

Review Article

Current Methods in the Molecular Typing of *Mycobacterium tuberculosis* and Other Mycobacteria

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In the epidemiology of tuberculosis (TB) and nontuberculous mycobacterial (NTM) diseases, as in all infectious diseases, the key issue is to define the source of infection and to disclose its routes of transmission and dissemination in the environment. For this to be accomplished, the ability of discerning and tracking individual *Mycobacterium* strains is of critical importance. Molecular typing methods have greatly improved our understanding of the biology of mycobacteria and provide powerful tools to combat the diseases caused by these pathogens. The utility of various typing methods depends on the *Mycobacterium* species under investigation as well as on the research question. For tuberculosis, different methods have different roles in phylogenetic analyses and person-to-person transmission studies. In NTM diseases, most investigations involve the search for environmental sources or phylogenetic relationships. Here, too, the type of setting determines which methodology is most suitable. Within this review, we summarize currently available molecular methods for strain typing of *M. tuberculosis* and some NTM species, most commonly associated with human disease. For the various methods, technical practicalities as well as discriminatory power and accomplishments are reviewed.

1. Introduction

The genus *Mycobacterium* contains more than 140 species [1], which are separated in three major groups, that is, *M. tuberculosis* complex (MTBC), *M. leprae*, and mycobacteria other than MTBC and *M. leprae*, collectively referred to as nontuberculous mycobacteria (NTM). *Mycobacterium tuberculosis*, the most prominent member of the MTBC, is an obligate human pathogen and the causative agent of tuberculosis (TB), which remains one of the leading global public health problems. According to the World Health Organization (WHO), over 9 million new cases of TB occur each year, resulting in

approximately 2 million deaths worldwide [2]. Conversely to *M. tuberculosis*, for which no environmental reservoir exists, NTM are ubiquitous organisms and are readily isolated from environmental sources, including soil and both natural and artificial water systems [3]. Despite reportedly low virulence of NTMs for immunocompetent human hosts, an increase in their isolation frequencies has been seen in the last decade, particularly in countries where TB incidence is on decrease [4, 5].

In the epidemiology of TB and other mycobacterioses, as in all infectious diseases, the key issue is to define the source of infection and to disclose its routes of transmission and

dissemination in the environment. For this to be accomplished, the ability of discerning and tracking individual *Mycobacterium* strains is of critical importance. The earliest discriminatory methods relied upon phenotypic characteristics, such as colony morphology, susceptibility to antituberculosis drugs, or mycobacterial phage typing. The usefulness of these methods is seriously limited by the phenotypic variability of mycobacteria. For instance, drug susceptibility patterns may change for the same isolate as it acquires resistance to specific drugs in the course of treatment. Otherwise, the limiting factor of phage typing is the low number of recognized mycobacteriophages [6–8]. Discrimination between the strains based on their biochemical and serological characteristics is even more unsuccessful [9, 10]. A turning point in this quest was the development of molecular biology tools in the mid 1980s. The DNA-based techniques have revolutionized the epidemiology of TB and other mycobacterial diseases, since they allow querying the whole genome that is unique and relatively stable for each strain.

In terms of genetic heterogeneity, the MTBC and NTM are vastly different and this has strong implications for the choice of typing methods and the achievable levels of discrimination.

Mycobacterium tuberculosis complex constitutes a remarkably genetically homogeneous group. This is perhaps best illustrated by the fact that, within the ribosomal DNA (rDNA) operon, not only genes encoding various types of rRNA but also regions between those genes, such as internal transcribed spacer (ITS) regions, which in many bacteria and fungi are highly polymorphic and thus are useful for identification and typing to subspecies or strain level, show complete conservation among members of the *M. tuberculosis* complex [11]. Likewise, many structural genes of *M. tuberculosis* complex show important sequence conservation, with an estimated rate of synonymous mutations of 0.01–0.03% [12–14]. Additionally, the lack of significant evidence for horizontal gene transfer between *M. tuberculosis* genomes speaks in favor of clonal evolution in *M. tuberculosis* complex [12–15]. This in turn renders strain-level discrimination of *M. tuberculosis* by means of molecular typing approaches challenging. However, recent studies have shown that the genetic diversity of the MTBC is much higher than previously assumed and that this genomic variance is attributed to the single nucleotide polymorphisms (SNPs), with potential impact on pathobiological phenotype [16, 17].

As for the NTM, the evolutionary time of divergence is believed to be much larger than that for the *M. tuberculosis* complex and this implies that less than a whole genome can be queried to obtain a highly discriminatory typing result [18].

Within this review, we summarize currently available molecular methods for strain typing of mycobacteria. For the various techniques, technical practicalities as well as discriminatory power and accomplishments are reviewed.

2. Restriction Site Analysis of Genomic DNA

First attempts of molecular typing of *M. tuberculosis* were based on restriction enzyme analysis of bacterial DNA (REA). In principle, chromosomal DNA of the analyzed strains is

digested using various restriction enzymes and the resulting fragments are separated and visualized by gel electrophoresis. The obtained pattern of DNA fragments (*genetic fingerprint*) is characteristic for each strain. However, using the original procedure, the sensitivity of the method is rather limited due to technical difficulties in providing a high-resolution electrophoretic separation of fragments within a broad range of sizes. Also, when more restriction enzymes are used, the high number of DNA fragments makes a reliable analysis impossible [19]. Therefore, new methods have been proposed for a more accurate separation of DNA molecules, such as REA-PFGE (pulsed-field gel electrophoresis), which guarantees high resolution of the restriction patterns. More commonly used methods, which are modifications of the traditional REA, employ DNA hybridization assay in the process of specific pattern detection. Specifically, after electrophoresis, the separated DNA fragments are denatured in situ and transferred onto a membrane, which is then incubated with a radiolabeled probe (Southern blot) for the target sequence. Hybridization signals are visualized on autoradiography. The Southern blot technique was first applied to the analysis of restriction fragment length polymorphism (RFLP), which is hence a combination of REA and hybridization technology. Early studies utilizing RFLP methodology indicated that strains of *M. tuberculosis* display a considerably low degree of genetic diversity. However, such interpretation of the results was actually misleading since probes used in those studies targeted highly conserved regions and were of low specificity [20–22]. The resolution of the RFLP method increased substantially when insertion sequences (IS) were identified and used in probe construction [23, 24]. Different repetitive sequences found in the genomes of *M. tuberculosis* and NTM are an important source of genetic polymorphism and provide reliable markers allowing the determination of genetic relationships at both species and strain levels.

3. Pulsed-Field Gel Electrophoresis (PFGE)

A method related to REA, pulsed-field gel electrophoresis (PFGE), enables the separation of large DNA fragments, up to 10 Mb. In contrast, by using conventional electrophoresis, the threshold length exists at about 50 kb. The principle of the PFGE system is based on the application of an electric field that periodically changes its orientation across a gel matrix. This is achieved by varying the duration of the electrical pulse and shifting the direction of the current frequently. In general, the PFGE procedure involves digestion of chromosomal DNA with rare cutting restriction endonucleases, followed by agarose gel electrophoresis and analysis of the resolved electrophoretic patterns. A crucial step is the preparation of genomic DNA. Since large DNA molecules are prone to shearing and crushing, DNA is isolated in a gentle manner by first embedding a suspension of the organism in agarose plugs, lysing the cells in situ, and digesting the chromosomal DNA with restriction enzymes. The plugs are then loaded into the gel wells and sealed into place with agarose. After the electrophoresis, the resulting banding patterns are compared, using a predefined set of criteria for strain relatedness [25]. Although the PFGE patterns are well reproducible and the

overall discriminatory power of the method is high, a number of limitations are apparent. Firstly, the method is technically demanding and cost intensive. Secondly, it requires intact DNA for restriction enzyme treatment. Thirdly, the PFGE method has a long turn-over time, as the whole protocol usually takes not less than a week. Finally, no standardized procedure for performing PFGE has yet been recommended. Despite these disadvantages, the PFGE typing was successfully used to differentiate between strains of *M. tuberculosis* [26], *M. bovis* [27], and *M. bovis* BCG [28]. However, the PFGE typing is rarely used in *M. tuberculosis* complex due to technical, time, and cost considerations, as mentioned above. Moreover, PFGE analysis does not always generate sufficient discrimination between the strains [26, 29, 30].

Quite oppositely, PFGE often remains the most powerful typing system for nontuberculous mycobacteria. The method has been applied, with different degrees of success, to both slow-growing NTM species, including *M. kansasii* [31, 32], *M. avium*-*M. intracellulare* complex [33], *M. goodii* [34], and *M. haemophilum* [35], and rapidly growing mycobacteria, such as *M. fortuitum* [36], *M. chelonae* [37], and *M. abscessus* [37, 38].

4. IS6110-RFLP Analysis

Study of the complete *M. tuberculosis* H₃₇Rv reference strain genome sequence revealed a relatively large amount of repetitive DNA elements [39]. Those elements vary in length, structure, and localization. Two main groups can be distinguished, that is, tandem repeats (TR) and interspersed repeats (IR). The first are short monomeric sequences (up to 100 bp) organized as head-to-tail arrays, whereas the latter are scattered as individual copies throughout the entire genome. An important class of IR sequences is insertion sequences (IS), which are mobile genetic elements.

The best known and investigated insertion sequence is IS6110 first recognized by Thierry et al. in the early 1990s [40–43]. The IS6110 sequence belongs to the IS3 family and is a 1,355 bp long with unique 28 bp terminal inverted repeats (TIR). The region between those repeats includes two overlapping reading frames, *orfA* and *orfB*, encoding for a transposase, an enzyme responsible for transposition of the insertion sequence. The IS6110 is found within the *M. tuberculosis* complex, and, in most members of the complex, the sequence is present at multiple copies, although *M. bovis* normally contains only one copy. In general, the copy number of IS6110 ranges from 0 to 25 and depends on the frequency of transposition, which is largely conditioned by the nature of the genomic region at which transposition occurs [44]. Although the IS6110 can be integrated into any place on the chromosome, there are regions with higher frequency of transposition. The so-called integration hot spots are usually located within coding regions of *M. tuberculosis* DNA [44, 45].

Differences in the copy number and locations within the genome, responsible for the high degree of IS6110 polymorphism, have predisposed this sequence to be used as a specific molecular marker for genotyping of *M. tuberculosis* strains [46, 47].

IS6110-based typing is still among the most widely applied genotyping methods in molecular epidemiological studies of *M. tuberculosis*. The method includes digestion of genomic DNA with *PvuII* restriction enzyme that cleaves the IS6110 sequence only once, generating DNA fragments that are separated through gel electrophoresis, then transferred onto a membrane, and hybridized with a peroxidase-labelled probe complementary to part of the 3'-end of the IS6110 sequence. As a result, every visualized fragment represents a single copy of IS6110 surrounded by different in length flanking DNA (Figure 1(1)) [48]. Since the IS6110-RFLP methodology has been standardized and published, recommendations have been adopted by most of the research groups; the fingerprints generated in different laboratories can be compared and catalogued [49, 50]. The method is highly discriminatory and reproducible. An important characteristic of IS6110-RFLP typing is the stability of its profiles over time, allowing distinguishing epidemiologically related from unrelated isolates. Specifically, the half-time of change in IS6110-RFLP pattern was estimated to be ca. 3-4 years [51, 52]. Stability of the IS6110-RFLP patterns depends on the transposition process frequency; the more common is the transposition, the less stable is the number of the IS6110 element in the genome. However, several important limitations exist for the IS6110-RFLP method. Firstly, there is a need for large amounts of high quality DNA (2 µg) for restriction enzyme digestion and therefore requires time consuming (up to several weeks) bacterial culturing. Secondly, the method is technically demanding and requires sophisticated and expensive computer software as well as experienced personnel of high technical expertise. Finally, the discriminatory power of IS6110-RFLP typing is insufficient for those strains whose copy number of IS6110 is 6 or less (the so-called low-copy strains are seen among *M. bovis* isolates from cattle or *M. tuberculosis* isolates from Asia) [53–56]. In addition, some NTM have multiple copies of sequences that are homologous to IS6110 and may thus hybridize with the IS6110 probe [57].

Despite these limitations, the IS6110-RFLP method remains one of the most commonly used approaches for *M. tuberculosis* typing and was long considered the gold standard technique in the molecular epidemiological investigations of TB.

5. IS6110-Based PCR Fingerprinting

The IS6110 is a target sequence in many methods currently used for molecular typing of *M. tuberculosis*. Among these, the most important is the mixed-linker PCR (ML-PCR) [58], ligation-mediated PCR (LM-PCR) [60], and fast ligation-mediated PCR (FliP) [59]. All these methods follow a similar four-step algorithm including genomic DNA fragmentation using restriction enzymes that generate protruding ends (i), ligation of those fragments with synthetic oligonucleotide linkers or adaptors (ii), amplification of the ligation products with one primer specific for the IS6110 and a second primer complementary to a linker (iii), and analysis of the amplicons (iv).

In ML-PCR and FliP methods the products are 3' fragments of IS6110, whereas in LM-PCR the IS6110-flanking

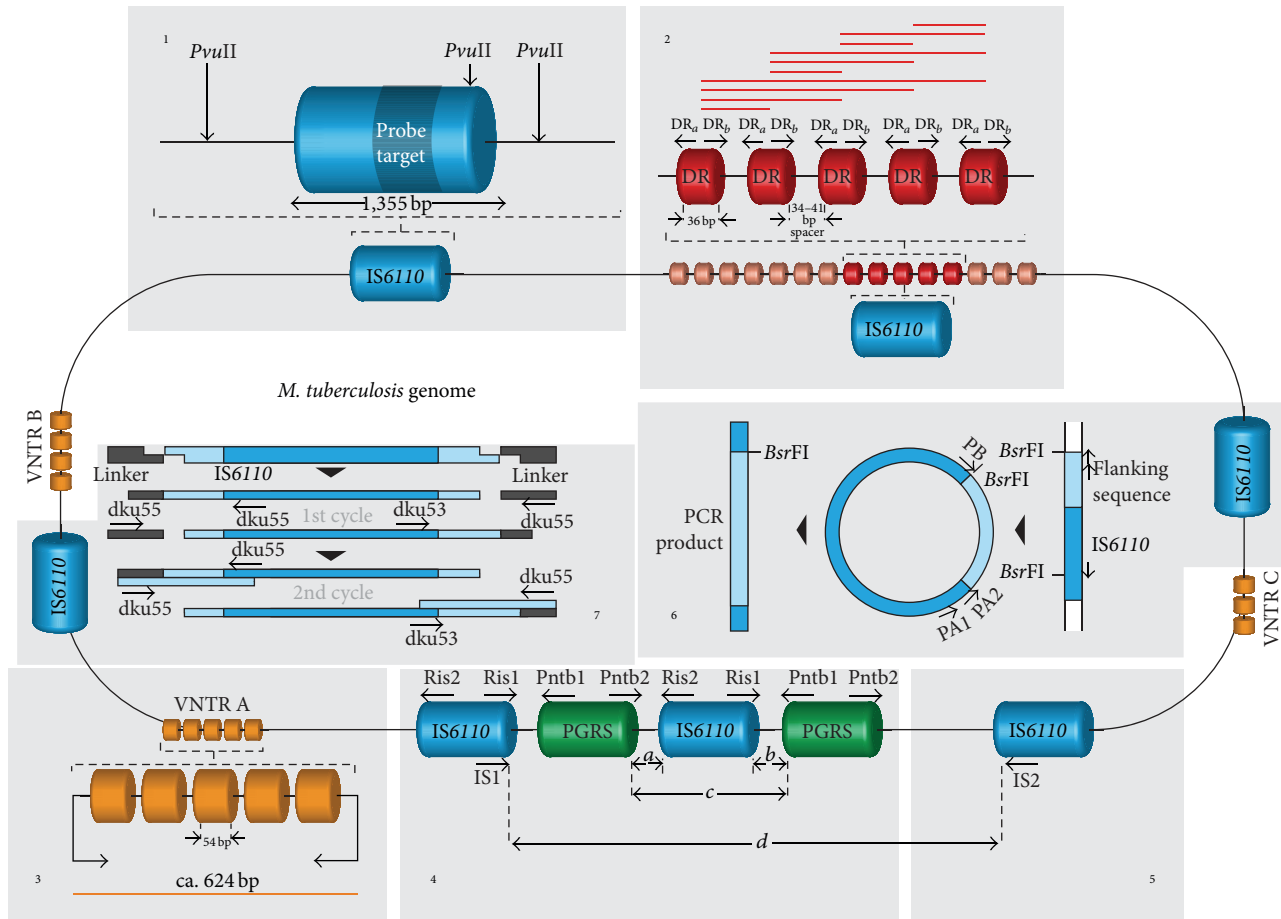


FIGURE 1: Schematic representation of the chromosome of a hypothetical *Mycobacterium tuberculosis* complex isolate with marked repetitive elements as targets for different typing methods. The principle of those methods is pictorially outlined. (1) In IS6110-RFLP typing, mycobacterial DNA is cleaved with the restriction endonuclease *PvuII*, and the resulting fragments are separated electrophoretically on an agarose gel, transferred onto a nylon membrane by Southern blotting, and hybridized to a probe complementary to the 3'-end of the IS6110 (probe target) yielding a characteristic banding pattern, in which every band represents a single IS6110 element. (2) Spoligotyping relies upon PCR amplification of a single direct repeat (DR) locus which harbours 36 bp direct repeats interspersed with unique 34–41 bp spacer sequences. The PCR products (red horizontal lines) are hybridized to a membrane containing 43 oligonucleotides corresponding to the spacers from *M. tuberculosis* H₃₇Rv and *Mycobacterium bovis* BCG. The presence or absence of each of those 43 spacers in the DR region of the analysed isolate will be represented as the pattern of positive or negative hybridization signals. (3) The variable numbers of tandem repeat loci (VNTR) or mycobacterial interspersed repetitive units (MIRU) are PCR-amplified and the obtained products (yellow horizontal line) are sized on agarose gels to deduce the number of repeats in each individual locus. (4, 5) Two PCR-based typing methods, that is, double-repetitive-element PCR (DRE-PCR) and amplityping, are designed to amplify DNA between clusters of IS6110 and polymorphic GC-rich sequences (PGRS) or between clusters of IS6110 elements, respectively. Different distances between the repetitive elements and their different copy numbers result in variability of banding patterns, composed of DNA fragments amplified (*a–d*) and produced for individual isolates. (6) A heminested inverse PCR (HIP) depends on the amplification of the 5'-end of the IS6110 sequence along with its upstream flanking sequence, bordered by the closest *BsrFI* site. The size and number of PCR amplicons generated depend on the number of copies of IS6110. (7) Ligation-mediated PCR (LM-PCR) procedure allows, by introducing specifically designed linkers, amplifying the flanking sequences on both sides of the IS6110 element. Names and positions of the PCR primers were excerpted from the original papers. For more details, read the text.

sequence on the 5' side is amplified. (In the original LM-PCR procedure flanking sequences on both sides of the IS6110 are amplified by using primers that are complementary to both termini of the IS6110 sequence and one of the primers is also complementary to the shorter strand of the linker (Figure 1(7)) [68].)

