

Daptomycin: a lipopeptide antibiotic for the treatment of serious Gram-positive infections

Judith N. Steenbergen*, Jeff Alder, Grace M. Thorne and Francis P. Tally

Cubist Pharmaceuticals, Inc., 65 Hayden Avenue, Lexington, MA 02421, USA

Infections caused by drug-resistant pathogens are on the rise. Daptomycin, a cyclic lipopeptide with activity against most Gram-positive pathogens, including vancomycin-resistant enterococci and methicillin-resistant *Staphylococcus aureus*, is a newly US-FDA approved antimicrobial for complicated skin and skin structure infections (cSSSI). Daptomycin has a unique mechanism of action that results in destruction of the membrane potential. The rapid bactericidal activity of daptomycin makes it an attractive antibiotic for serious Gram-positive infections.

Keywords: MRSA, cSSSI, vancomycin

Introduction

The increase in infections caused by Gram-positive pathogens and the rise in antibiotic-resistant bacterial strains have prompted the need for novel antibiotics.^{1,2} Recent reports indicate that more than 25% of *Staphylococcus aureus* infections in Europe are caused by methicillin-resistant *S. aureus* (MRSA), and the majority of these isolates are resistant to additional antibiotics.³ The incidence of MRSA varies greatly by country. Over 50% of *S. aureus* isolates in Portugal and Italy are methicillin-resistant, isolates in England, Greece, and France have MRSA rates around 25%, whereas the Netherlands and Switzerland have the lowest incidence of MRSA.³ Vancomycin has been an effective antibiotic against MRSA; however, the increased use of vancomycin has led to the development of isolates with reduced susceptibility. The mechanism of reduced susceptibility to vancomycin in *S. aureus* has not been fully elucidated and appears to be heterogeneous. Reduced susceptibility to vancomycin is correlated with alterations in the bacterial cell wall leading to significantly thicker and more disorganized cell walls.⁴ These thicker cell walls may sequester the vancomycin from reaching the target nascent cell wall precursors.⁴ Additional *in vitro* studies have linked development of vancomycin reduced susceptibility with phenotypic changes such as loss of haemolysis and the *mecA* gene, and genotypic changes such as the presence of either the group I or group II polymorphism in the *agr* gene locus.^{5–7}

To date, three vancomycin-resistant *S. aureus* strains (VRSA) have been isolated in the United States.^{8–11} Both the Pennsylvania and the New York strains were isolated from patients not on vancomycin therapy.^{9,12} Therefore, the need for new potent antimicrobial agents with MRSA activity is essential.

Mechanism of action

Daptomycin, a fermentation product produced by *Streptomyces roseosporus*, is a cyclic lipopeptide antibiotic with potent bactericidal activity against most Gram-positive organisms including multiple antibiotic-resistant and -susceptible strains.^{13–21} Daptomycin was recently approved in the United States for the treatment of complicated skin and skin structure infections (cSSSI) associated with *S. aureus* (methicillin-susceptible, MSSA, and methicillin-resistant, MRSA), *Streptococcus pyogenes*, *Streptococcus agalactiae*, *Streptococcus dysgalactiae* subsp. *equisimilis* and *Enterococcus faecalis* (vancomycin-susceptible only). Below, we discuss the *in vitro* potency of daptomycin against a range of other organisms including vancomycin-resistant *E. faecalis* and *Enterococcus faecium*.

The unique structure of daptomycin consists of a 13-member amino acid cyclic lipopeptide with a decanoyl side-chain (Figure 1). This distinctive structure confers a novel mechanism of action.²² The proposed mechanism involves insertion of the lipophilic daptomycin tail into the bacterial cell membrane, causing rapid membrane depolarization and a potassium ion efflux. This is followed by arrest of DNA, RNA and protein synthesis resulting in bacterial cell death (Figure 2).^{22–24} The bactericidal effect of daptomycin is rapid with greater than 99.9% of both MRSA and MSSA bacteria dead in less than 1 h.^{25,26} This rapid cell death does not result in rapid bacterial cell lysis.²⁴ Daptomycin also remains bactericidal (99.9% kill within 24 h) against stationary phase cultures of both MSSA and MRSA present at high density (10⁹ cfu) in a simulated endocardial vegetation model.²⁷

