

METHODOLOGY ARTICLE

Open Access



Development and evaluation of a broadly reactive reverse transcription recombinase polymerase amplification assay for rapid detection of murine norovirus

Lei Ma^{1†}, Fanwen Zeng^{2†}, Feng Cong¹, Bihong Huang¹, Yujun Zhu¹, Miaoli Wu¹, Fengjiao Xu¹, Wen Yuan¹, Ren Huang¹ and Pengju Guo^{1*}

Abstract

Background: Murine norovirus (MNV) is recognized as the most prevalent viral pathogen in captive mouse colonies. The rapid detection assay for MNV would be a useful tool for monitoring and preventing MNV infection. A recombinase polymerase amplification (RPA) assay was established in this study to provide a solution for rapid and sensitive detection of MNV.

Results: The detection limit of the RT-RPA assay for the detection of MNV was 1×10^2 copies of RNA molecules per reaction. The assay was specific since there was no cross-reaction with other common murine viruses. In addition, the broad reactivity of the RT-RPA assay was validated using the synthesized template carrying seven point mutations among several MNV strains. The MNV RT-RPA assay could detect as few as 1×10^2 copies of the mutant per reaction, suggesting the assay could be broadly reactive against a large diversity of MNV strains. Forty eight clinical samples including 16 gastric tissue specimens, 16 cecal tissue specimens and 16 fecal specimens were tested for the validation of the new developed RT-RPA assay. The detection results of RT-RPA and RT-qPCR for clinical samples were very similar, except that a gastric tissue sample which was positive by RT-qPCR, with a RNA titer of 27 copies, was negative by RT-RPA.

Conclusions: A broadly reactive RT-RPA assay was successfully established for MNV detection.

Keywords: Murine norovirus, Recombinase polymerase amplification, Genetic diversity, Detection

Background

Norovirus is a class of viral pathogens that could infect a variety of hosts [1]. Norovirus have been tentatively classified as five genogroups based on the sequences of major capsid protein (VP1) [1]. In a previous study, standard norovirus classification tree made with the alignment of 68 NoV sequences was proposed for nomenclature: 29 genetic clusters are classified in the 5 genogroups, 8 in GI, 17 in GII, 2

in GIII, and 1 each in GIV and GV [1]. GI, GII, and GIV strains, are found in humans, and GIII and GV strains are found in cows and mice, respectively. MNV, belongs to genogroup GIII, is one of the most common pathogens in laboratory mice [2, 3].

Since the prototype strain MNV-1 was firstly detected in immunocompromised mice [3], multiple strains of MNV have been discovered and/or isolated from the laboratory and wild rodents [4–6]. Although different MNV strains exhibited the considerable genetic and biological diversity, they belonged to a single serotype determined by cross-neutralization test [7]. Serological investigations in North America, Europe and Japan demonstrated the high prevalence of specific antibodies against MNV in laboratory rodents [8–10]. Currently,

* Correspondence: vetbio2016@hotmail.com

[†]Lei Ma and Fanwen Zeng contributed equally to this work.

¹Guangdong Key Laboratory of Laboratory Animals, Guangdong Laboratory Animals Monitoring Institute, Guangzhou, China

Full list of author information is available at the end of the article



MNV has become the most prevalent pathogen in laboratory mice worldwide.

The mice are the most commonly used laboratory animals in biomedical research. The high prevalence of MNV in laboratory mice may interfere in the result of biomedical research. Several experiments have been performed to investigate the effects of MNV infection on scientific research. For examples, experimental infection of the mice revealed that MNV-1 lead to the persistent infection and long fecal virus shedding [2]. MNV-4 infection induced multiple inflammatory hallmarks of human Crohn's disease in Atg16L1^{H^M} mice after dextran sodium sulfate administration [11]. Whereas, another study reported that transient or persistent norovirus infection didn't alter the pathology of intestinal inflammation and fibrosis induced by *Salmonella Typhimurium* infection in C57BL/6 mice [12]. As no clear conclusion can be made from the previous studies regarding to the effects of MNV infection on biomedical research in mice, the use of MNV-free mice was recommended for fundamental research and drug safety evaluation. Thus, the rapid and sensitive detection methods for MNV infection played a critical role in the quality control of the laboratory mice.

To monitor and detect MNV rapidly and specifically, several molecular methods such as reverse transcription polymerase chain reaction (RT-PCR) assays and RT-quantitative PCR (RT-qPCR) assays have been developed [8, 13–15]. Considering that the RT-PCR and RT-qPCR methods are time-consuming, the loop-mediated isothermal amplification (LAMP) method for detection of MNV was developed [16]. However four to six primers were required in the LAMP assay [16]. Recombinase polymerase amplification (RPA), another isothermal amplification method, which requires only two primers, has been recently used for pathogen detection and only takes less than 30 min to perform the amplification reaction [17–21]. RPA primers bind to DNA template by bacterial recombinase enzyme, then the extension and amplification reaction is initiated by an isothermal polymerase [22]. These enzymes are in concert with a reverse transcriptase and a fluorescent probe for real-time detection of the viruses containing RNA genomes [23].