All those methods are highly reliable and exhibit significant discriminatory potential, albeit slightly lower than that of IS6110-RFLP analysis [59, 69–71].

This potential can even be increased when using heminested inverse PCR (HIP). This method relies on amplification of a 5' part of the IS6110 and its flanking sequence up to the proximal *BsrFI* restriction site [61]. Briefly, chromosomal DNA is cut with the restriction endonuclease *BsrFI* and the restriction fragments are then self-ligated at low DNA concentration. The resulting circular molecules comprised of the 5'-end of the IS6110 and its flanking region are subjected to PCR using primers that anneal to the IS at sites between its

5'-end and the closest *Bsr*FI site. Products of amplification vary in length depending on the length of the flanking sequence and are visualized by agarose gel electrophoresis (Figure 1(6)). The HIP method is highly reproducible and its discriminatory power is equivalent to that of standard IS6110-RFLP analysis but is much simpler and faster in performance [61, 72].

Another PCR-based IS6110 typing method is amplotyping. Here, outward-oriented primers hybridize with the ends of the IS6110 sequence, so that DNA separating adjacent copies of this element on the genome is amplified (Figure 1(5)) [73–75]. A variant of the aforesaid method uses only a single primer, targeted to the terminal inverted repeat sequences of the IS6110 [76, 77]. Differences in the length of the amplicons reflect the distance between the IS elements and are analysed by standard gel electrophoresis. An important limitation of this typing approach is that production of the PCR amplicons is dependent on the priming sites within the ISs being close enough for efficient PCR.

All PCR-based typing assays, targeting the IS6110, are easy-to-perform, time saving and require relatively small amounts of genomic DNA, which makes them applicable to nonviable organisms or directly to clinical specimens, without culture. The resolution of the IS6110-PCR analysis may not be sufficient enough to differentiate among the isolates; however when supplemented with an additional restriction digestion step, the differentiation capacity of the method is comparable to IS6110-RFLP [78]. Apart from some technical difficulties, such as nonspecific priming, still the major drawback of the methods discussed is the lack of discriminatory power for typing of the isolates with low copy numbers of IS6110 [74–77].

6. IS-Based Typing of NTM

Since a number of different ISs have been described in various NTM species, these genetic mobile elements have widely been applied for strain-level typing.

Two closely related insertion sequences IS1245 and IS1311 were found very useful to differentiate within the *M. avium* complex.

Based on the IS1245 or IS1311-RFLP patterns, various clades of *M. avium* and its subspecies could be distinguished, related to infections in birds, pigs, and humans [79–81].

Other insertion elements that are known and have been used for identification rather than typing purposes of *M. avium* complex bacilli include IS900 present in *M. avium* subsp. *paratuberculosis* [82], IS901 present in *M. avium* subsp. *avium* [83], IS902 present in *M. avium* subsp. *silvaticum* [84], and IS666, IS1110, and IS1626, whose distribution among *M. avium* isolates has been ill-studied [85–87]. The insertion sequences: IS900, IS901, IS902, and IS1245, can be used for the identification of various *M. avium* subspecies as well as for differentiation within those subspecies.

One of the most important observations made upon IS1245 RFLP typing was that birds are infected by a genetically highly conserved type of *M. avium* strains invariably revealing the same three-band pattern, while the banding patterns of *M. avium* isolates of porcine and human origin revealed

highly variable and multibanded patterns. Consequently, it was proposed to reserve the naming *M. avium-avium* for the bird-type isolates and to introduce the designation *M. avium hominissuis* for typical isolates from humans and pigs [81].

Beyond the *M. avium* complex, RFLP typing has been pursued only sporadically. Yet potentially useful insertion sequences have been described in a variety of species. These include IS1407 in *M. celatum* [88], IS1395 in *M. xenopi* [89], IS1511/1512 in *M. gordonae* [90], IS2404 in *M. ulcerans*, IS2606 in *M. ulcerans* and *M. lentiflavum* [91], IS1652 in *M. kansasii* [92], and IS6120 in *M. smegmatis* [93].

7. Spoligotyping

Spoligotyping is currently one of the most frequently used PCR-based approaches for studying the phylogeography of *M. tuberculosis* complex. The spoligotyping method is based on the polymorphism at one particular genomic region, the so-called direct repeat (DR) locus, initially identified by Hermans et al. in the vaccine strain *M. bovis* BCG P3 [94]. The DR locus comprises a series of well-conserved 36 bp direct repeats (DRs) interspersed with unique, nonrepetitive spacer sequences of 34–41 bp. The DR and the adjacent variable sequence form a direct variant repeat (DVR). The DR locus belongs to the clustered regularly interspaced short palindromic repeats (CRISPRs) family of repetitive DNA. It is postulated that these elements are reminiscent of centromere-like structures with a possible role in replication partitioning [95]. In the spoligotyping method, the entire DR locus is amplified by PCR, using two inversely oriented primers complementary to the sequence of short DRs. The PCR products, of different sizes, are hybridized to a membrane with 43 covalently bound synthetic oligonucleotides representing the polymorphic spacers identified in *M. tuberculosis* H₃₇Rv (spacers 1–19, 22–32, and 37–43) and *M. bovis* BCG (spacers 20–21 and 33–36). The hybridization signals are detected by chemiluminescence through biotin labeling of the PCR products (one of the primers is biotinylated) and a streptavidin-peroxidase conjugate system and then visualized by autoradiography (Figure 1(2)). Individual strains are differentiated by the number of the spacers that are missing from the complete 43-spacer set [62]. The lack of spacers is most probably the result of deletions mediated by various genetic mechanisms, such as homologous recombination or transposition (the DR region is a hot spot for IS6110 integration) [96, 97].

Spoligotyping is a relatively simple, cost-effective, and high-throughput method, whose results are accurate and reproducible and are obtained in up to 2 days. The reliability of the results is linked to a high stability of the DR locus. The molecular clock of this genetic marker is believed to be very slow, since multiple *M. tuberculosis* isolates from the same patients corresponding to relapses or infections at different sites, even over time spans of several years, showed identical spoligotypes [98]. An important advantage of spoligotyping is its genuine sensitivity estimated at 10 fg of chromosomal DNA, equivalent to DNA from 2–3 bacterial cells [99], allowing the method to be applied directly in clinical samples, without the need for prior culture. Moreover, spoligotyping

has proven to be useful for typing on nonviable cultures, Ziehl-Neelsen smear slides, or paraffin-embedded tissue sections [100, 101].

Given the binary (present/absent) format of the data, the spoligotyping results can easily be interpreted, computerized, and compared between different laboratories [102].

In 2006, an international spoligotyping database (SpolDB4) was released. The database, which is accessible online (<http://www.pasteur-guadeloupe.fr:8081/SITVIT-Demo/>), describes 1,939 STs (shared types, i.e., spoligotype patterns shared by two or more isolates) and 3,370 orphan types (i.e., spoligotype patterns reported for only single isolates) from a total of 39,295 *M. tuberculosis* complex isolates, from 122 countries, classified temporarily into 62 clades/lineages [103]. In a recently erected publicly available multimer database named SITVIT, a total of 7105 spoligotype patterns (corresponding to 58,180 clinical isolates)—grouped into 2740 shared types containing 53,816 clinical isolates and 4364 orphan patterns—were incorporated (http://www.pasteur-guadeloupe.fr:8081/SITVIT_ONLINE/) [104].

Spoligotyping allows identification of *M. tuberculosis* complex isolates at the (sub)species level. For instance, the *M. tuberculosis* spoligotypes are characterized by the absence of spacers 33–36, whereas *M. bovis* spoligotypes usually lack spacers 39–43, and *M. bovis* BCG spoligotypes lack spacers 3, 9, and 16 [105]. Furthermore, spoligotyping allows identification of genotypes of significant, clinical, and epidemiological relevance. A clear example is the “Beijing” genotype, commonly encountered in the Beijing area, other regions of Asia, the former Soviet Union, and other geographical areas. Most of the “Beijing” genotype strains react only with the last 9 spacers (35–43) in the panel of 43 [105].

Spoligotyping has a lower level of discrimination than the IS6110 RFLP typing, as evidenced in several studies [70, 106–110]. The introduction of 51 novel spacer sequences, mostly [45] from the DR region from the *M. canetti* genome only, slightly improved the resolution of the method [97]. The 68-spacer format, with 25 out of 51 new spacers, was shown to improve the discrimination for the *M. africanum* subspecies and for the East African-Indian (EAI) clade of *M. tuberculosis* [111, 112].

The reason for the limited discriminatory capacity of the spoligotyping method is due to the fact that it targets only a single genetic locus, covering less than 0.1% of the *M. tuberculosis* complex genome. Nevertheless, spoligotyping can be effectively used for the differentiation of *M. tuberculosis* strains with low IS6110 copy numbers (≤ 5 bands in RFLP patterns) [54, 113]. As *M. tuberculosis* isolates with different spoligotypes invariably have distinct IS6110 RFLP profiles, a genotyping strategy has been proposed, in which spoligotyping could be performed as a first-line, screening test, to be followed by another typing method of greater discriminatory power [108]. Spoligotyping, when used alone, is not sufficient for epidemiological linking studies. Furthermore, contaminated isolates and multistrain infections may not be detected by performing spoligotyping directly on clinical samples. However, this remark can be extended to any PCR-based technology, which is applied directly in clinical material.

Spoligotyping, when applied for nontuberculous mycobacteria, produced no signal, indicating the specificity of the method solely for *M. tuberculosis* complex [108].

Since the description of the spoligotyping method in its original form, another two formulations have been proposed. The first benefits from the Luminex technology, where the synthetic spacer oligonucleotide probes are immobilized on microspheres by means of covalent coupling, and detection is achieved via fluorochromes attached to the beads and hybridized PCR product. The Luminex platform, by eliminating the membrane step with the subjective manual data interpretation, provides, greater robustness and reproducibility. It is also well suited for high-throughput analysis, since it allows 96 isolates to be assayed simultaneously, as opposed to 45 isolates in a standard spoligotyping approach [112, 114].

A more recent alternative to a conventional spoligotyping scheme is a new multiplexed primer extension-based spoligotyping assay using automated matrix-assisted laser desorption/ionization-time of flight mass spectrometry (MALDI-TOF MS). Spoligotyping by MALDI-TOF MS improves the classical reverse line blot hybridization assay with respect to reproducibility, throughput, process flow, ease of use, and data analysis [115]. An important limitation of these innovative, technologically refined spoligotyping assays is that they require advanced and expensive equipment which many laboratories, especially those in resource-constrained settings, cannot afford.

8. Methods Based on Minisatellite Sequences

Mycobacterium tuberculosis was among the first bacterial species in which tandem repeat loci resembling minisatellite loci in eukaryotic genomes were identified. The mycobacterial tandemly repeated sequences were, per analogiam to those in humans and animals, called variable number of tandem repeat (VNTR) loci. However, as new VNTR-type loci have been discovered, they have been referred to under different names.

The first described VNTRs were major polymorphic tandem repeat (MPTR) and exact tandem repeat (ETR), found in 5 (A–E) and 6 (A–F) loci, respectively [63]. The MPTR consists of a 10 bp repeated sequence separated by unique 5 bp spacer sequences. These repetitive DNA elements were identified in *M. tuberculosis* complex as well as in other mycobacteria, including *M. goodii*, *M. kansasii*, or *M. szulgai* [116]. The MPTR has been shown particularly useful in typing of *M. kansasii*. RFLP analysis with MPTR as a probe has revealed the existence of multiple subtypes within this species [92]. These subtypes have later been shown to have different degrees of pathogenicity in humans [117]. Interestingly, the MPTR sequences are part of the 3'-end of genes belonging to the PPE proteins (named after the conserved Pro-Pro-Glu (PPE) motifs near the N terminus of the molecule). The polymorphism of the PPE proteins in their C-terminal domains, linked to the presence of the MPTR motifs, is speculated to be the source of antigen variability in *M. tuberculosis* [118].

The ETR elements, found exclusively in *M. tuberculosis* complex strains, contain repeats ranging in size from 53 to

79 bp. Sequencing of the ETRs revealed that all of the ETR loci were variable. Contrastingly, among MPTR loci, only one (MPTR-A) showed some polymorphism, upon sequencing analysis [63]. Of eleven MPTR/ETR loci, only five (ETR-A-E) are routinely used for genotyping of *M. tuberculosis* strains. The principle of any typing system based on the polymorphism of VNTR loci is that each VNTR locus is PCR-amplified with specific primers complementary to the flanking regions, and the resulting PCR products are visualized by standard gel electrophoresis (Figure 1(3)). The number of tandem repeat units is determined by estimating the size of the amplicons, in relation to the known size of the repeat unit within the targeted VNTR locus. The results are expressed in a simple, digital format, in which each digit represents the number of copies at a particular locus [63]. Despite being fast, easy-to-perform, and highly reproducible, VNTR genotyping based on 5 ETR loci has a low discriminatory power, compared to IS6110-RFLP or spoligotyping [53, 70, 119, 120]. However, with the completion of the *M. tuberculosis* H₃₇Rv genome sequencing project, new VNTR-type loci have been identified. A specific class of these new VNTR elements is mycobacterial interspersed repetitive units (MIRUs). MIRUs were originally described by Supply et al. [121] as 46–101 bp tandem repeats scattered at 41 loci throughout the chromosome of *M. tuberculosis* H₃₇Rv. Based on the sequence analysis of each of those loci, 12 were found to display variations in tandem repeat copy numbers and were thus selected for genotyping of *M. tuberculosis* isolates [122, 123]. Among those 12 hypervariable loci, two (MIRU-4 and MIRU-31) are identical to formerly described ETR loci (i.e., ETR-D and ETR-E, resp.). MIRU-VNTR analysis, as every VNTR-based typing approach, involves PCR amplification of a specific MIRU locus, followed by determination of the sizes of the amplicons by gel electrophoresis or, after running multiplex PCRs, on an automated, fluorescence-based sequencer (Figure 1(3)). Since the length of the repeat units is known, the calculated sizes reflect the numbers of the amplified MIRU copies. The final result is a multidigit numerical code (the so-called MIRU-VNTR code), corresponding to the repeat number at each analyzed locus [64, 124]. This coding system allows the results to be readily compared across laboratories worldwide and facilitates the data to be deposited in the global databases via the Internet for large-scale epidemiological and population genetic studies [125, 126] (<http://www.miru-vntrplus.org/>). Recently, the biggest publicly available international database named SITVITWEB, which incorporates multimarker genotyping information (i.e., based on MIRU-VNTR typing and spoligotyping) on 62,582 *M. tuberculosis* complex clinical isolates from 153 countries of patient origin, has been released (http://www.pasteur-guadeloupe.fr:8081/SITVIT_ONLINE/) [104]. Furthermore, a 12-locus MIRU scheme, based on the minimum spanning tree (MST) method, has been proposed for classification of *M. tuberculosis* complex genotypic lineages [127].

