# Detection of Heterozygotes for Phenylketonuria by Column Chromatography and Discriminatory Analysis

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## Extract

Phenylalanine (100 mg/kg) was administered orally to 19 parents of patients with phenylketonuria and to 26 control subjects. Concentrations of phenylalanine and tyrosine in plasma were determined by column chromatography before phenylalanine administration and at 1, 2, 3, and 4 hours after loading. The values of two control subjects (C 21 and C 25) and of two parents (P 4 and P 6) were unusually low or high. Excluding subjects C21, C25, P4, and P6, the following calculations of the values after loading were made:

Mean and SD of mean of phenylalanine levels at each time: in spite of a significant difference, there was a slight overlap.

Sum of the phenylalanine values: significant difference, no overlap.

Sum of the phenylalanine/tyrosine ratios: significant difference, no overlap.

Using these criteria, control subjects C21 and C25 were considered to be heterozygotes. The values of subject P 4 (father of a known patient) were within the range of the control group; paternity was excluded by blood group analysis. The values of subject P 6 were unusually high and could not be explained satisfactorily.

The concentrations of phenylalanine and tyrosine after loading were evaluated by discriminatory analysis. Thus, subjects G21, G25, and P4 were excluded, but values were calculated both including and excluding subject P 6. To compare the different criteria of evaluation, the index of PENROSE (discriminatory power  $-D/\tilde{s}$ ) was calculated. A better separation was obtained with discriminatory analysis than with the sum of the phenylalanine values or of the phenylalanine/tyrosine ratios. The use of column chromatography and evaluation by discriminatory analysis seems to improve the possibilities of detection of heterozygotes for phenylketonuria.

## Speculation

With discriminatory analysis, the separation of heterozygotes for phenylketonuria from normal subjects was better than with other criteria of evaluation. We believe that this method could be used successfully for determination of heterozygosity in other inborn errors of metabolism with recessive inheritance when several variables are available for each subject examined.

#### Introduction

HSIA et al. [11, 12] first demonstrated that following an oral phenylalanine load, plasma concentrations of phenylalanine were significantly higher in parents of patients with phenylketonuria than in control subjects; however, there was some overlap in individual values. In an effort to obtain a better separation of heterozygotes and normal subjects, different conditions of testing and criteria of statistical analysis were subsequently used by several investigators [1, 3, 4, 9, 10, 13, 15, 21, 22, 23, 28–30], but separation was never completely satisfactory.

This report is a new study of oral phenylalanine loads in which the concentrations of phenylalanine and tyrosine in plasma were determined by column chromatography and evaluated by discriminatory analysis [26].

#### Subjects and Methods

Nineteen parents of patients with phenylketonuria were compared with 26 control subjects having negative family histories of phenylketonuria.

L-phenylalanine (100 mg/kg body weight) was administered in a mixture of 100 ml orange juice and 100 ml water after an overnight fast. Five samples of heparinized venous blood were obtained before and 1, 2, 3 and 4 hours after loading.

An 0.2 ml sample of plasma was deproteinized immediately with 0.8 ml of 5 percent sulfosalicylic acid. After centrifugation, the supernatant was frozen at  $-20^{\circ}$ . Analysis was carried out with an amino acid analyzer with an automatic programming device [31] according to the method of SPACKMAN et al. [27]. It was performed with a short program for phenylalaning and tyrosine based on methods reported previously [2, 16, 19, 25]. A  $42 \times 0.9$  cm column, packed with spheric Bio Rad -A5 cation exchange resin (height 22 cm) of particle size 12–15  $\mu$  was used. For elution, a 0.35 n sodium citrate buffer (pH 5.28) was used, and for regeneration, a 0.4 n NaOH was used. The flow rates of buffer and ninhydrine reagent were 80 and 40 ml/h, respectively, at a column temperature of 57°.

The expansion of the measuring range in a Honeywell recorder from 0-5 mV to 2.5-5.0 mV allowed the use of regular cuvettes. The programmed course of analysis was as follows:

Eluate	$_{ m pH}$ 5.28	NaOH	pH 5.28
Time (min)	0-7	7–15	15-41

The ninhydrine pump and the recorder were stopped at 29 minutes. Thus, time was saved by starting regeneration and equilibration of the column before the end of the analyses. Up to 10 analyses (plus one analysis of a standard mixture) were carried out in one working day.

For this short program, only one buffer was necessary. For good separation of tyrosine and phenylalanine, a high column temperature was more important than the pH of the buffer.

## Results and Statistical Evaluation

The values of phenylalanine and tyrosine before and after loading are shown in table I (controls) and table II (parents). As expected, the phenylalanine concentrations after loading were higher in the heterozygotes and the tyrosine concentrations were higher in the control subjects. In the control group, however, unusually high phenylalanine and low tyrosine concentrations were found in two control subjects, C21 and C25, after loading. Unusual values were also found in two parents. Subject P 4 had low phenylalanine and high tyrosine values and subject P 6 had very high phenylalanine and low tyrosine concentrations. For better analysis of these four subjects, the following calculations were made:

1. Mean values and standard deviations of the concentration of phenylalanine in the two groups each hour after load, excluding subjects C 21, C 25, P 4, and P 6 (table IV, fig. 1).

