2019

Dean-flow-coupled elasto-inertial particle and cell focusing in symmetric serpentine microchannels

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Publication Details

Yuan, D., Sluyter, R., Zhao, Q., Tang, S., Yan, S., Yun, G., Li, M., Zhang, J. & Li, W. (2019). Dean-flow-coupled elasto-inertial particle and cell focusing in symmetric serpentine microchannels. Microfluidics and Nanofluidics, 23 (3), 41-1-41-9.

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Abstract

This work investigates particle focusing under Dean-flow coupled elasto-inertial effects in symmetric serpentine microchannels. A small amount of polymers were added to the sample solution to tune the fluid elasticity, and allow particles to migrate laterally and reach their equilibriums at the centerline of a symmetric serpentine channel under the synthesis effect of elastic, inertial and Dean-flow forces. First, the effects of the flow rates on particle focusing in viscoelastic fluid in serpentine channels were investigated. Then, comparisons with particle focusing in the Newtonian fluid in the serpentine channel and in the viscoelastic fluid in the straight channel were conducted. The elastic effect and the serpentine channel structure could accelerate the particle focusing as well as reduce the channel length. This focusing technique has the potential as a pre-ordering unit in flow cytometry for cell counting, sorting, and analysis. Moreover, focusing behaviour of Jurkat cells in the viscoelastic fluid in this serpentine channel was studied. Finally, the cell viability in the culture medium containing a dissolved polymer and after processing through the serpentine channel was tested. The polymer within this viscoelastic fluid has a negligible effect on cell viability.

Keywords: Viscoelastic fluid, Dean-flow coupled elasto-inertial effects, viscoelastic force, cell viability

Introduction

Microfluidic particle focusing has been widely applied in numerous biomedical and chemical applications [1-3], since it is an essential step for downstream particle separation [4-6], counting [7], detection or analysis [8-10]. Particle focusing has been extensively studied in the Newtonian fluid using active and passive methods [11-14]. Active methods, such as dielectrophoresis (DEP) [15, 16], magnetophoresis [17, 18], acoustophoresis [19], and optical tweezers [20], are based on the external force fields. Whereas passive methods rely on intrinsic hydrodynamic forces induced in microchannels with specialized geometry or structures [21-23], such as pinched flow fractionation (PFF) [24], hydrodynamic filtration [25], inertial microfluidics [26, 27], and deterministic lateral displacement (DLD) [28].

The above microfluidic methods for particle focusing are all performed in Newtonian fluids, which employ either external force fields or specially designed complex channels. Recently, there is increasing research interest on particle manipulation in viscoelastic fluids because of its superior focusing performance and simpler channel design [29-35]. The viscoelastic particle focusing has been investigated extensively in straight channels with cylindrical [36-38] and rectangular cross-sections [39-42], spiral microchannels [43-45] and straight channel with side-walls [5, 46].

This work aimed to investigate particle focusing under Dean-flow coupled elasto-inertial effects in a serpentine microchannel. A small amount of polymers were added to the sample solution to allow particles to migrate laterally and reach equilibrium at the centreline of channel under the synthesis effect of elastic, inertial and Dean-flow forces. First, the effects of the flow rates on particle viscoelastic focusing in serpentine channels were investigated. Then, comparisons with particle focusing in the Newtonian fluid in the serpentine channel and in the viscoelastic fluid in the straight channel were conducted. It was found that the elastic effect plays an important role in efficient particle focusing, and the serpentine channel structure

could accelerate the particle focusing as well as reduce the channel length. This focusing technique has the potential as a pre-ordering unit in flow cytometry for cell counting, sorting, and analysis. Additionally, the focusing behaviour of human Jurkat cells in the viscoelastic fluid in this serpentine channel, as well as the cell viability in poly(ethylene oxide) (PEO) dissolved cell culture medium and in PEO dissolved PBS before and after microfluidic processing was tested. To the best knowledge of the authors, this is the first time that the biocompatibility of dissolved PEO polymer on cells in microfluidic focusing experiments has been reported.