The MIRU-VNTR method is a reliable and efficient typing system, whose discriminatory capacity approximates or even exceeds that of IS6110-RFLP profiling. In general, the discriminatory power of MIRU-VNTR analysis increases with the

number of loci evaluated. MIRU-VNTR typing based on 12 loci is slightly less discriminatory than IS6110-RFLP analysis for *M. tuberculosis* isolates with high copy number of IS6110 [64, 123, 128, 129] but at the same time more discriminatory than the IS6110-RFLP if low-copy-number IS6110 isolates are investigated [130, 131]. In principle, 12-locus MIRU-VNTR cannot be used as a sole typing method, as it may overestimate the number of true epidemiological links, especially in large, population-based studies [132, 133]. Consequently, it is suggested to use 12-locus MIRU-VNTR analysis in combination with other genotyping methods [128, 132]. An alternative way is to increase its resolution by expanding the investigation on other polymorphic VNTR loci.

The observed heterogeneity of the VNTR domains (they are still being discovered in the tubercle bacilli genome) provides great flexibility in designing new marker combinations that would enhance the discriminatory capacity of the genotyping method. In 2006, a new system employing 24 MIRU-VNTR loci (including 12 previously investigated) has been proposed [124]. Noteworthy, 15 (including 6 previously investigated) of those 24 loci account for 96% of all detected polymorphisms in *M. tuberculosis* strains. The discriminatory power of this new, 24-locus MIRU-VNTR typing system equals that of IS6110-RFLP profiling [125, 134, 135]. This has rendered 24-locus MIRU-VNTR typing the new gold standard in molecular typing of *M. tuberculosis* complex bacteria. Next to the proposed standardized MIRU-VNTR 15- or 24-loci sets, the use of another three loci, the so-called hypervariable loci (i.e., VNTRs 3232, 3820, and 4120), is recommended as a second-line typing step, particularly to differentiate Beijing genotype strains.

Overall, genotyping based on minisatellite sequences is a rapid, sensitive, and highly discriminating approach, which makes it well suited for large-scale investigations. A particular advantage of the VNTR genotyping, compared to the IS6110-RFLP typing, is its portability due to digitalization of the generated patterns and therefore simple intra- and interlaboratory comparability as well as the amenability to inclusion in web-based databases. Reproducibility of the method was expected to be good, due to the genetic stability of the targeted loci; yet, in the first worldwide proficiency study, both intra- and interlaboratory reproducibility proved to be suboptimal. Further harmonization of the laboratory methodology is still needed [136].

The occurrence of VNTR loci in the genomes of non-tuberculous mycobacteria remains largely obscure. Eight MIRU-VNTR-type loci have recently been described by Thibault et al. in *Mycobacterium avium* and *M. avium* subsp. *paratuberculosis* [137]. In Japan, a refined VNTR typing method using the *M. avium* tandem repeat (MATR) loci (MATR-VNTR) has been employed [138]. In *M. intracellulare* Dauchy et al. identified 45 potential MIRU-VNTR loci, 7 of which showed enough variability to be used as strain-level discriminatory markers [139]. The discriminatory power of the VNTR typing assays, based on the newly discovered loci in both species, was considered promising [80, 137, 139]. Recently, 13 VNTR loci have been described and applied to confirm clonal relationships between patient and environmental isolates of

M. ulcerans [140]. Twelve VNTR loci have been defined and tested against *M. marinum* isolates [141].

9. Methods Based on GC-Rich Sequences

The genome of *M. tuberculosis* has a particularly high GC content (>65.5%). The polymorphic GC-rich repetitive sequences (PGRS) are the most abundant type of repetitive DNA in the genome of *M. tuberculosis* complex. The PGRS elements occur at multiple loci and consist of several repeats of a 9 bp consensus sequence (5'-CGGCGGCAA-3'), tandemly arranged in up to 1.5 kb segments. Although the PGRS were initially identified in *M. tuberculosis* complex, they are now known to be present in other mycobacterial species, such as *M. kansasii*, *M. gastri*, and *M. szulgai* [142]. The PGRS bear important resemblance to the aforesaid MPTRs. The similarities between those repetitive elements include host range, structure, genetic stability, and copy number across the mycobacterial genome [143]. Furthermore, PGRS, like MPTR sequences, are part of protein-coding regions. Multiple tandem repetitions of the PGRS-encoded motif AsnGlyGlyAlaGlyGlyAla are found in glycine-rich proteins with a characteristic proline-glutamate (PE) residue group at the N terminus of the peptide. Members of the PE-PGRS family of proteins, analogously to PPE-MPTR protein family, are suspected to play a role in antigenic variability [118].

Since the number of the PGRS element and its distribution vary in different strains, it has been applied as a genetic marker for typing of *M. tuberculosis*. The most extensively used method utilizing this marker is PGRS-RFLP, the procedure of which is quite the same as that for the IS6110-RFLP, except that the chromosomal DNA is cut with *AluI* restriction endonuclease, instead of *PvuII*, and that a 3.4 kb fragment of the PGRS sequence, cloned in a recombinant plasmid pTBN12, is used as a probe in a hybridization step [142]. The PGRS-RFLP analysis or pTBN12-RFLP fingerprinting has been shown to have a relatively high discriminatory power, especially for IS6110 low-copy-number strains [56, 144, 145]. The pTBN12 fingerprinting method has similar limitations as the IS6110-RFLP typing, but additionally the hybridization patterns produced by PGRS typing are more complex and difficult in interpretation. A second method which benefits from the polymorphism of the PGRS sequence is double-repetitive-element PCR (DRE-PCR). This method relies upon amplification of the DNA segments located between IS6110 sequences and the PGRS sequences, by using primers directed outwards from the ends of both these repetitive elements (Figure 1(4)). Based on the distance between IS6110 and PGRS sequences and the copy number of these elements which differs between the strains, DRE-PCR yields strain-specific amplification patterns [65]. Although highly discriminative, the method suffers from poor reproducibility and a strong bias in the interpretation of the results [65, 146, 147].

A similar approach as that reported for DRE-PCR was used to design another typing method, called IS6110-ampliprinting [66]. In brief, the method measures the variability in the distances between IS6110 elements and copies

of MPTR sequences of *M. tuberculosis*, through unilateral-nested PCR with IS6110 and MPTR targeted primers, followed by hybridization with an IS6110-specific probe. The method, however, has not been widely adopted, mainly due to the limited number and size of generated PCR products [70].

A recent method that harbors GC-rich sequences, the IS6110-Mtb1-Mtb2 PCR typing, is guided by a similar principle as that involved in DRE-PCR profiling. Two PCR assays are performed with primers of two kinds, that is, primers complementary to terminal inverted repeats (TIR) flanking the IS6110 sequences and primers complementary to short (16 bp) GC-rich motifs, called Mtb1 (I PCR) and Mtb2 (II PCR). As a result, fragments of DNA located between Mtb1 and Mtb2 as well as between each of those elements and IS6110 sequences are amplified [67]. Although the IS6110-Mtb1-Mtb2 PCR method has not been exploited sufficiently, the results from the insofar performed studies suggest its usefulness in the molecular epidemiological investigations of TB. Most notably the discriminatory power of this method has been demonstrated as almost equivalent to that of IS6110-RFLP typing [67, 148, 149].

Although PGRS is not commonly being applied for strain typing of NTM, RFLP analysis with PGRS isolated from *M. tuberculosis* as a probe was successfully used to differentiate isolates of the species of *M. kansasii* [150] and *M. ulcerans* [151].

10. Repetitive Sequence-Based- (rep-) PCR

Every genotyping method, whose principle is based on the PCR amplification of DNA sequences between repetitive DNA elements, is referred to as repetitive sequence-based- (rep-) PCR. By using primers designed as extended away from the repetitive sequence elements, multiple amplified fragments are generated, depending on the sequence length between the repetitive elements. The amplified fragments produce a fingerprint pattern upon gel electrophoresis. IS6110 amplotyping, DRE-PCR, IS6110-ampliprinting, and IS6110-Mtb1-Mtb2 PCR, all these methods are representatives of rep-PCR technology. A long list of various repetitive elements that have been applied for genotyping of *M. tuberculosis* and nontuberculous mycobacteria includes also DNA sequences that lack species or even genus specificity, such as enterobacterial repetitive intergenic consensus (ERIC) and the (GTG)₅ sequences [152–155].

Recently, a commercially available rep-PCR system (DiversiLab System, bioMérieux, France) has been adapted for use on mycobacteria. The DiversiLab System, which takes advantage of various repetitive elements interspersed throughout different bacterial genomes, was evaluated on a collection of *M. tuberculosis* and *M. avium* complex isolates [156, 157] as well as *M. abscessus* isolates [158]. For *M. tuberculosis* and *M. avium* subsp. *avium*, the discriminatory ability of the assay equaled or exceeded that of IS6110-RFLP and IS1245-RFLP [156, 157, 159]. In more recent reports, rep-PCR was successfully used to disclose the source of infection in a patient with hypersensitivity pneumonitis caused by *M. avium* [160].

11. Random Amplified Polymorphic DNA (RAPD) Analysis

Random amplified polymorphic DNA (RAPD) analysis, also referred to as arbitrary primer PCR, depends on amplification of random fragments of genomic DNA using arbitrarily designed primers (5 to 50 bp) under low stringency conditions. The interstrain polymorphism is assessed by comparing the electrophoretic pattern of the PCR products. Although the method provides high discriminatory power, a number of limitations exist for this technique, of which the apparent lack of reproducibility is the most important. This is because differences between the strain patterns generated by RAPD are due to technical and operating parameters of the method rather than true interstrain genetic polymorphism. Variations in RAPD patterns are chiefly attributed to variations in the priming efficiency during early rounds of amplification, and these in turn depend on template concentration and purity, primer/template ratio, or the ramp times of the cyclers used. Nonetheless, the RAPD typing has been used to differentiate among isolates of *M. tuberculosis* complex isolates [161–164] as well as among isolates of numerous NTM species, including *M. abscessus* [165], *M. phocaicum* [166], *M. goodnae* [167], *M. szulgai* [168], and *M. malmoense* [169].

The RAPD analysis is usually performed on the entire genomic DNA; however in some modifications of the method only a fragment of the bacterial chromosome is used as a substrate for PCR. Such a strategy was employed for genotyping of *M. tuberculosis* strains by Abed et al., who used the 16–23S rDNA internal transcribed spacer (ITS) region within the rDNA operon as a target for PCR. The subsequent RAPD analysis of the amplified product resulted in highly polymorphic and easily interpretable profiles [170]. Still, the reproducibility of this RAPD-based method was found to be poor [171, 172].

12. Amplified Fragment Length Polymorphism (AFLP) Analysis

Amplified fragment length polymorphism (AFLP) analysis is a PCR-based method in which genomic DNA undergoes digestion with two restriction enzymes, a rare cutter and a frequent cutter, typically having six- and four-nucleotide-long recognitions sites, respectively (e.g., *EcoRI* and *MseI*, resp.), followed by ligation of two types of synthetic, double-stranded adaptors (10–30 bp) to the generated cohesive ends of DNA fragments. Amplification of a subset of restriction fragments is achieved by PCR with primers complementary to the adaptor sequence, the restriction site sequence, and a few nucleotides (1–3) extending beyond the restriction site. The amplicons are separated and visualized on denaturing polyacrylamide gels, usually through autoradiography, as the primers are radioactively labeled [173]. The conventional radioactive AFLP method has a lower discriminatory potential than IS6110-RFLP typing [174, 175]. However, the AFLP technology provides an extraordinary flexibility in designing the typing protocols of enhanced power of discrimination. Here, the choice of restriction endonucleases and the degree

of primer selectivity may largely determine the final resolution of the method.

A refinement of the traditional AFLP assay is the fluorescent (f)AFLP, where the subsets of the DNA digestion fragments are PCR-amplified with five primers, a single, nonselective, unlabeled forward primer targeting the *MseI* adaptor site and four reverse primers, targeting the *EcoRI* adaptor site, each containing the selective base A, G, C, or T, labeled with different fluorescent dyes. The amplified fragments are resolved by electrophoresis on an automated DNA sequencer and precisely sized using internal size standards [176]. Apart from the improved occupational safety, the discriminatory potential of fAFLP is higher than that of the radioactive AFLP and comparable to that achieved by IS6110-RFLP [176, 177].

The AFLP method has successfully been applied to various NTM species including the *M. avium* complex [178], *M. haemophilum* [179], and *M. marinum* and *M. ulcerans* [180], although its exact discriminatory power in larger sets of isolates has never been assessed.

Furthermore, AFLP analysis has clearly distinguished between *M. marinum* and *M. ulcerans*, conforming that these species which are difficult to distinguish by conventional methods, are genetically distinct [180, 181].

13. Multilocus Sequence Typing (MLST)

Genetic polymorphism between and within different mycobacterial species can also be investigated at the nucleotide sequence level. Such a concept gave rise to a new typing system, the so-called multilocus sequence typing (MLST), which evolved directly from the multilocus enzyme electrophoresis (MLEE or MEE) technique. Unlike MLEE, which targets the electrophoretic mobility of slowly evolving metabolic enzymes (usually 15 to 25), isolated from each bacterial isolate, MLST targets the sequences of a small number (up to 10) of housekeeping genes coding for vital enzymes and structural proteins. Although both MLEE and MLST schemes failed to reveal sufficient polymorphism among members of the *M. tuberculosis* complex [27, 182], these techniques were found useful for differentiation of various NTM species. For instance, by using MLEE, polymorphism was found among isolates of the *M. avium-M. intracellulare* complex [183–185]. On the other hand, the MLST analysis, based on 10 genetic loci, allowed the determination of the variability between subspecies and strains of *M. avium* and thus greatly improved our knowledge on genetic divergence and evolution of this group of NTM [186]. MLST has also proven to be valuable in investigations of NTM in laboratory outbreak settings [166].

Given the low degree of sequence polymorphisms in *M. tuberculosis* complex, standard MLST is poorly informative and inefficient. Its place has now been taken over by a new typing strategy based on analysis of single nucleotide polymorphisms (SNPs).

14. Single Nucleotide Polymorphism (SNP) Typing

Single nucleotide polymorphisms (SNPs) fall into two major groups: synonymous- (s-) SNPs and nonsynonymous- (ns-)

SNPs. The latter, if present in coding regions, introduce amino acid changes to the proteins. This in turn may influence the phenotype and be subjected to selection pressure. For instance, ns-SNPs are implicated in *M. tuberculosis* resistance to anti-TB drugs (drug resistance in *M. tuberculosis* is almost invariably associated with mutations (nonsynonymous point mutations, small deletions, and duplications) in specific, chromosomal loci) [187]. Screening for ns-SNPs in resistance-conferring genes provides important insights into the molecular mechanisms and dynamics of the development of drug resistance.

Contrariwise, s-SNPs do not alter the amino acid profiles and are thus phenotypically neutral. As s-SNPs are also believed to be evolutionary neutral, they are used for population genetics and for studying phylogenetic relationships among mycobacterial strains [188, 189]. A picture of the phylogenetic population structure of *M. tuberculosis* has recently been inferred by using a combination of s-SNP and non-SNP markers. Based on the SNPs at codon 463 of the *katG* gene and codon 95 of the *gyrA* gene, Sreevatsan et al. divided *M. tuberculosis* complex into 3 principal genetic groups (PGG1-PGG3) [14]. Those three PGG groups were further split into 9 major clusters (I–VIII and II.A) by analysis of additional 36 s-SNPs [189]. Recently, Dos Vultos et al. [190] have indicated SNPs in the 56 genes encoding 3R (DNA replication, recombination, and repair) system components as the key genetic markers to study the evolution of *M. tuberculosis*.

Although SNPs represent the most reliable markers for lineage classification of MTBC, their use is hampered by the need to test a large set of genes to achieve satisfactory resolution. Recently, Homolka et al. [16] have developed a SNP-based diagnostic algorithm allowing the identification of 17 MTBC phylogenetic lineages with high specificity. The algorithm involves sequence analysis of only five genes. The SNP typing approach is highly specific and sensitive, although SNP analysis has predominantly been used in genealogy, phylogenetic, and population genetics studies.

15. Deletion Mapping and Deligotyping

Comparative genomic studies of different strains of *M. tuberculosis* (e.g., H₃₇Rv, CDC1551) have proven that the loss of genetic material has stigmatized the evolutionary history of that species. Genomic deletions, also known as large-sequence polymorphisms (LSPs) or regions of difference (RD), have been detected across the *M. tuberculosis* genomes [191, 192]. For example, a total of 68 distinct deletions, ranging in size from 105 bp to approximately 12 kb, were found in 100 *M. tuberculosis* clinical isolates [192]. Deletions are not randomly distributed but rather appear in aggregations. They occur within both intra- and intergenic regions [191–194]. Noteworthy, almost half of the LSPs identified in *M. tuberculosis* H₃₇Rv and CDC1551 strains involved genes encoding PPE and PE family proteins [191]. Since LSPs have emerged as a significant source of interstrain genetic variability, they have been used as markers for genotyping. Analysis of chromosomal deletions has been shown as an extremely attractive approach for studying the phylogeny and evolution of

M. tuberculosis complex [192, 195, 196]. Deletion analysis, also referred to as deligotyping, can be performed either by a simple PCR-based method or by automated microarray techniques. The resolution of the method can greatly be increased if specific sequences flanking each side of the deletion element are known. Recently, a high-throughput method for distinguishing LSPs was invented. Both the concept and procedure of this method were patterned upon the spoligotyping technique. Here, deletion events are detected in 43 genomic loci by amplifying them in a multiplex PCR assay and subjecting the amplicons to hybridization with a set of 43 probes, whose sequences directly correspond to the targeted loci [197]. Deligotyping is a very sensitive and efficacious approach for rapid screening of clinical isolates of *M. tuberculosis*. The method is also well suited for constructing robust phylogenetic relationships [198].

