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Determination of lead by hydride generation atom trapping flame atomic absorption spectrometry

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An atom trapping approach is described for the determination of lead by hydride generation flame atomic absorption spectrometry. Lead hydride (PbH₄), generated on-line by reacting lead in hydrochloric acid–potassium ferricyanide medium with sodium borohydride (NaBH₄), was trapped on the interior walls of a slotted T-tube under highly oxidizing flame conditions. Atomization was achieved by aspirating 50 µL of methyl isobutyl ketone (MIBK) to the flame. Optimization of the experimental parameters for the generation and collection of lead hydride was described to achieve the highest peak height sensitivity. The detection limits were 0.075, 0.047 and 0.028 µg L⁻¹ for 2.6, 5.2 and 7.8 mL of blank solution (n = 13), respectively. Calibration was linear up to 3.0 µg L⁻¹ for a 30-s trapping period. The relative standard deviation was between 2 and 4%. The method was successfully applied to the determination of Pb in NIST certified reference materials, San Joaquin Soil (SRM 2709) and Apple Leaves (SRM 1515).

Introduction

Flame atomic absorption spectrometry (FAAS) is a robust, low cost and simple technique. However, the detection limits of conventional FAAS are not sufficiently low to accurately determine most heavy metals, such as lead (Pb), at trace and ultra-trace levels, because of the limited atom number density in the light path associated with poor nebulization and atomization. Thus, FAAS is usually interfaced to on-line preconcentration and hydride generation procedures to improve the detection limits for the determination of Pb in environmental and clinical samples.¹⁻³ On-line preconcentration via retention of Pb within a certain pH on a micro-column of a solid support improves the sensitivity and selectivity.^{4,5} Various procedures have been described for the determination of Pb by FAAS.⁶⁻¹⁵ Most frequently, C₁₈ bonded silica gel sorbent¹¹⁻¹⁵ is used as the support material, while other adsorbents, including activated carbon,¹⁶ alumina,¹⁷ cellulose,¹⁸ and Chromosorb 102,¹⁹ have also been used.

Hydride generation (HG) has been a powerful means of sample introduction for the determination of hydride-forming elements, such as arsenic (As), antimony (Sb), bismuth (Bi) and selenium (Se), by FAAS due to the efficient transport and atomization of gaseous hydride.^{20,21} Despite these advantages, the determination of Pb by hydride generation has been hindered by low chemical yield of lead hydride (plumbane, PbH₄) for which Pb²⁺ needs to be oxidized to the Pb⁴⁺ oxidation state. In acidic solutions, the chemical yield of PbH₄ is limited by the relatively low

redox potential of Pb^{2+}/Pb^{4+} (-1.455 V).^{20–22} A number of oxidant–acid mixtures, including (NH₄)₂S₂O₈–HNO₃ and H₂O₂–HNO₃,^{23–25} K₂Cr₂O₇–lactic acid,²⁶ H₂O₂–HCl,²⁷ and K₃Fe(CN)₆–HCl,^{28–31} have been used to improve the lead hydride generation efficiency. Flow injection hydride generation methodology has also been successfully utilized for the determination of lead in various samples, such as air filters,²⁴ water samples,²⁷ calcium supplements,²⁹ clinical samples,^{30,31} and wine³¹ by FAAS.

Atom trapping approaches that involve the collection and in situ preconcentration of analyte using a slotted silica and quartz tube were first demonstrated by Watling³² and Lau et al.,³³ and since then have attracted considerable interest to achieve lower detection limits by FAAS.³⁴⁻³⁶ The improvement in sensitivity was about 3- to 5-fold for a number of volatile elements, including Pb, Cd, Bi, Co, Mn, Ag, As, Sb, Cu and Se, when the mist of sample solution was continuously introduced to a slotted quartz tube atom trap.³² Trapping the analyte vapor over a period of time by condensation on the exterior walls of a water-cooled silica tube afforded better sensitivity by factor of 10-50. A different approach for atom trapping was reported by Huang et al.³⁷ with a slotted-quartz tube under highly oxidizing flame conditions followed by atomization by changing the fuel-to-oxidant ratio of the flame. In a comparative study of a bare slotted silica tube trap and a water-cooled silica trap in slotted silica tube, better sensitivity was achieved with the former in the determination of Cd and Pb from water samples.³⁸ Trapped species were atomized either by rapidly increasing the fuel flow rate or by aspirating an organic solvent to the flame.^{38,39}

