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Cytochrome P450 125 (CYP125) catalyses C26-hydroxylation to initiate sterol side-chain degradation in *Rhodococcus jostii* RHA1

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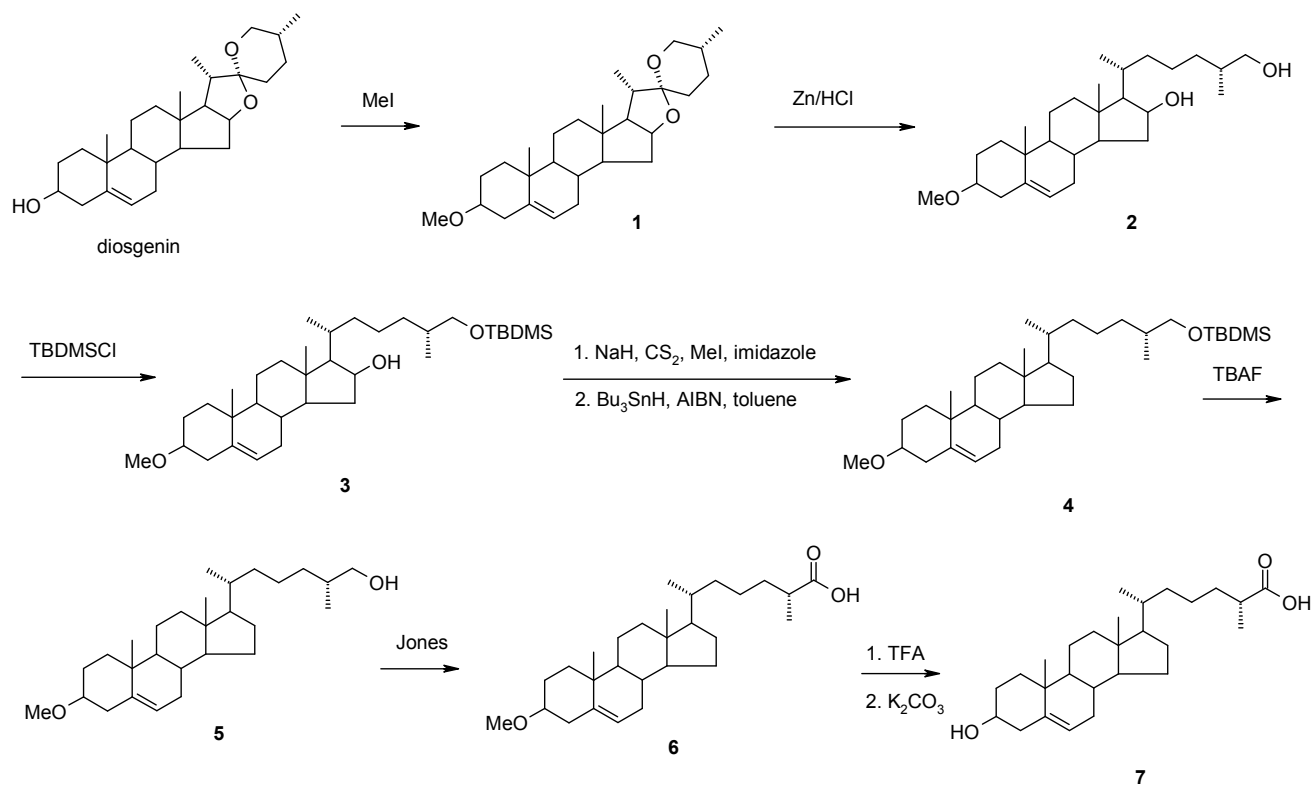
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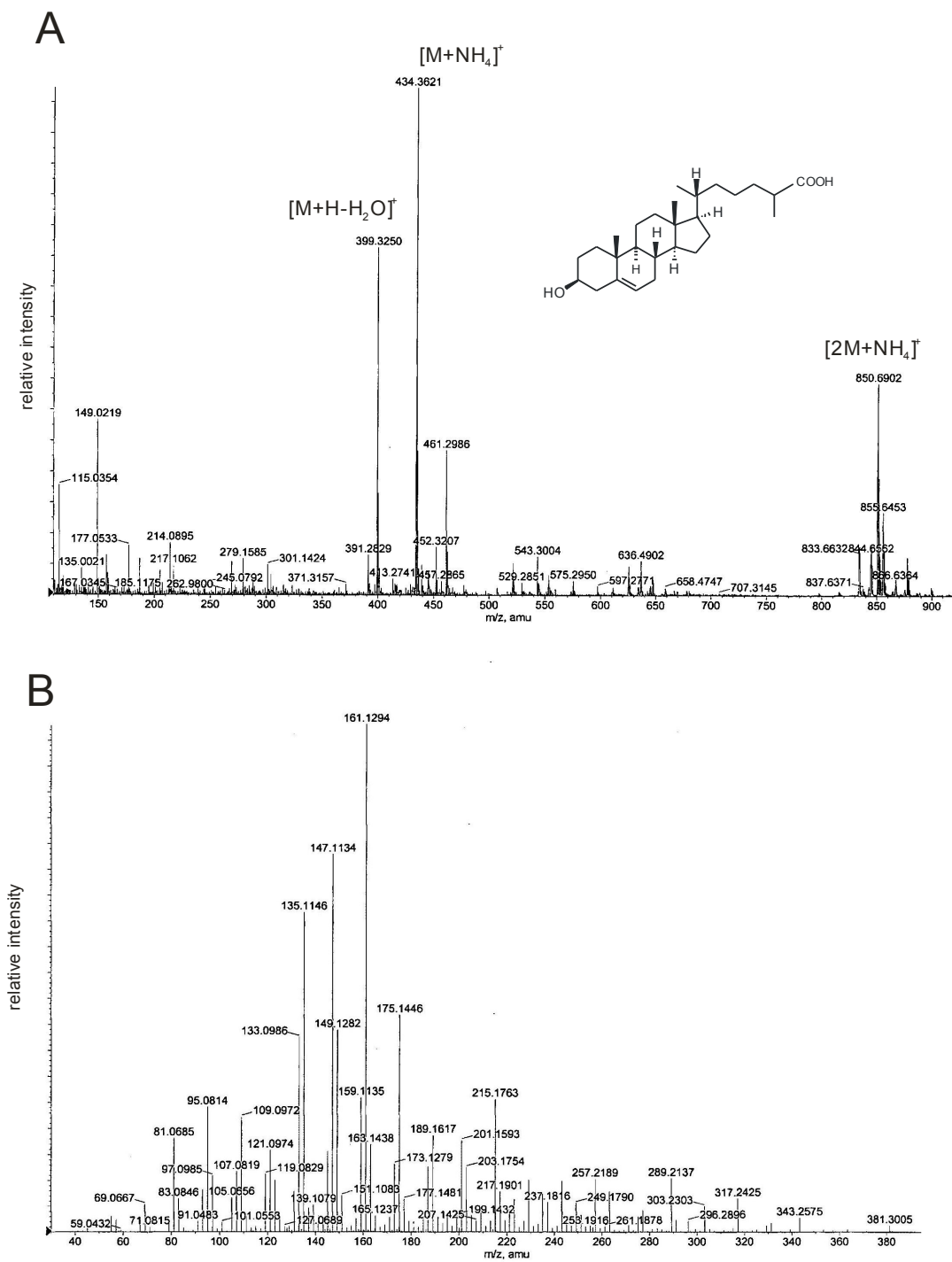
Supplemental Figure 1. Partial amino acid sequence alignment of actinobacterial and mammalian cytochrome P450 proteins: *R. jostii* RHA1 CYP125 (accession number ABG96465), *R. rhodochrous* DSM43269 CYP125 (FJ824698), *M. tuberculosis* H37Rv CYP125 (NP_218062), bovine CYP11A1 (NP_788817), human CYP27A1 (NP_000775), human CYP3A4 (P08684) and human CYP46A1 (NP_006659). The alignment was generated using ClustalW, MEGA version 3.1 (Kumar *et al.*, 2004). Specific residues discussed in the text in bold are indicated (V295, R300 and R343 of Cyp125_{RHA1}). Structure homology modeling of the CYP11A1:cholesterol complex revealed that the conserved arginine (R357, corresponding to R300 of CYP125_{RHA1}) points towards the docked cholesterol and is believed to be involved in catalysis (Storbeck *et al.*, 2007). Moreover, this residue was shown to be one of the active site residues in CYP46A1 (Mast *et al.*, 2008) and has been recently identified in P450s (e.g. CYP3A4) as a heme-interacting residue involved in substrate regio-selectivity and specificity (Seifert and Pleiss, 2009). For its part, the conserved arginine of CYP27A1 (R415, corresponding to R343 of CYP125_{RHA1}) was shown to be connected via a tripartite salt-bridge with the ExxR motif, forming an ERR-triad as part of the active site cavity (Prosser *et al.*, 2006; Masuda *et al.*, 2007). Finally, in CYP27A1, V367 (corresponding to V295 of CYP125_{RHA1}) is an active site residue and appears to be crucial for regioselectivity of hydroxylation of both cholesterol and 5 β -cholestane 3 α ,7 α ,12 α -triol (Mast *et al.*, 2006). Conserved motifs of the P450s super-family (ExxR and PxxF), the oxygen-binding domain (GxxT) and the heme binding domain (F[x]xxGxxxCxG) (Mestres, 2005; Huang *et al.*, 2008) are also shown.