Previous study has reported the RT-RPA assay for detection of human norovirus [24]. However, the whole genome sequence of human norovirus shared poor identity with MNV [2]. The RT-RPA assay used for human norovirus cannot be applied to the detection of MNV. Thus the aim of the present study is to establish a broadly reactive RT-RPA assay for rapid and sensitive detection of MNV which is widely present in laboratory mice across the world.

Results

Screening of the primer sets

Six primer sets were screened by the basic RT-RPA assay using the 10⁴ copies RNA standard as the template. As clearly showed in Fig. 2a, the product of the primer set f5/r5 emitted the strongest light under ultraviolet light. The probe was designed based on the target sequence of the primer set f5/r5. All the sequences of primers and probe used in this study were displayed in Table 1.

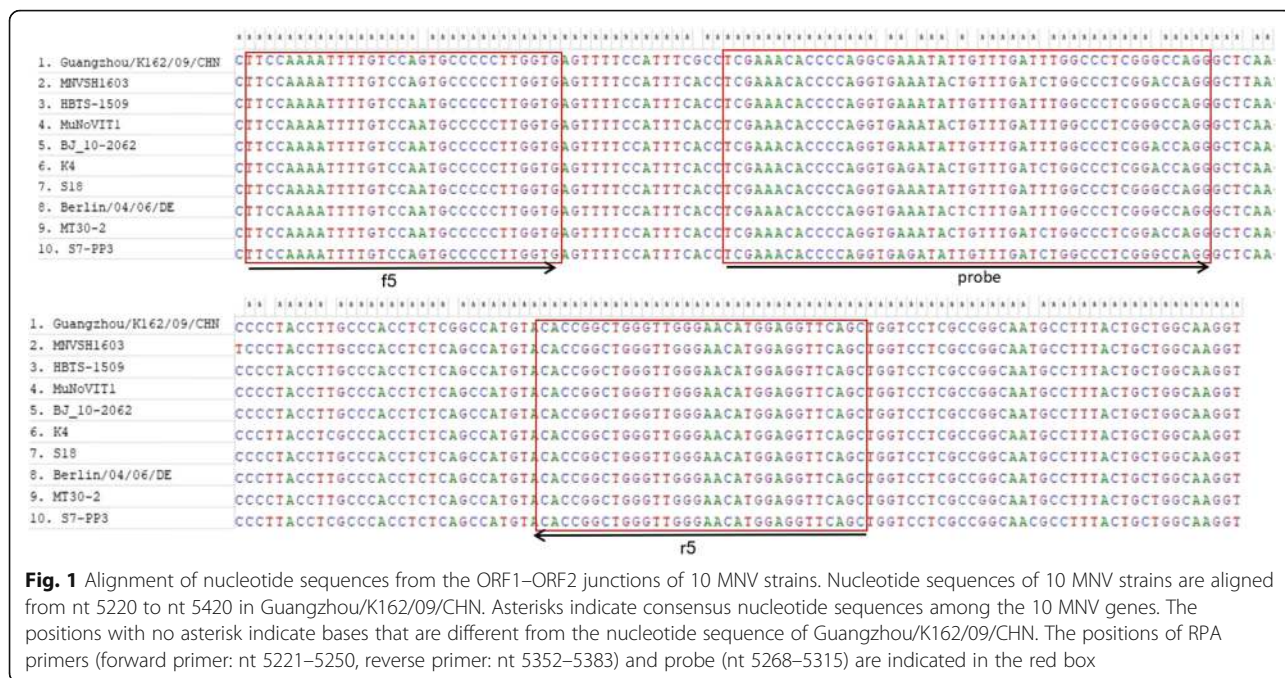
Sensitivity and specificity of the real-time RT-RPA

The detection limit of the RT-RPA assay was assessed using the 10-fold serially diluted RNA standards. As illustrated in Fig. 1b, the dynamic detection range of the assay ranged from 10⁶ to 10² copies per reaction, demonstrating that the limit of detection of the assay was 10² copies/μL molecular RNA. The RT-RPA assay was performed eight times on the serially diluted RNA standards, in which 8 out of 8 (8/8) runs were positive when viral load are 10⁶–10² copies per reaction; 5/8 runs were positive when 10 copies per reaction; and 0/8 positive when 1 copy per reaction was used. To evaluate the reproducibility of the RT-RPA assay, semi-logarithmic regression analysis was performed using the data from the 8 runs (Fig. 3a). A probit regression analysis using the results of 8 runs was performed to determine the exact detection limit, and the results showed that RT-RPA could detect 70 RNA molecules in 95% of cases (Fig. 3b). The synthesized templates carrying seven point mutations were