Initial studies of the atom trapping approach for the determination of hydride forming elements by atomic spectrometry involved *in situ* trapping in graphite furnaces.⁴⁰ Successful studies interfacing hydride generation to silica or quartz tube atom traps have been reported only recently.^{41–50} Korkmaz *et al.*⁴¹ used an electrically heated silica trap

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attached to the T-arm of a silica tube atomizer to collect PbH₄. Trapped species were released and swept into the flame-heated silica atomizer by raising the temperature of the trap rapidly; a detection limit of 19 ng L^{-1} was achieved for 60-s trapping (6 mL sample). Similarly, stibine (SbH₃) was trapped in an electrically-heated quartz tube trap;^{42,43} detection limits achieved for chemical⁴² and electrochemical⁴³ generation of stibine were 3.9 ng L^{-1} and 53 ng L^{-1} , respectively. Introduction of oxygen upstream to an externally heated quartz atomizer during trapping was found to increase the trapping efficiency of stibine⁴⁴ and bismuthine,⁴⁵ but not those of arsine, selenium hydride and tellurium hydride. Using the same approach, arsine and selenium hydride were trapped through the inlet arm of the quartz tube, which was modified to accommodate oxygen delivery to combust hydrogen during trapping.46 Very recently, Matusiewicz and Krawczyk collected hydrides of As and Se by using an integrated atom trap (IAT) assembly consisting of a water-cooled single silica tube and a double-slotted quartz tube.⁴⁷ Shortly after this report, the group extended the application of IAT approach to the determination of Cd and Pb,⁴⁸ tellurium (Te),⁴⁹ and indium (In) and thallium $(Tl)^{50}$ by hydride generation FAAS.

In this paper we described a conceptually similar approach of trapping lead hydride using a quartz tube atomizer for the determination of Pb by FAAS. Lead hydride was generated on-line by reaction of Pb in 0.3% v/v HCl–3% m/v K₃Fe(CN)₆ medium with 0.2% m/v NaBH₄. The gaseous hydride was trapped under fuel-lean flame conditions on the interior walls of a slotted T-tube placed on an air–acetylene flame. Atomization was performed by introducing 50 μ L of methyl isobutyl ketone (MIBK) through the nebulizer. The detection limit for a sample volume of 7.8 mL was 28 ng L⁻¹. The accuracy of the method for determination of Pb from environmental samples was demonstrated by analysis of soil and plant certified reference materials.

Experimental

Apparatus

All measurements were performed by use of a PerkinElmer Model 1100B atomic absorption spectrometer equipped with a 10-cm air-acetylene burner. A PerkinElmer Pb hollow cathode lamp (0.7 mA, 283.3 nm line) was used. The spectral bandpass was 0.7 nm. Deuterium lamp background correction was used throughout. Data were recorded using an Epson Model LQ-850 printer. The schematic diagrams of the flow injection manifold and the slotted quartz T-tube are shown in Figs. 1 and 2, respectively. Two peristaltic pumps (IsmaTec sa MS-REGLO) were used with Tygon pump tubes (1.5 mm id) delivering 5.2 mL min⁻¹. Connection tubings on the sample and reagent lines were made of 0.8 mm id PTFE. The mixing coil and the transfer line between the gas-liquid separator and the quartz T-tube were, respectively, 10- and 15-cm long PTFE (2 mm id). The tubing connecting argon supply to the gasliquid separator was also 2 mm id PTFE. The slotted quartz T-tube was 16 cm long (1.4 cm id). It had two slots: 11 cm long (lower) and 5 cm long (upper) at 180° with respect to each



Fig. 1 Schematic diagram of the flow injection hydride generation manifold.

other. The tube was held on the burner head by using the mount for the flame heated quartz tube.

