

Crystal and molecular structure of a histo-blood group antigen involved in cell adhesion: the Lewis x trisaccharide

Serge Pérez⁴, Nadine Mouhous-Riou,
Nikolay E. Nifant'ev¹, Yury E. Tsvetkov¹, Bernard Bachet²
and Anne Imberty^{3,4}

Ingenierie Moléculaire, Institut National de la Recherche Agronomique, BP 1627, F-44316 Nantes Cedex 03, France, ¹Zelinsky Institute of Organic Chemistry, Leninsky Prospect 47, Moscow B-334, 117913 Russia, ²Laboratoire de Minéralogie Cristallographie, Département des Macromolécules Biologiques, 4 Place Jussieu, 75252 Paris Cedex 05, France, and ³Laboratoire de Synthèse Organique, Centre National de la Recherche Scientifique, 2 Rue de la Houssinière, F-44072 Nantes Cedex 03, France

⁴To whom correspondence should be addressed

This work describes the first crystal structure ever reported of a histo-blood group carbohydrate antigen: Le^x. This study provides a detailed description of the conformation of two crystallographic independent molecules in a highly hydrated environment along with their hydrogen bonding properties and packing features. Some interactions observed between adjacent trisaccharides can provide the basis for involvement of Le^x–Le^x interactions in cell-cell adhesion.

Key words: histo-blood group/cell adhesion/carbohydrate-carbohydrate interactions/Lewis x/trisaccharide

Introduction

Histo-blood group carbohydrate-dependent antigens are borne by glycolipids and glycoproteins which are expressed on human erythrocytes and also in the epithelia of glandular tissues, primary sensory neurons, and exocrine secretions in man and other mammals (Oriol, 1995; Henry and Oriol, 1995). Two antigens families, the ABH(O) and the Lewis determinants, constitute the major histo-blood group carbohydrate determinant (Clausen and Hakomori, 1989). They exhibit a pattern of appearance and disappearance on defined cells in particular tissues, representing developmental and specific functions.

The Lewis x (Le^x) determinant (Figure 1) is a trisaccharide [Galβ1–4[Fucα1–3]GlcNAcβ-1 (Hakomori *et al.*, 1981). In mammals, this oligosaccharide is a stage-specific embryonic antigen (SSEA) (Solter and Knowles, 1978), implicated in the compaction of the morula (Bird and Kimber, 1984; Fenderson *et al.*, 1984) and is also a tumor-associated marker (Feizi, 1985; Hakomori, 1989). This trisaccharide has been recently found on the cell surface of *Helicobacter pylori*, a human pathogenic bacteria associated with gastric ulcer and cancer (Aspinall *et al.*, 1994). It is also present at the surface of the infecting phase of *Schistosoma mansoni* (Vellupillai and Harn, 1994), having a signaling role in the interaction between this parasitic worm and its human host (Van Dam *et al.*, 1994). The sialyl Le^x tetrasaccharide and its sulfated analog have also been recognized as important ligands in cellular adhesion, playing a

role in the inflammatory response through their interaction with selectins (Lasky, 1992).

Conformational studies of histo-blood group carbohydrates first appeared in the beginning of the 1980s (Biswas and Rao, 1980; Lemieux *et al.*, 1980). Important achievement in the chemistry of synthetic oligosaccharides, along with the availability of conformational analysis methods coupled to high resolution NMR characterizations, set the foundation for such studies (see Pérez *et al.*, 1994, for review). They have provided a general description of the molecular conformations in solution, but no direct information concerning their hydrogen bonding properties and packing features. At the moment, no such biologically active oligosaccharides have yet been crystallized.

Results

Description of the crystal

Starting from chemically synthesized Le^x trisaccharide methyl glycoside, single crystals could be grown. They were crystallized by slow evaporation of Le^x dissolved in a water/ethanol mixture. Over a period of 2 years, more than 20 crystals having dimensions suitable for x-ray investigations have been obtained. One single crystal having dimensions of 0.5 × 0.25 × 0.05 mm was mounted on a glass pin with its long dimension along the pin axis. Le^x crystallizes in the monoclinic space group P2₁ with unit cell parameters: $a = 12.147(6)$, $b = 27.552(9)$, $c = 8.662(6)$ Å, $\beta = 91.71^\circ(5)$. In such a unit cell, the asymmetric unit contains two independent molecules, and an unusually high number of water molecules. All hydrogen atoms of the O-H groups and of the water molecules could be located in this crystal structure, allowing a straightforward assignment of hydrogen bonds. The coordinates of all atoms are listed in Table 1 and Table 2

The trisaccharide conformation

The six monosaccharide units in the two Le^x molecules adopt the typical ⁴C₁ chair conformation, with no significant deviation away from classical pyranose ring shape. The N-acetyl group adopts a *trans* conformation, whereas the primary hydroxyl groups of the Gal and GlcNAc are *trans-gauche* and *gauche-trans*, respectively. The two Le^x molecules differ in their overall conformations (Figure 2). These differences are essentially located at the glycosidic torsion angles at the Galβ(1–4)GlcNAc linkage for which angle $\Phi 1$ differs by 10° (Table 3). Neither of the two trisaccharides exhibits any intramolecular hydrogen bonds. A strong interaction exists between the fucose and galactose ring, but only nonpolar van der Waals contacts are involved, each ring presenting its most hydrophobic face to the other one. Conformational studies using NMR and/or molecular modeling (Thogersen *et al.*, 1982; Ichikawa *et al.*, 1992; Miller *et al.*, 1992; Imberty *et al.*, 1995) generally

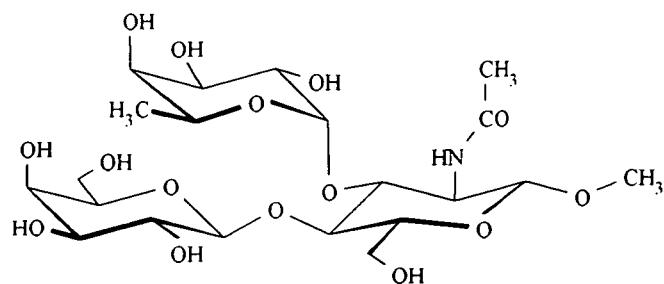


Fig. 1. Schematic representation of Le^x methyl glycoside: methyl 2-acetamido-4-O-(β -D-galactopyranosyl)-3-O-(α -L-fucopyranosyl)-2-deoxy- β -D-glucopyranoside.

agree on a single conformation for Lewis x in solution, corresponding closely to the one reported here.

