

Development of β -cyclodextrin-based hydrogel microparticles for solubility enhancement of rosuvastatin: an in vitro and in vivo evaluation

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Abstract: The aim of this study was to enhance the solubility of rosuvastatin (RST) calcium by developing β -cyclodextrin-g-poly(2-acrylamido-2-methylpropane sulfonic acid [AMPS]) hydrogel microparticles through aqueous free-radical polymerization technique. Prepared hydrogel microparticles were characterized for percent entrapment efficiency, solubility studies, Fourier transform infrared spectroscopy, differential scanning calorimetry, thermal gravimetric analysis, powder X-ray diffraction, scanning electron microscopy, zeta size and potential, swelling and release studies. Formulations (HS1–HS9) have shown entrapment efficiency between 83.50%±0.30% and 88.50%±0.25%, and optimum release was offered by formulation HS7 at both pH levels, ie, 1.2 (89%) and 7.4 (92%). The majority of microparticles had a particle size of less than 500 μ m and zeta potential of –37 mV. Similarly, optimum solubility, ie, 10.66-fold, was determined at pH 6.8 as compared to pure RST calcium, ie, 7.30-fold. In vivo studies on fabricated hydrogel microparticulate system in comparison to pure drug were carried out, and better results regarding pharmacokinetic parameters were seen in the case of hydrogel microparticles. A potential approach for solubility enhancement of RST calcium and other hydrophobic moieties was successfully developed. **Keywords:** hydrogel microparticles, rosuvastatin calcium, polymerization, β -cyclodextrin solubility

Introduction

Newly discovered as well as existing chemical substances have solubility issues due to their lipophilic nature that results in failure of new drug developments.¹ Drugs administered in low doses should be soluble in water to produce therapeutic effects.² Efforts are continuously made to improve solubility and bioavailability of drugs.^{3–5} Different approaches such as particle size reduction,^{6–8} solubilization by using a surfactant system,^{9–11} hydrosols, interpenetrating networks,¹² salt formation,¹³ rapid dissolving tablets,¹⁴ hydrogel microparticles and formation of water-soluble complexes, ie, solid dispersions and inclusion complexes with the aid of hydrophilic polymers, etc., have been tried in the literature successfully.¹⁵

Hydrogels are cross-linked three-dimensional polymeric networks having ability to imbibe large quantities of water in them while keeping their original geometry. These matrices are extensively employed in pharmaceutical and medical fields due to their special properties, such as excellent swelling, drug loading, degradability, gelling capacity, gel ability and biocompatibility. In the literature, hydrogel microparticles have been utilized for the delivery of insulin and solubility enhancement of acyclovir, methotrexate and other therapeutic agents.^{16,17}

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These systems are thermally more stable when compared to conventional complex systems, eg, solid dispersions and inclusion complexes, meaning that hydrogel microparticulate systems prevent rapid drug polymer decomplexation and rapid release and improve solubility to a greater extent while having weak interactions, eg, van der Waal forces and hydrogen bonding that promote fast dissociation and release of drug.^{18,20}

Rosuvastatin (RST) calcium, a hydroxy-methyl-glutaryl-coenzyme A (HMG-CoA) inhibitor, is one of the latest drugs of the statin class. It is widely used for the treatment of familial hyperlipidemia, dyslipidemia, triglyceridemia, atherosclerosis, osteoporosis, benign prostatic hyperplasia and Alzheimer's disease. It belongs to Biopharmaceutics Classification System-II (BCS-II), thereby presenting solubility issues. It is crystalline in nature. It particularly lowers low-density lipoprotein (LDL) level and usually increases the level of high-density lipoproteins (HDLs). It has low bioavailability of 20% due to its low water solubility and extensive metabolism offered by liver via oxidation, lactonization and glucuronidation.^{16,19,20}

β -Cyclodextrin (β -CD) is an acyclic oligosaccharide that has an inner hydrophobic cavity and hydrophilic outer rim. CDs are used in various sectors, ie, agriculture, cosmetics, chemical and pharmaceutical, as they possess innate ability to improve physiochemical properties of water drugs via complexation. They promote solubility, bioavailability and stability of the drugs. β -CD and γ -CD are listed in the generally regarded as safe category of the US Food and Drug Administration (FDA). β -CD has a prominent place in the pharmaceutical sector as it is economical and easily available and has dimensions of cavity that are suitable for most of the drug candidates. When CDs are polymerized with other polymers or monomers, their ability to enhance solubility and bioavailability increases, ie, hydrogels.²¹ Swelling depends upon the type of monomer used for polymerization, ie, pH-independent swelling is offered by 2-acrylamido-2-methylpropane sulfonic acid (AMPS) while methacrylic acid offers pH-dependent and ionic-responsive swelling behaviors.²²

AMPS is a white crystalline powder that is chemically produced from acrylonitrile and isobutylene by Ritter reaction in the presence of powerful sulfonic acid and aqueous media (water). It is freely soluble in water and dimethyl formamide (DMF) and has very limited solubility in polar organic liquids. AMPS-based pharmaceutical drug carriers dissociate over all pH ranges that give rise to pH-independent swelling.²⁴ Moreover, AMPS hydrogels have applications in skin-sensitive electrodes, carriers in biomedical engineering, muscle actuators and in drug delivery.²³

We present the development of β -CD- and AMPS-based hydrogel microparticulate system for solubility enhancement of RST calcium. The method could be considered a simple method for the development of hydrogel microparticles by chemical cross-linking with *N,N'*-methylene bisacrylamide (MBA) that was tuned and optimized for solubility enhancement purposes. Moreover, to the best of our knowledge, this hydrogel microparticulate system has not been used before for potentiating solubility of RST calcium.

Materials and methods

RST calcium was obtained as a generous gift from Getz Pharmaceuticals (Karachi, Pakistan). β -CD (97%), AMPS (99%), ammonium persulfate (APS), MBA and methanol were purchased from Sigma-Aldrich Co. (St Louis, MO, USA). Acetonitrile, diethyl ether, absolute ethanol, dichloromethane, chloroform and diethyl acetate were purchased from Sigma-Aldrich Co. The 0.45 μ m-sized membrane filters were purchased from Sartorius (Goettingen, Germany). High-performance liquid chromatography (HPLC) grade ultrapure water was prepared by Milli-Q[®] system (EMD Millipore, Billerica, MA, USA).

Synthesis of β -CD-g-poly(AMPS) hydrogel microparticles

New polymeric graft was prepared through aqueous free-radical polymerization technique. All the ingredients listed in Table 1 were accurately weighed. APS and MBA were used as initiator and crosslinker, respectively. Both polymer and monomer were separately taken, weighed and poured into beakers containing equal volumes of water. Both the solutions were heated at 50°C on a hot plate magnetic stirrer with continuous stirring until the formation of clear solutions. A weighed quantity of APS was poured into β -CD solution to initiate polymerization while MBA was added into the monomer

Table 1 Composition of β -CD-g-poly(AMPS) hydrogel microparticles

Code	β -CD (g/100 g)	AMPS (g/100 g)	MBA (g/100 g)	APS (g/100 g)
HS1	0.5	2	0.3	0.1
HS2	1	2	0.3	0.1
HS3	2	2	0.3	0.1
HS4	1	4	0.3	0.1
HS5	1	5	0.3	0.1
HS6	1	6	0.3	0.1
HS7	1	4	0.4	0.1
HS8	1	4	0.5	0.1
HS9	1	4	0.6	0.1

Abbreviations: AMPS, 2-acrylamido-2-methylpropane sulfonic acid; APS, ammonium persulfate; β -CD, β -cyclodextrin; MBA, *N,N'*-methylene bisacrylamide.