Table 1 Primers and probe

Name	Sequence	Amplicon length
MNV-F	ATGCATGGTGAAAAGTACTAT	788 bp
MNV-R	TAGAAAGAAGGCGACCAGAGA	
f1	CAGTCTTTGTGAATGAGGATGAGTGATG	189 bp
r1	AAAATTTTGGAGATCCAGGGGTCAATTT	
f2	CAAATCAACCAAATTGACCCCTGGATCTT	151 bp
r2	ACCTCCATGTCCCAACCCAGCCGGTGATACAT	
f3	CAAAATTTTGTCCAGTGCCCCCTTGGTGAGTT	135 bp
r3	AGGACCAGCTGAACCTCCATGTTCCCAACCCA	
f4	CACGCCACCGGTCTGTTCTGCGCTGGGTGC	154 bp
r4	AAGGAACAAGATCCTGGCCGCTGGCTTC	
f5	TTCCAAAATTTTGTCCAGTGCCCCCTTGGT	131 bp
r5	GCTGAACCTCCATGTTCCCAACCCAGCCGGTG	
f6	TTCAGCTGGTCTCGCCGCAATGCCTTTA	141 bp
r6	CCAGGGTGCGCACATCACACATGACATGTG	
probe	TCGAAACACCCAGGGCAAAATATTGTTGA (BHQ)(THF)(FAM)GGCCCTCGGGCCAGG	

BHQ1-dT dT-fluorophore, THF tetrahydrofuran, FAM-dT dT-quencher group



also tested by the RT-RPA assay to evaluate the broadly reactive capacity of the assay. The result showed that the RT-RPA assay can detect the mutant template as few as 10² copies/μL molecular RNA (data not shown), indicating that the point mutations had no effect on the sensitivity of RT-RPA assay.

RNA genomes from other common murine viral pathogens were tested by the RT-RPA assay. Figure 2c revealed that the amplification curve was only observed for MNV; no amplification signal was detected for TMEV, SeV, MHV, PVM, Reo-3 and distilled water.

Assay performance on clinical samples

The performance of RT-RPA was compared with probe-based RT-qPCR using RNA isolated from 48 murine samples. Eight of 16 cecal samples and 9 of 16 fecal samples were positive by both assays. In contrast, 6 of 16 gastric tissue samples were by RT-RPA while 7 of 16 gastric tissue samples were positive by RT-qPCR. The viral load in the gastric tissue sample which was negative by RT-RPA, was 27 copies per reaction determined by RT-qPCR, which was lower than the detection limit of the RT-RPA assay. Linear regression analysis demonstrated a poor correlation between C_T values determined by RT-qPCR and T_T values determined by RT-RPA (Fig. 3c), suggesting the RT-RPA assay could not be applied to quantitative analysis of MNV viral loads in the samples. This phenomenon was also reported in the RT-RPA assay for detection of avian influenza virus in clinical samples [25].

Discussion

In present study, a real-time RT-RPA assay was established for rapid and specific detection of MNV. The primer-probe set was designed based on the sequences of the highly conserved ORF1-ORF2 junction. This new assay could detect a broad range of genetically diverse MNV isolates with the same detection sensitivity.

MNV is considered the most prevalent viral pathogen in laboratory mice worldwide. MNV infection may affect the results of biomedical researches [16], so it is of vital importance to confirm the MNV-free status of the laboratory mice. Several real-time RT-PCR assays have been developed for detecting MNV. However, these techniques could not detect all the MNV strains due to the genetic diversity of MNV [5, 15, 26]. Thus two broadly reactive real-time RT-PCR assays were established to detect MNV infection [27, 28]. Although the real-time RT-PCR assay is sensitive and specific, it is expensive, time-consuming and labor-intensive. To overcome the shortcomings of the real-time RT-PCR, a broadly reactive LAMP assay was developed for detection of MNV. However, the LAMP assay required five primers, which increased the difficulty of primer design for MNV with genetic diversity. A portable and user-friendly method is therefore necessary for rapid detection of MNV on site. One promising solution is the RPA assay. Due to its convenience and portability, the RPA technique has been used for rapid detection of various pathogens, such as foot-and-mouth disease virus [29], Zika virus [30], human norovirus [24] and

