

REVIEW

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Dengue and COVID-19: two sides of the same coin



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Abstract

Background: Many countries in Asia and Latin America are currently facing a double burden of outbreaks due to dengue and COVID-19. Here we discuss the similarities and differences between the two infections so that lessons learnt so far from studying both infections will be helpful in further understanding their immunopathogenesis and to develop therapeutic interventions.

Main body: Although the entry routes of the SARS-CoV-2 and the dengue virus (DENV) are different, both infections result in a systemic infection, with some similar clinical presentations such as fever, headache, myalgia and gastrointestinal symptoms. However, while dengue is usually associated with a tendency to bleed, development of micro and macrothrombi is a hallmark of severe COVID-19. Apart from the initial similarities in the clinical presentation, there are further similarities between such as risk factors for development of severe illness, cytokine storms, endothelial dysfunction and multi-organ failure. Both infections are characterised by a delayed and impaired type I IFN response and a proinflammatory immune response. Furthermore, while high levels of potent neutralising antibodies are associated with protection, poorly neutralising and cross-reactive antibodies have been proposed to lead to immunopathology by different mechanisms, associated with an exaggerated plasmablast response. The virus specific T cell responses are also shown to be delayed in those who develop severe illness, while varying degrees of endothelial dysfunction leads to increased vascular permeability and coagulation abnormalities.

Conclusion: While there are many similarities between dengue and SARS-CoV-2 infection, there are also key differences especially in long-term disease sequelae. Therefore, it would be important to study the parallels between the immunopathogenesis of both infections for development of more effective vaccines and therapeutic interventions.

Keywords: Dengue, SARS-CoV-2, COVID-19, Severe illness, Immunopathology, Innate immunity, Antibodies, T cells, Endothelial dysfunction

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Background

Although SARS-CoV-2 is reported to have infected over 500 million individuals with at least 6.2 million individuals succumbing to COVID-19 by April 2022 [90], the true direct and indirect death toll due to COVID-19 is estimated to be much higher [126]. Despite the availability of several safe and effective vaccines for COVID-19, the emergence of SARS-CoV-2 variants that evade immunity has posed challenges in controlling outbreaks [33]. Due to the unprecedented cooperation between scientists, sharing of data and availability of funding, by early 2022

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ten vaccines had received emergency use licensing by the WHO for the prevention of COVID-19 [128]. This contrasts with many other neglected tropical infections such as dengue, despite causing deaths in 0.53 per 100,000 population in 2017 [139].

Although there are reports of epidemics caused by infection with the dengue virus (DENV) in the 1780s, epidemics resulting in dengue haemorrhagic fever (DHF) or dengue shock syndrome (DSS), which are severe forms of dengue infection, was initially reported in the 1950s in South East Asia [69]. However, dengue infections have gradually increased over time due to many factors such as climate change resulting in increase in temperatures, urbanization, increase mobility and overcrowding [108]. Although there is no specific treatment for dengue, intense monitoring to detect vascular leak and other supportive management has reduced case fatality rates (CFRs) to < 0.3% in most countries [19, 83], although in some countries such as in India the CFRs are estimated to be 2.6% [95]. The CFRs in patients with severe dengue was shown to be around 5.9% for younger children while it was as high as 32.6% in patients > 60 years of age in Brazil [79]. Therefore, it is evident that dengue is an important cause of morbidity and mortality in countries in the tropical and subtropical regions.

While SARS-CoV-2 infects individuals via the respiratory route, the DENV infects individuals following a bite of an infected mosquito of the Aedes species. However, it is well established that COVID-19 is not a mere respiratory infection but is a systemic illness. Furthermore, many of the initial clinical symptoms such as fever, myalgia, joint pain, headache, lethargy, abdominal pain, diarrhoea, vomiting and sometimes sore throat is frequently seen in both infections, making it difficult to clinically differentiate dengue from COVID-19 during early illness [86, 129]. Therefore, many countries that experienced regular outbreaks due to dengue are now faced with the double burden of dengue and COVID-19 [47, 52]. Apart from the initial similarities in the clinical presentation, there are many similarities between these two infections such as certain risk factors for severe illness, immunopathogenesis, antibody and T cell responses, cytokine storms, endothelial dysfunction and multi-organ failure. However, there are certain differences such as haemorrhage in dengue compared to thrombosis occurring in COVID-19. In this review, we discuss the similarities and differences between the two infections so that lessons learnt so far from studying both infections will be helpful in further understanding their immunopathogenesis and to develop therapeutic targets.

Risk factors for severe illness in dengue and COVID-19

While dengue was predominantly a childhood infection many years ago, there has been a gradual shift in the age of infection in many countries [63, 83, 115]. Therefore, currently more severe forms of dengue (DHF/DSS) are predominantly seen in the older population in some countries, where CFRs tend to be higher than in younger individuals [79, 83]. However, some studies have shown that CFRs are higher in children, and these differences possibly reflect the differences in disease epidemiology in different countries [119]. Men were significantly more likely to have severe illness in COVID-19, whereas no such associations have been seen with dengue [39, 60]. The presence of metabolic disease, diabetes, hypertension, chronic kidney disease and obesity have shown to be independently associated with the development of more severe illness [55, 72, 102, 110] (Fig. 1). Similarly, mortality rates are higher in elderly individuals and in those with comorbidities when infected with SARS-CoV-2 and

Dengue	Dengue and COVID-19	COVID-19
Age	Age	Gender
All ages	Older age	Male sex
Immune status	Physiological status	Comorbidities
Secondary infection	Pregnancy	Cardiovascular disease
,	J ,	Chronic lung disease
Viral factors	Comorbidities	Immunosuppression
All four dengue viruses cause severe	Diabetes	Cancers
dengue	Obesity	
DENV4 infection milder?	Hypertension	Viral factors
	Asthma	Wide differences in hospitalizations
	Chronic kidney disease	and mortality due to different variants
		in different countries.

Fig. 1 Risk factors for severe dengue and COVID-19. The common changes are highlighted in the middle box, while those specific to dengue (green box) and COVID-19 (blue box) are shown separately

influenza. However, in contrast to COVID-19, influenza and many other respiratory infections also cause severe disease in younger children [27, 95]. There are other differences in risk factors for COVID-19 compared to dengue. For instance, while those who were immunosuppressed or those with malignancies were significantly more likely to develop severe COVID-19, whereas such individuals are not at higher risk of severe dengue [39].

COVID-19 or dengue in pregnancy are also associated with a higher risk of severe disease and higher mortality rates [29, 101]. Maternal death was 450 times higher in pregnant women with DHF compared to dengue in non-pregnant females [101] and dengue in pregnancy was associated with a high incidence of acute renal and liver failure, acute respiratory distress and an increased need for ventilatory support [15]. Dengue in pregnancy was associated with a higher incidence in preterm delivery, still birth and low birth weight neonates, similar to COVID-19 in pregnancy, while both infections were not associated with congenital abnormalities of the fetus [100, 124].

Although mechanisms underlying the increased risk of severe dengue and COVID-19 in the elderly, those with comorbidities and in pregnant women are not entirely clear, it could be due to multiple causes such as immunosenescence, an aberrant immune response, pre-existing endothelial dysfunction worsening disease outcome and many other factors [49]. It would be important to further investigate the mechanisms by which these vulnerable groups are more prone to severe illness to provide better preventive methods and treatment modalities.

Interestingly, there has been there are geographical differences in mortality and morbidity due to both dengue and COVID-19. For instance, while the incidence of dengue is similar in many Asian countries and South America, the age standardized death rates and DALYs are significantly lower in South America [139]. Although the reasons for these differences in mortality and morbidity is not known, it could be due to the differences in the DENV that circulate in different regions, vector competence in transmission, force of infection and age of population affected. Similarly, the hospitalizations and mortality rates due to COVID-19 have shown to vary widely between many countries, which could be attributed to differences in COVID-19 vaccination rates, circulating variants, age of population and reporting of COVID-19 deaths [21]. Therefore, although many Asian and African countries have reported lower mortality rates than Europe and Northern America, despite lower vaccination coverage, this is possibly due to inaccurate reporting as many of these countries have reported high levels of excess mortality [21]. However, some countries in Asia (e.g. Sri Lanka) reported lower mortality rates during the omicron wave compared to Europe and North America, with lower excess mortality rates than these countries, despite significantly lower vaccine coverage [21, 46]. Therefore, it would be important to further investigate the reasons for differences in mortality rates in different populations in different geographical regions for both infections.

Infection characteristics due to dengue and COVID-19

Infections with the DENV occurs following feeding by an infected mosquito of the Aedes species, where the virus infects many innate immune cells and keratinocytes. The time from onset of infection to onset of symptoms (incubation period) for the four DENVs has shown to a mean of 5.9 days [35, 96]. Controlled human challenge models have shown that the duration of viraemia was a mean of 6.8 days, with those who were challenged with a high dose of virus having a longer viraemia than those challenged with a lower dose [35]. Although many different cells have shown to be permissive to infection by the DENV in cell culture [97], autopsy studies have shown that apart from immune cells such as monocytes, dendritic cells, mast cells, the DENV readily infected hepatocytes, kuffer cells, alveolar macrophages, and macrophage like cells in the lymph nodes and spleen [7, 11]. Although some autopsy studies have shown the presence of viral antigens in neurons, kidney cells and endothelial cells, evidence of viral replication within these cells have not been demonstrated [7, 11].

SARS-CoV-2 virus initiates infection by entering cells expressing ACE2, and engagement of the receptor binding domain (RBD) of the virus with ACE2 exposed the cleavage site in S2, which is subsequently cleaved by TMPRSS2 [53]. Following infection of the ciliated cells in the nasal epithelium and type II alveolar cells, the incubation period has shown to be on average 6.38 days, ranging from 2.33 to 17.6 days before patients show symptoms, based on a meta-analysis [34]. However, the incubation period of omicron BA.1 and BA.2 sub-lineages was shown to be shorter than delta and previous variants, which could have contributed to their higher transmissibility [91, 123]. ACE2 is expressed on many cells in addition to the ciliated cells in the nasal epithelium and type II alveolar cells in the lungs, which are initially infected with the virus. Due to the wide expression of ACE2, the SARS-CoV-2 virus has shown to infect the enterocytes, cells in the kidney, heart muscle and testis [53]. The increased susceptibility to severe disease has been attributed to different levels of expression of ACE2 in those with comorbidities [53]. Neuronal cells have been infected in vitro and in animal models, and cerebral atrophy and tissue damage in cortical areas of the brain

has been observed in SARS-CoV-2 infected individuals, suggesting that the virus may directly infect the brain [31, 107]. However, similar to the observations in dengue, although endothelium dysfunction plays a significant role in the pathogenesis of COVID-19, there is limited evidence that the virus infects the endothelium in acute SARS-CoV-2 infection [110].