(AMPS) solution. Stirring was continued till the clarity of both solutions was attained. Afterward, the monomer solution was added dropwise into the polymer solution. The whole mixture was then sonicated (5 min) and vortexed (3 min), and nitrogen gas was purged (20 min) to ensure complete removal of oxygen and proper mixing. This mixture was added into clean and dry labeled glass molds. Test tubes were sealed with aluminum foil, racked in test tube stand and shifted to a digital hot water bath (Memmert, Japan) for 24 h, ie, 55°C for 4 h, 60°C for 8 h, 65°C for 8 h and 70°C for 4 h. Upon solidification, test tubes were removed, cooled down to room temperature and broken. Hydrogel disks were cut with a sharp-edged blade into possible small size particles. Particles were soaked in hydroalcoholic solution, ie, methanol and water (50:50), to remove unreacted species. Washing was continued until stable pH value, ie, 6.5–7, of washing media was attained. Particles were separated from washing media using mesh, and were freeze dried at –55°C for 24 h (Christ Alpha, 1-4LD; Fuzhou Xing Shun Da Refrigeration Facility Project Co., Ltd., Japan). Freeze-dried particles were passed through sieve number 80 to obtain a micrometric size range. These were stored in air-tight containers for further analysis.²⁵ Schematic representation of β -CD-co-poly(AMPS) hydrogel microparticles is shown in Figure 1.

Drug loading

Diffusion-supported swelling method was adapted for loading of RST in hydrogel microparticles. About 1% RST solution was prepared in phosphate buffer (pH 6.8) by dissolving 1 g of RST in phosphate buffer with a final volume of 100 mL due to solubility concerns of RST at this pH value. Lyophilized hydrogel microparticles were weighed precisely

and soaked into drug solution for 24 h at room temperature. Swollen microparticles were separated on screen, and these were flushed with sufficient water to remove surface residues of RST. RST-loaded polymeric microparticles were dried at 40°C in a hot air oven (Memmert).²⁶

Entrapment efficiency (EE) and product yield

An extraction method was used to calculate EE and product yield. Prepared microparticles (RST =20 mg) were soaked into phosphate buffer of pH 6.8 (50 mL) to facilitate swelling and drug release. RST determination was performed by developing a calibration curve using a ultraviolet (UV)–visible spectrophotometer at 243 nm. EE and product yield were calculated by using the following equations:²⁷

$$\text{Product yield} = 100 - \text{Mass loss}$$

$$\text{Mass loss (\%)} = \frac{M_0 - M_1}{M_0} \times 100$$

where M_0 is the initial weight and M_1 is the final weight.

Entrapment efficiency =

$$\frac{\text{Absorbance of sample containing 20 mg RST}}{\text{Absorbance of 20 mg pure RST}} \times 100$$

Experimental techniques

Fourier transform infrared (FTIR) spectroscopy

Attenuated total reflectance (ATR) technology was used for the determination of functional groups and to record

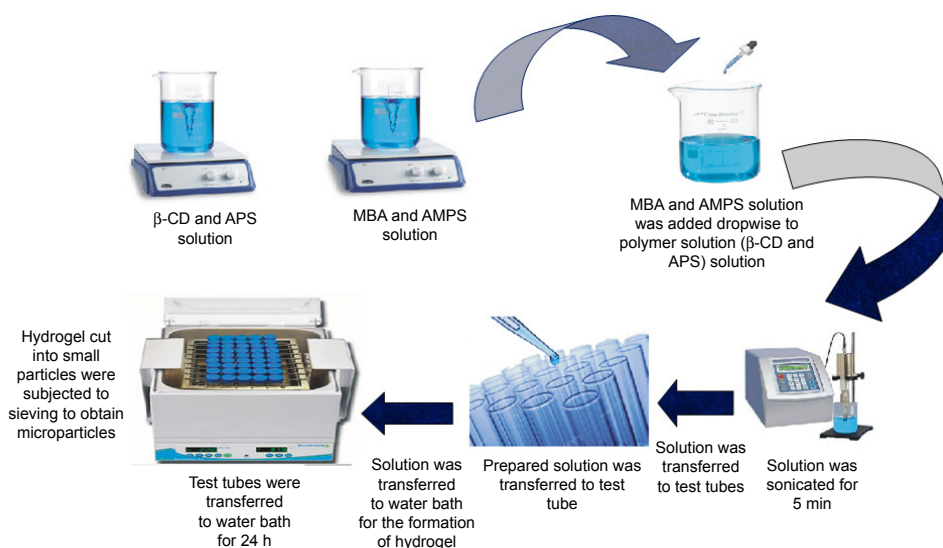


Figure 1 Schematic representation of hydrogel microparticle synthesis.

Abbreviations: AMPS, 2-acrylamido-2-methylpropane sulfonic acid; APS, ammonium persulfate; β -CD, β -cyclodextrin; MBA, *N,N'*-methylene bisacrylamide.

infrared (IR) spectra of individual ingredients, ie, APS, MBA, AMPS, RST, physical mixture of β -CD and RST, physical mixture of β -CD, APS, MBA, AMPS, RST and developed hydrogel microparticles. The FTIR spectrophotometer (Tensor 27; OPUS software) was operated after placing the sample on the crystal point of the instrument between 400 and 4,000 cm^{-1} .²⁸

Thermal analysis

To measure heat of fusion, melting point, mass loss with temperature rise and stability studies of β -CD, AMPS, APS, MBA, physical mixture and prepared hydrogel microparticles, differential scanning calorimetry (DSC) and thermal gravimetric analysis (TGA) studies were performed. Samples were triturated and wrapped in an aluminum pan. Simultaneous thermal analyzer (Q600 TA V8.3 Build 101 Thermal Analysis System; TA instruments, New Castle, DE, USA) was operated at 20°C/min under a stream of nitrogen, in triplicate.²⁸

Scanning electron microscopy (SEM)

SEM is a powerful and reliable tool to investigate morphology of hydrogels and their structure under JEOL Analytical Scanning Electron Microscope (JSM-6490A; JEOL, Tokyo, Japan). Dried microparticles were glued on an aluminum stub with conductive carbon tape, followed by ~300 Å thickness of gold coating under argon atmosphere using a gold sputter coater. Finally, samples were scanned, and photomicrographs were recorded and analyzed.²⁶

Powder X-ray diffraction (PXRD) studies

The nature of the individual ingredients and hydrogel microparticles, ie, crystalline or amorphous, was confirmed on RXD X'pert PRO with software Pan Analytical®. Results were obtained in triplicate using a scanning range of $2\theta=5^\circ-65^\circ$.

