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**Data paper: FoRAGE (Functional Responses from Around the Globe in all Ecosystems) database: a compilation of functional responses for consumers and parasitoids**

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**ABSTRACT**

Functional responses – the relationships between consumer foraging rate and resource (prey) density – provide key insights into consumer-resource interactions and predation mechanics while also being a major contributor to population dynamics and food web structure. We present a global database of standardized functional response parameters extracted from the published literature. We refit the functional responses with a Type II model using standardized methods and report the fitted parameters along with data on experimental conditions, consumer and resource taxonomy and type, as well as the habitat and dimensionality of the foraging interaction. The consumer and resource species covered here are taxonomically diverse, from protozoans filtering algae to wasps parasitizing moth larvae to wolves hunting moose. The FoRAGE database (doi:10.5063/F17H1GTQ) is a living data set that will be updated periodically as new functional responses are published.

**INTRODUCTION**

The strength of a consumer-resource interaction determines the importance of a given food web link (Novak and Wootton, 2010). Thus, interaction strength can give key insights into population dynamics and the structure and stability of food webs (Gilbert et al., 2014; McCann et al., 1998).
One way to measure consumer-resource interaction strength is through the functional response (Holling, 1959a). A consumer’s functional response describes foraging rate as a function of resource availability. The simplest form is Type I, where foraging increases linearly with resource density. Most consumers, however, must pay a time cost for each resource item utilized, resulting in an asymptotic, Type II functional response. A common form of Type II functional response is the Holling disc equation:

\[ f_{pc} = \frac{aR}{1+aRh} \]  

Equation 1.

where \( f_{pc} \) is the per capita foraging rate of the consumer (number of resources per time per predator), \( a \) is the space clearance rate (space per time per predator), \( R \) is the initial resource density (resources per space), and \( h \) is handling time (time per resource) (Holling, 1959a). Space clearance rate describe how quickly a consumer can remove resources from a given space, while handling time describes the loss in search time associated with the consumption of an individual resource item. Handling time reflects any activity that prevents the consumer from searching for additional resources (prey items) after catching a resource. This may include transporting the resource to a safe location, chewing the resource, and any time spent digesting (if the consumer cannot continue hunting while digesting).

Here, we present standardized functional response parameters and the associated experimental conditions for over 2,000 consumer-resource combinations from the literature. The database will be periodically revised and updated with new functional responses as they are published. The data set is housed at the Knowledge Network for Biocomplexity (https://knb.ecoinformatics.org/) and can be found through its DOI: doi:10.5063/F17H1GTQ. The data set contains two files, one containing the overall data set of functional response parameters.
and associated information (with ‘data set’ in the file name) and the other containing the raw foraging observations (with ‘original curves’ in the file name).

**METHODS**

We searched the literature using terms such as “functional response”, “predator-prey interaction”, and “biocontrol” to find papers reporting predator and parasitoid functional responses. We also searched within references of papers that contained functional responses, through other compilations of functional responses (DeLong et al., 2015; DeLong and Vasseur, 2012a, 2012b, 2011; Kalinoski and DeLong, 2016; Rall et al., 2012; Uiterwaal and DeLong, 2018), and through the websites of researchers that had done multiple functional response papers. Our search produced 2,083 functional responses across a wide range of taxa from all habitats and biomes and from around the world. See Appendix A for sources. Each individual functional response curve received a unique ID number associated with its functional response data, consumer and resource traits, and experimental conditions. Multiple distinct curves – due to experimental treatments, age and/or sex, consumer and/or resource species, temperature, number of consumers, substrate, or other category – from the same paper received different IDs.

**Functional response data**

**Data collection**

Due to discrepancies in terms of equations and techniques used to calculate functional responses across studies, species, and experimental approaches, we did not use the parameters reported in the original papers. Rather, we recorded original resource density and consumer foraging rate data as given in the paper. We preferentially recorded raw data in tables, but we digitized most data from figures. When raw data was not available, we recorded mean foraging rate at a given resource
density along with standard errors and sample size at that density. We converted error bars presented as 95% confidence intervals or standard deviation to standard error. In some cases where raw data was reported in figures, it was unclear how many actual observations were represented by a point on the graph. In these cases, we used the minimum possible number of replicates based on the reported sample size to obtain a conservative number of datapoints. If resource density was expressed in terms of carbon, protein, or chlorophyll concentration, we converted this to number of individuals using conversions provided in the paper (preferentially) or from outside sources. The data collection process is outlined in Figure 1. Additional functional response data can be included in the FoRAGE data set by making the corresponding author aware of a new source via email.

**Data fitting**

We made all data comparable by standardizing resource density to units of resources per cm$^2$ or m$^2$ or resources per cm$^3$ or m$^3$ and foraging rate to units of number of resources eaten (or parasitized) per consumer per day. For 2.5-dimensional foragers (See “Experimental conditions” below), we rescaled prey density by raising it to the power of 2.5/2 if density was reported as an area and to the power of 2.5/3 if density was reported as a volume. In this way, the units of space clearance rate for organisms foraging in a fractal dimension between a plane and a volume are m$^{2.5}$ per predator per day. However, the prey density data are retained in either two or three dimensional units within the ‘original curves’ file.