*Corresponding author. Tel: +1-781-860-8434; Fax: +1-781-861-1164; E-mail: Judith.steenbergen@cubist.com

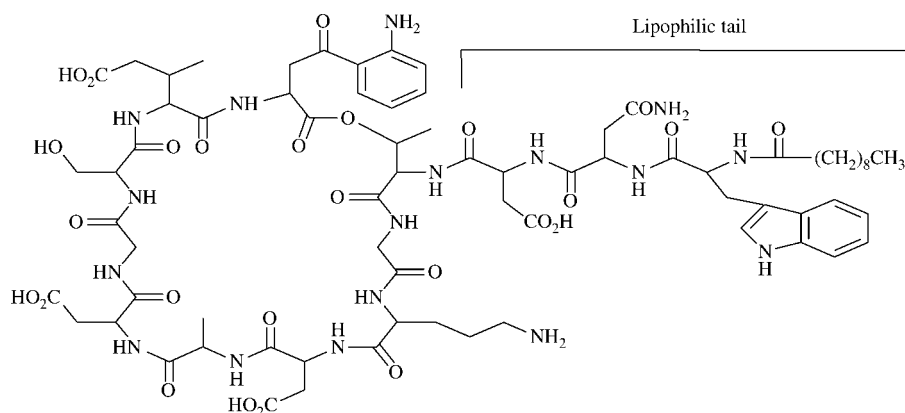


Figure 1. Daptomycin chemical structure.

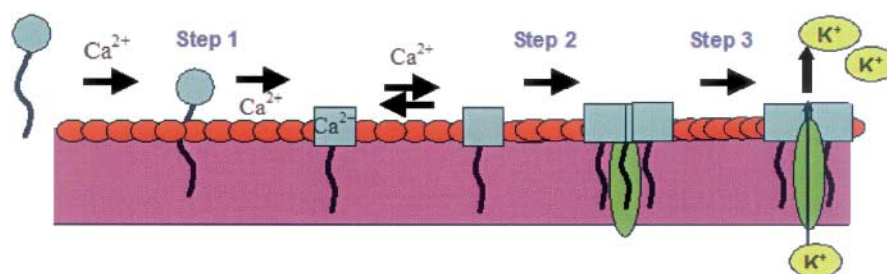


Figure 2. Daptomycin mechanism of action. Hypothetical steps: step 1, daptomycin binds to the cytoplasmic membrane in a calcium-dependent manner; step 2, daptomycin oligomerizes, disrupting the membrane; step 3, the release of intracellular ions and rapid cell death.

Microbiology

In vitro potency has been demonstrated for daptomycin against a range of aerobic and anaerobic Gram-positive bacteria including multidrug-resistant strains.^{13–21,28} MIC₉₀ values along with MIC ranges for select pathogens can be found in Table 1. The table shows data from two recent studies illustrating the conserved MIC ranges and values for both European strains and isolates collected worldwide. Daptomycin's spectrum of activity encompasses the difficult to treat antibiotic-resistant organisms including methicillin-resistant and -susceptible *Staphylococcus aureus* (MRSA, MSSA), glycopeptide-intermediate *S. aureus* (GISA), methicillin-resistant coagulase-negative *Staphylococcus* spp. (CoNS), and vancomycin-resistant enterococci (VRE).^{13–21} Daptomycin demonstrated potency against the recently isolated vancomycin-resistant *S. aureus* as well as linezolid and quinupristin/dalfopristin-resistant *S. aureus* and *E. faecium*.^{14,17–20,24} Furthermore, daptomycin is also effective against a variety of streptococcal groups such as the β -haemolytic streptococci including *S. pyogenes* (Group A) and *S. agalactiae* (Group B) as well as other *Streptococcus* spp.^{13–15,20,21} Along with the commonly isolated Gram-positive organisms, daptomycin is also potent against *Corynebacterium jeikeium*, and a variety of anaerobic species including *Peptostreptococcus* spp., *Clostridium perfringens*, *Clostridium difficile*, and *Propionibacterium acnes* (Table 1).²⁸ Drug synergy with daptomycin has been described *in vitro* with aminoglycosides and rifampicin antibiotics.²⁹

Resistance

Daptomycin's efficacy is enhanced by the near absence of antibiotic resistance as verified by both *in vitro* and clinical studies.³⁰ Resistance to daptomycin has been difficult to generate in the laboratory both in single passage and serial passage experiments.³⁰ The emergence of resistance was <0.2% across the entire set of Phase II and III clinical trials with over 1000 daptomycin-treated patients. The reason for this decrease in susceptibility is unknown and no transferable elements conferring daptomycin resistance have been isolated.