Hydration in the crystal

The nine water molecules present in the asymmetric unit are arranged in a cluster-like fashion. They establish hydrogen

bonds to other water molecules within the cluster and to the surrounding carbohydrate molecule. They are arranged to fill an empty space in the crystal packing. Six and seven water molecules, respectively, are involved in the hydration of the two trisaccharides (Figure 3). When comparing the two trisaccharides, the conserved hydration sites are the one accepting the amide hydrogen from the N-acetyl group and the one giving hydrogen to the oxygen O2fuc. In both trisaccharides, a hydrogen bond chain involving two water molecules links the oxygen atoms O6gln and O2gal but with different geometries.

Hydrogen bonds network

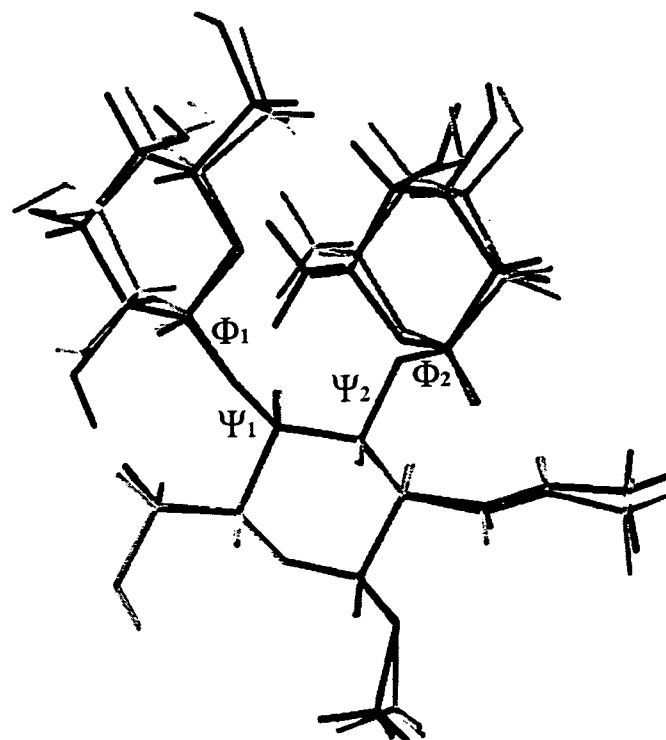
The two independent trisaccharides of the asymmetric unit are linked by four hydrogen bonds, two of them being part of a three-centered bond (O6gln \rightarrow O7gln', O3fuc' \rightarrow O7gln, O2fuc' \rightarrow O1gln, O2fuc' \rightarrow O5gln). Almost the same interaction is produced by one unit translation along the *a* axis. In fact, a pseudo-symmetry element exists between the two independent molecules, therefore creating this apparent high level of symmetry in the *ab* plane (Figure 4a).

Table I. Fractional atomic coordinates (with estimated standard deviations in parentheses) for Le^x