We generated 200 bootstrapped datasets per functional response. We used standard ransom sampling with replacement for data sets with raw data. For data sets that reported mean foraging rate at different densities, we generated simulated data sets with the same mean, standard error,
and sample size as the reported data set using the same bootstrapping procedure. Then, we fit foraging data to the Holling disc equation (Equation 1) if the resource was replenished by the experimenters as it was consumed throughout the experiment. Datasets from wild consumers were treated as having resources replenished. If resources were not replenished, we fit foraging data to the Roger’s random predator equation:

\[ f_{pc} = R - \frac{W(ahRe^{-a(t-hR)})}{ah} \]  

Equation 2.

where \( f_{pc} \) is the per capita number of resources eaten by the consumer in the time of the foraging trial \( t \), \( W \) is the Lambert W function, and \( R, a \), and \( h \) are as in Equation 1. This equation accounts for resource depletion as the consumer forages (Bolker, 2008; Rogers, 1972). We used nonlinear ordinary least squares regression in Matlab to conduct these fits. To obtain parameter estimates and confidence intervals, we used the medians and 95% quantiles of the bootstrapped parameter distributions, respectively. We did not account for prey or predator population growth in the fitting procedure (Rosenbaum and Rall, 2018), although most experiments were conducted for periods precluding changes in prey or predator through reproduction.

When sample size was not given for data presented as means, we assumed conservatively that there were three replicates, the minimum number required to obtain a standard error. To estimate error when none was given or when error type was not specified, we used a Taylor power law relationship between the mean and the foraging rate variance across all observations in the data set for which this was available. We estimated this relationship using ordinary least squares regression with the log of the standard error as the dependent variable and the log of the mean foraging rate as the independent variable (Figure 2). The fitted exponent was 0.83 (0.006 ± SE) and the intercept was -1.76 (± 0.028). When arena size was not given, we conducted the fits and reported the handling time in the standard units but space clearance rate with units of ‘arenas per
predator per day’. If handling time was not different from zero (95% confidence intervals overlapped zero), the functional response was assumed to be Type I and handling time was removed.

**Consumer and resource traits**

**Mass**

We recorded masses or lengths of consumer and resource species from the original papers if available. If size measurements were not available in the original paper, we used mass or length estimates from external sources. We converted lengths to masses using length-weight relationships (usually by order or family). When no species-level size estimates were available, we calculated size based on one or more closely related species (e.g., in the same genus). In some cases, we were unable to obtain sizes of juveniles. In these cases, we calculated juvenile:adult size ratios of a related organism for which we did have juvenile size data. We used this percentage to estimate the unknown juvenile size of the focal organism. When necessary, we used water content to calculate wet mass from dry mass. Because predators and parasitoids utilize live resources, wet mass provides a more accurate size estimate than dry mass for functional response experiments. We used volume estimates and the density of freshwater or saltwater to estimate masses of some aquatic organisms (e.g. single-celled algae). Our methods for recording consumer and resource masses are outlined in Figure 3.

**Other traits**

We recorded consumer and resource traits as reported in the paper, including developmental stage, sex, and interaction type (predation or parasitoidism). We also assigned each consumer and
resource to approximate taxonomic categories, including their level of cellularity (unicell or metazoan), vertebrate status (vertebrate, invertebrate, or protozoan), and other levels that represented major groupings of data such as ‘fish’ or ‘copepod’ or ‘dinoflagellate’.

**Experimental conditions**

We recorded temperature or average temperature (if a range was given) at which experiments with ectotherms were conducted. If the consumer was an endotherm, we recorded body temperature. We also reported the number of consumers per arena and the length of time consumers were deprived of resources before the experiment. We recorded notes on other experimental variables such as habitat complexity or chemical pretreatment of organisms. We also identified field-determined functional responses as ones where consumers were foraging in natural field settings rather than in a laboratory.

We determined whether the functional response occurred in two-dimensional (e.g. wolf spiders in a petri dish), three-dimensional (e.g. copepods and *Daphnia* in a tank), or 2.5-dimensional (e.g. insects crawling on whole plants, spiders on webs) space. In cases where the consumer moved in three dimensions but the resource moved in two dimensions (e.g. fish consuming bottom-dwelling crayfish), we determined the interaction to be occurring in two dimensions.

**ACKNOWLEDGEMENTS**

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Kalsi, M., Seal, D.R., Nuessly, G.S., Capinera, J.L., Martin, C.G., 2014. Distribution of Zelus longipes (Hemiptera: Reduviidae) in South Florida Corn Fields and Its Functional Response to Corn-


Figure 1. Flow diagram detailing the decisions for how to include and standardize functional response data from the literature.

*When data was presented in both tables and figures, we used data from tables.*
Figure 2. Taylor power law relationship between mean foraging rate and the standard error of the mean across all observations. This relationship was used to estimate the foraging rate standard error for studies where only the mean foraging rate at a resource level was given.
Figure 3. Flow diagram detailing the methods for obtaining body masses of consumers and resources represented in functional response data in the FoRAGE database.