Reagents and solutions

Deionized water was used in the preparation of all solutions. A $10 \ \mu g \ m L^{-1}$ Pb standard solution was prepared from a $1000 \ \mu g \ m L^{-1}$ Pb standard solution (Aldrich). All standard solutions were prepared by one-stage dilution of the $10 \ \mu g \ m L^{-1}$ Pb solution. Acidity was adjusted with HCl (Fisher Scientific). Solid K₃Fe(CN)₆ (EM Science) was used as the oxidant. Sodium borohydride (NaBH₄) solution was prepared daily by dissolving the necessary amount of NaBH₄ (Alfa) in 0.05% m/v sodium hydroxide (Fisher Scientific). Methyl isobutyl ketone (MIBK, Fisher Scientific) was used for atomization of the trapped species.

Procedure

A standard or sample solution prepared in 3% m/v $K_3Fe(CN)_6$ and 0.3% v/v HCl was continuously pumped at 5.2 mL min⁻¹ and mixed on-line with 0.2% m/v NaBH₄ solution also running at 5.2 mL min⁻¹ (Fig. 1). The volatile lead hydride generated along the mixing coil was separated within the gas–liquid separator and swept into the slotted T-tube. During the trapping of PbH₄ for a certain period of time, acetylene and air flow rates were set to 0.9 L min⁻¹ and 8.6 L min⁻¹, respectively, to produce an extremely oxidizing flame. Atomization of the trapped Pb species was achieved by aspirating 50 µL of methyl isobutyl ketone (MIBK) through the nebulizer. The solvent was delivered either by a micropipette or *via* a six-port flow injection valve. The rapid change in



Fig. 2 Schematic diagram of the slotted T-tube.

the flame characteristics from oxidizing (fuel-lean) to reducing (fuel-rich) flame resulted in a sharp transient signal

Method development

The experimental variables for the generation of PbH₄, trapping and atomization were optimized by a univariate method to obtain the greatest peak height absorbance. Five replicate measurements were made with either 1.0 or 2.0 μ g L⁻¹ Pb standard solution. The acidity of the solution containing 4% m/v K₃Fe(CN)₆ was varied from 0.1 to 0.8% v/v HCl while other parameters were kept constant. The concentration of K₃Fe(CN)₆ was increased from 0.5 to 4% m/v to affect the hydride generation efficiency for a series of standard solutions acidified to 0.3 v/v HCl. The length of the mixing coil was varied from 10 to 30 cm. For the transfer line, a 15 cm long (2 mm id) tube was used to minimize the distance between the gas-liquid separator and the slotted T-tube. The effect of NaBH₄ concentration was investigated between 0.05 and 2.0% m/v for a 1.0 μ g L⁻¹ Pb standard solution containing 0.3% v/v HCl and 3% m/v K₃Fe(CN)₆.

Argon gas was used to strip PbH₄ from the gas–liquid separator and pass it to the slotted quartz T-tube. The flow rate of argon was varied from 80 to 600 mL min⁻¹. Different flame conditions were studied to achieve maximum trapping efficiency. The flow rate of fuel (acetylene) was adjusted to a minimum value (0.9 L min⁻¹) to keep the flame lit and the flow rate of the oxidant (air) was varied from 8.0 to 9.2 L min⁻¹. For the atomization of the trapped species, the volume of organic solvent (MIBK) aspirated to the flame was varied from 15 to 100 µL.