2. Sum of the phenylalanine concentrations of all subjects 1, 2, 3, and 4 hours after loading (tables I and II, fig.2); mean and standard deviation, excluding subjects C 21, C 25, P 4, and P 6 (table IV).

3. Sum of the phenylalanine/tyrosine ratios of all subjects 1, 2, 3, and 4 hours after loading (tables I and II, fig.2); mean and standard deviation, excluding subjects C 21, C 25, P 4, and P 6 (table IV). The phenylalanine/tyrosine ratio = plasma phenylalanine mg/100 ml divided by plasma tyrosine mg/100 ml.

Excluding the four subjects with unusual values (C21, C25, P4, and P6), the 24 control subjects, whose values were analyzed by use of the three calculations, represented a homogenous group with a small variability; the variability was larger in the 17 heterozygotes. The difference between the two groups was significant. If calculation 1 was applied, there was some overlap between the two groups. If calculations 2 and 3 were used, the two groups were completely separated (table IV).

#### The Two Unusual Control Subjects

In subjects C 21 and C 25, concentrations of phenylalanine in plasma were definitely in the range of the heterozygotes (fig. 1). The phenylalanine concentrations at hours 2, 3, and 4 were more than 3 SD above the values of the other 24 controls. They were also in

		ars)	(kg)			nenylalan 100 l l					Tyrosina 00 ml pl			$\Sigma \operatorname{Phc}^1$	$rac{\Sigma}{( m Phc)}$ (Phc/		
Subject	Sex	Age (years)	Weight (kg)	0 h	l h	100 ml pl 2 h	3 h	4 h	0 h	1 h	2 h	3 h	4 h	mg/ 100 ml	- /	Formula 1 (with P6)	
C 1	m	26	78	0.94	8,07	7.54	5.00	3.35	1.19	2.57	3.10	3.28	3.22	23.96	8.13	2.094	1.219
C 2	f	23	56	1.06	6.42	8.22	6.06	4.74	1.05	2.28	2.61	2.95	2.88	25.44	9.67	1.867	3.149
C 3	f	20	48	0.68	11.24	7.11	5.07	2.72	1.06	2.24	2.23	2.46	1.70	26.14	11.87	2.814	4.346
G4	m	19	63	0.77	7.21	6.68	3.63	3.47	1.09	1.68	2.46	2.05	2.33	20.99	10.27	0.883	1.841
C 5	m	24	61	0.77	8.40	7.61	5.16	3.25	1.02	2.21	3.08	2,44	2.28	24,42	9.81	1.610	3.429
C 6	m	20	65	0.68	7.46	6.06	4.82	3.40	0.80	2.50	2.52	2.77	2.46	21.74	8.50	-1.094	0.064
C 7	III	32	76	0.82	14.78	5.59	4.42	3.28	1.13	2.50	2.46	2.44	2.46	28.07	11.32	5.776	5.328
C 8	m	19	64	0.75	11,47	6.50	4.44	3.25	0.97	2.66	3.24	2.44	2.64	25.66	9.37	2.502	2.520
C 9	f	30	52	0.75	8.43	5,43	3.02	2.06	1.09	3.04	3.00	2.12	1.92	18.94	7.07	-4.631	-4.880
G 10	m	42	51	0.78	10.69	6.06	3.40	2.28	1.26	2.79	2.70	2.88	2.28	22.43	8.25	-4.073	-3.396
C 11	f	20	49	0.78	11.12	7.95	5.30	2.89	1.09	2.39	2.72	2,70	2.15	27.26	10.87	3.685	4.443
C 12	m	36	65	0.90	11.30	6.62	3.63	2.64	0.79	2.28	2.33	I.77	1.50	24.19	11.61	1.066	1.766
C 13	ſ	19	53	0.82	11.09	9.90	6.06	3.27	1.08	2.60	3.26	3.35	2.66	30.32	10.35	2.997	4.050
C 14	f	20	57	1.08	8.43	8.68	6.24	5.21	1.46	2.53	3.04	3.31	3.57	28.56	9.54	3.862	3.998
C 15	f	22	45	1.10	10.56	5.86	3.96	2,56	1.28	2.75	2.75	2.44	1.81	22.94	9.00	-4.523	-1.987
G 16	f	18	52	1.06	10.89	8.10	5.11	3.50	1.42	2.64	3.22	3.53	3.17	27.60	9.20	0.173	1.140
C 17	ſ	35	58	1.14	12.64	6.47	4.82	3.66	1.65	3.31	3,62	3.14	3.04	27.59	8.36	-2.078	-0.827
C 18	ſ	23	61	1.14	9.08	6.85	5.46	3.70	1.41	2.46	3.08	3.40	3.28	25.09	8.65	1,805	2.673
C 19	f	23	45	0.91	7.89	5.12	3.98	3.25	0.91	1.74	2.17	2.14	2.39	20.24	10.11	3.612	3.767
C 20	<b>D</b> 1	38	60	1,26	13.15	6.55	5.61	3.30	1.62	3.46	2.95	3.40	2.57	28.61	8.95	-1.495	-0.139
G 21	m	34	75	1.78	14.12	15.71	11.62	9,72	1,38	1.74	1.61	1.83	1.86	51.17	29.46	19.559	23.628
C 22	f	25	54	1.01	7.06	6.24	4.59	3.22	1.23	2.50	2.79	3.10	2.69	21.11	7.74	-3.601	-1.729
C 23	$\mathbf{m}$	27	58	1.06	7.71	5.31	3.50	2.31	1.03	2.06	2.43	2.37	1.90	18.83	8.63	-3.667	-1.263
C 24	m	31	61	0.96	5.13	6.91	4.31	2.99	1.12	1.81	2.10	2.05	1.83	19.34	9.85	1.117	1.731
C 25	f	23	64	0.90	14.04	12.21	7.97	5.40	0.97	1.38	1.56	1.67	1,72	39.62	25.91	22.041	20.423
C 26	m	34	59	1.00	9.65	8.46	4.09	2.74	1.00	2.73	3.57	3.11	2.64	24.94	8.26	-4.670	-2.862