		GxxT	
CYP125_RHA1	235	LSPEEFGFFVILLAVAG NET TRNAITHGMMAFLDHPD	
CYP125_DSM43269	235	LAPEEFGFFFIVLAVAG NET TRNAITHGMAAFLDNPE	
CYP125_H37Rv	253	LSDDEFGFFVVMLAVAG NET TRNSITQGMMFAEHDP	
CYP11A1	272	MLLEDVKANITTEMLAG VNT TSMTLQWHLYEMARSLN	
CYP27A1h	287	LSPREAMGSLPELLMAG VDT TSNTLTWALYHLSKDPE	
CYP3A4h	290	LSDLELVAQSIIFIFAG YET TSSVLSFIMYELATHPD	
CYP46A1h	287	QDDEGLLDNFVTFFFIAG HETS SANHLAFTVMELSRQPE	
		ExxR	V295 R300
CYP125_RHA1	282	KTAD EIV RWATPVNS FQRT ALEDTELGGVQIKKGQRVVMLYGSANFDEDAFEN PEKFDIMR	PxxF R343
CYP125_DSM43269	282	KTAAD EII RWATPV TSFQRT ALEDTELGGQTIRKGERVVMLYASANNDEEVFEN PREFDILR	
CYP125_H37Rv	300	ETAAD EIV RWATPV TAFQRT ALRDYELSGVQIKKGQRVVMFYRSANFDEEVFQD PFTFNILR	
CYP11A1	339	KASIK ETLRL LHPISVTL QRY PESDLVLQDYLI PAKTLVQVAIYAMGRDPAFFSSPDKFDPTR	
CYP27A1h	354	KAVLK ETLRL LYPVVPTNS RI IEKEIEVDGFLFPKNTQFV FCHYVVS RDPTAF SEPESEFQPHR	
CYP3A4h	357	DMVV NETLRL FP FIAMRLER VCKKDVEINGMFI PKG VVMI PSYALHRDPKYWTEPEKFLPER	
CYP46A1h	354	SQVLK ESLRL LYPPAWGT FR LLEEETLIDGVRVPGNTPLLFSTYVMGRMDTYFED PLTFNPDR	
		FxxxGxxxCxG	
CYP125_RHA1	349	VG F GGT GAHFC LGANLARLEIDL	
CYP125_DSM43269	349	LA F GGT GAHYC LGANLARMEIDL	
CYP125_H37Rv	367	VG F GGT GAHYC IGANLARMTINL	
CYP11A1	414	LG F -GW G V RQ CV GRR IAELEM T L	
CYP27A1h	434	V F -GY G V RAC L GRR IAELEM Q L	
CYP3A4h	433	T F -GS G PR NC I GMR FALMN M KL	
CYP46A1h	428	F F -SL G HR SC I GQ QFAQ M EV K V	