Theory

Elastic force

In viscoelastic fluid, the polymer within the medium can induce an additional elastic force on particles. W_i was used to characterize the elastic effects of the viscoelastic fluid, which is the ratio of two time constants [40]:

$$W_i = \frac{\lambda}{t_f} = \lambda\dot{\gamma} = \lambda\frac{2V_m}{w} = \frac{2\lambda Q}{hw^2} \quad (1)$$

where λ defines the relaxation time of the fluid, and V_m and t_f define the average velocity and characteristic time of the channel flow, respectively. The characteristic time is $2V_m/w$ or $2\lambda Q/hw^2$ in a rectangular channel. In viscoelastic fluids, both the first normal stress N_1 ($=\tau_{xx}-\tau_{yy}$) and the second normal stress N_2 ($=\tau_{yy}-\tau_{zz}$) contribute to particle migration. τ_{xx}, τ_{yy}, and τ_{zz} are normal stresses that are exerted in the flow, for the velocity gradient and vorticity direction, respectively. However, the effects of N_2 can be neglected in the diluted polyethylene oxide (PEO) solutions [47, 48], because N_1 is much larger than N_2. The elastic force F_E, which originates from an imbalance in the distribution of N_1($=\tau_{xx}-\tau_{yy}$) over the size of the particle, can be expressed as [31]:

$$F_E = C_{eL}a^3\nabla N_1 = C_{eL}a^3(\nabla\tau_{xx}-\nabla\tau_{yy}) = -2C_{eL}a^3\eta_p\lambda\nabla\dot{\gamma}^2 \quad (2)$$

where C_{eL} is the non-dimensional elastic lift coefficient, a is the particle size, and η_p is the polymeric contribution to the solution viscosity.

Drag force

Assuming a spherical particle travelling in a uniform Stokes fluid, and there is a velocity difference between particle and fluid, a drag force can arise, which can be expressed as [11, 26]:

$$F_D = 3\pi\mu_f a(v_f - v_p) \quad (3)$$

where v_f and v_p are the velocities of the fluid element and particles, respectively, a is the particle size, and μ_f represents the fluid viscosity.

Schematic of the Dean-flow coupled elasto-inertial particle focusing in symmetric serpentine channel.

When particles in viscoelastic fluid flow in a symmetric serpentine channel under a certain flow rate, the particles are experiencing three forces: inertial lift force F_L, the resultant of the shear-gradient lift force (F_{LS}) and wall-repulsion force (F_{LW}); the Dean drag force F_D, resulting from the curved channel geometry; and elastic force F_E, induced by the nature of the viscoelastic medium. When particles in Newtonian fluid flow in straight channels, the shear gradient lift force pushes particles away from the centreline of the channel, while the wall lift force drives particles away from the channel wall. When the particles are not in straight channels but in a symmetric serpentine channel with a Newtonian fluid, two counter-rotating vortices are generated due to the non-uniform inertia of fluid elements within the channel cross-section. Therefore, the Dean drag force, which is perpendicular to the particle velocity, directing from the inner corner to the outer corner, acts in superposition on the particles, and the trajectory of these particles can be confined within a certain width after a certain distance. However, when particles are in the viscoelastic fluid, rather than Newtonian fluid, in a symmetric serpentine channel, the viscoelastic force F_E should arise, which is directing away

from the wall and declining with increasing distance from the wall. After introduction of the elastic force, the particle focusing can be improved. Compared with particle focusing in the Newtonian fluid in the serpentine channel, the addition of elastic force can accelerate the focusing process, as well as improve the focusing performance. With the synergetic effect of inertial lift force F_L, the Dean drag force F_D, and elastic force F_E, the Dean-flow coupled elasto-inertial particle focusing in symmetric serpentine channels can be realized easily in a relatively short length, Figure 1.