16. Concluding Remarks

As described above, there is a wide range of methods available for genotyping of *M. tuberculosis* complex and NTM species (Tables 1 and 2). Each method has its own benefits and shortfalls, and none of them have proven clearly superior to any of the others. The choice of the optimal typing system depends heavily on the sample under investigation, the setting in which typing is performed, and the expected outcome. For instance, spoligotyping is of particular value in population-based studies to define the phylogeographic specificity of circulating clades of tubercle bacilli. However, to assess the genetic relatedness and the epidemiological links among TB outbreak-related cases, the IS6110-RFLP or MIRU-VNTR typing is preferably chosen. Again, whereas spoligotyping is recommended as a preliminary screening test of a large number of *M. tuberculosis* isolates, disclosure of true genetic relationships between the isolates requires more discriminating methods, such as MIRU-VNTR typing, to be performed within the spoligotype-defined clusters.

An ideal molecular typing method should accommodate the requirements with respect to both performance and analytical criteria. The desired performance parameters include technical simplicity (easiness of performance), reproducibility, robustness, and time and cost effectiveness. An attractive feature of the method could be its applicability directly to clinical material. Another special advantage is a standardized and easily portable and interpretable format of the results (e.g., digital codes), facilitating databasing and interlaboratory comparative studies. As for the analytical parameters, the most important are the level of discrimination and stability of the genetic marker used. The general rule that the higher the discriminatory power of a given method, the more reliable the results obtained guides most of the molecular epidemiology investigations. However this may not always be the case. The validity of such assumption depends on several issues related to clustering, such as the characteristics of the study setting, the proportion of cases included (completeness of sampling), or the period of case recruitment (duration of the study). In other words, a number of important considerations have to be taken into account when choosing an appropriate typing methodology in terms of discrimination capacity.

TABLE 1: Selected typing methods for *Mycobacterium tuberculosis* complex and level of genetic polymorphism they reveal.

Typing method	DNA target	Polymorphism	References*
IS6110-RFLP		High	van Embden et al., 1993 [48]
ML-PCR		High	Haas et al., 1993 [58]
FliP	IS6110	High	Reisig et al., 2005 [59]
LM-PCR		High	Prod'hom et al., 1997 [60]
HIP		High	Kearns et al., 2000 [61]
Spoligotyping	DR locus	Low	Kamerbeek et al., 1997 [62]
VNTR typing	ETRs A-E	Low	Frothingham and Meeker-O'Connell, 1998 [63]
MIRU-VNTR typing	MIRUs	High	Supply et al., 2001 [64]
DRE-PCR	IS6110/PGRS	High	Friedman et al., 1995 [65]
IS6110-ampliprinting	IS6110/MPTR	High	Plikaytis et al., 1993 [66]
IS6110-Mtb1-Mtb2 PCR	IS6110/Mtb1/Mtb2	High	Kotlowski et al., 2004 [67]

*Papers with original description of a given method.

RFLP: restriction fragment length polymorphism; ML-PCR: mixed linker PCR; FliP: fast ligation-mediated PCR; LM-PCR: ligation-mediated PCR; HIP: heminested inverse PCR; VNTRs: variable numbers of tandem repeats; MIRU: mycobacterial interspersed repetitive units; DRE-PCR: double-repetitive-element PCR; DR: direct repeat; ETR: exact tandem repeat; PGRS: polymorphic GC-rich sequence; MPTR: major polymorphic tandem repeat.

TABLE 2: Discriminatory power of selected typing methods for *Mycobacterium tuberculosis* complex and nontuberculous mycobacteria and level of genetic polymorphism they reveal.

Typing method	MTBC	NTM
RFLP	High	Insufficient data
RAPD	Medium	High
PFGE	Medium	High
AFLP	High	Insufficient data
Spoligotyping	Low/diverse	None
MIRU-VNTR typing	High/diverse	Insufficient data

MTBC: *Mycobacterium tuberculosis* complex; NTM: nontuberculous mycobacteria; RFLP: restriction fragment length polymorphism; RAPD: random amplified polymorphic DNA; PFGE: pulsed-field gel electrophoresis; AFLP: amplified fragment length polymorphism; MIRU-VNTR: mycobacterial interspersed repetitive unit-variable number of tandem repeats.

The discriminatory power of molecular markers is directly linked to the genetic stability of each marker. There have been observed minor changes in DNA fingerprint patterns of *M. tuberculosis* strains isolated not only from epidemiologically related TB cases, but also from the same patient at different points of time [52, 199–202]. The stability of the genotypic patterns over time reflects the evolutionary rate, also referred to as a “molecular clock,” of each genetic marker. For example, a half-life of the IS6110-RFLP profiles has been shown to be much shorter than that of the spoligo-type profiles [70, 98]. Likewise, the combined molecular clock of the MIRU-VNTR loci has been shown to be slower than that of IS6110-RFLP [203]. In general, the best molecular marker is the one, whose “molecular clock” is, on the one hand, fast enough to distinguish unrelated cases and, on the other hand, sufficiently slow to capture epidemiologically linked cases [204]. Markers evolving rapidly and those evolving at slow rates may either underestimate or overestimate the amount of recent transmission of the disease, respectively.

The choice of a genotyping method, with respect to its discriminatory ability, depends on the type of research in

question. Whereas a highly discriminatory method, that is with a fast “molecular clock,” would be required to determine whether an infection is a reactivation of an infection acquired in the past (latent infection) or rather a reinfection with a new strain, a method with a slow “molecular clock” would be needed for global strain tracking and evolutionary studies.

The resolution power of different typing methods, yielding very diversified genetic patterns, has had an impact on the definition of clustering. Indeed, there persists a controversy in the literature on whether or not isolates whose patterns show subtle differences, that is, of 1-2 bands, in the IS6110-RFLP patterns or single locus variations (SLVs) in the MIRU-VNTR patterns may be regarded as part of a genetic cluster and thus constitute an ongoing chain of transmission. Whereas some authors apply a strict cluster definition including only isolates with identical genotypes [134, 135, 205, 206], the others advocate the use of slightly relaxed criteria for defining clusters, with a tolerance of a single- or double-band difference in the RFLP profiles and/or SLVs or double locus variations (DLVs) in the MIRU-VNTR profiles [52, 200, 207, 208]. With the latter approach, another important question arises, that is, to what extent the fingerprint patterns of two isolates may differ before they are no longer considered to be clustered. Since no clear cut-offs exist for defining a cluster, the decision which isolates should be included or excluded from clusters is largely a matter of arbitrariness. In general, the more lenient the assumed criteria, the higher the chance of detecting clusters, but the lower the likelihood that a cluster represents epidemiologically related cases [209].

One has also to be mindful that the identity or high similarity of DNA fingerprints from two individuals, obtained by using even the most discriminating techniques, may not always mean a recently transmitted infection. Other explanations are possible, including the simultaneous reactivation of a previously acquired infection with the same organism (coincidence of time), the regional predominance of a particular strain, circulating over a long time, or a laboratory cross-contamination [210].

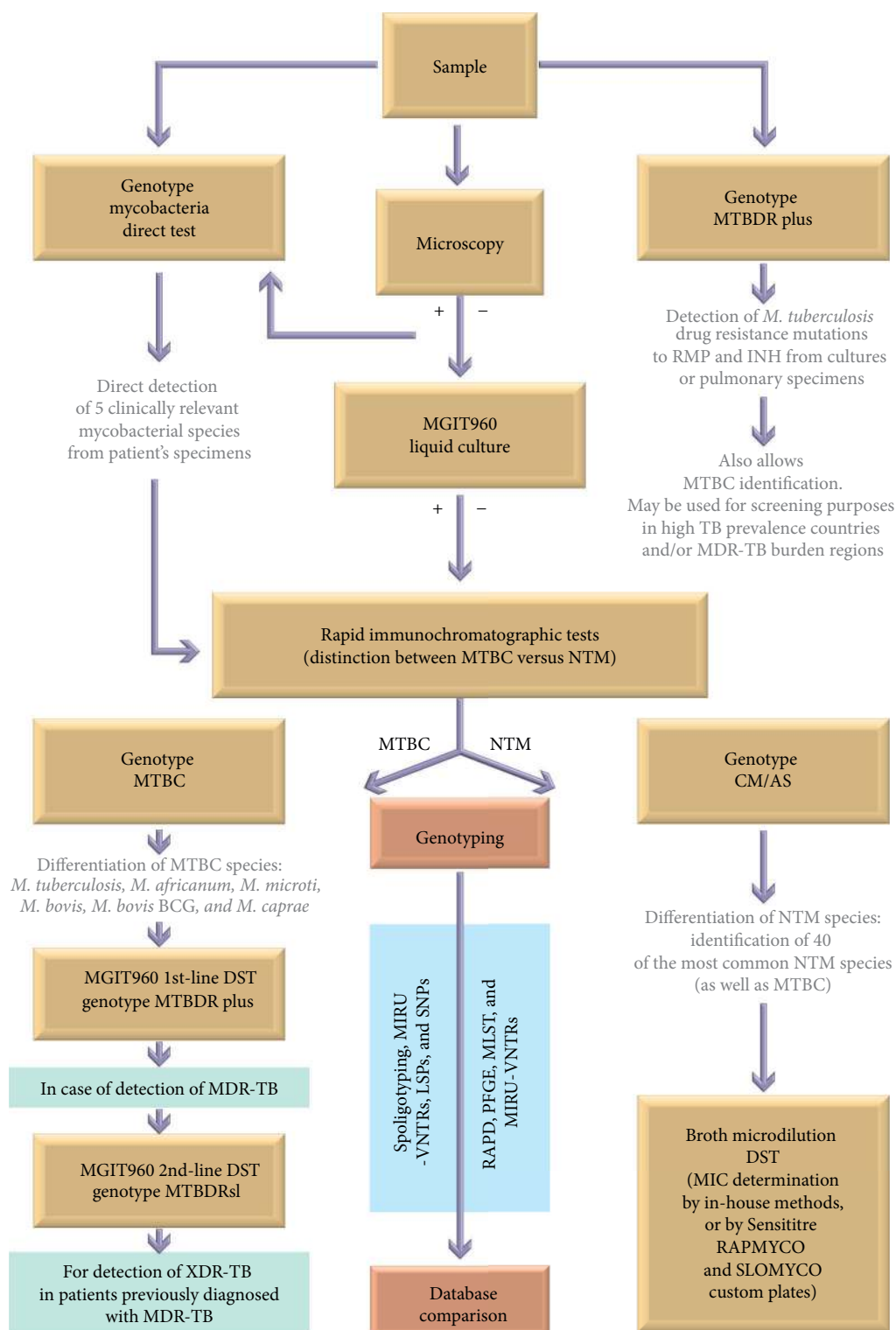


FIGURE 2: Schematic flow diagram illustrating processing of clinical samples for species identification, drug susceptibility testing (DST), and genotyping of mycobacteria belonging to *Mycobacterium tuberculosis* complex (MTBC) and nontuberculous mycobacteria (NTM). MIRU-VNTRs: mycobacterial interspersed repetitive units-variable number of tandem repeat loci; LSPs: large sequence polymorphisms; SNPs: single nucleotide polymorphisms; RAPD: random amplified polymorphic DNA; PFGE: pulsed-field gel electrophoresis; MLST: multilocus sequence typing.

Identical genotypic patterns may not indicate clonality, even when multiple genetic markers are employed. This was best evidenced by Niemann et al. who compared the complete genomes of two *M. tuberculosis* Beijing genotype isolates from a high-incidence region (Karakalpakstan, Uzbekistan), of which one was drug susceptible and the other was multidrug resistant. Both isolates shared the same IS6110-RFLP pattern and the same allele at 23 out of 24 MIRU-VNTR loci, yet they differed by 130 SNPs and one large deletion [211]. This finding implies some important messages. First, *M. tuberculosis* isolates exhibiting identical DNA fingerprinting profiles may still display substantial genomic diversity. This in turn may lead to misinterpretation of the extent of TB transmission in a community or invalid differentiation between disease relapse and exogenous reinfection, when using standard genotyping tools. It seems that the optimal option to fully explore the phylogenetic branching and variation on strain level and to justifiably draw epidemiological conclusions on the etiology, host-range, and transmission of TB disease would be the application of whole-genome sequencing (WGS) analysis. Although still an expensive solution, with the plummeting cost of DNA sequencing, the WGS may become an omnipotent approach to replace all previously known diagnostic tests for *M. tuberculosis*, including those for identification or drug susceptibility profiling [212]. As for the present, a status of the “gold standard” typing method for *M. tuberculosis* is still being held by the IS6110-RFLP. However, it is being increasingly replaced by the MIRU-VNTR typing, not only because its discriminatory power equals that of IS6110-RFLP but only because it has methodological and practical advantages over RFLP, most important of which is that the PCR-based typing method requires fewer bacteria and consequently shortens considerably delay in obtaining genotypes. Thus, the MIRU-VNTR typing can be regarded as the new standard for TB molecular epidemiology.

As for the NTM, a number of genotyping methods are available, most of which were first applied to *M. tuberculosis* complex and then turned out to work for specific NTM species, but the discriminatory power of those methods has not been fully examined. A method most commonly applied in various NTM species, with only slight species-specific modifications, has been PFGE typing. Due to its wide use, it could be considered the “gold standard” for all NTM species except for *M. avium*. For the latter, IS1245 RFLP typing is the widely recognized reference method [79]. MLST has also the potential to become a reference method, although its wider use is hampered by limited access and high costs of the sequencing facilities. However, with the level of genetic diversity in NTM (sub)species being ill-defined and the discriminatory power of most of the markers not fully established, for a proposal of the “gold standard” typing system for NTM species to be delivered, further studies are required (Figure 2).

Although there is currently no genotyping method that would work in diverse settings and population groups or to be equally effective at answering particular epidemiological questions, the application of molecular typing methods has significantly advanced our knowledge of the transmission and pathogenesis of mycobacteria.

Molecular typing methods have permitted investigation of the outbreaks [36, 61, 100, 139, 165, 166, 213, 214], discrimination between exogenous reinfection and endogenous reactivation [209, 210, 215–217], identification of mixed infections [215, 218, 219], and cases of misdiagnosis due to laboratory contamination [220–223]. Molecular markers have also extensively been used for tracking transmission patterns within specific populations and/or defined geographical settings [106–108, 153, 206, 224–227]. Finally, genotyping methods have shed important light on the phylogeny and evolutionary history of the mycobacterial species [12, 14, 189, 194].

Methods of molecular typing constitute an integral element of virtually all epidemiological studies on mycobacterial infections. They continue to substantially improve our understanding of the biology of mycobacteria and are believed to provide novel and powerful tools to combat and/or protect against the diseases caused by these pathogens.

Conflict of Interests

The authors report no conflict of interests.

References

- [1] J. van Ingen, “Diagnosis of nontuberculous mycobacterial infections,” *Seminars in Respiratory and Critical Care Medicine*, vol. 34, no. 1, pp. 103–109, 2013.
- [2] WHO, “Global tuberculosis control: epidemiology, strategy, financing,” WHO Report 2009, World Health Organization, Geneva, Switzerland, 2009.
- [3] J. O. Falkinham III, “Surrounded by mycobacteria: nontuberculous mycobacteria in the human environment,” *Journal of Applied Microbiology*, vol. 107, no. 2, pp. 356–367, 2009.
- [4] J. van Ingen, A. F. Verhagen, P. N. Dekhuijzen et al., “Surgical treatment of non-tuberculous mycobacterial lung disease: strike in time,” *International Journal of Tuberculosis and Lung Disease*, vol. 14, no. 1, pp. 99–105, 2010.
- [5] D. R. Prevots, P. A. Shaw, D. Strickland et al., “Nontuberculous mycobacterial lung disease prevalence at four integrated health care delivery systems,” *The American Journal of Respiratory and Critical Care Medicine*, vol. 182, no. 7, pp. 970–976, 2010.
- [6] H. W. Engel, “Mycobacteriophages and phage typing,” *Annales de Microbiologie*, vol. 129, no. 1, pp. 75–90, 1978.
- [7] H. Gruft, R. Johnson, R. Claffin, and A. Loder, “Phage-typing and drug-resistance patterns as tools in mycobacterial epidemiology,” *The American Review of Respiratory Disease*, vol. 130, no. 1, pp. 96–97, 1984.
- [8] T. A. Rado, J. H. Bates, and H. W. B. Engel, “World Health Organization studies in bacteriophage typing of mycobacteria: subdivision of the species *Mycobacterium tuberculosis*,” *The American Review of Respiratory Disease*, vol. 111, no. 4, pp. 459–468, 1975.
- [9] M. C. Roman and M. J. Sicilia, “Preliminary investigation of *Mycobacterium tuberculosis* biovars,” *Journal of Clinical Microbiology*, vol. 20, no. 5, pp. 1015–1016, 1984.
- [10] J. M. Grange and A. Laszlo, “Serodiagnostic tests for tuberculosis: a need for assessment of their operational predictive accuracy and acceptability,” *Bulletin of the World Health Organization*, vol. 68, no. 5, pp. 571–576, 1990.

