Check for updates

OPEN ACCESS

EDITED BY Francesco Ferrara, Santi Paolo e Carlo Hospital, Italy

REVIEWED BY Massimo Baudo, Spedali Civili Brescia, Italy Qian Cui, Guangdong Provincial People's Hospital, China Li Wang, Fudan University, China

*CORRESPONDENCE Yanru Qin yanruqin@zzu.edu.cn

[†]These authors have contributed equally to this work and share first authorship

SPECIALTY SECTION

This article was submitted to Surgical Oncology, a section of the journal Frontiers in Surgery

RECEIVED 30 May 2022 ACCEPTED 03 October 2022 PUBLISHED 31 October 2022

CITATION

Wang L, Han H, Feng L and Qin Y (2022) Development and validation of a nomogram for patients with stage II/III gastric adenocarcinoma after radical surgery. Front. Surg. 9:956256. doi: 10.3389/fsurg.2022.956256

COPYRIGHT

© 2022 Wang, Han, Feng and Qin. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

Development and validation of a nomogram for patients with stage II/III gastric adenocarcinoma after radical surgery

Lei Wang[†], Huiqiong Han[†], Liwen Feng and Yanru Qin^{*}

Department of Oncology, The First Affiliated Hospital of Zhengzhou University, Zhengzhou, China

Background: We aimed to construct nomograms based on clinicopathological features and routine preoperative hematological indices to predict cancer-specific survival (CSS) and disease-free survival (DFS) in patients with stage II/III gastric adenocarcinoma (GA) after radical resection.

Methods: We retrospectively analyzed 468 patients with stage II/III GA after curative gastrectomy between 2012 and 2018; 70% of the patients were randomly assigned to the training set (n = 327) and the rest were assigned to the validation set (n = 141). The nomogram was constructed from independent predictors derived from the Cox regression in the training set. Using the consistency index, the calibration and the time-dependent receiver operating characteristic curves were used to evaluate the accuracy of the nomogram. Decision curve analysis was used to assess the value of the model in clinical applications. Patients were further divided into low- and high-risk groups based on the nomogram risk score.

Results: Multivariate Cox model identified depth of invasion, lymph node invasion, tumor differentiation, adjuvant chemotherapy, CA724, and plateletalbumin ratio as covariates associated with CSS and DFS. CA199 is a risk factor unique to CSS. The nomogram constructed using the results of the multivariate analysis showed high accuracy with a consistency index of 0.771 (CSS) and 0.771 (DFS). Moreover, the area under the curve values for the 3and 5-year CSS were 0.868 and 0.918, and the corresponding values for DFS were 0.872 and 0.919, respectively. The nomogram had a greater clinical benefit than the TNM staging system. High-risk patients based on the nomogram had a worse prognosis than low-risk patients.

Conclusion: The prognostic nomogram for patients with stage II/III GA after radical gastrectomy established in this study has a good predictive ability, which is helpful for doctors to accurately evaluate the prognosis of patients to make more reasonable treatment plans.

KEYWORDS

nomogram, gastric adenocarcinoma, decision curve analysis, prognosis, survival, plateletalbumin ratio

Introduction

Gastric adenocarcinoma (GA), which accounts for 95% of gastric malignancies, is the fifth most common cancer and a major global health challenge (1). GA usually originates from the lining of the stomach, and its early symptoms are not obvious; therefore, many patients with GA are not diagnosed until the metastatic or advanced stage (2). Radical surgery with subsequent adjuvant chemotherapy is the main treatment for early stage GA; however, the overall survival rate remains poor due to the high frequency of metastasis and recurrence (3). Therefore, there is an urgent need to individually analyze independent risk factors and establish novel predictive models that can accurately identify high-risk patients with GA.

Multiple studies have shown that nutritional factors, inflammation, and coagulation are associated with cancer patient outcomes, including GA (4-6). Systemic inflammation and immune evasion are the cardinal features of malignancy, and various inflammatory factors contribute to tumor progression. Interestingly, in addition to neutrophils and lymphocytes, recent studies have revealed that platelets are potent immune modulators and effectors, including direct identification and elimination of pathogens or enhancement of leukocyte immunity, in addition to their central role in hemostasis (7). Additionally, nutritional status is also a critical part of cancer management, especially in gastrointestinal tumors with high prevalence of malnutrition (8). Feng et al. reported that lower preoperative serum albumin levels are associated with unfavorable prognosis in patients with gastric cancer (9). Furthermore, patients with cancer often have abnormal coagulation, which is closely related to tumor progression (10). Several peripheral blood markers have been shown to correlate with cancer prognosis, including the platelet-albumin ratio (PAR), platelet-lymphocyte ratio (PLR), albumin-fibrinogen ratio (AFR), lymphocyte-monocyte ratio (LMR), neutrophil-lymphocyte ratio (NLR), aspartate aminotransferase-alanine aminotransferase ratio (SLR), and D-dimer (9, 11). These indicators can directly reflect inflammation, nutritional level, liver function, and coagulation in patients with cancer. In addition, some classic tumor markers, including carcinoembryonic antigen (CEA), carbohydrate antigen 19-9 (CA19-9), and CA72-4, are widely used for prognostication of cancer patients (12). The hematological indicators mentioned above from routine testing are economical and readily available; therefore, we selected these indicators for evaluation as potential predictive factors.

The nomogram can visualize and integrate independent predictors, realize individualized prognostic assessment, and improve accuracy, and has been studied and applied to multiple cancer types (13, 14). In this study, we attempted to combine clinicopathological characteristics and preoperative routine laboratory indicators to construct and verify a nomogram for patients with stage II/III GA after radical surgery. This personalized prediction system can facilitate clinicians in identifying high-risk patients to develop more personalized treatments.

