

# Developmental Programming of Energy Balance and Its Hypothalamic Regulation

Floor Remmers and Henriette A. Delemarre-van de Waal

Institute of Physiological Chemistry (F.R.), University Medical Center of the Johannes Gutenberg University Mainz, 55128 Mainz, Germany; and Department of Pediatrics (H.A.D.-v.d.W.), Leiden University Medical Center, 2300 RC Leiden, The Netherlands

Developmental programming is an important physiological process that allows different phenotypes to originate from a single genotype. Through plasticity in early life, the developing organism can adopt a phenotype (within the limits of its genetic background) that is best suited to its expected environment. In humans, together with the relative irreversibility of the phenomenon, the low predictive value of the fetal environment for later conditions in affluent countries makes it a potential contributor to the obesity epidemic of recent decades. Here, we review the current evidence for developmental programming of energy balance. For a proper understanding of the subject, knowledge about energy balance is indispensable. Therefore, we first present an overview of the major hypothalamic routes through which energy balance is regulated and their ontogeny. With this background, we then turn to the available evidence for programming of energy balance by the early nutritional environment, in both man and rodent models. A wealth of studies suggest that energy balance can indeed be permanently affected by the early-life environment. However, the direction of the effects of programming appears to vary considerably, both between and within different animal models. Because of these inconsistencies, a comprehensive picture is still elusive. More standardization between studies seems essential to reach veritable conclusions about the role of developmental programming in adult energy balance and obesity. (*Endocrine Reviews* 32: 272–311, 2011)

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## I. Introduction

The concepts of “nutritional programming,” “fetal programming,” “fetal origins of adult disease,” “developmental origins of health and disease,” “developmental

induction,” and “developmental programming” (1–5) were all conceived to explain the same phenomenon: a detrimental environment during a critical period of development has persistent effects, whereas the same environmental stimulus outside that critical period induces only reversible changes. Many epidemiological studies have shown an association between low birth weight and an elevated risk of developing several chronic diseases in adulthood (reviewed in Refs. 2 and 6–9). The Dutch famine, a unique “natural experiment” with a well-defined period of food shortage in an otherwise well-nourished population, has shown that maternal undernutrition during gestation compromises health in later life and that these long-term effects depend on its timing during gestation (10). It is assumed that low-birth-weight infants, who are not small *per se* but rather are small for gestational age (SGA), suffered from intrauterine growth restriction (IUGR) due to a low availability of nutrients. As adults, these subjects have an increased risk for insulin resistance, hypertension, and cardiovascular disease, collectively

Abbreviations: AgRP, Agouti-related protein; ARC, arcuate nucleus; BMI, body mass index; CART, cocaine- and amphetamine-regulated transcript; DMN, dorsomedial nucleus; HPA axis, hypothalamic-pituitary-adrenal axis; HPT axis, hypothalamic-pituitary-thyroid axis; IUGR, intrauterine growth restriction; LHA, lateral hypothalamic area; MC, melanocortin; MCH, melanin-concentrating hormone;  $\alpha$ -MSH,  $\alpha$ -melanocyte-stimulating hormone; Ob-R, leptin receptor; ORX, orexin; NPY, neuropeptide Y; POMC, proopiomelanocortin; PVN, paraventricular nucleus; SGA, small for gestational age; VMN, ventromedial nucleus;  $Y_1$ R, NPY receptor.

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called the “metabolic syndrome” or “syndrome X” (11). How early malnutrition should lead to conditions normally related with affluent environments has been the subject of much debate. It is now believed that adaptations that helped these IUGR or SGA infants survive during pregnancy may become detrimental in later life when nutrients are no longer scarce. It is said that these individuals are “programmed” for the metabolic syndrome. Whether the obesity that is part of the metabolic syndrome is also programmed has long been a matter of debate (12–15). In addition to a low birth weight, high birth weight (*e.g.*, through maternal obesity or maternal diabetes) is also associated with an increased risk of developing obesity in later life (13, 16–19). In this review, developmental programming will be discussed, followed by energy balance, its regulation, its normal development, and what is known about its programming.

## II. Developmental Programming

The concept of developmental programming implies that characteristics of the environment encountered during early development can permanently alter physiology in later life (20). The perinatal level of nutrition has been proposed to be a particularly important feature (21). In early development, there is a window of plasticity, a period in which the organism can still develop in different directions. During this period, the developing organism has a large potential to adapt to its environment. Once the window of plasticity has closed, many of these adaptations will become fixed. Although epidemiological studies have mainly concentrated on the detrimental consequences of programming, it is not principally a harmful phenomenon. Being able to adjust your phenotype to the environment encountered in early life can be “evolutionary” adaptive if the environment is relatively stable (22, 23). A mismatch between the environment in early life and adult life may result in inappropriate adaptations in the organism.

This is thought to be the case for SGA infants in developed countries. The low nutrient availability during intrauterine development that causes these babies’ growth restriction is usually not due to a low maternal energy intake, but instead is due to other causes such as placental insufficiency (24), drug use (including caffeine, alcohol, and smoking), and stress or illness (25), and hence does not give an adequate prediction of nutrient availability in postnatal life. SGA subjects that develop the metabolic syndrome in later life are a good example of “developmental programming gone bad” (2, 26). In contrast, in infants born to obese mothers, the enhanced availability of nutrients to the fetus is thought to allow increased fetal adipogenesis and simultaneously alter the systems that reg-

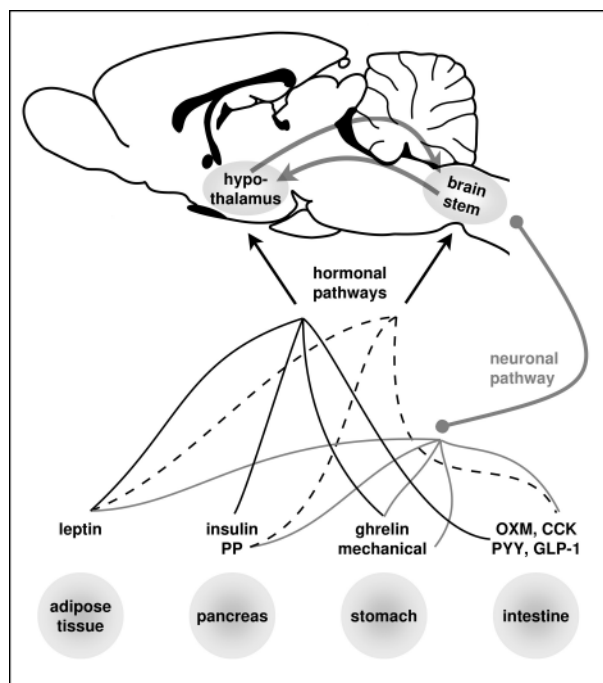
ulate energy balance while they are still plastic (26, 27). It is now well established that the perinatal environment can program similar changes in experimental animals (reviewed in Refs. 28–31).

## III. Energy Balance Regulation

Energy homeostasis, or the process whereby stable energy reserves are maintained over long periods of time, is tightly regulated. To maintain neutral energy balance, energy intake, thermogenesis, and activity need to be regulated. These components of energy balance are regulated by at least two separate, but interrelated systems: 1) a short-term system that controls the initiation and termination of meals depending on the contents of the gastrointestinal tract; and 2) a long-term system that defends the stability of the energy reserves and thereby that of body weight (32). In addition, higher brain functions, such as motivation and reward, as well as environmental factors such as social influences and food availability alter our food intake and activity levels (33, 34). It is the task of the short-term and long-term regulatory systems to balance the energy reserves in the face of a changing environment. Manipulations of the gastrointestinal peptides involved in the short-term regulation of hunger and satiety were mostly shown to have little effect on food intake and body weight over a longer period, and therefore we rely predominantly on the long-term system to maintain neutral energy balance.

The first indications that an important part of the long-term regulatory system resides in the hypothalamus of the brain came from early studies reporting severe anorexia or obesity after lesions of distinct areas of the hypothalamus (35, 36). The hypothalamus consists of several distinct nuclei that produce specific neuropeptides and perform different tasks in the homeostasis of temperature, water, energy, sleep, reproduction, and other functions. In the brain, there are extensive connections to, within, and from the hypothalamus (37). The regulation of energy balance by the hypothalamus is a complicated process, and the following explanation is a simplified summary. It is also important to keep in mind that in addition to energy balance, these nuclei and peptides are involved in other hypothalamic functions as well. Although the majority of the literature cited concerns research in rodents, most of this discussion is also applicable to humans (38–43).

In short, neurons in the mediobasally located arcuate nucleus (ARC) receive information about the status of the energy reserves (*e.g.*, adipose tissue) through peripheral hormones that circulate in amounts related to body fat stores; the information is integrated and passed on to several other hypothalamic nuclei, including the paraven-



**FIG. 1.** A simplified overview of the regulation of energy balance. Peripheral signals of energy reserves reach the hypothalamus and brainstem via hormonal and neuronal pathways. The former pathway is more important in transferring long-term adiposity signals, whereas the latter handles the rapid transmission of short-term satiety signals, mainly through the vagus nerve. Several brain areas, including the hypothalamus and brainstem, then interact with each other to regulate intake and expenditure of energy. This review will focus on the interaction between long-term adiposity signals and the hypothalamus. See the text for further information. CCK, Cholecystokinin; GLP-1, glucagon-like peptide 1; OXM, oxyntomodulin; PP, pancreatic polypeptide; PYY, peptide YY. [Adapted from G. Paxinos and C. Watson: *The rat brain in stereotaxic coordinates*, Academic Press, New York, 1997 (599). © Elsevier 1997].

tricular nucleus (PVN) and the lateral hypothalamic area (LHA), and from there on to output functions. The process is also influenced by satiety hormones from the gastrointestinal tract (44) (Fig. 1).

### A. Peripheral signals

Several hormones provide the brain with information on the status of energy balance. These can be divided into two categories: 1) hormones that are produced by the gastrointestinal tract and signal on satiety and hunger, or short-term information; and 2) hormones that signal the status of fat reserves of the body, the long-term signals. Leptin is the major peripheral hormone involved in long-term energy homeostasis.

#### 1. Leptin

Leptin was first identified as the product of the gene that is defective in obese *ob/ob* mice and was named after “leptos,” the Greek word for “thin” (45, 46). In both humans and rodents, leptin is produced by adipose tissue, in proportion to the body fat content, as an “adiposity

signal” (47–52). The leptin receptor (Ob-R) Ob-Rb is highly expressed in the ARC and is also found in some other hypothalamic areas (53–55). In addition, short forms of the Ob-R (a, c–f), which probably act as leptin transporters, exist in the choroid plexus and other areas where substances may cross the blood-brain barrier (53–56).

Leptin gene expression and serum levels decrease upon negative energy balance in both humans and rodents (48, 51, 52, 57–59). However, its levels are disproportionately reduced during acute depletion (60) to initiate immediate action to restore these reserves. The central effects of leptin include reductions in food intake and body weight gain and increased energy expenditure (61–63).

### 2. Other peripheral signals

Insulin, produced by the  $\beta$ -cells of the pancreas, controls blood glucose availability. In energy balance regulation, it acts both as a short-term and as a long-term signal (47). It is released upon acute changes in energy levels such as meal ingestion, but its circulating levels are also directly correlated with fat reserves. In the brain, it acts on receptors in the ARC to reduce food intake and increase energy expenditure (reviewed in Ref. 47).

Ghrelin, produced by the stomach, indicates negative energy balance (64). Its levels rise before meal onset and decrease with feeding. Over longer time periods, ghrelin levels are inversely correlated to energy stores. Ghrelin acts on receptors in the hypothalamus to influence ARC neuron activity. Upon peripheral or central injection, ghrelin stimulates food intake and decreases energy expenditure (reviewed in 64).

Other peripheral signals, which are predominantly involved in the short-term regulation of the initiation and termination of meals, are produced in different regions of the gastrointestinal tract and include cholecystokinin, glucagon-like peptide 1, oxyntomodulin, pancreatic polypeptide, and peptide YY (65, 66). These are mainly secreted after eating and inhibit further food intake. Their actions on food intake are exerted via the vagus nerve, the brainstem, and the hypothalamus. It has also been shown that several of these peripheral signals interact with each other to regulate food intake. Another important function of these gut peptides is to control the proper processing of the nutrients ingested in a meal (reviewed in Refs. 65 and 66).

### B. Peptides from the arcuate nucleus

The ARC is located mediobasally in the hypothalamus, close to the median eminence. This is a circumventricular organ where the blood-brain barrier is incomplete and blood-borne signals can easily reach the ARC neurons (67, 68). In addition, leptin, insulin, and ghrelin are actively

transported across the blood-brain barrier (69–71). Apart from these direct inputs from the periphery, the hypothalamus also receives information concerning energy balance from brainstem areas (66). The ARC integrates this information and drives other hypothalamic areas such as the PVN and the LHA (44).

The ARC contains two populations of neurons that are strongly involved in the regulation of energy balance. These two populations express different neuropeptides. A medial population coexpresses the orexigenic peptides neuropeptide Y (NPY) and agouti-related protein (AgRP) (72–75). The anorexigenic peptides  $\alpha$ -melanocyte-stimulating hormone ( $\alpha$ -MSH) and cocaine- and amphetamine-regulated transcript (CART) are produced by a more lateral population of ARC neurons (74, 76, 77). Both of these populations coexpress receptors for peripheral signals, including those for leptin (40, 78, 79), insulin (80, 81), and ghrelin (82, 83). Both the NPY/AgRP and the proopiomelanocortin (POMC)/CART neurons project widely throughout the hypothalamus and the brain (39, 72, 84, 85). Besides the ARC, CART is also expressed in several other hypothalamic nuclei, including the PVN and LHA (86).

These peptides exert their effects via various receptors. In the rat, four subtypes of the NPY receptor have been identified that have all been found in many different brain areas, including most hypothalamic nuclei (87, 88). Within the ARC, Y<sub>1</sub>R and Y<sub>5</sub>R mRNA and protein are found in many  $\alpha$ -MSH/CART neurons (89, 90), whereas the mRNA and protein for Y<sub>2</sub>R, which is believed to be an autoreceptor (91), was found in most NPY/AgRP neurons (89, 90). Of the five identified melanocortin (MC) receptors, the MC3 and MC4 receptors mediate the effects of  $\alpha$ -MSH in the regulation of energy balance (92–95), although some debate previously existed over the involvement of the MC3 receptor (93). Both are highly expressed in hypothalamic nuclei, but MC4 is more widely expressed throughout the brain (96, 97). AgRP is an inverse agonist of the constitutively active MC receptors (98, 99). A CART receptor has not been identified yet (100), but specific CART binding has been reported in cultured cells from the hypothalamus, hippocampus, and nucleus accumbens (101).

### 1. Neuropeptide Y

NPY is one of the most abundant peptides in the brain (84). Negative energy balance, as elicited by fasting, has been shown to increase NPY peptide and expression levels (102–106). Leptin injections, mimicking positive energy balance, have been reported to lower NPY expression (62, 102, 106, 107), as well as the activity of NPY neurons (82), NPY secretion by the hypothalamus (107), and NPY levels in the PVN (108). In the hypothalamus, NPY stimulates

food intake and body weight gain (109–111), increases white fat lipid storage, and reduces brown fat thermogenesis (112, 113). In contrast, NPY injections in most areas outside the hypothalamus did not have any effect on food intake (109).

### 2. Agouti-related protein

AgRP was discovered because of its resemblance to agouti, which in mice causes severe obesity when overexpressed (114, 115). As with NPY, levels of AgRP expression, peptide, and activity are increased by fasting and decreased by leptin (103, 106, 116–118). Administration of AgRP or other antagonists of MC receptors were shown to elevate food intake, body weight, and body fat and to reduce energy expenditure and brown fat thermogenesis (119–121). In contrast to the relatively short-lived effects of NPY, a single injection of AgRP will increase food intake for up to 1 wk (122, 123). These long-lasting effects of AgRP are proposed to be mediated by other routes than the MC receptors (122, 124).

### 3. $\alpha$ -Melanocyte-stimulating hormone

$\alpha$ -MSH is cleaved from the precursor polypeptide POMC, together with other peptides like  $\beta$ -endorphin and ACTH (125). POMC gene expression is reduced by fasting and stimulated by leptin (102–104, 106, 126, 127). In addition, leptin stimulates activity of POMC neurons and  $\alpha$ -MSH release (76, 128, 129). Central administration of  $\alpha$ -MSH or its agonist melanotan II decreases food intake, weight gain, and adiposity and increases energy expenditure, brown adipose tissue activity, and body temperature, but not locomotor activity (119, 120, 130–133). Within the hypothalamus, it reduced food intake when injected in the ARC, PVN, and LHA among others (134). In contrast to  $\alpha$ -MSH,  $\beta$ -endorphin has been shown to increase food intake (133).

### 4. Cocaine-and amphetamine-regulated transcript

CART was identified when its expression levels were shown to be increased after administration of cocaine or amphetamine (135), although it was sequenced as a peptide with unknown function long before that (136). ARC CART mRNA levels are decreased by fasting and increased by leptin (102, 103, 137, 138). In line with an anorexigenic role, intracerebroventricular CART injections decreased food intake (137, 139, 140), and intrahypothalamic injection increased gene expression of the thermogenic uncoupling protein-1 in brown adipose tissue (141, 142). Chronic injections also reduced body weight gain and increased lipid oxidation (143, 144). Injections into distinct hypothalamic nuclei, however, have produced either increased or decreased food intake (139–142). These contradictory results and colocalization with



both orexigenic and anorexigenic neuropeptides have been interpreted to suggest that CART may play a modulatory role, with different effects depending on its localization (100).

### C. Anorexigenic peptides from the paraventricular nucleus

The PVN expresses the anorexigenic peptides CRH and TRH in two distinct populations of parvocellular neurons (145, 146). A subpopulation of the latter coexpresses CART (40, 86). The PVN receives innervation from ARC NPY/AgRP, POMC, and CART terminals (85, 137, 147). In addition, receptors for these peptides are expressed in the PVN (88, 96), providing all the “machinery” for signaling from the ARC to the PVN. The PVN also receives some input from the LHA, dense projections from the dorsomedial nucleus (DMN), and indirect input from the amygdala (37).

Both peptides exert their effects via two receptors: the CRH<sub>1</sub>- and CRH<sub>2</sub>-receptors (148, 149) and the TRH<sub>1</sub>- and TRH<sub>2</sub>-receptors (150–152), respectively. CRH<sub>1</sub> (153, 154) and TRH<sub>2</sub> (155) are both widely expressed in different brain areas, whereas the expression of CRH<sub>2</sub> (156, 157) and TRH<sub>1</sub> (155) is mostly restricted to the hypothalamus.

#### 1. Corticotropin-releasing hormone

CRH, known for its role in the hypothalamic-pituitary-adrenal (HPA) or stress axis, is also involved in the regulation of energy balance (158). PVN CRH neurons project to the median eminence, where CRH regulates the release of ACTH and  $\beta$ -endorphin from the pituitary (146, 159), and to some cell groups in the brainstem and spinal cord (160).

PVN CRH expression and peptide levels are decreased by food deprivation and increased by leptin (62, 104, 127, 161, 162). Furthermore, CRH expression is increased by injections of both  $\alpha$ -MSH and CART (163, 164). Intracerebroventricular CRH has been shown to decrease food intake and body weight gain and to induce both locomotor activity and activity of brown adipose tissue (165–168). These data all point toward an anorexigenic and catabolic role for CRH.

#### 2. TSH-releasing hormone

TRH stimulates TSH release from the pituitary. TRH terminals are found throughout the hypothalamus, in the median eminence, and the pituitary (145). Via thyroid hormone, which stimulates energy expenditure and thermogenesis, this hypothalamic-pituitary-thyroid (HPT) axis plays an important role in energy homeostasis (169). TRH expression and release are reduced by fasting, NPY, and AgRP, and increased by leptin,  $\alpha$ -MSH, and CART

(129, 170–174). Central and peripheral injections of TRH decrease food intake and increase body temperature (175–177). Despite increased food intake (possibly a compensatory response), chronic oral TRH causes a reduction in body weight (178). These data point toward an anorexigenic and catabolic role for TRH.