In summary, both the DENV and SARS-CoV-2 infects many type of immune cells and many organs in the body, leading the a widespread infection. While evidence of active replication of the SARS-CoV-2 of the myocardium, kidney, spleen and intestines have been demonstrated, such evidence of active replication within these organs is not seen in dengue, due to the smaller number of studies which has explored this.

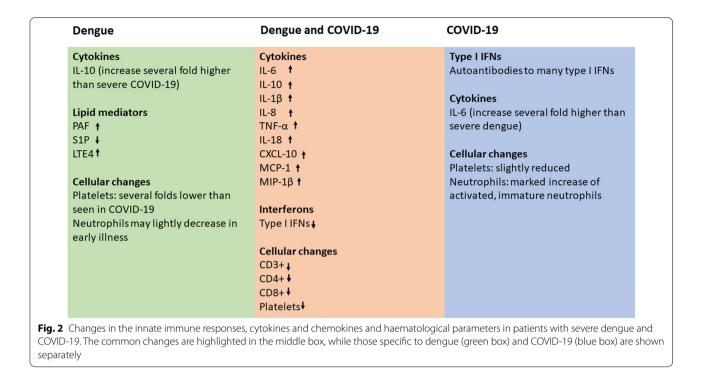
Innate immune responses and cytokine storms in dengue and COVID-19

Although SARS-CoV-2 initiates infection by infecting many different types of cells in the respiratory epithelium, it is known to cause a systemic infection in some individuals infecting the gastrointestinal tract, heart, brain and many other organs [114]. The DENV is also known to infect monocytes, dendritic cells, hepatocytes, keratinocytes and many other cells, while monocytes were shown to be the cell most susceptible to the virus [84]. A dysfunctional immune response by monocytes and other innate immune cells resulting in a delayed interferon response, an increase in proinflammatory cytokines and chemokines such as IL-1- β , TNF- α , CXCL-10, IL-10, IL-18, IL-8 with an increase in many inflammatory lipid mediators is seen in patients who progress to develop severe forms of dengue (DHF) [57, 61, 84]. An impaired and delayed IFN response has shown to associate with a prolonged viraemia and progression to severe disease in COVID-19, and the high levels of IFNs later in the disease were seen to worsen the immunopathology (Fig. 1) [102]. Many DENV proteins inhibit type I IFN production by inhibiting Tyk2 activation and STAT1 phosphorylation, downregulation of STAT2 phosphorylation and by inhibiting STAT2 phosphorylation [17]. In vitro, treatment of HepG2 cells with IFN α and IFN β prior to infection with the DENV was shown to significantly reduce viral loads in these cells [25]. Therefore, type I interferons appear to play an important role in inhibition of DENV replication and in fact, those who proceeded to develop severe dengue were shown to have reduced levels of plasma IFNa and IFN β [120]. As seen in dengue, type I IFN responses were shown to be impaired in those who progress to develop severe COVID-19 [72, 102]. Many SARS-CoV-2 structural and non-structural proteins inhibited the type I IFN response by Tyk2 activation, STAT1 and STAT2 phosphorylation and inhibition of IFN signaling [36, 38, 67]. Apart from the SARS-CoV-2 evading immunity by blocking type I IFN production, patients who progress to develop severe COVID-19 have shown to have autoan-tibodies against IFN α and other type I IFNs [9]. Neutralizing autoantibodies to IFNs, had not been detected in individuals with mild or asymptomatic COVID-19, highlighting the importance of type I IFNs in protection against severe COVID-19[9]. Therefore, an impaired and delayed type I IFN response leads to severe dengue and COVID-19.

Similar to dengue, those who proceed to develop severe COVID-19 have high levels of many proinflammatory cytokines such as IL-6, IL-1β, IL-10, CXCL-10, MCP-1 and the cytokine storm is shown to associate with both severe dengue and COVID-19 (Fig. 2) [137]. Although similar types of cytokines and chemokines are elevated in both dengue and COVID-19, there are many differences in the relative changes of these mediators [23]. For instance, IL-6 levels were shown to correlate with clinical disease severity in COVID-19 and IL-6 receptor antagonists were shown to improve outcomes including survival in critically ill patients [50, 140]. Although IL-6 levels were high in patients with dengue who proceeded to develop DHF, IL-6 levels were several fold lower in the critical phase in patients with dengue compared to COVID-19 (Fig. 1) [23]. High levels of IL-10 levels in early illness in patients with COVID-19 were shown to associate with poorer disease outcomes and was shown to be a predictor of severe disease along with IL-6 [45, 141]. IL-10, which is a potent immunosuppressive cytokine has also shown to act as a proinflammatory cytokine, when elevated with other cytokines [51]. IL-10 is thought to contribute to disease pathogenesis in COVID-19 by either due to its action as an immunostimulatory molecule or because of inability to suppress the hyperinflammation state [51]. However, dengue patients who proceeded to develop DHF had several fold higher levels of IL-10 (mean levels 1331 pg/ml) when compared to those who developed severe COVID-19 (mean 57.3 pg/ml) [23]. In dengue, IL-10 levels in early illness were an important predictor of developing severe disease [23]. IL-10 was shown to suppress DENV specific T cell responses, which could contribute to disease pathogenesis [82].

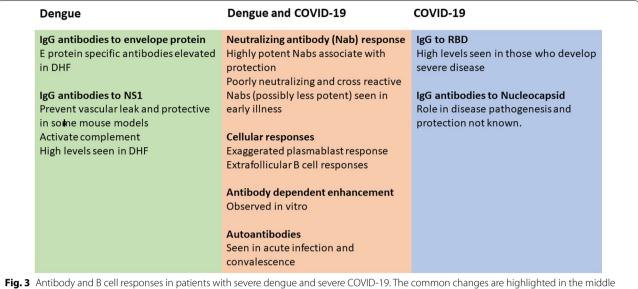
Changes in haematological parameters in dengue and COVID-19

The viraemic/febrile phase of dengue is characterized by a leucopenia with a slight decline in platelet counts [127]. Marked leucopenia with a drastic reduction in platelet counts is seen in those who progress to develop DHF, with a rise in the haematocrit due to fluid leakage [127].



Leucopenia is considered a warning sign of development of severe dengue and lymphopenia has shown to correlate with clinical disease severity [54, 127]. The lymphopenia in dengue is predominantly due to reduction in T cells due to apoptosis, although a reduction in B cells was also seen (Fig. 2) [81]. Although the mechanisms of T cell apoptosis are not clear, serum IL-10 correlated with T cell apoptosis, while inversely correlating with T cell numbers [81]. Lymphopenia is also seen in COVID-19 and is shown to correlate with clinical disease severity [20]. While lymphopenia is mainly due to reduction in CD8+ T cells, reduction in all types of lymphocytes (CD4+ T cells, B cells and natural killer cells), is seen in patients with severe COVID-19 (Fig. 2) [20]. In contrast to dengue, severe COVID-19 is associated with a marked increase in neutrophils in the nasopharynx, lung and in blood, which are highly activated and show an immature phenotype [105]. Neutrophils have been shown to contribute to disease pathogenesis in part by release of neutrophil extracellular traps (NET), which was a result of NLRP3 activation [8]. Although an increase in neutrophils is not observed in dengue, NLRP3 activation in many types of cells has been observed with an increase in NET components in the serum of patients with DHF, suggesting that activation of neutrophils is likely to play a role in severe dengue [48, 62, 99].

Antibody responses in patients with dengue and COVID-19 High levels of neutralizing antibodies (Nabs) following vaccination has shown to prevent infection with the SARS-CoV-2 to a certain degree and associate with protection [66]. DENV serotype specific Nabs have shown to protect against re-infection with the same serotype, while higher levels have also shown to offer protection against symptomatic disease for infection with other serotypes [65]. However, the Nabs antibodies and antibodies directed against the envelope and NS1 protein in dengue can also associate with disease pathogenesis as shown in some studies [24, 55]. The risk of developing DHF is substantially higher in a secondary dengue infection, in which the individual is infected with a different DENV serotype than the earlier infecting serotype [37]. This increase in disease severity is thought to be due to antibody dependent enhancement (ADE), where poorly neutralizing, highly cross-reactive antibodies enhance DENV infection in FcyR-expressing cells [64, 113]. Internalization of these antigen-antibody complexes further leads to disease pathogenesis by inducing IL-10 production by monocytes [118] and DENV-specific afucosylated IgG1 subclass of antibodies further enhanced infection by binding to the activating FcyRIIIA type Fc receptors [122]. Antibodies to NS1, which is a secretory protein of DENV has shown to be protective in some mouse studies [10], while other studies show that NS1 antibody levels are elevated in those who develop DHF [55], and that these contribute to vascular leak by cross-reacting with



box, while those specific to dengue (green box) and COVID-19 (blue box) are shown separately

endothelial cells and inducing apoptosis [75] and by activating complement [6] (Fig. 3). However, as most individuals develop asymptomatic or mild dengue despite the presence of antibodies to the previous DENV serotypes, the type, quality and quantity of DENV specific antibodies that associate with protection is not known.

ADE has been widely discussed in SARS-CoV-2 infection and shown to occur in vitro through multiple mechanisms such as C1q mediated ADE in Vero E6 cells [98], in FcyRIIB expressing B cells [121] and through FcyRIIA and FcyRIIIA receptors in monocytes [80]. However, the clinical significance of ADE in SARS-CoV-2 infection is not known. Although Nabs have shown to be protective in SARS-CoV-2 infection, high levels of virus specific Nabs antibodies and receptor binding domain specific antibodies during early illness have been associated with a worse disease outcome in some studies (Fig. 3) [59, 78, 138]. The extent to which the antibody levels are secondary to high levels of viral replication is not fully resolved. Nabs measured by the surrogate Nabs test (sVNT) showed that in fact, the antibody responses were highest and appeared earlier in those who succumbed to their illness [59]. However, other studies have shown that the presence of highly potent Nabs during early illness was associated with early virus clearance and improved survival [27, 41]. However, unlike in dengue, where infection with a different DENV serotype is a risk factor for severe disease, possibly due to ADE, this has not been seen with SARS-CoV-2 infection. Although there are several different SARS-CoV-2 variants of concern, and many individuals have been infected with a different variant than the one that cause the initial infection, this has not shown to predispose to severe clinical disease [4]. In fact, immune responses generated by natural infection were shown to be longer lasting that those induced by vaccination, and prior natural infection was shown to protect against development of severe clinical disease [4].