Method validation

Two NIST standard reference materials were used for method validation: SRM 2709 (San Joaquin Soil) and SRM 1515 (Apple Leaves). Approximately 0.1 g for the soil and 0.3 g for the apple leaves were weighed into PTFE beakers and digested in a mixture of 3.0 mL of concentrated HNO₃ (Fisher Scientific) and 1 mL of concentrated HF (EM Science) on a hot plate.¹⁹ After complete dissolution, the contents were heated to drvness to evaporate the acids. The residue was then dissolved and diluted to 25 mL with 0.3% v/v HCl. For the determination of Pb by HG-FAAS, subsequent dilutions were performed with 0.3% v/v HCl solutions so that the concentration of Pb in the analysis solutions was between 0.4 and 0.8 μ g L⁻¹. Solid K₃Fe(CN)₆ (3% m/v) was added to each sample solution and the solutions were analyzed for Pb with the optimized system. External calibration was used for quantification with standard solutions ranging from 0 to 2.0 μ g L⁻¹ in 0.3% v/v HCl. Standard solutions also contained 3% m/v K₃Fe(CN)₆.

Results and discussion

Optimum conditions for generation of PbH₄

The improvement in the generation of lead hydride in an acid-oxidant medium is attributed to the efficient oxidation of Pb^{2+} to metastable Pb^{4+} , from which PbH_4 is generated.^{20-31,51-53} Further, depending on the oxidant, the type

of acid used to acidify samples is critical to achieve desired sensitivity and precision.²² Both nitric and lactic acids improved efficiency in the presence of $(NH_4)_2S_2O_8$, $K_2Cr_2O_7$ or H_2O_2 as oxidants, but the best sensitivity was achieved with lactic acid– $K_2Cr_2O_7$ medium.²⁶ Oxalic acid was used with $Ce(NO_3)_4$,⁵¹ while tartaric acid was preferred in the presence of $(NH_4)_2S_2O_8$.⁵² Similarly, tartaric acid/Na₂S₂O₈ was used, although HNO₃–Na₂S₂O₈ did afford equal sensitivity.⁵³ Precision was poor when HCl was used with Na₂S₂O₈.⁵³ Conversely, HCl was the most suitable acid with $K_3Fe(CN)_6$, a relatively mild oxidant.^{28–31}

In this study, HCl-K₃Fe(CN)₆ acid-oxidant medium was used as it has been frequently reported to provide by far the highest sensitivity in the generation of lead hydride. The effect of HCl concentration on the peak height absorbance is illustrated in Fig. 3. The maximum signal was obtained within a narrow region between 0.25 and 0.4% v/v HCl, for which 0.3% v/v was chosen as the optimum value. The absorbance from 2 μ g L⁻¹ Pb in 0.3 v/v HCl increased rapidly with an increase in the concentration of K₃Fe(CN)₆ from 0.5 to 1.0 m/v that peaked at 1.5% m/v K₃Fe(CN)₆ (Fig. 4a). A gradual increase in the signal with higher concentrations of K_3 Fe(CN)₆ was apparently from a Pb impurity in the $K_3Fe(CN)_6$ solution, as displayed by the signal profile of the blank (Fig. 4b). However, the precision among the daily absorbances from freshly prepared Pb solutions (2 μ g L⁻¹) was poor with 1.5% m/v K₃Fe(CN)₆, which necessitated the use of 3% m/v K_3 Fe(CN)₆ to maintain the stability in the daily signals. For NaBH₄, a relatively low concentration (0.2% m/v) was adequate to successfully generate PbH₄ (Fig. 5). No significant increase in the signals occurred for higher concentrations of NaBH₄.

In the last stage of the hydride generation, the length of mixing coil and the flow rates of the solutions were optimized with 1 μ g L⁻¹ Pb solution. Increasing the length of the mixing coil from 10 to 30 cm had almost no effect on the absorbance signals, suggesting that PbH₄ was successfully generated along a 10-cm long tubing. The flow rates for sample and NaBH₄ solutions were 5.2 mL min⁻¹ during the optimization studies







Fig. 4 Effect of K_3 Fe(CN)₆ concentration on (a) peak height absorbance, (b) blank + background absorption, for a 2 µg L⁻¹ Pb solution in 0.3% v/v HCl.

described above. Final examination indicated that these were the optimum settings in terms of sensitivity *versus* sample consumption. Increasing the flow rates improved the sensitivity to some extent, but not proportionately to the volume of sample used: most of the sample was wasted without a significant gain in sensitivity.