Atom	x	y	z	Atom	x	y	z
C1gln	-0.1944(7)	0.0515(3)	1.049(1)	C1gln'	0.3053(7)	0.1167(3)	0.7283(9)
C2gln	-0.3186(7)	0.0553(3)	1.0447(9)	C2gln'	0.1807(6)	0.1121(3)	0.720(1)
C3gln	-0.3698(6)	0.0047(3)	1.0468(9)	G3gln'	0.1269(6)	0.1620(3)	0.7167(9)
C4gln	-0.3292(6)	-0.0238(3)	0.9068(9)	C4gln'	0.1704(7)	0.1912(3)	0.854(1)
C5gln	-0.2031(6)	-0.0232(3)	0.909(1)	C5gln'	0.2952(6)	0.1935(3)	0.8498(9)
C6gln	-0.1575(7)	-0.0466(3)	0.767(1)	C6gln'	0.3464(6)	0.2205(3)	0.985(1)
C7gln	-0.4069(7)	0.1265(3)	1.153(1)	C7gln'	0.0985(7)	0.0408(3)	0.602(1)
C8gln	-0.4552(8)	0.1490(4)	1.294(1)	C8gln'	0.0645(9)	0.0150(4)	0.455(1)
C9gln	-0.0360(7)	0.1001(4)	1.084(1)	C9gln'	0.4513(8)	0.0614(4)	0.682(1)
O1gln	-0.1496(5)	0.0971(2)	1.0383(7)	O1gln'	0.3491(5)	0.0705(2)	0.7521(8)
O5gln	-0.1624(4)	0.0250(2)	0.9153(6)	O5gln'	0.3396(4)	0.1453(2)	0.8572(7)
O6gln	-0.0411(4)	-0.0531(2)	0.7813(6)	O6gln'	0.4622(4)	0.2280(2)	0.9706(7)
O7gln	-0.4211(5)	0.1466(2)	1.0236(7)	O7gln'	0.0785(5)	0.0226(2)	0.7317(7)
N2gln	-0.3572(5)	0.0844(2)	1.1729(8)	N2gln'	0.1471(5)	0.0830(2)	0.5874(7)
C1gal	-0.4103(6)	-0.0942(3)	0.7807(9)	C1gal'	0.0835(7)	0.2614(3)	0.973(1)
C2gal	-0.4054(6)	-0.1481(3)	0.8060(9)	C2gal'	0.0660(6)	0.3139(3)	0.9314(9)
C3gal	-0.4694(7)	-0.1757(3)	0.684(1)	C3gal'	-0.0025(7)	0.3397(3)	1.048(1)
C4gal	-0.5855(7)	-0.1555(3)	0.668(1)	C4gal	-0.1046(7)	0.3116(3)	1.081(1)
C5gal	-0.5796(7)	-0.1015(3)	0.638(1)	C5gal'	-0.0774(7)	0.2595(3)	1.126(1)
C6gal	-0.6922(8)	-0.0761(3)	0.630(1)	C6gal'	-0.1779(8)	0.2261(3)	1.139(1)
O1gal	-0.3667(4)	-0.0724(2)	0.9180(6)	O1gal'	0.1276(4)	0.2398(2)	0.8397(6)
O2gal	-0.2923(5)	-0.1632(2)	0.8054(7)	O2gal'	0.1708(5)	0.3378(2)	0.9255(7)
O3gal	-0.4695(5)	-0.2262(2)	0.7135(7)	O3gal'	-0.0304(5)	0.3866(2)	0.9886(8)
O4gal	-0.6422(5)	-0.1630(2)	0.8074(7)	O4gal'	-0.1783(5)	0.3105(2)	0.9499(7)
O5gal	-0.5214(4)	-0.0769(2)	0.7611(6)	O5gal'	-0.0161(4)	0.2365(2)	1.0002(7)
O6gal	-0.7638(5)	-0.1000(2)	0.5189(7)	O6gal'	-0.2577(5)	0.2488(2)	1.2335(7)
C1fuc	-0.5411(7)	0.0011(3)	1.1795(9)	C1fuc'	-0.0502(7)	0.1606(3)	0.581(1)
C2fuc	-0.6591(7)	0.0175(3)	1.152(1)	C2fuc'	-0.1616(7)	0.1367(3)	0.600(1)
C3fuc	-0.7184(7)	-0.0152(3)	1.036(1)	C3fuc'	-0.2288(7)	0.1638(3)	0.715(1)
C4fuc	-0.7073(7)	-0.0676(3)	1.085(1)	C4fuc'	-0.2361(7)	0.2176(3)	0.670(1)
C5fuc	-0.5909(7)	-0.0815(3)	1.1225(9)	C5fuc'	-0.1230(7)	0.2386(3)	0.645(1)
C6fuc	-0.5767(8)	-0.1321(3)	1.191(1)	C6fuc'	-0.124(1)	0.2888(3)	0.581(1)
O1fuc	-0.4872(4)	0.0077(2)	1.0384(6)	O1fuc'	0.0096(4)	0.1557(2)	0.7247(6)
O2fuc	-0.6631(5)	0.0679(2)	1.1116(7)	O2fuc'	-0.1503(5)	0.0861(2)	0.6308(7)
O3fuc	-0.8321(5)	-0.0022(3)	1.0247(8)	O3fuc'	-0.3346(5)	0.1429(3)	0.7220(7)
O4fuc	-0.7759(5)	-0.0772(3)	1.2144(7)	O4fuc'	-0.3025(5)	0.2216(2)	0.5298(7)
O5fuc	-0.5383(5)	-0.0476(2)	1.2302(6)	O5fuc'	-0.0623(5)	0.2090(2)	0.5365(7)
Owat1	-0.1013(6)	0.3838(3)	0.4516(8)				
Owat2	0.2043(7)	0.1265(3)	1.3006(9)				
Owat3	-0.3239(7)	0.0381(3)	0.4668(8)				
Owat4	-0.3227	-0.0632	1.4045				
Owat5	-0.105(1)	-0.0773(3)	1.283(1)				
Owat6	0.154(1)	0.3094(3)	0.432(1)				
Owat7	0.2978(7)	0.3230(4)	0.671(1)				
Owat8	0.2810(9)	0.2201(3)	0.396(1)				
Owat9	0.4967(7)	0.2546(4)	0.655(1)				

Table II. Hydrogen atom fractional atomic coordinates for Le^x. Estimated standard deviations on x/a, y/b and z/c amount to 0.009, 0.004 and 0.009, respectively.