A high frequency of activated plasmablasts is seen in patients with severe COVID-19 [12], which is also a feature of severe dengue [40, 132, 136]. Nabs in COVID-19 were shown to be generated by extrafollicular B cells, which correlated with disease severity [135]. Therefore, while high antibody titre may be secondary to high levels of viral replication, it is also possible that antibodies that are generated by an extrafollicular B cell response are less potent and therefore, instead of neutralizing the virus efficiently, they may lead to disease pathogenesis by multiple mechanisms [135]. Extrafollicular B cell responses are also seen in systemic lupus erythematosus and such responses have shown to generate autoreactive antibodies, which is also a feature in COVID-19 [22, 134]. Although the presence of extrafollicular B cell responses have not been studies in dengue infection, poorly neutralizing Nabs have shown to associate with severe disease [24]. Furthermore, a high prevalence of antinuclear antibodies have been detected in the convalescent phase of patients with acute dengue [43]. Therefore, in both COVID-19 and in dengue, while certain types of virus specific antibodies may appear to contribute to disease pathogenesis by many different mechanisms, highly potent Nabs appear to be protective. In order to develop safer and effective vaccines, it would be important to

further study the type, quantity and quality of antibody responses that associate with protection, including the type and mechanisms of antibodies that lead to disease pathogenesis.

T cell responses in dengue and COVID-19

For many years DENV specific cross-reactive T cells were thought to be involved in disease pathogenesis [5, 30, 93]. Although patients with more severe forms of dengue had a higher magnitude of cross-reactive T cells, these were only detected during the convalescence period, and were either not detected or were seen in very low frequency in the critical phase [32]. Recent studies have highlighted the importance of DENV specific T cells in reducing disease severity and have shown that early appearance of virus specific T cells correlated with resolution of viraemia and with less severe disease (Fig. 4) [130]. Those with HLA types that were associated with more severe dengue had a lower frequency of DENV specific T cell responses, whereas T cell responses specific for the HLA alleles associated with protection were significantly higher dengue infections [125]. Furthermore, it was shown that the presence of multiple cytokine producing polyfunctional T cells was associated with milder dengue [131].

Acute SARS-CoV-2 infection is associated with a varied magnitude and functionality of the T cell response. Those who had severe disease had a preferential loss of CD8+ T cells compared to CD4+ T cells, a T cell phenotype characterized by activated CD4+ and CD8+ T cells, and CD8+ T cell displaying T cell exhaustion markers such as PD-1, CTLA-4, LAG3 and TIM-3, and a reduced frequency of follicular helper T cells (Fig. 4) [26, 70, 88]. Several negative T cell regulatory molecules such as CTLA-4, LAG3 and PD-1 have shown to be upregulated in patients with dengue [3, 18]. In both infections, CD8+ T cells of those with severe illness had a reduced cytokine production upon stimulation with mitogens and with peptides [18, 82, 142]. As patients with severe COVID-19 had either an unresponsive or suboptimal virus-specific T cells or an exaggerated T cell response, it would be important to understand the T cell responses that associate with protection for better vaccine design.

SARS-CoV-2 virus specific cross-reactive T cells have shown to be present in unexposed donors and are thought to be due to the presence of cross-reactive T cell responses for other seasonal human coronaviruses [87]. In SARS-CoV-2 infection, the presence of IL-2 producing cross-reactive T cells was shown to be protective in house-hold contacts and was associated with a negative PCR [68]. Furthermore, the presence of these immunodominant CD4+ T cell epitopes that cross react with SARS-CoV-2 and other human coronaviruses have shown to decline with age, which has been suggested as a contributory factor for severe disease in the elderly [77]. Although omicron and its emerging sub-lineages completely escaped antibody mediated immunity induced by two doses of many of the COVID-19 vaccines, they were still shown to be protected by severe disease and hospitalization due to the presence of robust T cell responses

Dengue	Dengue and COVID-19	COVID-19
Phenotype of T cells	Virus specific T cell response	T cell phenotype
Upregulation of exhaustion markers:	Early appearance associated with	Widely varied phenotype of T cells
do not correlate with clinical disease	virus clearance	seen in mild vs severe disease
severity	Delayed T cell response severe disease	Some patients with mild or severe
,	Polyfunctional T cell protective	disease had high activation of T cells
Cross reactive memory T cells	,	
Wide range of functionality shown	Phenotype of T cells	
Phenotype and functionality	Activation markers 1	
associated with protection not	Upregulation of exhaustion markers:	
known.	PD-1, LAG3, CTLA-4, TIM-3 1	
	10 1, 1, 1, 1, 1, 1, 1, 1, 1, 1, 1, 1, 1,	
	Response to antigen	
	Virus specific responses \downarrow	
	Mitogen responses 4	
	Wittogen responses ¥	
	Cross reactive T cells	
	T cells cross react with other	
	serotypes/ human coronaviruses	
	Magnitide of responses protective	
	magnitue of responses protective	

specific to dengue (green box) and COVID-19 (blue box) are shown separately

[94]. Previous infection and many of the COVID-19 vaccines, were shown to induce a high magnitude of T cell responses with a broad recognition of T cell epitopes of many of the viral proteins [94]. T cells have shown to play an important role in viral clearance especially in the context of low antibody levels in non-human primates [89]. Therefore, although the emergence of sub-lineages of omicron such as BA.4 and BA.5 are shown to be more immune evasive than all existing variants, as a result of further mutations in the Nab binding sites. However, due to the breadth of the T cell response induced following natural infection and vaccination, infection with these variants is likely to associate with a relative reduction in clinical disease severity, despite infection, as evidenced with the global decline in COVID-19 mortality rates [16].

Although many studies have been carried out in understanding the T cell responses in SARS-CoV-2 infection, the significance of DENV-specific cross-reactive memory T cell responses subsequent clinical disease severity when infected with the DENV has not been extensively studied. However, those with varying severity of past dengue, were shown to have varied frequencies of T cells and T cell functionality [56], possibly related to the timing of the previous infection and the number of previous DENV infections.

Endothelial dysfunction in dengue and COVID-19

Endothelial dysfunction leading to vascular leak in the hallmark of DHF, which occurs due to viral factors, cytokines and inflammatory mediators [85]. The vascular leak phase which lasts for 24 to 48 h in dengue, occurs around 3 to 5 days since the onset of illness, and this leads to pleural effusion, ascites and shock [85]. The dengue NS1 protein, which is a secretory protein has shown to contribute to endothelial dysfunction by disrupting the endothelial glycocalyx layer [104]. Many inflammatory lipid mediators such as platelet activating factor (PAF), vascular endothelial growth factor (VEGF), angiopoietin-2 (Ang-2) has shown to cause endothelial dysfunction and phospholipase A2 enzymes that generate PAF were shown to be elevated during early illness in dengue [57, 58, 85]. Dengue NS1 was also shown to induce phospholipase A2 enzymes, inflammatory cytokines and prostaglandins, further contributing to endothelial dysfunction [112].

Endothelial dysfunction is also a feature of severe COVID-19 with high levels of VEGF, Ang-2, osteopontin, although the vascular leak is not as prominent as seen in dengue [106]. In comparison to those who died of acute respiratory distress syndrome (ARDS) in influenza infection, those who died of COVID-19 had severe endothelial injury with membrane disruption along with the presence of virus within the endothelial cells [1, 14]. Autopsy

studies have shown the presence of venous and arterial platelet–fibrin microthrombi in many organs in many patients with COVID-19, which is thought to occur due to endothelial activation [1]. Many factors are thought to contribute to the endothelial dysfunction and occurrence of a prothrombotic state, such as direct infection of the endothelium with SARS-CoV-2 causing endothelial damage, inflammatory cytokines such as IL-6, hyperplasia of the endothelium due to lung tissue ischaemia and activation of neutrophils and monocytes along with platelets facilitating microthrombi formation [14, 106].

The coagulopathy in COVID-19 is associated with high levels of D-dimer, fibrinogen and von Willebrand factor, with modest reductions in platelet counts and slightly prolonged or normal prothrombin and activated partial thromboplastin times (APPT) [14, 116]. In contrast to COVID-19, dengue is associated with a bleeding tendency, with marked reductions in platelet counts, with prolonged prothrombin and APPT in patients with severe disease [2, 127]. However, those with DSS and severe COVID-19 had elevated levels of thrombomodulin, plasminogen activator inhibitor type 1 and von Willebrand factor antigen suggesting that like COVID-19, activation of procoagulant mechanisms also occur in severe dengue [28, 44, 106, 133]. Therefore, although the extent of endothelial dysfunction in both infections is a marker of clinical disease severity, the pathogenesis of endothelial dysfunction and the coagulation disturbances that occur as a result of this appear to be different.

Long term sequel of dengue and COVID-19

Many individuals report fatigue following dengue infections with 32% reporting fatigue at 2 months post-infection [111]. A smaller study showed that approximately 50% of individuals who had symptomatic dengue have persistent symptoms such as muscle and joint pain, headache and insomnia, 2 years post-infection, although there was no control group included in this study [42]. The persistence of symptoms was associated with polymorphisms of the FcyRIIa gene, presence of anti-nuclear antibodies and immune complexes [42]. An increased incidence of several different types of autoimmune disease such as Reiter's syndrome, myasthenia gravis, autoimmune encephalomyelitis and systemic vasculitis have been reported following dengue [73]. In Mexico, the annual burden due to persistent symptoms following dengue has shown to cost US\$ 22.6 million annually [117]. However, the proportion of individuals who develop chronic fatigue and the pathogenesis of these long-term complications in dengue has not been studied.

The long-term sequelae of COVID-19 is well recognized and the term 'long COVID (post-acute sequel of COVID-19)' is commonly used to describe the symptoms that occur following COVID-19. Although long COVID is more frequent following severe illness, it has also been reported in those with mild illness with chronic fatigue, persistent lung symptoms, olfactory symptoms, neurological, gastrointestinal symptoms and endocrine abnormalities being described, lasting for months postinfection [13, 92]. 52% of young adults with mild illness (home isolated) reported symptoms of fatigue, dyspnoea, cognitive dysfunctions and loss of taste and smell, at 6 months post-infection [13]. Abnormal lung function with fibrosis and structural changes, myocarditis, thromboembolism, chronic kidney disease, skin disease and structural changes in the brain [31, 92, 109]. Although the pathogenesis of long COVID appears to be multifactorial, persistence of the virus, alteration of immune homeostasis leading to persistent activation of the immune system, alternation of immunometabolic pathways and autoimmunity have been proposed as possible mechanisms [92, 103]. Although myocarditis (11.3%) [74], acute liver failure (0.31%) [71] and acute renal failure (2 to 5% of patients with severe dengue) [76] is reported to occur in dengue, long term organ dysfunction has not been reported in those who recover. Therefore, although chronic fatigue, the presence of certain autoantibodies and an increased risk of autoimmune diseases have been reported following dengue, dengue does not appear to associate with the occurrence of severe disabling and serious long-term sequelae seen following SARS-CoV-2 infection.