Pharmacology

Analysis of daptomycin pharmacodynamics determined that a once-daily dosing regimen increases the efficacy and safety of daptomycin.³¹ *In vitro* and *in vivo* analysis established that daptomycin is effective in a concentration-dependent manner, has a long half-life (8 h), and demonstrates a prolonged post-antibiotic effect up to 6.8 h (Table 2).³² These findings resulted in a once a day dosing regimen recommendation of 4 mg/kg for complicated skin and skin structure infections (cSSSI) in the United States.

Once-daily dosing of daptomycin results in linear pharmacokinetics with minimal drug accumulation.³¹ Daptomycin distributes primarily in the plasma, with penetration to vascular tissues (Table 3). Daptomycin does not cross the blood–brain barrier and does not penetrate the cerebrospinal fluid of normal individuals. However, there was a 5% penetration (relative to

Leading article

Table 1. *In vitro* activity of daptomycin against select Gram-positive bacteria

Organism	No. of strains	MIC range (mg/L)	MIC ₉₀ (mg/L)	Reference
Select aerobic pathogens				
<i>Staphylococcus aureus</i>				
oxacillin-resistant				
European isolates	334	0.12–1	0.5	14
worldwide isolates	1247	≤ 0.12–1	0.5	21
oxacillin-susceptible				
European isolates	888	≤ 0.015–1	0.5	14
worldwide isolates	1955	≤ 0.12–2	0.5	21
Coagulase-negative staphylococci ^a				
European isolates	1040	0.03–1	0.5	14
worldwide isolates	838	≤ 0.12–2	0.5	21
β-Haemolytic streptococci				
European isolates ^b	367	0.06–1	0.25	14
worldwide isolates	247	≤ 0.12–0.5	0.25	21
<i>Enterococcus faecalis</i>				
vancomycin-susceptible				
European isolates	1789	≤ 0.015–4	2	14
worldwide isolates	626	≤ 0.12–4	1	21
vancomycin-resistant				
European isolates	40	≤ 0.5–4	2	14
worldwide isolates	20	0.25–1	1	21
<i>Enterococcus faecium</i>				
vancomycin-susceptible				
European isolates	333	0.03–8	4	14
worldwide isolates	97	≤ 0.12–8	4	21
vancomycin-resistant				
European isolates	114	0.25–4	4	14
worldwide isolates	55	0.25–4	4	21
<i>Enterococcus</i> spp. ^c				
European isolates	160	≤ 0.015–4	4	14
worldwide isolates	21	0.5–4	2	21
<i>Corynebacterium jeikeium</i>	10	0.125–0.5	0.25	28
Select anaerobic pathogens				
<i>Actinomyces</i> group	22	0.06–16.0	4	28
<i>Bifidobacterium</i> spp.	13	< 0.03–1.0	0.5	28
<i>Clostridium difficile</i>	18	0.125–1.0	1	28
<i>Clostridium perfringens</i>	11	0.06–0.5	0.5	28
<i>Lactobacillus</i> spp. ^d	37	< 0.03–32.0	16	28
<i>Peptostreptococcus</i> spp.	14	0.125–1	1	28
<i>Propionibacterium</i> spp.	15	0.125–2	2	28

^aIncludes methicillin-resistant isolates vancomycin-resistant isolates.

^bEuropean isolates were *S. agalactiae* only.

^cIncludes vancomycin-resistant isolates.

^dAll *Lactobacillus* spp. were grown anaerobically.