Atom	x	y	z	Atom	x	y	z
H1gln	-0.160	0.037	1.145	H1gln'	0.339	0.131	0.633
H2gln	-0.344	0.078	0.950	H2gln'	0.154	0.093	0.812
H3gln	-0.353	-0.013	1.144	H3gln'	0.140	0.180	0.620
H4gln	-0.359	-0.010	0.806	H4gln'	0.142	0.177	0.954
H5gln	-0.175	-0.039	1.006	H5gln'	0.315	0.210	0.751
H61gln	-0.175	-0.026	0.673	H61gln'	0.341	0.198	1.079
H62gln	-0.192	-0.079	0.750	H62gln'	0.317	0.261	0.971
H81gln	-0.491	0.181	1.269	H81gln'	0.026	-0.017	0.478
H82gln	-0.512	0.127	1.339	H82gln'	0.130	0.007	0.391
H83gln	-0.396	0.155	1.377	H83gln'	0.011	0.035	0.390
H91gln	-0.011	0.135	1.073	H91gln'	0.478	0.028	0.702
H92gln	-0.025	0.090	1.194	H92gln'	0.510	0.085	0.721
H93gln	0.009	0.079	1.017	H93gln'	0.445	0.066	0.565
HO6gln	0.001	-0.023	0.780	HO6gln'	0.506	0.198	0.975
HN2gln	-0.348	0.066	1.283	HN2gln'	0.167	0.095	0.488
H1gal	-0.359	-0.083	0.688	H1gal'	0.131	0.256	1.069
H2gal	-0.437	-0.158	0.907	H2gal'	0.029	0.316	0.826
H3gal	-0.430	-0.172	0.586	H3gal'	0.043	0.343	1.145
H4gal	-0.631	-0.171	0.568	H4gal'	-0.147	0.330	1.162
H5gal	-0.541	-0.098	0.538	H5gal'	-0.033	0.261	1.226
H61gal	-0.682	-0.041	0.598	H61gal'	-0.156	0.193	1.179
H62gal	-0.726	-0.076	0.734	H62gal'	-0.213	0.221	1.030
HO2gal	-0.250	-0.156	0.901	HO2gal'	0.164	0.369	0.871
HO3gal	-0.489	-0.236	0.816	HO3gal'	0.028	0.410	1.011
HO4gal	-0.668	-0.196	0.814	HO4gal'	-0.197	0.341	0.912
HO6gal	-0.753	-0.091	0.410	HO6gal'	-0.268	0.232	1.329
H1fuc	-0.498	0.018	1.265	H1fuc'	-0.004	0.147	0.496
H2fuc	-0.703	0.018	1.251	H2fuc'	-0.211	0.136	0.486
H3fuc	-0.688	-0.009	0.930	H3fuc'	-0.194	0.159	0.822
H4fuc	-0.735	-0.088	0.995	H4fuc'	-0.274	0.243	0.752
H5fuc	-0.554	-0.080	1.020	H5fuc'	-0.089	0.239	0.751
H61fuc	-0.612	-0.157	1.121	H61fuc'	-0.166	0.312	0.649
H62fuc	-0.614	-0.134	1.294	H62fuc'	-0.047	0.301	0.571
H63fuc	-0.497	-0.140	1.207	H63fuc'	-0.162	0.289	0.474
HO2fuc	-0.640	0.067	1.000	HO2fuc'	-0.132	0.076	0.737
HO3fuc	-0.859	-0.009	0.923	HO3fuc'	-0.359	0.145	0.825
HO4fuc	-0.853	-0.075	1.184	HO4fuc'	-0.364	0.244	0.536
H1wat1	-0.087	0.390	0.458	H2wat1	0.004	0.397	0.351
H1wat2	0.251	0.112	1.227	H2wat2	0.233	0.158	1.315
H1wat3	-0.262	0.045	0.532	H2wat3	-0.346	0.006	0.501
H1wat4	-0.398	-0.063	1.356	H2wat4	-0.316	-0.098	1.408
H1wat5	-0.162	-0.054	1.328	H2wat5	-0.089	-0.095	1.380
H1wat6	0.195	0.329	0.354	H2wat6	0.083	0.317	0.392
H1wat7	0.279	0.314	0.775	H2wat7	0.226	0.319	0.619
H1wat8	0.356	0.231	0.397	H2wat8	0.246	0.248	0.349
H1wat9	0.505	0.2563	0.767	H2wat9	0.436	0.277	0.646

**Fig. 2.** Comparison of the crystalline conformations of the two independent Le^x molecules.

ones, and almost equivalent ones, exist between the two independent molecules and between one molecule and the translation about *a* of the other. These two interactions create a strong molecular chain along the *a* axis. The two second best interactions result from the rotation about the 2₁ axis. They also generate molecules in the *ab* plane. The *ab* plane displays therefore all the strongest interactions and can be seen as a sheet of Lewis x trisaccharide molecules with strong interconnections (Figure 4). A weaker but still energetically favorable interaction exists between each molecule and its related one along the *c* axis.

Discussion

The interaction between adjacent trisaccharides observed in the crystal structure can be related to several biological effects, which were thought to involve Le^x-Le^x interaction.

Table III. Comparison of the linkage torsion angles^a in the crystal and in solution^b

Linkage	Φ/Ψ		
	Crystal	Solution ^b	
Galβ(1-4)GlcNAc	-80.0/-104.6	-70.5/-107.7	-75/-104
Fucα(1-3)GlcNAc	-72.5/139.2	-76.7/139.0	-81/151

^aTorsion angles at the glycosidic linkages are defined as Ψ₁ = (O5gal-C1gal-O1gal-C4gln), Ψ₁ = (C1gal-O1gal-C4gln-C5gln), Φ₂ = (O5fuc-C1fuc-O1fuc-C3gln), Ψ₂ = (C1fuc-O1fuc-C3gln-C4gln).

^bLowest energy conformation (Imberty *et al.*, 1995) obtained with the MM3 force-field using a high dielectric constant in order to simulate the effect of water environment.

The crystal structure displays an extremely dense network of hydrogen bonds (Figure 5). Thirty-six hydrogen bonds are observed in the asymmetric unit, 30 of them implying water molecules. Each trisaccharide is involved in 21 hydrogen bonds, the ratio per glycosidic residue varying from 5 to 8. Such a high number of hydrogen bonds can be correlated to two facts: the high hydration level and the peculiar folding of the oligosaccharide, burying the hydrophobic faces of the residue and presenting the hydrophilic faces to the external part. The hydrogen bond network shown in Figure 5 has several features: one four-member ring of cooperative hydrogen bonds (→O2gal'→Owat5→Owat4→Owat7→) and three infinite chains. Whereas these features have been described previously in carbohydrate crystals (Jeffrey and Saenger, 1981), this is the first example where they are shown to occur simultaneously.

Description of the packing

Analysis of the packing has been attempted by evaluating the energy of interaction between pairs of neighbors. The best

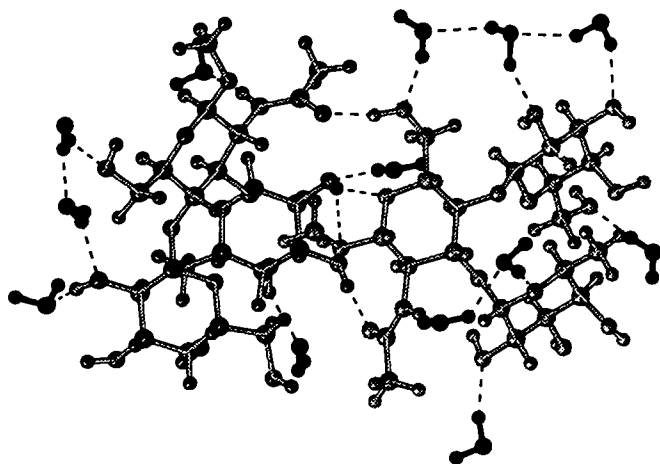


Fig. 3. Hydration shell of the dimer of Le^x trisaccharide in the crystalline form. The two independent molecules have been given a different gray shading, water molecules are shown in black, and dashed lines depict hydrogen bonds.