Conclusions

The concurrent occurrence of dengue and COVID-19 outbreaks in many Asian and Latin American countries are likely to cause a significant burden to the health care systems of these resource poor countries. Since the initial clinical presentations of these two infections are quite similar, it would be a challenge to clinically differentiate these two infections. In addition, there are many similarities in the immunopathogenesis between dengue and SARS-CoV-2 infection with a dysfunctional immune response leading to a cytokine storm and immunopathogenesis, delayed virus specific T cell responses failing to clear the virus, extra follicular B cell responses and exaggerated plasmablast responses associating with severe disease and endothelial dysfunction. However, while dengue is usually associated with a tendency to bleed, development of micro and macrothrombi is a hallmark of severe COVID-19. Apart from the differences in coagulation abnormalities and the differences in the cytokine storms and mediators that cause endothelial dysfunction, there are also many differences especially in long-term disease sequelae. Although both infections occur due to very different routes (respiratory infection vs mosquito borne infection), it would be important to study the parallels between the immunopathogenesis of both infections for development of more effective vaccines and to develop therapeutic interventions.

Abbreviations

DENV: Dengue virus; DF: Dengue fever; DHF: Dengue haemorrhagic fever; DSS: Dengue shock syndrome; CFRs: Case fatality rates; APPT: Activated partial thromboplastin times; IFN: Interferon; NET: Neutrophil extracellular traps; ADE: Antibody dependent enhancement; sVNT: Surrogate virus neutralizing test; VEGF: Vascular endothelial growth factor; Ang-2: Angiopoietin-2.

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Availability of data and materials

All data is available in the manuscript and figures.

Declarations

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Competing interests

Authors have no competing interests.

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References

- Ackermann M, Verleden SE, Kuehnel M, Haverich A, Welte T, Laenger F, Vanstapel A, Werlein C, Stark H, Tzankov A, Li WW, Li VW, Mentzer SJ, Jonigk D. Pulmonary vascular endothelialitis, thrombosis, and angiogenesis in Covid-19. N Engl J Med. 2020;383(2):120–8.
- Adane T, Getawa S. Coagulation abnormalities in Dengue fever infection: a systematic review and meta-analysis. PLoS Negl Trop Dis. 2021;15(8): e0009666.
- Adikari TN, Gomes L, Wickramasinghe N, Salimi M, Wijesiriwardana N, Kamaladasa A, Shyamali NL, Ogg GS, Malavige GN. Dengue NS1 antigen contributes to disease severity by inducing interleukin (IL)-10 by monocytes. Clin Exp Immunol. 2016;184(1):90–100.
- Altarawneh HN, Chemaitelly H, Ayoub HH, Tang P, Hasan MR, Yassine HM, Al-Khatib HA, Smatti MK, Coyle P, Al-Kanaani Z, Al-Kuwari E, Jeremijenko A, Kaleeckal AH, Latif AN, Shaik RM, Abdul-Rahim HF, Nasrallah GK, Al-Kuwari MG, Butt AA, Al-Romaihi HE, Al-Thani MH, Al-Khal A, Bertollini R, Abu-Raddad LJ. Effects of previous infection and vaccination on

symptomatic Omicron infections. N Engl J Med. 2022. https://doi.org/ 10.1056/NEJMoa2203965.

- Appanna R, Huat TL, See LL, Tan PL, Vadivelu J, Devi S. Cross-reactive T-cell responses to the nonstructural regions of dengue viruses among dengue fever and dengue hemorrhagic fever patients in Malaysia. Clin Vaccine Immunol. 2007;14(8):969–77.
- Avirutnan P, Punyadee N, Noisakran S, Komoltri C, Thiemmeca S, Auethavornanan K, Jairungsri A, Kanlaya R, Tangthawornchaikul N, Puttikhunt C, Pattanakitsakul SN, Yenchitsomanus PT, Mongkolsapaya J, Kasinrerk W, Sittisombut N, Husmann M, Blettner M, Vasanawathana S, Bhakdi S, Malasit P. Vascular leakage in severe dengue virus infections: a potential role for the nonstructural viral protein NS1 and complement. J Infect Dis. 2006;193(8):1078–88.
- Aye KS, Charngkaew K, Win N, Wai KZ, Moe K, Punyadee N, Thiemmeca S, Suttitheptumrong A, Sukpanichnant S, Prida M, Halstead SB. Pathologic highlights of dengue hemorrhagic fever in 13 autopsy cases from Myanmar. Hum Pathol. 2014;45(6):1221–33.
- Aymonnier K, Ng J, Fredenburgh LE, Zambrano-Vera K, Munzer P, Gutch S, Fukui S, Desjardins M, Subramaniam M, Baron RM, Raby BA, Perrella MA, Lederer JA, Wagner DD. Inflammasome activation in neutrophils of patients with severe COVID-19. Blood Adv. 2022;6(7):2001–13.
- Bastard P, Rosen LB, Zhang Q, Michailidis E, Hoffmann HH, Zhang Y, Dorgham K, Philippot Q, Rosain J, Beziat V, Manry J, Shaw E, Haljasmagi L, Peterson P, Lorenzo L, Bizien L, Trouillet-Assant S, Dobbs K, de Jesus AA, Belot A, Kallaste A, Catherinot E, Tandjaoui-Lambiotte Y, Le Pen J, Kerner G, Bigio B, Seeleuthner Y, Yang R, Bolze A, Spaan AN, Delmonte OM, Abers MS, Aiuti A, Casari G, Lampasona V, Piemonti L, Ciceri F, Bilguvar K, Lifton RP, Vasse M, Smadja DM, Migaud M, Hadjadj J, Terrier B, Duffy D, Quintana-Murci L, van de Beek D, Roussel L, Vinh DC, Tangye SG, Haerynck F, Dalmau D, Martinez-Picado J, Brodin P, Nussenzweig MC, Boisson-Dupuis S, Rodriguez-Gallego C, Vogt G, Mogensen TH, Oler AJ, Gu J, Burbelo PD, Cohen JI, Biondi A, Bettini LR, D'Angio M, Bonfanti P, Rossignol P, Mayaux J, Rieux-Laucat F, Husebye ES, Fusco F, Ursini MV, Imberti L, Sottini A, Paghera S, Quiros-Roldan E, Rossi C, Castagnoli R, Montagna D, Licari A, Marseglia GL, Duval X, Ghosn J, Lab H., Group N.-U.I.R.t.C., Clinicians C., Clinicians C.-S., Imagine C.G., French C.C.S.G., Milieu Interieur C., Co V.C.C., Amsterdam U.M.C.C.-B., Effort C.H.G., Tsang JS, Goldbach-Mansky R, Kisand K, Lionakis MS, Puel A, Zhang SY, Holland SM, Gorochov G, Jouanguy E, Rice CM, Cobat A, Notarangelo LD, Abel L, Su HC, Casanova JL. Autoantibodies against type I IFNs in patients with life-threatening COVID-19. Science. 2020;370(6515):eabd4585.
- Beatty PR, Puerta-Guardo H, Killingbeck SS, Glasner DR, Hopkins K, Harris E. Dengue virus NS1 triggers endothelial permeability and vascular leak that is prevented by NS1 vaccination. Sci Transl Med. 2015;7(304):304ra141.
- 11. Begum F, Das S, Mukherjee D, Mal S, Ray U. Insight into the tropism of dengue virus in humans. Viruses. 2019;11(12):1136.
- 12. Bernardes JP, Mishra N, Tran F, Bahmer T, Best L, Blase JI, Bordoni D, Franzenburg J, Geisen U, Josephs-Spaulding J, Kohler P, Kunstner A, Rosati E, Aschenbrenner AC, Bacher P, Baran N, Boysen T, Brandt B, Bruse N, Dorr J, Drager A, Elke G, Ellinghaus D, Fischer J, Forster M, Franke A, Franzenburg S, Frey N, Friedrichs A, Fuss J, Gluck A, Hamm J, Hinrichsen F, Hoeppner MP, Imm S, Junker R, Kaiser S, Kan YH, Knoll R, Lange C, Laue G, Lier C, Lindner M, Marinos G, Markewitz R, Nattermann J, Noth R, Pickkers P, Rabe KF, Renz A, Rocken C, Rupp J, Schaffarzyk A, Scheffold A, Schulte-Schrepping J, Schunk D, Skowasch D, Ulas T, Wandinger KP, Wittig M, Zimmermann J, Busch H, Hoyer BF, Kaleta C, Heyckendorf J, Kox M, Rybniker J, Schreiber S, Schultze JL, Rosenstiel P, Network H.C.A.L.B., Deutsche COI. Longitudinal multi-omics analyses identify responses of megakaryocytes, erythroid cells, and plasmablasts as hallmarks of severe COVID-19. Immunity. 2020;53(6):1296-1314.e1299.
- Blomberg B, Mohn KG, Brokstad KA, Zhou F, Linchausen DW, Hansen BA, Lartey S, Onyango TB, Kuwelker K, Saevik M, Bartsch H, Tondel C, Kittang BR, Bergen C-RG, Cox RJ, Langeland N. Long COVID in a prospective cohort of home-isolated patients. Nat Med. 2021;27(9):1607–13.
- Bonaventura A, Vecchie A, Dagna L, Martinod K, Dixon DL, Van Tassell BW, Dentali F, Montecucco F, Massberg S, Levi M, Abbate A. Endothelial dysfunction and immunothrombosis as key pathogenic mechanisms in COVID-19. Nat Rev Immunol. 2021;21(5):319–29.

