



## REVIEW ARTICLE

# Cyclo-oxygenase-2: pharmacology, physiology, biochemistry and relevance to NSAID therapy

\*<sup>1</sup>Jane A. Mitchell and <sup>2</sup>Timothy D. Warner

<sup>1</sup>Unit of Critical Care Medicine, Royal Brompton Hospital, Imperial College School of Medicine, Sydney Street, London SW3 6NP and <sup>2</sup>Vascular Inflammation, The William Harvey Research Institute, St Bartholomew's and the Royal London School of Medicine and Dentistry, Charterhouse Square, London EC1M 6BQ

Cyclo-oxygenase is expressed in cells in two distinct isoforms. Cyclo-oxygenase-1 is present constitutively whilst cyclo-oxygenase-2 is expressed primarily after inflammatory insult. The activity of cyclo-oxygenase-1 and -2 results in the production of a variety of potent biological mediators (the prostaglandins) that regulate homeostatic and disease processes. Inhibitors of cyclo-oxygenase include the nonsteroidal anti-inflammatory drugs (NSAIDs) aspirin, ibuprofen and diclofenac. NSAIDs inhibit cyclo-oxygenase-2 at the site of inflammation, to produce their therapeutic benefits, as well as cyclo-oxygenase-1 in the gastric mucosa, which produces gastric damage. Most recently selective inhibitors of cyclo-oxygenase-2 have been developed and introduced to man for the treatment of arthritis. Moreover, recent epidemiological evidence suggests that cyclo-oxygenase inhibitors may have important therapeutic relevance in the prevention of some cancers or even Alzheimer's disease. This review will discuss how the new advancements in NSAIDs research has led to the development of a new class of NSAIDs that has far reaching implications for the treatment of disease.

**Keywords:** Cyclo-oxygenase-2; nonsteroidal anti-inflammatory drugs; rofecoxib; celecoxib; inflammation; prostaglandins

**Abbreviations:** DFP, (3-(2-propyloxy)-4-(4-methylsulphonylphenyl)-5,5-dimethylfuranone); DFU, dimethyl-3-(3-fluorophenyl)-4-(4-methylsulphonylphenyl)-2(5H)-furanone; EPA, eicosapentaenoic acid; NSAID, nonsteroidal anti-inflammatory drugs; PPAR, peroxisome proliferator-activated receptor; PG, prostaglandin; TX, thromboxane

## Introduction

Cyclo-oxygenase is the first enzyme in the formation of prostaglandins (PG) and thromboxane (TX) from arachidonic acid. Cyclo-oxygenase metabolites have a wide variety of physiological and pathophysiological effects and are involved in a number of homeostatic processes. However, it is the role of these metabolites in cardiovascular homeostasis and inflammatory oedema and pain which has made cyclo-oxygenase a therapeutic target that potentially affects the majority of individuals at some time in their lives.

The nonsteroidal anti-inflammatory drugs (NSAIDs), which include aspirin, owe their therapeutic and side effects to inhibition of cyclo-oxygenase. Up until the early 1990s the beneficial and the deleterious effects of NSAIDs were thought to be caused by inhibition of a single cyclo-oxygenase enzyme; inhibition of cyclo-oxygenase at inflammatory sites explaining their therapeutic actions and inhibition of cyclo-oxygenase in the gastric mucosa explaining their gastrototoxic effects. However, we now know that two forms of cyclo-oxygenase exist. A constitutive form, cyclo-oxygenase-1, and an inducible form, cyclo-oxygenase-2; the latter form being preferentially expressed at sites of inflammation. Thus, a new hypothesis has been suggested to explain the effects of NSAIDs; 'inhibition of cyclo-oxygenase-1 accounts for the side effects whilst inhibition of cyclo-oxygenase-2 accounts for the therapeutic benefits of NSAIDs'. This review will discuss the respective similarities and differences in the biology of the two forms of cyclo-oxygenase. It will also pay particular attention to how recent

discoveries relating to the functioning of cyclo-oxygenase-2 has led to the development of new and potentially better NSAIDs.

## Cyclo-oxygenase isoforms

### *Cyclo-oxygenase-1; the constitutive isoform*

Cyclo-oxygenase-1 is expressed in most mammalian cells under physiological conditions (Seibert *et al.*, 1997). However endothelial cells, platelets and kidney tubule cells are notable in that they express particularly large amounts of cyclo-oxygenase-1. Cyclo-oxygenase-1 was initially identified and purified in the 1970s, using classical biochemical techniques, from bovine (Miyamoto *et al.*, 1976) and sheep (Hemler & Lands, 1976) vesicular glands and found to be a membrane bound homo-dimer of 70 kDa. The protein contained both the cyclo-oxygenase and peroxidase activities required to form, respectively, prostaglandin PGG<sub>2</sub> and PGH<sub>2</sub>. Either free or protein-bound heme was required for activity. The primary structure of cyclo-oxygenase-1 was later determined from the complementary DNA sequence of 2.7 kilobases (DeWitt & Smith, 1988).

### *Cyclo-oxygenase-2; the inducible isoform*

Cyclo-oxygenase metabolites are released in high amounts locally at the site of inflammation or systemically after infection. Initially it was believed that this was due to an increase in supply of arachidonic acid. However, in 1990 it was demonstrated that the increase in prostaglandin formation following exposure of isolated cells in culture to inflammatory

\*Author for correspondence; E-mail: [j.a.mitchell@ic.ac.uk](mailto:j.a.mitchell@ic.ac.uk)

stimuli (see De Witt, 1991) was due to an increase in cyclo-oxygenase enzyme expression (Fu *et al.*, 1990; Masferrer *et al.*, 1990). We now know that this increased cyclo-oxygenase is not cyclo-oxygenase-1 but an inducible isoform, cyclo-oxygenase-2.

The identification of cyclo-oxygenase-2 was, in many respects, a triumph of molecular biology. Early experiments demonstrated that cultured epithelial cells contained two distinct mRNA species, recognized under low stringency conditions by cDNA probes designed for the known cyclo-oxygenase (cyclo-oxygenase-1) (Rosen *et al.*, 1989). These probes hybridized to the predicted 2.8 kilobase mRNA but also to a novel 4.0 kilobase product. Importantly, these researchers also demonstrated that increases in the 4.0 kilobase product paralleled increases in enzymatic activity supporting the conclusion that this mRNA encoded for active protein. Later studies from the same group confirmed that these epithelial cells did indeed express two distinct forms of cyclo-oxygenase (Holtzman *et al.*, 1992). In separate studies, Xie and co-workers (Xie *et al.*, 1991) showed that mitogen stimulated chicken fibroblasts expressed a 4.1 kilobase mRNA which encoded a protein with 59% homology to the well characterized cyclo-oxygenase identified in sheep seminal vesicle (cyclo-oxygenase-1). Furthermore, phorbol esters were also shown to induce mouse fibroblasts to express an inducible cyclo-oxygenase (TIS10; Kujubu *et al.*, 1991) with striking similarities to the protein identified in chicken cells (Xie *et al.*, 1991). Finally, the inducible cyclo-oxygenase gene was directly demonstrated to encode a protein which had cyclo-oxygenase activities (Fletcher *et al.*, 1992; O'Banion *et al.*, 1992).