In several cases, the histo-blood group oligosaccharides have been shown to be more active when associated in clusters (Dean *et al.*, 1993; Varki, 1994). Such clusters are likely to exist at the surface of the cell, either due to high density of O-glycosylation sites on mucin-like proteins, or to the association of the glycosphingolipids in patches in the membrane. From the present crystal structure, the arrangement of trisaccharides along the *c* axis is compatible with this hypothesis (Figure 6). This translation maintains the orientation of the trisaccharide, therefore creating rows of identical molecules compatible with insertion of their carrier in a bilayer membrane. Calculations are in progress to model two-dimensional arrays of glycolipids, using this pairing as starting point.

The specific interaction of Le^x with Le^x has been proposed to be the basis of cell adhesion in morula compaction and

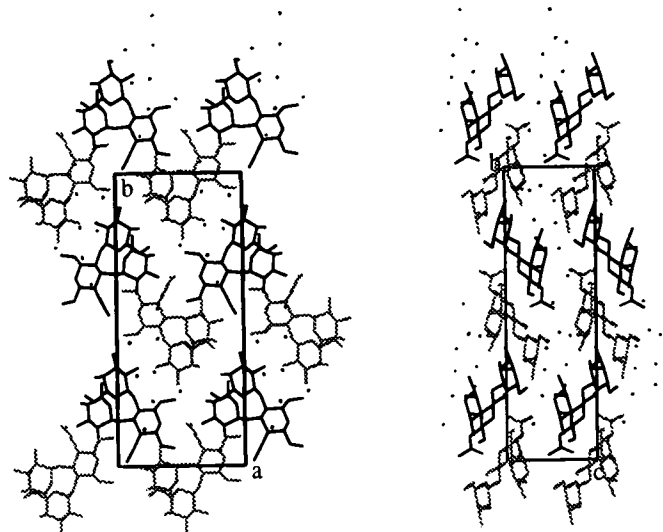


Fig. 4. Packing of Le^x trisaccharide in the *ab* and *bc* planes. The two independent trisaccharide molecules have been given different gray shading. Hydrogen atoms are not displayed.

autoaggregation of F9 teratocarcinoma (Eggens *et al.*, 1989). This hypothesis has been reinforced (Kojima *et al.*, 1994) based on autoaggregation studies of plastic beads coated with glycoconjugates bearing this trisaccharide determinant in the presence of Ca^{2+} . Indeed, this type of interaction, involving fucose and galactose residues, is observed in the crystal, between rows of trisaccharides (Figure 6). The two rows display a head-to-head arrangement which would be compatible with the arrangement required for a cell-cell recognition event. This interaction is one of the most favored in terms of energy since three hydrogen bonds are established ($O4fuc \rightarrow O3gal'$, $O4gal \rightarrow O6gal'$, $O4gal' \rightarrow O3fuc$).

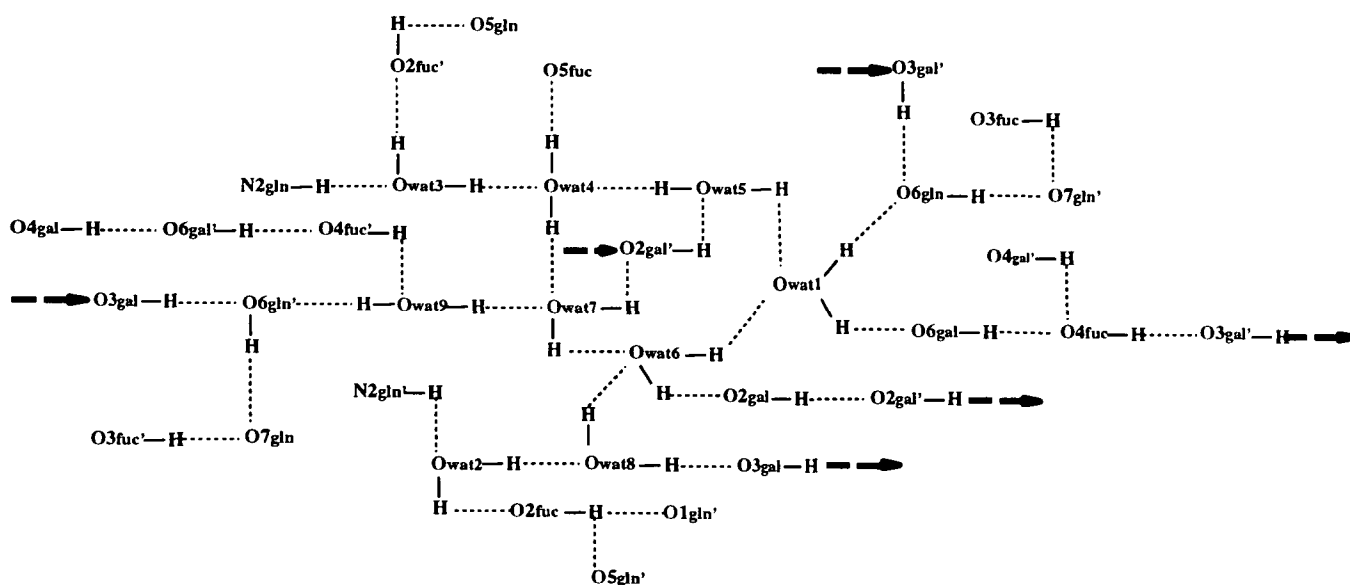


Fig. 5. Schematic representation of the hydrogen bond network in the crystal of Le^x trisaccharide. The three infinite chains are indicated by bold arrows. The atom names are followed by the letters gal, gln, and fuc according to whether they belong to the galactose, N-acetyl galactosamine, and fucose residue, respectively. The two independent molecules are referred to as unprimed and primed. Water molecules are labeled as wat, from 1 to 9.