- Brar R, Sikka P, Suri V, Singh MP, Suri V, Mohindra R, Biswal M. Maternal and fetal outcomes of dengue fever in pregnancy: a large prospective and descriptive observational study. Arch Gynecol Obstet. 2021;304(1):91–100.
- Cao Y, Yisimayi A, Jian F, Song W, Xiao T, Wang L, Du S, Wang J, Li Q, Chen X, Yu Y, Wang P, Zhang Z, Liu P, An R, Hao X, Wang Y, Wang J, Feng R, Sun H, Zhao L, Zhang W, Zhao D, Zheng J, Yu L, Li C, Zhang N, Wang R, Niu X, Yang S, Song X, Chai Y, Hu Y, Shi Y, Zheng L, Li Z, Gu Q, Shao F, Huang W, Jin R, Shen Z, Wang Y, Wang X, Xiao J, Xie XS. BA.2.12.1, BA.4 and BA.5 escape antibodies elicited by Omicron infection. Nature. 2022. https://doi.org/10.1038/s41586-022-04980-y.
- Castillo Ramirez JA, Urcuqui-Inchima S. Dengue virus control of type I IFN responses: a history of manipulation and control. J Interferon Cytokine Res. 2015;35(6):421–30.
- Chandele A, Sewatanon J, Gunisetty S, Singla M, Onlamoon N, Akondy RS, Kissick HT, Nayak K, Reddy ES, Kalam H, Kumar D, Verma A, Panda H, Wang S, Angkasekwinai N, Pattanapanyasat K, Chokephaibulkit K, Medigeshi GR, Lodha R, Kabra S, Ahmed R, Murali-Krishna K. Characterization of human CD8 T cell responses in dengue virus-infected patients from India, J Virol. 2016;90(24):11259–78.
- Chareonsook O, Foy HM, Teeraratkul A, Silarug N. Changing epidemiology of dengue hemorrhagic fever in Thailand. Epidemiol Infect. 1999;122(1):161–6.
- Chen Z, John Wherry E. T cell responses in patients with COVID-19. Nat Rev Immunol. 2020;20(9):529–36.
- Collaborators C-EM. Estimating excess mortality due to the COVID-19 pandemic: a systematic analysis of COVID-19-related mortality, 2020–21. Lancet. 2022;399(10334):1513–36.
- Damoiseaux J, Dotan A, Fritzler MJ, Bogdanos DP, Meroni PL, Roggenbuck D, Goldman M, Landegren N, Bastard P, Shoenfeld Y, Conrad K. Autoantibodies and SARS-CoV2 infection: the spectrum from association to clinical implication: report of the 15th Dresden Symposium on autoantibodies. Autoimmun Rev. 2022;21(3):103012.
- Dayarathna S, Jeewandara C, Gomes L, Somathilaka G, Jayathilaka D, Vimalachandran V, Wijewickrama A, Narangoda E, Idampitiya D, Ogg GS, Malavige GN. Similarities and differences between the "cytokine storms" in acute dengue and COVID-19. Sci Rep. 2020;10(1):19839.
- Dejnirattisai W, Jumnainsong A, Onsirisakul N, Fitton P, Vasanawathana S, Limpitikul W, Puttikhunt C, Edwards C, Duangchinda T, Supasa S, Chawansuntati K, Malasit P, Mongkolsapaya J, Screaton G. Cross-reacting antibodies enhance dengue virus infection in humans. Science. 2010;328(5979):745–8.
- Diamond MS, Roberts TG, Edgil D, Lu B, Ernst J, Harris E. Modulation of Dengue virus infection in human cells by alpha, beta, and gamma interferons. J Virol. 2000;74(11):4957–66.
- Diao B, Wang C, Tan Y, Chen X, Liu Y, Ning L, Chen L, Li M, Liu Y, Wang G, Yuan Z, Feng Z, Zhang Y, Wu Y, Chen Y. Reduction and functional exhaustion of T cells in patients with coronavirus disease 2019 (COVID-19). Front Immunol. 2020;11:827.
- Dispinseri S, Secchi M, Pirillo MF, Tolazzi M, Borghi M, Brigatti C, De Angelis ML, Baratella M, Bazzigaluppi E, Venturi G, Sironi F, Canitano A, Marzinotto I, Tresoldi C, Ciceri F, Piemonti L, Negri D, Cara A, Lampasona V, Scarlatti G. Neutralizing antibody responses to SARS-CoV-2 in symptomatic COVID-19 is persistent and critical for survival. Nat Commun. 2021;12(1):2670.
- Djamiatun K, van der Ven AJ, de Groot PG, Faradz SM, Hapsari D, Dolmans WM, Sebastian S, Fijnheer R, de Mast Q. Severe dengue is associated with consumption of von Willebrand factor and its cleaving enzyme ADAMTS-13. PLoS Negl Trop Dis. 2012;6(5):e1628.
- Donders GGG, Grinceviciene S, Haldre K, Lonnee-Hoffmann R, Donders F, Tsiakalos A, Adriaanse A, Martinez de Oliveira J, Ault K, Mendling W, On the behalf of The Covid-Isidog Guideline G. ISIDOG consensus guidelines on COVID-19 vaccination for women before, during and after pregnancy. J Clin Med. 2021;10(13):2902.
- Dong T, Moran E, Vinh Chau N, Simmons C, Luhn K, Peng Y, Wills B, Phuong Dung N, Thi Thu Thao L, Hien TT, McMichael A, Farrar J, Rowland-Jones S. High pro-inflammatory cytokine secretion and loss of high avidity cross-reactive cytotoxic T-cells during the course of secondary dengue virus infection. PLoS ONE. 2007;2(12):e1192.

- Douaud G, Lee S, Alfaro-Almagro F, Arthofer C, Wang C, McCarthy P, Lange F, Andersson JLR, Griffanti L, Duff E, Jbabdi S, Taschler B, Keating P, Winkler AM, Collins R, Matthews PM, Allen N, Miller KL, Nichols TE, Smith SM. SARS-CoV-2 is associated with changes in brain structure in UK Biobank. Nature. 2022;604(7907):697–707.
- Dung NT, Duyen HT, Thuy NT, Ngoc TV, Chau NV, Hien TT, Rowland-Jones SL, Dong T, Farrar J, Wills B, Simmons CP. Timing of CD8+ T cell responses in relation to commencement of capillary leakage in children with dengue. J Immunol. 2010;184(12):7281–7.
- Dyson L, Hill EM, Moore S, Curran-Sebastian J, Tildesley MJ, Lythgoe KA, House T, Pellis L, Keeling MJ. Possible future waves of SARS-CoV-2 infection generated by variants of concern with a range of characteristics. Nat Commun. 2021;12(1):5730.
- Elias C, Sekri A, Leblanc P, Cucherat M, Vanhems P. The incubation period of COVID-19: a meta-analysis. Int J Infect Dis. 2021;104:708–10.
- 35. Endy TP, Wang D, Polhemus ME, Jarman RG, Jasper LE, Gromowski G, Lin L, De La Barra RA, Friberg H, Currier JR, Abbott M, Ware L, Klick M, Paolino KM, Blair DC, Eckels K, Rutvisuttinunt W, Thomas SJ. A phase 1, open-label assessment of a dengue virus-1 live virus human challenge strain. J Infect Dis. 2021;223(2):258–67.
- Freitas RS, Crum TF, Parvatiyar K. SARS-CoV-2 spike antagonizes innate antiviral immunity by targeting interferon regulatory factor 3. Front Cell Infect Microbiol. 2021;11:789462.
- Fried JR, Gibbons RV, Kalayanarooj S, Thomas SJ, Srikiatkhachorn A, Yoon IK, Jarman RG, Green S, Rothman AL, Cummings DA. Serotypespecific differences in the risk of dengue hemorrhagic fever: an analysis of data collected in Bangkok, Thailand from 1994 to 2006. PLoS Negl Trop Dis. 2010;4(3):e617.
- Fung SY, Siu KL, Lin H, Chan CP, Yeung ML, Jin DY. SARS-CoV-2 NSP13 helicase suppresses interferon signaling by perturbing JAK1 phosphorylation of STAT1. Cell Biosci. 2022;12(1):36.
- Gao YD, Ding M, Dong X, Zhang JJ, Kursat Azkur A, Azkur D, Gan H, Sun YL, Fu W, Li W, Liang HL, Cao YY, Yan Q, Cao C, Gao HY, Bruggen MC, van de Veen W, Sokolowska M, Akdis M, Akdis CA. Risk factors for severe and critically ill COVID-19 patients: a review. Allergy. 2021;76(2):428–55.
- Garcia-Bates TM, Cordeiro MT, Nascimento EJ, Smith AP, Soares de Melo KM, McBurney SP, Evans JD, Marques ET Jr, Barratt-Boyes SM. Association between magnitude of the virus-specific plasmablast response and disease severity in dengue patients. J Immunol. 2013;190(1):80–7.
- Garcia-Beltran WF, Lam EC, Astudillo MG, Yang D, Miller TE, Feldman J, Hauser BM, Caradonna TM, Clayton KL, Nitido AD, Murali MR, Alter G, Charles RC, Dighe A, Branda JA, Lennerz JK, Lingwood D, Schmidt AG, lafrate AJ, Balazs AB. COVID-19-neutralizing antibodies predict disease severity and survival. Cell. 2021;184(2):476-488.e411.
- Garcia G, Gonzalez N, Perez AB, Sierra B, Aguirre E, Rizo D, Izquierdo A, Sanchez L, Diaz D, Lezcay M, Pacheco B, Hirayama K, Guzman MG. Long-term persistence of clinical symptoms in dengue-infected persons and its association with immunological disorders. Int J Infect Dis. 2011;15(1):e38-43.
- Gomes L, Gamage J, Wijesinghe A, Jeewandara C, Wijewickrama A, Ogg G, Malavige GN. High prevalence of autoantibodies in patients following acute dengue infection. 19th International Congress on Infectious Diseases. In: 19th international congress on infectious diseases, Virtual, 2020.
- 44. Goshua G, Pine AB, Meizlish ML, Chang CH, Zhang H, Bahel P, Baluha A, Bar N, Bona RD, Burns AJ, Dela Cruz CS, Dumont A, Halene S, Hwa J, Koff J, Menninger H, Neparidze N, Price C, Siner JM, Tormey C, Rinder HM, Chun HJ, Lee AI. Endotheliopathy in COVID-19-associated coagulopathy: evidence from a single-centre, cross-sectional study. Lancet Haematol. 2020;7(8):e575–82.
- 45. Han H, Ma Q, Li C, Liu R, Zhao L, Wang W, Zhang P, Liu X, Gao G, Liu F, Jiang Y, Cheng X, Zhu C, Xia Y. Profiling serum cytokines in COVID-19 patients reveals IL-6 and IL-10 are disease severity predictors. Emerg Microbes Infect. 2020;9(1):1123–30.
- Hannah Ritchie EM, Rodés-Guirao L, Appel C, Giattino C, Ortiz-Ospina E, Hasell J, Macdonald B, Beltekian D, Roser M. Coronavirus pandemic (COVID-19). 02/24/2022 ed. 2022.
- 47. Hasan MM, Sahito AM, Muzzamil M, Mohanan P, Islam Z, Billah MM, Islam MJ, Essar MY. Devastating dengue outbreak amidst COVID-19

pandemic in Bangladesh: an alarming situation. Trop Med Health. 2022;50(1):11.