Transmission electron microscopy (TEM)

Internal morphology and geometry was confirmed by TEM JOEL-2010 microscope at 120 kV using different magnifications.²⁹

Zeta size and zeta potential measurements

Particle size of the resultant particles was determined by using particle size analyzer (Zetasizer Ver System; Malvern Instruments, Malvern, UK). Zeta potential of the fabricated hydrogel microparticles was determined to ensure the stability of the polymeric microparticles.³⁰

Dissolution studies

Dissolution studies were performed in a calibrated six-station dissolution test apparatus (USP type-2; Pharma Test, Hainburg, Germany) at pH 1.2 and 6.8, respectively. The instrument was operated at 50 rpm, while the temperature was kept at $37^\circ\text{C}\pm 0.2^\circ\text{C}$ throughout the experiment. Hydrogel microparticles were enclosed in empty hard gelatin capsule shells and soaked in each basket. About 5 mL of samples was withdrawn from center, upper and lower portions of the basket by using a graduated pipette and was replaced with fresh dissolution media at predetermined time intervals. The average amount of drug released from hydrogel microparticles at each sampling time was determined by using a UV-visible spectrophotometer (Pharma Spec 1700; Shimadzu, Kyoto, Japan) at 243 nm.

Empirical calculations

Release data were inserted into empirical equations (kinetic models) by using drug dissolution solver Excel adds in program. Best fit model was confirmed from r^2 values, and the mechanism of release followed by hydrogel microparticles was confirmed by the value of “n,” ie, if $n=0.45$, then the drug diffusion mechanism is Fickian diffusion, and if $0.45 < n < 0.89$, then it is non-Fickian or anomalous diffusion. If “n” is equal to 0.89, then it will be case II transport or typical zero-order release.

Zero-order kinetics:

$$Q_t = Q_o - K_o t$$

First-order kinetics:

$$\text{Ln } Q_t = \text{Ln } Q_o - K_1 t$$

Higuchi kinetic model:

$$Q_t = K_h t^{\frac{1}{2}}$$

where Q_o is the initial amount of RST in hydrogel microparticles, Q_t is the amount of drug released at time “t” and K_o , K_1 and K_h are the rate constants. To determine the mode of drug release, the initial 60% drug release values were fitted to Korsmeyer–Peppas model:

$$\frac{M_t}{M_\infty} = K t^n$$

where M_t/M_∞ is the fraction of drug released at time t , K is the drug release rate constant and n is the release exponent. The n value is employed for the characterization of different release modes for cylindrical-shaped solid formulations. Drug release exponent (n) and drug release mode are related as: $n=0.45$, Fickian diffusion; $0.45 < n < 0.89$, anomalous (non-Fickian) diffusion; $n=0.89$, case II transport and $n > 0.89$, super case II transport. Dissolution data were evaluated using DDSolver.³¹

Solubility studies

Solubility studies of pure RST calcium and RST loaded in the microparticulate system were carried out at pH 1.2, pH 6.8 and in deionized water. Hydrogel microparticles were enclosed into screw-capped vials having 1 mL of buffer media. A thermo shaker incubator (MSC-100, Hangzhou Allsheng Instruments Co., Ltd, Allsheng, China) was used for shaking these vials at $37^\circ\text{C} \pm 0.5^\circ\text{C}$ at 150 rpm for 72 h. All samples were filtered through 0.45 μm membrane filters, diluted with buffer and analyzed for RST on a UV-visible spectrophotometer at 243 nm.³²

Swelling studies

Swellability of prepared hydrogel microparticles was determined at 37°C by enclosing the known amount of hydrogel microparticles (previously enclosed into empty tea bags) in 100 mL 0.5 M buffer solutions of pH 1.2 and 6.8. Tea bags were taken out at predefined time intervals to calculate dynamic swelling and weighed after removing the excess of water by blotting with tissue paper. Swelling studies were continued until constant weight of tea bags was not achieved. Degree of swelling and equilibrium water content were determined using the following equations:³³

$$Q = \frac{M_s}{M_d}$$

$$\text{EWC}\% = \frac{M_s - M_d}{M_d} \times 100$$

where EWC is the equilibrium water content, M_s is the mass of swollen hydrogel microparticles and M_d is the mass of dried hydrogel microparticles.

In vivo studies

In vivo studies were performed using Agilent 1100 series HPLC system to calculate different pharmacokinetic parameters, ie, C_{max} , T_{max} , area under the curve (AUC), mean residence time (MRT), volume of distribution (V_z), C_{last} and $t_{1/2}$ in rabbit plasma. The whole study was reviewed and approved by the

Pharmacy Research Ethics Committee (PREC) of the Islamia University of Bahawalpur (reference no 81-2014/PREC). Moreover, PREC guidelines were followed for the welfare of experimental animals (notification no 81-2014/PREC). PREC guidelines, for conducting the experiments on animals, follow the protocols of the US FDA that are prepared under the International Conference on Harmonisation guidelines.

Study design

A total of 12 male albino rabbits of average weight 1.5 ± 0.010 kg and age 6 months were used for this study. Rabbits were divided into two groups ($n=6$). All the rabbits were fasted overnight with ad libitum and free access to water during the experiment. One group of animals received a single dose of hydrogel microparticles containing 20 mg of RST calcium while the second group was administered commercially available 20 mg tablets of Rovista manufactured by Getz Pharmaceuticals (Karachi, Pakistan). Hair in the sampling area was removed by using depilatory cream, ie, EU-Cream from the neck area to expose the jugular vein, 3 h before dose administration. Samples were withdrawn at predetermined time intervals, ie, 0, 0.5, 1, 1.5, 2, 3, 4, 8 and 12 h from the jugular vein by cleansing the targeted area with methylated spirits, and 3 mL of blood was collected, using a sterile insulin syringe each time, in EDTA tubes, followed by 5–15 mL Glaxose-D solution. No sign of ailment was seen in any rabbit of either group. Each blood sample was centrifuged (Hitachi Zentrifugen EBA 20, Andreas Hettich GmbH & Co. KG (Hettich), Tuttlingen, Germany) at 5,000 rpm for 10 min. Plasma was carefully removed and collected in eppendorf tubes. Separated plasma was stored at -70°C in an ultralow freezer (maximum -86°C ; Sanyo, Japan).

Frozen plasma was taken and allowed to thaw at room temperature. About 1 mL of plasma was taken in glass centrifuge tubes; 0.2 mL of perchloric acid (20%) was used as protein precipitant and 20 mL of perchloric acid (20%) was prepared by taking 6.45 mL of 32.25% perchloric acid and the volume was made up to 20 mL with distilled water. After the addition of protein precipitant, tubes were vortexed (Mini shaker; IKA, Wilmington, NC, USA) for 3 min to attain proper mixing. Tubes were subjected to centrifugation at 5,000 rpm for 10 min. The separated layer was removed carefully by using a micropipette and collected in eppendorf tubes. It was filtered, and the volume of 20 μL was injected for analysis.

Statistical analysis

All the results were statistically evaluated by using one-way analysis of variance (ANOVA).

Results

Product yield, EE and drug loading

Results of product yield and EE were within the acceptable limits. All formulations (HS1–HS9) had a product yield of 87.00%–90.50% and EE from 83.50% to 88.50%. Maximum %EE was seen in the case of formulation HS3, ie, 88.50%. Moreover, the maximum amount of drug was loaded into formulation HS7, ie, 77%. All the results are summarized in Table 2.