Table 2. Mean (S.D.) daptomycin pharmacokinetic parameters in healthy volunteers on day 7

Dose (mg/kg)	C _{max} (mg/L)	T _{max} ^a (h)	AUC _{0–24} (mg·h/L)	t _{1/2} (h)	V (L/kg)	CL _T (mL/h/kg)	CL _R (mL/h/kg)	Ae ₂₄ (%)
4 (n = 6)	57.8 (3.0)	0.8 (0.5, 1.0)	494 (75)	8.1 (1.0)	0.096 (0.009)	8.3 (1.3)	4.8 (1.3)	53.0 (10.8)
6 (n = 6)	98.6 (12)	0.5 (0.5, 1.0)	747 (91)	8.9 (1.3)	0.104 (0.013)	8.1 (1.0)	4.4 (0.3)	47.4 (11.5)
8 (n = 6)	133 (13.5)	0.5 (0.5, 1.0)	1130 (117)	9.0 (1.2)	0.092 (0.012)	7.2 (0.8)	3.7 (0.5)	52.1 (5.19)

C_{max}, maximum plasma concentration; T_{max}, time to C_{max}; AUC_{0–24}, area under the concentration–time curve from 0 to 24 h; t_{1/2}, terminal elimination half-life; V, apparent volume of distribution; CL_T, systemic clearance; CL_R, renal clearance; Ae₂₄, percentage of dose recovered in urine over 24 h as unchanged daptomycin following the first dose.

^aMedian (minimum, maximum).

Table 3. Daptomycin tissue penetration

Tissue	Species	Maximum concentration	Percent relative to serum	Reference
Blister fluid	human	27.6 mg/L	68.4	42
Blood clot–tissue	rat, rabbit	3.5 µg/g	72.7	43
Peritoneal tissue chamber	rat	11.8 mg/L	35.1	44
Lung	mouse, rat	5 mg/L	9.3	45
BAL-ELF	mouse, rat, sheep	1 mg/L	2	45
CSF	rabbit	5.2 mg/L	5.97	33

BAL-ELF, bronchoalveolar lavage epithelial lining fluid.

serum) of daptomycin into the cerebrospinal fluid of rabbits with *Streptococcus pneumoniae* meningitis, resulting in clearance of the infection in this model.³³ Daptomycin is primarily renally excreted, with the majority of the drug remaining intact in the

urine.³¹ Since daptomycin is excreted through the kidneys, the dosing interval is increased to every 48 h in patients with severe renal impairment defined as a creatinine clearance of <30 mL/min. Because of daptomycin's unique mechanism of action and because it is not metabolized by cytochrome p450 or other hepatic enzymes, no antagonistic drug interactions have been observed.

Table 4. Incidence of adverse events that occurred in ≥ 2% of patients in either daptomycin or comparator treatment groups in Phase III cSSSI studies

Adverse event	Daptomycin % (n = 534)	Comparator ^a % (n = 558)
Gastrointestinal disorders		
constipation	6.2	6.8
nausea	5.8	9.5
diarrhoea	5.2	4.3
vomiting	3.2	3.8
dyspepsia	0.9	2.5
General disorders		
injection site reactions	5.8	7.7
fever	1.9	2.5
Nervous system disorders		
headache	5.4	5.4
insomnia	4.5	5.4
dizziness	2.2	2.0
Skin/subcutaneous disorders		
rash	4.3	3.8
pruritus	2.8	3.8
Diagnostic investigations		
abnormal liver function tests	3.0	1.6
elevated CPK	2.8	1.8
Infections		
fungal infections	2.6	3.2
urinary tract infections	2.4	0.5
Vascular disorders		
hypotension	2.4	1.4
hypertension	1.1	2.0
Renal/urinary disorders		
renal failure	2.2	2.7
Blood/lymphatic disorders		
anaemia	2.1	2.3
Respiratory disorders		
dyspnoea	2.1	1.6
Musculoskeletal disorders		
limb pain	1.5	2.0
arthralgia	0.9	2.2

^aComparators included vancomycin (1 g iv every 12 h) and antistaphylococcal penicillins (i.e. nafcillin, oxacillin, cloxacillin, flucloxacillin; 4–12 g/day in divided doses).

Pre-clinical studies

In pre-clinical studies, daptomycin treatment has been linked to fully reversible skeletal muscle toxicity with no effect on smooth or cardiac muscle. Animal studies determined that both degenerative and regenerative changes are observed in skeletal muscle with no rhabdomyolysis.³¹ These effects, which can be associated with elevated creatine phosphokinase (CPK) levels, are fully reversible after cessation of daptomycin use and were not statistically significant when compared with comparator.³¹ The numbers of side effects for patients receiving daptomycin were comparable to standard therapy and less than 2% of patients receiving daptomycin discontinued therapy.³⁴ The most common adverse events from the Phase III cSSSI clinical trials for daptomycin and comparator drugs are listed in Table 4.