#### *Differential induction of cyclo-oxygenase-1 and cyclo-oxygenase-2 transcription*

The human cyclo-oxygenase-2 gene is 8.3 kilobases whereas the cyclo-oxygenase-1 gene is much larger at 22 kilobases (see Wu, 1995; Vane *et al.*, 1998). As discussed above, the gene products also differ in size. Cyclo-oxygenase-1 mRNA is approximately 2.8 kilobases whilst cyclo-oxygenase-2 mRNA is approximately 4.0 kilobases. Analysis of the 5'-flanking untranslated regions of cyclo-oxygenase-1 and cyclo-oxygenase-2 show that the cyclo-oxygenase-1 gene exhibits the features of a housekeeping gene whereas the gene for cyclo-oxygenase-2 appears to be a primary response gene. For example, the 5'-flanking region of the cyclo-oxygenase-2 gene has a canonic TATA box 30 base pairs upstream from the transcription start site (Tazawa *et al.*, 1994) whereas the same region in the cyclo-oxygenase-1 gene has no canonic TATA box (Wu, 1995). The cyclo-oxygenase-2 gene also contains a number of putative regulatory sites, including cyclic AMP response element, IL-6 response element, CCAAT/enhancer binding proteins, AP-2, nuclear factor- $\kappa$ B (NF- $\kappa$ B), Sp-1, PEA-3, GATA-1, and glucocorticoid response element (Wu, 1995). Interestingly, the cyclo-oxygenase-1 gene also has some putative regulatory sites, including Sp-1, PEA-3, AP2, NF-IL6, GATA-1 and shear stress response element (Wu, 1995). Characterization of the cyclo-oxygenase-2 gene as being a primary response gene has led to a larger number of studies investigating potential regulatory factors. These include in various cell types: cytokines, e.g. interleukin-1 $\alpha$  (Ristimaki *et al.*, 1994), tumour necrosis factor- $\alpha$ , interleukin-6, bacterial endotoxin and PMA (Geng *et al.*, 1995); growth factors, e.g. epidermal growth factor (Hamasaki & Eling, 1995), platelet derived growth factor, serum (Xie & Herschman, 1996) and chorionic gonadotrophin (Han *et al.*,

1996); locally acting mediators, e.g. 5-hydroxytryptamine (Stroebel & Goppelt-Struebe, 1994) and endothelin (Hughes *et al.*, 1995); fatty acid related mediators, e.g. arachidonic acid (Barry *et al.*, 1999), thromboxane A<sub>2</sub> (Stroebel & Goppelt-Struebe, 1994) and platelet activating factor (Bazan *et al.*, 1994); mechanical forces, e.g. pulsatile flow (Klein-Nulend *et al.*, 1997) and cyclic stretch (Kato *et al.*, 1998); and other stimuli such as circulating hormones, e.g. parathyroid hormone (Tetradis *et al.*, 1996). *In vivo* local increases in cyclo-oxygenase-2 expression have been associated with inflammation (Vane *et al.*, 1994), rheumatoid arthritis (Kang *et al.*, 1996), seizures (Marcheselli & Bazan, 1996) and ischaemia (Planas *et al.*, 1995). Cyclo-oxygenase-2 expression in the spinal cord is also elevated following peripheral inflammation (Beiche *et al.*, 1996). The intracellular pathways regulating these events appear numerous and complicated, varying between cell types and cellular stimulus. These pathways include, but are not limited to, reactive oxygen intermediates (Feng *et al.*, 1995), NF- $\kappa$ B and NF-IL6 (Yamamoto *et al.*, 1995), Ras/Rac1/MEKK-1/JNK kinase/JNK signal transduction leading to phosphorylation of c-Jun (Xie & Herschman, 1996), ceramide (Hayakawa *et al.*, 1996), mitogen-activated protein kinase (Hwang *et al.*, 1997), AP-1 and CRE nuclear binding proteins (Miller *et al.*, 1998), CCAAT/enhancer-binding proteins (Kim & Fischer, 1998) and protein kinase C/p42/p44 MAPK/p38 kinase (Barry *et al.*, 1999). Despite the presence of many pathways regulating the expression of cyclo-oxygenase-2 it is widely found that cyclo-oxygenase-2 is down-regulated by glucocorticosteroids (Masferrer *et al.*, 1994), and also by related agents such as 17 $\beta$ -estradiol (Morisset *et al.*, 1998).

#### *Enzyme homology*

Interestingly, although the genes for cyclo-oxygenase-1 and cyclo-oxygenase-2 are clearly different (see above), the proteins share approximately 60% homology at the amino acid level (Xie *et al.*, 1991). Cyclo-oxygenase-1 and cyclo-oxygenase-2 also both catalyse the formation of prostaglandin (PG) G<sub>2</sub> followed by PGH<sub>2</sub> from arachidonic acid, have a molecular weight of 70 kDa, and are identical in length. Studies of the tertiary structures of cyclo-oxygenase-1 and cyclo-oxygenase-2 have demonstrated that the amino acid conformation for the substrate binding sites and catalytic regions are almost identical (Kurumbail *et al.*, 1996; Picot *et al.*, 1994). However, there are important differences in these regions, particularly the exchange of Ile in cyclo-oxygenase-1 for Val in cyclo-oxygenase-2 at positions 434 and 523. These substitutions result in a larger and more flexible substrate channel in cyclo-oxygenase-2 than in cyclo-oxygenase-1 and also in the inhibitor binding site in cyclo-oxygenase-2 being 25% larger than that in cyclo-oxygenase-1 (Kurumbail *et al.*, 1996). There are also differences in the amino acid sequences in the N and C terminus of these enzymes (Kurumbail *et al.*, 1996; Picot *et al.*, 1994). Cyclo-oxygenase-2 for instance, is lacking in a 17 amino acid sequence at the N terminus but has an extra 18 amino acid sequence at the C terminus. The functionality of these differences are yet not known.

Cyclo-oxygenase-1 and cyclo-oxygenase-2 are membrane bound proteins that reside, after synthesis and transport, primarily in the endoplasmic reticulum (Spencer *et al.*, 1998). After the crystal structures of both isoforms of cyclo-oxygenase has been elucidated, it appeared likely that four amphipathic helices near the amino termini of these proteins would act as membrane binding domains (Kurumbail *et al.*,

1996; Picot *et al.*, 1994), a hypothesis that has been confirmed by experiments using chimeric proteins (Li *et al.*, 1998).