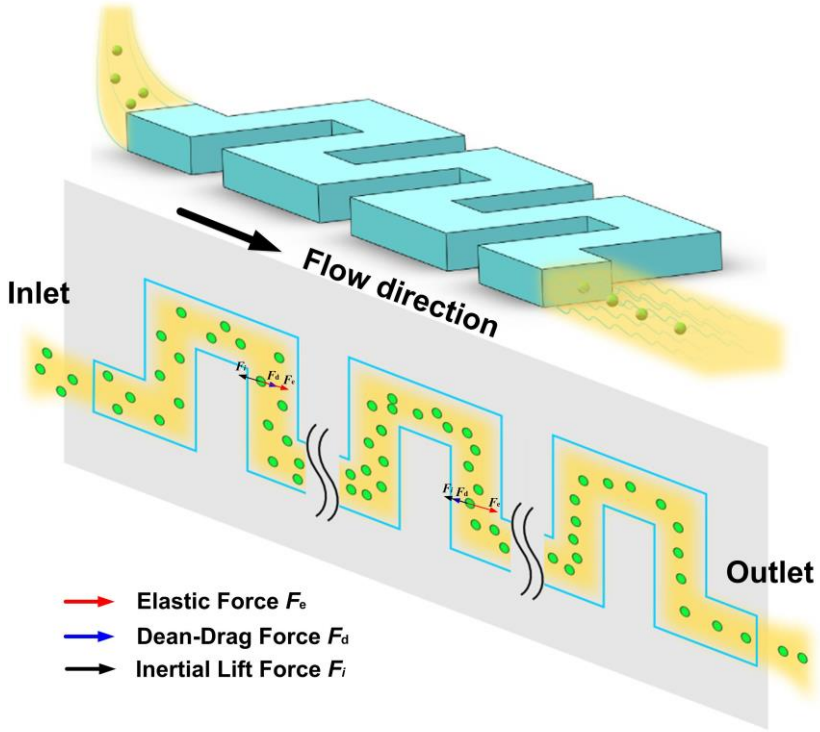

Figure 1: Schematic view of the Dean-flow coupled elasto-inertial particle focusing in the serpentine channels.

Materials and methods

Design and fabrication of a microfluidic device

Standard photolithography and soft lithographic techniques were used to fabricate the following device [49, 50]. The symmetric serpentine channel consisted of a 20 mm serpentine section with 16 zigzag periods. The width of micro-channel was 100 μm. The depth of the channel was uniform at 42 μm. As a comparison, a straight channel with the same width and

height (100 μm × 45 μm) but with longer length (40 mm) was used to investigate the effect of serpentine structure on particle focusing.

Preparation of the PEO medium

For particle focusing, PEO (2,000,000 Da; Sigma-Aldrich) was dissolved in deionized water (DI water) with a concentration of 1000 ppm to form the viscoelastic fluid. For comparisons to Newtonian fluid, the fluid was DI water. Tween 20 (Sigma-Aldrich) was added to DI water with a volume ratio of 0.01% (v/v) to prevent particle aggregation.

For Jurkat cell viability test, PEO was dissolved in cell culture medium (Roswell Park Memorial Institute (RPMI) 1640 medium (Thermo Fisher Scientific) containing 10% fetal bovine serum (Bovogen Biologicals) and 2 mM L-glutamine (Thermo Fisher Scientific)) at various concentrations. For Jurkat cell viscoelastic focusing and viability testing before and after each experiment, PEO was dissolved in phosphate-buffered saline (PBS) with a concentration of 1000 ppm.

Particle and cell preparation

For particle focusing experiments, particle suspensions were prepared by diluting 13 μm internally red dyed fluorescent polystyrene microspheres (Thermo Fisher Scientific, CV5%) in the 1000 ppm PEO medium.

For cell focusing and viability tests, Jurkat cells (ATCC), an immortalized human leukaemic T cell line (average diameter of approximately 15 μm), were cultured in complete (RPMI) 1640 medium in a humidified incubator (Thermo Scientific) at 37^{o}C and 95% air/5% CO_2. Before commencing each experiment, the particle mixture was re-suspended by vortexing and cell samples were manually stirred to provide uniform suspensions in a complete medium containing PEO.

Cell viability test

Live cells have intact membranes that exclude a variety of dyes that easily penetrate the damaged, permeable membranes of non-viable cells. In cell viability tests (including the viability tests of Jurkat cells in a polymer dissolved cell culture medium with different PEO concentrations after incubation, and before and after the viscoelastic microfluidic experiment), fluorochrome 7-amino actinomycin D (7-AAD) (Thermo Scientific) was used to stain non-viable cells. 1µl 7AAD was added to each 300µl Jurkat cell sample, and after 30s, the 7-AAD fluorescence was determined by flow cytometry (BD Accuri C6, BD biosciences). Therefore, the number of viable and non-viable cells can be analyzed based on the 7-AAD fluorescence.

Experimental setup

The sample suspension was transferred to the chip from two 1 ml syringes, with silicon tube connected between them. The flow rate was controlled by syringe pumps (Legato 100, Kd Scientific, USA). An inverted microscope (CKX41, Olympus, Japan) mounted with a CCD camera (Optimos, Q-imaging, Australia) was used to observe and capture images of the fluorescent particles, cells and fluids. The fluorescent images were post-processed and analysed with Q-Capture Pro 7 (Q-imaging, Australia) software.