The efficacy of daptomycin against a range of infections has been demonstrated in animal studies. Using a variety of antibiotic-resistant and -sensitive Gram-positive bacteria, daptomycin eradicated infections in the blood, muscle, kidney, heart and bone tissues of animals.^{35–41} These results show promise for daptomycin therapy for further clinical indications. An ongoing Phase III clinical trial is in progress to determine the efficacy of 6 mg/kg daptomycin once a day for endocarditis and bacteraemia caused by *S. aureus*.

Clinical studies

Daptomycin was evaluated in two large investigator-blinded, randomized, multicentre cSSSI studies in Europe, South Africa and the United States.³⁴ Adults with cSSSI of known or suspected Gram-positive aetiology were enrolled. The predominant cSSSI infections studied included wound infections, major abscesses and ulcer infections. Daptomycin was compared with conventional therapy of a semi-synthetic penicillin (e.g. nafcillin, oxacillin, cloxacillin, or flucloxacillin) or vancomycin (for suspected MRSA). The clinical success rates for each treatment group (intent to treat, modified intent to treat, clinically evaluable, and microbiologically evaluable) are shown in Table 5.³⁴

Table 5. Clinical success rates by treatment group, in Phase III cSSSI studies

Population	Daptomycin		Comparator ^a		
	<i>n</i>	% success	<i>n</i>	% success	95% CI ^b
Intent-to-treat	534	71.5	558	71.1	(−5.8, 5.0)
Modified intent-to-treat	428	74.5	471	74.7	(−5.5, 5.9)
Clinically evaluable	446	83.4	456	84.2	(−4.0, 5.6)
Microbiologically evaluable	365	84.7	396	85.9	(−3.8, 6.3)

^aCloxacillin, flucloxacillin, nafcillin, oxacillin or vancomycin.

^b95% confidence interval around the difference in success rate (comparator—daptomycin).

Table 6. Clinical success rates by infecting pathogen, in Phase III cSSSI studies

Pathogen	Success rate, <i>n/N</i> (%)	
	Daptomycin	comparator ^a
Methicillin-susceptible <i>Staphylococcus aureus</i> (MSSA) ^b	170/198 (85.9)	180/207 (87.0)
Methicillin-resistant <i>Staphylococcus aureus</i> (MRSA) ^b	21/28 (75.0)	25/36 (69.4)
<i>Streptococcus pyogenes</i>	79/84 (94.0)	80/88 (90.9)
<i>Streptococcus agalactiae</i>	23/27 (85.2)	22/29 (75.9)
<i>Streptococcus dysgalactiae</i> subsp. <i>equisimilis</i>	8/8 (100)	9/11 (81.8)
<i>Enterococcus faecalis</i> (vancomycin-susceptible only) ^b	27/37 (73.0)	40/53 (75.5)

^aVancomycin or semi-synthetic penicillins (e.g. nafcillin, oxacillin, cloxacillin or flucloxacillin).

^bAs determined by the central laboratory.

The study was designed to determine whether daptomycin was comparable to standard therapy and was not powered to show superiority. Therefore, statistical analysis determined that in the clinical trials, daptomycin was non-inferior to comparator therapy leading to daptomycin approval by the FDA in the United States.³⁴ Results of the microbiologically evaluable population are detailed by pathogen in Table 6.³⁴ Over 1000 patients were evaluated, and the following pathogens were the predominant organisms isolated; MSSA, MRSA, *S. pyogenes*, *S. agalactiae*, *S. dysgalactiae* subsp. *equisimilis*, and *E. faecalis*. The results from these Phase III trials confirmed the efficacy and safety of daptomycin.

Conclusions

In summary, daptomycin is a rapidly bactericidal antibiotic that is active against clinically relevant Gram-positive bacteria including antibiotic-resistant strains. Clinical data demonstrate that daptomycin is highly effective against cSSSI and ongoing clinical trials including infectious endocarditis caused by *S. aureus*, should expand treatment indications. The low occurrence of side effects, low resistance rates, and high potency demonstrate that daptomycin has significant clinical utility in the treatment of Gram-positive infections, including those caused by MRSA.