### D. Orexigenic peptides from the lateral hypothalamic area

The LHA was recognized early on as a “feeding center” (35, 36). Two distinct cell populations express the orexigenic peptides melanin-concentrating hormone (MCH) and the orexins (ORXs), the latter especially in the perifornical area (179). Some MCH cells also contain CART (40, 86). The LHA is innervated by nerve terminals containing NPY, AgRP, and  $\alpha$ -MSH (39, 179) and expresses receptors for these peptides (88, 96, 97). Apart from this input from the ARC, the LHA also receives input from the hypothalamic PVN and DMN and from some higher brain areas including anterior limbic cortical areas, the nucleus accumbens, and indirectly from the hippocampus (37). From the cell bodies in the LHA, MCH and ORX fibers project throughout the hypothalamus and to many different brain areas (180–182). MCH has been identified as the ligand for an orphan receptor by several groups simultaneously (183–187). Humans, but not rodents, have an additional MCH<sub>2</sub>-receptor (188). The distribution of MCH<sub>1</sub> and the ORX<sub>1</sub>- and ORX<sub>2</sub>-receptors corresponds well with that of MCH (189, 190) and ORX (191, 192) fibers, respectively.

#### 1. Melanin-concentrating hormone

MCH was first discovered in fish, as the peptide that causes melanosomes to contract and thereby has a skin-lightening effect (193). Thereafter, a similar peptide was found in rat brain (194), and the homologous peptide was identified (195). MCH expression was either decreased (as would be expected for an orexigenic peptide) or increased by injections of leptin (196, 197). Fasting has been shown to increase MCH expression (103, 197–199), but MCH neurons appear to be inhibited by NPY (200). Most studies have reported increased food intake after MCH injections (199, 201–203), although the opposite has also been reported (198). When injected chronically, the orexigenic effect of MCH faded, and body weight was not affected (204). Mice that lack MCH or its receptor show hyperactivity (205, 206). This suggests that MCH suppresses activity, in accordance with an orexigenic, anabolic role.

#### 2. The orexins

ORX-A (or hypocretin 1) and ORX-B (hypocretin 2) were discovered and named simultaneously by two groups (207, 208). Negative energy balance has been shown to

increase LHA ORX expression and peptide levels (208–211), although unchanged expression levels were also reported (103, 197, 212). Leptin lowers LHA ORX expression and ORX-A levels (210, 213, 214). Although AgRP injections were shown to increase activity of ORX neurons (215), NPY either decreased or failed to affect their activity in two different paradigms (216, 217). Nevertheless, Y<sub>1</sub>R and Y<sub>5</sub>R antagonists suppress ORX-induced feeding, which is again more in line with the orexigenic role of the ORXs. Central ORX injections stimulate feeding (208, 218–220), supporting an orexigenic role. However, not all ORX aspects fit the profile for a truly anabolic protein. The results for ORX-B have been less consistent, and the outcome generally depends on the time of day (221). Furthermore, metabolic rate and activity seem to be stimulated rather than decreased by ORX (222–225), and continuous infusion of ORX altered the circadian rhythm of feeding without affecting total food intake or body weight (219, 226). Apart from its complicated role in the regulation of energy balance, ORX is also involved in the regulation of arousal and vigilance, and its primary function in energy balance may be to synchronize feeding behavior with other essential behaviors and the environment (227–229).

#### E. Downstream events

There are basically three output pathways through which hypothalamic signaling can eventually alter the intake and expenditure of energy. The first pathway influences behavior through integration of signals from many brain areas and, ultimately, the activation of motor neurons (33). A final effect of increased hypothalamic orexigenic activity may be the initiation of a meal. The second pathway, the neuroendocrine route, influences energy balance through the secretion of hormones. The HPA and HPT axes are part of this pathway. In the HPA axis, CRH from the PVN stimulates the release of ACTH from the pituitary, which in turn induces glucocorticoid release from the adrenal glands (37). These can indirectly influence feeding behavior and energy expenditure (reviewed in Ref. 230). In the HPT axis, TRH from the PVN, via the release of TSH from the pituitary, induces the release of thyroid hormone by the thyroid gland, which stimulates energy expenditure and thermogenesis (reviewed in Ref. 169). The third pathway is via the autonomic nervous system. Several hypothalamic nuclei, especially the PVN, innervate neurons in the brainstem and the spinal cord that are part of the autonomic nervous system, both sympathetic and parasympathetic (37). Via these sympathetic and parasympathetic pathways, energy expenditure can be regulated, for example by influencing the heart rate and thermogenesis by adipose tissue and skeletal muscle (reviewed in Refs. 32, 231, and 232).

#### F. Mutual connections

When reflecting on the regulation of energy balance, it is important to bear in mind that the pathways and processes described above are much simplified. The main route for information is from the periphery to the ARC and in turn via the PVN or LHA to the output systems (Fig. 2A). However, other brain areas (hypothalamic and otherwise) are involved in the routing of peripheral signals as well, with ample feedback between the different brain areas. In several respects, the regulation of energy balance is much more complex than the relatively straightforward pathways described above. A few examples are given below.

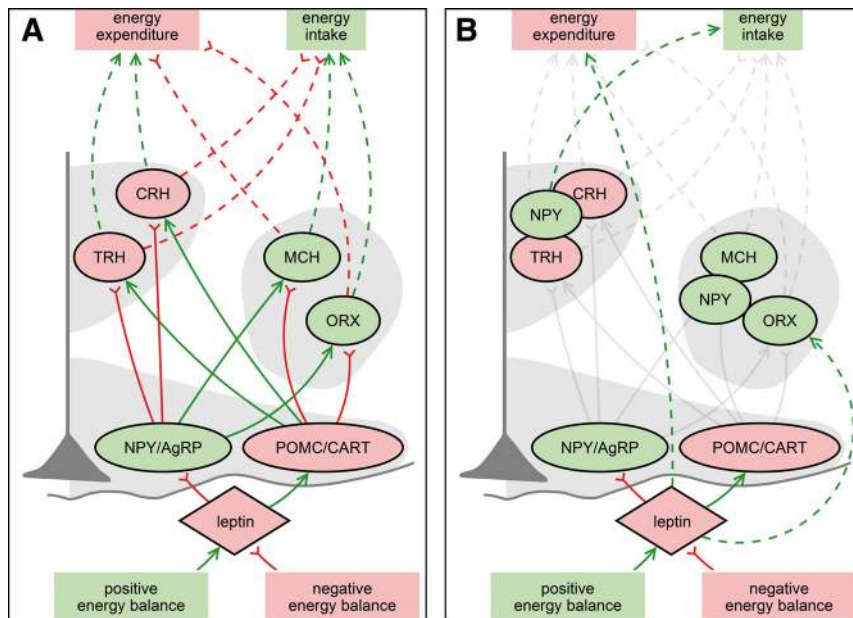
First, the different populations of cells in the ARC influence their own and each other's activity. Both NPY/AgRP and POMC/CART cells express receptors for NPY and MCs, albeit different subtypes: NPY/AgRP neurons express Y<sub>2</sub>R and the MC3 receptor, whereas POMC/CART neurons express Y<sub>1</sub>R, Y<sub>5</sub>R, and the MC4 receptor (89, 90, 233). Through these receptors, NPY inhibits the POMC/CART cells (128, 234); NPY and AgRP stimulate each other's release and (at least *in vitro*) can be stimulated by  $\alpha$ -MSH and CART (235); whereas CART reduces  $\alpha$ -MSH release (140).

Second, the PVN and LHA may also receive peripheral input directly, through leptin receptors in these nuclei, although these receptors are not necessarily colocalized with the four peptides of our interest: CRH, TRH, MCH, and ORX (236–239). For example, leptin also influences motivational brain areas directly, via receptors in these areas (reviewed in Refs. 33 and 240).

Third, there is feedback within and between the PVN and LHA. CRH and TRH neurons are contacted by each other's axons (241), as are MCH and ORX neurons (242), and ORX stimulates both ORX and MCH neurons (200, 243). Furthermore, MCH and ORX have been shown to stimulate CRH neurons and release (244, 245), whereas MCH reduces TRH release (246). CRH, in turn, has been shown to activate ORX neurons (247).

Fourth, besides this mutual influence of the PVN and LHA peptides, they also project back onto the ARC neurons. ORX axons, for example, terminate on both NPY and POMC neurons (220, 248). Via these terminals, ORX stimulates NPY neurons and inhibits POMC neurons (82, 220). MCH has been shown to have similar effects on the ARC as ORX (201). Moreover, CRH receptors have been identified in NPY neurons in the ARC (249), and a CRH receptor agonist has been shown to inhibit medial ARC neurons (250).

In addition, as mentioned before, there are many more brain areas involved in the regulation of energy balance than these three hypothalamic nuclei. Among these are the



**FIG. 2.** Simplified diagrams of the hypothalamic regulation of energy balance in adult (A) and juvenile (B) life. A, The main pathway for energy balance regulation is from peripheral input (leptin) to the ARC (with NPY, AgRP, POMC, and CART), via the PVN (CRH and TRH) and LHA (MCH and ORX), to the output systems. B, Energy balance does not appear to be tightly regulated in the neonatal period. The few connections that have been reported to be in place are summarized in this figure. The adult connections are not yet present and/or active and are shown in light gray. Green arrow, Positive effect; red inverted arrow, negative effect; solid line, direct connection; dashed line, indirect connection; green (red) nodes lead to more positive (negative) energy balance when stimulated/active. See the text for details.

ventromedial nucleus (VMN), the DMN, and the medial preoptic area (251–255). Furthermore, the caudal brainstem is not only a relay station between the periphery and the hypothalamus; it is also known to be capable of performing part of the regulation of food intake independently of the forebrain (256). In addition, the higher brain areas that deal with the reward, cognitive, and social aspects of food intake are not only output areas for the hypothalamus, but also send information back to the hypothalamus (33).

Lastly, although the peptides that have been mentioned here do play an important role in the regulation of energy balance, many other substances are involved. Some of these are hypothalamic neuropeptides, such as galanin, galanin-like peptide, malonyl-coenzyme A, neurotensin, and neuromedin U (257–261). Naturally, the classical neurotransmitters, glutamate and  $\gamma$ -aminobutyric acid, are also present and functional in the hypothalamus (262–264). Moreover, the hypothalamic nuclei and peptides discussed here are involved in many other processes besides energy balance. These include the immune system (leptin, the MCs), bone formation and remodeling (leptin, NPY, CART, MCH), blood pressure and cardiovascular regulation (leptin, NPY), kidney function (NPY), reproduction (NPY, MCH), stress (NPY, CART, CRH, MCH), pigmentation and pain sensation (the MCs), reward and

addiction (CART, MCH), anxiety (MCH), and the wake-sleep cycle (MCH and ORX) (100, 158, 227, 265–273).

This section is not meant to be exhaustive, but intends merely to give an impression of the complexity of the regulation of energy balance (summarized in Fig. 3). However, despite the many interconnections, the main pathway is believed to be from the peripheral input to the ARC (with NPY, AgRP, POMC, and CART), via the PVN (CRH and TRH) and LHA (MCH and ORX), to the output systems (as depicted in Fig. 2A). Although at first sight these peptides all seem to fulfill one of two functions (orexigenic or anorexigenic), subtle differences between these peptides are revealed upon closer inspection. We have concentrated on evidence from rodents.

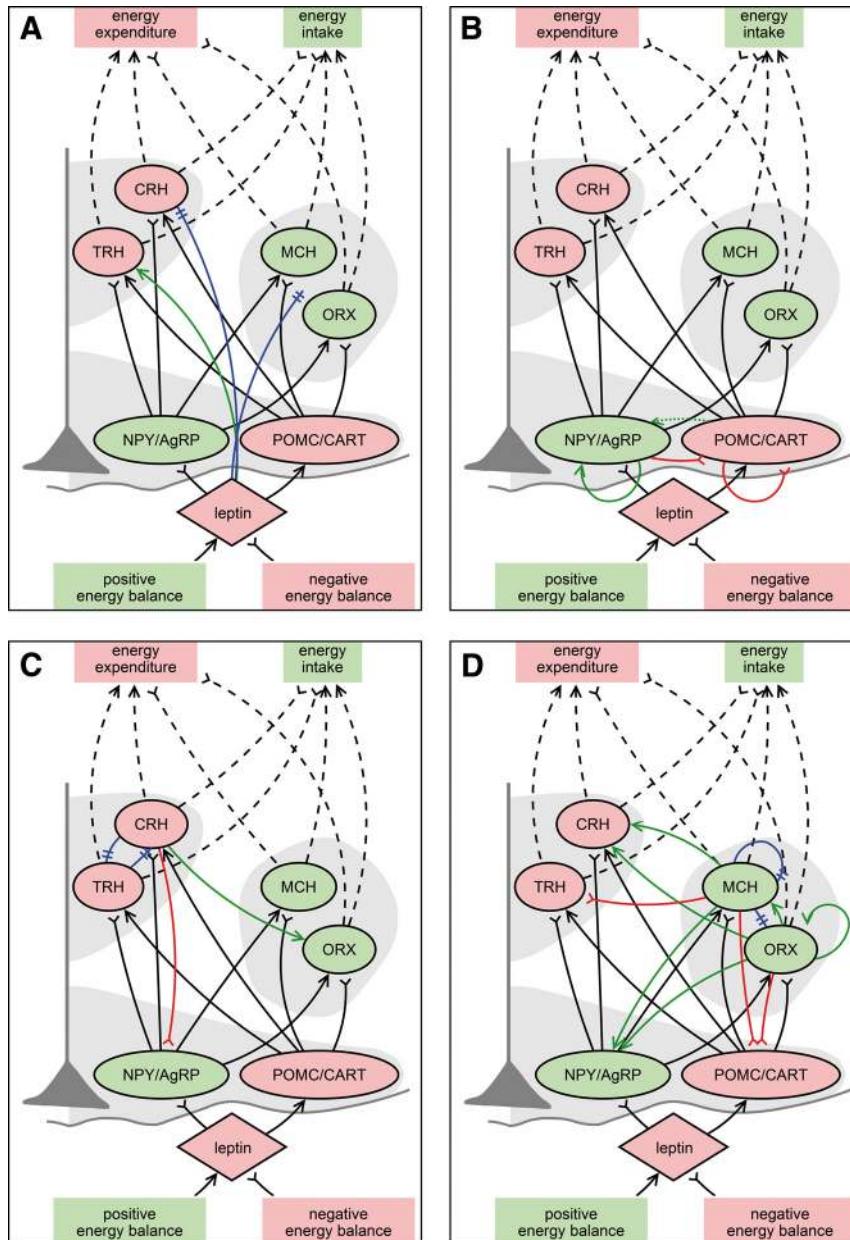
Although some variation in the details exists (38, 274), the regulation of energy balance is very similar in different animal species, including humans.

**G. Ontogeny**

To program a certain system or function, an environmental stimulus must occur during a period in development when the system or function is still plastic. In rodents, the energy balance-regulating system is structurally and functionally immature at the start of postnatal life. The basic anatomy of the rat hypothalamus is established prenatally, with its nuclei expressing specific neuropeptides being recognizable before birth (275), but the majority of connections between the hypothalamus and its input and output systems (276, 277), and those within the hypothalamus itself (27, 278), develop only in the first weeks after birth. This rapid postnatal development is also reflected in overall brain growth: in neonatal rats, total brain weight increases by a factor 5 between birth and weaning (279).

Developing rat pups go through some major transitions. Whereas the fetus receives mainly glucose, lactate, and amino acids via the placenta, at birth the source of energy changes to high-fat mother’s milk (280). Only a few weeks later, the pups are weaned and make a more gradual transition to the high-carbohydrate, low-fat adult diet





**FIG. 3.** Simplified diagrams of the hypothalamic regulation of energy balance. Besides the main pathway (see Fig. 2), these schematics show additional connections from leptin (A), the ARC (B), the PVN (C), and the LHA (D). Note that in A, the connection to the LHA does not contact MCH or ORX neurons directly. *Green arrow*, Positive effect; *red inverted arrow*, negative effect; *blue crossed line*, unspecified effect; *solid line*, direct connection; *dashed line*, indirect connection; *dotted line*, probable connection; *green (red) nodes* lead to more positive (negative) energy balance when stimulated/active. See the text for details.

(280, 281). At the same time, the pups have to make the transition from obtaining all energy and fluids from the dam by suckling to the two separate processes of feeding and drinking (282). As will be described below, different mechanisms appear to regulate these different types of ingestive behavior.

Rat pups as young as 1 d old already regulate their milk intake according to how deprived they are (283, 284). The only cue that suckling rats have been shown to use to regulate their milk intake is the distension by gastrointes-

tinal fill (285, 286). This response is mediated primarily by vagus nerve activity (285, 287), and hence by the brainstem rather than by the hypothalamus. Other signals that influence food intake in adult rats, such as the nutritional value of the stomach contents, serum leptin levels, and manipulations of levels of glucose and free fatty acids, do not affect intake in suckling rats (286, 288, 289). It appears that the regulation of energy balance in the suckling pup is limited to optimizing energy intake for growth, and intake is only restricted by a full stomach to prevent gross overeating. Therefore, in suckling pups there seems to be only short-term regulation of milk intake, with no long-term regulation (290).

Thermoregulation and the regulation of adult forms of ingestion then develop in the early postnatal period. From d 1 on, pups can already regulate their temperature by moving toward or away from a heat source (291), whereas mechanisms for thermogenesis develop over the first 2 wk of life (288). In the first 10 d of life, gastric distension is the only cue that terminates intake. From then on, the nutritive value of the gastric content starts to play a role (reviewed in Refs. 276 and 290). At this same age, pups first start to ad-

just their intake according to their level of fatty acids (reviewed in Ref. 282), whereas a similar response to glucose levels does not appear until the age of 4 to 5 wk (284, 290). Another major development event is the differentiation between feeding and drinking; young pups simply increase their intake when they are dehydrated, and only from around the age of 20 d they will reduce their milk intake when dehydrated, a phenomenon called dehydration anorexia (reviewed in Refs. 276 and 290).



The development of the regulation of energy balance is accompanied by changes in mRNA and protein levels of the reviewed peptides. The ontogeny of these peptides is summarized in *Sections III.G.1–4* and depicted in Fig. 2B.

### 1. Leptin

In rats, leptin can be detected in fetal plasma on d 19 of gestation (292). In nearly full-term fetuses (d 20–21), leptin levels strongly resembled those of the pregnant dams (292, 293). Leptin mRNA is already expressed by rat adipose tissue at birth, and its expression and serum levels are immediately regulated by the nutritional status of the neonatal pup (294). In addition, Ob-Rb, the leptin receptor, has been shown to be expressed in the fetal brain as early as d 14 of gestation (295, 296).

During the lactation period, leptin undergoes some major changes. A first, relatively small increase in serum leptin levels can be detected in rat pups 1 to 2 d old (293, 297), followed by a second and larger peak around d 7 to 12 (297–299). Interestingly, this leptin surge is unrelated to changes in body weight and fat content in the neonatal period (297, 300). The high leptin levels do coincide with elevated leptin mRNA in neonatal adipose tissue, suggesting that the peak originates from the pups' own leptin production (301). Both the neonatal pituitary, which has high leptin expression during this period (298), and the dam's milk (302) may contribute to the leptin surge. In concert with the changes in leptin levels, hypothalamic levels of Ob-Rb and its mRNA rise significantly between birth and weaning (297, 303).

Leptin's functionality in the regulation of energy balance appears to be partial in the neonatal period. In rats as young as 1 wk old, leptin injections are found to reduce gain in body weight and especially in fat mass, without any effect on milk intake (289, 304–306). Instead, these effects seem to be the result of an increase in energy expenditure (306, 307). Leptin is effective in increasing POMC and decreasing NPY mRNA in the ARC of rats in this neonatal period (305), and a robust positive relation between leptin levels and fat mass has been reported on d 10 (308). The exact timing of the development of this system seems to differ between mice and rats. In mice, serum leptin levels were not altered after milk deprivation on d 8 (300), energy expenditure was not yet increased by leptin injections on d 9 (309), and daily leptin injections in the second week of postnatal life were not found to affect hypothalamic neuropeptide expression (310).

During the period of partial functionality in energy balance regulation, leptin has a neurotrophic role. In the absence of leptin, general brain development and that of the hypothalamic circuitry specifically are impaired (311, 312). Leptin shares this property with insulin, which is also implicated in brain development. The neurotrophic

actions of insulin include stimulation of neurite outgrowth, protein synthesis, and neuronal survival (313–315). Leptin's neurotrophic effects may actually persist until adulthood because leptin administration in *ob/ob* mice significantly alters the synaptic input on both NPY and POMC neurons in the ARC (316). Furthermore, the lining of the third ventricle has been shown to contain neural progenitor cells that can be induced by neurotrophic factors to proliferate and differentiate into functional hypothalamic neurons (317, 318). This residual plasticity of the hypothalamic circuitry in adulthood provides an additional route by which environmental signals (including leptin) can regulate energy balance (319).