## Cyclo-oxygenase substrate, products and receptors

### Substrate

The first step in the formation of prostaglandins is the liberation of arachidonic acid from membrane bound phospholipids. This usually follows the action of phospholipase enzymes, primarily phospholipase A<sub>2</sub>. Like cyclo-oxygenase, phospholipase A<sub>2</sub> is expressed in a number of different isoforms, some of which are present constitutively, and some of which appear after inflammatory insult (see Cirino, 1998). However, it is the cytosolic 85 kDa phospholipase A<sub>2</sub> that most commonly supplies the arachidonic acid for prostaglandin production. Unlike cyclo-oxygenase, this form of phospholipase A<sub>2</sub> requires calcium and calmodulin for activation. Thus, cells expressing cyclo-oxygenase-1 require only the calcium activation of phospholipase A<sub>2</sub> for prostaglandin production to follow. Indeed, it has long been considered that phospholipase and not cyclo-oxygenase is the rate limiting step in prostaglandin production (see Cirino, 1998). More recently, however, with so much research attention being paid to cyclo-oxygenase-2, the role of phospholipases in the liberation of prostaglandins by inflamed cells and tissues expressing cyclo-oxygenase-2 has often been overlooked. However, as is the case for cyclo-oxygenase-1, the ability of cells and tissues expressing cyclo-oxygenase-2 *in vitro* (Saunders *et al.*, 1999) and *in vivo* (Hamilton *et al.*, 1999) to release prostanoids is greatly enhanced when substrate is provided or phospholipase activated. It is not yet clear exactly how phospholipase activity synchronises with cyclo-oxygenase in cells expressing both cyclo-oxygenase isoforms. However, it has been suggested that some level of compartmentalization is present in cells and that distinct pools of arachidonic acid are utilized by cyclo-oxygenase-1 and cyclo-oxygenase-2 (Reddy *et al.*, 1992). Because of the relationship between substrate liberation and prostanoid release, we have hypothesized that the cyclo-oxygenase-1-dependent release of products from cells

represents a one component system (Figure 1) whilst cyclo-oxygenase-2 dependent release represents a two component model (Hamilton *et al.*, 1999; Saunders *et al.*, 1999).

### Products

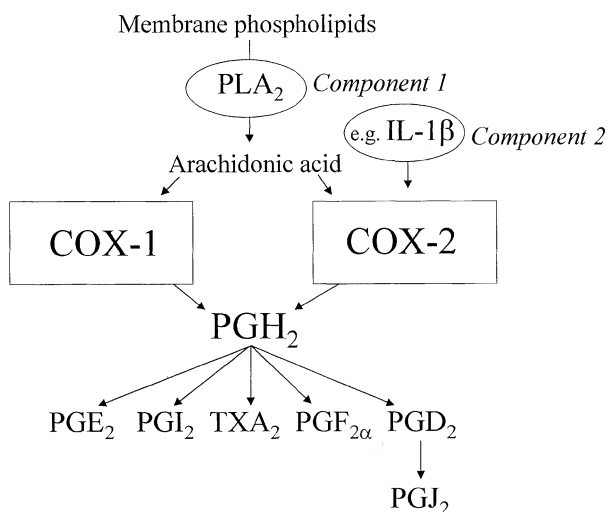
Once arachidonic acid has been supplied both isoforms of cyclo-oxygenase form PGG<sub>2</sub> and PGH<sub>2</sub> via identical enzymatic processes. Following these steps, PGH<sub>2</sub> can be metabolized by different enzyme pathways to a range of products with potent biological effects (Figure 1). Prostacyclin (PGI<sub>2</sub>), the main cyclo-oxygenase product of endothelial cells, is formed by prostacyclin synthetase. Thromboxane (TXA<sub>2</sub>), the main product of platelets, is formed by thromboxane synthetase. PGD<sub>2</sub> and PGF<sub>2</sub> are formed from PGH<sub>2</sub> by their respective synthase enzymes in a variety of cell types. PGE<sub>2</sub> is formed both by enzymatic (PGE<sub>2</sub> isomerase) and non-enzymatic pathways. The profile of products made by cells expressing cyclo-oxygenase-1 or cyclo-oxygenase 2 is therefore determined by the presence of different 'down stream' enzymes. However, when cells express large amounts of cyclo-oxygenase-2 the PGH<sub>2</sub> formed appears to be saturating for the PG synthase enzymes resulting in the formation of proportionately larger amounts of PGE<sub>2</sub> (Bishop-Bailey *et al.*, 1997a), possibly by non-enzymatic conversion. This is probably explained by the apparent lack of regulation of synthase enzymes at sites of inflammation.

The isoprostanes are a group of prostaglandin-like compounds formed under conditions of oxidative stress (see Morrow & Roberts, 1997). Although originally it was thought that these products were formed independently of cyclo-oxygenase, it is now clear that there are also cyclo-oxygenase-dependent pathways of formation. Thus, isoprostanes can be formed along with other arachidonic acid metabolites when cyclo-oxygenase-2 is induced (Jourdan *et al.*, 1999). Moreover, there is also evidence that platelets form isoprostanes, presumably *via* the action of cyclo-oxygenase-1 (Pratico *et al.*, 1995). Clearly, however, isoprostanes formed during oxidative stress are not dependent upon the activity of cyclo-oxygenase (Jourdan *et al.*, 1999).

One interesting observation has been that the products formed by cyclo-oxygenase-1 and cyclo-oxygenase-2 diverge following treatment with aspirin. Aspirin acetylates serine 530 in cyclo-oxygenase, which has the effect of irreversibly inactivating cyclo-oxygenase-1. This explains the anti-platelet activity of aspirin. By contrast, aspirin-acetylated cyclo-oxygenase-2 produces R15HETE instead of PGH<sub>2</sub> (Holtzman *et al.*, 1992; Meade *et al.*, 1993). The biological significance of this property of aspirin and cyclo-oxygenase-2 is not known. However, R15HETE is further metabolized by 5 lipoxygenase to form novel lipoxins (Clària & Sherhan, 1995), which may have effects in inflammation (Mitchell & Belvisi, 1997). These observations give rise to the intriguing possibility that part of the beneficial effects of aspirin could be attributed to the production of anti-inflammatory products.