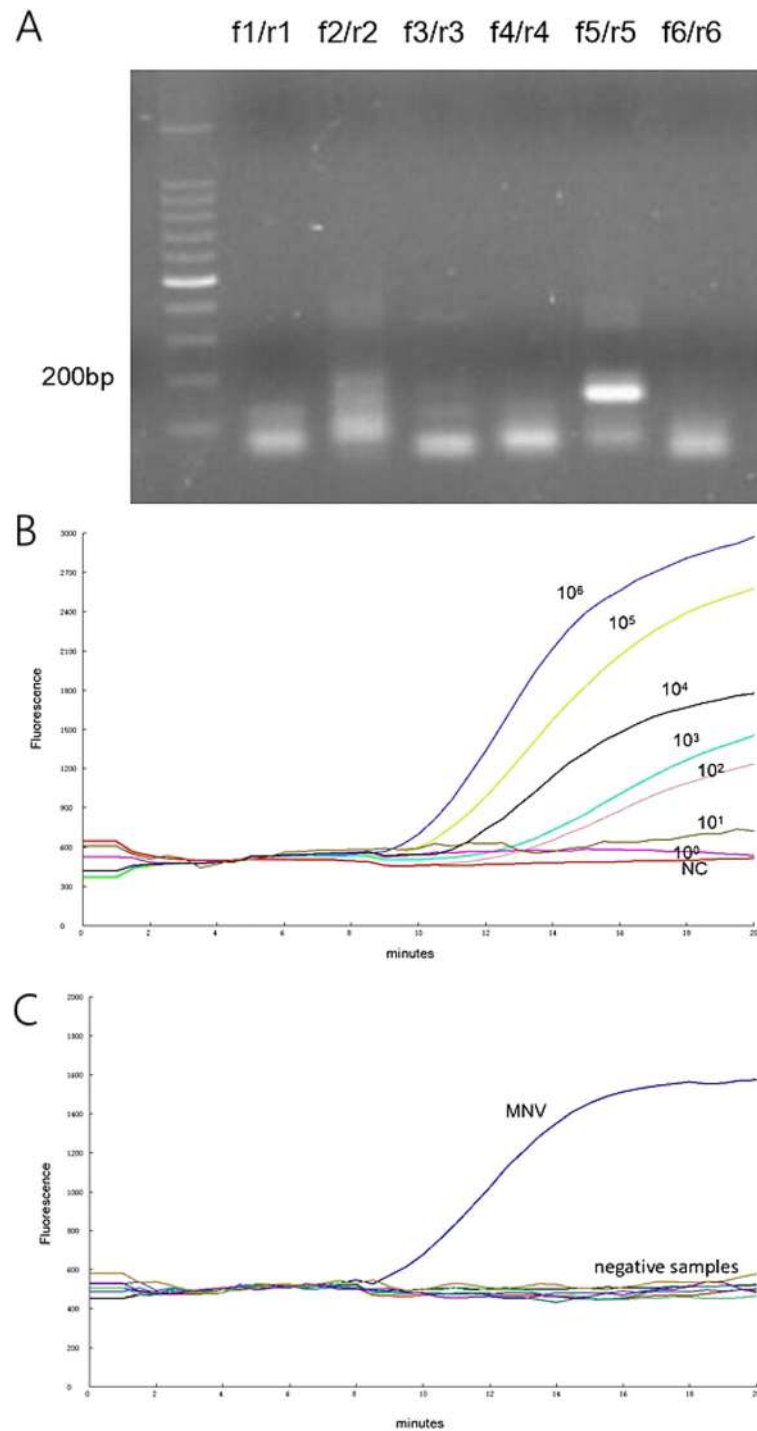


Fig. 2 Primers sets screening; sensitivity and specificity of the RT-RPA assay. **a** The products of RT-RPA using six primers pairs were separated on agarose gel electrophoresis. **b** Fluorescence signal over time using the RNA standards ranging from 10⁶ to 10⁰ copies. **c** Specificity of the RT-RPA assay. TMEV, SeV, MHV, MPV, Reo-3 and distilled water were the negative samples

pathogenic bacteria [31]. Up to date, there is only one report about the RPA assay for the laboratory animal health monitoring [32].

Design of the primer set plays a vital role in the performance of the RPA assay. Up to now, no software is available for RPA primer design. The primers used in

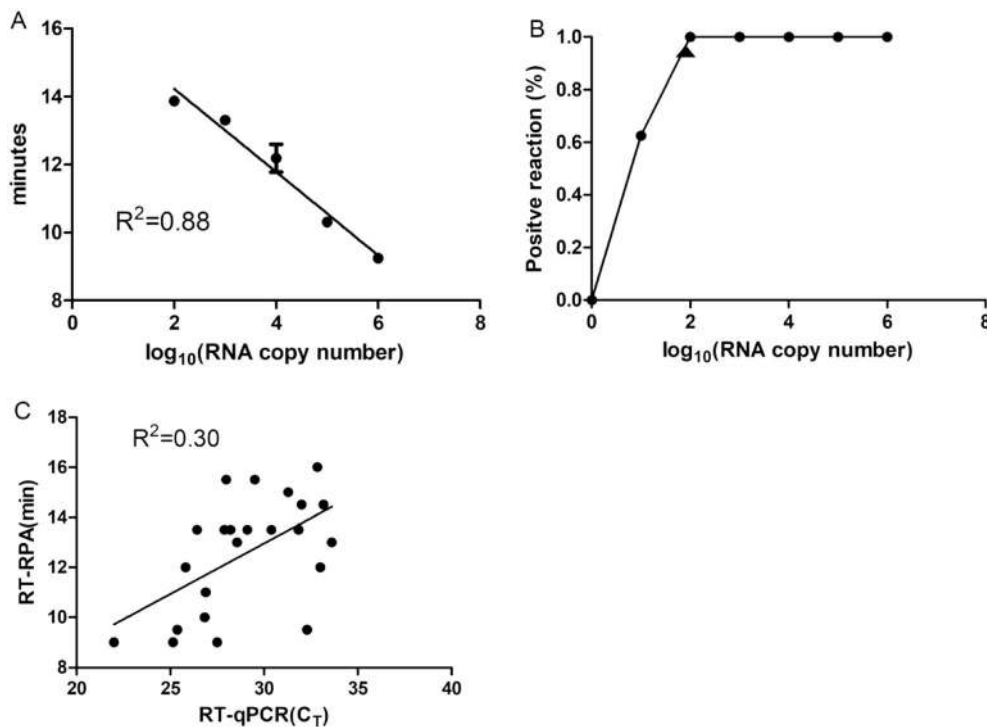


Fig. 3 Performance of the RT-RPA assay. **a** Semi-logarithmic regression of the data collected from 8 runs using the RNA standard analyzed by GraphPad Prism 5.0. **b** Probit regression analysis using the data of the 8 runs. The detection limit at 95% probability (95 molecules) is depicted by a triangle. **c** Linear regression analysis of RT-RPA threshold time (TT, y axis) and RT-qPCR cycle threshold (CT) values (x axis) were performed by Prism software. R² value was 0.3