Materials and methods

Patients and data collection

We retrospectively collected the data of 468 patients with stage II/III GA who underwent radical gastrectomy at the First Affiliated Hospital of Zhengzhou University between May 2012 and May 2018. Seventy percent of the patients were randomly selected as the training set (n = 327), and the rest were assigned to the validation set (n = 141). The inclusion criteria were as follows: (1) histologically confirmed stage II/ III GA; (2) R0 resection; (3) no antitumor therapy before surgery; (4) complete clinicopathological and follow-up data, and all hematological parameters to be assessed should be measured within 1 week before surgery; (5) no other malignancies; (6) no parenteral nutrition, acute inflammation, or significant organ damage within 1 week before surgery; and (7) no cause of death other than GA.

Widely accepted thresholds for grouping continuous variables: D-Dimer (0.3 mg/L), CA199 (35 U/ml), CA724 (6.9 U/ml), and CEA (5 U/ml). The optimal cutoff values for age (68), tumor size (3.5 cm), PAR (6.4), AFR (9.7), NLR (2.3), LMR (3.1), SLR (1.8), and PLR (202.9) were determined using X-tile (15) analysis because of the lack of a defined threshold. Studies involving human participants were reviewed and approved by the Medical Ethics Committee of the First Affiliated Hospital of Zhengzhou University. According to the Declaration of Helsinki, patient data were anonymized and kept confidential. Due to the retrospective nature of this study, informed consent was not obtained.

Follow-up and outcome

Follow-up was done *via* different methods such as medical records and telephone surveys. Patients were observed after curative gastrectomy every 3 months during the first year, every 6 months for 2–3 years, and annually thereafter for up to 5 years post-surgery. Each follow-up included physical examination, laboratory testing, electronic gastroscopy, as clinically indicated, and chest/abdomen/pelvic enhanced computed tomography. The primary endpoint of this study was cancer-specific survival (CSS), which was defined as the time from the date of surgery to cancer-related death. Disease-free survival (DFS) was defined as the time from curative surgery to death, recurrence, or the final follow-up.

Variables	NO. (%)		X^2	Р	Variables	N	NO. (%)		Р
	Training Cohort (<i>n</i> = 327)	Validation Cohort (n = 141)	_			Training Cohort (n = 327)	Validation Cohort (<i>n</i> = 141)		
Sex			0.5	0.480	Adjuvant			0.002	0.962
Male	240 (73.4)	99 (70.2)			chemotherapy	142 (42 7)	(2, (44, 0))		
Female	87 (26.6)	42 (29.8)			No	143 (43.7)	62 (44.0)		
Age, years			3.754	0.053	Yes CA199, U/ml	184 (56.3)	79 (56.0)	0.306	0.580
≤68	263 (80.4)	102 (72.3%)				257 (78 6)	114 (80.0)	0.300	0.380
>68	64 (19.6)	39 (27.7)			<u>≤</u> 35	257 (78.6)	114 (80.9)		
Family history			0.41	0.522	>35	70 (21.4)	27 (19.1)	0.017	0.898
No	268 (82.0)	119 (84.5)			CA724, U/ml	251 (76.9)	100 (77.2)	0.017	0.898
Yes	59 (18.0)	22 (15.5)			≤6.9	251 (76.8)	109 (77.3)		
Diabetes			0.653	0.419	>6.9	76 (23.2)	32 (22.7)	0.000	0.622
No	312 (95.4)	132 (93.6)			CEA, U/ml	257 (79.6)	100 (7(()	0.229	0.632
Yes	15 (4.6)	9 (6.4)			≤5	257 (78.6)	108 (76.6)		
Hypertension			0.044	0.843	>5	70 (21.4)	33 (23.4)		
No	264 (80.7)	115 (81.6)			D-Dimer, mg/l	222 (54.2)		0.065	0.798
Yes	63 (19.3)	26 (18.4)			≤0.3	238 (74.3)	101 (71.6)		
Tobacco			0.076	0.783	>0.3	89 (27.2)	40 (28.4)	0.514	0.453
No	236 (72.2)	100 (70.9)			PLR	245 (54.0)	110 (50.0)	0.514	0.473
Yes	91 (27.8)	41 (29.1)			≤202.9	245 (74.9)	110 (78.0)		
Alcohol			0.217	0.641	>202.9	82 (25.1)	31 (22.0)		
No	263 (80.4)	116 (82.3)			PAR		05 (60 2)	1.515	0.218
Yes	64 (19.6)	25 (17.7)			≤6.4	177 (54.1)	85 (60.3)		
Depth of invasion			2.041	0.153	>6.4	150 (45.9)	56 (39.7)		
T1-2	80 (24.5)	26 (18.4)			AFR	(15.0)	10 (12 5)	0.181	0.671
T3-4	247 (75.5)	115 (81.6)			<9.7	49 (15.0)	19 (13.5)		
Lymph node			0.126	0.722	≥9.7	278 (85.0)	122 (86.5)		
invasion					NLR		/ >	0.224	0.636
N0	83 (25.4)	38 (27.0)			≤2.3	184 (56.3)	76 (53.9)		
N1-3	244 (74.6)	103 (73.0)			>2.3	143 (43.7)	65 (46.1)		
TNM stage			0.066	0.798	LMR			0.061	0.805
II	113 (34.6)	47 (33.3)			<3.1	119 (36.4)	53 (37.6)		
III	214 (65.4)	94 (66.7)			≥3.1	208 (63.6)	88 (62.4)		
Tumor differentiation			0.661	0.416	SLR ≤1.8	280 (85.6)	119 (84.4)	0.119	0.731
	70 (21.4)	25 (24.8)			>1.8	47 (14.4)	22 (15.6)		
Middle or high	70 (21.4)	35 (24.8)			>1.0	47 (14.4)	22 (13.0)		
low Size cm	257 (78.6)	106 (75.2)	0.247	0.610					
Size, cm	105 (22 1)	12 (20 %)	0.247	0.619	Statistical	analysis			
≤3.5 >3.5	105 (32.1)	42 (29.8)							
>3.5 Turner lesstion	222 (67.9)	99 (70.2)	4 5	0.104	Statistical	analysis was pe	erformed using S	PSS Sta	tistics
Tumor location	166 (50.0)	75 (52.2)	4.5	0.104	(version 26.0,	IBM, USA) and	R software (versic	on 4.1.2)	. The
Upper 1/3	166 (50.8)	75 (53.2)			optimal cut-or	ff value was det	ermined using X-	tile sof	tware.
Middle 1/3	73 (22.3)	20 (14.2)			The continuo	us variables wer	e transformed int	to categ	orical

TABLE 1 Continued

 $\ensuremath{\mathsf{TABLE}}\xspace$ 1 Baseline characteristics of patients in the training and validation cohorts.