### Receptors

As outlined above the effects resulting from induction of cyclo-oxygenase-2 at a particular site will be partly dependent upon the distribution of synthetase enzymes and the oxidative state of the cell; i.e. these will regulate the mix of arachidonic acid metabolites released. Clearly, however, another factor regulating the overall biological consequence of cyclo-oxygenase-2 induction is the distribution of prostanoid receptors on local target tissues and cells. There



**Figure 1** One and two component models of prostanoid production by, respectively, cyclo-oxygenase-1 (COX-1) and cyclo-oxygenase-2 (COX-2). PLA<sub>2</sub>, phospholipase A<sub>2</sub>; PG, prostaglandin.

are a number of different prostanoid receptor types, and yet more subtypes, linked to a range of signal transduction pathways. These prostanoid receptors are cell membrane spanning G-protein coupled receptors that mediate the physiological actions of the principal prostanoid metabolites (see Figure 1). Five major subdivisions of the prostanoid receptor family have been defined pharmacologically, and these correspond to each of the metabolites. The main endogenous receptor for PGI<sub>2</sub> is the IP receptor. IP receptors are linked to activation of adenylate cyclase and so the action of PGI<sub>2</sub> upon IP receptors is to elevate cyclic AMP, generally leading in many cells to an inhibition of active processes. For instance, in vascular smooth muscle activation of IP receptors promotes vasodilatation and in platelets leads to a reduction in aggregation and adhesion (Armstrong, 1996). TXA<sub>2</sub> is the principle ligand for TP receptors. TP receptors are linked to activation of the inositolphosphate pathway with subsequent increases in intracellular calcium, generally leading to activation of target cells. For instance, in direct contrast to IP receptors, activation of TP receptors on vascular smooth muscle causes contraction and on platelets causes aggregation and adhesion (Negishi *et al.*, 1995). There are four characterized receptors that PGE<sub>2</sub> preferentially activates, EP1-4. Each EP receptor is linked to a different transduction pathway giving rise to activation or inhibition of cellular responses (Pierce & Regan, 1998). PGF<sub>2α</sub> and PGD<sub>2</sub> activate FP and DP receptors, respectively. FP and DP receptors are linked to activation of inositolphosphates and generally result in activation of cellular processes including smooth muscle contraction.

Recently much interest has been paid to the role of the nuclear hormone receptor, peroxisome proliferator-activated receptor- $\gamma$  (PPAR- $\gamma$ ), in the biological actions of PGJ<sub>2</sub> and related metabolites (Forman *et al.*, 1995; Kliewer *et al.*, 1995). In particular PGJ<sub>2</sub> has been found to activate PPAR- $\gamma$  leading to changes in cell proliferation, such as promoting adipogenesis. Recently, it has been shown that PGF<sub>2α</sub> can negatively regulate PPAR- $\gamma$  (Reginato *et al.*, 1998), probably *via* an indirect action following activation of cell surface FP receptors. Interestingly, it appears that cyclo-oxygenase-2 and PPAR- $\gamma$  are co-localized under some circumstances. This is because, cyclo-oxygenase-2 is located on the nuclear membrane during synthesis and transport to the endoplasmic reticulum. It is, therefore, possible that cyclo-oxygenase-2 products preferentially activate PPAR or even conventional prostanoid receptors located on the nuclear membrane.

#### Other substrates

As mentioned above, the main substrate for cyclo-oxygenase-1 or cyclo-oxygenase-2 is the n-6 polyunsaturated fatty acid, arachidonic acid (20:4). However, other fatty acids can be utilized by both cyclo-oxygenase-1 and cyclo-oxygenase-2 under certain conditions. For instance, the inhibitory effects of n-3 polyunsaturated fatty acids on platelet function are thought to be due to inhibition of thromboxane production following their competition with arachidonic acid for the active site of cyclo-oxygenase-1 in platelets. Indeed, the n-3 fatty acid, eicosapentaenoic acid (EPA), can be metabolized by platelets, at the expense of arachidonic acid, to produce TXA<sub>3</sub> (see Goodnight *et al.*, 1992). TXA<sub>3</sub> has considerably weaker biological actions than the arachidonic acid derivative TXA<sub>2</sub>. Thus, the anti-platelet effects of dietary EPA are probably exerted *via* effects on cyclo-oxygenase-1 metabolites. In addition to its effects on platelets, EPA also appears to

substitute as a cyclo-oxygenase substrate in other cells as well. For example in isolated endothelial cells in culture (cyclo-oxygenase-1; Mitchell *et al.*, 1993) treatment with EPA leads to the formation of PGI<sub>3</sub> (Bordet *et al.*, 1986; Takahata & Yamanaka, 1987). *In vivo*, however, a number of studies have shown that humans taking EPA experience decreases in TXA<sub>2</sub> production without any change (von Shacky and Weber, 1985) or even an increase in PGI<sub>2</sub> formation (DeCaterina *et al.*, 1990; Honstra *et al.*, 1990). Thus, in man, EPA further tips the PGI<sub>2</sub>:TXA<sub>2</sub> balance in the favour of cardio-protection. The ability of purified preparations of cyclo-oxygenase-2 and cyclo-oxygenase-1 to metabolize other substrates (Percival *et al.*, 1994) has recently been compared. Both cyclo-oxygenase-1 and -2 catabolized substrates with the following rank order of efficiency, arachidonic acid > dihomo-gamma-linolenate > linoleate > alpha-linolenate. Gamma linolenate, alpha linolenate and EPA were very poor substrates for both forms, but in both cases were better metabolized by cyclo-oxygenase-2 than by cyclo-oxygenase-1 (Laneuville *et al.*, 1995). The greater promiscuity of cyclo-oxygenase-2 is thought to be due to the larger substrate channel in this isoform (see Vane *et al.*, 1998).

## Cyclo-oxygenase-2 as a therapeutic target

### Inflammation

Cyclo-oxygenase products, mainly PGE<sub>2</sub>, modulate the classical signs of inflammation. The two best studied inflammatory roles of cyclo-oxygenase products are induction of swelling and pain. In the case of 'swelling' PGEs are thought to cause plasma exudation in a synergistic fashion with other mediators such as complement factor 5a (Williams & Peak, 1977). One of the most important inflammatory disease targets associated with cyclo-oxygenase-2 is arthritis. In animal models of arthritis cyclo-oxygenase-2 is induced and thought to be responsible for the associated increase in PG production (Anderson *et al.*, 1996). Cyclo-oxygenase-2 expression has been identified in human osteoarthritis affected cartilage (Amin *et al.*, 1997) as well as in synovial tissue taken from patients with rheumatoid arthritis (Kang *et al.*, 1996).