Results and discussion

Effect of flow rates

Figure 2 shows the 13 µm particle distribution along the channel length from the 1st zigzag turn to the 16th zigzag turn under different flow rates. As seen from the fluorescent images of particle focusing (Figure 2 (a)), when the flow rate ranges from 5 µl/min to 20 µl/min, the particles can be focused to a single line at the last zigzag turn under the effect of Dean-flow coupled elasto-inertial effects. As the flow rate increases to 30 µl/min, the particles become dispersed, which is attributed to the fact that inertial effects become dominant gradually. Besides, the channel length for particles to reach a single line is different for flow rates of 5 µl/min, 10 µl/min and 20 µl/min. Among the three flow rates, particles flowing at 20µl/min first reach their

equilibrium positions after the 6^{th} zigzag turn. This indicates that 20µl/min is the optimal flow rate for particle focusing and this flow rate yields the shortest channel length. Furthermore, as the channel length increases, the particle focusing performance can be improved. Figure 2 (b) is the corresponding fluorescent intensity profiles at the middle of each zigzag turn under different flow rates. Therefore, it is concluded that as the channel length increases, the focusing performance improves; as the flow rate increases, but below 30 µl/min, the particles are focused more quickly; as the flow rate increases to 30 µl/min or above, the particle focusing is broken by the increasing inertial effects.