this study were manually designed following the manufacturer's instruction. In this study, six primer sets were tested by the RPA assay, only one primer set (f5/r5) produced strong gene amplification signal. The probe was designed based on the target region of the primers set (f5/r5). The primer-probe set produced satisfied result (Fig. 2b). This result demonstrated that it is efficient and cost-effective to screen the primer sets through basic RPA assay, followed by gel electrophoresis.

Although the target region of the RT-RPA assay is highly conserved within the whole MNV genome sequence, it still possesses several point mutations among different strains that may affect the diagnostic sensitivity of the RPA assay. The sequences of 10 represent MNV strains in different countries were aligned. There are six different nucleotides at the binding site of the forward primer and the probe between the MNV strain Guangzhou/K162/09/CHN and strain K4. In addition, one more point mutation was present at other strains. To determine the effect of all the mutations on the performance of the RT-RPA assay, the templates containing all the seven point mutations were synthesized and tested by the RT-RPA assay. The result demonstrated that the mutations have no effect on the performance of the RT-RPA assay, which was consistent with previous

studies proving that up to nine point mutations were tolerated [33, 34], suggesting that the RT-RPA assay could detect a diverse array of MNV strains.

The clinical performance of the RT-RPA assay was evaluated using 48 field samples. The MNV positive rate was 47.9% (23/48) determined by RT-RPA and 50.0% determined by RT-qPCR. The sensitivity of the RT-RPA technique for detection of MNV in clinical samples was 95.8% when compared with the RT-qPCR assay. Only one sample containing low RNA titer (27 RNA copies) was tested negative by RT-RPA. We believe that this was due to the low RNA copies (27 RNA copies) in the sample and low analytical sensitivity of the RT-RPA technique. However, the MNV RT-RPA assay was 10 times more sensitive than the RT-RPA for detection of human norovirus reported in a previous study [24]. It is worthy to note that the limit of detection of the human norovirus RT-RPA assay was performed on serially diluted RNA extract of stool. The sensitivity of present study was determined using a purified in vitro transcribed RNA rather than RNA extract of stool which may contain a lot of other RNA that interacts with recombinase proteins and thus can harm sensitivity. This may explained the difference of sensitivity between the two RPA assays. The RT-RPA assay could detect human

norovirus in directly boiled stool, and displayed better resistance to inhibitors than a commonly used RT-qPCR assay [24]. It is promising to detect MNV in directly boiled animal tissue or stool samples by the RT-RPA assay in the future. This procedure would be valuable to reduce sample processing time and increase portability.

Conclusion

A rapid, sensitive and specific method for detection of MNV was successfully established based on RPA assay. The RT-RPA had several advantages over RT-qPCR, including: (i.) a cost-effective portable device is sufficient; (ii) quicker time-to-result; (iii) the use of enzyme pellet in one tube reduces the laborious work and the likelihood of contaminant; (iv.) tolerance to common PCR/qPCR inhibitors [24]; (v.) the potential of point-of-care detection of the clinical samples. Thus, the assay provides an easy-to-use platform for rapid detection and monitoring of MNV infection which circulates in laboratory mice across the world.

Methods

Virus

MNV Guangzhou/K162/09/CHN strain was isolated and preserved in our laboratory [35], Theiler's mouse encephalomyelitis virus (TMEV) BeAn8386 strain, sendai virus (SeV) Sendai/52 strain, mouse hepatitis virus A59 strain (MHV), reovirus type 3 (Reo-3) Dearing strain, pneumonia virus of mice (PVM) number 15 strain were purchased from American Type Culture Collection (ATCC). Nucleic acid for lymphocytic choriomeningitis virus (LCMV) Armstrong strain were kindly provided by Dr. Zheng Ming He, from the Laboratory Animal Institute, National Institutes for Food and Drug Control, China.

Generation of MNV RNA standard

The ORF1-ORF2 junction sequence on MNV genome was demonstrated to be highly conserved [16]. A pair of primers (MNV-F: 5'-ATGCATGGTGAAAAGTACT AT-3', MNV-R: 5'-TAGAAAGAAGGCGACCAGAGA3') were designed and used to amplify the highly conserved ORF1-ORF2 junction. RNA of MNV strain Guangzhou/K162/09/CHN (Accession No: HQ317203) was used to amplify the target region by One Step RT-PCR kit (Takara, China). The reaction condition was set up as follows: reverse transcription (RT) at 50 °C for 30 min, denaturation at 95 °C for 5 min, 30 cycles of 94 °C for 30 s, 50 °C for 30 s and 72 °C for 45 s, a final extension step at 72 °C for 5 min. The amplicon was subjected to gel electrophoresis and recovered. The purified amplicon was cloned into the pGEM-T vector (Promega, USA) and designated as pGEM-MNV. The positive plasmid was confirmed by Sanger sequencing and in vitro transcribed using

RiboMAX™ Large Scale RNA Production Systems (Promega, USA) according to the manufacturer's instruction. The in vitro transcribed RNA was quantified by Nano2000 (GE, USA) and converted to copy number.