In the case of pain, PGEs appear to 'sensitize' peripheral sensory nerve endings located at the site of inflammation (Bley *et al.*, 1998). In addition, cyclo-oxygenase products are thought to act in the spinal cord to facilitate the transmission of pain responses (Beiche *et al.*, 1996; Yamamoto *et al.*, 1996). However, despite a clear role for cyclo-oxygenase-2 in causing inflammatory swelling in animal models (see Chan & Rodger, 1997), the relative role of the two isoforms in pain is more complex. Clearly the perception of acute pain is more likely to be modulated by cyclo-oxygenase-1 as time for induction must elapse for cyclo-oxygenase-2. However, a number of studies have strongly implicated a role for cyclo-oxygenase-2 in inflammatory pain. For instance, highly selective cyclo-oxygenase-2 inhibitors (e.g. DFU) inhibit hyperalgesia in rats (Riendeau *et al.*, 1997). Moreover, the process of sensing pain may actually lead to the induction of cyclo-oxygenase-2 induction in the spinal cord, as has been demonstrated in rats exposed to inflammatory stimuli in the paw (Beiche *et al.*, 1996; Gardiner *et al.*, 1997). Importantly, selective inhibitors of cyclo-oxygenase-2, e.g. rofecoxib, have been shown to be analgesic in humans when used for post-dental surgery pain (Morrison *et al.*, 1999). The relative contributions of cyclo-oxygenase-1 and cyclo-oxygenase-2 in chronic pain (i.e. associated with rheumatoid or osteo-arthritis) remain to be established.

## Cancer

One of the exciting observations associated with the use of NSAIDs is the association with a reduction in the incidence of colon cancer. Indeed, a retro-prospective study revealed the startling findings that patients taking relatively low doses of aspirin, a maximum effect being seen at four to six tablets per week, for long periods of time had substantially reduced risks of developing colon cancers (Giovannucci *et al.*, 1995). It is not entirely clear how this protective effect of NSAIDs is exerted. However, adenocarcinomas in human subjects appear associated with marked increases in cyclo-oxygenase-2 expression (Smalley & DuBois, 1997) and evidence from studies with isolated cells in culture (DuBois *et al.*, 1998) or animal models (Williams *et al.*, 1997) similarly points to cyclo-oxygenase-2 being the level at which the beneficial effects of NSAIDs are exerted. The process underlying these effects is thought to be the ability of prostaglandins produced by cyclo-oxygenase-2 to slow down the rate of apoptosis in cancerous cells. This response is reduced by exposure to NSAIDs of the colon cancer cells (See DuBois *et al.*, 1998). The effect of cyclo-oxygenase-2 selective NSAIDs on colon cancer in man is currently being investigated.

Cyclo-oxygenase-2 has now been identified as present in a number of other cancers including, oesophageal (Zimmerman *et al.*, 1999), gastric (Murata *et al.*, 1999), and pancreatic (Tucker *et al.*, 1999) cancer. Whether or not cyclo-oxygenase-2 inhibitors can provide some level of protection in these forms of cancer remains to be established.

## Alzheimer's disease

In addition to colon cancer, epidemiological evidence has suggested that in patients taking NSAIDs there is a reduced risk of developing Alzheimer's disease (see Pasinetti, 1998). Indeed, in 1997 an inverse correlation was drawn between onset and severity of Alzheimer's and the intake of NSAIDs, usually ibuprofen. The protective effects of NSAIDs in this setting are most likely related to reduction of inflammation as paracetamol is without effect (Stewart *et al.*, 1997). Several groups have published supporting evidence for a role of cyclo-oxygenase-2 in this disease. Pasinetti & Aisen (1998) have shown that cyclo-oxygenase-2 expression is increased in the frontal cortex of brains from patients with Alzheimer's disease. Moreover, both isoforms of cyclo-oxygenase, as well as PPAR- $\gamma$  receptor, expression are increased in the temporal cortex of brains from these patients (Kitamura *et al.*, 1999). The link between the inflammatory properties of prostaglandins and Alzheimer's is not completely understood. However, animal studies have shown that after kainic acid induced seizures, cyclo-oxygenase-2 is induced in neurones that are susceptible to apoptosis (Tocco *et al.*, 1997). Nevertheless, it is possible that the epidemiological data showing that NSAIDs reduce Alzheimer's may have nothing to do with cyclo-oxygenase-2 inhibition. Indeed, the anti-platelet properties of NSAIDs may also relieve or prevent Alzheimer's. For example, de la Torre and co-workers (de la Torre *et al.*, 1997) hypothesised that Alzheimer's disease could be caused by the inappropriate development and invasion of capillaries in and around the brain tissue. The possibility of coagulation leading to ischaemic damage *via* blockade of these capillaries would be greatly reduced by aspirin and related drugs *via* an action on platelet cyclo-oxygenase-1.

The introduction of cyclo-oxygenase-2 selective NSAIDs into the general population for the treatment of arthritis and pain will allow, in time, epidemiological comparisons to be

made with traditional drugs for their effects on the development of Alzheimer's disease.

## Nonsteroidal anti-inflammatory drugs (NSAIDs) and cyclo-oxygenase isoforms

NSAID is a 'catch all' name for a large number of chemically distinct drugs. Together they represent the single most important group of self-prescribed pharmaceuticals and the most widely used drug class. The therapeutic benefits of all NSAIDs include inhibition of swelling and/or pain at the site of inflammation. In addition, aspirin also offers protection against stroke and thrombosis, Alzheimer's disease and cancer (see above). There are, however, side effects of NSAIDs that limit their use in some patients. Most common among these side effects is irritation and damage to the gastro-intestinal mucous, particularly the gastric mucosa. Each member of the NSAID family has some level of individual effects, however, the unifying common mechanism of action of all is inhibition of cyclo-oxygenase (Vane, 1971). With identification of two distinct forms of cyclo-oxygenase came a new hypothesis to explain the effects of NSAIDs; 'inhibition of cyclo-oxygenase-2 accounts for the therapeutic benefits and inhibition of cyclo-oxygenase-1 for the side-effects of NSAIDs' (Mitchell *et al.*, 1994). The following explains why this hypothesis was presented and how its validity was proven.

### *Relationship between the selectivity of NSAIDs for cyclo-oxygenase-1 and gastric side effects*

As a class the NSAIDs represent a major risk for morbidity and mortality from gastro-intestinal damage, perforation, ulcers and bleeding. In the U.S.A. the number of deaths per year due to NSAIDs is approximately 16,500 with 107,000 hospitalizations in the same period (Fries *et al.*, 1998). In the U.K., it is estimated that 12,000 ulcer complications and 1200 deaths per year are directly linked to NSAID intake (Hawkey, 1996). However, some NSAIDs cause more gastro-intestinal side effects than others (Henry *et al.*, 1996). A number of different experimental assays have been used to compare the potencies of NSAIDs on cyclo-oxygenase<sup>-1</sup> and cyclo-oxygenase<sup>-2</sup>. On the whole, studies have shown that the ability of a given NSAID to inhibit cyclo-oxygenase-1 correlates with the degree of side effects it causes.