# Table I. Oral L-phenylalanine load. Results in 26 controls

<sup>1</sup> Sum of the phenylalanine concentrations 1, 2, 3, and 4 hours after load. <sup>2</sup> Sum of the phenylalanine/tyrosine ratios 1, 2, 3, and 4 hours after load.

<sup>3</sup> For explanation see text and table III.

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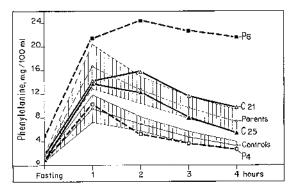


Fig. 1. Results of oral L-phenylalanine load (100 mg/kg). Plasma phenylalanine concentrations. Hatched areas: mean  $\pm 1$  SD at each time in 24 of 26 controls (C 21 and C 25 excluded) and in 17 of 19 parents (P 4 and P 6 excluded). The values of C 21, C 25, P 4 and P 6 are shown individually.

	Σ Ph (1,2,3	e and 4 h		C(Phe/1 ooding)	[yr]
	С	Р		C	P 114 ∎ P6
90_		■ P6			着 P6
80_			40_		
70_			35_		••
60_		1	30_	с 4.21	
50_	C ∆21	i	25_	▲25	:
40.	<b>▲</b> 25	.i.	20		7
30_		t •	15.		:
20_	*	🗆 P4	10.	X.	•
10_	-		5_	.xp.	£0 P4
•					

Fig. 2. Results of oral L-phenylalanine load (100 mg/kg). 26 controls (C) and 19 parents (P). Left; sum of the plasma phenylalanine concentrations 1, 2, 3, and 4 hours after load. Right: sum of the phenylalanine/tyrosine ratios 1, 2, 3, and 4 hours after load.

Fig. 3. Results of oral L-phenylalanine load (100 mg/kg). Individual discriminant scores in all 26 controls (C) and in all 19 parents (P). C 21, C 25, and P 4 were excluded from the discriminatory analysis. Formula 1: P 6 included in the discriminatory analysis. Formula 2: P 6 excluded. The line between the controls and the heterozygotes represents the critical line according to PENROSE (see table IV).

the range of the heterozygotes and more than 3 SD above the values of the other 24 controls if calculations 2 and 3 were applied (fig. 2).

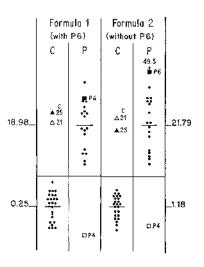
The frequency of the heterozygotes for phenylketonuria in the general population is believed to be 1:50 [5-8, 17, 18]. Each individual in the control group had therefore a probability of 1:50 of being a heterozygote. By binomial distribution, it can be calculated that a random sample of 26 subjects will include two heterozygotes with a probability of 8 % [32]. Since the values of subjects C 21 and C 25 were well within the range for heterozygotes, these subjects were considered as such and were excluded from the sample on which discriminatory analysis was performed.

#### The Two Unusual Parents

The values of subject P 4 (father of a known patient) were definitely within the control range using all three calculations (figs. 1 and 2). Paternity could be excluded with practical certainty by blood group analysis in this family [33]. Subject P 4 was therefore omitted for discriminatory analysis. In subject P6 (mother of a known patient, but not married to subject P 4), the initial increase of phenylalanine concentration after loading was not unusually high, but the values remained considerably elevated at the 2nd, 3rd, and 4th hour after loading (fig. 1). In this case, the fasting phenylalanine concentration was already unusually high (4.97 mg/ 100ml). Tyrosine concentrations after loading remained very low and the sums of the phenylalanine values and of the phenylalanine/tyrosine ratios were excessively high (table II, fig. 2). This behavior remains unexplained.