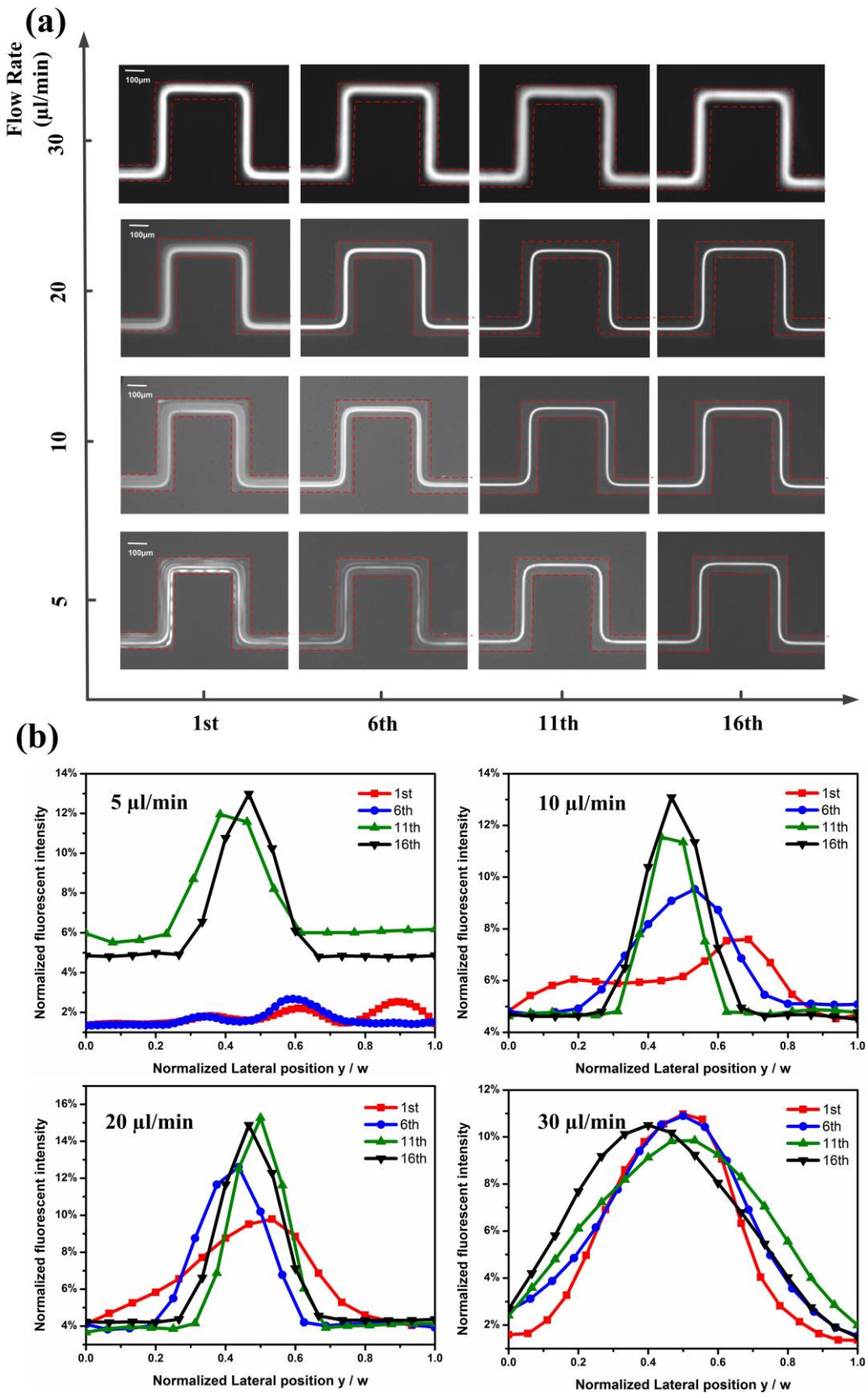

Figure 2. Effects of flow rate on particle viscoelastic focusing in the serpentine channel. (a) Fluorescent images of particle focusing at different zigzag turns under different flow rates. (b) Fluorescent intensity profile at the middle of each zigzag turn under different flow rates (as indicated).

Effect of elasticity

To investigate the fluid elasticity on particle migration, experiments were also carried out in the Newtonian fluid in this serpentine channel. Figure 3 shows the 13 μm particle focusing in the Newtonian fluid in a serpentine channel under the flow rate of 5μl/min, 10μl/min, 20μl/min, and 30μl/min at the 16th zigzag turn and corresponding fluorescence intensity profiles. The particles remain dispersed under all flow conditions. Compared with particle flowing conditions in Figure 2, the only difference is the carrying medium. In viscoelastic fluid, the particles can be focused under the synthesis effects of elastic, inertial and Dean flow forces, while in Newtonian fluid the particles cannot be focused under the effects of inertial and Dean flow forces. This indicates that the elastic effect plays an important role in efficient particle focusing.

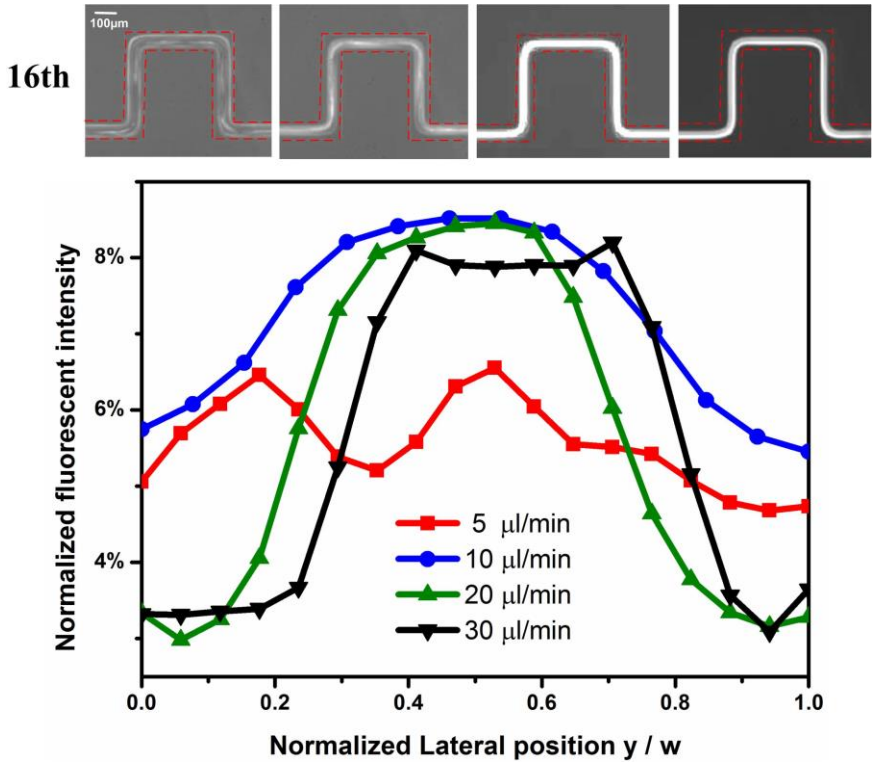

Figure 3. Particle focusing in Newtonian fluid in serpentine channel under flow rates of 5μl/min, 10μl/min, 20μl/min, 30μl/min at the 16th zigzag turn and corresponding fluorescence intensity profiles.