Primer and probe design

There is no software available for RPA primer and probe design up to now. The primer sets were manually designed following the manufacturer recommendation (TwistDx), based on the sequence of MNV RNA molecular standard. Basic RPA assay was performed to screen the primer set with the best amplification efficiency. Thereafter, the probe was designed based on the target sequence of the primer pair.

RT-RPA assay

Basic RT-RPA reaction was carried out in a 50 µl volume using the TwistAmp Basic RT kit (TwistDx, UK), which consisted of 30.1 µl of rehydration buffer, 1 µl of template, 2.1 µl of each primer (10 µM), 12.2 µl of water and 2.5 µl of magnesium acetate (280 mM). The microtubes were immediately placed in the heating block and incubated at 39 °C for 20 min. The RPA product was cleaned using a clean-up kit (Beyotime, China) to remove the inhibitors that might affect the agarose electrophoresis and separated on a 2% agarose gel.

Real time RT-RPA assay was performed using the TwistAmp exo RT kit. The reaction conditions for real-time RT-RPA were the same as for the basic RT-RPA, except that 0.6 µl of water was replaced by 0.6 µl of probe. The microtubes were placed in the Deaou-308C tubescanner (DEAOU Biotechnology, China) set to 39 °C to start the reaction. After incubating for 4 min, the microtubes were mixed, spun down, and placed back on the tubescanner for 16 min at 39 °C. A sample was considered positive when it generated an exponential amplification curve above the threshold of the negative control.

Sensitivity and specificity of the RT-RPA assay

The transcribed RNA standard was 10-fold serially diluted by distilled water and used to evaluate the detection limit of the MNV real-time RT-RPA assay. Each RNA dilution was tested in 8 replicates. A semi-log regression was performed by plotting the threshold time against the log₁₀ RNA copy numbers using Prism 5.0 software (GraphPad, USA). To determine the analytical sensitivity of the RT-RPA technique, a probit regression was performed using Prism 5.0 software (GraphPad, USA).

Due to the high genetic diversity of different MNV strains, the broad reactivity of the RT-RPA technique was investigated. Multiple alignments of sequences from 10 isolates retrieved from Genbank were performed.

These 10 MNV strains were isolated from different countries and are the dominant strains in these countries, the point mutations at the primer-probe set may represent a large diversity of MNV. Six point mutations were present at the binding site of the probe and one mutation was present at the RPA forward primer (Fig. 1). The sequence containing all the seven point mutations were synthesized by Sangon Biotech (Sangon, China) and cloned in pGEM-T vector. The sensitivity of the RT-RPA assay using the mutants as the template was evaluated by the above method.

The specificity of the MNV RT-RPA assay was evaluated by testing a panel of viruses including TMEV, Reo-3, SeV, PVM and MHV.

Detection of clinical samples

Forty eight frozen clinical samples (16 gastric tissue specimens, 16 cecal tissue specimens and 16 fecal specimens) from previous experiment stored at -80°C at our laboratory were subjected to the RT-RPA assay [36]. Homogenates of the tissues were produced with Dulbecco's Modified Eagle Medium (DMEM). The homogenates after two freezing-thawing cycles were clarified by high-speed centrifugation and the supernatant fluids were collected for RNA extraction and RT-RPA assay. All the samples were also tested by the RT-qPCR method previously described [27]. Linear regression analysis of RT-RPA threshold time (TT) and RT-qPCR cycle threshold (C_t) values were performed by Prism 5.0 software (GraphPad, USA).

Abbreviations

MNV: Murine norovirus; RT-PCR: Reverse transcriptase polymerase chain reaction; RT-RPA: Reverse transcriptase recombinase polymerase amplification

Acknowledgments

We would like to thank LetPub (www.letpub.com) for providing linguistic assistance during the preparation of this manuscript.

Funding

This work was supported by grants from Science and Technology Program of Guangdong Province, China (2017A030303026, 2017B030314171, 2017A070702001, 2017A030303025, 2016A030303016) and Science and Technology Program of Guangzhou, China (201707010440). The funders had no role in the design of the study, collection, analysis, and interpretation of data, nor writing of the manuscript.

Availability of data and materials

The datasets contained in this study are available from the corresponding author upon request.

Authors' contributions

PJG and LM designed the study and drafted the manuscript. LM and FWZ conducted the experiments, data analysis, and participated in drafting of the manuscript. FC, BHH, YJZ, MLW, FJX, RH helped with the experiments. WY contributed to the revision of the manuscript. All the authors read and approved the final manuscript.