#### Discriminatory Analysis

For the purpose of this study, only the unquestionable subjects were considered, and subjects C 21, C 25, and P4 were therefore excluded. Discriminatory anal-



		(sı	kg)			henylalan					Tyrosin			$\Sigma$ Phe <sup>1</sup>	$\Sigma$ (Phe/		ant score <sup>3</sup>
ct		vea	ot (		mg/	(100  ml pl)	lasma				00 ml pl			mg/	Tyr) ²		Formula 2
Subject	Sex	Age (years)	Weight (kg)	0 h	1 h	2 h	3 h	4 h	0 h	1 h	2 h	3 h	4 h	100 ml		(with P6)	(without P 6)
P 1	f	26	62	1.02	12.75	9.14	7,13	4.64	1.23	2.21	2,72	2.79	2.82	33.66	13.34	12.232	12.000
Р2	m	30	83	1.53	11.65	11.04	8.86	8.35	1.21	1.85	1.63	1.73	1,99	39.90	22.39	10.560	14.894
Р3	f	25	52	1,23	18.85	11.14	8.63	5.69	0.66	1.70	2.63	2.76	2.52	44.31	20.70	16.822	20.018
P 4	m	36	75	1.27	10.26	5.26	3.70	2.61	1.32	2,90	3.22	2.82	2.41	21.83	7.56	-6.381	3.788
₽5	f	39	55	1.03	11.53	11.24	8.51	5.33	0.78	1.76	2.41	2.66	2.30	36.61	16.73	13.304	15.374
P 6	f	28	62	4.97	21.58	24,45	22.69	21.58	1.01	0.80	0.80	0.80	0.76	90.30	114.29	25.089	49.539
P 7	m	32	55	1.31	21.83	13.32	10.41	7.46	1.20	1.56	1.77	1.83	1.65	53.02	31.73	24.465	27.753
P 8	f	34	64	1.35	22.97	16.81	12.24	8.58	1.10	1.77	2.33	2.50	2.33	60.60	28.77	28.744	31.377
P 9	m	33	85	1.25	16.96	13.05	8.79	6.30	1.04	1.45	1.68	1.97	1.90	45,10	27.25	22.012	22.283
P 10	f	39	51	1.33	15.38	10.65	6.88	4.80	1.32	2.01	2.32	2.14	1.94	37.71	17.92	12.495	13.344
Р 11	m	39	73	1.61	20.39	15.81	10.03	9.31	1.28	1.59	1,74	1.34	1.67	55.54	34.97	21.175	24.826
P 12	f	38	54	1.42	15.91	15.91	12.69	9.83	1.33	1.48	1.92	1.99	1.99	54.34	30.36	23.035	28.506
P 13	m	67	74	1.48	22.54	14.70	11.70	10.54	1.38	1.59	1.72	1.72	1.94	59.48	34.96	21.623	28.621
P 14	f	30	65	1.17	14.32	15.13	12.49	9.92	0.71	1.36	1.76	1.76	1.86	51.86	31.56	21.812	27.736
P 15	m	36	75	1,23	14.70	11.07	8.04	6.42	1.01	1.92	2.73	2.73	2.70	40.23	17.04	9.873	13.851
P 16	f	33	60	1.25	18,15	10.21	7.66	5.03	0.97	1.70	1.97	2.10	1.90	41.05	22.16	18.026	19.006
P 17	m	34	80	1.28	18.63	12.44	10.03	7.11	0.82	1.25	1.63	1.70	1.70	48.21	32.61	24.565	27.188
P 18	f	42	57	1.39	15.11	12.29	10.03	8.40	1.28	1.63	2.17	2.50	2,88	45.83	21.86	18.092	21.419
Р 19	m	51	80	1.41	12.21	11.60	11.76	8.66	1.57	2.05	2.10	2.32	2,37	44.23	20.20	17.683	22.207

 $^1$  Sum of the phenylalanine concentrations 1, 2, 3, and 4 hours after load.  $^2$  Sum of the phenylalanine/tyrosine ratios 1, 2, 3, and 4 hours after load.

<sup>3</sup> For explanation see text and table III.

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ysis [26] was then performed with and without subject P 6. With this statistical method, the individual variables were combined by coefficients to a discriminant score for each subject, and the best possible separation of two groups was obtained. In each subject, 10 measurements or 10 variables (5 phenylalanine and 5 tyrosine values) were available. Various combinations of the variables were chosen. In the present context, only the results of discriminatory analysis using all phenylalanine and tyrosine concentrations after loading (a total of 8) are presented.

The coefficients arc shown in table III. Formula 1 was obtained including subject P 6 and formula 2 was obtained excluding this subject. The signs of the coefficients agreed in both formulae, but size showed considerable difference.

The discriminant scores (for calculation, see table III) of the 26 control subjects and of the 19 parents are shown in tables I and II and in figure 3. In figure 3, the critical line separating the scores of the two groups (see table IV) is also shown. The scores of control subjects C 21 and C 25 were again definitely in the range of those of heterozygotes and the score of subject P4 was in the range of that of the control group. Subject P 6 showed an excessively high score when calculated by use of formula 2 only.