Trapping and atomization conditions

In continuous mode hydride generation, the atom number density and the residence time of the atoms in the light path are mainly determined by the speed with which the particular hydride is transported to the atomizer cell, which consequently influences the peak shape and peak height sensitivity. In atom trapping hydride generation, however, the peak height sensitivity is dependent on both trapping and atomization efficiency. It was proposed that analytes accumulate either as metals or oxides on the surface of silica or quartz traps,^{38,39,54} suggesting that direct contact of volatile hydride with the surface of the atom trap cell is essential for a certain period



Fig. 5 Effect of NaBH₄ concentration on peak height absorbance. Lead solution (1 μ g L⁻¹) is in 0.3% v/v HCl-3% m/v K₃Fe(CN)₆.

of time to achieve efficient collection. While the argon flow rate determines the duration of the contact, the flame conditions (*e.g.*, temperature and composition) impact on the stability of trapped species. Thus, prior to examining the effect of argon flow rate, the flame composition of the acetylene–air flame was optimized on the trapping efficiency of plumbane. The optimum settings for the flame gases were 0.9 L min⁻¹ for acetylene (fuel) and 8.6 L min⁻¹ for air (oxidant), which ensured a highly fuel-lean (oxidizing) flame during trapping. The acetylene flow rate was the lowest value to maintain the flame burning. Further increase in the air flow rate resulted in a noisy and unstable flame that went out shortly after it was lit. Stable flame conditions were also obtained for 1.0 L min⁻¹ acetylene and 9.2 L min⁻¹ air flow rates, but these conditions did not provide any significant improvement in trapping efficiency.

The optimum flow rate of argon carrier gas ranged between 200 and 320 mL min⁻¹: this ensured efficient trapping of Pb species. The flow rate was adjusted to 250 mL min⁻¹ thereafter. At lower argon flow rates (*e.g.*, 100–120 mL min⁻¹), signals were significantly low, which was most likely due to the loss of PbH₄ before reaching the slotted T-tube trap. The signals gradually decreased with increasing argon flow rate above 320 mL min⁻¹ since the volatile plumbane was swept rapidly without sufficient physical contact with the slotted T-tube.

Atomization was performed by nebulizing methyl isobutyl ketone (MIBK) to the fuel-lean flame. A volume of 50 μ L of MIBK was sufficient to produce a hot, fuel-rich flame, allowing rapid release and atomization of the trapped species from the walls of the slotted tube. Greater volumes up to 100 μ L did not show any enhancement in the signals, but rather increased the background absorption up to 0.07 for 30-s trapping. Alternatively, atomization was performed by manually switching the air and acetylene flow rates to the stoichiometric flame settings. Though this procedure eliminated the background absorption significantly, it was not practical because of the speed and operator skill required to maintain the precision between replicate atomizations.

Analytical figures of merit

Calibration was performed with a number of Pb standard solutions ranging from 0 to 2.0 μ g L⁻¹ Pb. The relative standard deviation (RSD) ranged between 2 and 4% for 30-s trapping. The detection limits and characteristic concentrations were measured for a period of 30, 60 and 90 s trapping using a sample volume of 2.6, 5.2 and 7.8 mL, respectively. The results are given in Table 1. A comparison of analytical performances is also provided in Table 2. The HG-AT procedure afforded significant improvements in the detection limits with respect to those reported for ETAAS and HG-FAAS.

Table 1 Effect of trapping time on detection limit (3s, n = 13) and characteristic concentration

Trapping period/s	Sample volume/mL	$\begin{array}{c} Characteristic \\ concentration/\mu g \ L^{-1} \end{array}$	Detection limit/µg L^{-1}
30	2.6	0.049	0.075
60	5.2	0.025	0.047
90	7.8	0.019	0.028

Table 2Comparison of detection limits for the determination of Pbby different atomic spectrometry techniques. The values are based onthe peak area absorbance for ETAAS and peak height absorbance forFAAS