FTIR spectroscopy

FTIR spectra of pure drug, polymer, monomer and developed hydrogel microparticles were recorded to confirm the development of graft polymer. FTIR spectrum of the physical mixture was also recorded to observe any interaction. Spectra were recorded between 500 and 4,000 cm^{-1} . IR spectrum of RST calcium showed that characteristic peaks were present at 3,337.90, 2,968.23 and 1,435.48 cm^{-1} corresponding to cyclic amines, C–H stretching, C=O stretching and O–H bending. The spectrum of β -CD had alcoholic O–H stretching at 3,292.74 cm^{-1} , methyl and methylene C–H stretching vibration at 2,927.62 cm^{-1} and C=O ether stretching vibration at 1,021.63 cm^{-1} . RST calcium was also physically mixed with APS, AMPS and MBA to see any interaction among ingredients. However, results did not reveal any interaction among ingredients when they were physically mixed. The results are shown in Figure 2.

Thermal analysis (DSC and TGA)

In the DSC thermogram of β -CD, two endothermic peaks were seen at 106.89°C and 329.46°C that correspond to moisture loss and the melting point of β -CD. Upon complete combustion, an exothermic event was observed at 382.31°C. RST showed a sharp endothermic peak at 143.71°C that was near to the melting point of RST. Variation in melting point

Table 2 EE, product yield and drug loading

Code	EE (%)	Product yield ^a (%)	Drug loading (%)
HS1	83.50±0.30	89.00±1.30	65±0.35
HS2	84.60±1.15	88.90±0.15	69±0.50
HS3	88.50±0.50	90.00±0.25	73±1.05
HS4	87.50±0.25	89.50±1.15	71±0.75
HS5	87.50±0.30	89.10±0.25	69±0.65
HS6	88.50±0.25	87.00±1.30	74±0.73
HS7	87.80±0.25	89.10±0.50	77±0.75
HS8	88.40±0.15	90.50±0.25	73±0.43
HS9	86.60±1.30	88.70±0.30	68±0.55

Note: ^aAverage of three determinations; \pm standard deviation.

Abbreviation: EE, entrapment efficiency.

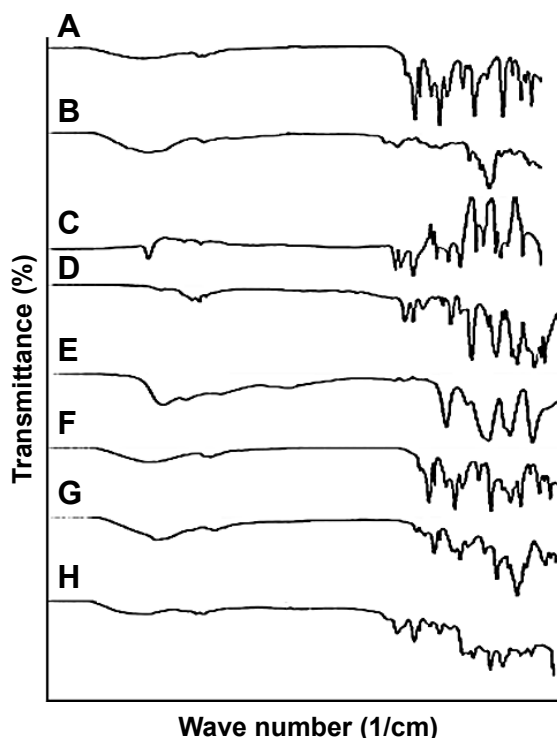


Figure 2 FTIR spectra of (A) RST calcium, (B) β -cyclodextrin, (C) MBA, (D) AMPS, (E) APS, (F) physical mixture of β -CD and RST calcium, (G) physical mixture of RST, APS, AMPS and MBA, (H) hydrogel microparticles.

Abbreviations: AMPS, 2-acrylamido-2-methylpropane sulfonic acid; APS, ammonium persulfate; β -CD, β -cyclodextrin; FTIR, Fourier transform infrared; MBA, *N,N'*-methylene bisacrylamide; RST, rosuvastatin.

of RST may be due to %age purity of RST and source of supply. Another endothermic peak at 243.71°C was seen upon further increase in temperature, and on final combustion at 378.77°C an exothermic event occurred. In the case of the grafted polymeric network, two sharp prominent peaks were observed that indicated that the newly polymerized network was thermally stable.

TGA curve of pure RST calcium presented gradual reduction in mass with the increase in temperature. This loss in mass occurred in three steps, and up to 331.72°C, 47.92% of mass was lost. TGA thermogram of pure β -CD presented mass loss of 84.28% when heated up to 359.21°C. In the case of the grafted polymeric network, DSC studies showed endothermic peaks at 318.47°C and 372.01°C. TGA of developed hydrogel microparticles offered greater stability to RST as there was only 32.75% mass loss observed at 372.01°C. DSC and TGA results are shown in Figure 3.

SEM

SEM micrographs showed that the majority of polymeric microparticles had irregular geometry and variable diameter from each dimension while few had spherical geometry, as shown in Figure 4.

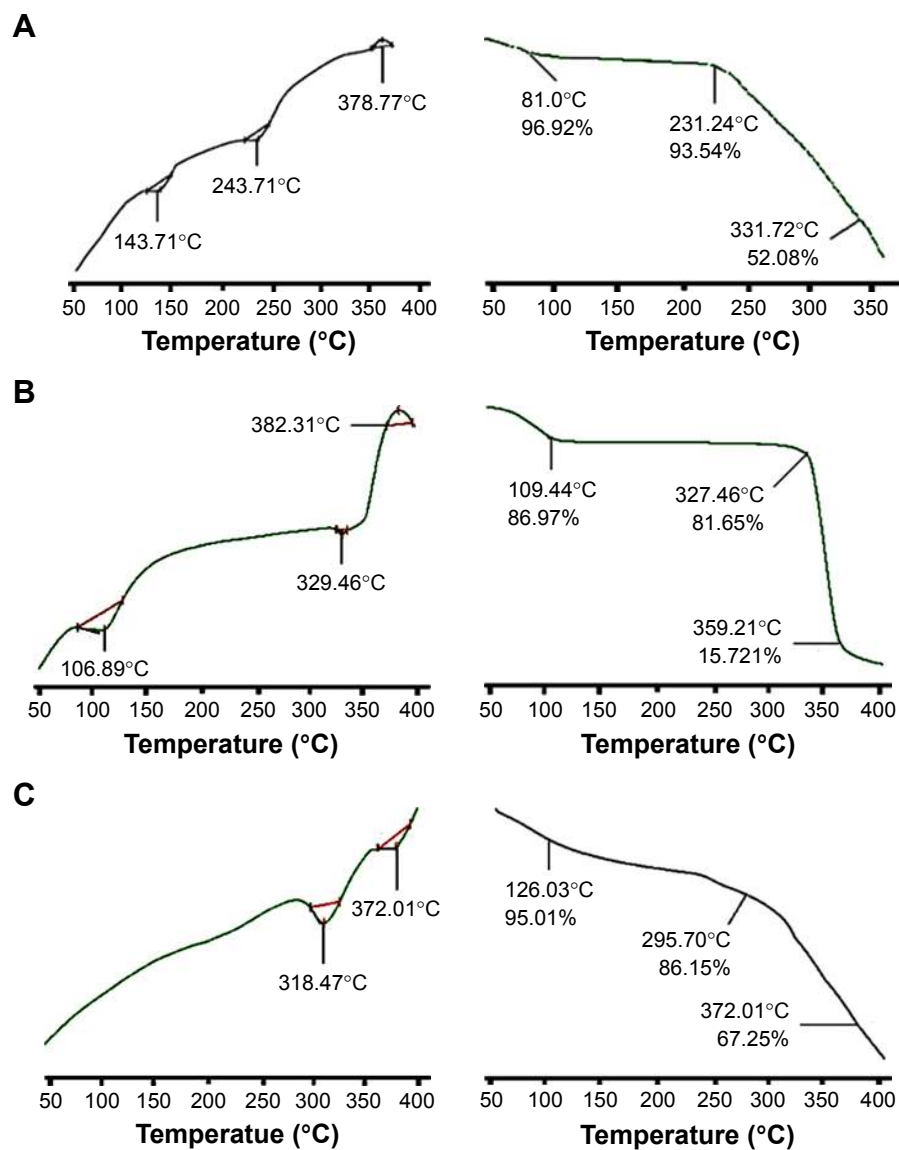


Figure 3 DSC (left) and TGA (right) curves of (A) RST calcium, (B) β -CD, (C) hydrogel microparticles.