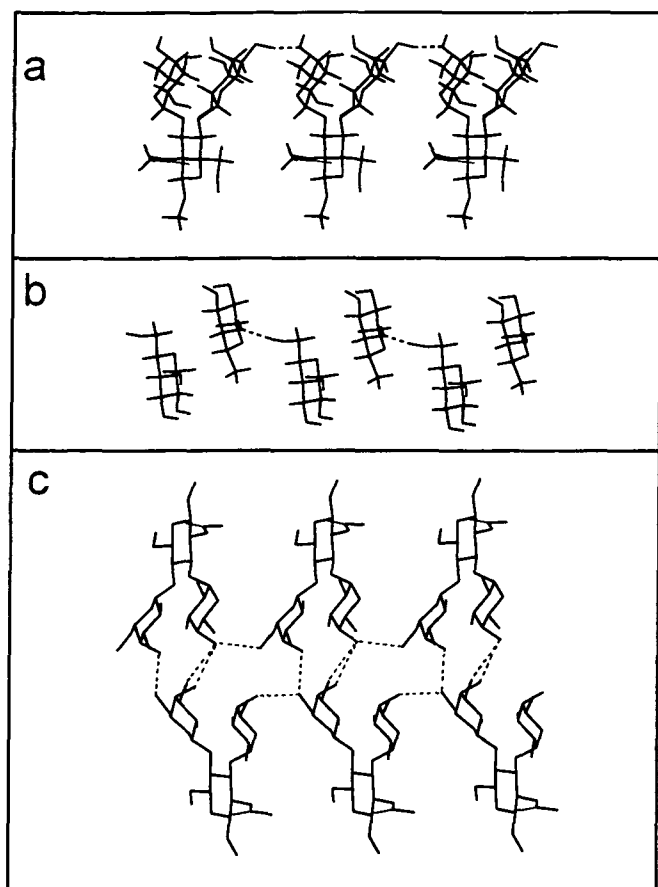


Fig. 6. Packing of Le^x trisaccharide in the crystalline state which may mimic Le^x - Le^x contact in biological conditions. (a) Row along c axis, (b) orthogonal representation of (a) without displaying the GlcNAc residues; (c) head-to-head contacts between two rows of parallel molecules (hydrogen atoms are not displayed).

Material and methods

Synthesis of Lewis x trisaccharide methyl glycoside

Synthesis of Lewis x trisaccharide methyl glycoside (**1**) was accomplished by stepwise elongation of the oligosaccharide chain starting from the formation of lactosamine fragment (see Figure 7). Methyl 6-O-benzoyl-3-O-benzyl-2-deoxy-2-phthalimido- β -D-glucopyranoside (**4**) was used as first glycosyl acceptor. It was prepared by acid hydrolysis of methyl 3-O-benzyl-4,6-O-benzylidene-2-deoxy-2-phthalimido- β -D-glucopyranoside (**2**) (Alais and David, 1990) and subsequent regio-selective 6-O-benzoylation of diol (**3**) (Schwartz *et al.*, 1985). Presence of Bz-group in **4** at O-6 was confirmed by low-field location of the signals of H-6a and H-6b protons and up-field location of the signal of H-4 in 1H -NMR spectrum.

Glycosylation of acceptor **4** by 2,3,4,6-tetra-O-acetyl- α -D-galactopyranosyl bromide (**5**) under promotion with silver triflate gave selectively substituted methyl lactosaminide derivative (**6**) in 83% yield. β -Configuration of the inter-residual linkage was confirmed by the value of coupling constant $J_{1,2}$ of 7.8 Hz. Glycosylation of **4** by 1,2,3,4,6-penta-O-acetyl- β -D-galactopyranose catalyzed by trimethylsilyl triflate (Ogawa *et al.*, 1981; Nifant'ev *et al.*, 1996) and by ethyl 2,3,4,6-tetra-O-acetyl-1-thio- β -D-galactopyranoside under promotion with nitrosyl tetrafluoroborate (Pozsgay and Jennings, 1987) were less effective (yield of **6** was 40%) and accompanied by O-6 \rightarrow O-4 migration of benzoate in **4** and formation of (1 \rightarrow 6)-linked allo-lactosaminide product (Nivant'ev *et al.*, 1988).

Hydrazinolysis of disaccharide **6**, followed by complete acetylation and debenzoylation gave lactosaminide derivative **8** with free OH-group at C-3. Glycosylation of **8** by fucosyl bromide **9** under halide-ion catalysis and subsequent removal of protecting groups gave target trisaccharide **1**.