References

1. Bell, J. M. & Turnidge, J. D. (2002). High prevalence of oxacillin-resistant *Staphylococcus aureus* isolates from hospitalized patients in Asia-Pacific and South Africa: results from SENTRY antimicrobial surveillance program, 1998–1999. *Antimicrobial Agents and Chemotherapy* **46**, 879–81.
2. Stefani, S. & Varaldo, P. E. (2003). Epidemiology of methicillin-resistant staphylococci in Europe. *Clinical Microbiology and Infection* **9**, 1179–86.
3. Fluit, A. C., Wielders, C. L., Verhoef, J. *et al.* (2001). Epidemiology and susceptibility of 3,051 *Staphylococcus aureus* isolates from 25 university hospitals participating in the European SENTRY study. *Journal of Clinical Microbiology* **39**, 3727–32.
4. Cui, L., Ma, X., Sato, K. *et al.* (2003). Cell wall thickening is a common feature of vancomycin resistance in *Staphylococcus aureus*. *Journal of Clinical Microbiology* **41**, 5–14.
5. Verdier, I., Reverdy, M. E., Etienne, J. *et al.* (2004). *Staphylococcus aureus* isolates with reduced susceptibility to glycopeptides belong to accessory gene regulator group I or II. *Antimicrobial Agents and Chemotherapy* **48**, 1024–7.
6. Sakoulas, G., Eliopoulos, G. M., Moellering, R. C. *et al.* (2003). *Staphylococcus aureus* accessory gene regulator (agr) group II: is there a relationship to the development of intermediate-level glycopeptide resistance? *Journal of Infectious Diseases* **187**, 929–38.
7. Adhikari, R. P., Scales, G. C., Kobayashi, K. *et al.* (2004). Vancomycin-induced deletion of the methicillin resistance gene *mecA* in *Staphylococcus aureus*. *Journal of Antimicrobial Chemotherapy* **54**, 360–3.
8. Centers for Disease Control and Prevention. (2002). *Staphylococcus aureus* resistant to vancomycin—United States, 2002. In *Morbidity and Mortality Weekly Report*, pp. 565–7.
9. Centers for Disease Control and Prevention. (2002). Public Health Dispatch: vancomycin-resistant *Staphylococcus aureus*—Pennsylvania, 2002. In *Morbidity and Mortality Weekly Report* 902.
10. Centers for Disease Control and Prevention. (2004). Brief report: vancomycin-resistant *Staphylococcus aureus*—New York, 2004. In *Morbidity and Mortality Weekly Report* 322.
11. Mutnick, A. H., Biedenbach, D. J. & Jones, R. N. (2003). Geographic variations and trends in antimicrobial resistance among *Enterococcus faecalis* and *Enterococcus faecium* in the SENTRY Antimicrobial Surveillance Program (1997–2000). *Diagnostic Microbiology and Infectious Disease* **46**, 63–8.
12. Kacica, M. A., Scott, C., Johnson, G. *et al.* (2004). Vancomycin-resistant *Staphylococcus aureus* (VRSA) in a resident of a long-term care facility (LTCF). In *Program and Abstracts of the 42nd Annual Meeting of the Infectious Diseases Society of America*, Boston, MA, USA. Abstract 530, p. 143. Infectious Diseases Society of America, Alexandria, VA, USA.
13. Barry, A. L., Fuchs, P. C. & Brown, S. D. (2001). *In vitro* activities of daptomycin against 2,789 clinical isolates from 11 North

American medical centers. *Antimicrobial Agents and Chemotherapy* **45**, 1919–22.

14. Critchley, I. A., Draghi, D. C., Sahm, D. F. *et al.* (2003). Activity of daptomycin against susceptible and multidrug-resistant Gram-positive pathogens collected in the SECURE study (Europe) during 2000–2001. *Journal of Antimicrobial Chemotherapy* **51**, 639–49.

15. Fluit, A. C., Schmitz, F. J., Verhoef, J. *et al.* (2004). Daptomycin *in vitro* susceptibility in European Gram-positive clinical isolates. *International Journal of Antimicrobial Agents* **24**, 59–66.