### 2. ARC peptides

The four reviewed peptides that are expressed by the ARC (NPY, AgRP, POMC, and CART) are already expressed in the prenatal rat brain (295). However, ARC projections to other hypothalamic nuclei only develop during the early postnatal period (320). During this period, there are also dynamic changes in the levels of the peptides and their gene expression.

*a. Orexigenic ARC peptides.* NPY peptide is detected in the rat fetal midbrain as early as d 13 or 14 of gestation (321–323). NPY mRNA levels rise during gestation to reach near adult levels around birth (75, 324). Like leptin, NPY gene expression is elevated during the lactation period, with a peak around d 16 (324, 325). At the same time, NPY mRNA is transiently expressed in hypothalamic areas that do not produce NPY in adulthood. Suckling rat pups express NPY mRNA in the DMN, PVN, LHA, and perifornical area, albeit at lower levels than in the ARC (324, 326). Alongside the developmental changes in NPY mRNA, NPY peptide levels show a rapid postnatal rise and in the ARC reach adult levels by the time of weaning (327, 328). Immunohistochemistry studies have shown that the number of cell bodies containing NPY peptide rises gradually until birth, with declining numbers afterward (321, 323). After d 10, NPY cell bodies can only be visualized when axonal transport is chemically blocked by colchicine administration—a finding that is consistent with the simultaneous increase in NPY-immunoreactive fibers throughout the hypothalamus (321–323). In a more recent study, by staining for NPY and AgRP peptide simultaneously, the origin of these postnatally developing fibers was proven to be the ARC (329). Indeed, the developmental pattern of AgRP resembles that of NPY, with increasing expression during the first postnatal weeks and a peak around d 16 (329, 330).

In the neonatal period, NPY and AgRP already appear to have some functionality. Maternal deprivation has been shown to increase expression in the ARC already on d 2

(NPY) and at least from d 11 (AgRP) (326, 331). Furthermore, NPY injections into the PVN increased intake of water and milk as early as d 2; on d 15 the pups showed a preferential increase in milk intake (332). As mentioned, intrahypothalamic fibers in the neonatal rat are still incomplete, and NPY is expressed in several hypothalamic nuclei. Therefore, NPY may exert most of its actions locally at the site of expression, rather than after being axonally transported from the ARC to other hypothalamic regions.

**b. Anorexigenic ARC peptides.** POMC mRNA is first detected in the midbrain on d 13 of gestation (333). During the lactation period, hypothalamic POMC expression is either stable (334) or increases toward weaning (310). ARC POMC expression then increases significantly between weaning and young adulthood (335, 336). Hypothalamic POMC peptide has been detected as early as d 12 of gestation (337, 338), with  $\alpha$ -MSH, the cleaved product, only appearing between d 15 and d 19.5 of gestation (337, 339, 340). Postnatally, POMC and  $\alpha$ -MSH protein in the ARC go through a rapid increase, to peak around d 21 to 28 (337, 340). There is only limited information about early CART ontogeny. One study in mice has reported low levels of hypothalamic mRNA on postnatal d 5, with near adult levels on d 10 and 22 (310). However, the developmental patterns reported by this study for NPY, AgRP, and POMC were different from those found in most other studies.

In contrast to NPY,  $\alpha$ -MSH does not seem to have much functionality early in life. In 1-wk-old rat pups, many PVN neurons are responsive to NPY, whereas only a few show a response after administration of an  $\alpha$ -MSH agonist (341). At the age of 4 to 5 wk, however, the number of PVN neurons responsive to NPY has decreased, whereas the number of neurons responsive to melanotan II has increased dramatically (341). This phenomenon may ensure a high intake in neonatal life by minimizing anorexigenic signaling in early life.

### 3. PVN and LHA peptides

Less detailed information is available about the development of the peptides of interest in the PVN and the LHA: CRH, TRH, MCH, and ORX. Gene expression is detected in the fetal rat brain for all four peptides (342–346). The peptide is generally also detected in the hypothalamus before birth (343, 346–348). Neonatally, there is a gradual increase in expression and protein levels of most peptides, and adult levels are generally reached around the time of weaning (207, 344–346, 349–353), although ORX and TRH peptide levels may keep on rising between weaning and young adulthood (347, 354).

Functional tests are reported for ORX. In the neonatal period, leptin administration increases ORX mRNA in the

LHA (353), where the normal effect in adults would be inhibition of expression (210). Interestingly, the neonatal leptin administration that increases ORX expression does not affect body weight and blood glucose levels, whereas 24 h of milk deprivation reduces body weight and blood glucose levels but does not affect ORX expression levels (353). Therefore, the neonatal leptin effect on ORX mRNA may be interpreted to reflect a developmental role, rather than an effect on energy balance regulation (353).

### 4. Development in humans and rats

If we want to extrapolate data and conclusions from animal studies to the human situation, it is important to consider the respective timing of the ontogeny of the relevant systems in humans and rats. At birth, humans are further in their development, and many developmental events that occur in the early postnatal period in rats take place in the third trimester of human pregnancy (27, 355–357). NPY immunoreactivity is first detected in the human ARC at about 21 wk gestation (41), and in nonhuman primates, NPY/AgRP projections to the PVN increase dramatically during the third trimester of gestation (358) and seem to be nearly complete by birth (27, 278). Therefore, caution is needed in extrapolating findings from one species to another.

## IV. Energy Balance Programming

As has been shown in *Section III.G*, a large part of the development of the energy balance-regulating system occurs in the perinatal period in both man and rat, although the exact timing of developmental events differs between the two species. With the knowledge of the previous section, one can imagine that the perinatal period with its rapid development may be a critical period and that during this critical time-window, the organism is vulnerable to environmental influences. One can also imagine that different timing of an external stimulus, relative to the stage of development of the organism, can produce different outcomes. Also, different types of stimulus (*e.g.*, undernutrition *vs.* overnutrition, global *vs.* specific nutrients, maternal *vs.* fetal/neonatal) may produce different outcomes. Therefore, in this section, we will discuss developmental programming of energy balance according to the type and timing of the stimulus.

### A. Indicators of developmental programming of energy balance

To identify programming of energy balance, different approaches have been taken. There are basically three types of outcome that can be measured to investigate this phenomenon. An indirect way of looking at energy bal-

ance is to measure body dimensions and body composition. Because positive energy balance results in fat deposition and allows growth, these measurements can give an indication of enduring positive or negative energy balance in the (recent) past. Relevant parameters are body weight, body length, body mass index (BMI), fat mass and lean mass, and whether or not there is complete catch-up growth. These parameters are most apparent, and in humans are often the first indication that energy balance may be disturbed. Another way of investigating energy balance programming is to examine components of energy balance directly. Energy intake, resting energy expenditure, and activity-related energy expenditure together determine energy balance. These parameters may be somewhat less explicit in everyday life, but they can be studied relatively easily, also in the human situation. The third approach to investigate energy balance programming is to study the peptides and hormones that are responsible for the regulation of energy balance. Properties like gene expression, peptide levels, epigenetic modifications, and functional changes can be studied. Because these measurements require invasive techniques, this approach is less suitable for use in the human situation.

Naturally, a combination of the three approaches will generate the most complete description of the phenomenon of developmental programming of energy balance. With many new studies on the subject, our understanding of this phenomenon has much advanced in recent years. Now, various influences of the perinatal environment on energy balance parameters will be discussed—first, briefly for the human situation, and then in different rat models.

## B. Programming of energy balance in humans

Epidemiological evidence suggests that the early environment can have a profound influence on energy balance. With these studies, it must be kept in mind, however, that in the human situation, the underlying cause of low birth weight or restricted fetal growth varies and is often unknown (25, 359). In addition, there are many confounding factors (*e.g.*, the living conditions) that may obscure the real effects of the early environmental influence.

### 1. Body dimensions and body composition

Although higher adult body weight and BMI have repeatedly been reported with increasing birth weight (13, 16–19), the notion that low birth weight and impaired fetal growth may also program increased adiposity is gaining recognition. Over the last decade or so, researchers have increasingly investigated effects on more refined indicators of obesity, such as body composition (lean *vs.* fat mass) and fat distribution (*e.g.*, waist-to-hip ratio, skin-fold ratios). These studies have shown that the positive

relationship between birth weight and adult BMI results mostly from a positive relationship with lean mass, but not with fat mass (18, 360–365). Moreover, low birth weight and impaired fetal growth have now been shown to be associated with a higher fat percentage in later life (10, 365–369) and with a detrimental distribution of fat (*i.e.*, more central, abdominal, and visceral) (363, 370–380). The fact that these studies were performed in diverse populations [from different European countries (Belgium, Finland, France, The Netherlands, Spain, and the United Kingdom), the United States (non-Hispanic white, non-Hispanic black, and Mexican-American), Brazil, Guatemala, and Jamaica], with different ages (from young children to old age), and in both sexes underlines the robustness of these associations. It is becoming more and more clear that low birth weight is not always a reliable proxy for impaired fetal growth. When, for example, early-gestation growth impairment is followed by prenatal catch-up growth, adult health can be affected without an effect on birth weight (see Ref. 10). Furthermore, the significance of the rapid postnatal catch-up that often follows perinatal undernutrition, rather than that of the undernutrition *per se*, has been stressed in recent years. Several studies have shown that rapid early growth (with the definition of early ranging from the first week of postnatal life to about 3 yr) increases the risk for later adiposity and obesity (364, 381–385). This at least partly removes the apparent paradox of the association of both low and high birth weight with metabolic syndrome and obesity. When both situations are characterized by perinatal overfeeding (even if this is postnatal-only in the case of SGA subjects and may be both pre- and postnatal after maternal obesity), the underlying mechanisms may also share some similarities.

Maternal obesity and gestational diabetes are increasingly common problems (386, 387). The newborns of those affected usually have greater birth weights than infants born to control mothers (388–392). Greater gestational weight gain is also associated with higher birth weight (393, 394). Even when their birth weight is not altered, the offspring of diabetic mothers often have an increased fat percentage (395). In older children, ranging from 2 to 10 yr of age in the different studies, more obesity was found in those that were born to obese or diabetic mothers (393, 396–401). Interestingly, this obesity-prone profile improved dramatically after bariatric weight loss surgery. Children that were born to obese mothers with substantial weight loss after surgery had lower birth weights without a higher risk for SGA, and their obesity rates in the ages of 2 to 25 yr were reduced to normal population levels (402). To summarize, more obese phe-



notypes with detrimental adiposity have been found after both prenatal undernutrition and overnutrition.

## 2. Energy intake and expenditure

Relatively few studies have directly assessed energy balance parameters in low- and high-birth-weight subjects. For energy expenditure, mostly neonatal data are available. These suggest that infants that are born SGA have higher energy expenditure than both premature appropriate-for-gestational-age very low-birth-weight infants (403–406) and at-term appropriate-for-gestational-age infants (407, 408). In a study on prepubertal children on the other hand, SGA subjects were reported to have reduced resting energy expenditure compared with at-term appropriate-for-gestational-age children (409). Energy intake was generally similar to that of premature infants of the same body weight (403, 405, 406). One study reported a higher intake per kilogram body weight in SGA infants, whereas those large for gestational age had a lower relative intake compared with control infants of the same postnatal age (410). In a more long-term study, a sample of prepubertal SGA children that did not catch up had a food intake below the recommended energy intake for their age (411). After gestational famine exposure, middle-aged subjects had a higher energy intake, consumed diets with a higher fat density, and had lower levels of physical activity than nonexposed persons (412, 413).

## 3. Peptides and hormones

In humans, measurements of the third category (that of the peptides and hormones that are involved in the regulation of energy balance) have largely been limited to the circulating hormones. Serum leptin levels have been investigated most thoroughly.

In neonates, several studies have found positive correlations of leptin with birth weight, birth length, and BMI (414–419). Because the strongest correlation was usually found with BMI, these associations most likely reflect the deficit in fat deposition in low-birth-weight infants and the excess in those born after fetal hypernutrition, respectively. However, a programming effect is suggested by the fact that subjects that were born with a low birth weight were found to have high leptin levels with respect to their BMI at several different ages (ranging from 4 months to adulthood) (420–423). Another report that suggests programming of leptin levels studied the influence of early nutrition in preterm infants (424). It was shown that adolescents that had received preterm formula had more leptin per kilogram fat mass than adolescents that had received a control diet in infancy (424). Besides altered leptin levels, a few studies have shown increased ghrelin levels in SGA subjects at birth (417, 425, 426) but not at the age of

1 yr (427). In contrast, high-birth-weight newborns were reported to have normal ghrelin levels (425). Children 2 to 25 yr of age that were born to obese mothers after bariatric weight loss surgery had higher ghrelin levels and lower leptin levels than those born before such surgery, a beneficial profile that corresponded to their improved body composition (402). Lastly, there is also some evidence (in neonates and children) that the HPT axis may be disturbed in SGA subjects (428, 429).

## 4. Evidence for developmental programming

Summarizing, there is quite some evidence that the early nutritional environment can have a permanent effect on the body dimensions of humans. The long-term effects observed at both sides of the birth weight spectrum seem to share their general direction: after the initial period of catch-up growth after perinatal undernutrition, both are associated with more obese phenotypes. Although direct measurements of energy balance and its regulation are still scarce, disturbances have been found, some of which seem to persist into adult life. Because these kinds of measurements are more invasive and some can only be performed postmortem, they are obviously not employed in humans on a large scale. That is why different animal models were designed to study these effects more closely.

### C. Programming of energy balance in animal models

The use of experimental animal models has some substantial advantages over studies in humans. In contrast to the human situation, with animal models for perinatal restriction of growth and nutrition, the exact cause of the observed symptoms is known, and the degree of control over the subsequent environment is far greater. In addition, animal models permit the use of more invasive methods than in humans.

Experimental animal models for developmental programming have been designed in various species, including primates, sheep, guinea pigs, and rats (28, 31, 387, 430). In this review, we will focus on studies in the rat, although a few studies in mice are also included. In rats, both prenatal and postnatal manipulations of nutrition have been used to induce developmental programming of energy balance, including ligation of the uterine arteries; maternal diets with altered protein, fat, or energy content; and manipulations of litter size (28, 31, 387, 430). These different models produce different phenotypes. Here, we will first describe effects on the body dimensions and body composition of the major models that have been used in rodents. Then, the effects on energy balance and its regulation will be discussed.

## 1. Body dimensions and body composition

*a. Nutritional manipulation of the dam.* Prenatal manipulations of fetal nutrition, via the diet of the pregnant dam, exert long-term effects on the body dimensions of the offspring, with or without an immediate effect on birth weight of the pups. Two major types of this kind of manipulation are maternal low-protein diets and global maternal food restriction to different degrees (ranging from 30 to 70% of control intake). Perinatal overfeeding, on the other hand, can be induced by feeding the dams high-fat or high-energy (high on both fat and sugar) diets.

Whether a maternal low-protein diet actually reduces birth weight of the pups appears to depend on the exact composition of the diet and other details in the methodology because some studies (mostly using a low-protein Hope Farms diet) report lower birth weights (431–435), whereas others (mostly using the Southampton diet) have reported normal birth weight after maternal low-protein diet during gestation (436–439). After a maternal low-protein diet, body weight either stays reduced or normalizes to control levels, with the outcome apparently independent of birth weight and the experimental diet used during pregnancy (431–437, 439–442). Two studies have reported rapid catch-up growth with increased body weight (438, 443). Adult body composition after a maternal low-protein diet has mostly been reported to be normal (439–442), although some of these studies did report an altered fat percentage in either males or females. One study found increased leptin and triglyceride levels in males, but not females, with otherwise normal body weight and fat mass (442). This suggests that, although the body composition may be normal, its regulation can still be disturbed in these animals.

Maternal food restriction usually reduces birth weight of the resulting pups (301, 444–451), except when the food restriction is limited to the first 2 wk of pregnancy or in some cases when intake is only mildly restricted to 70% of control intake (452–455). After maternal food restriction, rats show either complete or incomplete catch-up growth (446–448, 452, 456) so that in rats with a low birth weight, adult body weight was reduced, normal, or elevated compared with that of controls (445, 449, 450, 457). Several studies have found normal body composition after prenatal maternal food restriction (301, 445–447, 453–455, 458–460). However, increased and decreased adiposity has also been reported. Within studies, these different outcomes can be attributed to sex differences, different effects at different ages, strain differences, and timing of the food restriction (445, 452, 453, 455, 458, 460). Between studies, the method of determining body composition (*e.g.*, BMI, weight of different fat pads, total lipid determination by carcass analysis, dual-energy

x-ray absorptiometry) and the severity of the food restriction may explain a large part of the variation in outcome. One group that uses severe maternal food restriction (to 30% of control levels) has consistently found a persistent lower body weight, combined with increased fat mass and leptin levels in both males and females (449–451, 456, 457). Leptin levels usually reflected body composition (301, 445, 447, 449–451, 457, 459, 460), although in one study increased leptin levels appeared to precede the increased fat percentage (445). In summary, although studies using the Vickers model present a constant exception, most studies have found normal body composition after prenatal maternal food restriction.

Because a considerable part of the developmental events that occur *in utero* in humans take place after birth in rats, postnatal manipulations are also frequently used as a model. When the same maternal dietary manipulations that are used prenatally are either started or continued in the lactation period, the reductions in body weight are generally longer lasting, and less catch-up growth is reported (15, 431, 433, 434, 442–447, 461–467). Concomitantly, an obese phenotype is observed less frequently than with strictly prenatal manipulations (434, 442, 445–447, 460–462). There may be less catch-up growth after these postnatal manipulations because the condition is too severe to recover from (especially when prenatal and postnatal malnutrition are combined), or at weaning the animals may have reached the end of the time-window in which complete catch-up is possible. Alternatively, the fact that these animals do not seem to be “programmed for obesity” may reflect a different type of programming than with exclusive prenatal maternal dietary manipulation.

Besides maternal underfeeding paradigms, maternal overfeeding and gestational diabetes have also been induced in rodents. After maternal overfeeding before and/or during gestation, birth weight in the offspring can be either higher (468) or lower (469) than in control animals, but it is more often found to be unchanged (470–476). Interestingly, one study reported a lower birth weight specifically after a pregestational-only cafeteria diet (477). With maternal overfeeding continued into lactation, a substantial number of studies reported increased body weight by the time of weaning (468–471, 476, 478–480), although a reduced body weight was found in a study where the high-fat-fed dams lost more weight during lactation than the control dams (472). In later life, animals born to overfed dams had normal (475, 479, 481, 482) or elevated (469, 471, 476, 478, 480, 483, 484) body weight when fed on chow. A higher body weight was usually accompanied by increased adiposity (469–471, 476, 479, 480, 483, 484). In one study, the development of overweight was specific to animals that were born to control

dams but then cross-fostered to dams fed on hypercaloric diets (485). When transferred to an obesogenic diet themselves, some (471, 475, 481, 482), but not all (476, 478, 481, 482), of these animals showed an increased susceptibility to diet-induced obesity. In the studies by Levin and colleagues (481, 482), the adverse consequences of the maternal diet were mostly specific to animals from a strain bred for diet-induced obesity, demonstrating the importance of the interaction between perinatal nutrition and genetic factors.

In rodents, gestational diabetes can be induced by glucose injections in early pregnancy or injections of the pancreatic islet toxin streptozotocin, but it also occurs in the female offspring of rats that underwent uterine artery ligation (see *Section IV.C.1.b*) and in *db/+* mice that are heterozygous for a silencing mutation in the leptin receptor (486–489). Mostly, birth weight is found to be increased in these models (486, 487, 489), although normal birth weight has been reported after maternal streptozotocin injections (488). Around weaning, body weight remained higher in the offspring of ligated dams (486), remained normal, or increased slightly after streptozotocin injections (488, 490). Cross-fostering to normal dams after birth did not influence growth (486), but normal pups that were cross-fostered to diabetic dams had lower body weights (491). Offspring of diabetic mothers was reported to be overweight with increased adiposity in adulthood (489, 492, 493).