Abbreviations: β -CD, β -cyclodextrin; DSC, differential scanning calorimetry; RST, rosuvastatin; TGA, thermal gravimetric analysis.

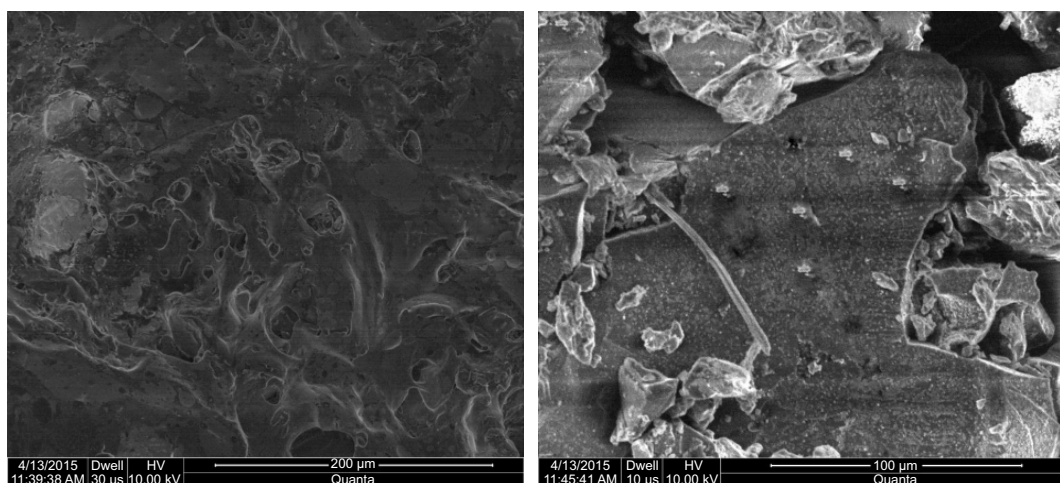


Figure 4 Scanning electron micrographs of lyophilized hydrogel microparticles.

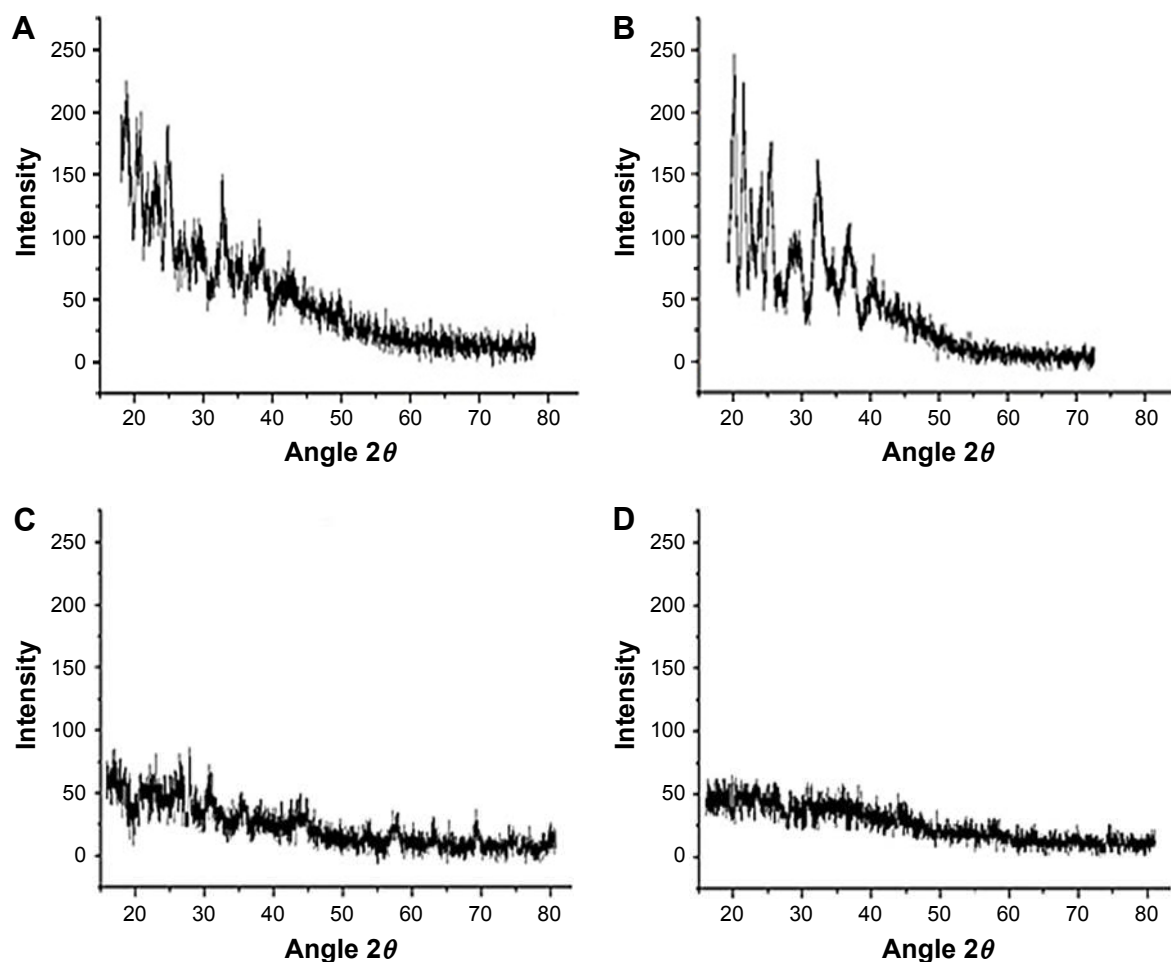


Figure 5 PXRD pattern of (A) RST calcium, (B) β -CD, (C) physical mixture and (D) hydrogel microparticles.
Abbreviations: β -CD, β -cyclodextrin; PXRD, powder X-ray diffraction; RST, rosuvastatin.

PXRD studies

The presence of sharp peaks at $2\theta=16.04^\circ$, 22.45° and 34.3° and $2\theta=20.99^\circ$, 26.85° , 34.63° , 40.03° and 43.85° confirmed the crystalline nature of RST and β -CD, respectively, as shown in Figure 5.