Crystallography

Lattice constants were determined by a least-squares fit to the setting angles at high 2θ values measured with a Philips PW1100 diffractometer on an x-ray generator, Ni-filtered $Cu\alpha$ radiation, wavelength $\lambda = 1.541 \text{ \AA}$. The intensities of 4144 independent reflections were measured inside the sphere limited by $2\theta < 125^\circ$ of which 3139 such as $I > 3\sigma(I)$ were considered as observed. Lorentz and polarization corrections were applied, but no correction was made for absorption. The atomic scattering factors used were taken from the International Tables for X-Ray Crystallography (1974). The structure was solved by direct methods, allowing the location of all, C, O, and N atoms. The last refinement cycles were performed using an anisotropic thermal temperature factor for the nonhydrogen atoms. All the hydrogen atoms of the two trisaccharide molecules were located by successive difference Fourier maps and isotropic refinement. Half of the hydrogen atoms of the water molecules were

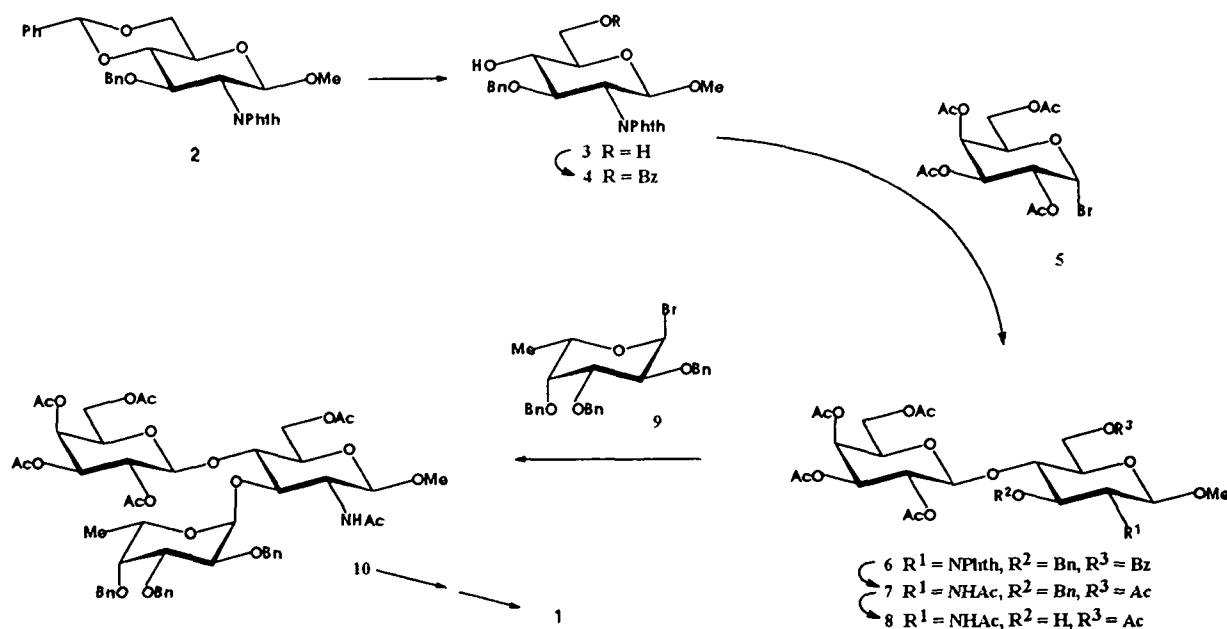


Fig. 7. Synthetic route for Le^x trisaccharide methyl glycoside.

located using the same procedure: consideration of these atomic positions in relation to possible hydrogen bonding schemes were sufficient to define the locations of the remaining hydrogen atoms without ambiguity. Introduction of these geometrically defined atoms at the final stage of the refinement did not increase the magnitude of the reliability index. The final *R* value was 0.051 and *R_w* = 0.054. A final electron density map showed no significant residual density. Averaged standard deviations are 0.01 Å for bond lengths, 0.7° for bond angles, and 1.5° for torsion angles.

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Abbreviations

Le^x, Lewis x trisaccharide; Gal, galactose; GlcNAc, N-acetylglucosamine; Fuc, fucose.