**b. Nutritional manipulation of the offspring.** Uterine artery ligation in the pregnant dam reduces the blood flow to the fetuses (494) and is frequently used as a model for placental insufficiency, the most common cause of low birth weight in westernized countries (24). To approach the human IUGR situation as closely as possible, often only pups that are growth restricted according to similar criteria as those used in humans are selected for studies (495). This obviously results in a birth weight that is by definition reduced (496–500). Nevertheless, studies that did not use pup selection have also reported a lower birth weight in rats born after uterine artery ligation (494, 500–502). The long-term effects on body weight seem to be dependent on the exact timing of the ligation. When performed on d 17 of gestation, the weight deficit is usually persistent (496, 503–506), whereas after ligation on d 19 of gestation, complete catch-up growth has been reported (498, 507, 508). Some studies also found a return to normal body weight after ligation on d 16 or 17 (500, 509). Newborn pups that were growth restricted by uterine artery ligation were shown to have a fat percentage that was either reduced or comparable to that of control pups (497, 501). Juveniles and adults that do not completely catch up in

body weight have been shown to have normal BMI, fat percentage, and serum leptin levels (496, 510). The ones that do catch up to control body weight also have normal leptin levels when young (at an age when their body weight is still reduced) (498, 511). Rats that stay at the same body weight as control rats after catch-up have elevated leptin levels and increased fat mass in adulthood (498). The group that reported overweight in adulthood found normal or increased fat mass at the age that body weights were similar to those of controls (508, 512) and increased fat mass afterward (512, 513). In summary, when there is complete (or even overcomplete) catch-up in body weight, the animals' body composition is disturbed and shifted toward a more obese phenotype. If the catch-up growth stays limited, however, body composition remains normal. It seems likely that the capacity for true growth of organs and other lean tissue is curbed by the early growth restriction, and if there is catch-up beyond a certain point, any additional "growth" is in fat only.

A method to manipulate early postnatal nutrition that targets the offspring directly (rather than indirectly via the diet of the dam) is to manually adjust the number of pups nursed in a litter (514, 515). In this way, both neonatal under- and overnutrition can be achieved. By definition, birth weight is not affected by these manipulations because they take place after birth. Shortly after redistribution into litters of different sizes, differences in body weight become apparent. Rats that are raised in a small litter of only two to five pups receive more milk, resulting in a higher growth rate and body weight before weaning (250, 516–525). Although a few studies report normalization of body weight (518, 519, 526–528), this elevated body weight is generally found to persist into adulthood and middle-age (250, 516, 520, 521, 523, 526, 529–543). The opposite is true for rats that are raised in a large litter of 14 to 24 pups, which has less milk available per pup. These rats grow much slower during the lactation period and have a significantly lower body weight (496, 503, 516–522, 544, 545). Again, some studies report normalization (519, 526, 546), but most researchers find that body weight is persistently reduced (496, 504, 516, 518–521, 526, 533, 535, 539, 540, 542, 544, 545, 547). Already during the lactation period, the two models show marked effects on body composition: overfed small-litter pups have an increased fat percentage and leptin levels, whereas these are both decreased in underfed large-litter pups (517, 519, 520, 522, 524, 525, 545). Thus, a disproportionate part of the added growth in small-litter pups can be ascribed to adipose tissue. After weaning, when all animals are transferred to a normal feeding regime, body composition remains disturbed. In most small- and large-litter rats with persistent changes in body



weight, fat percentage and leptin levels also remain altered into adulthood and middle-age (496, 510, 516, 518, 520, 527, 529, 533–535, 537, 538, 540–543, 545). One study even reported an increased fat percentage in small-litter rats at an age when their body weight was no longer elevated (527). Apart from a few exceptions, the effects of neonatal litter manipulations are long-lasting and also rather consistent between studies. Neonatal overfeeding by raising rats in small litters causes an immediate rise in growth velocity, with persistent higher body weight and fat mass in adulthood, resulting in an obese phenotype. Neonatal underfeeding by raising rats in large litters, on the other hand, acutely reduces growth rate and causes a permanently lower body weight and fat mass, resulting in a leaner phenotype.

*c. Response to a dietary challenge.* This section has demonstrated that diverse manipulations of perinatal nutrition can bring forth different phenotypes. Even seemingly comparable manipulations have been shown to generate different long-term effects on body dimensions and body composition. What's more, some of these manipulations have been shown to alter the animals' susceptibility to diet-induced obesity (which is induced by feeding a hypercaloric diet, usually a high-fat diet). Again, there is considerable variation in the reports on this effect. A maternal low-protein diet either did not affect (548) or increased (15, 549, 550) the susceptibility to diet-induced obesity when the manipulation was prenatal. When the manipulation was restricted to the lactation period, less obesity was observed on a highly palatable diet (15). Several studies have reported a higher susceptibility to diet-induced obesity after prenatal maternal food restriction (301, 453, 454, 456, 457, 459, 551, 552), but unchanged obesity has also been reported (450, 451, 453, 454, 551, 553). Here, there seems to be a difference in susceptibility between the sexes, although this sex difference may be strain-dependent; Jones (453, 454, 551) reported increased diet-induced obesity in Sprague-Dawley males but not females, whereas Vickers (450, 451, 456, 457, 459, 552, 553) found higher susceptibility in Wistar females but not males. In rats that were neonatally overfed or underfed by raising them in small or large litters, conflicting results have also been reported. In rats with persistent differences in body weight, some studies found no difference between the two models in their susceptibility to diet-induced obesity (518, 533). One study, however, reported that diet-induced obesity was augmented in small-litter rats and diminished in large-litter rats (540). From these data, we can conclude that the effects of a dietary challenge are mostly consistent with the general phenotype. More diet-induced obesity is observed in those models that un-

der baseline conditions showed more catch-up growth and increased adiposity.

## 2. Energy intake and expenditure

In the above-mentioned rodent models, energy intake and energy expenditure have been studied using a range of different parameters. Expenditure-related parameters include resting and total energy expenditure, (locomotor) activity, body temperature, and measurements of thyroid function and cellular metabolism. For energy intake, the variety is more in how the data are represented. Daily food intake is given per animal (raw data), per kilogram body weight (or some other approximation of body size), or adjusted for body size in a statistical test. The results of these different representations are not always easily compared. Especially when intake is divided by body size, the results can be distorted. Because energy requirements per kilogram body weight fall with increasing body size, this calculation systematically underestimates energy utilization by larger individuals (554). Therefore, such studies are excluded from this review; only studies that report raw food intake data or intake adjusted for body size in a statistical test are included.

*a. Nutritional manipulation of the dam.* One study that induced prenatal underfeeding by a maternal low-protein diet reported normal food intake in the adult offspring (442). The same study found reduced food intake when the underfeeding was (continued) during the lactation period. This was confirmed by others (433, 467), although some have also reported normal levels of food intake in these (prenatally and) postnatally malnourished rats (432, 461, 464). These data suggest a subtle decrease in food intake after protein malnutrition in the lactation period, whereas prenatal-only malnutrition probably does not affect long-term energy intake. On the expenditure side, in rats with postnatal exposure, increased thyroid function (pointing to increased basal metabolism) was found (463, 465), and normal-to-low activity levels have been reported after prenatal exposure (441). Taken together, these studies suggest that in prenatally malnourished animals normal levels of intake and reduced activity may lead to positive energy balance, whereas in postnatally malnourished animals a negative balance may result from their lower food intake and increased basal metabolism.

After maternal food restriction, food intake was usually found to be similar to that of control animals. However, when body size is taken into account, the effects on energy intake differ according to the timing of the malnutrition: prenatally or postnatally. When pups were exposed to the maternal diet postnatally, they often had reduced body size combined with normal food intake (445, 447, 464, 467), which results in an elevated relative energy intake.

With prenatal-only maternal food restriction, both body size and food intake were usually normal (301, 447, 452–454, 458), leading to a normal relative food intake. In a few cases, both body size and food intake were elevated (445, 453, 454), which also may point to a fairly normal relative energy intake. Measurements of energy expenditure were mostly performed in prenatally underfed rats; in postnatally underfed rats, one study reported a normal thyroid function (463). Using the Vickers' model of prenatal maternal undernutrition, female rats (that have a low body weight and high fat mass) were found to have reduced activity levels in adulthood (552, 555). Other studies using prenatal undernutrition have reported normal levels of activity (453) and normal body temperature and resting energy expenditure (301). These data are suggestive of normal total energy expenditure, which together with an unaltered food intake points to a normal energy balance for these rats that are prenatally exposed to maternal undernutrition.

In juvenile pups born after maternal overfeeding, food intake may be normal (476, 480, 484), although dramatic overfeeding has also been reported (468). In adulthood, these animals are usually hyperphagic (469, 471, 476, 478, 480, 484). Moreover, rats born to cafeteria-diet-fed mothers showed a stronger preference to fatty and sugary foods themselves (483). In offspring of high-energy diet-fed dams, reduced activity levels and slightly increased diet-induced thermogenesis have been reported (479, 484). This will probably lead to lower total energy expenditure, with the reduction in locomotor activity only enhancing the obesogenic effects of the elevated food intake. An elevated food intake was also reported in rats born after gestational diabetes (493).

**b. Nutritional manipulation of the offspring.** Food intake was not widely studied after uterine artery ligation; one study reported decreased food intake (498), whereas another found an unaltered intake per kilogram body weight (508). In both studies, the experimental animals had similar body weight as controls (which nullifies the interpretational problems of the per kilogram representation). In both juvenile and adult rats, cellular metabolism was reduced (513, 556, 557), whereas locomotor activity has been reported to be normal (499, 500). Taken together, a reduced or normal food intake, lower basal metabolic rate, and probably normal activity-related energy expenditure suggest that energy balance may be either approximately normal (intake and expenditure both reduced) or more positive than in control animals (normal intake with reduced expenditure), respectively.

In virtually all small-litter rats that were heavier than controls, food intake was reported to be elevated through-

out life (523, 526, 529–532, 536, 538, 539, 541, 543) although this was not always the case (537). In rats that would later lose their overweight, unchanged food intake was found in juvenile life (519, 527). Fewer studies have reported on the expenditure side of the balance. Rats raised in small litters were found to have a higher body temperature and resting expenditure (521), and in young animals, elevated total energy expenditure was reported (543). The latter study found that the elevation in energy expenditure was appropriate for the larger body size of the small-litter rats. Because both energy intake and expenditure are increased in these animals, the overall effect on energy balance depends on the relative sizes of the effects on intake and expenditure. These are difficult to compare between studies. On the other hand, large-litter rats were generally reported to have lower energy intake and expenditure than controls (521, 526, 539, 545, 558). Again, the fact that these measurements were taken in separate studies complicates interpretations about the overall effect on energy balance in these animals.

The foregoing paragraphs have shown that different models of perinatal malnutrition can have different effects on adult energy balance. They have also shown that, although there is a lot of information about the effects of these manipulations on components of the energy balance, the exact information needed to assess a directional change in energy balance is not always available. Furthermore, in the interpretation of these studies, it is vital to distinguish absolute measurements from adjusted data. Comparisons should only be made between data that are expressed in the same dimensions.

**c. Response to leptin administration.** A related parameter that marks the transition to the subject of the next paragraph is the anorexigenic effect of leptin. Peripheral leptin administration acutely reduces food intake in control animals, but not in adult rats that were previously subjected to prenatal or postnatal maternal food restriction or a postnatal maternal low-protein diet (456, 464, 559). In young adult small-litter rats, central injections of leptin are effective, in contrast to peripheral injections (537). This suggests that this leptin resistance may be due to impaired leptin transport, rather than an altered hypothalamic response (537).

### 3. Hypothalamic regulation

It has been known for quite some time that perinatal malnutrition can have profound effects on brain development (560). Nevertheless, studies investigating programming effects on the hypothalamic peptides that regulate energy balance are relatively scarce (compared with the other two categories of measurements discussed above). Most of these have studied relatively short-term effects.

*a. Nutritional manipulation of the dam.* Weanling rats subjected to a maternal low-protein diet during gestation and lactation were shown to have a reduced number of NPY immunoreactive cells in the ARC (561). This was combined with an increase of the concentration of NPY protein in the PVN and LHA and a tendency for an increased concentration in the ARC, whereas the NPY content of other hypothalamic nuclei was unaltered (562). This is suggestive of an increased orexigenic drive in these animals, provided that the PVN and LHA are fully responsive to NPY. In view of the slightly hypophagic phenotype of these animals (see *Section IV.C.2.a*), the responsiveness of these areas (or other regions further downstream) is probably reduced. Rats that were only exposed to a low-protein diet prenatally did not show changes in ARC gene expression of Ob-Rb, NPY, AgRP, POMC, and CART at weaning (563). In contrast, weanling pups that were subjected to the diet postnatally had increased expression of Ob-Rb, NPY, and AgRP and decreased expression of the anorexigenic POMC and CART (563), again suggesting an increased orexigenic drive. After fasting, NPY and AgRP mRNA were increased relative to control levels in weanling rats that were exposed either prenatally only or both pre- and postnatally, although the effects were stronger in the latter group (432). In adulthood, CART mRNA was found to be increased in animals that were exposed prenatally (but not in those exposed both pre- and postnatally), with no changes in expression of NPY, AgRP, and POMC (432).

Prenatal maternal food restriction has been shown to drastically increase hypothalamic mRNA levels of Ob-Rb at birth (559), an effect that was reversed by weaning to levels below normal (559, 564). In adulthood, hypothalamic Ob-Rb expression had normalized (459), but Ob-Ra expression was lower than in control animals (301), which points to a reduction in leptin transport. The latter is supported by a normal reaction to central injections of leptin, with a reduced reaction to peripheral leptin (301). In weanling rats, reductions in leptin, ghrelin, NPY, and  $\alpha$ -MSH peptide levels, as well as NPY and POMC mRNA levels have been reported (564). Adult hypothalamic expression of the ARC peptide AgRP was reduced, whereas that of NPY and POMC was normal (459). Despite this, the PVN in these adult animals did receive a larger number of NPY and CART terminals (301). This was not reflected in a change in PVN CRH expression (565), although the PVN in juvenile rats did show increased neuronal activity and CRH mRNA levels (448, 566). When the maternal diet was continued postnatally, juvenile pups showed very low serum levels of leptin (567). This was accompanied by reduced POMC expression and axons, but surprisingly, hypothalamic NPY expression and its protein levels in the

PVN were normal (567). There does not seem to be a predominant direction in which energy balance regulation is shifted, which is in line with the variation in the general phenotype described above for these animals.

After perinatal maternal overfeeding, changes have also been found in energy balance regulation. In two studies that found normal birth weight after maternal high-fat diet, serum leptin levels and hypothalamic gene expression for Ob-R, NPY, and POMC were either up-regulated or down-regulated at birth (473, 474). In the former case, hypothalamic Ob-R peptide and AgRP and MC4 receptor mRNA were also elevated (473). After a maternal cafeteria diet, a much more pronounced neonatal leptin surge was reported (480). By the time of weaning (when these animals were heavier than controls), serum leptin levels were increased (468, 470, 476, 478). In the hypothalamus, this resulted in down-regulated Ob-Rb mRNA (468), with normal to reduced NPY and AgRP and increased POMC expression (468, 470). But although the ARC response to elevated leptin signaling seemed roughly normal, the VMN showed reduced responsiveness (568). Fasting revealed more changes that were not seen under basal conditions: pups born to high-fat diet-fed dams showed increased elevations in mRNA levels of NPY, AgRP, and the  $Y_1$  receptor but lacked the decrease in MC4 receptor expression that is found in control animals upon fasting (468). From weaning on, leptin levels in these animals were found to be normal to elevated (469, 471, 476, 478, 484), where the larger increases in leptin were usually in more overweight animals. Shortly after weaning, at a point that the rats born to overfed mothers had normal body weights, the number of projections from the ARC to the PVN containing AgRP was reduced, whereas projections containing  $\alpha$ -MSH were normal (480). In addition, gene expression for the MC4 receptor was up-regulated in the VMN, and that for the leptin receptor Ob-R was down-regulated in the ARC (479), accompanied by a reduction in leptin sensitivity that remained at least until the age of 3 months (480). In adulthood, offspring of high-energy-fed dams showed enlargement of the VMN and DMN nuclei (482). In addition, hypothalamic gene expression was either reported to be normal for Ob-R and NPY and reduced for AgRP and POMC (471), or was up-regulated for NPY with normal expression of AgRP and POMC (476). Because in the former study, the reduction of POMC expression seemed stronger than that of AgRP, despite the conflicting details, both profiles could be expected to lead to more orexigenic signaling.

Hypothalamic alterations have also been found in pups born to or cross-fostered to dams with gestational diabetes due to streptozotocin injections. At weaning, despite normal serum leptin levels, postnatal-only exposure increased



the size of the PVN (491), whereas combined pre- and postnatal exposure led to a reduction in PVN and VMN size (490). At the same time, ARC peptide levels of NPY and AgRP were elevated, and those of POMC and  $\alpha$ -MSH were reduced (488, 491), suggestive of increased orexigenic signaling. The up-regulation of NPY levels was also found in middle-aged animals (493).

**b. Nutritional manipulation of the offspring.** In rats that were prenatally growth restricted by uterine artery ligation, NPY mRNA and protein were both increased at weaning, whereas CRH levels were unaffected (511). In young adulthood, the number of ARC cells expressing NPY mRNA was normal, but the levels of expression were reduced (504). This suggests an increased orexigenic drive in the juvenile animals, which is in accordance with the complete catch-up growth reported for these animals (see *Section IV.C.1.b*). Lower NPY expression in the adult rats is concurrent with the incomplete catch-up growth that these animals display (see *Section IV.C.1.b*).

Weanling small-litter rats were shown to have reduced Ob-Rb expression (520), which is in agreement with the high serum leptin levels found in these rats. ARC NPY, AgRP, and CART mRNA levels were all increased, but this predominantly orexigenic signal did not seem to reach the PVN and LHA because expression of TRH, MCH, and ORX was unaltered (520). This was also suggested by the fact that NPY peptide levels in both the ARC and the PVN were normal (522). In young adulthood, expression of the ARC peptides NPY, AgRP, and CART is normal, as well as CRH and TRH expression in the PVN and MCH and ORX expression in the LHA (529, 537). At this age, leptin transport across the blood-brain barrier appears to be impaired (537), which indicates a state of leptin resistance. This leptin resistance seems to develop only after weaning because weanling small-litter rats are still responsive to peripheral injections of leptin (524). Taken together, these studies suggest that the obese phenotype of adult small-litter rats (see *Section IV.C.1.b*) may be at least partly attributable to central leptin resistance caused by high neonatal leptin levels and the resulting hyperproductivity of the ARC. Additionally, studies by Davidowa and colleagues (reviewed in Refs. 27, 278) suggest that in these rats, neurons of several hypothalamic nuclei have an altered response to many of the orexigenic and anorexigenic signals.

Interestingly, juvenile large-litter rats also show more orexigenic signaling. In the ARC, the balance between orexigenic and anorexigenic gene expression was shifted (569), or AgRP and NPY expression and NPY peptide levels were increased with unchanged CART expression (520, 522). This resulted in elevated PVN NPY peptide

levels (522), but did not affect its expression of CRH or TRH, nor LHA ORX expression (520, 569). LHA MCH gene expression was shown to be transiently increased, with elevated levels at d 10, but not at 25 d of age (569). Unlike small-litter rats, juvenile rats raised in large litters had normal hypothalamic Ob-Rb expression (520). Instead, some of the short forms of Ob-R were expressed at increased levels (520). One study suggested that ARC NPY mRNA was no longer elevated in young adulthood (504), although there still seemed to be a small tendency toward higher expression levels. These results generally appear to be in agreement with the phenotype described above. The acute effects of juvenile food restriction seem to be mostly orexigenic, although apparently not enough to achieve full catch-up growth. Information on the long-term effects of this model is still largely missing.

Although the studies described here have all used nutritional manipulations, perinatal nonnutritional manipulations have also been shown to program hypothalamic (an)orexigenic signaling. One example is neonatal stress, which has been shown to have long-term effects on levels of POMC, CRH, ORX, and ORX receptors (570, 571).

**c. The neonatal role of leptin.** As has been mentioned in *Section III.G.1*, leptin is not fully functional in the regulation of energy balance during the neonatal period. Instead, it seems to play a more developmental role. It is responsible for the proper development of intrahypothalamic connections (312, 572) that occurs during the early postnatal period (320). Even general brain development seems to depend on leptin because the brains of leptin-deficient mice show a variety of abnormalities that can be rescued by juvenile leptin treatment (311). In recent years, several researchers have hypothesized that altered neonatal leptin levels may play a key role in developmental programming (12, 15, 573–578). This hypothesis is supported by several recent studies that manipulated perinatal leptin levels.

Interestingly, the direction of the reported effects differed between these studies. Some researchers have found a beneficial effect of perinatal leptin administration on adult body adiposity (579, 580). Moreover, one study reported the absence of an anorexigenic reaction to peripheral leptin in adulthood when leptin action was blocked neonatally (581). In contrast, others have reported increased fat mass, leptin levels, and/or food intake (451, 582–584) and leptin resistance (301, 583, 584) in adult rodents subjected to perinatal leptin administration. Similarly, different effects of perinatal leptin on susceptibility to diet-induced obesity were reported. Some studies found perinatal leptin to be protective against diet-induced obesity (581, 585), whereas others reported in-

creased weight gain on a high-fat diet after neonatal leptin injections (301, 451).