Effect of Dean flow induced by zigzag turns in the symmetric serpentine channel

In order to investigate the effects of Dean flow induced by zigzag turns in symmetric serpentine channel, the 13 μm particle distribution in a straight rectangular channel in the same viscoelastic fluid with a cross-section of 100 μm × 50 μm (width × height) (Figure 4) was compared with that in the serpentine channel (Figure 2). The sole difference between the two channels is the structure. With the zigzag turns, at 20μl/min, the particles are focused into a single line at centerline under the combined effect of Dean-flow coupled elasto-inertial effects, while in straight channels, without the Dean flow effect, the single line focusing cannot be achieved. The particles are dispersed widely at 20 mm from the inlet, and have multiple focusing positions at 30 mm and 40 mm, evidenced by the fluorescent intensity profiles (Figure 4). In comparison, the total length of the serpentine channel is 20 mm, and the particles are focused at the 6th zigzag turns under the same flow rate and viscoelastesity. This indicates that this serpentine channel allows efficient particle focusing in the viscoelastic fluid with the aid of Dean flow.

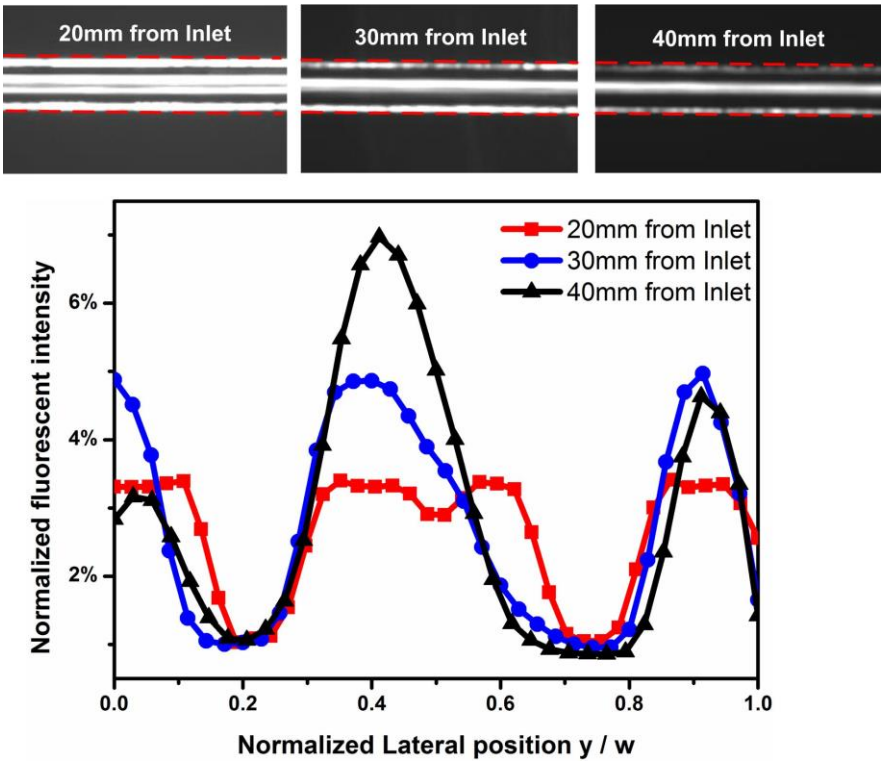

Figure 4. Particle focusing in a viscoelastic fluid in a straight channel under a flow rate of 20μl/min.

Cell viability test

Figure 5 shows the viability of Jurkat cells in cell culture medium containing polymer PEO, with PEO concentrations of 500 ppm and 1000 ppm after 2 h and 4 h incubation, compared to Jurkat cells in cell culture medium alone (0 ppm). The viability of Jurkat cells in 0 ppm, 500 ppm and 1000 ppm PEO dissolved in cell culture medium was similar at each time point, with no reduction in viability observed for up to 4 h incubation. Thus, PEO has little effect on Jurkat cell viability within 4 h of exposure, displaying high biocompatibility over this time period. This allows sufficient time for carrying out viscoelastic focusing or separation experiments, as well as potential downstream analysis.