Ethics approval and consent to participate

Not applicable

Consent for publication

Not applicable

Competing interests

The authors declare that they have no competing interests.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Author details

¹Guangdong Key Laboratory of Laboratory Animals, Guangdong Laboratory Animals Monitoring Institute, Guangzhou, China. ²College of Veterinary Medicine, South China Agricultural University, Guangzhou 510640, China.

Received: 2 September 2018 Accepted: 5 December 2018

Published online: 14 December 2018

References

- Zheng DP, Ando T, Fankhauser RL, Beard RS, Glass RI, Monroe SS. Norovirus classification and proposed strain nomenclature. *Virology*. 2006;346(2):312–23.
- Hsu CC, Riley LK, Livingston RS. Molecular characterization of three novel murine noroviruses. *Virus Genes*. 2007;34(2):147–55.
- Karst SM, Wobus CE, Lay M, Davidson J, Virgin HW. STAT1-dependent innate immunity to a Norwalk-like virus. *Science*. 2003;299(5612):1575–8.
- Hsu CC, Riley LK, Wills HM, Livingston RS. Persistent infection with and serologic cross-reactivity of three novel murine noroviruses. *Comp Med*. 2006;56(4):247–51.
- Muller B, Klemm U, Mas Marques A, Schreier E. Genetic diversity and recombination of murine noroviruses in immunocompromised mice. *Arch Virol*. 2007;152(9):1709–19.
- Smith DB, McFadden N, Blundell RJ, Meredith A, Simmonds P. Diversity of murine norovirus in wild-rodent populations: species-specific associations suggest an ancient divergence. *J Gen Virol*. 2012;93(Pt 2):259–66.
- Thackray LB, Wobus CE, Chachu KA, Liu B, Alegre ER, Henderson KS, Kelley ST. Virgin HW: murine noroviruses comprising a single genogroup exhibit biological diversity despite limited sequence divergence. *J Virol*. 2007;81(19):10460–73.
- Hsu CC, Wobus CE, Steffen EK, Riley LK, Livingston RS. Development of a microsphere-based serologic multiplexed fluorescent immunoassay and a reverse transcriptase PCR assay to detect murine norovirus 1 infection in mice. *Clin Diagn Lab Immunol*. 2005;12(10):1145–51.
- Pritchett-Corning KR, Cosentino J, Clifford CB. Contemporary prevalence of infectious agents in laboratory mice and rats. *Lab Anim*. 2009;43(2):165–73.
- Kitagawa Y, Tohya Y, Ike F, Kajita A, Park SJ, Ishii Y, Kyuwa S, Yoshikawa Y. Indirect ELISA and indirect immunofluorescent antibody assay for detecting the antibody against murine norovirus S7 in mice. *Exp Anim*. 2010;59(1):47–55.
- Cadwell K, Patel KK, Maloney NS, Liu TC, Ng AC, Storer CE, Head RD, Xavier R, Stappenbeck TS, Virgin HW. Virus-plus-susceptibility gene interaction determines Crohn's disease gene Atg16L1 phenotypes in intestine. *Cell*. 2010;141(7):1135–45.
- Higgins PD, Johnson LA, Sauder K, Moons D, Blanco L, Taube S, Wobus CE. Transient or persistent norovirus infection does not alter the pathology of Salmonella typhimurium induced intestinal inflammation and fibrosis in mice. *Comp Immunol Microbiol Infect Dis*. 2011;34(3):247–57.
- Compton SR. Prevention of murine norovirus infection in neonatal mice by fostering. *J Am Assoc Lab Anim Sci*. 2008;47(3):25–30.
- Tajima M, Kotani Y, Kurosawa T, Miyasaka M. Pitfalls in mouse norovirus (MNV) detection in fecal samples using RT-PCR, and construction of new MNV-specific primers. *Exp Anim*. 2013;62(2):127–35.
- Baert L, Wobus CE, Van Coillie E, Thackray LB, Debevere J, Uyttendaele M. Detection of murine norovirus 1 by using plaque assay, transfection assay, and real-time reverse transcription-PCR before and after heat exposure. *Appl Environ Microbiol*. 2008;74(2):543–6.
- Hanaki K, Ike F, Kajita A, Yasuno W, Yanagiba M, Goto M, Sakai K, Ami Y, Kyuwa S. Detection of murine norovirus by reverse transcription loop-mediated isothermal amplification. *J Virol Methods*. 2014;204:17–24.
- Lillis L, Siverson J, Lee A, Cantera J, Parker M, Piepenburg O, Lehman DA, Boyle DS. Factors influencing recombinase polymerase amplification (RPA) assay outcomes at point of care. *Mol Cell Probes*. 2016;30(2):74–8.
- Yang Y, Qin X, Wang G, Zhang Y, Shang Y, Zhang Z. Development of a fluorescent probe-based recombinase polymerase amplification assay for rapid detection of Orf virus. *Viol J*. 2015;12:206.