Based on some of the positive effects mentioned above, several groups have investigated whether perinatal leptin administration might rescue the obesity-prone phenotype of rats that were programmed by perinatal nutritional manipulations. Their results have been mixed. Rats that were malnourished by a maternal low-protein diet throughout gestation and lactation had lower body weight in adulthood, similar leptin levels, and similar susceptibility to diet-induced obesity as controls (466). When the low-protein dams were infused with leptin during the perinatal period, weight gain on the high-fat diet was abolished (466). The effect of perinatal leptin on control rats was not investigated in this study, which hampers the interpretation of the results. Notably, the body weight of saline-treated low-protein pups appears to reach normal control levels on the high-fat diet. Another group has investigated the effects of neonatal leptin injections on the obese phenotype of rats subjected to prenatal maternal food restriction. These underfed rats have a higher baseline adiposity (at least the males), and both sexes are more susceptible to diet-induced obesity than control rats (451, 552). When prenatal undernutrition was followed by neonatal leptin injections, baseline adiposity was reduced in males, with no effect on diet-induced obesity (451), whereas in females, neonatal leptin reduced the effects of the high-fat diet to that found in controls, without an effect on baseline adiposity (552). Notably, neonatal leptin injections aggravated diet-induced obesity in control males, but not in control females. A third group attempted to rescue the obese phenotype of weanling rats raised in small litters by using neonatal leptin injections. In female small-litter rats, leptin injections reduced the fat percentage to that of control females raised in normal litters (524). In male rats, however, the fat percentage at weaning was not altered by leptin injections in small-litter animals, whereas it was significantly reduced by leptin in normal-litter males (525). Thus, neonatal leptin rescued the obese phenotype in weanling female small-litter rats, but not in males.

Summarizing, perinatal leptin supplementation can have beneficial or detrimental effects on energy balance and body composition in both normal and programmed rats. The outcome is probably determined by the exact timing and levels of leptin, as well as the phenotypic background of the animal. Therefore, we recommend extreme caution when investigating the option of providing infants with supplemental leptin as a proposed obesity-protective agent (586). An additional concern is the reduction in skeletal growth that was reported in some of the studies, resulting in reduced body length (451, 579), which is usually undesirable in the human situation.

*d. Other programming candidates.* Of course, leptin cannot be the sole factor responsible for programming. For that, the phenomenon is too widespread and its consequences too diverse. Examples of other proposed candidates are thyroid hormone, the IGFs, insulin, and glucocorticoids (587–589). These hormones are important regulators of fetal growth and metabolism, and their levels in the fetus depend on environmental conditions. Besides their own effects on fetal development, levels of thyroid hormone and the IGFs have been shown to be affected by IUGR (587, 588).

In the fetus, insulin correlates strongly with nutrient supplies and is mostly produced by the fetus itself in response to maternal glucose. The effects on the development of energy balance regulation in offspring of diabetic mothers mentioned earlier indicate a developmental role for insulin. Together with its previously mentioned neurotrophic actions (313–315) and the observation that early postnatal intrahypothalamic insulin administration induces altered hypothalamic organization (590), these data have identified insulin as an important candidate for developmental programming (591–593).

In normal pregnancy, the fetus is protected from maternal and environmental glucocorticoids by the placental  $11\beta$ -hydroxysteroid dehydrogenase type 2, which catalyzes the transition of the biologically active cortisol into inactive metabolites (594, 595). Many factors that have been associated with low birth weight and later disease risk (such as hypoxia and stress) also reduce the placental activity of this enzyme and hence increase fetal exposure to glucocorticoids. Many studies have now found programming effects after excess glucocorticoid exposure (594, 595).

In addition to environmental factors that can influence programming, genetic factors have long been suggested to be causal in both low birth weight and later-life disease risks (596). Because of the technical difficulties of such studies, finding evidence for this hypothesis has been problematic. Recently however, large-scale studies have provided compelling support for a common genetic origin of alterations in birth weight and in later health (597, 598). Moreover, as we have seen above in the rare studies that have used animal models with different genetic backgrounds (482), the phenotype observed after nutritional manipulations can be modulated considerably by genetic factors.

In conclusion, leptin is an important candidate, but certainly not the only one, for the mechanistic underpinnings of developmental programming. It seems likely that the relative importance of the different candidates varies according to the environmental conditions. Whether experimental manipulation or naturally occurring, each set of

conditions is likely to elicit specific responses of the different candidates and hence bring about its specific phenotypical outcome.

#### 4. Summary and implications

In this section, we have seen ample evidence for developmental programming of energy balance and its hypothalamic regulation in experimental animals. Studies have investigated outcomes in all three categories of measurements of energy balance (*i.e.*, body size and composition, energy intake and expenditure, and hypothalamic neuropeptides). Persistent changes have been found in all of these parameters in various rodent models with perinatal manipulations of nutrition. Nevertheless, the precise effects that have been reported differed considerably, not only between the models, but also between studies using similar models. Many of these apparent discrepancies can be explained by (small) differences between studies in the timing, nature, and severity of the manipulation or other subtle variations in their methods, such as the genetic background, sex, and age of the experimental animals. When we focus on the similarities between the studies discussed in this section, rather than their inconsistencies, a few generalizations can be made.

The long-term effects initiated by perinatal overnutrition seem to be quite consistent, independent of whether the exposure to overfeeding starts prenatally (*e.g.*, maternal diet) or postnatally (small litter size). Although the details vary, when an effect is found, it is generally a shift toward overweight, higher adiposity, more food intake, and more orexigenic signaling.

In the case of undernutrition, the exact perinatal timing combined with the degree of undernutrition seems to play an important role. Overweight in adulthood is seen more often after prenatal-only than after postnatal undernutrition. The thrifty phenotype, with a higher food intake, energy storage, and concomitant susceptibility to obesity, is proposed to be the result of an inappropriate “adaptive” response to an adult environment that turns out to be richer than expected (2, 6). There are several possible reasons why this thrifty phenotype is seen less often after postnatal undernutrition. First, postnatal underfeeding may fail to induce a predictive adaptive response, because of an insufficient degree of undernutrition to elicit such a response or maybe because the degree of undernutrition is so severe that it abolishes any adaptive response by disruption of developmental processes in the pups (such severe damage can also be imagined when prenatal undernutrition is continued into the neonatal period). Alternatively, it might be that in rodents the critical time-window of plasticity already closes before the postnatal undernutrition could induce any adaptive response. In case of prenatal malnutrition, postnatal growth may show

either catch-up or lack of catch-up, presumably dependent on the adaptation of the organism to the insult. Unfortunately, in epidemiological and experimental studies alike, it is difficult to identify the contributions of different underlying causes to the observed phenotype.

The obesogenic effect of rapid catch-up growth and of perinatal overfeeding models may originate from programming of the set-points of energy balance regulation according to the circulating (high) levels in perinatal life of regulators such as insulin and leptin (589, 592). In most models of perinatal underfeeding, the hypothalamic effects observed in juvenile life (despite variations in the details) seem to be directed at promoting positive energy balance, and hence may be aimed at catch-up growth. These hypothalamic adaptations are also seen in the absence of adult overweight. The lack of obesogenic effects of this increased orexigenic signaling might be the result of irreversible damage to the hypothalamic or downstream circuitry that interferes with the putative predictive adaptive response. This could also be true for the disagreement between the studies using uterine artery ligation, where later ligation (*i.e.*, milder damage) results more often in an obese phenotype than ligation at an earlier time-point (*i.e.*, stronger damage) in gestation.

Although developmental events in the early postnatal period in rats resemble those in the third trimester of human pregnancy (355), rapid neonatal growth seems to program for later obesity in both species. Therefore, it might be postulated that the time-windows for hypothalamic plasticity may be similar (or at least extend until similar time-points) for both species. A striking difference between the evidence in humans and rodents is that in humans both extremes of perinatal nutrition (under- and overfeeding) seem to elicit consistent obesogenic effects, whereas in rodents the directions of long-term effects vary between higher and lower rates of overweight and obesity. This species difference may result from (interaction with) the obesogenic environment that most humans (unlike most experimental animals) encounter in later life. However, the fact that not all perinatally over- or underfed rodents are more susceptible to diet-induced obesity suggests that other factors must also play a role. Experiments by Levin and colleagues (482) suggest that the three-way interaction between perinatal nutrition, later obesogenic environment, and genetic background may explain many of the observed effects.

Because of its distinct role during development, leptin has been hypothesized to play a major role in programming of energy balance regulation. Evidence supporting this hypothesis has been published in recent years, but there is also strong evidence for the involvement of other factors besides leptin. With regard to the effects of neo-



natal leptin injections, the direction of its effects seems to vary strongly with a number of factors, some of which may even be unknown at present. Among these, the exact timing of administration, the initial state of energy balance, and the genetic background may be especially important. With so many aspects still unknown, proposals to supplement leptin as an obesity-protective agent seem rather premature and should be considered with extreme caution.

## V. Concluding Remarks

In this review, we have presented the concept of developmental programming. It explains how changes in the environment during a critical time-window in early development can permanently alter the phenotype of an organism. In this manner, individuals can be “fine-tuned” to their expected future environment. Although there has been controversy on this subject, it is now generally believed that energy balance and its regulation can also be programmed. We have discussed a substantial number of studies that have investigated developmental programming of energy balance in different species, using different techniques, and from different angles. From these studies, we can conclude that early nutrition can truly program energy balance and its regulation in both humans and animals. The direction of the programming effects that were reported appears to be variable and dependent on the environment—both the perinatal and the adult environment. One thing that becomes apparent from the discussed animal studies is that developmental programming of energy balance does not necessarily entail detrimental changes; in some cases, the programmed changes were favorable, such as reductions in fat mass. In contrast, mostly adverse effects on adult body composition were reported in humans. This striking disparity, between metabolic effects with different directions in animal models on one hand and consistent detrimental effects in humans on the other, may result from (interaction with) the obesogenic environment that most humans (unlike most experimental animals) encounter in later life.

With the inconsistencies between animal studies, a comprehensive picture of the impact of perinatal nutrition on energy balance in later life has thus far remained elusive. If we intend to extrapolate conclusions between studies, and from animal models to the human situation, it is vital to identify the exact circumstances leading to each outcome and to standardize the variable methodology that researchers have used to investigate this subject.

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Address all correspondence and requests for reprints to: Floor Remmers, Institute of Physiological Chemistry, University Medical Center of the Johannes Gutenberg University Mainz, Duesbergweg 6, 55128 Mainz, Germany. E-mail: remmersf@uni-mainz.de.

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## References

1. Barker DJ 1992 The fetal origins of diseases of old age. *Eur J Clin Nutr* 46(Suppl 3):S3–S9
2. Gluckman PD, Hanson MA 2004 The developmental origins of the metabolic syndrome. *Trends Endocrinol Metab* 15:183–187
3. Gluckman PD, Beedle AS, Hanson MA, Vickers MH 2007 Leptin reversal of the metabolic phenotype: evidence for the role of developmental plasticity in the development of the metabolic syndrome. *Horm Res* 67(Suppl 1):115–120
4. Hales CN, Barker DJ 1992 Type 2 (non-insulin-dependent) diabetes mellitus: the thrifty phenotype hypothesis. *Diabetologia* 35:595–601
5. Lucas A 1991 Programming by early nutrition in man. *Ciba Found Symp* 156:38–50; discussion 50–55
6. Barker DJ 2007 The origins of the developmental origins theory. *J Intern Med* 261:412–417
7. Dunger DB, Ong KK 2005 Endocrine and metabolic consequences of intrauterine growth retardation. *Endocrinol Metab Clin North Am* 34:597–615, ix
8. Fernandez-Twinn DS, Ozanne SE 2006 Mechanisms by which poor early growth programs type-2 diabetes, obesity and the metabolic syndrome. *Physiol Behav* 88:234–243
9. Levy-Marchal C, Jaquet D 2004 Long-term metabolic consequences of being born small for gestational age. *Pediatr Diabetes* 5:147–153
10. Roseboom T, de Rooij S, Painter R 2006 The Dutch famine and its long-term consequences for adult health. *Early Hum Dev* 82:485–491
11. Cornier MA, Dabelea D, Hernandez TL, Lindstrom RC, Steig AJ, Stob NR, Van Pelt RE, Wang H, Eckel RH 2008 The metabolic syndrome. *Endocr Rev* 29:777–822
12. Cripps RL, Martin-Gronert MS, Ozanne SE 2005 Fetal and perinatal programming of appetite. *Clin Sci (Lond)* 109:1–11
13. Martorell R, Stein AD, Schroeder DG 2001 Early nutrition and later adiposity. *J Nutr* 131:874S–880S
14. Ong KK 2006 Size at birth, postnatal growth and risk of obesity. *Horm Res* 65(Suppl 3):65–69
15. Ozanne SE, Lewis R, Jennings BJ, Hales CN 2004 Early programming of weight gain in mice prevents the induction of obesity by a highly palatable diet. *Clin Sci (Lond)* 106:141–145
16. Curhan GC, Chertow GM, Willett WC, Spiegelman D, Colditz GA, Manson JE, Speizer FE, Stampfer MJ 1996 Birth weight and adult hypertension and obesity in women. *Circulation* 94:1310–1315
17. Curhan GC, Willett WC, Rimm EB, Spiegelman D, Ascherio AL, Stampfer MJ 1996 Birth weight and adult hypertension, diabetes mellitus, and obesity in US men. *Circulation* 94:3246–3250
18. Kahn HS, Narayan KM, Williamson DF, Valdez R 2000

- Relation of birth weight to lean and fat thigh tissue in young men. *Int J Obes Relat Metab Disord* 24:667–672
19. Ravelli GP, Stein ZA, Susser MW 1976 Obesity in young men after famine exposure in utero and early infancy. *N Engl J Med* 295:349–353
  20. Hales CN, Barker DJ 2001 The thrifty phenotype hypothesis. *Br Med Bull* 60:5–20
  21. Harding JE 2001 The nutritional basis of the fetal origins of adult disease. *Int J Epidemiol* 30:15–23
  22. Metcalfe NB, Monaghan P 2001 Compensation for a bad start: grow now, pay later? *Trends Ecol Evol* 16:254–260
  23. West-Eberhard MJ 1989 Phenotypic plasticity and the origins of diversity. *Annu Rev Ecol Syst* 20:249–278
  24. Henriksen T, Clausen T 2002 The fetal origins hypothesis: placental insufficiency and inheritance versus maternal malnutrition in well-nourished populations. *Acta Obstet Gynecol Scand* 81:112–114
  25. Fowden AL, Giussani DA, Forhead AJ 2006 Intrauterine programming of physiological systems: causes and consequences. *Physiology (Bethesda)* 21:29–37
  26. Gluckman PD, Hanson MA 2008 Developmental and epigenetic pathways to obesity: an evolutionary-developmental perspective. *Int J Obes (Lond)* 32(Suppl 7):S62–S71
  27. Grove KL, Grayson BE, Glavas MM, Xiao XQ, Smith MS 2005 Development of metabolic systems. *Physiol Behav* 86:646–660
  28. Bertram CE, Hanson MA 2001 Animal models and programming of the metabolic syndrome. *Br Med Bull* 60:103–121
  29. Holemans K, Aerts L, Van Assche FA 2003 Fetal growth restriction and consequences for the offspring in animal models. *J Soc Gynecol Investig* 10:392–399
  30. Nathanielsz PW 2006 Animal models that elucidate basic principles of the developmental origins of adult diseases. *ILAR J* 47:73–82
  31. Ozanne SE 2001 Metabolic programming in animals. *Br Med Bull* 60:143–152
  32. Spiegelman BM, Flier JS 2001 Obesity and the regulation of energy balance. *Cell* 104:531–543
  33. Berthoud HR, Morrison C 2008 The brain, appetite, and obesity. *Annu Rev Psychol* 59:55–92
  34. Saper CB, Chou TC, Elmquist JK 2002 The need to feed: homeostatic and hedonic control of eating. *Neuron* 36:199–211
  35. Anand BK, Brobeck JR 1951 Localization of a “feeding center” in the hypothalamus of the rat. *Proc Soc Exp Biol Med* 77:323–324
  36. Hetherington AW, Ranson SW 1940 Hypothalamic lesions and adiposity in the rat. *Anat Rec* 78:149–172
  37. Luiten PG, ter Horst GJ, Steffens AB 1987 The hypothalamus, intrinsic connections and outflow pathways to the endocrine system in relation to the control of feeding and metabolism. *Prog Neurobiol* 28:1–54
  38. Dumont Y, Jacques D, Bouchard P, Quirion R 1998 Species differences in the expression and distribution of the neuropeptide Y Y1, Y2, Y4, and Y5 receptors in rodents, guinea pig, and primates brains. *J Comp Neurol* 402:372–384
  39. Elias CF, Saper CB, Maratos-Flier E, Tritos NA, Lee C, Kelly J, Tatro JB, Hoffman GE, Ollmann MM, Barsh GS, Sakurai T, Yanagisawa M, Elmquist JK 1998 Chemically defined projections linking the mediobasal hypothalamus and the lateral hypothalamic area. *J Comp Neurol* 402:442–459
  40. Elias CF, Lee CE, Kelly JF, Ahima RS, Kuhar M, Saper CB, Elmquist JK 2001 Characterization of CART neurons in the rat and human hypothalamus. *J Comp Neurol* 432:1–19
  41. Koutcherov Y, Mai JK, Paxinos G 2003 Hypothalamus of the human fetus. *J Chem Neuroanat* 26:253–270
  42. Menyhárt J, Wittmann G, Hrabovszky E, Keller E, Liposits Z, Fekete C 2006 Interconnection between orexigenic neuropeptide Y- and anorexigenic  $\alpha$ -melanocyte stimulating hormone-synthesizing neuronal systems of the human hypothalamus. *Brain Res* 1076:101–105
  43. Mihály E, Fekete C, Tatro JB, Liposits Z, Stopa EG, Lechan RM 2000 Hypophysiotropic thyrotropin-releasing hormone-synthesizing neurons in the human hypothalamus are innervated by neuropeptide Y, agouti-related protein, and  $\alpha$ -melanocyte-stimulating hormone. *J Clin Endocrinol Metab* 85:2596–2603
  44. Schwartz MW, Woods SC, Porte Jr D, Seeley RJ, Baskin DG 2000 Central nervous system control of food intake. *Nature* 404:661–671
  45. Halaas JL, Gajiwala KS, Maffei M, Cohen SL, Chait BT, Rabinowitz D, Lallone RL, Burley SK, Friedman JM 1995 Weight-reducing effects of the plasma protein encoded by the obese gene. *Science* 269:543–546
  46. Zhang Y, Proenca R, Maffei M, Barone M, Leopold L, Friedman JM 1994 Positional cloning of the mouse obese gene and its human homologue. *Nature* 372:425–432
  47. Benoit SC, Clegg DJ, Seeley RJ, Woods SC 2004 Insulin and leptin as adiposity signals. *Recent Prog Horm Res* 59:267–285
  48. Considine RV, Sinha MK, Heimann ML, Kriauciunas A, Stephens TW, Nyce MR, Ohannesian JP, Marco CC, McKee LJ, Bauer TL, Caro JF 1996 Serum immunoreactive-leptin concentrations in normal-weight and obese humans. *N Engl J Med* 334:292–295
  49. Frederich RC, Hamann A, Anderson S, Löllmann B, Lowell BB, Flier JS 1995 Leptin levels reflect body lipid content in mice: evidence for diet-induced resistance to leptin action. *Nat Med* 1:1311–1314
  50. Landt M, Gingerich RL, Havel PJ, Mueller WM, Schoner B, Hale JE, Heiman ML 1998 Radioimmunoassay of rat leptin: sexual dimorphism reversed from humans. *Clin Chem* 44:565–570
  51. Maffei M, Halaas J, Ravussin E, Pratley RE, Lee GH, Zhang Y, Fei H, Kim S, Lallone R, Ranganathan S, Kern PA, Friedman JM 1995 Leptin levels in human and rodent: measurement of plasma leptin and ob RNA in obese and weight-reduced subjects. *Nat Med* 1:1155–1161
  52. Ostlund Jr RE, Yang JW, Klein S, Gingerich R 1996 Relation between plasma leptin concentration and body fat, gender, diet, age, and metabolic covariates. *J Clin Endocrinol Metab* 81:3909–3913
  53. Elmquist JK, Bjørbaek C, Ahima RS, Flier JS, Saper CB 1998 Distributions of leptin receptor mRNA isoforms in the rat brain. *J Comp Neurol* 395:535–547
  54. Fei H, Okano HJ, Li C, Lee GH, Zhao C, Darnell R, Friedman JM 1997 Anatomic localization of alternatively spliced leptin receptors (Ob-R) in mouse brain and other tissues. *Proc Natl Acad Sci USA* 94:7001–7005
  55. Guan XM, Hess JF, Yu H, Hey PJ, van der Ploeg LH 1997