(continued)

cs ıe e. The continuous variables were transformed into categorical variables. The chi-square test was used to compare categorical data. Statistical significance was set at P < 0.05. For continuous

Lower 1/3

88 (26.9)

46 (32.6)

variables, the Kolmogorov-Smirnov test was first performed. If an approximately normal distribution was displayed, the data were described using the mean and standard deviation. Otherwise, the median with the interquartile range (IQR) was used. First, the proportional hazards hypothesis test was performed using the Cox regression model. If the hypothesis was not satisfied, a Cox regression model with timedependent covariates was used (16). Significant factors in the univariate analysis will be included in the Cox regression equation to identify independent factors that will be used to construct nomograms. Nomograms for CSS and DFS were constructed using the rms and survival packages in the R software. Internal validation was performed to demonstrate the reliability and repeatability of the nomograms. The consistency index (C-index), calibration curve, and timedependent receiver operating characteristic curve were used to evaluate the accuracy and discriminative ability of the prediction map in the training and validation sets, respectively. The ggDCA package in the R software was used

to construct the decision curve analysis (DCA) to further evaluate the clinical benefit of the nomogram. Each patient was scored using the survival package in R software and divided into high- and low-risk groups based on the median risk score. The Kaplan–Meier method was used to draw the CSS and DFS survival curves, and the log-rank test was used for statistical analysis.

Results

Clinicopathological characteristics

Table 1 shows the clinicopathological characteristics of the 468 patients with GA, including 327 patients in the training set and 141 patients in the validation set. In the training set, the age of the patients at diagnosis ranged from 25 to 88 years, with a median age of 61 years (IQR, 52–66 years). Most patients were male (n = 240, 73.4%), and the rest were female (n = 87,

TABLE 2 Univariate and multivariate cox regression analyses of prognostic factors for cancer-specific survival.

Variables	Univariate A	nalysis	Multivariate Analysis		
	HR (95% CI)	P-value	HR (95% CI)	P-value	
Sex (male)	0.929 (0.674–1.283)	0.656			
Age (>68)	1.721 (1.239–2.390)	0.001	0.953 (0.660-1.377)	0.797	
Family history (yes)	0.959 (0.659-1.398)	0.829			
Diabetes (yes)	1.458 (0.793-2.681)	0.225			
Hypertension (yes)	1.000 (0.695-1.439)	0.999			
Tobacco (yes)	0.750 (0.536-1.049)	0.093			
Alcohol (yes)	0.720 (0.487-1.065)	0.100			
Depth of invasion (T3-4)	4.981 (2.979-8.326)	<0.001	6.495 (3.763-11.208)	<0.001	
Lymph node invasion (N1-3)	3.111 (2.037-4.753)	<0.001	4.762 (2.995-7.573)	<0.001	
Differentiation (low)	1.720 (1.169-2.531)	0.005	1.657 (1.102-2.492)	0.015	
Tumor size (>3.5)	1.852 (1.327-2.585)	<0.001	1.042 (0.731-1.486)	0.819	
Tumor location					
Upper 1/3	1				
Middle 1/3	1.102 (0.771-1.574)	0.515			
Lower 1/3	0.853 (0.601-1.212)	0.375			
Adjuvant chemotherapy (yes)	0.599 (0.449-0.799)	<0.001	0.582 (0.426-0.796)	0.001	
CA199 (>35)	2.385 (1.740-3.269)	<0.001	1.436 (1.019–2.023)	0.038	
CA724 (>6.9)	2.800 (2.056-3.812)	<0.001	1.704 (1.223–2.375)	0.002	
CEA (>5)	1.362 (0.974-1.906)	0.071			
D-Dimer (>0.3)	1.575 (1.160-2.139)	0.004	1.165 (0.843-1.609)	0.355	
PLR (>202.9)	1.839 (1.355-2.496)	<0.001	0.878 (0.620-1.242)	0.461	
PAR (>6.4)	2.660 (1.980-3.575)	<0.001	1.822 (1.299-2.556)	0.001	
AFR (<9.7)	2.469 (1.741-3.500)	<0.001	1.081 (0.734-1.592)	0.693	
NLR (>2.3)	0.814 (0.608-1.090)	0.168			
LMR (<3.1)	0.745 (0.548-1.014)	0.061			
SLR (>1.8)	1.728 (1.194-2.499)	0.004	1.010 (0.682-1.498)	0.959	

26.6%). The median tumor size was 4.5 cm (IQR, 3.5-6.0 cm). According to the 8th edition of the American Joint Committee on Cancer staging system, it is more common in patients with pT3-4 (n = 247, 75.5%), and most patients have lymph node invasion (n = 244, 74.6%). Most patients with GA were classified as stage III (n = 214, 65.4%). Poor differentiation (n = 257, 78.6%) was the most common tumor grade. In total, 184 patients (56.3%) received adjuvant chemotherapy. Furthermore, the baseline characteristics did not differ between the training and validation groups (P >0.05). For all patients, the final follow-up period ranged from 0.5 to 96 months, with a median of 36.0 months. In the training set, the 1-, 3-, and 5-year CSS rates were 82.9, 48.6, and 40.3%, respectively. The 1-, 3-, and 5-year DFS rates were 75.5, 46.8, and 39.5%, respectively. In the validation set, the 1-, 3-, and 5-year CSS rates were 81.6, 51.1, and 44.6%, respectively. The 1-, 3-, and 5-year DFS rates were 75.2, 48.9, and 42.9%, respectively.

