014 Carbon dioxide flux in the ablation area of Koxkar Glacier, western Tien Shan, China Ann. Glaciol. 55 231–8
- Wientjes I G M, Van de Wal R S W, Reichart G J, Sluijs A and Oerlemans J 2010 Dust from the dark region in the western ablation zone of the Greenland ice sheet *Cryosphere* 5 589–601
- Wynn P M, Hodson A J and Heaton T 2006 Chemical and isotopic switching within the subglacial environment of a High Arctic glacier *Biogeochem.* **78** 173–93
- Yallop M L et al 2012 Photophysiology and albedo-changing potential of the ice algal community on the surface of the Greenland ice sheet ISME J. 6 2302–13
- Yde J C, Finster K W and Bárcena T G 2011 Subglacial and proglacial ecosystem responses to climate change ed J Blanco and H Kheradmand *Climate Change—Geophysical Foundations and Ecological Effects* (Rijeka, Croatia: INTECH) ch 23
- Zarsky J D, Stibal M, Hodson A, Sattler B, Schostag M, Hansen L H, Jacobsen C S and Psenner R 2013 Large cryoconite aggregates on a Svalbard glacier support a diverse microbial community including ammonia-oxidizing archaea *Environ. Res. Lett.* 8 035044
- Zumsteg A, Luster J, Göransson H, Smittenberg R H, Brunner I, Bernasconi S M, Zeyer J and Frey B 2012 Bacterial, archaeal and fungal succession in the forefield of a receding glacier *Microbial Ecol.* 63 552–64