- Differential expression of mRNA for leptin receptor isoforms in the rat brain. *Mol Cell Endocrinol* 133:1–7
56. Bjørbaek C, Elmquist JK, Michl P, Ahima RS, van Bueren A, McCall AL, Flier JS 1998 Expression of leptin receptor isoforms in rat brain microvessels. *Endocrinology* 139:3485–3491
  57. Boden G, Chen X, Mozzoli M, Ryan I 1996 Effect of fasting on serum leptin in normal human subjects. *J Clin Endocrinol Metab* 81:3419–3423
  58. Frederich RC, Löllmann B, Hamann A, Napolitano-Rosen A, Kahn BB, Lowell BB, Flier JS 1995 Expression of ob mRNA and its encoded protein in rodents. Impact of nutrition and obesity. *J Clin Invest* 96:1658–1663
  59. Mizuno TM, Bergen H, Funabashi T, Kleopoulos SP, Zhong YG, Bauman WA, Mobbs CV 1996 Obese gene expression: reduction by fasting and stimulation by insulin and glucose in lean mice, and persistent elevation in acquired (diet-induced) and genetic (yellow agouti) obesity. *Proc Natl Acad Sci USA* 93:3434–3438
  60. Weigle DS, Duell PB, Connor WE, Steiner RA, Soules MR, Kuijper JL 1997 Effect of fasting, refeeding, and dietary fat restriction on plasma leptin levels. *J Clin Endocrinol Metab* 82:561–565
  61. Mistry AM, Swick AG, Romsos DR 1997 Leptin rapidly lowers food intake and elevates metabolic rates in lean and ob/ob mice. *J Nutr* 127:2065–2072
  62. Schwartz MW, Seeley RJ, Campfield LA, Burn P, Baskin DG 1996 Identification of targets of leptin action in rat hypothalamus. *J Clin Invest* 98:1101–1106
  63. Seeley RJ, van Dijk G, Campfield LA, Smith FJ, Burn P, Nelligan JA, Bell SM, Baskin DG, Woods SC, Schwartz MW 1996 Intraventricular leptin reduces food intake and body weight of lean rats but not obese Zucker rats. *Horm Metab Res* 28:664–668
  64. Cummings DE 2006 Ghrelin and the short- and long-term regulation of appetite and body weight. *Physiol Behav* 89:71–84
  65. Murphy KG, Bloom SR 2006 Gut hormones and the regulation of energy homeostasis. *Nature* 444:854–859
  66. Stanley S, Wynne K, McGowan B, Bloom S 2005 Hormonal regulation of food intake. *Physiol Rev* 85:1131–1158
  67. Faouzi M, Leshan R, Björnholm M, Hennessey T, Jones J, Münzberg H 2007 Differential accessibility of circulating leptin to individual hypothalamic sites. *Endocrinology* 148:5414–5423
  68. Peruzzo B, Pastor FE, Blázquez JL, Schöbitz K, Peláez B, Amat P, Rodríguez EM 2000 A second look at the barriers of the medial basal hypothalamus. *Exp Brain Res* 132:10–26
  69. Banks WA, Kastin AJ, Huang W, Jaspan JB, Maness LM 1996 Leptin enters the brain by a saturable system independent of insulin. *Peptides* 17:305–311
  70. Banks WA, Tschöp M, Robinson SM, Heiman ML 2002 Extent and direction of ghrelin transport across the blood-brain barrier is determined by its unique primary structure. *J Pharmacol Exp Ther* 302:822–827
  71. Schwartz MW, Bergman RN, Kahn SE, Taborsky Jr GJ, Fisher LD, Sipols AJ, Woods SC, Steil GM, Porte Jr D 1991 Evidence for entry of plasma insulin into cerebrospinal fluid through an intermediate compartment in dogs. Quantitative aspects and implications for transport. *J Clin Invest* 88:1272–1281
  72. Broberger C, Johansen J, Johansson C, Schalling M, Hökfelt T 1998 The neuropeptide Y/agouti gene-related protein (AGRP) brain circuitry in normal, anorectic, and monosodium glutamate-treated mice. *Proc Natl Acad Sci USA* 95:15043–15048
  73. Gehlert DR, Chronwall BM, Schafer MP, O'Donohue TL 1987 Localization of neuropeptide Y messenger ribonucleic acid in rat and mouse brain by in situ hybridization. *Synapse* 1:25–31
  74. Hahn TM, Breininger JF, Baskin DG, Schwartz MW 1998 Coexpression of Agrp and NPY in fasting-activated hypothalamic neurons. *Nat Neurosci* 1:271–272
  75. Larhammar D, Ericsson A, Persson H 1987 Structure and expression of the rat neuropeptide Y gene. *Proc Natl Acad Sci USA* 84:2068–2072
  76. Elias CF, Lee C, Kelly J, Aschkenasi C, Ahima RS, Couceyro PR, Kuhar MJ, Saper CB, Elmquist JK 1998 Leptin activates hypothalamic CART neurons projecting to the spinal cord. *Neuron* 21:1375–1385
  77. Gee CE, Chen CL, Roberts JL, Thompson R, Watson SJ 1983 Identification of proopiomelanocortin neurons in rat hypothalamus by in situ cDNA-mRNA hybridization. *Nature* 306:374–376
  78. Baskin DG, Schwartz MW, Seeley RJ, Woods SC, Porte Jr D, Breininger JF, Jonak Z, Schaefer J, Krouse M, Burghardt C, Campfield LA, Burn P, Kochan JP 1999 Leptin receptor long-form splice-variant protein expression in neuron cell bodies of the brain and co-localization with neuropeptide Y mRNA in the arcuate nucleus. *J Histochem Cytochem* 47:353–362
  79. Cheung CC, Clifton DK, Steiner RA 1997 Proopiomelanocortin neurons are direct targets for leptin in the hypothalamus. *Endocrinology* 138:4489–4492
  80. Benoit SC, Air EL, Coolen LM, Strauss R, Jackman A, Clegg DJ, Seeley RJ, Woods SC 2002 The catabolic action of insulin in the brain is mediated by melanocortins. *J Neurosci* 22:9048–9052
  81. Könnner AC, Janoschek R, Plum L, Jordan SD, Rother E, Ma X, Xu C, Enriori P, Hampel B, Barsh GS, Kahn CR, Cowley MA, Ashcroft FM, Brüning JC 2007 Insulin action in AgRP-expressing neurons is required for suppression of hepatic glucose production. *Cell Metab* 5:438–449
  82. van den Top M, Lee K, Whyment AD, Blanks AM, Spanswick D 2004 Orexin-sensitive NPY/AgRP pacemaker neurons in the hypothalamic arcuate nucleus. *Nat Neurosci* 7:493–494
  83. Willeesen MG, Kristensen P, Rømer J 1999 Co-localization of growth hormone secretagogue receptor and NPY mRNA in the arcuate nucleus of the rat. *Neuroendocrinology* 70:306–316
  84. Allen YS, Adrian TE, Allen JM, Tatemoto K, Crow TJ, Bloom SR, Polak JM 1983 Neuropeptide Y distribution in the rat brain. *Science* 221:877–879
  85. Bagnol D, Lu XY, Kaelin CB, Day HE, Ollmann M, Gantz I, Akil H, Barsh GS, Watson SJ 1999 Anatomy of an endogenous antagonist: relationship between Agouti-related protein and proopiomelanocortin in brain. *J Neurosci* 19:RC26
  86. Broberger C 1999 Hypothalamic cocaine- and amphetamine-regulated transcript (CART) neurons: histochemical



- relationship to thyrotropin-releasing hormone, melanin-concentrating hormone, orexin/hypocretin and neuropeptide Y. *Brain Res* 848:101–113
87. Fetissov SO, Kopp J, Hökfelt T 2004 Distribution of NPY receptors in the hypothalamus. *Neuropeptides* 38:175–188
  88. Parker RM, Herzog H 1999 Regional distribution of Y-receptor subtype mRNAs in rat brain. *Eur J Neurosci* 11:1431–1448
  89. Broberger C, Landry M, Wong H, Walsh JN, Hökfelt T 1997 Subtypes Y1 and Y2 of the neuropeptide Y receptor are respectively expressed in pro-opiomelanocortin- and neuropeptide-Y-containing neurons of the rat hypothalamic arcuate nucleus. *Neuroendocrinology* 66:393–408
  90. Fetissov SO, Byrne LC, Hassani H, Ernfors P, Hökfelt T 2004 Characterization of neuropeptide Y Y2 and Y5 receptor expression in the mouse hypothalamus. *J Comp Neurol* 470:256–265
  91. King PJ, Widdowson PS, Doods HN, Williams G 1999 Regulation of neuropeptide Y release by neuropeptide Y receptor ligands and calcium channel antagonists in hypothalamic slices. *J Neurochem* 73:641–646
  92. Chen AS, Marsh DJ, Trumbauer ME, Frazier EG, Guan XM, Yu H, Rosenblum CI, Vongs A, Feng Y, Cao L, Metzger JM, Strack AM, Camacho RE, Mellin TN, Nunes CN, Min W, Fisher J, Gopal-Truter S, MacIntyre DE, Chen HY, Van der Ploeg LH 2000 Inactivation of the mouse melanocortin-3 receptor results in increased fat mass and reduced lean body mass. *Nat Genet* 26:97–102
  93. Harrold JA, Widdowson PS, Williams G 1999 Altered energy balance causes selective changes in melanocortin-4 (MC4-R), but not melanocortin-3 (MC3-R), receptors in specific hypothalamic regions: further evidence that activation of MC4-R is a physiological inhibitor of feeding. *Diabetes* 48:267–271
  94. Huszar D, Lynch CA, Fairchild-Huntress V, Dunmore JH, Fang Q, Berkemeier LR, Gu W, Kesterson RA, Boston BA, Cone RD, Smith FJ, Campfield LA, Burn P, Lee F 1997 Targeted disruption of the melanocortin-4 receptor results in obesity in mice. *Cell* 88:131–141
  95. Marks DL, Hruby V, Brookhart G, Cone RD 2006 The regulation of food intake by selective stimulation of the type 3 melanocortin receptor (MC3R). *Peptides* 27:259–264
  96. Mountjoy KG, Mortrud MT, Low MJ, Simerly RB, Cone RD 1994 Localization of the melanocortin-4 receptor (MC4-R) in neuroendocrine and autonomic control circuits in the brain. *Mol Endocrinol* 8:1298–1308
  97. Roselli-Reh fuss L, Mountjoy KG, Robbins LS, Mortrud MT, Low MJ, Tatro JB, Entwistle ML, Simerly RB, Cone RD 1993 Identification of a receptor for  $\gamma$  melanotropin and other proopiomelanocortin peptides in the hypothalamus and limbic system. *Proc Natl Acad Sci USA* 90:8856–8860
  98. Haskell-Luevano C, Monck EK 2001 Agouti-related protein functions as an inverse agonist at a constitutively active brain melanocortin-4 receptor. *Regul Pept* 99:1–7
  99. Nijenhuis WA, Oosterom J, Adan RA 2001 AgRP(83–132) acts as an inverse agonist on the human-melanocortin-4 receptor. *Mol Endocrinol* 15:164–171
  100. Vicentic A, Jones DC 2007 The CART (cocaine- and amphetamine-regulated transcript) system in appetite and drug addiction. *J Pharmacol Exp Ther* 320:499–506
  101. Jones DC, Kuhar MJ 2008 CART receptor binding in primary cell cultures of the rat nucleus accumbens. *Synapse* 62:122–127
  102. Ahima RS, Kelly J, Elmquist JK, Flier JS 1999 Distinct physiologic and neuronal responses to decreased leptin and mild hyperleptinemia. *Endocrinology* 140:4923–4931
  103. Bertile F, Oudart H, Criscuolo F, Maho YL, Raclot T 2003 Hypothalamic gene expression in long-term fasted rats: relationship with body fat. *Biochem Biophys Res Commun* 303:1106–1113
  104. Brady LS, Smith MA, Gold PW, Herkenham M 1990 Altered expression of hypothalamic neuropeptide mRNAs in food-restricted and food-deprived rats. *Neuroendocrinology* 52:441–447
  105. Kalra SP, Dube MG, Sahu A, Phelps CP, Kalra PS 1991 Neuropeptide Y secretion increases in the paraventricular nucleus in association with increased appetite for food. *Proc Natl Acad Sci USA* 88:10931–10935
  106. Korner J, Savontaus E, Chua Jr SC, Leibel RL, Wardlaw SL 2001 Leptin regulation of *Agrp* and *Npy* mRNA in the rat hypothalamus. *J Neuroendocrinol* 13:959–966
  107. Stephens TW, Basinski M, Bristow PK, Bue-Valleskey JM, Burgett SG, Craft L, Hale J, Hoffmann J, Hsiung HM, Kriauciunas A 1995 The role of neuropeptide Y in the antiobesity action of the obese gene product. *Nature* 377:530–532
  108. Crowley WR, Ramoz G, Torto R, Kalra SP 2004 Role of leptin in orexigenic neuropeptide expression during lactation in rats. *J Neuroendocrinol* 16:637–644
  109. Stanley BG, Chin AS, Leibowitz SF 1985 Feeding and drinking elicited by central injection of neuropeptide Y: evidence for a hypothalamic site(s) of action. *Brain Res Bull* 14:521–524
  110. Stanley BG, Kyrkouli SE, Lampert S, Leibowitz SF 1986 Neuropeptide Y chronically injected into the hypothalamus: a powerful neurochemical inducer of hyperphagia and obesity. *Peptides* 7:1189–1192
  111. Stanley BG, Magdalin W, Seirafi A, Thomas WJ, Leibowitz SF 1993 The perifornical area: the major focus of (a) patchily distributed hypothalamic neuropeptide Y-sensitive feeding system(s). *Brain Res* 604:304–317
  112. Billington CJ, Briggs JE, Harker S, Grace M, Levine AS 1994 Neuropeptide Y in hypothalamic paraventricular nucleus: a center coordinating energy metabolism. *Am J Physiol* 266:R1765–R1770
  113. Egawa M, Yoshimatsu H, Bray GA 1991 Neuropeptide Y suppresses sympathetic activity to interscapular brown adipose tissue in rats. *Am J Physiol* 260:R328–R334
  114. Ollmann MM, Wilson BD, Yang YK, Kerns JA, Chen Y, Gantz I, Barsh GS 1997 Antagonism of central melanocortin receptors in vitro and in vivo by agouti-related protein. *Science* 278:135–138
  115. Shutter JR, Graham M, Kinsey AC, Scully S, Lüthy R, Stark KL 1997 Hypothalamic expression of ART, a novel gene related to agouti, is up-regulated in obese and diabetic mutant mice. *Genes Dev* 11:593–602
  116. Li JY, Finniss S, Yang YK, Zeng Q, Qu SY, Barsh G, Dickinson C, Gantz I 2000 Agouti-related protein-like immunoreactivity: characterization of release from hypothalamic tissue and presence in serum. *Endocrinology* 141:1942–1950
  117. Mizuno TM, Mobbs CV 1999 Hypothalamic agouti-re-

lated protein messenger ribonucleic acid is inhibited by leptin and stimulated by fasting. *Endocrinology* 140:814–817

118. Takahashi KA, Cone RD 2005 Fasting induces a large, leptin-dependent increase in the intrinsic action potential frequency of orexigenic arcuate nucleus neuropeptide Y/agouti-related protein neurons. *Endocrinology* 146:1043–1047
119. Hwa JJ, Ghibaudi L, Gao J, Parker EM 2001 Central melanocortin system modulates energy intake and expenditure of obese and lean Zucker rats. *Am J Physiol Regul Integr Comp Physiol* 281:R444–R451
120. Jonsson L, Skarphedinsson JO, Skuladottir GV, Watanobe H, Schiöth HB 2002 Food conversion is transiently affected during 4-week chronic administration of melanocortin agonist and antagonist in rats. *J Endocrinol* 173:517–523
121. Small CJ, Liu YL, Stanley SA, Connoley IP, Kennedy A, Stock MJ, Bloom SR 2003 Chronic CNS administration of Agouti-related protein (Agrp) reduces energy expenditure. *Int J Obes Relat Metab Disord* 27:530–533
122. Hagan MM, Benoit SC, Rushing PA, Pritchard LM, Woods SC, Seeley RJ 2001 Immediate and prolonged patterns of agouti-related peptide-(83–132)-induced c-Fos activation in hypothalamic and extrahypothalamic sites. *Endocrinology* 142:1050–1056
123. Lu XY, Nicholson JR, Akil H, Watson SJ 2001 Time course of short-term and long-term orexigenic effects of Agouti-related protein (86–132). *Neuroreport* 12:1281–1284
124. Pritchard LE, White A 2005 Agouti-related protein: more than a melanocortin-4 receptor antagonist? *Peptides* 26:1759–1770
125. Mains RE, Eipper BA 1979 Synthesis and secretion of corticotropins, melanotropins, and endorphins by rat intermediate pituitary cells. *J Biol Chem* 254:7885–7894
126. Hagan MM, Rushing PA, Schwartz MW, Yagaloff KA, Burn P, Woods SC, Seeley RJ 1999 Role of the CNS melanocortin system in the response to overfeeding. *J Neurosci* 19:2362–2367
127. van Dijk G, Seeley RJ, Thiele TE, Friedman MI, Ji H, Wilkinson CW, Burn P, Campfield LA, Tenenbaum R, Baskin DG, Woods SC, Schwartz MW 1999 Metabolic, gastrointestinal, and CNS neuropeptide effects of brain leptin administration in the rat. *Am J Physiol* 276:R1425–R1433
128. Cowley MA, Smart JL, Rubinstein M, Cerdán MG, Diano S, Horvath TL, Cone RD, Low MJ 2001 Leptin activates anorexigenic POMC neurons through a neural network in the arcuate nucleus. *Nature* 411:480–484
129. Kim MS, Small CJ, Stanley SA, Morgan DG, Seal LJ, Kong WM, Edwards CM, Abusnana S, Sunter D, Ghatei MA, Bloom SR 2000 The central melanocortin system affects the hypothalamo-pituitary thyroid axis and may mediate the effect of leptin. *J Clin Invest* 105:1005–1011
130. Haynes WG, Morgan DA, Djalali A, Sivitz WI, Mark AL 1999 Interactions between the melanocortin system and leptin in control of sympathetic nerve traffic. *Hypertension* 33:542–547
131. Li G, Zhang Y, Wilsey JT, Scarpace PJ 2004 Unabated anorexic and enhanced thermogenic responses to melanotan II in diet-induced obese rats despite reduced melanocortin 3 and 4 receptor expression. *J Endocrinol* 182:123–132
132. Murphy B, Nunes CN, Ronan JJ, Hanaway M, Fairhurst AM, Mellin TN 2000 Centrally administered MTII affects feeding, drinking, temperature, and activity in the Sprague-Dawley rat. *J Appl Physiol* 89:273–282
133. Tsujii S, Bray GA 1989 Acetylation alters the feeding response to MSH and  $\beta$ -endorphin. *Brain Res Bull* 23:165–169
134. Kim MS, Rossi M, Abusnana S, Sunter D, Morgan DG, Small CJ, Edwards CM, Heath MM, Stanley SA, Seal LJ, Bhatti JR, Smith DM, Ghatei MA, Bloom SR 2000 Hypothalamic localization of the feeding effect of agouti-related peptide and  $\alpha$ -melanocyte-stimulating hormone. *Diabetes* 49:177–182
135. Douglass J, McKinzie AA, Couceyro P 1995 PCR differential display identifies a rat brain mRNA that is transcriptionally regulated by cocaine and amphetamine. *J Neurosci* 15:2471–2481
136. Spiess J, Villarreal J, Vale W 1981 Isolation and sequence analysis of a somatostatin-like polypeptide from ovine hypothalamus. *Biochemistry* 20:1982–1988
137. Kristensen P, Judge ME, Thim L, Ribel U, Christjansen KN, Wulff BS, Clausen JT, Jensen PB, Madsen OD, Vrang N, Larsen PJ, Hastrup S 1998 Hypothalamic CART is a new anorectic peptide regulated by leptin. *Nature* 393:72–76
138. Wortley KE, Chang GQ, Davydova Z, Fried SK, Leibowitz SF 2004 Cocaine- and amphetamine-regulated transcript in the arcuate nucleus stimulates lipid metabolism to control body fat accrual on a high-fat diet. *Regul Pept* 117:89–99
139. Abbott CR, Rossi M, Wren AM, Murphy KG, Kennedy AR, Stanley SA, Zollner AN, Morgan DG, Morgan I, Ghatei MA, Small CJ, Bloom SR 2001 Evidence of an orexigenic role for cocaine- and amphetamine-regulated transcript after administration into discrete hypothalamic nuclei. *Endocrinology* 142:3457–3463
140. Stanley SA, Small CJ, Murphy KG, Rayes E, Abbott CR, Seal LJ, Morgan DG, Sunter D, Dakin CL, Kim MS, Hunter R, Kuhar M, Ghatei MA, Bloom SR 2001 Actions of cocaine- and amphetamine-regulated transcript (CART) peptide on regulation of appetite and hypothalamo-pituitary axes in vitro and in vivo in male rats. *Brain Res* 893:186–194
141. Kong WM, Stanley S, Gardiner J, Abbott C, Murphy K, Seth A, Connoley I, Ghatei M, Stephens D, Bloom S 2003 A role for arcuate cocaine and amphetamine-regulated transcript in hyperphagia, thermogenesis, and cold adaptation. *FASEB J* 17:1688–1690
142. Wang C, Billington CJ, Levine AS, Kotz CM 2000 Effect of CART in the hypothalamic paraventricular nucleus on feeding and uncoupling protein gene expression. *Neuroreport* 11:3251–3255
143. Larsen PJ, Vrang N, Petersen PC, Kristensen P 2000 Chronic intracerebroventricular administration of recombinant CART(42–89) peptide inhibits and causes weight loss in lean and obese Zucker (fa/fa) rats. *Obes Res* 8:590–596
144. Rohner-Jeanrenaud F, Craft LS, Bridwell J, Suter TM, Tinsley FC, Smiley DL, Burkhardt DR, Statnick MA, Heiman ML, Ravussin E, Caro JF 2002 Chronic central infusion of