### *Comparison of the selectivity's of NSAIDs for cyclo-oxygenase-1 versus cyclo-oxygenase-2*

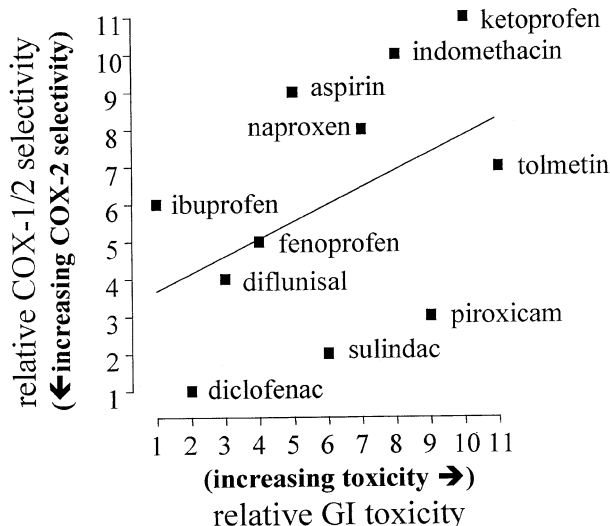
In 1993, two studies showed that a range of contemporary NSAIDs preferentially inhibited cyclo-oxygenase-1 (Meade *et al.*, 1993; Mitchell *et al.*, 1993). Meade and co-workers used semi-purified preparations of murine cyclo-oxygenase-1 and -2 and monitored activity by an oxygen electrode. In our study (Mitchell *et al.*, 1993), the production of PGs from intact cells expressing cyclo-oxygenase-1 (bovine endothelial cells) or cyclo-oxygenase-2 (LPS-activated J774 murine macrophages) was used as an index of enzyme activity. In addition, we compared NSAIDs potencies in whole cells with those of broken cell membranes and purified protein (Mitchell *et al.*, 1994). In both of the early reports, the rank orders of selectivity of NSAIDs for cyclo-oxygenase-1 were found to be associated with the risk of these agents producing gastro-intestinal side effects. Thus, the hypothesis that cyclo-oxygenase-1 inhibition was linked to NSAID side effects was suggested. Since these studies

were made a number of variations, and improvements have been made to *in vitro* assays used to screen novel NSAIDs, with the intention of identifying cyclo-oxygenase-2 selective compounds (see Vane *et al.*, 1998). A number of important criteria have been identified for optimum reliability of cyclo-oxygenase screens that are; (1), the use of both isoforms from a common species (preferably human); (2), common incubation times of drugs with both cyclo-oxygenase systems; (3), presence of realistic levels of plasma; (4), equivalent levels of substrate being available to both isoforms. To these ends, we have recently developed an improved cyclo-oxygenase-1–cyclo-oxygenase-2 screen that takes all of the points into consideration (Warner *et al.*, 1999).

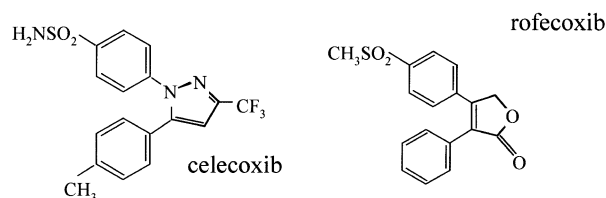
The classical NSAIDs currently used fall into three categories for the induction of gastro-intestinal side effects (see Figure 2). Ketorolac, ketoprofen, indomethacin, tolmetin and piroxicam constitute the first and most dangerous group of drugs. Fenoprofen, aspirin, naproxen and sulindac represent a mid-range gastro-intestinal risk. By contrast, ibuprofen, diclofenac and diflunisal appear to produce relatively fewer gastro-intestinal events (Garcia Rodriguez *et al.*, 1998; Henry *et al.*, 1998). Comparison of the relative activities of all these NSAIDs as inhibitors of human cyclo-oxygenase-1 versus cyclo-oxygenase-2 supports the idea that it is inhibition of cyclo-oxygenase-1 rather than inhibition of cyclo-oxygenase-2 that underlies the gastrotoxic effects of the NSAIDs (Figure 2).

#### Development of cyclo-oxygenase-2 selective compounds

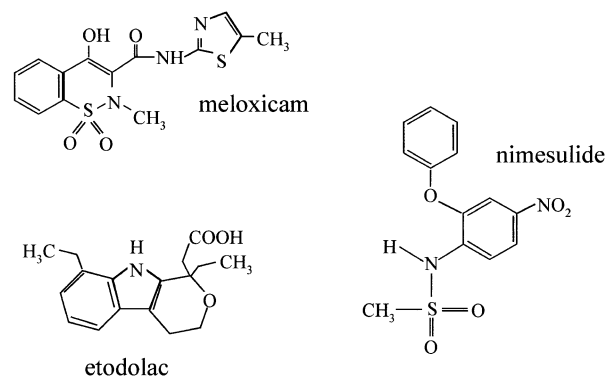
The use of cyclo-oxygenase screens *in vitro* has been remarkably successful and has led to the development of a number of experimental cyclo-oxygenase-2 selective compounds. These include from Merck Frosst (Canada), L-745,337 (Chan *et al.*, 1995), DFU (5,5-dimethyl-3-(3-fluorophenyl)-4-(4-methylsulphonyl)phenyl-2 (5H)-furanone; Riendeau *et al.*, 1997) and DFP (3-(2-propyloxy)-4-(4-methylsulphonylphenyl)-5,5-dimethylfuranone; Black *et al.*, 1999), and from Monsanto-Searle (U.S.A.), SC58125 (Guo *et al.*, 1996). Furthermore this approach has led to the



**Figure 2** Relationship for NSAIDs between relative activities against cyclo-oxygenase-1 versus cyclo-oxygenase-2 ( $IC_{80}$  ratios, Warner *et al.*, 1999) and risk of producing gastrointestinal toxicity (Henry *et al.*, 1996).



**Figure 3** Chemical structures of celecoxib and rofecoxib.



**Figure 4** Chemical structures of etodolac, meloxicam and nimesulide.

development of two cyclo-oxygenase-2 selective NSAIDs with FDA approval that are currently or about to be available in the U.S.A. These are rofecoxib (Vioxx<sup>TM</sup>; Chan *et al.*, 1999) from Merck-Frosst and celecoxib (Celebrex<sup>TM</sup>; Geiss, 1999) from Monsanto (Figure 3). In addition to the development of new compounds by these screens, it has been possible to identify currently prescribed NSAIDs that are cyclo-oxygenase-2 selective. For example, using whole blood assays compounds displaying more than 5 fold selectivity for cyclo-oxygenase-2 would include etodolac, meloxicam and nimesulide (Warner *et al.*, 1999) (Figure 4).