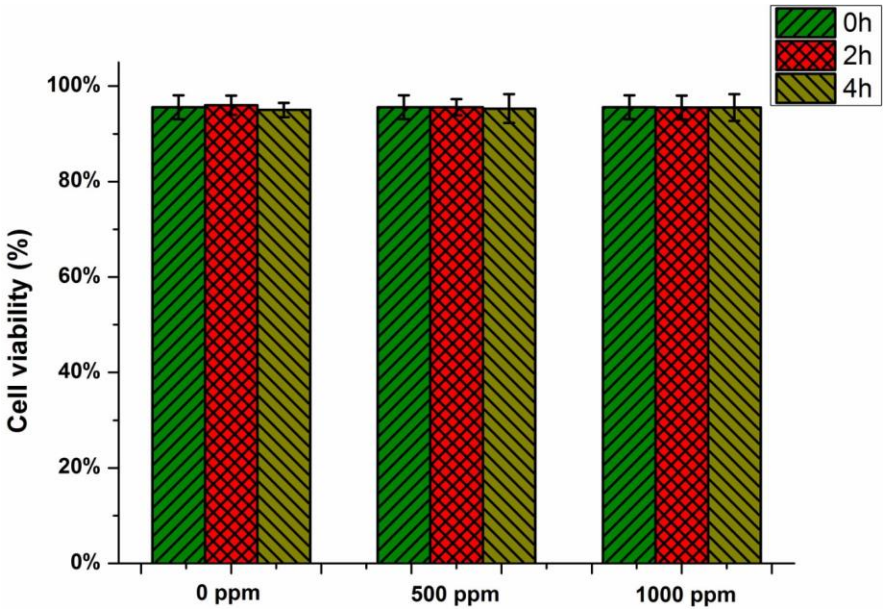

Figure 5. Cell viability test in polymer dissolved cell culture medium with respect to different culture time and polymer (PEO) concentration.

After establishing that PEO has negligible effects on Jurkat cell viability, Jurkat cell migration behaviour in viscoelastic fluids in the serpentine channel, as well as cell viability before and after microfluidic experiments were investigated (Figure 6). PBS containing polymer (1000ppm) was used in place of cell culture medium containing the polymer (since most of the microfluidic experiments involving cells are performed in PBS based fluid). The Jurkat cell distribution at the outlet of the serpentine channel under different flow rates was captured and shown in Figure 6 (a). At flow rates of 5 µl/min and 10 µl/min, cells were focused

to a single line after the 16th zigzag turn, flowing as single events under the Dean-flow coupled elasto-inertial effects. As the flow rate increases to 20 µl/min or higher, the cells could not be focused well and started to disperse due to the increasing inertial effects. Notably, the optimal focusing flow rates for particles and Jurkat cells are different, despite the Jurkat cells being similar in shape and size to the 13 µm particles. The may be attributed to the differences in deformability of polystyrene particles and cells.

The cell viability was tested before and after the microfluidic focusing experiments (Figure 6 (b)). The focusing experiments took approximately 1 hour. Cell viability was tested in PEO dissolved PBS medium before the experiment at 0 h from the inlet for comparison (the left figure), at approximately 1 h from the inlet (the middle figure), and 1 h from the outlet after focusing experiment (the right figure). The viabilities were 88.6%, 84.7% and 76.7% for cells before the experiment at 0 h from the inlet, 1 h from the inlet, and 1h from the outlet after focusing experiment, respectively. It can be seen that without running the microfluidic experiment, Jurkat cell viability was reduced slightly from 88.6% to 84.7% after 1 h exposure to PEO polymer in PBS medium. After the cell focusing experiment, the cell viability was reduced further slightly from 84.7% to 76.7% compared to cells not subject to the microfluidic experiments at corresponding time points. This reduction in viability may be due to the mechanical forces that cells experienced during the microfluidic channels rather than exposure to the polymer dissolved in PBS.

From the viability tests of Jurkat cells in polymer dissolved cell culture medium with different PEO concentrations after incubation, and before and after the viscoelastic microfluidic experiment, we concluded that dissolved polymer has a negligible effect on cell viability. Some other researchers have tested the cell viability in other polymer dissolved mediums. Cha et al. [51] tested whether PVP affects the viability of the stem cells by MTS assay. The results clearly showed no distinctive difference in viability between the PVP

solution and PBS without PVP after 24 h; Besides, Del Giudice et al. [52] evaluated the viability of both Jurkat and NIH 3T3 cells in aqueous HA 0.8 wt % and without PBS, over 90 min, Cell viability for both cell types in HA 0.8 wt % does not change significantly within 90 min. In our experiments, cell viability test in the polymer dissolved cell culture medium with respect to different culture time (0h, 2h, 4h) and polymer (PEO) concentration (0ppm, 500 ppm, 1000 ppm), as well as in polymer dissolved PBS before and after viscoelastic microfluidic focusing experiments were examined. To the best knowledge of the authors, this is the first time that the biocompatibility of dissolved PEO polymer on cells in microfluidic focusing experiments has been reported.