- [11] R. Frothingham, H. G. Hills, and K. H. Wilson, "Extensive DNA sequence conservation throughout the *Mycobacterium tuberculosis* complex," *Journal of Clinical Microbiology*, vol. 32, no. 7, pp. 1639–1643, 1994.
- [12] M. M. Gutacker, J. C. Smoot, C. A. Migliaccio et al., "Genome-wide analysis of synonymous single nucleotide polymorphisms in *Mycobacterium tuberculosis* complex organisms: resolution of genetic relationships among closely related microbial strains," *Genetics*, vol. 162, no. 4, pp. 1533–1543, 2002.
- [13] J. M. Musser, A. Amin, and S. Ramaswamy, "Negligible genetic diversity of *Mycobacterium tuberculosis* host immune system protein targets: evidence of limited selective pressure," *Genetics*, vol. 155, no. 1, pp. 7–16, 2000.
- [14] S. Sreevatsan, X. Pan, K. E. Stockbauer et al., "Restricted structural gene polymorphism in the *Mycobacterium tuberculosis* complex indicates evolutionarily recent global dissemination," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 94, no. 18, pp. 9869–9874, 1997.
- [15] P. Supply, R. M. Warren, A.-L. Bañuls et al., "Linkage disequilibrium between minisatellite loci supports clonal evolution of *Mycobacterium tuberculosis* in a high tuberculosis incidence area," *Molecular Microbiology*, vol. 47, no. 2, pp. 529–538, 2003.
- [16] S. Homolka, M. Projahn, S. Feuerriegel et al., "High resolution discrimination of clinical *Mycobacterium tuberculosis* complex strains based on single nucleotide polymorphisms," *PLoS ONE*, vol. 7, no. 7, Article ID e39855, 2012.
- [17] R. Hershberg, M. Lipatov, P. M. Small et al., "High functional diversity in *Mycobacterium tuberculosis* driven by genetic drift and human demography," *PLoS Biology*, vol. 6, no. 12, article e311, 2008.
- [18] T. P. Stinear, T. Seemann, P. F. Harrison et al., "Insights from the complete genome sequence of *Mycobacterium marinum* on the evolution of *Mycobacterium tuberculosis*," *Genome Research*, vol. 18, no. 5, pp. 729–741, 2008.
- [19] D. M. Collins and G. W. de Lisle, "DNA restriction endonuclease analysis of *Mycobacterium tuberculosis* and *Mycobacterium bovis* BCG," *Journal of General Microbiology*, vol. 130, no. 4, pp. 1019–1021, 1984.
- [20] P. Palittapongarnpim, S. Rienthong, and W. Panbangred, "Comparison of restriction fragment length polymorphism of *Mycobacterium tuberculosis* isolated from cerebrospinal fluid and sputum: a preliminary report," *Tubercle and Lung Disease*, vol. 74, no. 3, pp. 204–207, 1993.
- [21] B. C. Ross, K. Raios, K. Jackson, A. Sievers, and B. Dwyer, "Differentiation of *Mycobacterium tuberculosis* strains by use of a nonradioactive southern blot hybridization method," *Journal of Infectious Diseases*, vol. 163, no. 4, pp. 904–907, 1991.
- [22] S. A. Shoemaker, J. H. Fisher, W. D. Jones Jr., and C. H. Scoggin, "Restriction fragment analysis of chromosomal DNA defines different strains of *Mycobacterium tuberculosis*," *The American Review of Respiratory Disease*, vol. 134, no. 2, pp. 210–213, 1986.
- [23] K. D. Eisenach, J. T. Crawford, and J. H. Bates, "Genetic relatedness among strains of the *Mycobacterium tuberculosis* complex. Analysis of restriction fragment heterogeneity using cloned DNA probes," *The American Review of Respiratory Disease*, vol. 133, no. 6, pp. 1065–1068, 1986.
- [24] K. D. Eisenach, J. T. Crawford, and J. H. Bates, "Repetitive DNA sequences as probes for *Mycobacterium tuberculosis*," *Journal of Clinical Microbiology*, vol. 26, no. 11, pp. 2240–2245, 1988.
- [25] F. C. Tenover, R. D. Arbeit, R. V. Goering et al., "Interpreting chromosomal DNA restriction patterns produced by pulsed-field gel electrophoresis: criteria for bacterial strain typing," *Journal of Clinical Microbiology*, vol. 33, no. 9, pp. 2233–2239, 1995.
- [26] M. M. Feizabadi, I. D. Robertson, R. Edwards, D. V. Cousins, and D. J. Hampson, "Genetic differentiation of Australian isolates of *Mycobacterium tuberculosis* by pulsed-field gel electrophoresis," *Journal of Medical Microbiology*, vol. 46, no. 6, pp. 501–505, 1997.
- [27] M. M. Feizabadi, I. D. Robertson, D. V. Cousins, and D. J. Hampson, "Genomic analysis of *Mycobacterium bovis* and other members of the *Mycobacterium tuberculosis* complex by isoenzyme analysis and pulsed-field gel electrophoresis," *Journal of Clinical Microbiology*, vol. 34, no. 5, pp. 1136–1142, 1996.
- [28] Y. Zhang, R. J. Wallace Jr., and G. H. Mazurek, "Genetic differences between BCG substrains," *Tubercle and Lung Disease*, vol. 76, no. 1, pp. 43–50, 1995.
- [29] S. P. Singh, H. Salamon, C. J. Lahti, M. Farid-Moyer, and P. M. Small, "Use of pulsed-field gel electrophoresis for molecular epidemiologic and population genetic studies of *Mycobacterium tuberculosis*," *Journal of Clinical Microbiology*, vol. 37, no. 6, pp. 1927–1931, 1999.
- [30] Y. Zhang, G. H. Mazurek, M. D. Cave et al., "DNA polymorphisms in strains of *Mycobacterium tuberculosis* analyzed by pulsed-field gel electrophoresis: a tool for epidemiology," *Journal of Clinical Microbiology*, vol. 30, no. 6, pp. 1551–1556, 1992.
- [31] Y. Iinuma, S. Ichiyama, Y. Hasegawa, K. Shimokata, S. Kawahara, and T. Matsushima, "Large-restriction-fragment analysis of *Mycobacterium kansasii* genomic DNA and its application in molecular typing," *Journal of Clinical Microbiology*, vol. 35, no. 3, pp. 596–599, 1997.
- [32] Y. Zhang, L. B. Mann, R. W. Wilson et al., "Molecular Analysis of *Mycobacterium kansasii* isolates from the United States," *Journal of Clinical Microbiology*, vol. 42, no. 1, pp. 119–125, 2004.
- [33] G. H. Mazurek, S. Hartman, Y. Zhang et al., "Large DNA restriction fragment polymorphism in the *Mycobacterium avium-M. intracellulare* complex: a potential epidemiologic tool," *Journal of Clinical Microbiology*, vol. 31, no. 2, pp. 390–394, 1993.
- [34] V. Lalande, F. Barbut, A. Varnerot et al., "Pseudo-outbreak of *Mycobacterium gordonae* associated with water from refrigerated fountains," *Journal of Hospital Infection*, vol. 48, no. 1, pp. 76–79, 2001.
- [35] M. A. Yakrus and W. L. Straus, "DNA polymorphisms detected in *Mycobacterium haemophilum* by pulsed-field gel electrophoresis," *Journal of Clinical Microbiology*, vol. 32, no. 4, pp. 1083–1084, 1994.
- [36] J. S. Hector, Y. Pang, G. H. Mazurek, Y. Zhang, B. A. Brown, and R. J. Wallace Jr., "Large restriction fragment patterns of genomic *Mycobacterium fortuitum* DNA as strain-specific markers and their use in epidemiologic investigation of four nosocomial outbreaks," *Journal of Clinical Microbiology*, vol. 30, no. 5, pp. 1250–1255, 1992.
- [37] R. J. Wallace Jr., Y. Zhang, B. A. Brown, V. Fraser, G. H. Mazurek, and S. Maloney, "DNA large restriction fragment patterns of sporadic and epidemic nosocomial strains of *Mycobacterium chelonae* and *Mycobacterium abscessus*," *Journal of Clinical Microbiology*, vol. 31, no. 10, pp. 2697–2701, 1993.
- [38] Y. Zhang, M. A. Yakrus, E. A. Graviss et al., "Pulsed-field gel electrophoresis study of *Mycobacterium abscessus* isolates previously affected by DNA degradation," *Journal of Clinical Microbiology*, vol. 42, no. 12, pp. 5582–5587, 2004.
- [39] S. T. Cole, R. Brosch, J. Parkhill et al., "Deciphering the biology of *Mycobacterium tuberculosis* from the complete genome sequence," *Nature*, vol. 393, no. 6685, pp. 537–544, 1998.

- [40] P. W. Hermans, D. van Soolingen, J. W. Dale et al., "Insertion element IS986 from *Mycobacterium tuberculosis*: a useful tool for diagnosis and epidemiology of tuberculosis," *Journal of Clinical Microbiology*, vol. 28, no. 9, pp. 2051–2058, 1990.
- [41] R. A. McAdam, P. W. Hermans, D. van Soolingen et al., "Characterization of a *Mycobacterium tuberculosis* insertion sequence belonging to the IS3 family," *Molecular Microbiology*, vol. 4, no. 9, pp. 1607–1613, 1990.
- [42] D. Thierry, A. Brisson-Noel, V. Vincent-Levy-Frebault, S. Nguyen, J.-L. Guesdon, and B. Gicquel, "Characterization of a *Mycobacterium tuberculosis* insertion sequence, IS6110, and its application in diagnosis," *Journal of Clinical Microbiology*, vol. 28, no. 12, pp. 2668–2673, 1990.
- [43] D. Thierry, M. D. Cave, K. D. Eisenach et al., "IS6110, an IS-like element of *Mycobacterium tuberculosis* complex," *Nucleic Acids Research*, vol. 18, no. 1, article 188, 1990.
- [44] S. Wall, K. Ghanekar, J. McFadden, and J. W. Dale, "Context-sensitive transposition of IS6110 in mycobacteria," *Microbiology*, vol. 145, no. 11, pp. 3169–3176, 1999.
- [45] H. Yesilkaya, J. W. Dale, N. J. C. Strachan, and K. J. Forbes, "Natural transposon mutagenesis of clinical isolates of *Mycobacterium tuberculosis*: how many genes does a pathogen need?" *Journal of Bacteriology*, vol. 187, no. 19, pp. 6726–6732, 2005.
- [46] M. D. Cave, K. D. Eisenach, P. F. McDermott, J. H. Bates, and J. T. Crawford, "IS6110: conservation of sequence in the *Mycobacterium tuberculosis* complex and its utilization in DNA fingerprinting," *Molecular and Cellular Probes*, vol. 5, no. 1, pp. 73–80, 1991.
- [47] D. van Soolingen, P. W. Hermans, P. E. de Haas, D. R. Soll, and J. D. A. van Embden, "Occurrence and stability of insertion sequences in *Mycobacterium tuberculosis* complex strains: evaluation of an insertion sequence-dependent DNA polymorphism as a tool in the epidemiology of tuberculosis," *Journal of Clinical Microbiology*, vol. 29, no. 11, pp. 2578–2586, 1991.
- [48] J. D. van Embden, M. D. Cave, J. T. Crawford et al., "Strain identification of *Mycobacterium tuberculosis* by DNA fingerprinting: recommendations for a standardized methodology," *Journal of Clinical Microbiology*, vol. 31, no. 2, pp. 406–409, 1993.
- [49] J. D. van Embden, D. van Soolingen, H. F. Heersma et al., "Establishment of a European network for the surveillance of *Mycobacterium tuberculosis*, MRSA and penicillin-resistant pneumococci," *Journal of Antimicrobial Chemotherapy*, vol. 38, no. 5, pp. 905–907, 1996.
- [50] H. F. Heersma, K. Kremer, and J. D. van Embden, "Computer analysis of IS6110 RFLP patterns of *Mycobacterium tuberculosis*," *Methods in Molecular Biology*, vol. 101, pp. 395–422, 1998.
- [51] A. S. de Boer, M. W. Borgdorff, P. E. de Haas, N. J. D. Nagelkerke, J. D. van Embden, and D. van Soolingen, "Analysis of rate of change of IS6110 RFLP patterns of *Mycobacterium tuberculosis* based on serial patient isolates," *Journal of Infectious Diseases*, vol. 180, no. 4, pp. 1238–1244, 1999.
- [52] R. W. Yeh, A. P. de Leon, C. B. Agasino et al., "Stability of *Mycobacterium tuberculosis* DNA genotypes," *Journal of Infectious Diseases*, vol. 177, no. 4, pp. 1107–1111, 1998.
- [53] R. E. Barlow, D. M. Gascoyne-Binzi, S. H. Gillespie, A. Dickens, S. Qamer, and P. M. Hawkey, "Comparison of variable number tandem repeat and IS6110-restriction fragment length polymorphism analyses for discrimination of high- and low-copy-number IS6110 *Mycobacterium tuberculosis* isolates," *Journal of Clinical Microbiology*, vol. 39, no. 7, pp. 2453–2457, 2001.
- [54] J. Bauer, Å. B. Andersen, K. Kremer, and H. Miørner, "Usefulness of spoligotyping to discriminate IS6110 low-copy-number *Mycobacterium tuberculosis* complex strains cultured in Denmark," *Journal of Clinical Microbiology*, vol. 37, no. 8, pp. 2602–2606, 1999.
- [55] J. T. Rhee, M. M. Tanaka, M. A. Behr et al., "Use of multiple markers in population-based molecular epidemiologic studies of tuberculosis," *International Journal of Tuberculosis and Lung Disease*, vol. 4, no. 12, pp. 1111–1119, 2000.
- [56] Z. H. Yang, K. Ijaz, J. H. Bates, K. D. Eisenach, and M. D. Cave, "Spoligotyping and polymorphic GC-rich repetitive sequence fingerprinting of *Mycobacterium tuberculosis* strains having few copies of IS6110," *Journal of Clinical Microbiology*, vol. 38, no. 10, pp. 3572–3576, 2000.
- [57] T. D. McHugh, L. E. Newport, and S. H. Gillespie, "IS6110 homologs are present in multiple copies in Mycobacteria other than tuberculosis: causing mycobacteria," *Journal of Clinical Microbiology*, vol. 35, no. 7, pp. 1769–1771, 1997.
- [58] W. H. Haas, W. R. Butler, C. L. Woodley, and J. T. Crawford, "Mixed-linker polymerase chain reaction: a new method for rapid fingerprinting of isolates of the *Mycobacterium tuberculosis* complex," *Journal of Clinical Microbiology*, vol. 31, no. 5, pp. 1293–1298, 1993.
- [59] F. Reisig, K. Kremer, B. Amthor, D. van Soolingen, and W. H. Haas, "Fast ligation-mediated PCR, a fast and reliable method for IS6110-based typing of *Mycobacterium tuberculosis* complex," *Journal of Clinical Microbiology*, vol. 43, no. 11, pp. 5622–5627, 2005.
- [60] G. Prod'Hom, C. Guilhot, M. C. Gutierrez, A. Varnerot, B. Gicquel, and V. Vincent, "Rapid discrimination of *Mycobacterium tuberculosis* complex strains by ligation-mediated PCR fingerprint analysis," *Journal of Clinical Microbiology*, vol. 35, no. 12, pp. 3331–3334, 1997.
- [61] A. M. Kearns, A. Barrett, C. Marshall et al., "Epidemiology and molecular typing of an outbreak of tuberculosis in a hostel for homeless men," *Journal of Clinical Pathology*, vol. 53, no. 2, pp. 122–124, 2000.
- [62] J. Kamerbeek, L. Schouls, A. Kolk et al., "Simultaneous detection and strain differentiation of *Mycobacterium tuberculosis* for diagnosis and epidemiology," *Journal of Clinical Microbiology*, vol. 35, no. 4, pp. 907–914, 1997.
- [63] R. Frothingham and W. A. Meeker-O'Connell, "Genetic diversity in the *Mycobacterium tuberculosis* complex based on variable numbers of tandem DNA repeats," *Microbiology*, vol. 144, no. 5, pp. 1189–1196, 1998.
- [64] P. Supply, S. Lesjean, E. Savine, K. Kremer, D. van Soolingen, and C. Locht, "Automated high-throughput genotyping for study of global epidemiology of *Mycobacterium tuberculosis* based on mycobacterial interspersed repetitive units," *Journal of Clinical Microbiology*, vol. 39, no. 10, pp. 3563–3571, 2001.
- [65] C. R. Friedman, M. Y. Stoeckle, W. D. Johnson Jr., and L. W. Riley, "Double-repetitive-element PCR method for subtyping *Mycobacterium tuberculosis* clinical isolates," *Journal of Clinical Microbiology*, vol. 33, no. 5, pp. 1383–1384, 1995.
- [66] B. B. Plikaytis, J. T. Crawford, C. L. Woodley et al., "Rapid, amplification-based fingerprinting of *Mycobacterium tuberculosis*," *Journal of General Microbiology*, vol. 139, no. 7, pp. 1537–1542, 1993.
- [67] R. Kotłowski, I. C. Shamputa, N. A. El Aila et al., "PCR-Based Genotyping of *Mycobacterium tuberculosis* with New GC-Rich Repeated Sequences and IS6110 Inverted Repeats Used as Primers," *Journal of Clinical Microbiology*, vol. 42, no. 1, pp. 372–377, 2004.