16. Fluit, A. C., Schmitz, F. J., Verhoef, J. *et al.* (2004). *In vitro* activity of daptomycin against Gram-positive European clinical isolates with defined resistance determinants. *Antimicrobial Agents and Chemotherapy* **48**, 1007–11.

17. Petersen, P. J., Bradford, P. A., Weiss, W. J. *et al.* (2002). *In vitro* and *in vivo* activities of tigecycline (GAR-936), daptomycin, and comparative antimicrobial agents against glycopeptide-intermediate *Staphylococcus aureus* and other resistant gram-positive pathogens. *Antimicrobial Agents and Chemotherapy* **46**, 2595–601.

18. Richter, S. S., Kealey, D. E., Murray, C. T. *et al.* (2003). The *in vitro* activity of daptomycin against *Staphylococcus aureus* and *Enterococcus* species. *Journal of Antimicrobial Chemotherapy* **52**, 123–7.

19. Rybak, M. J., Hershberger, E., Moldovan, T. *et al.* (2000). *In vitro* activities of daptomycin, vancomycin, linezolid, and quinupristin–dalbopristin against staphylococci and enterococci, including vancomycin-intermediate and -resistant strains. *Antimicrobial Agents and Chemotherapy* **44**, 1062–6.

20. Snyderman, D. R., Jacobus, N. V., McDermott, L. A. *et al.* (2000). Comparative *in vitro* activities of daptomycin and vancomycin against resistant Gram-positive pathogens. *Antimicrobial Agents and Chemotherapy* **44**, 3447–50.

21. Streit, J. M., Jones, R. N. & Sader, H. S. (2004). Daptomycin activity and spectrum: a worldwide sample of 6737 clinical Gram-positive organisms. *Journal of Antimicrobial Chemotherapy* **53**, 669–74.

22. Silverman, J. A., Perlmutter, N. G. & Shapiro, H. M. (2003). Correlation of daptomycin bactericidal activity and membrane depolarization in *Staphylococcus aureus*. *Antimicrobial Agents and Chemotherapy* **47**, 2538–44.

23. Canepari, P., Boaretti, M., del Mar Lleo, M. *et al.* (1990). Lipoteichoic acid as a new target for activity of antibiotics: mode of action of daptomycin (LY146032). *Antimicrobial Agents and Chemotherapy* **34**, 1220–6.

24. Silverman, J., Harris, B., Cotroneo, N., *et al.* (2003). Daptomycin (DAP) treatment induces membrane and cell wall alterations in *Staphylococcus aureus*. In *Interscience Conference on Antimicrobial Agents and Chemotherapy*, Chicago, IL. Abstract C1–2135, p. 103. American Society for Microbiology, Washington, DC, USA.

25. Cha, R. & Rybak, M. J. (2004). Influence of protein binding under controlled conditions on the bactericidal activity of daptomycin in an *in vitro* pharmacodynamic model. *Journal of Antimicrobial Chemotherapy* **54**, 259–62.

26. Fuchs, P. C., Barry, A. L. & Brown, S. D. (2002). *In vitro* bactericidal activity of daptomycin against staphylococci. *Journal of Antimicrobial Chemotherapy* **49**, 467–70.

27. Tedesco, K. L. & Rybak, M. J. (2003). Impact of high inoculum *Staphylococcus aureus* on the activities of nafcillin (NAF), vancomycin (VAN), linezolid (LZD), gentamicin (GEN), and daptomycin (DAP) in an *in vitro* pharmacodynamic model (IVD). In *Interscience Conference on Antimicrobial Agents and Chemotherapy*, Chicago, IL. Abstract A-1151, p. 14. American Society for Microbiology, Washington, DC, USA.

28. Goldstein, E. J., Citron, D. M., Merriam, C. V. *et al.* (2003). *In vitro* activities of daptomycin, vancomycin, quinupristin–dalbopristin, linezolid, and five other antimicrobials against 307 gram-positive

anaerobic and 31 *Corynebacterium* clinical isolates. *Antimicrobial Agents and Chemotherapy* **47**, 337–41.

29. Rand, K. H. & Houck, H. (2004). Daptomycin synergy with rifampicin and ampicillin against vancomycin-resistant enterococci. *Journal of Antimicrobial Chemotherapy* **53**, 530–2.