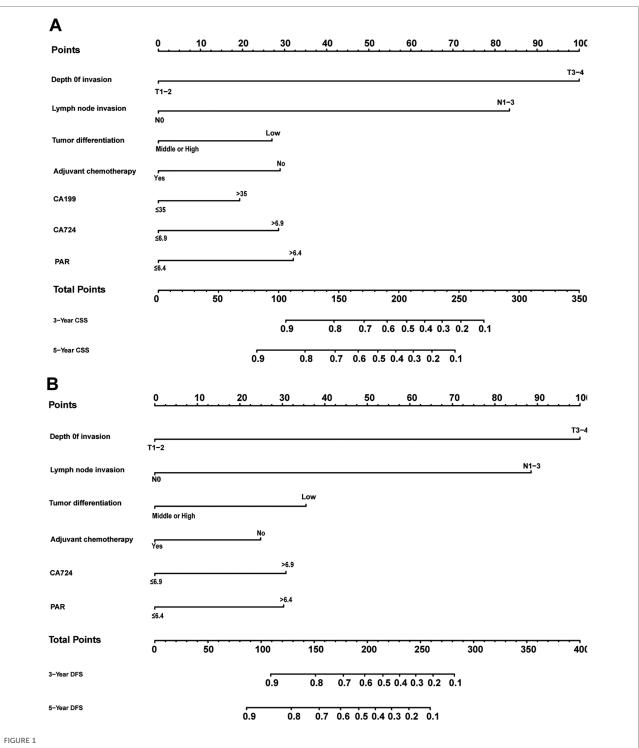
Independent predictors

Tables 2, 3 show the results of univariate and multivariate Cox analyses on the training set data, including hazard ratios and 95% confidence intervals (CI). Univariate analysis showed that age, depth of invasion, lymph node invasion, tumor size, differentiation, adjuvant chemotherapy, CA199, CA724, Ddimer, PLR, PAR, SLR, and AFR were related to CSS and DFS, while CEA was only relevant for DFS (P < 0.05). Significant factors in the univariate analysis were included in multivariate analysis. Depth of invasion (P < 0.001), lymph node invasion (P < 0.001), tumor differentiation (P = 0.015), adjuvant chemotherapy (P = 0.001), CA199 (P = 0.038), CA724 (P = 0.002), and PAR (P = 0.001) were independent predictors of CSS, while depth of invasion (P < 0.001), lymph node invasion (P < 0.001), tumor differentiation (P = 0.003), adjuvant chemotherapy (P = 0.004), CA724 (P = 0.001), and PAR (P = 0.002) were independent predictors of DFS.

TABLE 3 Univariate and multivariate cox regression analyses of prognostic factors for disease-free survival.

Variables	Univariate Ar	nalysis	Multivariate Analysis		
	HR (95% CI)	P-value	HR (95% CI)	<i>P</i> -value	
Sex (male)	0.926 (0.675-1.271)	0.636			
Age (>68)	1.631 (1.78-1.2.258)	0.003	0.930 (0.648-1.336)	0.695	
Family history (yes)	0.952 (0.657-1.379)	0.793			
Diabetes (yes)	1.359 (0.739-2.499)	0.323			
Hypertension (yes)	1.074 (0.755-1.526)	0.692			
Tobacco (yes)	0.719 (0.489-1.056)	0.093			
Alcohol (yes)	0.718 (0.488-1.055)	0.091			
Depth of invasion (T3-4)	4.265 (2.652-6.858)	<0.001	5.951 (3.583-9.886)	<0.001	
Lymph node invasion (N1-3)	3.180 (2.096-4.824)	<0.001	4.848 (3.067-7.662)	<0.001	
Differentiation (low)	1.712 (1.175-2.493)	0.005	1.887 (1.241-2.870)	0.003	
Tumor size (>3.5)	1.908 (1.372-2.654)	<0.001	1.091 (0.767-1.551)	0.630	
Tumor location					
Upper 1/3	1				
Middle 1/3	1.147 (0.809-1.627)	0.442			
Lower 1/3	0.831 (0.586-1.179)	0.300			
Adjuvant chemotherapy (yes)	0.650 (0.489-0.863)	0.003	0.642 (0.473-0.870)	0.004	
CA199 (>35)	2.426 (1.776-3.315)	<0.001	1.315 (0.924-1.871)	0.128	
CA724 (>6.9)	2.905 (2.138-3.948)	<0.001	1.735 (1.235-2.438)	0.001	
CEA (>5)	1.424 (1.024-1.979)	0.036	1.390 (0.953-2.029)	0.087	
D-Dimer (>0.3)	1.473 (1.086-1.996)	0.013	1.065 (0.768-1.476)	0.706	
PLR (>202.9)	1.863 (1.378-2.519)	<0.001	0.964 (0.684-1.359	0.834	
PAR (>6.4)	2.542 (1.899-3.401)	<0.001	1.718 (1.226-2.407)	0.002	
AFR (<9.7)	2.356 (1.6663-3.338)	<0.001	1.097 (0.753-1.599)	0.628	
NLR (>2.3)	0.808 (0.605-1.078)	0.147			
LMR (<3.1)	0.764 (0.564-1.035)	0.082			
SLR (>1.8)	1.750 (1.216-2.518)	0.003	1.041 (0.708-1.529)	0.839	

05



Nomogram predicting cancer-specific survival (A) and disease-free survival (B) of patients with stage II/III gastric adenocarcinoma who underwent gastrectomy.