Other discriminatory analysis that included control subjects C 21 and C 25 was performed. The coefficients obtained are shown in table III (formulas 3 and 4). The results of statistical evaluation of the discriminant scores obtained with these formulas are shown in table IV. The discriminant scores of subjects C 21 and C 25 (8.01 and 9.78 with formula 3, and 10.29 and 7.73 with formula 4, respectively) were the highest of the control group and were the only ones that overlapped with those of the parents.

A final discriminatory analysis was performed including subjects C21 and C25 in the parent (or hetero

Table III. Oral L-phenylalanine load. Results of discriminatory analysis
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Variables:		Phenyl	alanine	Tyrosine				
concentration after load (h)	Ī	2	3	4	1	2	3	4
Coefficients				•				
A. C 21, C 25, and P 4 excluded								
Formula 1 (with P 6)	0.63	1.06	4.24	-4.92	-0.39	-5.76	-10.77	11.95
Formula 2 (without P 6)	0.63	0.21	3.57	-2.11	-3.43	-1.76	-5.44	4.93
B. C 21 and C 25 included,								
P 4 excluded								
Formula 3 (with P 6)	0.72	-0.71	2.56	-1.66	-4.42	1.30	-3.73	4.17
Formula 4 (without P 6)	0.74	-1.48	2.07	0.62	-7.30	4.81	0.63	-1.69
C. Formula 5								
(for explanation see text)	0.65	0.36	3.86	-2.56	-3.10	-2.30	-6.54	5.82

*Remarks:* With these formulae one calculates a discriminant score for each subject. It is obtained by multiplying the phenylalanine and tyrosine concentrations with the respective coefficient and by adding these products.

#### Notes table IV:

- <sup>1</sup> 24 controls: C 21 and C 25 excluded
  - 26 controls: C 21 and C 25 included
  - 17 parents: P 4 and P 6 excluded
  - 18 parents: P 4 excluded

25 'controls' (for formula 5 only): 24 controls (C 21 and C 25 excluded), P 4 included in the 'control' group. 19 'heterozygotes' (for formula 5 only): 17 parents (P 4 and P 6 excluded), C 21 and C 25 included in the 'heterozygote' group.

<sup>2</sup> t according to SNEDECOR and COCHRAN ([26], chapter 4.10, p. 104). P of all ts  $\ll 0.001$ .

<sup>3</sup> Discriminatory power according to PENROSE [20].

<sup>4</sup> Critical line according to Penrose [20] (C =  $[M_1s_2+M_2s_1]/[s_1+s_2]$ ) at 7.05.

<sup>6</sup> Critical line at 7.56.

<sup>6</sup> Critical line at 7.88.