TEM

TEM micrographs of fabricated microparticles from cross-sectional area were recorded. Micrographs proved that microparticles had porous surface. RST was transported through these pores and appeared as a whitish mass. All the results are shown in Figure 6.

Zeta size and zeta potential measurements

Values of the zeta size of formulations were in micron size ranging from 200 to 500 μm but the majority of microparticles had a size of 300 μm , while zeta potential value was near to -37 mV , as shown in Figure 7A and B.

Dissolution studies

Results of dissolution studies showed that there was a marked difference in the release of drug from hydrogel microparticles when compared with the marketed brand of

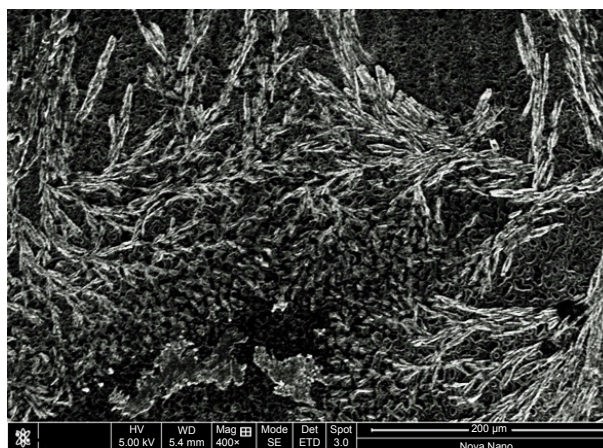


Figure 6 TEM micrograph of lyophilized hydrogel microparticles.
Abbreviation: TEM, transmission electron microscopy.

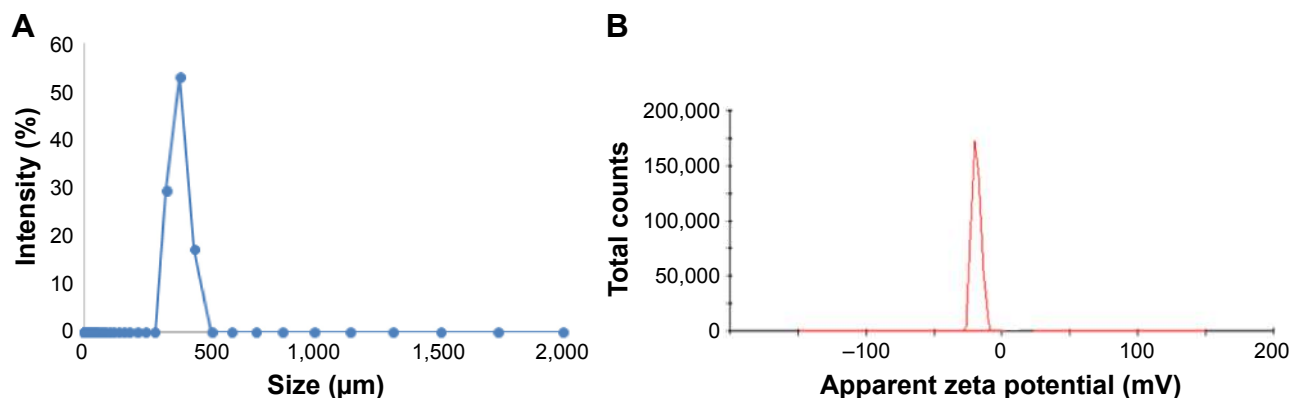


Figure 7 Zeta size and zeta potential.

Notes: (A) Zeta size measurement of hydrogel microparticles. (B) Zeta potential measurements of hydrogel microparticles.

RST, ie, tablet Rovista. Better release results were observed at pH 6.8 as compared to pH 1.2 due to the acidic nature of RST calcium. It is a weak acid and becomes more ionized in basic environment thus more soluble in basic media. From formulation HS7, 92% of drug was released within 1 h while from RST tablets only 44% of drug was released in 3 h at pH 6.8, as shown in Figure 8A. Slightly less amount of drug was released from hydrogel microparticles at pH 1.2, as shown in Figure 8B, but the optimized formulation was HS7.

Solubility studies

Solubility of hydrogel microparticles was potentiated at acidic, basic and in distilled water. This was due to pH-independent swelling behavior of AMPS. The solubility of hydrogel microparticles was enhanced up to 7.5-fold at pH 1.2, while 10.66-fold solubility was enhanced at pH 6.8. In distilled water, the solubility of RST-loaded hydrogel

microparticles was increased up to 7.30-fold, as shown in Figure 9.

Swelling studies

Release of drug from hydrogel microparticles is dependent upon swelling behavior. If higher swelling is present within hydrogel microparticles, then more rapid drug will be released. Developed hydrogel microparticles have higher swelling at pH 7.4 as compared to pH 1.2. At pH 1.2, equilibrium swelling percent (ES%) was between 5.45% and 76.14%, while at pH 7.4 ES% was between 7.93% and 84.12%, as shown in Figure 10.

In vivo pharmacokinetic studies

Different pharmacokinetic parameters such as C_{max} (ng/mL), T_{max} (h), $t_{1/2}$ (h), AUC_{0-t} (ng/mL·h), MRT (h), C_{last} (ng/mL) and T_{last} (h) were calculated. These pharmacokinetic parameters

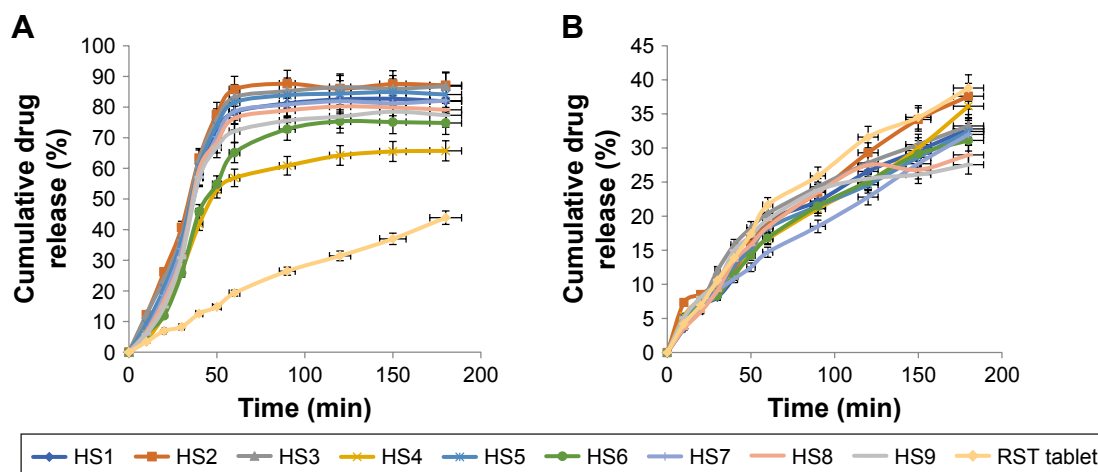


Figure 8 In vitro drug release profiles.

Notes: (A) Cumulative% drug release from hydrogel microparticles and RST tablets at pH 6.8. (B) Cumulative% drug release from hydrogel microparticles and RST tablets at pH 1.2.

Abbreviation: RST, rosuvastatin.