Construction and verification of nomogram models

Figure 1 shows the nomogram predicting CSS and DFS that was constructed based on the results of the multivariate analysis with hazard ratios. The models can score each patient, and the

higher the score, the worse is the prognosis. The C-index for predicting CSS and DFS was 0.771 (95%CI, 0.738–0.804) and 0.771 (95%CI, 0.740–0.802), suggesting that the constructed nomogram had an accurate predictive ability. In addition, as shown in **Figure 2**, the calibration curve is close to the diagonal line, suggesting that the nomogram predicts the

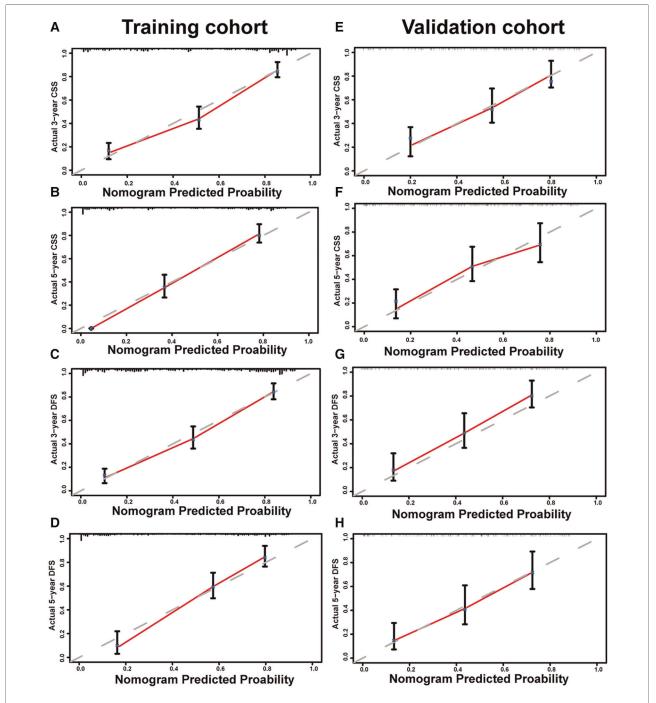


FIGURE 2

Calibration curves of nomogram for predicting 3-year cancer-specific survival (CSS) (A), 5-year CSS (B), 3-year disease-free survival (DFS) (C), and 5-year DFS (D) in the training set. Calibration curves of nomogram for predicting 3-year CSS (E), 5-year CSS (F), 3-year DFS (G), and 5-year DFS (H) in the validation set.

patients' 3- and 5-year CSS and DFS to be similar to the actual situation, further illustrating the predictive accuracy of the model. In the nomogram model, the area under the curve (AUC) for the 3- and 5-year CSS were 0.868 (95%CI, 0.829-0.907) and 0.918 (95%CI, 0.884-0.953), respectively, and the AUC for the 3- and 5-year DFS was 0.872 (95%CI, 0.834-0.910) and 0.919 (95%CI, 0.885-0.952), respectively. In TNM staging, the AUC for 3- and 5-year CSS were 0.696 (95%CI, 0.648-0.743) and 0.749 (95%CI, 0.683-0.816), respectively, and the AUC for 3- and 5-year DFS were 0.712 (95%CI, 0.664-0.760) and 0.747 (95%CI, 0.678-0.816), respectively. These results showed that the nomogram model had a higher accuracy than TNM staging (Figure 3). Similarly, in the validation set, the C-indices for CSS and DFS were 0.734 (95%CI, 0.679-0.789) and 0.731 (95%CI, 0.680-0.782), respectively, and the calibration curve predicted survival probability with high agreement, which also proved the reliability of the model.

Decision curve analysis and survival curves based on the nomograms

As shown in **Figure 4**, DCA was used to analyze the net benefit rate of the nomogram and TNM staging at different threshold probabilities. The results showed that the nomogram had a higher clinical benefit than the TNM staging system when the probability threshold was 0.2 to 0.8, whether it was 3-year or 5-year CSS, or 3-year or 5-year DFS. The results of the validation set were consistent with those of the training set. **Figure 5** shows that patients in the high-risk group, based on the median nomogram score, had a worse prognosis than those in the low-rank group (P < 0.001).

Discussion

GA is a complex disease and surgical or endoscopic resection remains the only cure. However, the current survival rate of patients with GA remains low, even with the combined efforts of multidisciplinary teams. Over the years, tumor microenvironment has been the focus of our research (17, 18), and chronic inflammation and immune dysfunction are its most important features (19). The release of various pro-inflammatory cytokines and inflammatory substances such as interleukin-1ß, interleukin-8, and tumor necrosis factor- α promotes GA progression (19). Neutrophils and lymphocytes are the most important immune cells that can intuitively reflect the level of inflammation in the human body, and abnormal changes in these cells are thought to be related to tumor progression. Recently, in addition to their known hemostatic effects, researchers found that platelets are potent immunomodulators that play an important role in regulating systemic inflammation and immunity (20).