- cocaine- and amphetamine-regulated transcript (CART 55–102): effects on body weight homeostasis in lean and high-fat-fed obese rats. *Int J Obes Relat Metab Disord* 26:143–149
145. Lechan RM, Jackson IM 1982 Immunohistochemical localization of thyrotropin-releasing hormone in the rat hypothalamus and pituitary. *Endocrinology* 111:55–65
  146. Swanson LW, Sawchenko PE, Rivier J, Vale WW 1983 Organization of ovine corticotropin-releasing factor immunoreactive cells and fibers in the rat brain: an immunohistochemical study. *Neuroendocrinology* 36:165–186
  147. Broberger C, Visser TJ, Kuhar MJ, Hökfelt T 1999 Neuropeptide Y innervation and neuropeptide-Y-Y1-receptor-expressing neurons in the paraventricular hypothalamic nucleus of the mouse. *Neuroendocrinology* 70:295–305
  148. Lovenberg TW, Liaw CW, Grigoriadis DE, Clevenger W, Chalmers DT, De Souza EB, Oltersdorf T 1995 Cloning and characterization of a functionally distinct corticotropin-releasing factor receptor subtype from rat brain. *Proc Natl Acad Sci USA* 92:836–840
  149. Perrin MH, Donaldson CJ, Chen R, Lewis KA, Vale WW 1993 Cloning and functional expression of a rat brain corticotropin releasing factor (CRF) receptor. *Endocrinology* 133:3058–3061
  150. Cao J, O'Donnell D, Vu H, Payza K, Pou C, Godbout C, Jakob A, Pelletier M, Lembo P, Ahmad S, Walker P 1998 Cloning and characterization of a cDNA encoding a novel subtype of rat thyrotropin-releasing hormone receptor. *J Biol Chem* 273:32281–32287
  151. Itadani H, Nakamura T, Itoh J, Iwaasa H, Kanatani A, Borkowski J, Ihara M, Ohta M 1998 Cloning and characterization of a new subtype of thyrotropin-releasing hormone receptors. *Biochem Biophys Res Commun* 250:68–71
  152. Straub RE, Frech GC, Joho RH, Gershengorn MC 1990 Expression cloning of a cDNA encoding the mouse pituitary thyrotropin-releasing hormone receptor. *Proc Natl Acad Sci USA* 87:9514–9518
  153. Potter E, Sutton S, Donaldson C, Chen R, Perrin M, Lewis K, Sawchenko PE, Vale W 1994 Distribution of corticotropin-releasing factor receptor mRNA expression in the rat brain and pituitary. *Proc Natl Acad Sci USA* 91:8777–8781
  154. Wong ML, Licinio J, Pasternak KI, Gold PW 1994 Localization of corticotropin-releasing hormone (CRH) receptor mRNA in adult rat brain by *in situ* hybridization histochemistry. *Endocrinology* 135:2275–2278
  155. Heuer H, Schäfer MK, O'Donnell D, Walker P, Bauer K 2000 Expression of thyrotropin-releasing hormone receptor 2 (TRH-R2) in the central nervous system of rats. *J Comp Neurol* 428:319–336
  156. Lovenberg TW, Chalmers DT, Liu C, De Souza EB 1995 CRF2  $\alpha$  and CRF2  $\beta$  receptor mRNAs are differentially distributed between the rat central nervous system and peripheral tissues. *Endocrinology* 136:4139–4142
  157. Van Pett K, Viau V, Bittencourt JC, Chan RK, Li HY, Arias C, Prins GS, Perrin M, Vale W, Sawchenko PE 2000 Distribution of mRNAs encoding CRF receptors in brain and pituitary of rat and mouse. *J Comp Neurol* 428:191–212
  158. Richard D, Huang Q, Timofeeva E 2000 The corticotropin-releasing hormone system in the regulation of energy balance in obesity. *Int J Obes Relat Metab Disord* 24(Suppl 2):S36–S39
  159. Jones MT, Gillham B 1988 Factors involved in the regulation of adrenocorticotrophic hormone/ $\beta$ -lipotropic hormone. *Physiol Rev* 68:743–818
  160. Sawchenko PE, Swanson LW 1985 Localization, colocalization, and plasticity of corticotropin-releasing factor immunoreactivity in rat brain. *Fed Proc* 44:221–227
  161. Suemaru S, Hashimoto K, Hattori T, Inoue H, Kageyama J, Ota Z 1986 Starvation-induced changes in rat brain corticotropin-releasing factor (CRF) and pituitary-adrenocortical response. *Life Sci* 39:1161–1166
  162. Uehara Y, Shimizu H, Ohtani K, Sato N, Mori M 1998 Hypothalamic corticotropin-releasing hormone is a mediator of the anorexigenic effect of leptin. *Diabetes* 47:890–893
  163. Fekete C, Légrádi G, Mihály E, Tatro JB, Rand WM, Lechan RM 2000  $\alpha$ -Melanocyte stimulating hormone prevents fasting-induced suppression of corticotropin-releasing hormone gene expression in the rat hypothalamic paraventricular nucleus. *Neurosci Lett* 289:152–156
  164. Smith SM, Vaughan JM, Donaldson CJ, Rivier J, Li C, Chen A, Vale WW 2004 Cocaine- and amphetamine-regulated transcript activates the hypothalamic-pituitary-adrenal axis through a corticotropin-releasing factor receptor-dependent mechanism. *Endocrinology* 145:5202–5209
  165. Arase K, York DA, Shimizu H, Shargill N, Bray GA 1988 Effects of corticotropin-releasing factor on food intake and brown adipose tissue thermogenesis in rats. *Am J Physiol* 255:E255–E259
  166. Britton DR, Koob GF, Rivier J, Vale W 1982 Intraventricular corticotropin-releasing factor enhances behavioral effects of novelty. *Life Sci* 31:363–367
  167. Buwalda B, Van Kalkeren AA, de Boer SF, Koolhaas JM 1998 Behavioral and physiological consequences of repeated daily intracerebroventricular injection of corticotropin-releasing factor in the rat. *Psychoneuroendocrinology* 23:205–218
  168. LeFeuvre RA, Rothwell NJ, Stock MJ 1987 Activation of brown fat thermogenesis in response to central injection of corticotropin releasing hormone in the rat. *Neuropharmacology* 26:1217–1221
  169. Lechan RM, Fekete C 2006 Role of melanocortin signaling in the regulation of the hypothalamic-pituitary-thyroid (HPT) axis. *Peptides* 27:310–325
  170. Fekete C, Légrádi G, Mihály E, Huang QH, Tatro JB, Rand WM, Emerson CH, Lechan RM 2000  $\alpha$ -Melanocyte-stimulating hormone is contained in nerve terminals innervating thyrotropin-releasing hormone-synthesizing neurons in the hypothalamic paraventricular nucleus and prevents fasting-induced suppression of prothyrotropin-releasing hormone gene expression. *J Neurosci* 20:1550–1558
  171. Fekete C, Mihály E, Luo LG, Kelly J, Clausen JT, Mao Q, Rand WM, Moss LG, Kuhar M, Emerson CH, Jackson IM, Lechan RM 2000 Association of cocaine- and amphetamine-regulated transcript-immunoreactive elements with thyrotropin-releasing hormone-synthesizing neurons in the hypothalamic paraventricular nucleus and its role in the regulation of the hypothalamic-pituitary-thyroid axis during fasting. *J Neurosci* 20:9224–9234
  172. Fekete C, Kelly J, Mihály E, Sarkar S, Rand WM, Légrádi G, Emerson CH, Lechan RM 2001 Neuropeptide Y has a central inhibitory action on the hypothalamic-pituitary-thyroid axis. *Endocrinology* 142:2606–2613



173. Fekete C, Sarkar S, Rand WM, Harney JW, Emerson CH, Bianco AC, Lechan RM 2002 Agouti-related protein (AGRP) has a central inhibitory action on the hypothalamic-pituitary-thyroid (HPT) axis; comparisons between the effect of AGRP and neuropeptide Y on energy homeostasis and the HPT axis. *Endocrinology* 143:3846–3853
174. Fekete C, Singru PS, Sanchez E, Sarkar S, Christoffolete MA, Riberio RS, Rand WM, Emerson CH, Bianco AC, Lechan RM 2006 Differential effects of central leptin, insulin, or glucose administration during fasting on the hypothalamic-pituitary-thyroid axis and feeding-related neurons in the arcuate nucleus. *Endocrinology* 147:520–529
175. Choi YH, Hartzell D, Azain MJ, Baile CA 2002 TRH decreases food intake and increases water intake and body temperature in rats. *Physiol Behav* 77:1–4
176. Suzuki T, Kohno H, Sakurada T, Tadano T, Kisara K 1982 Intracranial injection of thyrotropin releasing hormone (TRH) suppresses starvation-induced feeding and drinking in rats. *Pharmacol Biochem Behav* 17:249–253
177. Vijayan E, McCann SM 1977 Suppression of feeding and drinking activity in rats following intraventricular injection of thyrotropin releasing hormone (TRH). *Endocrinology* 100:1727–1730
178. Iglesias R, Llobera M, Montoya E 1986 Long-term effects of TRH administration on food intake and body weight in the rat. *Pharmacol Biochem Behav* 24:1817–1819
179. Broberger C, De Lecea L, Sutcliffe JG, Hökfelt T 1998 Hypocretin/orexin- and melanin-concentrating hormone-expressing cells form distinct populations in the rodent lateral hypothalamus: relationship to the neuropeptide Y and agouti gene-related protein systems. *J Comp Neurol* 402:460–474
180. Bittencourt JC, Presse F, Arias C, Peto C, Vaughan J, Nahon JL, Vale W, Sawchenko PE 1992 The melanin-concentrating hormone system of the rat brain: an immunohistochemical characterization. *J Comp Neurol* 319:218–245
181. Date Y, Ueta Y, Yamashita H, Yamaguchi H, Matsukura S, Kangawa K, Sakurai T, Yanagisawa M, Nakazato M 1999 Orexins, orexigenic hypothalamic peptides, interact with autonomic, neuroendocrine and neuroregulatory systems. *Proc Natl Acad Sci USA* 96:748–753
182. Peyron C, Tighe DK, van den Pol AN, de Lecea L, Heller HC, Sutcliffe JG, Kilduff TS 1998 Neurons containing hypocretin (orexin) project to multiple neuronal systems. *J Neurosci* 18:9996–10015
183. Bächner D, Kreienkamp H, Weise C, Buck F, Richter D 1999 Identification of melanin concentrating hormone (MCH) as the natural ligand for the orphan somatostatin-like receptor 1 (SLC-1). *FEBS Lett* 457:522–524
184. Chambers J, Ames RS, Bergsma D, Muir A, Fitzgerald LR, Hervieu G, Dytko GM, Foley JJ, Martin J, Liu WS, Park J, Ellis C, Ganguly S, Konchar S, Cluderay J, Leslie R, Wilson S, Sarau HM 1999 Melanin-concentrating hormone is the cognate ligand for the orphan G-protein-coupled receptor SLC-1. *Nature* 400:261–265
185. Lembo PM, Grazzini E, Cao J, Hubatsch DA, Pelletier M, Hoffert C, St-Onge S, Pou C, Labrecque J, Groblewski T, O'Donnell D, Payza K, Ahmad S, Walker P 1999 The receptor for the orexigenic peptide melanin-concentrating hormone is a G-protein-coupled receptor. *Nat Cell Biol* 1:267–271
186. Saito Y, Nothacker HP, Wang Z, Lin SH, Leslie F, Civelli O 1999 Molecular characterization of the melanin-concentrating-hormone receptor. *Nature* 400:265–269
187. Shimomura Y, Mori M, Sugo T, Ishibashi Y, Abe M, Kurokawa T, Onda H, Nishimura O, Sumino Y, Fujino M 1999 Isolation and identification of melanin-concentrating hormone as the endogenous ligand of the SLC-1 receptor. *Biochem Biophys Res Commun* 261:622–626
188. Tan CP, Sano H, Iwaasa H, Pan J, Sailer AW, Hreniuk DL, Feighner SD, Palyha OC, Pong SS, Figueroa DJ, Austin CP, Jiang MM, Yu H, Ito J, Ito M, Ito M, Guan XM, MacNeil DJ, Kanatani A, Van der Ploeg LH, Howard AD 2002 Melanin-concentrating hormone receptor subtypes 1 and 2: species-specific gene expression. *Genomics* 79:785–792
189. Hervieu GJ, Cluderay JE, Harrison D, Meakin J, Maycox P, Nasir S, Leslie RA 2000 The distribution of the mRNA and protein products of the melanin-concentrating hormone (MCH) receptor gene, slc-1, in the central nervous system of the rat. *Eur J Neurosci* 12:1194–1216
190. Saito Y, Cheng M, Leslie FM, Civelli O 2001 Expression of the melanin-concentrating hormone (MCH) receptor mRNA in the rat brain. *J Comp Neurol* 435:26–40
191. Marcus JN, Aschkenasi CJ, Lee CE, Chemelli RM, Saper CB, Yanagisawa M, Elmquist JK 2001 Differential expression of orexin receptors 1 and 2 in the rat brain. *J Comp Neurol* 435:6–25
192. Trivedi P, Yu H, MacNeil DJ, Van der Ploeg LH, Guan XM 1998 Distribution of orexin receptor mRNA in the rat brain. *FEBS Lett* 438:71–75
193. Kawauchi H, Kawazoe I, Tsubokawa M, Kishida M, Baker BI 1983 Characterization of melanin-concentrating hormone in chum salmon pituitaries. *Nature* 305:321–323
194. Skofitsch G, Jacobowitz DM, Zamir N 1985 Immunohistochemical localization of a melanin concentrating hormone-like peptide in the rat brain. *Brain Res Bull* 15:635–649
195. Vaughan JM, Fischer WH, Hoeger C, Rivier J, Vale W 1989 Characterization of melanin-concentrating hormone from rat hypothalamus. *Endocrinology* 125:1660–1665
196. Huang Q, Viale A, Picard F, Nahon J, Richard D 1999 Effects of leptin on melanin-concentrating hormone expression in the brain of lean and obese Lep(ob)/Lep(ob) mice. *Neuroendocrinology* 69:145–153
197. Tritos NA, Mastaitis JW, Kokkotou E, Maratos-Flier E 2001 Characterization of melanin concentrating hormone and preproorexin expression in the murine hypothalamus. *Brain Res* 895:160–166
198. Presse F, Sorokovsky I, Max JP, Nicolaidis S, Nahon JL 1996 Melanin-concentrating hormone is a potent anorectic peptide regulated by food-deprivation and glucopenia in the rat. *Neuroscience* 71:735–745
199. Qu D, Ludwig DS, Gammeltoft S, Piper M, Pellemounter MA, Cullen MJ, Mathes WF, Przyspek R, Kanarek R, Maratos-Flier E 1996 A role for melanin-concentrating hormone in the central regulation of feeding behaviour. *Nature* 380:243–247
200. van den Pol AN, Acuna-Goycolea C, Clark KR, Ghosh PK 2004 Physiological properties of hypothalamic MCH neurons identified with selective expression of reporter gene after recombinant virus infection. *Neuron* 42:635–652
201. Abbott CR, Kennedy AR, Wren AM, Rossi M, Murphy KG, Seal LJ, Todd JF, Ghatei MA, Small CJ, Bloom SR