19. Yang Y, Qin X, Wang G, Jin J, Shang Y, Zhang Z. Development of an isothermal amplification-based assay for rapid visual detection of an Orf virus. *Virology*. 2016;13:46.
20. Yang Y, Qin X, Zhang X, Zhao Z, Zhang W, Zhu X, Cong G, Li Y, Zhang Z. Development of real-time and lateral flow dipstick recombinase polymerase amplification assays for rapid detection of goatpox virus and sheeppox virus. *Virology*. 2017;14(1):131.
21. Yang Y, Qin X, Song Y, Zhang W, Hu G, Dou Y, Li Y, Zhang Z. Development of real-time and lateral flow strip reverse transcription recombinase polymerase amplification assays for rapid detection of peste des petits ruminants virus. *Virology*. 2017;14(1):24.
22. Piepenburg O, Williams CH, Stemple DL, Armes NA. DNA detection using recombination proteins. *PLoS Biol*. 2006;4(7):e204.
23. Amer HM, Abd El Wahed A, Shalaby MA, Almajhdi FN, Hufert FT, Weidmann M. A new approach for diagnosis of bovine coronavirus using a reverse transcription recombinase polymerase amplification assay. *J Virol Methods*. 2013;193(2):337–40.
24. Moore MD, Jaykus LA. Development of a recombinase polymerase amplification assay for detection of epidemic human noroviruses. *Sci Rep*. 2017;7:40244.
25. Yehia N, Arafa AS, Abd El Wahed A, El-Sanousi AA, Weidmann M, Shalaby MA. Development of reverse transcription recombinase polymerase amplification assay for avian influenza H5N1 HA gene detection. *J Virol Methods*. 2015;223:45–9.
26. Bae J, Schwab KJ. Evaluation of murine norovirus, feline calicivirus, poliovirus, and MS2 as surrogates for human norovirus in a model of viral persistence in surface water and groundwater. *Appl Environ Microbiol*. 2008;74(2):477–84.
27. Kitajima M, Oka T, Takagi H, Tohya Y, Katayama H, Takeda N, Katayama K. Development and application of a broadly reactive real-time reverse transcription-PCR assay for detection of murine noroviruses. *J Virol Methods*. 2010;169(2):269–73.
28. Hanaki K, Ike F, Kajita A, Yasuno W, Yanagiba M, Goto M, Sakai K, Ami Y, Kyuwa S. A broadly reactive one-step SYBR green I real-time RT-PCR assay for rapid detection of murine norovirus. *PLoS One*. 2014;9(5):e98108.
29. Wang HM, Zhao GM, Hou PL, Yu L, He CQ, He HB. Rapid detection of foot-and-mouth disease virus using reverse transcription recombinase polymerase amplification combined with a lateral flow dipstick. *J Virol Methods*. 2018;261:46–50.
30. Vasileva Wand NI, Bonney LC, Watson RJ, Graham V, Hewson R. Point-of-care diagnostic assay for the detection of Zika virus using the recombinase polymerase amplification method. *J Gen Virol*. 2018;99(8):1012–26.
31. Chen J, Xu Y, Yan H, Zhu Y, Wang L, Zhang Y, Lu Y, Xing W. Sensitive and rapid detection of pathogenic bacteria from urine samples using multiplex recombinase polymerase amplification. *Lab Chip*. 2018;18(16):2441–52.
32. Ma L, Cong F, Zhu Y, Wu M, Xu F, Huang R, Moore RJ, Guo P. Development of a reverse transcription recombinase polymerase amplification assay for rapid detection of Theiler's murine encephalomyelitis virus. *Mol Cell Probes*. 2018;41:27–31.
33. Abd El Wahed A, El-Deeb A, El-Tholoth M, Abd El Kader H, Ahmed A, Hassan S, Hoffmann B, Haas B, Shalaby MA, Hufert FT, et al. A portable reverse transcription recombinase polymerase amplification assay for rapid detection of foot-and-mouth disease virus. *PLoS One*. 2013;8(8):e71642.
34. Boyle DS, Lehman DA, Lillis L, Peterson D, Singhal M, Armes N, Parker M, Piepenburg O, Overbaugh J. Rapid detection of HIV-1 proviral DNA for early infant diagnosis using recombinase polymerase amplification. *mBio*. 2013;4(2):e00135–13. <https://doi.org/10.1128/mBio.00135-13>.
35. Yuan W, Zhang Y, Wang J, Liu XM, Zhao WB, Huang R. Isolation, identification and genetic analysis of a murine norovirus strain. *Bing Du Xue Bao = Chin J Virol*. 2014;30(4):359–68.
36. Yuan W, Wang J, Xu F, Huang B, Lian Y, Rao D, Yin X, Wu M, Zhu Y, Zhang Y, et al. Development of a duplex real-time RT-PCR for the simultaneous detection and differentiation of Theiler's murine encephalomyelitis virus and rat theilovirus. *J Virol Methods*. 2016;236:139–46.

Ready to submit your research? Choose BMC and benefit from:

- fast, convenient online submission
- thorough peer review by experienced researchers in your field
- rapid publication on acceptance
- support for research data, including large and complex data types
- gold Open Access which fosters wider collaboration and increased citations
- maximum visibility for your research: over 100M website views per year

At BMC, research is always in progress.

Learn more biomedcentral.com/submissions