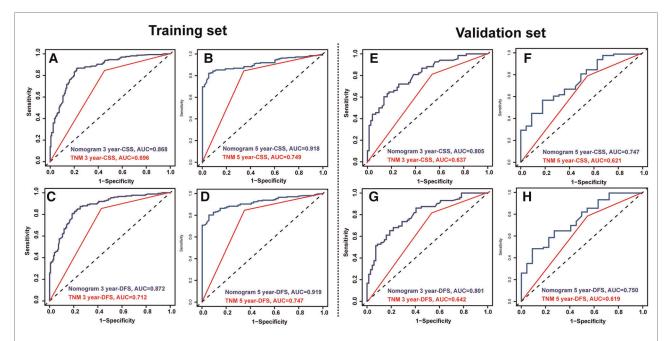
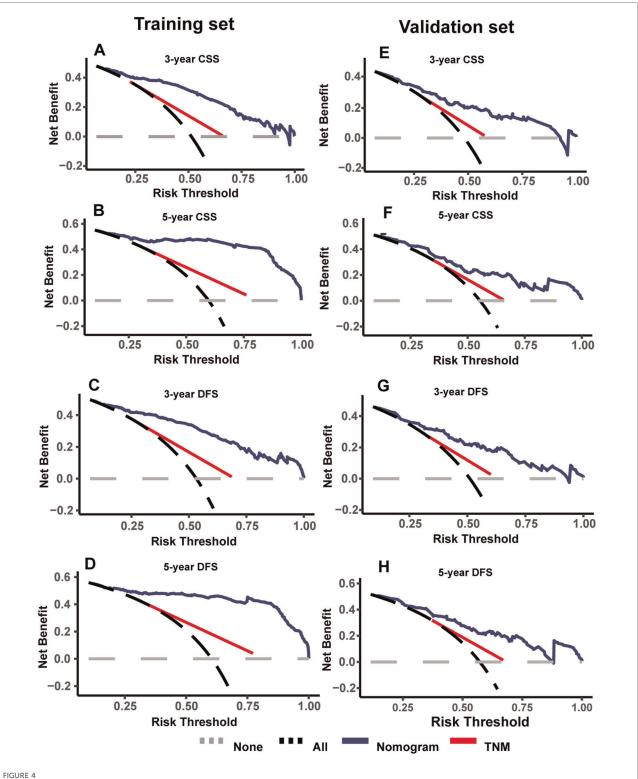
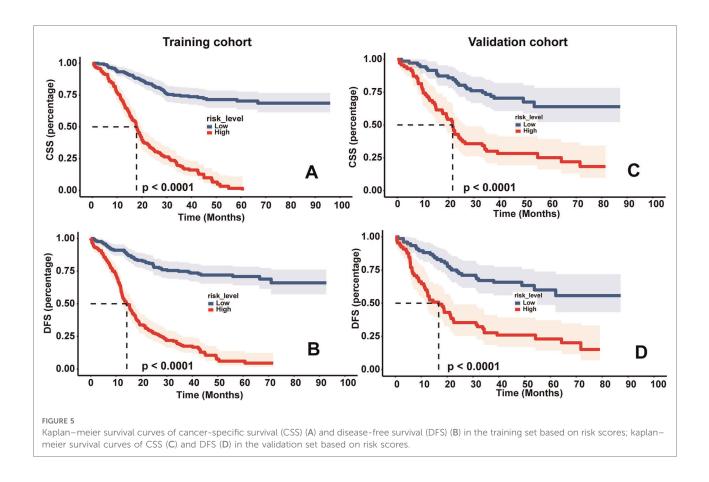


FIGURE 3

Time-dependent reciever operating characteristics (ROC) curves of nomograms and TNM staging were used to test the predictive power of the 3-year cancer-specific survival (CSS) (A) and 5-year CSS (B), 3- year disease-free survival (DFS) (C) and 5-year DFS (D) in the training set. Time-dependent ROC curves of nomograms and TNM staging were used to test the predictive power of the 3-year CSS (E) and 5-year CSS (F), 3- year DFS (G) and 5-year DFS (H) in the validation set.



Decision curve analysis were used to compare the clinical benefit of nomogram and TNM staging in cancer-specific survival (CSS) (A,B) and diseasefree survival (DFS) (C,D) in the training set; decision curve analysis were used to compare the clinical benefit of nomogram and TNM staging in CSS (E,F) and DFS (G,H) in the validation set.



Moreover, tumors release a variety of cytokines including vascular endothelial growth factor, platelet-derived growth factor, and transforming growth factor-\beta1 that can promote platelet production, and the increased platelets can shield tumor cells in peripheral blood and interfere with immune cells to enhance the metastatic potential of tumors (21, 22). However, the nutritional status of cancer patients is an important part of cancer management. The prevalence of malnutrition in patients with cancer is high, particularly in those with gastrointestinal tumors. According to one study, the prevalence of malnutrition in digestive malignancies is approximately 52% (23). Albumin is widely recognized as an indicator of nutritional levels, and the mechanism of hypoalbuminemia is related to the increase in two inflammatory cytokines, tumor necrosis factor- α and interleukin-6, which inhibit the synthesis of albumin (24). A study by Feng et al. indicated that malnutrition is significantly associated with poor prognosis in patients with gastric cancer (9). Hypercoagulability is an important physiological characteristic of patients with malignant tumors. Abnormalities in indicators that can represent coagulation, such as D-dimer and fibrinogen, are often associated with poor prognosis in patients with tumors (25, 26).

In our study, we mainly focused on PLR, PAR, NLR, LMR, STR, AFR, and D-dimer levels, which can reflect inflammation, nutritional status, liver function, and coagulation function in patients. These routine hematological markers are readily available and economical, and previous studies have shown that these indicators are associated with the prognosis of malignancies (11). The NLR and LMR are indicators of systemic inflammation. Previous studies have shown that an increase in NLR or decrease in LMR indicates that the inflammatory response promotes tumor development, indicating a poor prognosis for patients (27, 28). The relationship between PLR and cancer prognosis is unclear, and the underlying mechanism may be related to platelet and lymphocyte functions (28). SLR is an indicator of liver function. Previous studies have indicated that people with an elevated SLR have an increased risk of gastric cancer (29). PAR has also been explored in a variety of other cancers, such as pancreatic, esophageal, and liver cancers; however, its significance in GA has not been explored (30, 31). The study of Huang et al. pointed out that PAR is an independent predictor of esophageal squamous cell carcinoma and can accurately predict the prognosis of patients with this cancer (32). Similar to PAR, AFR also reflects multiple patient metrics, including nutritional levels and coagulation. Feng et al. confirmed its association with gastric cancer prognosis (9). A lower AFR appears to indicate a worse

prognosis. In our study, compared to PAR, the other remaining indicators showed no statistical significance as independent prognostic factors in the multivariable Cox regression analysis. This result suggests that we may need to pay more attention to the interaction of platelets with tumor cells and other immune cells. However, preoperative attention to the nutritional status of patients and timely intervention may help prolong patient survival. A study by Bang et al. showed that postoperative platinum-based chemotherapy in patients with gastric cancer can effectively prolong the survival time of patients (33). Our study is consistent with previous findings that timely adjuvant chemotherapy is one of the protective factors for patient outcomes.