- Hottz ED, Lopes JF, Freitas C, Valls-de-Souza R, Oliveira MF, Bozza MT, Da Poian AT, Weyrich AS, Zimmerman GA, Bozza FA, Bozza PT. Platelets mediate increased endothelium permeability in dengue through NLRP3-inflammasome activation. Blood. 2013;122(20):3405–14.
- Huertas A, Montani D, Savale L, Pichon J, Tu L, Parent F, Guignabert C, Humbert M. Endothelial cell dysfunction: a major player in SARS-CoV-2 infection (COVID-19)? Eur Respir J. 2020;56(1):2001634.
- 50. Investigators R-C, Gordon AC, Mouncey PR, Al-Beidh F, Rowan KM, Nichol AD, Arabi YM, Annane D, Beane A, van Bentum-Puijk W, Berry LR, Bhimani Z, Bonten MJM, Bradbury CA, Brunkhorst FM, Buzgau A, Cheng AC, Detry MA, Duffy EJ, Estcourt LJ, Fitzgerald M, Goossens H, Haniffa R, Higgins AM, Hills TE, Horvat CM, Lamontagne F, Lawler PR, Leavis HL, Linstrum KM, Litton E, Lorenzi E, Marshall JC, Mayr FB, McAuley DF, McGlothlin A, McGuinness SP, McVerry BJ, Montgomery SK, Morpeth SC, Murthy S, Orr K, Parke RL, Parker JC, Patanwala AE, Pettila V, Rademaker E, Santos MS, Saunders CT, Seymour CW, Shankar-Hari M, Sligl WI, Turgeon AF, Turner AM, van de Veerdonk FL, Zarychanski R, Green C, Lewis RJ, Angus DC, McArthur CJ, Berry S, Webb SA, Derde LPG. Interleukin-6 receptor antagonists in critically ill patients with covid-19. N Engl J Med. 2021;384(16):1491–502.
- Islam H, Chamberlain TC, Mui AL, Little JP. Elevated Interleukin-10 levels in COVID-19: potentiation of pro-inflammatory responses or impaired anti-inflammatory action? Front Immunol. 2021;12:677008.
- Islam Z, Mohanan P, Bilal W, Hashmi T, Rahmat Z, Abdi I, Riaz MMA, Essar MY. Dengue Virus cases surge amidst COVID-19 in Pakistan: challenges efforts and recommendations. Infect Drug Resist. 2022;15:367–71.
- 53. Jackson CB, Farzan M, Chen B, Choe H. Mechanisms of SARS-CoV-2 entry into cells. Nat Rev Mol Cell Biol. 2022;23(1):3–20.
- Jayaratne SD, Atukorale V, Gomes L, Chang T, Wijesinghe T, Fernando S, Ogg GS, Malavige GN. Evaluation of the WHO revised criteria for classification of clinical disease severity in acute adult dengue infection. BMC Res Notes. 2012;5:645.
- 55. Jayathilaka D, Gomes L, Jeewandara C, Jayarathna GSB, Herath D, Perera PA, Fernando S, Wijewickrama A, Hardman CS, Ogg GS, Malavige GN. Role of NS1 antibodies in the pathogenesis of acute secondary dengue infection. Nat Commun. 2018;9(1):5242.
- Jeewandara C, Adikari TN, Gomes L, Fernando S, Fernando RH, Perera MK, Ariyaratne D, Kamaladasa A, Salimi M, Prathapan S, Ogg GS, Malavige GN. Functionality of dengue virus specific memory T cell responses in individuals who were hospitalized or who had mild or subclinical dengue infection. PLoS Negl Trop Dis. 2015;9(4):e0003673.
- Jeewandara C, Gomes L, Udari S, Paranavitane SA, Shyamali NL, Ogg GS, Malavige GN. Secretory phospholipase A2 in the pathogenesis of acute dengue infection. Immun Inflamm Dis. 2017;5(1):7–15.
- Jeewandara C, Gomes L, Wickramasinghe N, Gutowska-Owsiak D, Waithe D, Paranavitane SA, Shyamali NL, Ogg GS, Malavige GN. Platelet activating factor contributes to vascular leak in acute dengue infection. PLoS Negl Trop Dis. 2015;9(2):e0003459.
- Jeewandara C, Jayathilaka D, Gomes L, Wijewickrama A, Narangoda E, Idampitiya D, Guruge D, Wijayamuni R, Manilgama S, Ogg GS, Tan CW, Wang LF, Malavige GN. SARS-CoV-2 neutralizing antibodies in patients with varying severity of acute COVID-19 illness. Sci Rep. 2021;11(1):2062.
- Kaeuffer C, Le Hyaric C, Fabacher T, Mootien J, Dervieux B, Ruch Y, Hugerot A, Zhu YJ, Pointurier V, Clere-Jehl R, Greigert V, Kassegne L, Lefebvre N, Gallais F, Covid Alsace Study G, Meyer N, Hansmann Y, Hinschberger O, Danion F, Group C.A.S. Clinical characteristics and risk factors associated with severe COVID-19: prospective analysis of 1,045 hospitalised cases in North-Eastern France, March 2020. Euro Surveill. 2020;25(48):2000895.
- Kamaladasa A, Gomes L, Jeewandara C, Shyamali NL, Ogg GS, Malavige GN. Lipopolysaccharide acts synergistically with the dengue virus to induce monocyte production of platelet activating factor and other inflammatory mediators. Antivir Res. 2016;133:183–90.
- 62. Kamaladasa A, Gomes L, Wijesinghe A, Jeewandara C, Toh YX, Jayathilaka D, Ogg GS, Fink K, Malavige GN. Altered monocyte response to the dengue virus in those with varying severity of past dengue infection. Antivir Res. 2019;169:104554.
- 63. Katzelnick LC, Ben-Shachar R, Mercado JC, Rodriguez-Barraquer I, Elizondo D, Arguello S, Nunez A, Ojeda S, Sanchez N, Lopez Mercado B,

Gresh L, Burger-Calderon R, Kuan G, Gordon A, Balmaseda A, Harris E. Dynamics and determinants of the force of infection of dengue virus from 1994 to 2015 in Managua, Nicaragua. Proc Natl Acad Sci USA. 2018;115(42):10762–7.

- Katzelnick LC, Gresh L, Halloran ME, Mercado JC, Kuan G, Gordon A, Balmaseda A, Harris E. Antibody-dependent enhancement of severe dengue disease in humans. Science. 2017;358(6365):929–32.
- Katzelnick LC, Montoya M, Gresh L, Balmaseda A, Harris E. Neutralizing antibody titers against dengue virus correlate with protection from symptomatic infection in a longitudinal cohort. Proc Natl Acad Sci USA. 2016;113(3):728–33.
- Khoury DS, Cromer D, Reynaldi A, Schlub TE, Wheatley AK, Juno JA, Subbarao K, Kent SJ, Triccas JA, Davenport MP. Neutralizing antibody levels are highly predictive of immune protection from symptomatic SARS-CoV-2 infection. Nat Med. 2021;27(7):1205–11.
- Kumar A, Ishida R, Strilets T, Cole J, Lopez-Orozco J, Fayad N, Felix-Lopez A, Elaish M, Evseev D, Magor KE, Mahal LK, Nagata LP, Evans DH, Hobman TC. SARS-CoV-2 nonstructural protein 1 inhibits the interferon response by causing depletion of key host signaling factors. J Virol. 2021;95(13):e0026621.
- Kundu R, Narean JS, Wang L, Fenn J, Pillay T, Fernandez ND, Conibear E, Koycheva A, Davies M, Tolosa-Wright M, Hakki S, Varro R, McDermott E, Hammett S, Cutajar J, Thwaites RS, Parker E, Rosadas C, McClure M, Tedder R, Taylor GP, Dunning J, Lalvani A. Cross-reactive memory T cells associate with protection against SARS-CoV-2 infection in COVID-19 contacts. Nat Commun. 2022;13(1):80.
- Kuno G. Emergence of the severe syndrome and mortality associated with dengue and dengue-like illness: historical records (1890 to 1950) and their compatibility with current hypotheses on the shift of disease manifestation. Clin Microbiol Rev. 2009;22(2):186–201 (Table of Contents).
- Kuri-Cervantes L, Pampena MB, Meng W, Rosenfeld AM, Ittner CAG, Weisman AR, Agyekum RS, Mathew D, Baxter AE, Vella LA, Kuthuru O, Apostolidis SA, Bershaw L, Dougherty J, Greenplate AR, Pattekar A, Kim J, Han N, Gouma S, Weirick ME, Arevalo CP, Bolton MJ, Goodwin EC, Anderson EM, Hensley SE, Jones TK, Mangalmurti NS, Luning Prak ET, Wherry EJ, Meyer NJ, Betts MR. Comprehensive mapping of immune perturbations associated with severe COVID-19. Sci Immunol. 2020;5(49):e7114.
- KyeMon K, Nontprasert A, Kittitrakul C, Tangkijvanich P, Leowattana W, Poovorawan K. Incidence and clinical outcome of acute liver failure caused by dengue in a hospital for tropical diseases, Thailand. Am J Trop Med Hyg. 2016;95(6):1338–44.
- 72. Lee JS, Shin EC. The type I interferon response in COVID-19: implications for treatment. Nat Rev Immunol. 2020;20(10):585–6.
- Li HM, Huang YK, Su YC, Kao CH. Increased risk of autoimmune diseases in dengue patients: A population-based cohort study. J Infect. 2018;77(3):212–9.
- Li Y, Hu Z, Huang Y, Li J, Hong W, Qin Z, Tong Y, Li J, Lv M, Li M, Zheng X, Hu J, Hua J, Zhang F, Xu DL. Characterization of the Myocarditis during the worst outbreak of dengue infection in China. Medicine (Baltimore). 2016;95(27):e4051.
- Lin CF, Lei HY, Shiau AL, Liu CC, Liu HS, Yeh TM, Chen SH, Lin YS. Antibodies from dengue patient sera cross-react with endothelial cells and induce damage. J Med Virol. 2003;69(1):82–90.
- Lizarraga KJ, Nayer A. Dengue-associated kidney disease. J Nephropathol. 2014;3(2):57–62.
- 77. Loyal L, Braun J, Henze L, Kruse B, Dingeldey M, Reimer U, Kern F, Schwarz T, Mangold M, Unger C, Dorfler F, Kadler S, Rosowski J, Gurcan K, Uyar-Aydin Z, Frentsch M, Kurth F, Schnatbaum K, Eckey M, Hippenstiel S, Hocke A, Muller MA, Sawitzki B, Miltenyi S, Paul F, Mall MA, Wenschuh H, Voigt S, Drosten C, Lauster R, Lachman N, Sander LE, Corman VM, Rohmel J, Meyer-Arndt L, Thiel A, Giesecke-Thiel C. Cross-reactive CD4(+) T cells enhance SARS-CoV-2 immune responses upon infection and vaccination. Science. 2021;374(6564):eabh1823.
- Luo YR, Chakraborty I, Yun C, Wu AHB, Lynch KL. Kinetics of severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) antibody avidity maturation and association with disease severity. Clin Infect Dis. 2021;73(9):e3095–7.
- Macias AE, Werneck GL, Castro R, Mascarenas C, Coudeville L, Morley D, Recamier V, Guergova-Kuras M, Etcheto A, Puentes-Rosas E, Baurin

N, Toh ML. Mortality among hospitalized dengue patients with comorbidities in Mexico, Brazil, and Colombia. Am J Trop Med Hyg. 2021;105(1):102–9.