#### Confirmation of the 'cyclo-oxygenase-2 hypothesis' in animal models

The effects of experimental cyclo-oxygenase-2 selective inhibitors have been studied in a number of animal models of inflammation and hyperalgesia. As a general rule inhibitors with more than 10 fold selectivity for cyclo-oxygenase-2, have proven to inhibit inflammatory responses without causing gastro-intestinal side effects. For example in 1995, Chan and co-workers showed that the L-745,337, inhibited cyclo-oxygenase-2 with more than a 500 fold selectivity. This compound was compared with indomethacin and found to be equally effective at inhibiting carageenin-induced paw oedema. However, unlike indomethacin, L745-337 did not cause any gastric lesions (Chan *et al.*, 1995). Similarly, clinical trials with cyclo-oxygenase-2 selective agents such as celecoxib and rofecoxib have indicated that when used at therapeutic doses these produce significantly less gastrointestinal damage than traditional NSAIDs (Geiss, 1999; Lanza *et al.*, 1999). Other cyclo-oxygenase-2 selective inhibitors have been used to implicate a role for this isoform in hyperalgesia associated with thermal trauma (Dirig *et al.*, 1998) or intraplantar Freund's complete adjuvant (Hay *et al.*, 1997). In addition, cyclo-oxygenase-2 activity may contribute to hyperexcitability of sensory nerves in the spinal chord (Willingale *et al.*, 1997) and also to the genesis

of fever (Schwartz *et al.*, 1999). At the same time, however, it must also be noted that other studies have demonstrated pro-inflammatory roles for prostanoids generated by cyclo-oxygenase-1 (Wallace *et al.*, 1998b).

## Homeostatic/beneficial roles of cyclo-oxygenase-2

### *Constitutive expression of cyclo-oxygenase-2*

**Kidney** In addition to its expression at sites of inflammation cyclo-oxygenase-2 is also expressed constitutively in a number of tissues including the kidney and the brain. At these sites cyclo-oxygenase-2 activity is likely to be beneficial to the body forming prostanoids important to homeostatic processes. For example, in animal kidneys cyclo-oxygenase-1 and cyclo-oxygenase-2 are co-localized in the macula densa (Harris *et al.*, 1994), although in man cyclo-oxygenase-2 appears more associated with podocytes. In addition, cyclo-oxygenase-2 has been identified in a subset of thick ascending limb cells, where it has been suggested to be involved in the handling of ions (Vio *et al.*, 1997). People taking NSAIDs are at an increased risk of side effects in the kidney, but the incidence of these are much lower than those in the gastrointestinal tract (see Vane *et al.*, 1998). Indeed, it seems that formation of PGs generally only become important when the kidney is 'stressed', following for instance volume depletion, and PGI<sub>2</sub> in particular becomes essential to sustaining renal vasodilatation and blood flow. Despite the kidney expressing cyclo-oxygenase-2 constitutively the clinical trials to date have not identified any deleterious effects of cyclo-oxygenase-2-selective inhibitors. However, in salt-depleted individuals the cyclo-oxygenase-2-selective inhibitor, celecoxib, has been reported to cause sodium and potassium retention, leading to the conclusion that cyclo-oxygenase-2 selectivity does not spare the kidney under all circumstances (Rossat *et al.*, 1999).

**Central Nervous System (CNS)** Cyclo-oxygenase-1 and cyclo-oxygenase-2 are both localized in the brain and spinal cord. However, there appears to be specific regions of these structures where either isoform is expressed predominately. For instance cyclo-oxygenase-2 is mainly expressed in the cortex, hypothalamus and hippocampus (Breder *et al.*, 1995; Breder & Soper, 1996). By contrast, cyclo-oxygenase-1 is expressed in almost all structures of the central nervous system and is particularly highly localized in the forebrain. It is not yet clear what the specific roles of the two isoforms are in the central nervous system. However, it has been known for some time that PGs modulate activities such as thermoregulation and sleep by actions on or within nerves.

**Vas deferens** Recently, using immunohistochemical techniques, McKanna and co-workers (1998) have reported that cyclo-oxygenase-2 is the predominant isoform present in the vas deferens of the rat. In particular, cyclo-oxygenase-2 appears highly localized to the epithelium lining this structure. As may be expected, cyclo-oxygenase-1 and not cyclo-oxygenase-2 was predominately expressed in the seminal vesicles. Cyclo-oxygenase-2 expression was regulated by androgen and suggested to be involved in penile erection (McKanna *et al.*, 1998).

**Pancreas** It has been known for some time that PGE<sub>2</sub> inhibits the glucose-dependent release of insulin (see Robertson, 1998). However, it has recently been suggested that cyclo-oxygenase-2

is expressed constitutively in the pancreatic islets and that this isoform mediates PG production by these cells. In fact, this isoform predominates in islet cells isolated from hamster and human pancreas (Sorli *et al.*, 1998). These observations suggest that specific cyclo-oxygenase-2 inhibitors may have effects on glucose regulation in some patients.

**Gastrointestinal tract** As discussed above, gastric damage is the main side effect associated with inhibition of cyclo-oxygenase-1. Thus, cyclo-oxygenase-2 inhibitors are expected to cause fewer gastric side effects. Recent studies conducted by Zimmerman and co-workers (1998) suggests that cyclo-oxygenase-2, along with cyclo-oxygenase-1, is expressed in human gastric mucosa. Furthermore cyclo-oxygenase-2 selective inhibitors suppress the formation of PGs from samples of human gastric (Zimmerman *et al.*, 1998) and colonic tissue (McCartney *et al.*, 1999). Thus, a number of lines of evidence suggest that cyclo-oxygenase-2 is expressed throughout the human gastrointestinal tract. This 'constitutive' expression may be a result of low level trauma and/or infection that often occurs without obvious macroscopic symptoms. Whether cyclo-oxygenase-2 activity in the gut acts in the same way as cyclo-oxygenase-1, to protect it from damage, remains to be established. Nevertheless, these observations suggest that some patients may experience gastric side effects when taking cyclo-oxygenase-2 selective inhibitors (see Wallace *et al.*, 1998a). Indeed, use of selective inhibitors of cyclo-oxygenase-2 in animal models of ulceration and gastrointestinal damage have suggested that cyclo-oxygenase-2 products promote gastrointestinal healing (Reuter *et al.*, 1996; Mizuno *et al.*, 1997; Ukawa *et al.*, 1998).