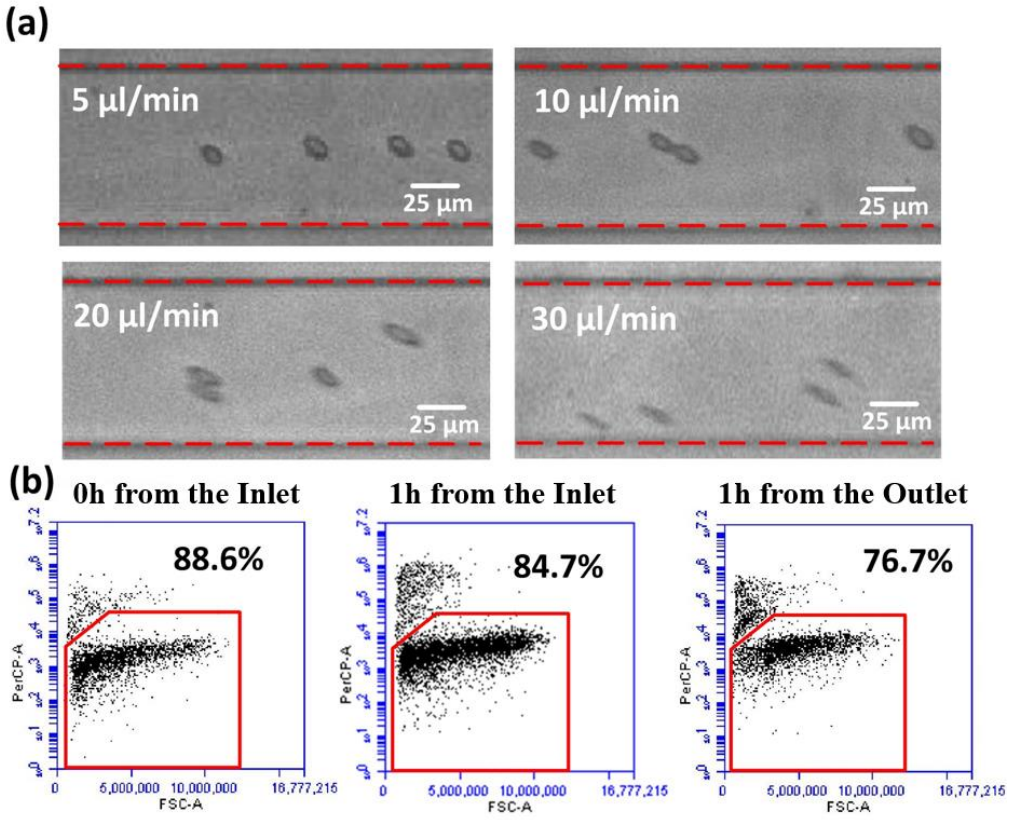

Figure 6. Jurkat cell focusing in the PEO viscoelastic fluid in a serpentine channel. (a) Jurkat cell focusing in the viscoelastic fluid in a serpentine channel under the effect of different flow rates. (b) Cell viability test before and after viscoelastic microfluidic focusing experiments.

Concluding remarks

Particle and cell distribution in a symmetric serpentine microchannel in the viscoelastic

fluid was investigated. Under Dean-flow coupled elasto-inertial effects, particles and cells can be focused into a single line. From the experiments of effects of the flow rates on particle/cell focusing, it is concluded that as the flow rate exceeds its optimal focusing value, the inertial effect becomes dominant and disrupts particle focusing. Comparison of particle focusing in the Newtonian fluid in the symmetric serpentine channel, and in the viscoelastic fluid in the straight channel demonstrated the fluid elasticity and Dean flow induced by zigzag turns in the serpentine channel both play important roles in efficient particle focusing. This focusing technique has the potential as a pre-ordering unit in flow cytometry for cell counting, sorting, and analysis. Additionally, the Jurkat cell viability test in the polymer dissolved in cell culture medium, and the cell viability before and after microfluidic experiments indicated that the dissolved PEO polymer within the viscoelastic fluid has minimal effect on cell viability, while the microfluidic focusing may have a minor negative effect on cell viability. The cell viability tests with regard to polymer in viscoelastic fluid provide general information regarding the biocompatibility of the polymer PEO in aqueous solutions.

Acknowledgements

This work is supported by the National Natural Science Foundation of China (Grant No.51705257), the Australian Research Council (ARC) Discovery Project (Grant No. DP180100055), and the Natural Science Foundation of Jiangsu Province (Grant No. BK20170839).

The authors have declared no conflict of interest.

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