Technique	Sample volume/mL	Detection limit/µg L^{-1}	Reference
ETAAS	0.02	0.4	PE 4100ZL catalogue
HG-AT-ETAAS	1.0	0.03	29
HG-FAAS	0.5	0.7	30
HG-AT-FAAS	6.0	0.019	41
HG-IAT-FAAS	2.0	0.40	48
HG-AT-FAAS	7.8	0.028	This study

Detection limits were similar to those reported for HG-AT-ETAAS²⁹ and HG-AT-FAAS⁴¹ and were significantly lower than that for HG-IAT-FAAS.⁴⁸

The relationship between detection limit and sample volume (Table 1) was consistent with the equation " $C_{\rm L} = b/V + a$ " proposed by Tyson⁵⁵ for flow-based systems, where " C_L " is the detection limit, "V" is the sample volume, and "a" and "b" are constants. The data fit a detection limit equation of $C_{\rm L} = 0.174 \ (\pm 0.023)/V + 0.009 \ (\pm 0.004)$ with a correlation coefficient (r^2) of 0.985. The values in the parenthesis are uncertainties for coefficients "b" and "a", respectively. A plot of the detection limit versus sample volume is therefore consistent with a rectangular hyperbola (Fig. 6), suggesting that the improvement in the detection limit would be slight for sample volumes greater than 6 mL due to the impurities in the reagents; the lowest detection limit would be $C_{\rm L} = 0.009 \ \mu g$ L^{-1} as $V = \infty$. In this study, the major source of blank contamination was from the impurities in $K_3Fe(CN)_6$. Blank absorbance was about 0.07 for 30-s trapping (2.6 mL sample); therefore, it was concluded that better detection limits could be achieved by using high purity potassium ferricyanide.

Application to analysis of samples for Pb

The accuracy of procedure was validated by analysis of two NIST standard reference materials, SRM 2709 (San Joaquin Soil) and SRM 1515 (Apple Leaves). The results for Pb are summarized in Table 3. No significant differences were found at the 95% confidence level (p = 0.05) between experimental and certificate values since the confidence interval of the experimental values included the mean of the certificate values. A number of transition elements, including Co, Cu, Mn, Ni and V, were reported to increase the lead signal by about 20–60% when plumbane was generated in H₂O₂–HNO₃ medium.³ The interference from Fe was even more complicated: the Pb signal increased in H₂O₂–HNO₃ medium at a Pb:Fe ratio of 1:0.016, but was suppressed at a Pb:Fe ratio of 1:1

Table 3 Results obtained from certified reference materials by HG-AT-FAAS. Results are given as mean $\pm 95\%$ confidence interval of 4 replicates. Certified values are given as mean $\pm 95\%$ confidence interval

	San Joaquin Soil SRM 2709	Apple Leaves SRM 1515
Found/ μ g g ⁻¹	18.6 ± 1.3	0.49 ± 0.04
Certified/ $\mu g g^{-1}$	18.9 ± 0.5	0.47 ± 0.024
Accuracy (%)	98.2 ± 6.1	105.2 ± 4.9



Fig. 6 The effect of sample volume on the detection limit for Pb by HG-AT-FAAS. Blank solution (n = 13) contained 0.3% v/v HCl and 3% m/v K₃Fe(CN)₆.

when $S_2O_8^{2-}$ was the oxidant.³ In this study, the soil SRM contained considerably higher levels of these interfering elements: Cu (34.6 µg g⁻¹), Fe (3.50%), Mn (538 µg g⁻¹), Ni (88 µg g⁻¹), V (112 µg g⁻¹). The lowest value was for Cu for which Pb-to-Cu ratio was about 1:2 in the analysis solutions; nevertheless, the procedure was highly robust against the concomitant ions in the soil matrix. No significant enhancement or suppression was observed in Pb signals. The acidity of the samples was, however, a critical point that deserved careful adjustment to achieve high precision and accuracy. In the preparation of the samples by HNO₃–HF digestion, the contents were heated to dryness after dissolution to completely remove excess acids and then diluted to the appropriate volume with 0.3% v/v HCl.