- Maemura T, Kuroda M, Armbrust T, Yamayoshi S, Halfmann PJ, Kawaoka Y. Antibody-Dependent enhancement of SARS-CoV-2 infection is mediated by the IgG Receptors FcgammaRIIA and FcgammaRIIIA but does not contribute to aberrant cytokine production by macrophages. MBio. 2021;12(5):e0198721.
- Malavige GN, Huang LC, Salimi M, Gomes L, Jayaratne SD, Ogg GS. Cellular and cytokine correlates of severe dengue infection. PLoS ONE. 2012;7(11):e50387.
- Malavige GN, Jeewandara C, Alles KM, Salimi M, Gomes L, Kamaladasa A, Jayaratne SD, Ogg GS. Suppression of virus specific immune responses by IL-10 in acute dengue infection. PLoS Negl Trop Dis. 2013;7(9):e2409.
- Malavige GN, Jeewandara C, Ghouse A, Somathilake G, Tissera H. Changing epidemiology of dengue in Sri Lanka-challenges for the future. PLoS Negl Trop Dis. 2021;15(8):e0009624.
- Malavige GN, Jeewandara C, Ogg GS. Dysfunctional innate immune responses and severe dengue. Front Cell Infect Microbiol. 2020;10:590004.
- Malavige GN, Ogg GS. Pathogenesis of vascular leak in dengue virus infection. Immunology. 2017;151(3):261–9.
- Malavige GN, Ranatunga PK, Velathanthiri VG, Fernando S, Karunatilaka DH, Aaskov J, Seneviratne SL. Patterns of disease in Sri Lankan dengue patients. Arch Dis Child. 2006;91(5):396–400.
- Mateus J, Grifoni A, Tarke A, Sidney J, Ramirez SI, Dan JM, Burger ZC, Rawlings SA, Smith DM, Phillips E, Mallal S, Lammers M, Rubiro P, Quiambao L, Sutherland A, Yu ED, da Silva Antunes R, Greenbaum J, Frazier A, Markmann AJ, Premkumar L, de Silva A, Peters B, Crotty S, Sette A, Weiskopf D. Selective and cross-reactive SARS-CoV-2 T cell epitopes in unexposed humans. Science. 2020;370(6512):89–94.
- Mathew D, Giles JR, Baxter AE, Oldridge DA, Greenplate AR, Wu JE, Alanio C, Kuri-Cervantes L, Pampena MB, D'Andrea K, Manne S, Chen Z, Huang YJ, Reilly JP, Weisman AR, Ittner CAG, Kuthuru O, Dougherty J, Nzingha K, Han N, Kim J, Pattekar A, Goodwin EC, Anderson EM, Weirick ME, Gouma S, Arevalo CP, Bolton MJ, Chen F, Lacey SF, Ramage H, Cherry S, Hensley SE, Apostolidis SA, Huang AC, Vella LA, Unit U.P.C.P., Betts MR, Meyer NJ, Wherry EJ. Deep immune profiling of COVID-19 patients reveals distinct immunotypes with therapeutic implications. Science. 2020;369(6508):eabc8511.
- McMahan K, Yu J, Mercado NB, Loos C, Tostanoski LH, Chandrashekar A, Liu J, Peter L, Atyeo C, Zhu A, Bondzie EA, Dagotto G, Gebre MS, Jacob-Dolan C, Li Z, Nampanya F, Patel S, Pessaint L, Van Ry A, Blade K, Yalley-Ogunro J, Cabus M, Brown R, Cook A, Teow E, Andersen H, Lewis MG, Lauffenburger DA, Alter G, Barouch DH. Correlates of protection against SARS-CoV-2 in rhesus macaques. Nature. 2021;590(7847):630–4.
- Medicine J.H.U.a. Coronavirus Resource Centre. In: CRITICAL TRENDS: TRACKING CRITICAL DATA John Hopkins University. 2021. https://coron avirus.jhu.edu/. Accessed 28th June 2021.
- Mefsin Y, Chen D, Bond HS, Lin Y, Cheung JK, Wong JY, Ali ST, Lau EHY, Wu P, Leung GM, Cowling BJ. Epidemiology of infections with SARS-CoV-2 Omicron BA.2 variant in Hong Kong, January–March 2022. medR xiv:2022.2004.2007.22273595. 2022.
- 92. Mehandru S, Merad M. Pathological sequelae of long-haul COVID. Nat Immunol. 2022;23(2):194–202.
- Mongkolsapaya J, Duangchinda T, Dejnirattisai W, Vasanawathana S, Avirutnan P, Jairungsri A, Khemnu N, Tangthawornchaikul N, Chotiyarnwong P, Sae-Jang K, Koch M, Jones Y, McMichael A, Xu X, Malasit P, Screaton G. T cell responses in dengue hemorrhagic fever: are crossreactive T cells suboptimal? J Immunol. 2006;176(6):3821–9.
- 94. Moss P. The T cell immune response against SARS-CoV-2. Nat Immunol. 2022;23(2):186–93.
- 95. Murhekar MV, Kamaraj P, Kumar MS, Khan SA, Allam RR, Barde P, Dwibedi B, Kanungo S, Mohan U, Mohanty SS, Roy S, Sagar V, Savargaonkar D, Tandale BV, Topno RK, Sapkal G, Kumar CPG, Sabarinathan R, Kumar VS, Bitragunta S, Grover GS, Lakshmi PVM, Mishra CM, Sadhukhan P, Sahoo PK, Singh SK, Yadav CP, Bhagat A, Srivastava R, Dinesh ER, Karunakaran T, Govindhasamy C, Rajasekar TD, Jeyakumar A, Suresh A, Augustine D, Kumar PA, Kumar R, Dutta S, Toteja GS,

Gupta N, Mehendale SM. Burden of dengue infection in India, 2017: a cross-sectional population based serosurvey. Lancet Glob Health. 2019;7(8):e1065–73.

- 96. Nishiura H, Halstead SB. Natural history of dengue virus (DENV)-1 and DENV-4 infections: reanalysis of classic studies. J Infect Dis. 2007;195(7):1007–13.
- Noisakran S, Onlamoon N, Songprakhon P, Hsiao HM, Chokephaibulkit K, Perng GC. Cells in dengue virus infection in vivo. Adv Virol. 2010;2010:164878.
- Okuya K, Hattori T, Saito T, Takadate Y, Sasaki M, Furuyama W, Marzi A, Ohiro Y, Konno S, Hattori T, Takada A. Multiple routes of antibodydependent enhancement of SARS-CoV-2 infection. Microbiol Spectr. 2022;10(2):e0155321.
- 99. Opasawatchai A, Amornsupawat P, Jiravejchakul N, Chan-In W, Spoerk NJ, Manopwisedjaroen K, Singhasivanon P, Yingtaweesak T, Suraamornkul S, Mongkolsapaya J, Sakuntabhai A, Matangkasombut P, Loison F. Neutrophil activation and early features of NET formation are associated with dengue virus infection in human. Front Immunol. 2018;9:3007.
- Paixao ES, Campbell OM, Teixeira MG, Costa MC, Harron K, Barreto ML, Leal MB, Almeida MF, Rodrigues LC. Dengue during pregnancy and live birth outcomes: a cohort of linked data from Brazil. BMJ Open. 2019;9(7):e023529.
- Paixao ES, Harron K, Campbell O, Teixeira MG, Costa M, Barreto ML, Rodrigues LC. Dengue in pregnancy and maternal mortality: a cohort analysis using routine data. Sci Rep. 2018;8(1):9938.
- 102. Park A, Iwasaki A. Type I and Type III interferons—induction, signaling, evasion, and application to combat COVID-19. Cell Host Microbe. 2020;27(6):870–8.
- Proal AD, VanElzakker MB. Long COVID or post-acute sequelae of COVID-19 (PASC): an overview of biological factors that may contribute to persistent symptoms. Front Microbiol. 2021;12:698169.
- Puerta-Guardo H, Glasner DR, Harris E. Dengue virus NS1 disrupts the endothelial glycocalyx leading to hyperpermeability. PLoS Pathog. 2016;12(7):e1005738.
- Reusch N, De Domenico E, Bonaguro L, Schulte-Schrepping J, Bassler K, Schultze JL, Aschenbrenner AC. Neutrophils in COVID-19. Front Immunol. 2021;12: 652470.
- 106. Ruhl L, Pink I, Kuhne JF, Beushausen K, Keil J, Christoph S, Sauer A, Boblitz L, Schmidt J, David S, Jack HM, Roth E, Cornberg M, Schulz TF, Welte T, Hoper MM, Falk CS. Endothelial dysfunction contributes to severe COVID-19 in combination with dysregulated lymphocyte responses and cytokine networks. Signal Transduct Target Ther. 2021;6(1):418.
- 107. Rutkai I, Mayer MG, Hellmers LM, Ning B, Huang Z, Monjure CJ, Coyne C, Silvestri R, Golden N, Hensley K, Chandler K, Lehmicke G, Bix GJ, Maness NJ, Russell-Lodrigue K, Hu TY, Roy CJ, Blair RV, Bohm R, Doyle-Meyers LA, Rappaport J, Fischer T. Neuropathology and virus in brain of SARS-CoV-2 infected non-human primates. Nat Commun. 2022;13(1):1745.
- Ryan SJ, Carlson CJ, Mordecai EA, Johnson LR. Global expansion and redistribution of Aedes-borne virus transmission risk with climate change. PLoS Negl Trop Dis. 2019;13(3):e0007213.
- Salamanna F, Veronesi F, Martini L, Landini MP, Fini M. Post-COVID-19 syndrome: The persistent symptoms at the post-viral stage of the disease a systematic review of the current data. Front Med (Lausanne). 2021;8:653516.
- 110. Schimmel L, Chew KY, Stocks CJ, Yordanov TE, Essebier P, Kulasinghe A, Monkman J, Dos Santos Miggiolaro AFR, Cooper C, de Noronha L, Schroder K, Lagendijk AK, Labzin LI, Short KR, Gordon EJ. Endothelial cells are not productively infected by SARS-CoV-2. Clin Transl Immunol. 2021;10(10):e1350.
- 111. Sigera PC, Rajapakse S, Weeratunga P, De Silva NL, Gomes L, Malavige GN, Rodrigo C, Fernando SD. Dengue and post-infection fatigue: findings from a prospective cohort-the Colombo Dengue Study. Trans R Soc Trop Med Hyg. 2020;115:669–76.
- 112. Silva T, Gomes L, Jeewandara C, Ogg GS, Malavige GN. Dengue NS1 induces phospholipase A2 enzyme activity, prostaglandins, and inflammatory cytokines in monocytes. Antivir Res. 2022;202:105312.
- Syenina A, Jagaraj CJ, Aman SA, Sridharan A, St John AL. Dengue vascular leakage is augmented by mast cell degranulation mediated by immunoglobulin Fcgamma receptors. Elife. 2015;4:e05291.