$\begin{array}{c ccccc} \mbox{Phenylalanine concentration} & 1 \mbox{hours} & 17 \mbox{ parents} & 16.70 & 3.76 & 11.5-23.0 \\ \mbox{after load (mg/100 ml plasma)} & 1 \mbox{hours} & 17 \mbox{ parents} & 16.97 & 3.83 & 11.5-23.0 \\ \mbox{11 parents} & 12.68 & 2.26 & 9.1-14.8 \\ \mbox{2 hours} & 17 \mbox{ parents} & 12.68 & 2.26 & 9.1-16.8 \\ \mbox{2 hours} & 17 \mbox{ parents} & 12.68 & 2.26 & 9.1-16.8 \\ \mbox{2 hours} & 17 \mbox{ parents} & 13.33 & 3.54 & 9.1-24.5 \\ \mbox{2 hours} & 17 \mbox{ parents} & 13.33 & 3.54 & 9.1-24.5 \\ \mbox{2 hours} & 17 \mbox{ parents} & 19.76 & 1.90 & 6.9-12.7 \\ \mbox{2 hours} & 17 \mbox{ parents} & 10.48 & 3.56 & 6.9-22.7 \\ \mbox{2 hours} & 17 \mbox{ parents} & 10.48 & 3.56 & 6.9-22.7 \\ \mbox{2 hours} & 17 \mbox{ parents} & 10.48 & 3.56 & 6.9-22.7 \\ \mbox{2 hours} & 17 \mbox{ parents} & 10.48 & 3.56 & 6.9-22.7 \\ \mbox{2 hours} & 17 \mbox{ parents} & 1.43 & 1.94 & 4.6-10.5 \\ \mbox{2 hours} & 17 \mbox{ parents} & 1.43 & 1.94 & 4.6-10.5 \\ \mbox{2 hours} & 17 \mbox{ parents} & 2.435 & 3.35 & 18.8-30.3 \\ \mbox{2 hours} & 17 \mbox{ parents} & 24.35 & 3.35 & 18.8-30.3 \\ \mbox{2 hours after load} & 17 \mbox{ parents} & 24.97 & 6.97 & 13.3-35.0 \\ \mbox{2 hours after load} & 17 \mbox{ parents} & 24.97 & 6.97 & 13.3-35.0 \\ \mbox{2 hours after load} & 17 \mbox{ parents} & 24.97 & 6.97 & 13.3-35.0 \\ \mbox{2 hours after load} & 12 \mbox{ parents} & 24.97 & 6.97 & 13.3-35.0 \\ \mbox{2 hours after load} & 24 \mbox{ controls} & 9.39 & 1.24 & 7.1-11.9 \\ \mbox{2 hours after load} & 12 \mbox{ parents} & 18.98 & 5.54 & 9.67-28.74 \\ \mbox{4 hours after load} & 12 \mbox{ parents} & 18.98 & 5.54 & 9.67-28.74 \\ \mbox{4 hours after load} & 12 \mbox{ parents} & 18.98 & 5.54 & 9.67-28.74 \\ \mbox{4 hours after load} & 12 \mbox{4 controls} & 0.25 & 3.15 & -467-5.78 & -4 & 13.87 & 4.3 & 1.6 \\ \mbox{1 hours after load} & 12 \mbox{ controls} & 1.94 & 2.88 & -2.91-9.78 & + & 10.43 & 3.2 & 5.5 \\ \mbox{1 hours after load} & 12 \mbox{4 controls} & 1.94 & 2.88 & -2.91-9.78 & + & 10.43 & 3.2 & 5.5 \\ \mbox{1 hours after load} & 12 \mb$			Subjects <sup>1</sup>	Mean	SD	Range	Overlap	test <sup>2</sup>	D/s²	Classifica- tion error (%)
after load (mg/100 ml plasma) 24 controls 9.58 2.34 $5.1-14.8 > + 7.74$ 2.4 11.6 18 parents 16.97 3.83 $11.5-23.0 > + 7.74$ 2.4 11.6 2 hours 17 parents 12.68 2.26 9.1-16.8 24 controls 6.91 1.20 $5.1-9.9 > + 8.32$ 2.7 8.8 3 hours 17 parents 13.33 3.54 9.1-24.5 $+ 8.32$ 2.7 8.8 3 hours 17 parents 9.76 1.99 $6.9-12.7 > - 11.53$ 3.7 3.4 18 parents 10.48 3.56 $6.9-22.7 > - 7.72$ 2.6 9.5 4 hours 17 parents 7.43 1.94 $4.6-10.5 > + 9.85$ $3.2$ 2.5 18 parents 8.22 3.83 $4.6-21.6 > + 6.30$ 2.2 13.5 Sum of phenylalanine concentrations 17 parents 46.57 8.10 33.7 $-60.6 > - 8.96$ 3.0 $-6.2 > - 8.96$ 3.0 $-6.6 > - 8.96$ 3.0 $-1.2 > - 8.96$ 3.0 $-6.6 > - 8.96$ 3.0 $-6.6 > - 8.96$ 3.0 $-6.6 > - 8.96$ 3.0 $-6.6 > - 8.96$ 3.0 $-6.6 > - 8.96$ 3.0 $-6.6 > - 8.96 = - 1.6 = - 8.96 = - 1.6 = - 8.96 = - 1.6 = - 8.96 = - 1.6 = - 8.96 = - 1.6 = - 8.96 = - 1.6 =$	Phenylalanine concentration	l hour	17 parents	16.70	3.76	11.5- 23.0			_	<u> </u>
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$\begin{array}{c c c c c c c c c c c c c c c c c c c $	·····						> +	7.74	2.4	11.6
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$		2 hours								
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$										
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$			18 parents				> +	8.32	2.7	8.8
$\begin{array}{cccccccccccccccccccccccccccccccccccc$		3 hours	н		1.90					
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$			*	•	> —					
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$			18 parents		3.56		> —	7.72	2.6	9.5
$\begin{array}{c ccccccccccccccccccccccccccccccccccc$		4 hours	-	7.43	1.94			0.05	0.0	
$\begin{array}{c c c c c c c c c c c c c c c c c c c $			24 controls	3.21	0.70	2.1- 5.2				
$\begin{array}{c c c c c c c c c c c c c c c c c c c $			18 parents	8.22	3.83	4.6-21.6	> +	6.30	2.2	13.5
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Sum of phenylalanine concentra	tions	17 parents	46.57	8.10	33.7- 60.6				
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	(1, 2, 3, and 4 hours after load)		24 controls	24.35	3.35	18.8- 30.3	>			
$ \begin{array}{c} (1,2,3,\text{ and 4 hours after load}) \\ (1,2,3,\text{ and 4 hours after load}) \\ \hline 24 \text{ controls} \\ 18 \text{ parents} \\ \hline 29.94 \\ 29.94 \\ 22.11 \\ 13.3-114.3 \\ \hline \end{array} \\ \begin{array}{c} > - \\ - \\ - \\ - \\ - \\ - \\ - \\ - \\ - \\ -$			18 parents	49.00	12.96		> —	8.96	3.0	0.0
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	Sum of phenylalanine/tyrosine ra	atios	17 parents	24.97	6.97	13.3- 35.0		10.70		
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	(1, 2, 3, and 4 hours after load)		24 controls	9.39	1.24	7.1-11.9	> —			
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$			18 parents	29.94	22.11	13.3 - 114.3	> —	4.56	1.8	18.9
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$	Discriminant scores	Formula 1	24 controls	0.25	3.15	-4.67- 5.78	4	13.87	4.3	1.6
17 parents       21.79       6.24       12.00-31.38         Formula 3       26 controls       1.94       2.88       -2.91-9.78       +       10.43       3.2       5.5         18 parents       12.17       3.61       5.20-18.24       5.20-18.24       5.2       5.2         Formula 4       26 controls       1.95       2.87       -3.95-10.29       +       10.98       3.3       4.7         17 parents       13.68       4.15       7.39-20.14       5.5       -6       15.39       4.7       0.9         Formula 5       25 'controls'       0.59       3.09       -5.19-5.25       -6       15.39       4.7       0.9			18 parents	18.98	5.54	9.87-28.74				
Formula 3         26 controls         1.94         2.88         -2.91-9.78         +         10.43         3.2         5.5           18 parents         12.17         3.61         5.20-18.24<		Formula 2	24 controls	1.18	2.80	-4.88~ 5.33	5	14.31	4.6	1.1
$ \begin{array}{c ccccccccccccccccccccccccccccccccccc$			17 parents	21.79	6,24	12.00-31.38				
Formula 4 26 controls 1.95 2.87 -3.95-10.29 + 10.98 3.3 4.7 17 parents 13.68 4.15 7.39-20.14 Formula 5 25 'controls' 0.59 3.09 -5.19- 5.256 15.39 4.7 0.9		Formula 3	26 controls	1.94	2.88	-2.91- 9.78	+	10.43	3.2	5.5
17 parents 13.68 4.15 7.39–20.14 Formula 5 25 'controls' 0.59 3.09 –5.19– 5.25 — 6 15.39 4.7 0.9	•		18 parents	12.17	3.61	5.20 - 18.24				
17 parents 13.68 4.15 7.39-20.14 Formula 5 25 'controls' 0.59 3.09 -5.19- 5.25 6 15.39 4.7 0.9		Formula 4	26 controls	1.95	2.87	-3.95 - 10.29	+	10.98	3.3	4.7
			17 parents	13.68	4.15	7.3920.14				
19 'hetero- 22.52 6.20 12.29–32.63		Formula 5	25 'controls'	0.59	3.09	-5.19- 5.25	6	15.39	4.7	0.9
			19 'hetero-	22.52	6.20	12.29-32.63				
zygotes'			zygotes'							