References

- Alais, J. and David, S. (1990) Preparation of disaccharides having a β-D-mannopyranosyl group from N-phthaloyllactosamine derivatives by double or triple S_N2 substitution. *Carbohydr. Res.*, **201**, 69–77.
- Aspinall, G.O., Monteiro, M.A., Pang, H., Walsh, E.J. and Moran, A.P. (1994) O antigen chains in the lipopolysaccharide of *H.pylori*. *Carbohydr. Lett.*, **1**, 151–156.
- Bird, J.M. and Kimber, S.J. (1984) Oligosaccharides containing fucose linked α(1–3) and α(1–4) to N-acetylglucosamine cause decompaction of mouse morulae. *Dev. Biol.*, **104**, 449–460.
- Biswas, M. and Rao, V.S.R. (1980) Conformational studies on the ABH and Lewis blood group oligosaccharides. *Biopolymers*, **19**, 1555–1565.
- Clausen, H. and Hakomori, S.-I. (1989) ABH and related histo-blood group antigens: immunochemical differences in carrier isotypes and their distribution. *Vox Sang.*, **56**, 1–20.
- Dean, B., Oguchi, H., Cai, S., Otsuji, E., Tasgiri, K., Hakomori, S.-i. and Toyokuni, T. (1993) Synthesis of multivalent β-lactosyl clusters as potential tumor metastasis inhibitors. *Carbohydr. Res.*, **245**, 175–192.
- Eggs, I., Fenderson, B., Toyokuni, T., Dean, B., Stroud, M. and Hakomori, S. (1989) Specific interaction between Lex and Le^x determinants. A possible basis for cell recognition in preimplantation embryos and in embryonal carcinoma cells. *J. Biol. Chem.*, **264**, 9476–9484.
- Feizi, T. (1985) Demonstration by monoclonal antibodies that carbohydrate structures of glycoproteins and glycolipids are onco-developmental antigens. *Nature*, **314**, 53–57.
- Fenderson, B., Zehavi, U. and Hakomori, S. (1984) A multivalent lacto-N-fuco pentaose III-lyssyllysine conjugate decompacts preimplantation mouse embryos, while the free oligosaccharide is ineffective. *J. Exp. Med.*, **160**, 1591–1596.
- Hakomori, S. (1989) Aberrant glycosylation in tumors and tumor-associated carbohydrate antigens. *Adv. Cancer Res.*, **52**, 257–331.
- Hakomori, S.-I., Nudelman, E., Levery, S., Solter, D. and Knowles, B.B. (1981) The hapten structure of a developmentally regulated glycolipid antigen (SSEA-1) isolated from human erythrocytes and adenocarcinoma. *Biochem. Biophys. Res. Commun.*, **100**, 1578–1586.
- Henry, S., Oriol, R. and Samuelsson, B. (1995) Lewis-histo-blood group system and associated secretory phenotypes. *Vox Sang.*, **69**, 166–182.
- Ichikawa, Y., Lin, Y.-C., Dumas, D.P., Shen, G.-J., Garcia-Junceda, E., Williams, M.A., Bayer, R., Ketcham, C., Walker, L.E., Paulson, J.C. and Wong, C.-H. (1992) Chemical-enzymatic synthesis and conformational analysis of sialyl Lewis x and derivatives. *J. Am. Chem. Soc.*, **114**, 9283–9298.
- Imberty, A., Mikros, E., Koca, J., Mollicone, R., Oriol, R. and Pérez, S. (1995) Computer simulation of histo-blood group oligosaccharides: energy maps of all constituting disaccharides and potential energy surfaces of 14 ABH and Lewis carbohydrate antigens. *Glycoconj. J.*, **12**, 331–349.
- International Tables for X-Ray Crystallography (1974) Vol. 4. Kyoich Press, Birmingham.
- Jeffrey, G.A. and Saenger, W. (1991) *Hydrogen Bonding in Biological Structures*. Springer-Verlag, Berlin.
- Kojima, N., Fenderson, B.A., Stroud, M.R., Goldberg, R.I., Habermans, R., Toyokuni, T. and Hakomori, S. (1994) Further studies on cell adhesion based on Le^x-Le^x interaction, with new approaches: embryoglycan aggregation of F9 teratocarcinoma cells, and adhesion of various tumour cells base on Le^x expression. *Glycoconj. J.*, **11**, 238–248.
- Lasky, L.A. (1992) Selectins: interpreters of cell-specific carbohydrate information during inflammation. *Science*, **258**, 964–969.
- Lemieux, R.U., Bock, K., Delbaere, L.T.J., Koto, S. and Rao, V.S. (1980) The conformations of oligosaccharides related to the ABH and Lewis human blood group determinants. *Can. J. Chem.*, **58**, 631–653.
- Miller, K.E., Mukhopadhyay, C., Cagas, P. and Bush, C.A. (1992) Solution structure of the Lewis x oligosaccharide determined by NMR spectroscopy and molecular dynamics simulations. *Biochemistry*, **31**, 6703–6709.
- Nivant'ev, N.E., Backinowsky, L.V. and Kochetkov, N.K. (1988) Synthesis of derivatives of 2-amino-2-deoxy-4-O-(α and β-D-galactopyranosyl)-D-glucose. *Carbohydr. Res.*, **174**, 61–72.
- Nivant'ev, N.E., Khatuntseva, E.A., Shashkov, A.S. and Bock, K. (1995) *Carbohydr. Lett.*, **2**, 399–406.
- Ogawa, T., Beppu, K. and Nakabayashi, S. (1981) Trimethylsilyl trifluoromethanesulfonate as an effective catalyst of glycoside synthesis. *Carbohydr. Res.*, **93**, c6–c9.
- Oriol, R. (1995) ABO, Hh, Lewis, and secretion. Serology, genetics, and tissue distribution. In Cartron, J.P. and Rouger, P. (eds.), *Blood Cell Biochemistry*, Vol. 6. Plenum, New York, pp. 161–171.
- Pérez, S., Imberty, A. and Carver, J.P. (1994) Molecular modeling: an essential component in the structure determination of oligosaccharides and polysaccharides. *Adv. Comput. Biol.*, **1**, 146–202.
- Pozsgay, V. and Jennings, H.J. (1987) A new method for the synthesis of O-glycosides from S-glycosides. *J. Org. Chem.*, **52**, 4635–4637.
- Schwartz, D.E., Lee, H.H., Carver, J.P. and Krepinsky, J.J. (1985) Synthesis of model oligosaccharides of biological significance. 4. Synthesis of a fucosylated N,N'-diacetylchitobioside and related oligosaccharides. *Can. J. Chem.*, **63**, 1073–1079.
- Solter, D. and Knowles, B.B. (1978) Monoclonal antibodies defining a stage specific mouse embryonic antigen. *Proc. Natl. Acad. Sci. USA*, **75**, 5565–5569.
- Thogersen, H., Lemieux, R.U., Bock, K. and Meyer, B. (1982) Further justification for the exo-anomeric effect. Conformational analysis based on nuclear magnetic resonance spectroscopy of oligosaccharides. *Can. J. Chem.*, **60**, 44–57.
- Van Dam, G.J., Bergwerff, A.A., Thomas-Oates, J.E., Rotmans, J.P., Kamerling, J.P., Vliegthart, J.F.G. and Deedler, A.M. (1994) The immunologically reactive O-linked polysaccharide chains derived from circulating cathodic antigen isolated from the human blood fluke *Schistosoma mansoni* have Lewis^x as repeating unit. *Eur. J. Biochem.*, **225**, 467–482.
- Varki, A. (1994) Selectin ligands. *Proc. Natl. Acad. Sci. USA*, **91**, 7390–7397.
- Velupillai, P. and Harn, D.A. (1994) Oligosaccharide specific induction of interleukin 10 production by B220+ cells from schistosoma-infected mice: a mechanism for regulation of CD4+ T-cell subsets. *Proc. Natl. Acad. Sci. USA*, **91**, 18–22.
- Yvelin, F., Zhang, Y.-M., Mallet, J.M., Robert, F., Jeannin, Y. and Sinay, P. *Carbohydr. Lett.*, **1**, 475–482.

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Note added in proof

While our manuscript was processed, a paper describing an independent X-ray determination of the crystal structure of the Lewis x trisaccharide was published (Yvelin *et al.*, 1996).