- [68] P. Palittapongarnpim, S. Chomyc, A. Fanning, and D. Kunitomo, "DNA fingerprinting of *Mycobacterium tuberculosis* isolates by ligation-mediated polymerase chain reaction," *Nucleic Acids Research*, vol. 21, no. 3, pp. 761–762, 1993.
- [69] S. Bonora, M. C. Gutierrez, G. di Perri et al., "Comparative evaluation of ligation-mediated PCR and spoligotyping as screening methods for genotyping of *Mycobacterium tuberculosis* strains," *Journal of Clinical Microbiology*, vol. 37, no. 10, pp. 3118–3123, 1999.
- [70] K. Kremer, D. van Soolingen, R. Frothingham et al., "Comparison of methods based on different molecular epidemiological markers for typing of *Mycobacterium tuberculosis* complex strains: interlaboratory study of discriminatory power and reproducibility," *Journal of Clinical Microbiology*, vol. 37, no. 8, pp. 2607–2618, 1999.
- [71] H. Mardassi, A. Namouchi, A. Karbouli et al., "Usefulness of ligation-mediated PCR genotyping in tracking outbreak-associated *Mycobacterium tuberculosis* strains," *Archives de l'Institut Pasteur de Tunis*, vol. 82, no. 1–4, pp. 23–29, 2005.
- [72] S. Patel, S. Wall, and N. A. Saunders, "Heminested inverse PCR for IS6110 fingerprinting of *Mycobacterium tuberculosis* strains," *Journal of Clinical Microbiology*, vol. 34, no. 7, pp. 1686–1690, 1996.
- [73] H. Neimark, M. Ali Baig, and S. Carleton, "Direct identification and typing of *Mycobacterium tuberculosis* by PCR," *Journal of Clinical Microbiology*, vol. 34, no. 10, pp. 2454–2459, 1996.
- [74] B. C. Ross and B. Dwyer, "Rapid, simple method for typing isolates of *Mycobacterium tuberculosis* by using the polymerase chain reaction," *Journal of Clinical Microbiology*, vol. 31, no. 2, pp. 329–334, 1993.
- [75] K. Y. Yuen, C. M. Chan, K. S. Chan, W. C. Yam, P. L. Ho, and P. Y. Chau, "IS6110 based amplotyping assay and RFLP fingerprinting of clinical isolates of *Mycobacterium tuberculosis*," *Journal of Clinical Pathology*, vol. 48, no. 10, pp. 924–928, 1995.
- [76] S. M. Wilson, S. Goss, and F. Drobniowski, "Evaluation of strategies for molecular fingerprinting for use in the routine work of a *Mycobacterium* reference unit," *Journal of Clinical Microbiology*, vol. 36, no. 11, pp. 3385–3388, 1998.
- [77] I. Otal, S. Samper, M. P. Asensio et al., "Use of a PCR method based on IS6110 polymorphism for typing *Mycobacterium tuberculosis* strains from BACTEC cultures," *Journal of Clinical Microbiology*, vol. 35, no. 1, pp. 273–277, 1997.
- [78] M. D. Yates, F. A. Drobniowski, and S. M. Wilson, "Evaluation of a rapid PCR-based epidemiological typing method for routine studies of *Mycobacterium tuberculosis*," *Journal of Clinical Microbiology*, vol. 40, no. 2, pp. 712–714, 2002.
- [79] D. van Soolingen, J. Bauer, V. Ritacco et al., "IS1245 restriction fragment length polymorphism typing of *Mycobacterium avium* isolates: proposal for standardization," *Journal of Clinical Microbiology*, vol. 36, no. 10, pp. 3051–3054, 1998.
- [80] N. Radomski, V. C. Thibault, C. Karoui et al., "Determination of genotypic diversity of *Mycobacterium avium* subspecies from human and animal origins by mycobacterial interspersed repetitive-unit-variable-number tandemrepeat and IS1311 restriction fragment length polymorphism typing methods," *Journal of Clinical Microbiology*, vol. 48, no. 4, pp. 1026–1034, 2010.
- [81] W. Mijs, P. de Haas, R. Rossau et al., "Molecular evidence to support a proposal to reserve the designation *Mycobacterium avium* subsp. *avium* for bird-type isolates and "*M. avium* sub sp. *hominissuis*" for the human/porcine type of *M. avium*," *International Journal of Systematic and Evolutionary Microbiology*, vol. 52, no. 5, pp. 1505–1518, 2002.
- [82] D. Whipple, P. Kapke, and C. Vary, "Identification of restriction fragment length polymorphisms in DNA from *Mycobacterium paratuberculosis*," *Journal of Clinical Microbiology*, vol. 28, no. 11, pp. 2561–2564, 1990.
- [83] Z. M. Kunze, F. Portaeals, and J. J. McFadden, "Biologically distinct subtypes of *Mycobacterium avium* differ in possession of insertion sequence IS901," *Journal of Clinical Microbiology*, vol. 30, no. 9, pp. 2366–2372, 1992.
- [84] M. T. Moss, Z. P. Malik, M. L. V. Tizard, E. P. Green, J. D. Sanderson, and J. Hermon-Taylor, "IS902, an insertion element of the chronic-enteritis-causing *Mycobacterium avium* sub sp. *silvaticum*," *Journal of General Microbiology*, vol. 138, no. 1, pp. 139–145, 1992.
- [85] M. H. Perez, N. G. Fomukong, T. Hellyer, I. N. Brown, and J. W. Dale, "Characterization of IS1110, a highly mobile genetic element from *Mycobacterium avium*," *Molecular Microbiology*, vol. 12, no. 5, pp. 717–724, 1994.
- [86] X. Puyang, K. Lee, C. Pawlichuk, and D. Y. Kunitomo, "IS1626, a new IS900-related *Mycobacterium avium* insertion sequence," *Microbiology*, vol. 145, no. 11, pp. 3163–3168, 1999.
- [87] F. J. Sangari, M. Bächli, L. E. Bermudez, and T. Bodmer, "Characterization of IS666, a newly described insertion element of *Mycobacterium avium*," *Microbial and Comparative Genomics*, vol. 5, no. 4, pp. 181–188, 2000.
- [88] M. Picardeau, T. J. Bull, G. Prod'hom, A. L. Pozniak, D. C. Shanson, and V. Vincent, "Comparison of a new insertion element, IS1407, with established molecular markers for the characterization of *Mycobacterium celatum*," *International Journal of Systematic Bacteriology*, vol. 47, no. 3, pp. 640–644, 1997.
- [89] M. Picardeau, A. Varnerot, J. Rauzier, B. Gicquel, and V. Vincent, "*Mycobacterium xenopi* IS1395, a novel insertion sequence expanding the IS256 family," *Microbiology*, vol. 142, no. 9, pp. 2453–2461, 1996.
- [90] M. Picardeau, T. J. Bull, and V. Vincent, "Identification and characterization of IS-like elements in *Mycobacterium gordonae*," *FEMS Microbiology Letters*, vol. 154, no. 1, pp. 95–102, 1997.
- [91] T. Stinear, B. C. Ross, J. K. Davies et al., "Identification and characterization of IS2404 and IS2606: two distinct repeated sequences for detection of *Mycobacterium ulcerans* by PCR," *Journal of Clinical Microbiology*, vol. 37, no. 4, pp. 1018–1023, 1999.
- [92] M. Picardeau, G. Prod'hom, L. Raskine, M. P. LePennec, and V. Vincent, "Genotypic characterization of five subspecies of *Mycobacterium kansasii*," *Journal of Clinical Microbiology*, vol. 35, no. 1, pp. 25–32, 1997.
- [93] C. Guilhot, B. Gicquel, J. Davies, and C. Martin, "Isolation and analysis of IS6120, a new insertion sequence from *Mycobacterium smegmatis*," *Molecular Microbiology*, vol. 6, no. 1, pp. 107–113, 1992.
- [94] P. W. Hermans, D. van Soolingen, E. M. Bik, P. E. W. de Haas, J. W. Dale, and J. D. A. van Embden, "Insertion element IS987 from *Mycobacterium bovis* BCG is located in a hot-spot integration region for insertion elements in *Mycobacterium tuberculosis* complex strains," *Infection and Immunity*, vol. 59, no. 8, pp. 2695–2705, 1991.
- [95] R. A. Skuce and S. D. Neill, "Molecular epidemiology of *Mycobacterium bovis*: exploiting molecular data," *Tuberculosis*, vol. 81, no. 1–2, pp. 169–175, 2001.

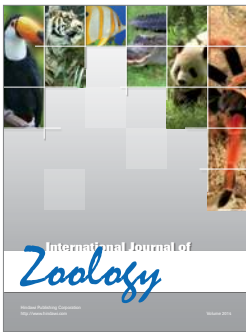
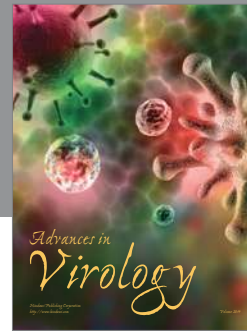
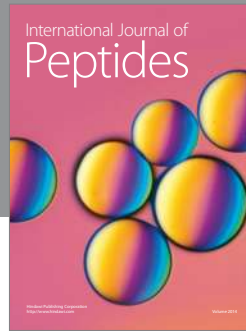
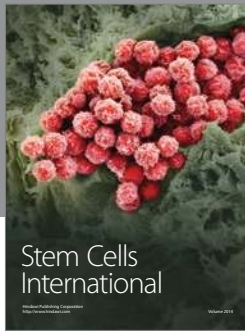
- [96] P. M. Groenen, A. E. Bunschoten, D. van Soolingen, and J. D. A. van Embden, "Nature of DNA polymorphism in the direct repeat cluster of *Mycobacterium tuberculosis*: application for strain differentiation by a novel typing method," *Molecular Microbiology*, vol. 10, no. 5, pp. 1057–1065, 1993.
- [97] J. D. van Embden, T. van Gorkom, K. Kremer, R. Jansen, B. A. M. van der Zeijst, and L. M. Schouls, "Genetic variation and evolutionary origin of the direct repeat locus of *Mycobacterium tuberculosis* complex bacteria," *Journal of Bacteriology*, vol. 182, no. 9, pp. 2393–2401, 2000.
- [98] S. Niemann, E. Richter, and S. Rüscher-Gerdes, "Stability of *Mycobacterium tuberculosis* IS6110 restriction fragment length polymorphism patterns and spoligotypes determined by analyzing serial isolates from patients with drug-resistant tuberculosis," *Journal of Clinical Microbiology*, vol. 37, no. 2, pp. 409–412, 1999.
- [99] S. Kulkarni, C. Sola, I. Filliol, N. Rastogi, and G. Kadival, "Spoligotyping of *Mycobacterium tuberculosis* isolates from patients with pulmonary tuberculosis in Mumbai, India," *Research in Microbiology*, vol. 156, no. 4, pp. 588–596, 2005.
- [100] J. R. Driscoll, M. A. McGarry, and H. W. Taber, "DNA typing of a nonviable culture of *Mycobacterium tuberculosis* in a homeless shelter outbreak," *Journal of Clinical Microbiology*, vol. 37, no. 1, pp. 274–275, 1999.
- [101] A. G. van der Zanden, A. H. Hoentjen, F. G. Heilmann, E. F. Weltevreden, L. M. Schouls, and J. D. A. van Embden, "Simultaneous detection and strain differentiation of *Mycobacterium tuberculosis* complex in paraffin wax embedded tissues and in stained microscopic preparations," *Journal of Clinical Pathology*, vol. 51, no. 4, pp. 209–214, 1998.
- [102] J. W. Dale, D. Brittain, A. A. Cataldi et al., "Spacer oligonucleotide typing of bacteria of the *Mycobacterium tuberculosis* complex: recommendations for standardised nomenclature," *International Journal of Tuberculosis and Lung Disease*, vol. 5, no. 3, pp. 216–219, 2001.
- [103] K. Brudey, J. R. Driscoll, L. Rigouts et al., "*Mycobacterium tuberculosis* complex genetic diversity: mining the fourth international spoligotyping database (SpolDB4) for classification, population genetics and epidemiology," *BMC Microbiology*, vol. 6, article 23, 2006.
- [104] C. Demay, B. Liens, T. Burguière et al., "SITVITWEB—a publicly available international multimarker database for studying *Mycobacterium tuberculosis* genetic diversity and molecular epidemiology," *Infection, Genetics and Evolution*, vol. 12, no. 4, pp. 755–766, 2012.
- [105] D. van Soolingen, "Molecular epidemiology of tuberculosis and other mycobacterial infections: main methodologies and achievements," *Journal of Internal Medicine*, vol. 249, no. 1, pp. 1–26, 2001.
- [106] R. Diaz, K. Kremer, P. E. de Haas et al., "Molecular epidemiology of tuberculosis in Cuba outside of Havana, July 1994–June 1995: utility of spoligotyping versus IS6110 restriction fragment length polymorphism," *International Journal of Tuberculosis and Lung Disease*, vol. 2, no. 9, pp. 743–750, 1998.
- [107] P. Farnia, F. Mohammadi, M. R. Masjedi et al., "Evaluation of tuberculosis transmission in Tehran: using RFLP and spoligotyping methods," *Journal of Infection*, vol. 49, no. 2, pp. 94–101, 2004.
- [108] Y.-O. G. de La Salmonière, H. M. Li, G. Torrea, A. Bunschoten, J. van Embden, and B. Gicquel, "Evaluation of spoligotyping in a study of the transmission of *Mycobacterium tuberculosis*," *Journal of Clinical Microbiology*, vol. 35, no. 9, pp. 2210–2214, 1997.
- [109] A. Gori, A. Bandera, G. Marchetti et al., "Spoligotyping and *Mycobacterium tuberculosis*," *Emerging Infectious Diseases*, vol. 11, no. 8, pp. 1242–1248, 2005.
- [110] A. Gori, A. D. Esposti, A. Bandera et al., "Comparison between spoligotyping and IS6110 restriction fragment length polymorphisms in molecular genotyping analysis of *Mycobacterium tuberculosis* strains," *Molecular and Cellular Probes*, vol. 19, no. 4, pp. 236–244, 2005.
- [111] K. Brudey, M. C. Gutierrez, V. Vincent et al., "*Mycobacterium africanum* genotyping using novel spacer oligonucleotides in the direct repeat locus," *Journal of Clinical Microbiology*, vol. 42, no. 11, pp. 5053–5057, 2004.
- [112] J. Zhang, E. Abadia, G. Refregier et al., "*Mycobacterium tuberculosis* complex CRISPR genotyping: improving efficiency, throughput and discriminative power of 'spoligotyping' with new spacers and a microbead-based hybridization assay," *Journal of Medical Microbiology*, vol. 59, no. 3, pp. 285–294, 2010.
- [113] W. A. Cronin, J. E. Golub, L. S. Magder et al., "Epidemiologic usefulness of spoligotyping for secondary typing of *Mycobacterium tuberculosis* isolates with low copy numbers of IS6110," *Journal of Clinical Microbiology*, vol. 39, no. 10, pp. 3709–3711, 2001.
- [114] L. S. Cowan, L. Diem, M. C. Brake, and J. T. Crawford, "Transfer of a *Mycobacterium tuberculosis* genotyping method, Spoligotyping, from a reverse line-blot hybridization, membrane-based assay to the Luminex multianalyte profiling system," *Journal of Clinical Microbiology*, vol. 42, no. 1, pp. 474–477, 2004.
- [115] C. Honisch, M. Mosko, C. Arnold, S. E. Gharbia, R. Diel, and S. Niemann, "Replacing reverse line blot hybridization spoligotyping of the *Mycobacterium tuberculosis* complex," *Journal of Clinical Microbiology*, vol. 48, no. 5, pp. 1520–1526, 2010.
- [116] P. W. Hermans, D. van Soolingen, and J. D. van Embden, "Characterization of a major polymorphic tandem repeat in *Mycobacterium tuberculosis* and its potential use in the epidemiology of *Mycobacterium kansasii* and *Mycobacterium goodii*," *Journal of Bacteriology*, vol. 174, no. 12, pp. 4157–4165, 1992.
- [117] C. Taillard, G. Greub, R. Weber et al., "Clinical implications of *Mycobacterium kansasii* species heterogeneity: Swiss national survey," *Journal of Clinical Microbiology*, vol. 41, no. 3, pp. 1240–1244, 2003.
- [118] S. T. Cole, "Comparative and functional genomics of the *Mycobacterium tuberculosis* complex," *Microbiology*, vol. 148, no. 10, pp. 2919–2928, 2002.
- [119] I. Filliol, S. Ferdinand, L. Negroni, C. Sola, and N. Rastogi, "Molecular typing of *Mycobacterium tuberculosis* based on variable number of tandem DNA repeats used alone and in association with spoligotyping," *Journal of Clinical Microbiology*, vol. 38, no. 7, pp. 2520–2524, 2000.
- [120] S. Roring, A. Scott, D. Brittain et al., "Development of variable-number tandem repeat typing of *Mycobacterium bovis*: comparison of results with those obtained by using existing exact tandem repeats and spoligotyping," *Journal of Clinical Microbiology*, vol. 40, no. 6, pp. 2126–2133, 2002.
- [121] P. Supply, J. Magdalena, S. Himpens, and C. Locht, "Identification of novel intergenic repetitive units in a mycobacterial two-component system operon," *Molecular Microbiology*, vol. 26, no. 5, pp. 991–1003, 1997.
- [122] P. Supply, E. Mazars, S. Lesjean, V. Vincent, B. Gicquel, and C. Locht, "Variable human minisatellite-like regions in the