# Table IV. Oral L-phenylalanine load. Results of statistical evaluation by different criteria

Authors		Subjects C = controls P = parents	Phenylalanine load : mg/kg	Criterion $\langle Phc = phenylalanine, Tyr = tyrosine \rangle$	Overlap	D/s	Classifica- tion error (%)
HSIA et al. [11, 12]	1956	<b>C</b> : 19	100 p.o.	Phe 1 h	+ 2	3.2 2	5.5
		P: 19		$\Sigma$ Phe (1+2+4 h)	- - <sup>2</sup>	3.7²	3.2
BERRY et al. [3]	1957	C: 10 P: 10	100 p.o.	Area under phenylalanine curve (sq.in.)	3	3.03	6.7
LIPPMAN et al. [15]	1960	C: 19 P: 17	î.v.	Phe 3 h	—		
Jervis [13]	1960	C: 20 P: 21	330 p.o.	Tyr 2 h	—	3.2	5.5
RENWICK et al. [23]	1960	C: 45 P: 39	100 p.o.	$Log_{10}$ (Phe 4 h), corrected for weight	<u> </u>	3.3	4.9
WANG et al. [29]	1961	C: 19 P: 17	100 p.o.	discriminant score from: Phe 0, 1 $\frac{1}{2}$ , 2 and 3 h		3.6	3.6
ANDERSON et al. [1]	1962	C: 29	100–200 p.o.	discriminant score from:			
		P: 14	(two doses load)	– Tyr 0, 1 and 2 h	+	3.1	5.6
			· · · · ·	– Tyr 1 and 2 h	+	3.5	3.9
BREMER and NEUMANN [4]	1966	C:17 P:9	50 i.v.	biological half-time of phenylalanine elimination		2.8	8.0
WOOLF et al. [30]	1967	C: 34 P: 31	80 i.v.	index of rate of phenylalanine elimination in relation to total body water		2.8	8.0
				discriminant on the basis of phenylalanine and		\$ 2.7	8.8
				tyrosine concentration		Ç 4,8	0.8
PERRY et al. [21]	1967	C: 77 P: 43	no load	Phe/Tyr ratio fasting	+	2.554	10.1
Present study	1968		100 p.o.	discriminatory analysis			
,		C: 24 <sup>5</sup> P: 18		- formula 1 (with P 6)		4.3	1.6
		C: 24 <sup>5</sup> P: 17		– formula 2 (without P 6)		4.6	1.1
		C: 26 P: 18		– formula 3 (with P 6)	÷	3.2	5.5
		P: 18 C: 26 P: 17		– formula 4 (without P 6)	+	3.3	4.7

Table V. Detection of heterozygotes for phenylketonuria. Discriminatory power  $(D/\bar{s})$  and classification error in earlier studies<sup>1</sup> and in the present investigation  $V_{++}^{0}$ 

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zygote) group and subject P 4 in the control group. The coefficients obtained in this way (formula 5, table III) were similar to those of formula 2 (table III). Statistical evaluation of the scores calculated with this formula is shown in table IV.