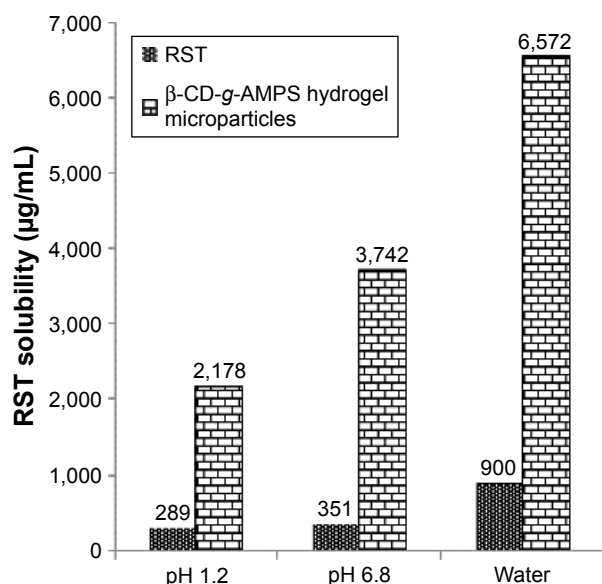


Figure 9 Solubility studies of pure drug and hydrogel microparticles.
Abbreviations: AMPS, 2-acrylamido-2-methylpropane sulfonic acid; β-CD, β-cyclodextrin; RST, rosuvastatin.

of RST calcium were analyzed after the administration of hydrogel microparticles and commercially available tablets of RST calcium. Non-compartmental pharmacokinetic analysis was performed using software package kinetica v 4.4. Peak plasma concentration (C_{max}) and time required to reach peak plasma concentration (T_{max}) were calculated from the visual inspection of plasma concentration–time curves. The area under the plasma concentration–time curve (AUC_{0-t}) was calculated by using trapezoidal rule.

Results of mean plasma concentration after the administration of hydrogel microparticles and commercially available tablets of RST calcium was calculated. Mean values of different pharmacokinetic parameters of RST calcium after oral administration of hydrogel microparticles and

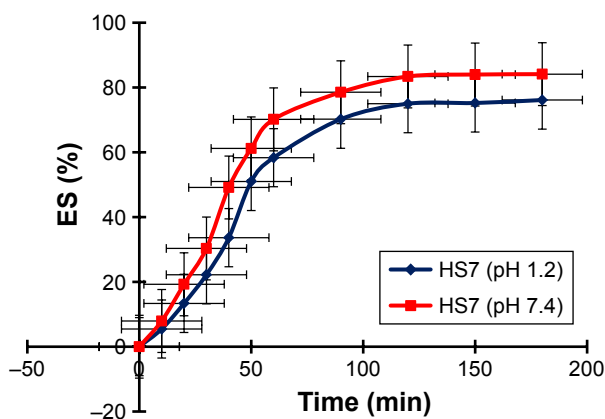


Figure 10 ES% at pH 1.2 and pH 7.4 (HS7).
Abbreviation: ES%, equilibrium swelling percent.

Table 3 Mean values of pharmacokinetic parameters

Parameters	Hydrogel microparticles	Commercial tablet
C_{max} (ng/mL)	90.543	41.263
T_{max} (h)	2.00	4.00
AUC_{0-t} (ng/mL-h)	480.09	366.79
$AUC_{0-\infty}$ (ng/mL-h)	567.26	1,200.093789
$AUMC_{0-\infty}$	3,728.439942	35,029.79044
MRT (h)	6.57	29.18
C_{last} (ng/mL)	10.385	28.728
T_{last} (h)	4.00	12.00
$t_{1/2}$ (h)	4.20	18.80
V_z (µg/mL)	8.82581294	18.64547677
Cl (µg/mL/h)	1.454359553	0.687446271

Notes: C_{max} , maximum concentration; T_{max} , maximum time; $t_{1/2}$, half-life; V_z , volume of distribution.

Abbreviations: AUC, area under the curve; AUMC, area under the first moment curve; Cl, clearance; MRT, mean residence time.

commercially available tablets of RST calcium are given in Table 3. Pharmacokinetic data were statistically analyzed using one-way ANOVA.

Pharmacokinetic studies also proved the rapid release of drug from hydrogel microparticles compared to commercially available tablets of RST. Values of C_{max} were 90 and 41 ng/mL for hydrogel microparticles and commercially available tablets of RST calcium, respectively, as shown in Figure 11.

Discussion

EE is a vital parameter that is used to determine the efficiency of a prepared carrier system to evaluate a suitable dosage regimen. It was noticed that with the increase in polymeric content %EE was increased from 83.50% to 88.50%, not up to 100%, because RST transport from the drug solution (1% RST solution, ie, 1 g in buffer solution up to volume 100 mL)

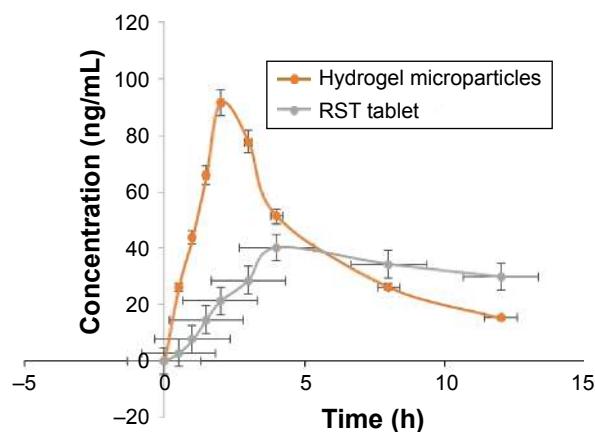


Figure 11 Combined plasma profile of RST calcium after the administration of hydrogel microparticles and commercially available RST tablets.
Abbreviation: RST, rosuvastatin.

was diffusion assisted and during washing of the hydrogel microparticles surface RST was removed. Similarly, product yield was also higher in this formulation due to high polymeric content. High polymeric content offer more channeling and hydrophilic character to developed carrier system leading to more and more penetration of the RST. Our results have justified that the prepared carrier system was suitable for the loading of RST and its accurate delivery at the target site.

FTIR spectra of each participant of hydrogel microparticle formulation (HS1–HS9) were recorded to investigate any possible interaction in formulation component. FTIR spectrum of β -CD-*co*-poly(AMPS) cross-linked hydrogel microparticles was entirely different from that of pure drug, ie, RST and polymer, ie, β -CD. It was evident from the IR spectra of hydrogel microparticles that shifting of characteristic peaks was from 3,337.90 and 2,968.23 cm^{-1} to 3,306.87 and 2,930.97 cm^{-1} , respectively. The peak present at 1,438.03 cm^{-1} was shifted toward higher values of 1,655.91. Shifting, reduction in intensity and broadening of peaks confirmed that there was a new polymeric complex formation. Riekes et al³⁴ prepared complexes of β -CD with amiodarone for solubility and dissolution improvements. They studied FTIR spectra of individual ingredients. β -CD bands were present in the physical mixture while in the case of complexes these bands were absent thus indicating complex formation as proved in our study.