Conclusion

The hydride generation atom trapping approach has been proven to be highly suitable to achieve better sensitivity for accurate determination of hydride forming elements at trace levels since the traps made up of quartz or silica tubes act, in a sense, like an enrichment device.41-50 In this study, we have demonstrated in situ trapping of lead hydride species on the inner walls of a flame-heated quartz tube under highly oxidizing flame conditions followed by the release of trapped species by introducing MIBK. The method is simple, cost-effective and yet offers the capability for the determination of lead at trace levels by FAAS. The acidity of the sample solution and the concentration of the oxidant, K₃Fe(CN)₆, require close control to maintain precision and accuracy. While the former is influential in the efficacy of the generation of plumbane, the latter mostly impacts on the detection limits as it increases the blank signals notably due to the impurities of Pb. Thus, it is expected that detection limits could be further improved with the use of high purity potassium ferricyanide.

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References

- 1 S. J. Hill, J. B. Dawson, W. J. Price, I. L. Shuttler, C. M. M. Smith and J. F. Tyson, J. Anal. At. Spectrom., 1998, 13, 131R.
- 2 A. Taylor, S. Branch, D. Halls, M. Patriarca and M. White, J. Anal. At. Spectrom., 2002, 17, 414.
- 3 M. R. Cave, O. Butler, S. R. N. Chenery, J. M. Cook, M. S. Cresser and D. L. Miles, *J. Anal. At. Spectrom.*, 2001, **16**, 194.
- 4 Z. Fang, *Flow Injection Atomic Absorption Spectrometry*, Wiley, New York, 1995.
- 5 J. F. Tyson, Anal. Chim. Acta, 1990, 234, 3.
- 6 Z. Fang, M. Sperling and B. Welz, J. Anal. At. Spectrom., 1991, 6, 301.
- 7 D. Rodriguez, P. Fernandez, C. Perez-Conde, A. Gutierrez and C. Camara, *Fresenius' J. Anal. Chem.*, 1994, **349**, 442.
- 8 R. L. Ma, V. Van Mol and F. Adams, *Anal. Chim. Acta*, 1994, 285, 33.
- 9 F. M. Fernandez, J. D. Stripeikis, M. B. Tudino and O. T. Troccoli, *Analyst*, 1997, **122**, 679.
- 10 E. Bulska, M. Walcerz, W. Jedral and A. Hulanicki, *Anal. Chim. Acta*, 1997, **357**, 133.
- 11 H. Lancester, G. D. Marshall, E. L. Gazola, J. Ruzicka and G. D. Christian, *Analyst*, 1994, **119**, 1459.
- 12 J. Ruzicka and A. Arndal, Anal. Chim. Acta, 1989, 216, 243.
- 13 Z. Fang, M. Sperling and B. Welz, J. Anal. At. Spectrom., 1990, 5, 639
- 14 M. Sperling, X. Yin and B. Welz, J. Anal. At. Spectrom., 1991, 6, 295.
- 15 R. Lima, K. C. Leandro and R. E. Santelli, Talanta, 1996, 43, 977.
- 16 R. E. Santelli, M. Gallego and M. Valcarcel, *Talanta*, 1994, 41, 817.
- 17 Y. A. Zhang, P. Riby, A. G. Cox, C. W. McLeod, A. R. Date and Y. Y. Cheung, *Analyst*, 1988, **113**, 125.
- 18 A. M. Naghmush, K. Pyrzynska and M. Trojanowicz, *Talanta*, 1995, 42, 851.
- 19 L. Elçi, Z. Arslan and J. F. Tyson, *Spectrochim. Acta, Part B*, 2000, 55, 1109.
- 20 J. Dedina and D. L. Tsalev, Hydride Generation Atomic Absorption Spectrometry, Wiley, New York, 1995.
- 21 X.-P. Yan and Z.-M. Ni, Anal. Chim. Acta, 1994, 291, 89.
- 22 C. Camara and Y. Madrid, Analyst, 1994, 119, 1647.
- 23 K. Jin and M. Taga, Anal. Chim. Acta, 1982, 143, 229.