- Mycobacterium tuberculosis* genome,” *Molecular Microbiology*, vol. 36, no. 3, pp. 762–771, 2000.
- [123] E. Mazars, S. Lesjean, A.-L. Banuls et al., “High-resolution minisatellite-based typing as a portable approach to global analysis of *Mycobacterium tuberculosis* molecular epidemiology,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 98, no. 4, pp. 1901–1906, 2001.
- [124] P. Supply, C. Allix, S. Lesjean et al., “Proposal for standardization of optimized mycobacterial interspersed repetitive unit-variable-number tandem repeat typing of *Mycobacterium tuberculosis*,” *Journal of Clinical Microbiology*, vol. 44, no. 12, pp. 4498–4510, 2006.
- [125] C. Allix-Béguet, D. Harmsen, T. Weniger, P. Supply, and S. Niemann, “Evaluation and strategy for use of MIRU-VNTRplus, a multifunctional database for online analysis of genotyping data and phylogenetic identification of *Mycobacterium tuberculosis* complex isolates,” *Journal of Clinical Microbiology*, vol. 46, no. 8, pp. 2692–2699, 2008.
- [126] T. Weniger, J. Krawczyk, P. Supply, S. Niemann, and D. Harmsen, “MIRU-VNTRplus: a web tool for polyphasic genotyping of *Mycobacterium tuberculosis* complex bacteria,” *Nucleic Acids Research*, vol. 38, supplement 2, pp. W326–W331, 2010.
- [127] V. Hill, T. Zozio, S. Sadikalay et al., “MLVA based classification of *Mycobacterium tuberculosis* complex lineages for a robust phylogeographic snapshot of its worldwide molecular diversity,” *PLoS ONE*, vol. 7, no. 9, Article ID e41991, 2012.
- [128] K. S. Blackwood, J. N. Wolfe, and A. M. Kabani, “Application of mycobacterial interspersed repetitive unit typing to *Manitoba tuberculosis* cases: can restriction fragment length polymorphism be forgotten?” *Journal of Clinical Microbiology*, vol. 42, no. 11, pp. 5001–5006, 2004.
- [129] P. M. Hawkey, E. G. Smith, J. T. Evans et al., “Mycobacterial interspersed repetitive unit typing of *Mycobacterium tuberculosis* compared to IS6110-based restriction fragment length polymorphism analysis for investigation of apparently clustered cases of tuberculosis,” *Journal of Clinical Microbiology*, vol. 41, no. 8, pp. 3514–3520, 2003.
- [130] L. S. Cowan, L. Mosher, L. Diem, J. P. Massey, and J. T. Crawford, “Variable-number tandem repeat typing of *Mycobacterium tuberculosis* isolates with low copy numbers of IS6110 by using mycobacterial interspersed repetitive units,” *Journal of Clinical Microbiology*, vol. 40, no. 5, pp. 1592–1602, 2002.
- [131] A. S. Lee, L. L. Tang, I. H. Lim, R. Bellamy, and S.-Y. Wong, “Discrimination of single-copy IS6110 DNA fingerprints of *Mycobacterium tuberculosis* isolates by high-resolution minisatellite-based typing,” *Journal of Clinical Microbiology*, vol. 40, no. 2, pp. 657–659, 2002.
- [132] L. S. Cowan, L. Diem, T. Monson et al., “Evaluation of a two-step approach for large-scale, prospective genotyping of *Mycobacterium tuberculosis* isolates in the United States,” *Journal of Clinical Microbiology*, vol. 43, no. 2, pp. 688–695, 2005.
- [133] A. N. Scott, D. Menzies, T.-N. Tannenbaum et al., “Sensitivities and specificities of spoligotyping and mycobacterial interspersed repetitive unit-variable-number tandem repeat typing methods for studying molecular epidemiology of tuberculosis,” *Journal of Clinical Microbiology*, vol. 43, no. 1, pp. 89–94, 2005.
- [134] N. Alonso-Rodríguez, M. Martínez-Lirola, M. Herránz et al., “Evaluation of the new advanced 15-loci MIRU-VNTR genotyping tool in *Mycobacterium tuberculosis* molecular epidemiology studies,” *BMC Microbiology*, vol. 8, article 34, 2008.
- [135] M. C. Oelemann, R. Diel, V. Vatin et al., “Assessment of an optimized mycobacterial interspersed repetitive-unit-variable-number tandem-repeat typing system combined with spoligotyping for population-based molecular epidemiology studies of tuberculosis,” *Journal of Clinical Microbiology*, vol. 45, no. 3, pp. 691–697, 2007.
- [136] J. L. de Beer, K. Kremer, C. Kodmon et al., “First worldwide proficiency study on variable-number tandem-repeat typing of *Mycobacterium tuberculosis* complex strains,” *Journal of Clinical Microbiology*, vol. 50, no. 3, pp. 662–669, 2012.
- [137] V. C. Thibault, M. Grayon, M. L. Boschirola et al., “New variable-number tandem-repeat markers for typing *Mycobacterium avium* subsp. paratuberculosis and *M. avium* strains: comparison with IS900 and IS1245 restriction fragment length polymorphism typing,” *Journal of Clinical Microbiology*, vol. 45, no. 8, pp. 2404–2410, 2007.
- [138] T. Inagaki, K. Nishimori, T. Yagi et al., “Comparison of a variable-number tandem-repeat (VNTR) method for typing *Mycobacterium avium* with mycobacterial interspersed repetitive-unit-VNTR and IS1245 restriction fragment length polymorphism typing,” *Journal of Clinical Microbiology*, vol. 47, no. 7, pp. 2156–2164, 2009.
- [139] F.-A. Dauchy, S. Dégrange, A. Charron et al., “Variable-number tandem-repeat markers for typing *Mycobacterium intracellulare* strains isolated in humans,” *BMC Microbiology*, vol. 10, article 93, 2010.
- [140] C. J. Lavender, T. P. Stinear, P. D. Johnson et al., “Evaluation of VNTR typing for the identification of *Mycobacterium ulcerans* in environmental samples from Victoria, Australia,” *FEMS Microbiology Letters*, vol. 287, no. 2, pp. 250–255, 2008.
- [141] G. Sun, C. Chen, J. Li et al., “Discriminatory potential of a novel set of variable number of tandem repeats for genotyping *Mycobacterium marinum*,” *Veterinary Microbiology*, vol. 152, no. 1–2, pp. 200–204, 2011.
- [142] B. C. Ross, K. Raios, K. Jackson, and B. Dwyer, “Molecular cloning of a highly repeated DNA element from *Mycobacterium tuberculosis* and its use as an epidemiological tool,” *Journal of Clinical Microbiology*, vol. 30, no. 4, pp. 942–946, 1992.
- [143] S. Poulet and S. T. Cole, “Characterization of the highly abundant polymorphic GC-rich-repetitive sequence (PGRS) present in *Mycobacterium tuberculosis*,” *Archives of Microbiology*, vol. 163, no. 2, pp. 87–95, 1995.
- [144] F. Chaves, Z. Yang, H. E. L. Hajj et al., “Usefulness of the secondary probe pTBN12 in DNA fingerprinting of *Mycobacterium tuberculosis*,” *Journal of Clinical Microbiology*, vol. 34, no. 5, pp. 1118–1123, 1996.
- [145] Z. Yang, F. Chaves, P. F. Barnes et al., “Evaluation of method for secondary DNA typing of *Mycobacterium tuberculosis* with pTBN12 in epidemiologic study of tuberculosis,” *Journal of Clinical Microbiology*, vol. 34, no. 12, pp. 3044–3048, 1996.
- [146] A. Dela, A. Sajduda, I. Pawłowska, and J. Dziadek, “Molecular characterization of *Mycobacterium tuberculosis* isolates from Łódź, Poland: analysis by IS6110 restriction fragment length polymorphism and double-repetitive-element PCR,” *Journal of Infection*, vol. 52, no. 5, pp. 346–353, 2006.
- [147] E. Montoro, J. Valdivia, and S. C. Leão, “Molecular fingerprinting of *Mycobacterium tuberculosis* isolates obtained in Havana, Cuba, by IS6110 restriction fragment length polymorphism analysis and by the double-repetitive-element PCR method,” *Journal of Clinical Microbiology*, vol. 36, no. 10, pp. 3099–3102, 1998.

- [148] A. Sajduda, J. Dziadek, R. Kotłowski, and F. Portaels, "Evaluation of multiple genetic markers for typing drug-resistant *Mycobacterium tuberculosis* strains from Poland," *Diagnostic Microbiology and Infectious Disease*, vol. 55, no. 1, pp. 59–64, 2006.
- [149] T. Jagielski, E. Augustynowicz-Kopec, K. Pawlik, J. Dziadek, Z. Zwolska, and J. Bielecki, "A two-step strategy for molecular typing of multidrug-resistant *Mycobacterium tuberculosis* clinical isolates from Poland," *Polish Journal of Microbiology*, vol. 60, no. 3, pp. 229–232, 2011.
- [150] B. C. Ross, K. Jackson, M. Yang, A. Sievers, and B. Dwyer, "Identification of a genetically distinct subspecies of *Mycobacterium kansasii*," *Journal of Clinical Microbiology*, vol. 30, no. 11, pp. 2930–2933, 1992.
- [151] K. Jackson, R. Edwards, D. E. Leslie, and J. Hayman, "Molecular method for typing *Mycobacterium ulcerans*," *Journal of Clinical Microbiology*, vol. 33, no. 9, pp. 2250–2253, 1995.
- [152] L. A. Sechi, S. Zanetti, I. Dupré, G. Delogu, and G. Fadda, "Enterobacterial repetitive intergenic consensus sequences as molecular targets for typing of *Mycobacterium tuberculosis* strains," *Journal of Clinical Microbiology*, vol. 36, no. 1, pp. 128–132, 1998.
- [153] L. A. Sechi, S. Zanetti, G. Delogu, B. Montinaro, A. Sanna, and G. Fadda, "Molecular epidemiology of *Mycobacterium tuberculosis* strains isolated from different regions of Italy and Pakistan," *Journal of Clinical Microbiology*, vol. 34, no. 7, pp. 1825–1828, 1996.
- [154] J. L. M. Sampaio, C. Viana-Niero, D. de Freitas, A. L. Höfling-Lima, and S. C. Leão, "Enterobacterial repetitive intergenic consensus PCR is a useful tool for typing *Mycobacterium chelonae* and *Mycobacterium abscessus* isolates," *Diagnostic Microbiology and Infectious Disease*, vol. 55, no. 2, pp. 107–118, 2006.
- [155] A. Wojtasik, M. Majchrzak, W. Adamus-Bialek et al., "Trinucleotide repeat sequence-based PCR as a potential approach for genotyping *Mycobacterium gordonae* strains," *Journal of Microbiological Methods*, vol. 85, no. 1, pp. 28–32, 2011.
- [156] G. A. Cangelosi, R. J. Freeman, K. N. Lewis et al., "Evaluation of a high-throughput repetitive-sequence-based PCR system for DNA fingerprinting of *Mycobacterium tuberculosis* and *Mycobacterium avium* complex strains," *Journal of Clinical Microbiology*, vol. 42, no. 6, pp. 2685–2693, 2004.
- [157] M. H. Jang, G. E. Choi, B.-M. Shin et al., "Comparison of an automated repetitive sequence-based pcr microbial typing system with IS6110-restriction fragment length polymorphism for epidemiologic investigation of clinical *Mycobacterium tuberculosis* isolates in Korea," *Korean Journal of Laboratory Medicine*, vol. 31, no. 4, pp. 282–284, 2011.
- [158] A. M. Zelazny, J. M. Root, Y. R. Shea et al., "Cohort study of molecular identification and typing of *Mycobacterium abscessus*, *Mycobacterium massiliense*, and *Mycobacterium bolletii*," *Journal of Clinical Microbiology*, vol. 47, no. 7, pp. 1985–1995, 2009.
- [159] P. H. S. de Groot, J. van Ingen, R. de Zwaan, A. Mulder, M. J. Boeree, and D. van Soolingen, "Disseminated *Mycobacterium avium* subsp. *avium* infection in a cat, the Netherlands," *Veterinary Microbiology*, vol. 144, no. 3-4, pp. 527–529, 2010.
- [160] R. J. C. van der Zanden, C. Magis-Escurra, W. C. M. de Lange et al., "Hypersensitivity pneumonitis caused by *Mycobacterium avium* subsp. *hominissuis* in a hot tub, as proven by IS1245 RFLP and rep-PCR typing," *International Journal of Mycobacteriology*, vol. 1, no. 3, pp. 152–154, 2012.
- [161] J. G. Rodriguez, G. A. Mejia, P. del Portillo, M. E. Patarroyo, and L. A. Murillo, "Species-specific identification of *Mycobacterium bovis* by PCR," *Microbiology*, vol. 141, no. 9, pp. 2131–2138, 1995.
- [162] C. J. Linton, H. Jalal, J. P. Leeming, and M. R. Millar, "Rapid discrimination of *Mycobacterium tuberculosis* strains by random amplified polymorphic DNA analysis," *Journal of Clinical Microbiology*, vol. 32, no. 9, pp. 2169–2174, 1994.
- [163] C. J. Linton, A. D. Smart, J. P. Leeming et al., "Comparison of random amplified polymorphic DNA with restriction fragment length polymorphism as epidemiological typing methods for *Mycobacterium tuberculosis*," *Journal of Clinical Pathology*, vol. 48, no. 3, pp. M133–M135, 1995.
- [164] P. Palittapongarnpim, S. Chomyc, A. Fanning, and D. Kunitomo, "DNA fragment length polymorphism analysis of *Mycobacterium tuberculosis* isolates by arbitrarily primed polymerase chain reaction," *Journal of Infectious Diseases*, vol. 167, no. 4, pp. 975–978, 1993.
- [165] Y. Zhang, M. Rajagopalan, B. A. Brown, and R. J. Wallace Jr., "Randomly amplified polymorphic DNA PCR for comparison of *Mycobacterium abscessus* strains from nosocomial outbreaks," *Journal of Clinical Microbiology*, vol. 35, no. 12, pp. 3132–3139, 1997.
- [166] R. C. Cooksey, M. A. Jhung, M. A. Yakrus et al., "Multiphasic approach reveals genetic diversity of environmental and patient isolates of *Mycobacterium mucogenicum* and *Mycobacterium phocaicum* associated with an outbreak of bacteremias at a Texas hospital," *Applied and Environmental Microbiology*, vol. 74, no. 8, pp. 2480–2487, 2008.
- [167] E. Vogiatzakis, S. Stefanou, A. Skroubelou, S. Anagnostou, E. Marinis, and P. Matsiota-Bernard, "Molecular markers for the investigation of *Mycobacterium gordonae* epidemics," *Journal of Hospital Infection*, vol. 38, no. 3, pp. 217–222, 1998.
- [168] J. van Ingen, M. J. Boeree, W. C. M. de Lange, P. E. de Haas, P. N. Dekhuijzen, and D. van Soolingen, "Clinical relevance of *Mycobacterium szulgai* in The Netherlands," *Clinical Infectious Diseases*, vol. 46, no. 8, pp. 1200–1205, 2008.
- [169] J. Kauppinen, R. Mantjarvi, and M.-L. Katila, "Random amplified polymorphic DNA genotyping of *Mycobacterium malmoense*," *Journal of Clinical Microbiology*, vol. 32, no. 7, pp. 1827–1829, 1994.
- [170] Y. Abed, A. Davin-Regli, C. Bollet, and P. de Micco, "Efficient discrimination of *Mycobacterium tuberculosis* strains by 16S-23S spacer region-based random amplified polymorphic DNA analysis," *Journal of Clinical Microbiology*, vol. 33, no. 5, pp. 1418–1420, 1995.
- [171] R. Frothingham, Y. Abed, and C. Bollet, "Discrimination of *Mycobacterium tuberculosis* strains by PCR," *Journal of Clinical Microbiology*, vol. 33, no. 10, pp. 2801–2802, 1995.
- [172] M. Glennon, T. Smith, Y. Abed, and C. Bollet, "Can random amplified polymorphic DNA analysis of the 16S-23S spacer region of *Mycobacterium tuberculosis* differentiate between isolates?" *Journal of Clinical Microbiology*, vol. 33, no. 12, pp. 3359–3360, 1995.
- [173] P. Janssen, R. Coopman, G. Huys et al., "Evaluation of the DNA fingerprinting method AFLP as a new tool in bacterial taxonomy," *Microbiology*, vol. 142, no. 7, pp. 1881–1893, 1996.
- [174] G. Huys, L. Rigouts, K. Chemlal, F. Portaels, and J. Swings, "Evaluation of amplified fragment length polymorphism analysis for inter- and intraspecific differentiation of *Mycobacterium bovis*, *M. tuberculosis*, and *M. ulcerans*," *Journal of Clinical Microbiology*, vol. 38, no. 10, pp. 3675–3680, 2000.