Thermal analysis was performed to check the stability of β -CD-*co*-poly(AMPS) hydrogel microparticles. In the case of β -CD-*co*-poly(AMPS) hydrogel microparticle thermogram, shifting of endothermic peaks to higher temperatures confirmed that polymer and monomer individual ingredients are cross-linked and a stable hydrogel network in the form of microparticles has been developed successfully. DSC and TGA thermograms have different patterns of thermal stability of β -CD-*co*-poly(AMPS) hydrogel microparticles. Cross-linking improves thermal stability as well as provides excellent delivery ability for various drugs. Our results are in good agreement with a previously conducted study by Chen et al,³⁵ where they prepared chitosan-based hydrogel beads and concluded that disappearance or shifting of peaks is the indication of complex formation and drugs become more soluble when they form a complex with other hydrophilic moieties.

In SEM photomicrographs, the presence of white spots confirmed that RST was successfully distributed and loaded within the microparticles. Microparticles had porous surfaces that assisted rapid penetration of water and high swelling and ultimately promoted rapid faster release. Borba et al³⁶

prepared microparticles of less water-soluble drug telmisartan. They analyzed morphology and particle size of microparticles. Their results showed that developed microparticles had irregular shapes and variable sizes but in the micrometric range they offered large surface area, fast wetting and, ultimately, more solubility as observed in our SEM results.

X-ray diffraction studies were conducted to confirm the nature of ingredients as well as fabricated crosslinked networks. Presence of peaks in XRD diffractograms of drug and polymer proved crystalline nature due to which these both have solubility issues. While, in the diffractogram of hydrogel microparticles, these characteristic peaks were markedly reduced and fused with each other to ensure the conversion of the crystalline form into the amorphous form with better solubility profile. Thus, the solubility of RST was enhanced in hydrogel microparticles, because amorphous forms are more soluble than crystalline. Similarly, results have been proved in a work conducted by Zhang et al.³⁷ They proved the loss of crystalline nature of berberine hydrochloride when processed with β -CD.

TEM studies revealed that microparticles had porous surface that facilitated rapid uptake of media and swelling and release of the drug from the developed carrier. White appearances confirmed successful loading and distribution of RST. Small size and large surface area lead to dissolution enhancement of RST. These findings agree with the findings of Sinha et al.³⁸

All the prepared hydrogel microparticles had size within the micrometric size range that was desired in this work. Zeta potential values near to -40 mV usually confirm the stability of a particulate system. All the formulations (HS1–HS9) had zeta potential values near to -40 mV, ie, -37 mV confirmed the stability of the developed hydrogel microparticles. Negative charge imparts repulsion among microparticles and resists aggregation of microparticles.³⁹

Maximum release (92%) was observed at pH 6.8 by formulation HS7. This was due to the hydrophilic nature of β -CD, micrometric size range and porous nature confirmed from SEM studies and AMPS contents. As the concentration of AMPS increases, it offers more channeling and promotes release of drug. Various concentrations of AMPS and MBA were studied to optimize the best concentration to enhance dissolution of RST. When 4 g/100 g of AMPS was used, it produced the best release rate as compared to 2, 5 and 6 g/100 g of formulation. Similarly, MBA produced the best results when 0.4 g/100 g was used. The rate and percent release of RST from the cross-linked system is dependent on swelling behavior, composition of copolymeric system, environmental conditions and pH of dissolution medium. A

study conducted by Chen et al supported our findings that β -CD enhances dissolution profile of BCS-II drugs by complex formation.⁴⁰ Kinetic models were applied on release data to determine the best fit model. Drug release followed first-order kinetics in the case of hydrogel microparticles while RST tablets followed zero-order kinetics, as given in Table 4. Formulations HS4 and HS7 followed anomalous diffusion of RST, which means that dual mode of drug release was diffusion and erosion based. This was due to the presence of β -CD hydrogel microparticles that enhanced the release of the drug from the carrier system.

RST calcium is a weak acid. Due to its acidic nature, it has more solubility in basic media as compared to acidic pH. So, it becomes ionized at higher pH values and therefore more soluble in solutions having high pH. However, AMPS gives more swelling at higher pH values that have also contributed to more and more uptake of solvent. Moreover, solubility in phosphate buffer solution (pH 6.8) was due to the weak acidic nature of RST. These findings confirmed that hydrophilic complexation of RST with β -CD containing hydrogel microparticles potentiated the solubility of RST. The order in which RST had more solubility was distilled water > phosphate buffer of pH 6.8 > HCl buffer of pH 1.2.

All prepared formulations exhibited good swelling behavior. AMPS is a hydrophilic monomer that possesses both ionic and non-ionic functional groups. As the number of ionizable groups ($-\text{SO}_3^-$) is enhanced, swelling capacity increases due to the availability of ionizable groups. Previously, Elliott et al also observed the same kind of swelling behavior from AMPS and concluded that AMPS had pH-independent swelling.³⁹ In another work, conducted by Atta,⁴¹ similar findings for AMPS-based hydrogels were

seen. Polymeric complexes that have higher AMPS content offer more swelling.

From in vivo studies, it was confirmed that there was rapid release of drug from prepared formulations that ultimately improved the bioavailability of the drug in rabbit plasma. Similarly, AUC_{0-24} was also higher for prepared formulations of hydrogel microparticles. This means that a larger amount of drug was released from prepared formulations due to complex formation with β -CD and the drug became more soluble in gastric media. Results of C_{max} and AUC_{0-24} were significant ($p < 0.05$). Hydrogel microparticles have a value of T_{max} 2 h while RST tablets have 4 h which meant that prepared formulations released RST more rapidly as compared to RST tablets. This rapid release of drug was due to more solubility. Elimination half-life ($t_{1/2}$) of hydrogel microparticles was 4.20 h ($p < 0.05$), while commercially available RST tablets had a half-life of 18.80 h. This confirmed that more rapid bioavailability was achieved from hydrogel microparticles as compared to commercial tablets. These results confirmed that the solubility of the drug increased in prepared formulations due to complexation with the carrier system that ultimately enhanced its bioavailability.⁴²

Conclusion

In the current study, β -CD-*g*-poly(AMPS) hydrogel microparticles with different ratios of polymer, monomer and cross-linking agent were successfully prepared and tuned for solubility enhancement of RST calcium. Prominent rise in solubility of RST was observed at pH 6.8. RST release behavior from β -CD-*g*-poly(AMPS) hydrogel microparticles was observed and found swelling-dependent release. Slightly higher swelling was seen at pH 7.4. Pharmacokinetic parameters also confirmed rapid release and excellent bioavailability offered by this developed polymeric carrier system. This system can be utilized for solubility and bioavailability enhancement of other hydrophobic drugs.

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Disclosure

The authors report no conflicts of interest in this work.

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Table 4 In vitro drug release kinetics of hydrogel microparticles

Models	Parameters	HS4	HS7	RST tablet
Zero-order kinetics	K_0	0.720	0.737	0.259
	R^2	0.7769	0.7990	0.9573
	T_{25}	34.726	33.935	89.438
	T_{50}	69.452	67.870	192.875
	T_{75}	104.178	101.805	289.313
First-order kinetics	K_1	0.025	0.025	0.003
	R^2	0.9988	0.9884	0.9682
	T_{25}	11.684	11.389	89.357
	T_{50}	28.153	27.442	215.299
	T_{75}	56.305	54.883	430.598
Higuchi	K_H	8.286	8.459	2.744
	R^2	0.8957	0.9131	2.744
Korsmeyer–Peppas	K_{kp}	14.148	14.154	0.600
	R^2	0.9108	0.9711	0.9978
	N	0.383	0.9265	0.827

Abbreviation: RST, rosuvastatin.

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