- 24 H. M. F. Tavares, M. T. S. D. Vasconcelos, A. A. S. C. Machado and P. A. P. Silva, *Analyst*, 1993, **118**, 1433.
- 25 C. M. Mena, C. Cabrera, M. L. Lorenzo and M. C. Lopez, J. Agric. Food Chem., 1997, 45, 1812.
- 26 Y. Madrid, D. Chakraborti and C. Camara, *Microchim. Acta*, 1995, **120**, 63.
- 27 M. Chikuma and H Aoki, J. Anal. At. Spectrom., 1993, 8, 415.
- 28 R. Thao and H. Zhou, Fenxi Huaxue, 1985, 13, 283.
- 29 J. F. Tyson, R. I. Ellis, G. Carnrick and F. Fernandez, *Talanta*, 2000, **52**, 403.
- 30 A. Berkkan and N. Ertaş, Talanta, 2004, 64, 423.
- 31 W. Chuachuad and J. F. Tyson, J. Anal. At. Spectrom., 2005, 20, 282.
- 32 R. J. Watling, Anal. Chim. Acta, 1977, 94, 181.
- 33 C. Lau, A. Held and R. Stephens, *Can. J. Spectrosc.*, 1976, 21, 100.
 34 C. M. Lau, A. M. Ure and T. S. West, *Anal. Chim. Acta*, 1982, 141, 213
- 35 C. M. Lau, A. M. Ure and T. S. West, *Anal. Chim. Acta*, 1983, **146**, 171.
- 36 H. Matusiewicz, Spectrochim. Acta, Part B, 1997, 52, 1711.
- 37 G. Huang, S. Qian and H. Yang, Fenxi Huaxue, 1995, 41, 707.
- 38 N. Ertaş, D. K. Korkmaz, S. Kumser and O. Y. Ataman, J. Anal. At. Spectrom., 2002, 17, 1415.
- 39 D. Korkmaz, S. Kumser, N. Ertaş, M. Mahmuta and O. Y. Ataman, J. Anal. At. Spectrom., 2002, 17, 1610.
- 40 H. Matusiewicz and R. E. Sturgeon, Spectrochim. Acta, Part B, 1996, 51, 377.
- 41 D. K. Korkmaz, N. Ertaş and O. Y. Ataman, Spectrochim. Acta, Part B, 2002, 57, 571.
- 42 D. Korkmaz, J. Dědina and O. Y. Ataman, J. Anal. At. Spectrom., 2004, 19, 255.
- 43 İ. Menemenlioglu, D. Korkmaz and O. Y. Ataman, Spectrochim. Acta, Part B, 2007, 62, 40.
- 44 J. Kratzer and J. Dedina, Spectrochim. Acta, Part B, 2005, 60, 859.
- 45 J. Kratzer and J. Dedina, J. Anal. At. Spectrom., 2006, 21, 208.
- 46 J. Kratzer and J. Dedina, Anal. Bioanal. Chem., 2007, 388, 793.
- 47 H. Matusiewicz and M. Krawczyk, Anal. Sci., 2006, 22, 249.
- 48 H. Matusiewicz and M. Krawczyk, Microchem. J., 2006, 83, 17.
- 49 H. Matusiewicz and M. Krawczyk, Spectrochim. Acta, Part B, 2007, 62, 309.
- 50 H. Matusiewicz and M. Krawczyk, J. Braz. Chem. Soc., 2007, 18, 304.
- 51 J. Li, F. Lu, T. Umemura and K.-I. Tsunoda, *Anal. Chim. Acta*, 2000, **419**, 65.
- 52 J. F. N. Carrijo, L. C. Brasil and N. M. M. Coelho, J. Braz. Chem. Soc., 2005, 16, 520.
- 53 N. Maleki, A. Safavi and Z. Ramezani, J. Anal. At. Spectrom., 1999, 14, 1227.
- 54 T. S. West, Anal. Proc., 1988, 25, 240.
- 55 J. F. Tyson, J. Anal. At. Spectrom., 1999, 14, 169.