- [175] M. Y. Krishnan, I. Radhakrishnan, B. V. Joseph, L. G. K. Madhavi, K. R. Ajay, and S. Mundayoor, "Combined use of amplified fragment length polymorphism and IS6110-RFLP in fingerprinting clinical isolates of *Mycobacterium tuberculosis* from Kerala, South India," *BMC Infectious Diseases*, vol. 7, article 86, 2007.
- [176] J. N. Goulding, J. Stanley, N. Saunders, and C. Arnold, "Genome-sequence-based fluorescent amplified-fragment length polymorphism analysis of *Mycobacterium tuberculosis*," *Journal of Clinical Microbiology*, vol. 38, no. 3, pp. 1121–1126, 2000.
- [177] E. J. Sims, M. Goyal, and C. Arnold, "Experimental versus in silico fluorescent amplified fragment length polymorphism analysis of *Mycobacterium tuberculosis*: improved typing with an extended fragment range," *Journal of Clinical Microbiology*, vol. 40, no. 11, pp. 4072–4076, 2002.
- [178] S. L. Pfaller, T. W. Aronson, A. E. Holtzman, and T. C. Covert, "Amplified fragment length polymorphism analysis of *Mycobacterium avium* complex isolates recovered from southern California," *Journal of Medical Microbiology*, vol. 56, no. 9, pp. 1152–1160, 2007.
- [179] L. E. B. van Coppenraet, P. H. M. Savelkoul, N. Buffing et al., "Amplified fragment length polymorphism analysis of human clinical isolates of *Mycobacterium haemophilum* from different continents," *Clinical Microbiology and Infection*, vol. 15, no. 10, pp. 924–930, 2009.
- [180] K. Chemlal, G. Huys, P.-A. Fonteyne et al., "Evaluation of PCR-restriction profile analysis and IS2404 restriction fragment length polymorphism and amplified fragment length polymorphism fingerprinting for identification and typing of *Mycobacterium ulcerans* and *M. marinum*," *Journal of Clinical Microbiology*, vol. 39, no. 9, pp. 3272–3278, 2001.
- [181] K. Chemlal, G. Huys, F. Laval et al., "Characterization of an unusual mycobacterium: a possible missing link between *Mycobacterium marinum* and *Mycobacterium ulcerans*," *Journal of Clinical Microbiology*, vol. 40, no. 7, pp. 2370–2380, 2002.
- [182] M. Achtman, "Evolution, population structure, and phylogeography of genetically monomorphic bacterial pathogens," *Annual Review of Microbiology*, vol. 62, pp. 53–70, 2008.
- [183] C. F. Wasem, C. M. McCarthy, and L. W. Murray, "Multilocus enzyme electrophoresis analysis of the *Mycobacterium avium* complex and other mycobacteria," *Journal of Clinical Microbiology*, vol. 29, no. 2, pp. 264–271, 1991.
- [184] M. A. Yakrus, M. W. Reeves, and S. B. Hunter, "Characterization of isolates of *Mycobacterium avium* serotypes 4 and 8 from patients with AIDS by multilocus enzyme electrophoresis," *Journal of Clinical Microbiology*, vol. 30, no. 6, pp. 1474–1478, 1992.
- [185] M. M. Feizabadi, I. D. Robertson, D. V. Cousins, D. J. Dawson, and D. J. Hampson, "Use of multilocus enzyme electrophoresis to examine genetic relationships amongst isolates of *Mycobacterium intracellulare* and related species," *Microbiology*, vol. 143, no. 4, pp. 1461–1469, 1997.
- [186] C. Y. Turenne, D. M. Collins, D. C. Alexander, and M. A. Behr, "*Mycobacterium avium* subsp. paratuberculosis and *M. avium* subsp. *avium* are independently evolved pathogenic clones of a much broader group of *M. avium* organisms," *Journal of Bacteriology*, vol. 190, no. 7, pp. 2479–2487, 2008.
- [187] S. Ramaswamy and J. M. Musser, "Molecular genetic basis of antimicrobial agent resistance in *Mycobacterium tuberculosis*: 1998 update," *Tubercle and Lung Disease*, vol. 79, no. 1, pp. 3–29, 1998.
- [188] L. Baker, T. Brown, M. C. Maiden, and F. Drobniowski, "Silent nucleotide polymorphisms and a phylogeny for *Mycobacterium tuberculosis*," *Emerging Infectious Diseases*, vol. 10, no. 9, pp. 1568–1577, 2004.
- [189] M. M. Gutacker, B. Mathema, H. Soini et al., "Single-nucleotide polymorphism-based population genetic analysis of *Mycobacterium tuberculosis* strains from 4 geographic sites," *Journal of Infectious Diseases*, vol. 193, no. 1, pp. 121–128, 2006.
- [190] T. Dos Vultos, J. Blázquez, J. Rauzier, I. Matic, and B. Gicquel, "Identification of nudix hydrolase family members with an antimutator role in *Mycobacterium tuberculosis* and *Mycobacterium smegmatis*," *Journal of Bacteriology*, vol. 188, no. 8, pp. 3159–3161, 2006.
- [191] R. D. Fleischmann, D. Alland, J. A. Eisen et al., "Whole-genome comparison of *Mycobacterium tuberculosis* clinical and laboratory strains," *Journal of Bacteriology*, vol. 184, no. 19, pp. 5479–5490, 2002.
- [192] A. G. Tsolaki, A. E. Hirsh, K. DeRiemer et al., "Functional and evolutionary genomics of *Mycobacterium tuberculosis*: insights from genomic deletions in 100 strains," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 101, no. 14, pp. 4865–4870, 2004.
- [193] M. Kato-Maeda, J. T. Rhee, T. R. Gingeras et al., "Comparing genomes within the species *Mycobacterium tuberculosis*," *Genome Research*, vol. 11, no. 4, pp. 547–554, 2001.
- [194] R. Brosch, S. V. Gordon, M. Marmiesse et al., "A new evolutionary scenario for the *Mycobacterium tuberculosis* complex," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 99, no. 6, pp. 3684–3689, 2002.
- [195] D. Alland, D. W. Lacher, M. H. Hazbón et al., "Role of large sequence polymorphisms (LSPs) in generating genomic diversity among clinical isolates of *Mycobacterium tuberculosis* and the utility of LSPs in phylogenetic analysis," *Journal of Clinical Microbiology*, vol. 45, no. 1, pp. 39–46, 2007.
- [196] A. G. Tsolaki, S. Gagneux, A. S. Pym et al., "Genomic deletions classify the Beijing/W strains as a distinct genetic lineage of *Mycobacterium tuberculosis*," *Journal of Clinical Microbiology*, vol. 43, no. 7, pp. 3185–3191, 2005.
- [197] Y.-O. L. Goguet de la Salmonière, C. C. Kim, A. G. Tsolaki, A. S. Pym, M. S. Siegrist, and P. M. Small, "High-throughput method for detecting genomic-deletion polymorphisms," *Journal of Clinical Microbiology*, vol. 42, no. 7, pp. 2913–2918, 2004.
- [198] M. Cardoso Oelemann, H. M. Gomes, E. Willery et al., "The forest behind the tree: phylogenetic exploration of a dominant *Mycobacterium tuberculosis* strain lineage from a high tuberculosis burden country," *PLoS ONE*, vol. 6, no. 3, Article ID e18256, 2011.
- [199] R. M. Warren, G. D. van der Spuy, M. Richardson et al., "Calculation of the stability of the IS6110 banding pattern in patients with persistent *Mycobacterium tuberculosis* disease," *Journal of Clinical Microbiology*, vol. 40, no. 5, pp. 1705–1708, 2002.
- [200] M. D. Cave, Z. H. Yang, R. Stefanova et al., "Epidemiologic import of tuberculosis cases whose isolates have similar but not identical IS6110 restriction fragment length polymorphism patterns," *Journal of Clinical Microbiology*, vol. 43, no. 3, pp. 1228–1233, 2005.
- [201] K. Ijaz, Z. Yang, H. S. Matthews, J. H. Bates, and M. D. Cave, "*Mycobacterium tuberculosis* transmission between cluster members with similar fingerprint patterns," *Emerging Infectious Diseases*, vol. 8, no. 11, pp. 1257–1259, 2002.

- [202] S. A. Al-Hajj, O. Akkerman, I. Parwati et al., "Microevolution of *Mycobacterium tuberculosis* in a tuberculosis patient," *Journal of Clinical Microbiology*, vol. 48, no. 10, pp. 3813–3816, 2010.
- [203] E. Savine, R. M. Warren, G. D. van der Spuy et al., "Stability of variable-number tandem repeats of mycobacterial interspersed repetitive units from 12 loci in serial isolates of *Mycobacterium tuberculosis*," *Journal of Clinical Microbiology*, vol. 40, no. 12, pp. 4561–4566, 2002.
- [204] G. D. van der Spuy, R. M. Warren, M. Richardson, N. Beyers, M. A. Behr, and P. D. van Helden, "Use of genetic distance as a measure of ongoing transmission of *Mycobacterium tuberculosis*," *Journal of Clinical Microbiology*, vol. 41, no. 12, pp. 5640–5644, 2003.
- [205] R. Diel, S. Schneider, K. Meywald-Walter, C.-M. Ruf, S. Rüscher, and S. Niemann, "Epidemiology of tuberculosis in Hamburg, Germany: long-term population-based analysis applying classical and molecular epidemiological techniques," *Journal of Clinical Microbiology*, vol. 40, no. 2, pp. 532–539, 2002.
- [206] T. N. Quitugua, B. J. Seaworth, S. E. Weis et al., "Transmission of drug-resistant tuberculosis in Texas and Mexico," *Journal of Clinical Microbiology*, vol. 40, no. 8, pp. 2716–2724, 2002.
- [207] N. Alonso-Rodriguez, M. Martínez-Lirola, M. L. Sánchez et al., "Prospective universal application of mycobacterial interspersed repetitive-unit-variable-number tandem-repeat genotyping to characterize *Mycobacterium tuberculosis* isolates for fast identification of clustered and orphan cases," *Journal of Clinical Microbiology*, vol. 47, no. 7, pp. 2026–2032, 2009.
- [208] T. N. Buu, D. van Soolingen, M. N. T. Huyen et al., "Tuberculosis acquired outside of households, rural Vietnam," *Emerging Infectious Diseases*, vol. 16, no. 9, pp. 1466–1468, 2010.
- [209] J. R. Glynn, J. Bauer, A. S. de Boer et al., "Interpreting DNA fingerprint clusters of *Mycobacterium tuberculosis*," *International Journal of Tuberculosis and Lung Disease*, vol. 3, no. 12, pp. 1055–1060, 1999.
- [210] A. Seidler, A. Nienhaus, and R. Diel, "The transmission of tuberculosis in the light of new molecular biological approaches," *Occupational and Environmental Medicine*, vol. 61, no. 2, pp. 96–102, 2004.
- [211] S. Niemann, C. U. Köser, S. Gagneux et al., "Genomic diversity among drug sensitive and multidrug resistant isolates of *Mycobacterium tuberculosis* with identical DNA fingerprints," *PLoS ONE*, vol. 4, no. 10, Article ID e7407, 2009.
- [212] A. C. Schürch and D. van Soolingen, "DNA fingerprinting of *Mycobacterium tuberculosis*: from phage typing to whole-genome sequencing," *Infection, Genetics and Evolution*, vol. 12, no. 4, pp. 602–609, 2012.
- [213] B. R. Edlin, J. I. Tokars, M. H. Grieco et al., "An outbreak of multidrug-resistant tuberculosis among hospitalized patients with the acquired immunodeficiency syndrome," *The New England Journal of Medicine*, vol. 326, no. 23, pp. 1514–1521, 1992.
- [214] P. D. McElroy, T. R. Sterling, C. R. Driver et al., "Use of DNA fingerprinting to investigate a multiyear, multistate tuberculosis outbreak," *Emerging Infectious Diseases*, vol. 8, no. 11, pp. 1252–1256, 2002.
- [215] A. van Rie, T. C. Victor, M. Richardson et al., "Reinfection and mixed infection cause changing *Mycobacterium tuberculosis* drug-resistance patterns," *The American Journal of Respiratory and Critical Care Medicine*, vol. 172, no. 5, pp. 636–642, 2005.
- [216] J. A. Caminero, M. J. Pena, M. I. Campos-Herrero et al., "Exogenous reinfection with tuberculosis on a European Island with a moderate incidence of disease," *The American Journal of Respiratory and Critical Care Medicine*, vol. 163, no. 3, pp. 717–720, 2001.
- [217] M.-L. Lambert, E. Hasker, A. van Deun, D. Roberfroid, M. Boelaert, and P. van der Stuyft, "Recurrence in tuberculosis: relapse or reinfection?" *The Lancet Infectious Diseases*, vol. 3, no. 5, pp. 282–287, 2003.
- [218] K. Mallard, R. McNerney, A. C. Crampin et al., "Molecular detection of mixed infections of *Mycobacterium tuberculosis* strains in sputum samples from patients in Karonga District, Malawi," *Journal of Clinical Microbiology*, vol. 48, no. 12, pp. 4512–4518, 2010.
- [219] P. Suffys, A. da Silva Rocha, A. Brandão et al., "Detection of mixed infections with *Mycobacterium lentiflavum* and *Mycobacterium avium* by molecular genotyping methods," *Journal of Medical Microbiology*, vol. 55, no. 1, pp. 127–131, 2006.
- [220] M. D. C. Ramos, H. Soini, G. C. Roscanni, M. Jaques, M. C. Villares, and J. M. Musser, "Extensive cross-contamination of specimens with *Mycobacterium tuberculosis* in a reference laboratory," *Journal of Clinical Microbiology*, vol. 37, no. 4, pp. 916–919, 1999.
- [221] L. Fitzpatrick, C. Braden, W. Cronin et al., "Investigation of Laboratory cross-contamination of *Mycobacterium tuberculosis* cultures," *Clinical Infectious Diseases*, vol. 38, no. 6, pp. e52–54, 2004.
- [222] J.-J. Yan, R. Jou, W.-C. Ko, J.-J. Wu, M.-L. Yang, and H.-M. Chen, "The use of variable-number tandem-repeat mycobacterial interspersed repetitive unit typing to identify laboratory cross-contamination with *Mycobacterium tuberculosis*," *Diagnostic Microbiology and Infectious Disease*, vol. 52, no. 1, pp. 21–28, 2005.
- [223] D. B. Blossom, K. A. Alelis, D. C. Chang et al., "Pseudo-outbreak of *Mycobacterium abscessus* infection caused by laboratory contamination," *Infection Control and Hospital Epidemiology*, vol. 29, no. 1, pp. 57–62, 2008.
- [224] L. Nabyonga, D. P. Kateete, F. A. Katabazi et al., "Determination of circulating *Mycobacterium tuberculosis* strains and transmission patterns among pulmonary TB patients in Kawempe municipality, Uganda, using MIRU-VNTR," *BMC Research Notes*, vol. 4, article 280, 2011.
- [225] S. Mandal, L. Bradshaw, L. F. Anderson et al., "Investigating transmission of *Mycobacterium bovis* in the United Kingdom in 2005 to 2008," *Journal of Clinical Microbiology*, vol. 49, no. 5, pp. 1943–1950, 2011.
- [226] J. Mutsvangwa, K. A. Millington, K. Chaka et al., "Identifying recent *Mycobacterium tuberculosis* transmission in the setting of high HIV and TB burden," *Thorax*, vol. 65, no. 4, pp. 315–320, 2010.
- [227] E. Augustynowicz-Kopeć, T. Jagielski, M. Kozińska et al., "Transmission of tuberculosis within family-households," *Journal of Infection*, vol. 64, no. 6, pp. 596–608, 2012.



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