30. Silverman, J. A., Oliver, N., Andrew, T. *et al.* (2001). Resistance studies with daptomycin. *Antimicrobial Agents and Chemotherapy* **45**, 1799–802.

31. Oleson, F. B., Jr, Berman, C. L., Kirkpatrick, J. B. *et al.* (2000). Once-daily dosing in dogs optimizes daptomycin safety. *Antimicrobial Agents and Chemotherapy* **44**, 2948–53.

32. Safdar, N., Andes, D. & Craig, W. A. (2004). *In vivo* pharmacodynamic activity of daptomycin. *Antimicrobial Agents and Chemotherapy* **48**, 63–8.

33. Cottagnoud, P., Pfister, M., Acosta, F. *et al.* (2004). Daptomycin is highly efficacious against penicillin-resistant and penicillin- and quinolone-resistant pneumococci in experimental meningitis. *Antimicrobial Agents and Chemotherapy* **48**, 3928–33.

34. Arbeit, R. D., Maki, D., Tally, F. P. *et al.* (2004). The safety and efficacy of daptomycin for the treatment of complicated skin and skin-structure infections. *Clinical Infectious Diseases* **38**, 1673–81.

35. Alder, J., Li, T., Yu, D. *et al.* (2003). Analysis of daptomycin efficacy and breakpoint standards in a murine model of *Enterococcus faecalis* and *Enterococcus faecium* renal infection. *Antimicrobial Agents and Chemotherapy* **47**, 3561–6.

36. Cha, R., Grucz, R. G., Jr & Rybak, M. J. (2003). Daptomycin dose–effect relationship against resistant gram-positive organisms. *Antimicrobial Agents and Chemotherapy* **47**, 1598–603.

37. Dandekar, P. K., Tessier, P. R., Williams, P. *et al.* (2003). Pharmacodynamic profile of daptomycin against *Enterococcus* species and methicillin-resistant *Staphylococcus aureus* in a murine thigh infection model. *Journal of Antimicrobial Chemotherapy* **52**, 405–11.

38. Kaatz, G. W., Seo, S. M., Reddy, V. N. *et al.* (1990). Daptomycin compared with teicoplanin and vancomycin for therapy of experimental *Staphylococcus aureus* endocarditis. *Antimicrobial Agents and Chemotherapy* **34**, 2081–5.

39. Louie, A., Kaw, P., Liu, W. *et al.* (2001). Pharmacodynamics of daptomycin in a murine thigh model of *Staphylococcus aureus* infection. *Antimicrobial Agents and Chemotherapy* **45**, 845–51.

40. Mader, J. T. & Adams, K. (1989). Comparative evaluation of daptomycin (LY146032) and vancomycin in the treatment of experimental methicillin-resistant *Staphylococcus aureus* osteomyelitis in rabbits. *Antimicrobial Agents and Chemotherapy* **33**, 689–92.

41. Smith, K., Cobbs, G., Dill, R. *et al.* (1990). Daptomycin versus vancomycin treatment for *Staphylococcus aureus* bacteremia in a murine model. *Chemotherapy* **36**, 428–34.

42. Wise, R., Gee, T., Andrews, J. M. *et al.* (2002). Pharmacokinetics and inflammatory fluid penetration of intravenous daptomycin in volunteers. *Antimicrobial Agents and Chemotherapy* **46**, 31–3.

43. Michiels, M. J. & Bergeron, M. G. (1996). Differential increased survival of staphylococci and limited ultrastructural changes in the core of infected fibrin clots after daptomycin administration. *Antimicrobial Agents and Chemotherapy* **40**, 203–11.

44. Vaudaux, P., Francois, P., Bisognano, C. *et al.* (2003). Comparative efficacy of daptomycin and vancomycin in the therapy of experimental foreign body infection due to *Staphylococcus aureus*. *Journal of Antimicrobial Chemotherapy* **52**, 89–95.

45. Alder, J., Arbeit, R., Eisenstein, B., *et al.* (2004). Pulmonary epithelial lining fluid (ELF) as a privileged site: daptomycin in pulmonary infections. In *International Congress of Infectious Diseases, Cancun, Mexico. 11th ICID Abstracts*, Abstract 60.005, p. S195. International Society for Infectious Diseases, Brighton, UK.