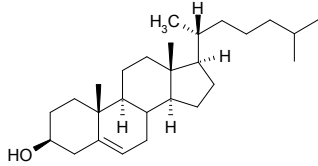
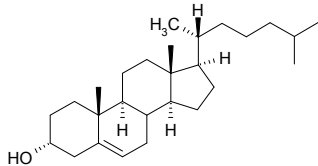
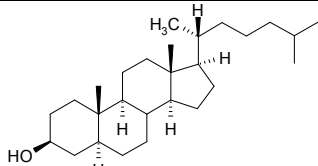
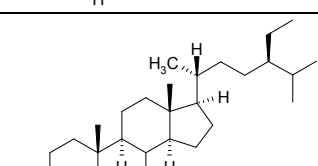
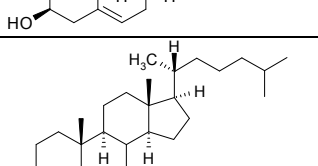
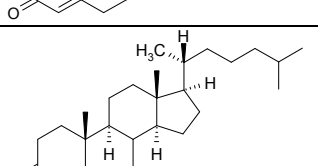
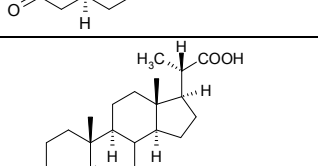
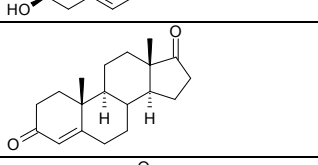
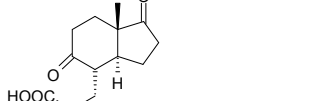
Supplemental Figure 2. Scheme of the chemical synthesis of 5-cholestene-26-oic-acid-3 β -ol from diosgenin.

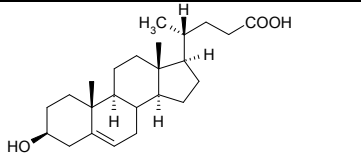
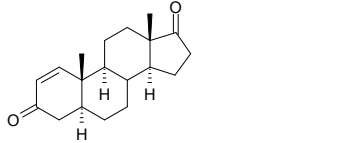
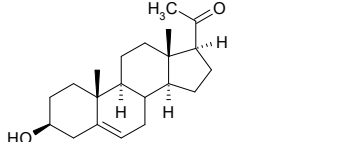
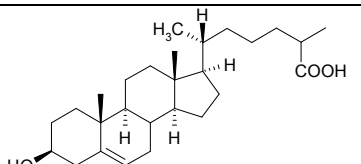


Supplemental Figure 3. (A) MS spectrum with fragmentation pattern of chemically synthesized 5-cholestene-26-oic acid-3 β -ol. (B) MS-MS spectrum with fragmentation pattern of ion peak with m/z 399.



Supplemental Table 1. Growth of wild type *Rhodococcus jostii* strain RHA1 and mutant strain RHA1 Δ *cyp125* on different sterols and related organic substrates. Growth is indicated with (+), whereas no growth is indicated with (-).