#### Discussion

In this study, concentrations of phenylalanine and tyrosine in plasma after an oral phenylalanine load were determined by column chromatography and the results were evaluated by discriminatory analysis. Column chromatography seemed to be more accurate and advantageous than other techniques because both amino acids could be determined in one step.

In control subjects G 21 and G 25, the values (phenylalanine and tyrosine concentrations, sum of the phenylalanine/tyrosine ratios after loading) were very unusual compared with those of the other 24 controls. Since the values were definitely within the range of those of the heterozygotes, these subjects were considered as such. We thought, therefore, that for examination of a possible heterozygote, it would be appropriate to use the calculations that were performed, excluding these two controls as well as the parent P 4 (paternity excluded by blood group analysis).

To determine whether discriminatory analysis allows a better separation of the heterozygotes from the control subjects than other criteria, the index of PEN-ROSE [20] was used:

$$D/\bar{s} = \frac{M_1 - M_2}{V_2 (s_1 + s_2)}$$
 where  $\frac{M}{s} = mean value and s = standard deviation.$ 

This equation represents the difference of the mean values divided by the mean of the standard deviations of the two groups. With this index, the discriminatory power of a criterion is expressed as a number. The  $D/\bar{s}$  of the criteria used in this study, as well as other statistical data, is shown in table IV. Subjects C 21, C 25, and P 4 were excluded from these calculations. Sub-

Notes table V:

- <sup>1</sup> Only PERRY *et al.* [21] used column chromatography.
- <sup>2</sup> l control excluded (probable heterozygote).
- <sup>3</sup> 1 control excluded (probable heterozygote).
- <sup>4</sup> 3 controls excluded (probable heterozygotes).
- <sup>5</sup> 2 controls excluded (probable heterozygotes).

jects C 21 and C 25 were included in the control group for calculation of formulas 3 and 4 only. In contrast, the calculations were made with and without the parent P 6 (mother with unusually high phenylalanine and low tyrosine values after loading). The index of PENROSE of the discriminatory analysis was superior to that of the other criteria, especially if subject P 6 was included in the calculations (formulas 1 and 2).

As indicated by the classification error (table IV) and by figure 3, even with these formulas (subjects C 21 and C 25 excluded), some overlapping might be expected in a larger number of subjects. The variability in the heterozygotes was larger than in the controls, who represented a homogenous group (figs. 1, 2, and 3). It is conceivable that this observation indicates a genetic heterogeneity of the alleles on the phenyl-ketonuria locus [24, 30] or the presence of other modifying genes.

The results of other investigators and of our study are summarized in table V. If formula 1 or preferably the more logical formula 2, which we recommend for practical use, is applied, only the results of WOOLF [30], who used an intravenous load, were partially better (in females) than the results in the present study. If formulas 3 and 4, which were calculated including the unusual control subjects C 21 and C 25, are applied, the results are in the range of those of other authors. In some of these studies, unusual controls were also excluded.

The use of column chromatography and of discriminatory analysis seems, therefore, to improve the possibility of detection of heterozygotes for the gene of phenylketonuria.

#### Summary

Phenylalanine (100 mg/kg) was given orally to 26 controls and 19 heterozygotes. Concentrations of phenylalanine and tyrosine in plasma were determined by column chromatography before and 1, 2, 3, and 4 hours after loading. Using as criteria the phenylalanine concentrations, the sum of the phenylalanine values and the sum of the phenylalanine/tyrosine ratios after loading, two controls were considered to be heterozygotes. A father of a patient had, on the contrary, values within the range of the controls; paternity was excluded by blood group analysis. These three subjects were therefore excluded from discriminatory analysis. Another heterozygote had unusually high values. Discriminatory analysis was performed with and without this subject. The index of PENROSE  $(D/\bar{s})$  shows that a better separation of the heterozygotes from the control subjects is obtained with discriminatory analysis than with other criteria [34].

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- 31. Bio Cal Instruments GmbH, Munich/Germany, model BC 200.
- 32. Probability to include 0 heterozygotes = 59.2 %, l = 31.4 %, 3 = 1.3 % and 4 or more 0.2 %.
- 33. Father (P4):  $A_1$ , MsNs,  $P_1+$ , ccddee, K -, Fy<sup>a</sup>-. Mother:  $A_1$ , MsMs,  $P_1+$ , CC<sup>w</sup>DEe, K -, Fy<sup>a</sup>-. Child:  $A_1$ , MsNs,  $P_1+$ , CC<sup>w</sup>Dec, K -, Fy<sup>a</sup>+.
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- 35. Requests for reprints should be addressed to: SER-GIO RAMPINI, M.D., Universitäts-Kinderklinik, Steinwicsstrasse 75, 8032 Zurich (Switzerland).