### *Protective/beneficial effects of induced cyclo-oxygenase-2*

When discussing the role of cyclo-oxygenase-2, it should not be forgotten that the 'inflammatory response' is in itself a defence mechanism to protect the body from noxious stimuli. In this way, when cyclo-oxygenase-2 is induced at sites of inflammation and the response is appropriately limited, it can be considered a beneficial process. It is only when the response continues unnecessarily that cyclo-oxygenase-2 activity becomes deleterious. There are now emerging a number of other areas in which cyclo-oxygenase-2 induction has beneficial or protective properties including in the cardiovascular and respiratory systems as well as in the reproductive tract.

### *Cyclo-oxygenase-2 in the cardiovascular system*

The activity of cyclo-oxygenase-1 in endothelial cells is thought to be beneficial, contributing to the normal functioning of the cardiovascular system, *via* the release of PGI<sub>2</sub>. PGI<sub>2</sub> is a vasodilator, with potent inhibitory actions on platelet function (Moncada *et al.*, 1976; Radomski *et al.*, 1976; Sneddon & Vane, 1988), and is an endogenous anti-lipidemic agent (Willis *et al.*, 1986). Indeed, it has been suggested that increasing PGI<sub>2</sub> production or applying exogenous PGI<sub>2</sub>-mimetics could be therapeutically beneficial in cardiovascular diseases such as atherosclerosis (Willis *et al.*, 1986). Recently it has been shown that cyclo-oxygenase-2 becomes induced in animal arterial vessels after physical damage or exposure to pro-inflammatory cytokines (Rimarachin *et al.*, 1994). Similarly, we have shown that in human vessels *in vitro* inflammatory agents including interleukin (IL)-1 $\beta$ , tumour necrosis factor (TNF) $\alpha$ , interferon (INF) $\gamma$  or endotoxin induce cyclo-oxygenase-2 within the vascular smooth muscle component. Moreover, this induction is able to rectify the loss of PG production that follows

endothelial damage (Bishop-Bailey *et al.*, 1997b; 1998; Jourdan *et al.*, 1997). Induction of cyclo-oxygenase-2 in human vascular smooth muscle cells appears to have protective functions. Indeed, our studies show that cyclo-oxygenase-2 expression in the smooth muscle of damaged vessels suppresses inflammatory events such as cell proliferation (Bishop-Bailey *et al.*, 1997b), cytokine release (see Mitchell and Evans, 1998) and adhesion receptor expression (Bishop-Bailey *et al.*, 1998). These observations strongly suggest that cyclo-oxygenase-2 is induced as a protective defence enzyme in human cardiovascular disease (see Mitchell & Evans, 1998). Most recently, the group of Fitzgerald have shown that two cyclo-oxygenase-2 selective inhibitors, celecoxib (McAdam *et al.*, 1999) and rofecoxib (Catella-Lawson *et al.*, 1999) reduce circulating levels of prostacyclin in healthy human volunteers. This data suggests that cyclo-oxygenase-2 is a feature of the human cardiovascular system under physiological conditions. However, despite its selectivity in broken cell systems (Gierse *et al.*, 1999) it should be noted that celecoxib has also been shown to inhibit cyclo-oxygenase-1 in intact human cells (Warner *et al.*, 1999).

#### *Cyclo-oxygenase-2 in the respiratory tract*

Cyclo-oxygenase-2 is induced by cytokines in a number of airway cells including the epithelium (Mitchell *et al.*, 1994) and underlying smooth muscle (Belvisi *et al.*, 1998). Asthma and related diseases are characterized by excessive proliferation of airway cells, which contributes to airway narrowing in some patients. Cyclo-oxygenase-2 induction inhibits proliferation of human airway smooth muscle cells, suggesting a protective role of this enzyme in diseases such as asthma (Belvisi *et al.*, 1998).

A subset of asthmatic patients experience symptoms after taking aspirin and related drugs (Kowalski, 1995). This form of asthma is termed 'aspirin-sensitive asthma'. Although the biochemical mechanisms underlying aspirin-sensitive asthma are still being debated, there is a general level of acceptance that cyclo-oxygenase activity suppresses leukotriene production (Kowalski, 1995). Thus when cyclo-oxygenase is blocked by aspirin in sensitive patients, leukotriene production increases and asthma symptoms ensue. Aspirin sensitive asthmatics express cyclo-oxygenase-2 in their airways (Sousa *et al.*, 1997; Cowburn *et al.*, 1998). It is not yet clear whether cyclo-oxygenase-2 activity contributes to the 'leukotriene brake' associated with PG production in these patients. However, it is tempting to speculate that cyclo-oxygenase-2 induction in airway cells leads to beneficial products that limit the production of leukotrienes.

#### *Cyclo-oxygenase-2 in reproduction*

There are several lines of evidence that suggest that cyclo-oxygenase-2 is important in positive feed-back mechanisms

associated with reproduction. Both cyclo-oxygenase-1 and cyclo-oxygenase-2 are expressed in the uterine epithelium at different times in early pregnancy. For example, cyclo-oxygenase-1 is expressed preferentially in the uterine epithelium prior to implantation. After the embryo attaches, cyclo-oxygenase-1 is then down regulated (Chakraborty *et al.*, 1996). Cyclo-oxygenase-2 is expressed in the luminal epithelium and subepithelial stromal cells at the time of attachment of the blastocyst. Thus, expression of cyclo-oxygenase-2 may be important for localized increased uterine permeability and the attachment reaction (Chakraborty *et al.*, 1996).

### Summary and conclusions

This year is the centenary of aspirin. It remains one of the most popular self-prescribed preparations world wide. There have been a number of important scientific observations made during the first 100 years of aspirin that have led us to the identification of better drugs and identified new therapeutic targets. We now know that aspirin and related drugs inhibit the two identified forms of cyclo-oxygenase. Inhibition of the constitutively expressed cyclo-oxygenase-1 contributes significantly to the side effects of these drugs. Inhibition of the inducible cyclo-oxygenase-2 accounts for the beneficial anti-inflammatory and perhaps the analgesic, effects of these drugs. Using this principle as an incentive, assays have been developed and used to screen for new and better aspirin-like drugs. Indeed, several selective inhibitors of cyclo-oxygenase-2 have been identified. As predicted, the selective cyclo-oxygenase-2 inhibitors are anti-inflammatory without being ulcerogenic.

In addition to arthritis, there now seems to be two important diseases where cyclo-oxygenase-2 activity predominates and is deleterious, these are cancer (colon) and Alzheimer's. Thus, new selective inhibitors of cyclo-oxygenase-2 may have far reaching therapeutic potential.

There are clearly identified areas where cyclo-oxygenase-2 induction may be of benefit to the body. These are in major blood vessels and airways as well as in the gastro-intestinal and urogenital tract. Thus, we could anticipate potential unexpected side effects of cyclo-oxygenase-2 selective drugs in some patients.

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