Steroid name	Steroid formula	RHA1	RHA1 Δ <i>cyp125</i>
cholesterol (5-cholestene-3 β -ol)		+	-
Epicholesterol (5-cholestene-3 α -ol)		+	-
5 α -cholestanol (5 α -cholestane-3 β -ol)		+	-
β -sitosterol* (5-cholestene-24 β -ethyl-3 β -ol)		+	-
Cholestenone (4-cholestene-3-one)		+	+
cholestanone (5 α -cholestane-3-one)		+	+
23,24-bisnor-5-cholenic acid-3 β -ol		+	+
4-androstene-3,17-dione		+	+
9,17-dioxo-1,2,3,4,10,19-hexanorandrostane-5-oic acid		+	+

5-cholenic acid-3 β -ol		+	+
1-5 α -androstene-3,17-dione		+	+
5-pregnene-3 β -ol-20-one		+	+
5-cholestene-26-oic acid-3 β -ol		+	+

* Mixture of β -sitosterol (>75 %) with β -sitostanol (10-14 %) and campesterol (6-9 %) was used.

Supplemental Table 2. Strains and plasmids used in this study.

Strain	Characteristics	Reference
<i>Rhodococcus jostii</i> RHA1	Wild type strain, PCB degrader	(Masai <i>et al.</i> , 1995)
<i>Rhodococcus rhodochrous</i> DSM43269	Wild type strain; potent sterol degrader; identical to strain IFO3338	DSMZ culture collection
<i>Rhodococcus rhodochrous</i> RG32	Mutant strain of DSM43269 capable of selective sterol side chain degradation; carries 5-fold unmarked <i>kshA</i> gene deletion	Wilbrink <i>et al.</i> , unpublished
<i>E. coli</i> DH5 α	General host for cloning	Bethesda Res. Laboratories
<i>E. coli</i> S17-1	Host strain for conjugal mobilization of pK18mobsacB derivatives to <i>Rhodococcus</i> strains	DSMZ culture collection
<i>Rhodococcus jostii</i> RHA1 Δ <i>cyp125</i>	<i>cyp125</i> gene deletion mutant strain of RHA1; blocked side chain degradation	This study
<i>Rhodococcus rhodochrous</i> RG32 Ω <i>cyp125</i>	<i>cyp125</i> gene disruption mutant strain of RG32; blocked side chain degradation	This study
Plasmid		
pK18mobsacB	Conjugative plasmid for gene mutagenesis in <i>Rhodococcus</i> ; <i>aphII sacB oriT</i> (RP4) <i>lacZ</i>	(Van der Geize <i>et al.</i> , 2001)
pBlueScript KS(II)	General <i>E. coli</i> cloning vector; <i>bla lacZ</i>	Stratagene
pTip-QC1	<i>Rhodococcus</i> expression vector; Chlr <i>bla PtipA repAB</i> (pRE2895)	(Nakashima and Tamura, 2004)
pRESQ	<i>E.coli-Rhodococcus</i> shuttle-vector; <i>aphII, lacZ-ccdB rep</i> (pMVS301)	(Van der Geize <i>et al.</i> , 2002)
pBs-Apra-ori	Conjugative pBlueScript KS(II) derivative; <i>aacIV, oriT</i> (RP4) <i>lacZ</i>	(Van der Geize <i>et al.</i> , 2008)
pRRE1	<i>E. coli- R. rhodochrous</i> shuttle-vector	This study
pDEL <i>cyp125</i> _{RHA1}	pK18mobsacB-derived mutagenic plasmid for <i>cyp125</i> _{RHA1} disruption in RHA1	This study
pTip-QC1 <i>cyp125</i> _{RHA1}	<i>cyp125</i> _{RHA1} expression plasmid used for CYP125 _{RHA1} production and functional complementation of RHA1 Δ <i>cyp125</i>	This study
pRESQ4679	Genomic library clone DSM43269 carrying <i>cyp125</i> _{DSM43269}	This study
p Ω <i>cyp125</i>	pK18mobsacB derived mutagenic plasmid for <i>cyp125</i> gene disruption in RG32	This study
pCOMP <i>cyp125</i> _{DSM43269}	pRRE1-derived plasmid carrying <i>cyp125</i> _{DSM43269} ; functional complementation of RG32 Ω <i>cyp125</i>	This study

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