A nomogram is an effective predictive tool that quantifies individual risk, and its intuitive and visual features make predictive models more readable and facilitate clinical application (13). In this study, we constructed a nomogram to predict the prognosis of patients with stage II/III GA after curative gastrectomy. The results showed that TNM stage, tumor differentiation, postoperative adjuvant chemotherapy, tumor markers, and PAR were independent patient predictors. In addition, high-risk patients identified using the model had a poorer prognosis. Therefore, timely interventions for high-risk patients, including improved nutrition and inhibition of inflammation, may improve patient outcomes. Compared with the nomogram for stage II/III gastric cancer patients after curative gastrectomy followed by adjuvant chemotherapy constructed by Li-tong Shi (11), our model is different; we only focused on one pathological type, GA; we included adjuvant chemotherapy as a variable and incorporated different hematological indicators while constructing receiver operator characteristic and DCA curves to evaluate the clinical applicability of the model. In our study, the relationship between PAR and GA prognosis was demonstrated, which has not been previously reported. This further confirms that immunity and nutritional status are closely related to GA prognosis. However, we acknowledge that this study has some limitations. First, this was a retrospective single-center study, and the preliminary results need to be further validated in prospective clinical trials. Second, we only performed internal validation of the nomogram, and external validation was required to extend the applicability of the model. Third, we mainly focused on economical and convenient routine laboratory tests; therefore, we did not conduct research on EGFR mutations, Her-2 expression, ERBB2 expression, or microsatellite instability status. Some indicators, such as Helicobacter pylori and PG I/II, were not routinely detected; therefore, they were not included in this study. These metrics will be the focus of future research.

Conclusion

In conclusion, the nomogram constructed in this study for patients with stage II/III GA after curative gastrectomy could accurately predict CSS and DFS. In addition, our models are more accurate than traditional TNM staging for predicting GA prognosis and may bring more clinical benefits to patients. For patients with poor outcomes, shortening the follow-up time and timely intervention can be used to prevent the occurrence of adverse events.

Data availability statement

The raw data supporting the conclusions of this article will be made available by the authors, without undue reservation.

Ethics statement

The studies involving human participants were reviewed and approved by the medical ethics committee of the First Affiliated Hospital of Zhengzhou University. Written informed consent for participation was not required for this study in accordance with the national legislation and the institutional requirements.

Author contributions

LW, HQH, and LWF were involved in the study design, data collection, and statistical analysis. LW was responsible for writing the manuscript. YRQ designed the study and revised the manuscript critically. All authors contributed to the article and approved the submitted version.

Funding

This study was supported by a grant from the National Natural Science Foundation of China (No. 81872264).

Acknowledgments

We thank our families for supporting this work.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Publisher's note

All claims expressed in this article are solely those of the authors and do not necessarily represent those of their

References

1. Smyth EC, Nilsson M, Grabsch HI, van Grieken NC, Lordick F. Gastric cancer. *Lancet (London, England)*. (2020) 396(10251):635-48. doi: 10.1016/S0140-6736(20)31288-5

2. Sitarz R, Skierucha M, Mielko J, Offerhaus GJA, Maciejewski R, Polkowski WP. Gastric cancer: epidemiology, prevention, classification, and treatment. *Cancer Manag Res.* (2018) 10:239–48. doi: 10.2147/CMAR.S149619

3. Gambardella V, Cervantes A. Precision medicine in the adjuvant treatment of gastric cancer. *Lancet Oncol.* (2018) 19(5):583–4. doi: 10.1016/S1470-2045(18) 30131-1

4. Pabinger I, van Es N, Heinze G, Posch F, Riedl J, Reitter EM, et al. A clinical prediction model for cancer-associated venous thromboembolism: a development and validation study in two independent prospective cohorts. *Lancet Haematology.* (2018) 5(7):e289–e98. doi: 10.1016/S2352-3026(18)30063-2

5. Shinko D, Diakos CI, Clarke SJ, Charles KA. Cancer-related systemic inflammation: the challenges and therapeutic opportunities for personalized medicine. *Clin Pharmacol Ther.* (2017) 102(4):599–610. doi: 10.1002/cpt.789

6. Gupta D, Lis CG. Pretreatment serum albumin as a predictor of cancer survival: a systematic review of the epidemiological literature. *Nutr J.* (2010) 9:69. doi: 10.1186/1475-2891-9-69

7. Jenne CN, Kubes P. Platelets in inflammation and infection. *Platelets*. (2015) 26(4):286–92. doi: 10.3109/09537104.2015.1010441

8. Bouleuc C, Anota A, Cornet C, Grodard G, Thiery-Vuillemin A, Dubroeucq O, et al. Impact on health-related quality of life of parenteral nutrition for patients with advanced cancer cachexia: results from a randomized controlled trial. *Oncologist.* (2020) 25(5):e843–e51. doi: 10.1634/theoncologist.2019-0856

 Feng LW, Li J, Liang LF, Guo QQ, Li J, Wu J, et al. A predictive scoring system based on inflammatory and tumor markers for gastric cancer patients undergoing curative resection. *Cancer Manag Res.* (2020) 12:3937–48. doi: 10. 2147/CMAR.S250408

10. Nasser NJ, Fox J, Agbarya A. Potential mechanisms of cancer-related hypercoagulability. Cancers (Basel). (2020) 12(3):566. doi: 10.3390/cancers12030566

11. Shi L, Wang Z, Wang L, Jia Y, Li J, Qin Y. A prognostic nomogram and heat map to predict survival in stage II/III gastric cancer patients after curative gastrectomy followed by adjuvant chemotherapy. *Cancer Manag Res.* (2022) 14:287–301. doi: 10.2147/CMAR.S348890

12. Locker GY, Hamilton S, Harris J, Jessup JM, Kemeny N, Macdonald JS, et al. ASCO 2006 Update of recommendations for the use of tumor markers in gastrointestinal cancer. *J Clin Oncol.* (2006) 24(33):5313–27. doi: 10.1200/JCO. 2006.08.2644