- 114. Synowiec A, Szczepanski A, Barreto-Duran E, Lie LK, Pyrc K. Severe Acute respiratory syndrome coronavirus 2 (SARS-CoV-2): a systemic infection. Clin Microbiol Rev. 2021;34(2):e00133-20.
- 115. Tan LK, Low SL, Sun H, Shi Y, Liu L, Lam S, Tan HH, Ang LW, Wong WY, Chua R, Teo D, Ng LC, Cook AR. Force of infection and true infection rate of dengue in Singapore: implications for dengue control and management. Am J Epidemiol. 2019;188(8):1529–38.
- Tang N, Li D, Wang X, Sun Z. Abnormal coagulation parameters are associated with poor prognosis in patients with novel coronavirus pneumonia. J Thromb Haemost. 2020;18(4):844–7.
- 117. Tiga DC, Undurraga EA, Ramos-Castaneda J, Martinez-Vega RA, Tschampl CA, Shepard DS. Persistent symptoms of dengue: estimates of the incremental disease and economic burden in Mexico. Am J Trop Med Hyg. 2016;94(5):1085–9.
- Tsai TT, Chuang YJ, Lin YS, Chang CP, Wan SW, Lin SH, Chen CL, Lin CF. Antibody-dependent enhancement infection facilitates dengue virusregulated signaling of IL-10 production in monocytes. PLoS Negl Trop Dis. 2014;8(11):e3320.
- Tsheten T, Clements ACA, Gray DJ, Adhikary RK, Furuya-Kanamori L, Wangdi K. Clinical predictors of severe dengue: a systematic review and meta-analysis. Infect Dis Poverty. 2021;10(1):123.
- 120. Upasani V, Scagnolari C, Frasca F, Smith N, Bondet V, Vanderlinden A, Lay S, Auerswald H, Heng S, Laurent D, Ly S, Duong V, Antonelli G, Dussart P, Duffy D, Cantaert T. Decreased type I interferon production by plasmacytoid dendritic cells contributes to severe dengue. Front Immunol. 2020;11:605087.
- 121. Wang S, Wang J, Yu X, Jiang W, Chen S, Wang R, Wang M, Jiao S, Yang Y, Wang W, Chen H, Chen B, Gu C, Liu C, Wang A, Wang M, Li G, Guo C, Liu D, Zhang J, Zhang M, Wang L, Gui X. Antibody-dependent enhancement (ADE) of SARS-CoV-2 pseudoviral infection requires FcgammaRIIB and virus-antibody complex with bivalent interaction. Commun Biol. 2022;5(1):262.
- 122. Wang TT, Sewatanon J, Memoli MJ, Wrammert J, Bournazos S, Bhaumik SK, Pinsky BA, Chokephaibulkit K, Onlamoon N, Pattanapanyasat K, Taubenberger JK, Ahmed R, Ravetch JV. IgG antibodies to dengue enhanced for FcgammaRIIIA binding determine disease severity. Science. 2017;355(6323):395–8.
- 123. Wassie GT, Azene AG, Bantie GM, Dessie G, Aragaw AM. Incubation period of severe acute respiratory syndrome novel coronavirus 2 that causes coronavirus disease 2019: a systematic review and meta-analysis. Curr Ther Res Clin Exp. 2020;93:100607.
- Wei SQ, Bilodeau-Bertrand M, Liu S, Auger N. The impact of COVID-19 on pregnancy outcomes: a systematic review and meta-analysis. CMAJ. 2021;193(16):E540–8.
- 125. Weiskopf D, Angelo MA, de Azeredo EL, Sidney J, Greenbaum JA, Fernando AN, Broadwater A, Kolla RV, De Silva AD, de Silva AM, Mattia KA, Doranz BJ, Grey HM, Shresta S, Peters B, Sette A. Comprehensive analysis of dengue virus-specific responses supports an HLA-linked protective role for CD8+ T cells. Proc Natl Acad Sci USA. 2013;110(22):E2046-2053.
- WHO. 14.9 million excess deaths associated with the COVID-19 pandemic in 2020 and 2021. In: WHO, editor. World Health Organization;2022.
- 127. World Health Organization. Comprehensive guidelines for prevention and control of dengue fever and dengue haemorrhagic fever. New Delhi: World Health Organization, SEARO; 2011.
- 128. WHO. 2022 29th of April. World Health Organization, COVID-19 vaccine tracker. VIPER Group COVID19 Vaccine Tracker Team https://covid19. trackvaccines.org/agency/who/. Accessed 29th Apr 2022.
- World Health Organization. Coronavirus disease (COVID-19) technical guidance: early investigations. Geneva: World Health Organization; 2020.
- 130. Wijeratne DT, Fernando S, Gomes L, Jeewandara C, Ginneliya A, Samarasekara S, Wijewickrama A, Hardman CS, Ogg GS, Malavige GN. Quantification of dengue virus specific T cell responses and correlation with viral load and clinical disease severity in acute dengue infection. PLoS Negl Trop Dis. 2018;12(10):e0006540.
- 131. Wijeratne DT, Fernando S, Gomes L, Jeewandara C, Jayarathna G, Perera Y, Wickramanayake S, Wijewickrama A, Ogg GS, Malavige GN. Association of dengue virus-specific polyfunctional T-cell responses with clinical disease severity in acute dengue infection. Immun Inflamm Dis. 2019;7:276–85.

- 132. Wijesinghe A, Gamage J, Goonewardena H, Gomes L, Jayathilaka D, Wijeratne DT, de Alwis R, Jeewandara C, Wijewickrama A, Ogg GS, Malavige GN. Phenotype and functionality of follicular helper T cells in patients with acute dengue infection. J Biomed Sci. 2020;27(1):50.
- 133. Wills BA, Oragui EE, Stephens AC, Daramola OA, Dung NM, Loan HT, Chau NV, Chambers M, Stepniewska K, Farrar JJ, Levin M. Coagulation abnormalities in dengue hemorrhagic fever: serial investigations in 167 Vietnamese children with Dengue shock syndrome. Clin Infect Dis. 2002;35(3):277–85.
- 134. Woodruff M, Ramonell R, Cashman K, Nguyen D, Ley A, Kyu S, Saini A, Haddad N, Chen W, Howell JC, Ozturk T, Lee S, Estrada J, Morrison-Porter A, Derrico A, Anam F, Wu H, Le S, Jenks S, Hu W, Lee FE, Sanz I. Critically ill SARS-CoV-2 patients display lupus-like hallmarks of extrafollicular B cell activation. medRxiv. 2020.
- 135. Woodruff MC, Ramonell RP, Nguyen DC, Cashman KS, Saini AS, Haddad NS, Ley AM, Kyu S, Howell JC, Ozturk T, Lee S, Suryadevara N, Case JB, Bugrovsky R, Chen W, Estrada J, Morrison-Porter A, Derrico A, Anam FA, Sharma M, Wu HM, Le SN, Jenks SA, Tipton CM, Staitieh B, Daiss JL, Ghosn E, Diamond MS, Carnahan RH, Crowe JE Jr, Hu WT, Lee FE, Sanz I. Extrafollicular B cell responses correlate with neutralizing antibodies and morbidity in COVID-19. Nat Immunol. 2020;21:1506–16.
- 136. Wrammert J, Onlamoon N, Akondy RS, Perng GC, Polsrila K, Chandele A, Kwissa M, Pulendran B, Wilson PC, Wittawatmongkol O, Yoksan S, Angkasekwinai N, Pattanapanyasat K, Chokephaibulkit K, Ahmed R. Rapid and massive virus-specific plasmablast responses during acute dengue virus infection in humans. J Virol. 2012;86(6):2911–8.
- Yang L, Xie X, Tu Z, Fu J, Xu D, Zhou Y. The signal pathways and treatment of cytokine storm in COVID-19. Signal Transduct Target Ther. 2021;6(1):255.
- 138. Young BE, Ong SWX, Ng LFP, Anderson DE, Chia WN, Chia PY, Ang LW, Mak TM, Kalimuddin S, Chai LYA, Pada S, Tan SY, Sun L, Parthasarathy P, Fong SW, Chan YH, Tan CW, Lee B, Rotzschke O, Ding Y, Tambyah P, Low JGH, Cui L, Barkham T, Lin RTP, Leo YS, Renia L, Wang LF, Lye DC, Singapore Novel Coronavirus Outbreak Research T. Viral dynamics and immune correlates of coronavirus disease 2019 (COVID-19) severity. Clin Infect Dis. 2021;73(9):e2932–42.
- Zeng Z, Zhan J, Chen L, Chen H, Cheng S. Global, regional, and national dengue burden from 1990 to 2017: a systematic analysis based on the global burden of disease study 2017. EClinicalMedicine. 2021;32:100712.
- Zhang J, Hao Y, Ou W, Ming F, Liang G, Qian Y, Cai Q, Dong S, Hu S, Wang W, Wei S. Serum interleukin-6 is an indicator for severity in 901 patients with SARS-CoV-2 infection: a cohort study. J Transl Med. 2020;18(1):406.
- 141. Zhao Y, Qin L, Zhang P, Li K, Liang L, Sun J, Xu B, Dai Y, Li X, Zhang C, Peng Y, Feng Y, Li A, Hu Z, Xiang H, Ogg G, Ho LP, McMichael A, Jin R, Knight JC, Dong T, Zhang Y. Longitudinal COVID-19 profiling associates IL-1RA and IL-10 with disease severity and RANTES with mild disease. JCI Insight. 2020;5(13):e139834.
- 142. Zheng M, Gao Y, Wang G, Song G, Liu S, Sun D, Xu Y, Tian Z. Functional exhaustion of antiviral lymphocytes in COVID-19 patients. Cell Mol Immunol. 2020;17(5):533–5.

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