*When both mass and length were available, we used mass. When both wet mass and dry mass were available, we used wet mass.

**For some aquatic organisms, we used length to calculate volume, which we converted to mass using the density of saltwater or freshwater, as appropriate.
Figure 4. Pie charts showing the breakdown of functional responses in the FoRAGE database by A) habitat, B) foraging type, C) predator major grouping, and minor groupings within the major groupings, connected by lines, for D) invertebrates, E) protozoa, and F) vertebrates.
Appendix A: Publications containing original data on the functional responses in the FoRAGE database. Sources are grouped by Predator Major or Minor group.

<table>
<thead>
<tr>
<th>Predator major or minor group</th>
<th>Sources</th>
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<td>Insect</td>
<td>(Agarwala et al., 2001; Akre and Johnson, 1979; Anderson, 2016; Asante, 1995; Atlihan et al., 2010; Bailey, 1989; Ball et al., 2015; Barbosa et al., 2014; Cabral et al., 2009; Chang et al., 1993; Chenaux et al., 2011; Chesson, 1989; Chong and Oetting, 2006; Chow et al., 1983; Claver et al., 2003; Coll and Ridgway, 1995; Cothran and Thorp, 1985; Crowley and Martin, 1989; De Clercq et al., 1998, 2000; Ding-Xu et al., 2007; Dinis et al., 2016; Donnelly and Phillips, 2001; El-Basha et al., 2012; Elliott, 2003, 2003; Enkegaard et al., 2001; Farahani and Goldansaz, 2013; Farhadi et al., 2010; Faria et al., 2004; Farrokhi et al., 2010; Flinn, 1991; Foglar et al., 1990; Folsom and Collins, 1984; Fox and Murdoch, 1978; Frewin et al., 2010; Gers et al., 2010; Gers and Ratte, 2009; Gilioli et al., 2005; Gitonga et al., 2002; Gotoh et al., 2004; Gresens et al., 1982; Gupta et al., 2012; Hassanpour et al., 2011; Hassell, 1971; Hassell et al., 1977, 1977; He et al., 2012; Heidarian et al., 2012; Heimpel and Hough-Goldstein, 1994; Hildrew and Townsend, 1977; Holling, 1965; Hossie and Murray, 2010; Hunt and Swift, 2010; Isenhour and Yeargan, 1981; Işikber, 2005; Jeschke and Tollrian, 2005; Jones et al., 2007; Kalsi et al., 2014; Khan, 2009; Khan et al., 2016; Kharboutli and Mack, 1993; Koch et al., 2003; Krishnaraj and Pritchard, 1995; Kumar et al., 2014; LeCato, 1978; LeCato and Arbogast, 1979; Lee and Kang, 2004; Losey and Denno, 1998; Loughridge and Luff, 1983; Madadi et al., 2011; Mandour et al., 2006; Maselou et al., 2015; McCaffrey and Horsburgh, 1986; McCoul et al., 1998; McCoy et al., 2011; Messina and Hanks, 1998; Mills and Lacan, 2004; Moayeri et al., 2013; Mohaghegh et al., 2001; Mondal et al., 2014; Montserrat et al., 2000; Moore, 1988; Morales and Burandt, 1985; Mross et al., 2013; Munyaneza and Obrycki, 1997; Murdie and Hassell, 1973; Nachappa et al., 2006; Nagai and Yano, 2000; Nordlund and Morrison, 1990; Ofuya and Akingbohungbe, 1988; Opit et al., 1997; Papanikolaou et al., 2011; Parajulee et al., 1994, 2006; Pervez and Omkar, 2005; Queiroz et al., 2015; Rocha and Redaelli, 2004; Rohlf and Mack, 1984; Sabaghi et al., 2011; Sahayarak et al., 2015; Sahayarak and Ambrose, 1994; Saleh et al., 2010; Saljoqi et al., 2015, 2016; Sarmento et al., 2007; Seko and Miura, 2008; Sentis et al., 2012, 2017; Sharifian et al., 2015; Shrestha and Enkegaard, 2013; Shukla et al., 1990; Siddiqui et al., 2015; Sing and Arbogast, 2008; Sohrabi and Shishehbor, 2007; Soluk, 1993; Spitze, 1985; Stewart et al., 2002; Sueldo et al., 2010; Taylor, 1988; Tellez et al., 2013; Tenhumberg, 1995; Thompson, 1975, 1978; Timms et al., 2008; Trichilo and Leigh, 1986; Uiterwaal et al., 2017; van Lenteren et al., 2016; Veeravel and Baskaran, 1997; Vinyard and Menger, 1980; Wasserman et al., 2016b; Wells and McPherson, 1999; Wiedenmann and O’Neil, 1991; Xue et al., 2009; Yaşar and Özge, 2005; Yaşar and Özge, 2005; Zarghami et al., 2016; Zhang et al., 2013)</td>
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