13. Park SY. Nomogram: an analogue tool to deliver digital knowledge. J Thorac Cardiovasc Surg. (2018) 155(4):1793. doi: 10.1016/j.jtcvs.2017.12.107

14. Iasonos A, Schrag D, Raj GV, Panageas KS. How to build and interpret a nomogram for cancer prognosis. *J Clin Oncol.* (2008) 26(8):1364–70. doi: 10. 1200/JCO.2007.12.9791

15. Camp RL, Dolled-Filhart M, Rimm DL. X-tile: a new bio-informatics tool for biomarker assessment and outcome-based cut-point optimization. *Clin Cancer Res.* (2004) 10(21):7252–9. doi: 10.1158/1078-0432.CCR-04-0713

16. Zhang Z, Reinikainen J, Adeleke KA, Pieterse ME, Groothuis-Oudshoorn CGM. Time-varying covariates and coefficients in cox regression models. *Ann Transl Med.* (2018) 6(7):121. doi: 10.21037/atm.2018.02.12

17. Arneth B. Tumor microenvironment. Medicina (Kaunas, Lithuania). (2019) 56 (1):15. doi: 10.3390/medicina56010015

18. Wang L, Han H, Wang Z, Shi L, Yang M, Qin Y. Targeting the microenvironment in esophageal cancer. *Front Cell Dev Biol.* (2021) 9:684966. doi: 10.3389/fcell.2021.684966

affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

19. Lee K, Hwang H, Nam KT. Immune response and the tumor microenvironment: how they communicate to regulate gastric cancer. *Gut Liver*. (2014) 8(2):131–9. doi: 10.5009/gnl.2014.8.2.131

 Boilard E, Nigrovic PA, Larabee K, Watts GF, Coblyn JS, Weinblatt ME, et al. Platelets amplify inflammation in arthritis via collagen-dependent microparticle production. *Science (New York, NY)*. (2010) 327(5965):580–3. doi: 10.1126/science.1181928

21. Palumbo JS, Talmage KE, Massari JV, La Jeunesse CM, Flick MJ, Kombrinck KW, et al. Platelets and fibrin(ogen) increase metastatic potential by impeding natural killer cell-mediated elimination of tumor cells. *Blood.* (2005) 105 (1):178–85. doi: 10.1182/blood-2004-06-2272

22. Stoiber D, Assinger A. Platelet-leukocyte interplay in cancer development and progression. *Cells.* (2020) 9(4):855. doi: 10.3390/cells9040855

23. Attar A, Malka D, Sabaté JM, Bonnetain F, Lecomte T, Aparicio T, et al. Malnutrition is high and underestimated during chemotherapy in gastrointestinal cancer: an AGEO prospective cross-sectional multicenter study. *Nutr Cancer.* (2012) 64(4):535–42. doi: 10.1080/01635581.2012.670743

24. Tanaka T, Narazaki M, Kishimoto T. IL-6 in inflammation, immunity, and disease. *Cold Spring Harbor Perspect Biol.* (2014) 6(10):a016295. doi: 10.1101/cshperspect.a016295

25. Diao D, Wang Z, Cheng Y, Zhang H, Guo Q, Song Y, et al. D-dimer: not just an indicator of venous thrombosis but a predictor of asymptomatic hematogenous metastasis in gastric cancer patients. *PloS one*. (2014) 9(7):e101125. doi: 10.1371/ journal.pone.0101125

26. Suzuki T, Shimada H, Nanami T, Oshima Y, Yajima S, Ito M, et al. Hyperfibrinogenemia is associated with inflammatory mediators and poor prognosis in patients with gastric cancer. *Surg Today*. (2016) 46(12):1394–401. doi: 10.1007/s00595-016-1339-z

27. Stotz M, Szkandera J, Stojakovic T, Seidel J, Samonigg H, Kornprat P, et al. The lymphocyte to monocyte ratio in peripheral blood represents a novel prognostic marker in patients with pancreatic cancer. *Clin Chem Lab Med.* (2015) 53(3):499–506. doi: 10.1515/cclm-2014-0447

28. Luo Z, Jiao B, Huang T, Zhao H, Zhang G. What is the role of the preoperative blood-based inflammation biomarkers in the prognosis of upper tract urothelial carcinoma with radical nephroureterectomy? A single-centre retrospective study. *Technol Cancer Res Treat.* (2022) 21:15330338221095667. doi: 10.1177/15330338221095667

29. Kobayashi D, Yamamoto K, Kimura T, Shimbo T. Aspartate aminotransferase/alanine aminotransferase ratio and subsequent cancer development. *Cancer Med.* (2022) 11(3):798-814. doi: 10.1002/cam4.4473

30. Fan Z, Luo G, Gong Y, Xu H, Qian Y, Deng S, et al. Prognostic value of the C-reactive protein/lymphocyte ratio in pancreatic cancer. *Ann Surg Oncol.* (2020) 27(10):4017–25. doi: 10.1245/s10434-020-08301-3

31. Xu B, Li XL, Ye F, Zhu XD, Shen YH, Huang C, et al. Development and validation of a nomogram based on perioperative factors to predict post-hepatectomy liver failure. *J Clin Transl Hepatol.* (2021) 9(3):291–300. doi: 10. 14218/JCTH.2021.00013

32. Huang Z, Zheng Q, Yu Y, Zheng H, Wu Y, Wang Z, et al. Prognostic significance of platelet-to-albumin ratio in patients with esophageal squamous cell carcinoma receiving definitive radiotherapy. *Sci Rep.* (2022) 12(1):3535. doi: 10.1038/s41598-022-07546-0

33. Bang YJ, Kim YW, Yang HK, Chung HC, Park YK, Lee KH, et al. Adjuvant capecitabine and oxaliplatin for gastric cancer after D2 gastrectomy (CLASSIC): a phase 3 open-label, randomised controlled trial. *Lancet (London, England).* (2012) 379(9813):315–21. doi: 10.1016/S0140-6736(11)61873-4