- 2003 Identification of hypothalamic nuclei involved in the orexigenic effect of melanin-concentrating hormone. *Endocrinology* 144:3943–3949
202. Ludwig DS, Mountjoy KG, Tatro JB, Gillette JA, Frederich RC, Flier JS, Maratos-Flier E 1998 Melanin-concentrating hormone: a functional melanocortin antagonist in the hypothalamus. *Am J Physiol* 274:E627–E633
203. Rossi M, Beak SA, Choi SJ, Small CJ, Morgan DG, Ghatei MA, Smith DM, Bloom SR 1999 Investigation of the feeding effects of melanin concentrating hormone on food intake—action independent of galanin and the melanocortin receptors. *Brain Res* 846:164–170
204. Rossi M, Choi SJ, O’Shea D, Miyoshi T, Ghatei MA, Bloom SR 1997 Melanin-concentrating hormone acutely stimulates feeding, but chronic administration has no effect on body weight. *Endocrinology* 138:351–355
205. Marsh DJ, Weingarh DT, Novi DE, Chen HY, Trumbauer ME, Chen AS, Guan XM, Jiang MM, Feng Y, Camacho RE, Shen Z, Frazier EG, Yu H, Metzger JM, Kuca SJ, Shearman LP, Gopal-Truter S, MacNeil DJ, Strack AM, MacIntyre DE, Van der Ploeg LH, Qian S 2002 Melanin-concentrating hormone 1 receptor-deficient mice are lean, hyperactive, and hyperphagic and have altered metabolism. *Proc Natl Acad Sci USA* 99:3240–3245
206. Segal-Lieberman G, Bradley RL, Kokkotou E, Carlson M, Trombly DJ, Wang X, Bates S, Myers Jr MG, Flier JS, Maratos-Flier E 2003 Melanin-concentrating hormone is a critical mediator of the leptin-deficient phenotype. *Proc Natl Acad Sci USA* 100:10085–10090
207. de Lecea L, Kilduff TS, Peyron C, Gao X, Foye PE, Danielson PE, Fukuhara C, Battenberg EL, Gautvik VT, Bartlett 2nd FS, Frankel WN, van den Pol AN, Bloom FE, Gautvik KM, Sutcliffe JG 1998 The hypocretins: hypothalamus-specific peptides with neuroexcitatory activity. *Proc Natl Acad Sci USA* 95:322–327
208. Sakurai T, Amemiya A, Ishii M, Matsuzaki I, Chemelli RM, Tanaka H, Williams SC, Richardson JA, Kozlowski GP, Wilson S, Arch JR, Buckingham RE, Haynes AC, Carr SA, Annan RS, McNulty DE, Liu WS, Terrett JA, Elshourbagy NA, Bergsma DJ, Yanagisawa M 1998 Orexins and orexin receptors: a family of hypothalamic neuropeptides and G protein-coupled receptors that regulate feeding behavior. *Cell* 92:573–585
209. Cai XJ, Widdowson PS, Harrold J, Wilson S, Buckingham RE, Arch JR, Tadayyon M, Clapham JC, Wilding J, Williams G 1999 Hypothalamic orexin expression: modulation by blood glucose and feeding. *Diabetes* 48:2132–2137
210. López M, Seoane L, García MC, Lago F, Casanueva FF, Señaris R, Diéguez C 2000 Leptin regulation of prepro-orexin and orexin receptor mRNA levels in the hypothalamus. *Biochem Biophys Res Commun* 269:41–45
211. Mondal MS, Nakazato M, Date Y, Murakami N, Yanagisawa M, Matsukura S 1999 Widespread distribution of orexin in rat brain and its regulation upon fasting. *Biochem Biophys Res Commun* 256:495–499
212. Swart I, Overton JM, Houpt TA 2001 The effect of food deprivation and experimental diabetes on orexin and NPY mRNA levels. *Peptides* 22:2175–2179
213. Beck B, Richy S 1999 Hypothalamic hypocretin/orexin and neuropeptide Y: divergent interaction with energy depletion and leptin. *Biochem Biophys Res Commun* 258:119–122
214. Yamanaka A, Beuckmann CT, Willie JT, Hara J, Tsujino N, Mieda M, Tominaga M, Yagami K, Sugiyama F, Goto K, Yanagisawa M, Sakurai T 2003 Hypothalamic orexin neurons regulate arousal according to energy balance in mice. *Neuron* 38:701–713
215. Zheng H, Corkern MM, Crousillac SM, Patterson LM, Phifer CB, Berthoud HR 2002 Neurochemical phenotype of hypothalamic neurons showing Fos expression 23 h after intracranial AgRP. *Am J Physiol Regul Integr Comp Physiol* 282:R1773–R1781
216. Campbell RE, Smith MS, Allen SE, Grayson BE, Ffrench-Mullen JM, Grove KL 2003 Orexin neurons express a functional pancreatic polypeptide Y4 receptor. *J Neurosci* 23:1487–1497
217. Fu LY, Acuna-Goycolea C, van den Pol AN 2004 Neuropeptide Y inhibits hypocretin/orexin neurons by multiple presynaptic and postsynaptic mechanisms: tonic depression of the hypothalamic arousal system. *J Neurosci* 24:8741–8751
218. Dube MG, Kalra SP, Kalra PS 1999 Food intake elicited by central administration of orexins/hypocretins: identification of hypothalamic sites of action. *Brain Res* 842:473–477
219. Haynes AC, Jackson B, Overend P, Buckingham RE, Wilson S, Tadayyon M, Arch JR 1999 Effects of single and chronic intracerebroventricular administration of the orexins on feeding in the rat. *Peptides* 20:1099–1105
220. Muroya S, Funahashi H, Yamanaka A, Kohno D, Uramura K, Nambu T, Shibahara M, Kuramochi M, Takigawa M, Yanagisawa M, Sakurai T, Shioda S, Yada T 2004 Orexins (hypocretins) directly interact with neuropeptide Y, POMC and glucose-responsive neurons to regulate Ca<sup>2+</sup> signaling in a reciprocal manner to leptin: orexigenic neuronal pathways in the mediobasal hypothalamus. *Eur J Neurosci* 19:1524–1534
221. Kotz CM 2006 Integration of feeding and spontaneous physical activity: role for orexin. *Physiol Behav* 88:294–301
222. Ida T, Nakahara K, Katayama T, Murakami N, Nakazato M 1999 Effect of lateral cerebroventricular injection of the appetite-stimulating neuropeptide, orexin and neuropeptide Y, on the various behavioral activities of rats. *Brain Res* 821:526–529
223. Kiwaki K, Kotz CM, Wang C, Lanningham-Foster L, Levine JA 2004 Orexin A (hypocretin 1) injected into hypothalamic paraventricular nucleus and spontaneous physical activity in rats. *Am J Physiol Endocrinol Metab* 286:E551–E559
224. Kotz CM, Teske JA, Levine JA, Wang C 2002 Feeding and activity induced by orexin A in the lateral hypothalamus in rats. *Regul Pept* 104:27–32
225. Lubkin M, Stricker-Krongrad A 1998 Independent feeding and metabolic actions of orexins in mice. *Biochem Biophys Res Commun* 253:241–245
226. Yamanaka A, Sakurai T, Katsumoto T, Yanagisawa M, Goto K 1999 Chronic intracerebroventricular administration of orexin-A to rats increases food intake in daytime, but has no effect on body weight. *Brain Res* 849:248–252
227. Sakurai T 2005 Roles of orexin/hypocretin in regulation of sleep/wakefulness and energy homeostasis. *Sleep Med Rev* 9:231–241

228. Sutcliffe JG, de Lecea L 2002 The hypocretins: setting the arousal threshold. *Nat Rev Neurosci* 3:339–349
229. Willie JT, Chemelli RM, Sinton CM, Yanagisawa M 2001 To eat or to sleep? Orexin in the regulation of feeding and wakefulness. *Annu Rev Neurosci* 24:429–458
230. Nieuwenhuizen AG, Rutters F 2008 The hypothalamic-pituitary-adrenal-axis in the regulation of energy balance. *Physiol Behav* 94:169–177
231. Fliers E, Kreier F, Voshol PJ, Havekes LM, Sauerwein HP, Kalsbeek A, Buijs RM, Romijn JA 2003 White adipose tissue: getting nervous. *J Neuroendocrinol* 15:1005–1010
232. Pénicaud L, Cousin B, Leloup C, Lorsignol A, Casteilla L 2000 The autonomic nervous system, adipose tissue plasticity, and energy balance. *Nutrition* 16:903–908
233. Mounien L, Bizet P, Boutelet I, Vaudry H, Jégou S 2005 Expression of melanocortin MC3 and MC4 receptor mRNAs by neuropeptide Y neurons in the rat arcuate nucleus. *Neuroendocrinology* 82:164–170
234. Roseberry AG, Liu H, Jackson AC, Cai X, Friedman JM 2004 Neuropeptide Y-mediated inhibition of proopiomelanocortin neurons in the arcuate nucleus shows enhanced desensitization in ob/ob mice. *Neuron* 41:711–722
235. Dhillon WS, Small CJ, Stanley SA, Jethwa PH, Seal LJ, Murphy KG, Ghatgei MA, Bloom SR 2002 Hypothalamic interactions between neuropeptide Y, agouti-related protein, cocaine- and amphetamine-regulated transcript and  $\alpha$ -melanocyte-stimulating hormone in vitro in male rats. *J Neuroendocrinol* 14:725–730
236. Håkansson M, de Lecea L, Sutcliffe JG, Yanagisawa M, Meister B 1999 Leptin receptor- and STAT3-immunoreactivities in hypocretin/orexin neurons of the lateral hypothalamus. *J Neuroendocrinol* 11:653–663
237. Håkansson ML, Brown H, Ghilardi N, Skoda RC, Meister B 1998 Leptin receptor immunoreactivity in chemically defined target neurons of the hypothalamus. *J Neurosci* 18:559–572
238. Harris M, Aschkenasi C, Elias CF, Chandrankunnel A, Nillni EA, Bjørbaek C, Elmquist JK, Flier JS, Hollenberg AN 2001 Transcriptional regulation of the thyrotropin-releasing hormone gene by leptin and melanocortin signaling. *J Clin Invest* 107:111–120
239. Leininger GM, Jo YH, Leshan RL, Louis GW, Yang H, Barrera JG, Wilson H, Opland DM, Faouzi MA, Gong Y, Jones JC, Rhodes CJ, Chua Jr S, Diano S, Horvath TL, Seeley RJ, Becker JB, Münzberg H, Myers Jr MG 2009 Leptin acts via leptin receptor-expressing lateral hypothalamic neurons to modulate the mesolimbic dopamine system and suppress feeding. *Cell Metab* 10:89–98
240. Figlewicz DP, Benoit SC 2009 Insulin, leptin, and food reward: update 2008. *Am J Physiol Regul Integr Comp Physiol* 296:R9–R19
241. Hisano S, Fukui Y, Chikamori-Aoyama M, Aizawa T, Shibasaki T 1993 Reciprocal synaptic relations between CRF-immunoreactive- and TRH-immunoreactive neurons in the paraventricular nucleus of the rat hypothalamus. *Brain Res* 620:343–346
242. Guan JL, Uehara K, Lu S, Wang QP, Funahashi H, Sakurai T, Yanagisawa M, Shioda S 2002 Reciprocal synaptic relationships between orexin- and melanin-concentrating hormone-containing neurons in the rat lateral hypothalamus: a novel circuit implicated in feeding regulation. *Int J Obes Relat Metab Disord* 26:1523–1532
243. Li Y, Gao XB, Sakurai T, van den Pol AN 2002 Hypocretin/orexin excites hypocretin neurons via a local glutamate neuron-A potential mechanism for orchestrating the hypothalamic arousal system. *Neuron* 36:1169–1181
244. Kennedy AR, Todd JF, Dhillon WS, Seal LJ, Ghatgei MA, O'Toole CP, Jones M, Witty D, Winborne K, Riley G, Hervieu G, Wilson S, Bloom SR 2003 Effect of direct injection of melanin-concentrating hormone into the paraventricular nucleus: further evidence for a stimulatory role in the adrenal axis via SLC-1. *J Neuroendocrinol* 15:268–272
245. Sakamoto F, Yamada S, Ueta Y 2004 Centrally administered orexin-A activates corticotropin-releasing factor-containing neurons in the hypothalamic paraventricular nucleus and central amygdaloid nucleus of rats: possible involvement of central orexins on stress-activated central CRF neurons. *Regul Pept* 118:183–191
246. Kennedy AR, Todd JF, Stanley SA, Abbott CR, Small CJ, Ghatgei MA, Bloom SR 2001 Melanin-concentrating hormone (MCH) suppresses thyroid stimulating hormone (TSH) release, *in vivo* and *in vitro*, via the hypothalamus and the pituitary. *Endocrinology* 142:3265–3268
247. Winsky-Sommerer R, Yamanaka A, Diano S, Borok E, Roberts AJ, Sakurai T, Kilduff TS, Horvath TL, de Lecea L 2004 Interaction between the corticotropin-releasing factor system and hypocretins (orexins): a novel circuit mediating stress response. *J Neurosci* 24:11439–11448
248. Guan JL, Saotome T, Wang QP, Funahashi H, Hori T, Tanaka S, Shioda S 2001 Orexinergic innervation of POMC-containing neurons in the rat arcuate nucleus. *Neuroreport* 12:547–551
249. Campbell RE, Grove KL, Smith MS 2003 Distribution of corticotropin releasing hormone receptor immunoreactivity in the rat hypothalamus: coexpression in neuropeptide Y and dopamine neurons in the arcuate nucleus. *Brain Res* 973:223–232
250. Davidowa H, Plagemann A 2004 Hypothalamic neurons of postnatally overfed, overweight rats respond differentially to corticotropin-releasing hormones. *Neurosci Lett* 371:64–68
251. Bingham NC, Anderson KK, Reuter AL, Stallings NR, Parker KL 2008 Selective loss of leptin receptors in the ventromedial hypothalamic nucleus results in increased adiposity and a metabolic syndrome. *Endocrinology* 149:2138–2148
252. Kawano H, Masuko S 2000  $\beta$ -Endorphin-, adrenocorticotrophic hormone- and neuropeptide Y-containing projection fibers from the arcuate hypothalamic nucleus make synaptic contacts on to nucleus preopticus medianus neurons projecting to the paraventricular hypothalamic nucleus in the rat. *Neuroscience* 98:555–565
253. Mihály E, Fekete C, Légrádi G, Lechan RM 2001 Hypothalamic dorsomedial nucleus neurons innervate thyrotropin-releasing hormone-synthesizing neurons in the paraventricular nucleus. *Brain Res* 891:20–31
254. Singru PS, Fekete C, Lechan RM 2005 Neuroanatomical evidence for participation of the hypothalamic dorsomedial nucleus (DMN) in regulation of the hypothalamic paraventricular nucleus (PVN) by  $\alpha$ -melanocyte stimulating hormone. *Brain Res* 1064:42–51
255. Stenerson SM, Shepherd GM, Friedman JM 2005 Topographic mapping of VMH  $\rightarrow$  arcuate nucleus microcircuits



- and their reorganization by fasting. *Nat Neurosci* 8:1356–1363
256. Grill HJ, Kaplan JM 2002 The neuroanatomical axis for control of energy balance. *Front Neuroendocrinol* 23:2–40
  257. Bergonzelli GE, Pralong FP, Glauser M, Cavadas C, Grouzmann E, Gaillard RC 2001 Interplay between galanin and leptin in the hypothalamic control of feeding via corticotropin-releasing hormone and neuropeptide Y. *Diabetes* 50:2666–2672
  258. Hanada R, Teranishi H, Pearson JT, Kurokawa M, Hosoda H, Fukushima N, Fukue Y, Serino R, Fujihara H, Ueta Y, Ikawa M, Okabe M, Murakami N, Shirai M, Yoshimatsu H, Kangawa K, Kojima M 2004 Neuromedin U has a novel anorexigenic effect independent of the leptin signaling pathway. *Nat Med* 10:1067–1073
  259. Krasnow SM, Fraley GS, Schuh SM, Baumgartner JW, Clifton DK, Steiner RA 2003 A role for galanin-like peptide in the integration of feeding, body weight regulation, and reproduction in the mouse. *Endocrinology* 144:813–822
  260. Sahu A 1998 Evidence suggesting that galanin (GAL), melanin-concentrating hormone (MCH), neurotensin (NT), proopiomelanocortin (POMC) and neuropeptide Y (NPY) are targets of leptin signaling in the hypothalamus. *Endocrinology* 139:795–798
  261. Wolf G 2006 The regulation of food intake by hypothalamic malonyl-coenzyme A: the MalCoA hypothesis. *Nutr Rev* 64:379–383
  262. Bäckberg M, Ultenius C, Fritschy JM, Meister B 2004 Cellular localization of GABA<sub>A</sub> receptor  $\alpha$  subunit immunoreactivity in the rat hypothalamus: relationship with neurons containing orexigenic or anorexigenic peptides. *J Neuroendocrinol* 16:589–604
  263. Hentges ST, Nishiyama M, Overstreet LS, Stenzel-Poore M, Williams JT, Low MJ 2004 GABA release from proopiomelanocortin neurons. *J Neurosci* 24:1578–1583
  264. van den Pol AN, Trombley PQ 1993 Glutamate neurons in hypothalamus regulate excitatory transmission. *J Neurosci* 13:2829–2836
  265. Allison SJ, Baldock PA, Herzog H 2007 The control of bone remodeling by neuropeptide Y receptors. *Peptides* 28:320–325
  266. Cone RD, Lu D, Koppula S, Vage DI, Klungland H, Boston B, Chen W, Orth DN, Pouton C, Kesterson RA 1996 The melanocortin receptors: agonists, antagonists, and the hormonal control of pigmentation. *Recent Prog Horm Res* 51:287–317; discussion 318
  267. Heilig M, Thorsell A 2002 Brain neuropeptide Y (NPY) in stress and alcohol dependence. *Rev Neurosci* 13:85–94
  268. Karsenty G 2006 Convergence between bone and energy homeostases: leptin regulation of bone mass. *Cell Metab* 4:341–348
  269. Mukherjee R, Villarreal D, Reams GP, Freeman RH, Tchoukina I, Spear RM 2006 Leptin as a common link to obesity and hypertension. *Timely Top Med Cardiovasc Dis* 10:E1
  270. Pedrazzini T, Pralong F, Grouzmann E 2003 Neuropeptide Y: the universal soldier. *Cell Mol Life Sci* 60:350–377
  271. Pissios P, Bradley RL, Maratos-Flier E 2006 Expanding the scales: the multiple roles of MCH in regulating energy balance and other biological functions. *Endocr Rev* 27:606–620
  272. Wikberg JE 1999 Melanocortin receptors: perspectives for novel drugs. *Eur J Pharmacol* 375:295–310
  273. Zhang F, Chen Y, Heiman M, Dimarchi R 2005 Leptin: structure, function and biology. *Vitam Horm* 71:345–372
  274. Menyhárt J, Wittmann G, Lechan RM, Keller E, Liposits Z, Fekete C 2007 Cocaine- and amphetamine-regulated transcript (CART) is colocalized with the orexigenic neuropeptide Y and agouti-related protein and absent from the anorexigenic  $\alpha$ -melanocyte-stimulating hormone neurons in the infundibular nucleus of the human hypothalamus. *Endocrinology* 148:4276–4281
  275. Markakis EA 2002 Development of the neuroendocrine hypothalamus. *Front Neuroendocrinol* 23:257–291
  276. Rinaman L 2006 Ontogeny of hypothalamic-hindbrain feeding control circuits. *Dev Psychobiol* 48:389–396
  277. Rinaman L 2007 Visceral sensory inputs to the endocrine hypothalamus. *Front Neuroendocrinol* 28:50–60
  278. Grove KL, Smith MS 2003 Ontogeny of the hypothalamic neuropeptide Y system. *Physiol Behav* 79:47–63
  279. Sugita N 1917 Comparative studies on the growth of the cerebral cortex. I. On the changes in the size and shape of the cerebrum during the postnatal growth of the brain. Albino rat. *J Comp Neurol* 28:495–510
  280. Ferré P, Decaux JF, Issad T, Girard J 1986 Changes in energy metabolism during the suckling and weaning period in the newborn. *Reprod Nutr Dev* 26:619–631
  281. Babický A, Parížek J, Ostádalová I, Kolár J 1973 Initial solid food intake and growth of young rats in nests of different sizes. *Physiol Bohemoslov* 22:557–566
  282. Swithers SE 2003 Do metabolic signals stimulate intake in rat pups? *Physiol Behav* 79:71–78
  283. Cramer CP, Blass EM 1985 Nutritive and nonnutritive determinants of milk intake of suckling rats. *Behav Neurosci* 99:578–582
  284. Haupt KA, Epstein AN 1973 Ontogeny of controls of food intake in the rat: GI fill and glucoprivation. *Am J Physiol* 225:58–66
  285. Lorenz DN, Ellis SB, Epstein AN 1982 Differential effects of upper gastrointestinal fill on milk ingestion and nipple attachment in the suckling rat. *Dev Psychobiol* 15:309–330
  286. Phifer CB, Sikes CR, Hall WG 1986 Control of ingestion in 6-day-old rat pups: termination of intake by gastric fill alone? *Am J Physiol* 250:R807–R814
  287. Lorenz DN 1983 Effects of gastric filling and vagotomy on ingestion, nipple attachment, and weight gain by suckling rats. *Dev Psychobiol* 16:469–483
  288. Lorenz DN 1992 Suckling physiology and behavior of rats: an integrated theory of ingestion and satiety. *Prog Psychobiol Physiol Psychol* 15:1–83
  289. Proulx K, Clavel S, Nault G, Richard D, Walker CD 2001 High neonatal leptin exposure enhances brain GR expression and feedback efficacy on the adrenocortical axis of developing rats. *Endocrinology* 142:4607–4616
  290. Hall WG 1985 What we know and don't know about the development of independent ingestion in rats. *Appetite* 6:333–356
  291. Kleitman N, Satinoff E 1982 Thermoregulatory behavior in rat pups from birth to weaning. *Physiol Behav* 29:537–541

292. Kawai M, Yamaguchi M, Murakami T, Shima K, Murata Y, Kishi K 1997 The placenta is not the main source of leptin production in pregnant rat: gestational profile of leptin in plasma and adipose tissues. *Biochem Biophys Res Commun* 240:798–802
293. Herrera E, Lasunción MA, Huerta L, Martín-Hidalgo A 2000 Plasma leptin levels in rat mother and offspring during pregnancy and lactation. *Biol Neonate* 78:315–320
294. Dessolin S, Schalling M, Champigny O, Lönnqvist F, Ailhaud G, Dani C, Ricquier D 1997 Leptin gene is expressed in rat brown adipose tissue at birth. *FASEB J* 11:382–387
295. Beloosesky R, Gayle DA, Amidi F, Ahanya SN, Desai M, Ross MG 2006 Ontogenic expression of putative feeding peptides in the rat fetal brain and placenta. *Nutr Neurosci* 9:33–40
296. Carlo AS, Meyerhof W, Williams LM 2007 Early developmental expression of leptin receptor gene and [<sup>125</sup>I]leptin binding in the rat forebrain. *J Chem Neuroanat* 33:155–163
297. Smith JT, Waddell BJ 2003 Developmental changes in plasma leptin and hypothalamic leptin receptor expression in the rat: peripubertal changes and the emergence of sex differences. *J Endocrinol* 176:313–319
298. Morash B, Wilkinson D, Murphy P, Ur E, Wilkinson M 2001 Developmental regulation of leptin gene expression in rat brain and pituitary. *Mol Cell Endocrinol* 185:151–159
299. Rayner DV, Dalglish GD, Duncan JS, Hardie LJ, Hoggard N, Trayhurn P 1997 Postnatal development of the ob gene system: elevated leptin levels in suckling fa/fa rats. *Am J Physiol* 273:R446–R450
300. Ahima RS, Prabakaran D, Flier JS 1998 Postnatal leptin surge and regulation of circadian rhythm of leptin by feeding. Implications for energy homeostasis and neuroendocrine function. *J Clin Invest* 101:1020–1027
301. Yura S, Itoh H, Sagawa N, Yamamoto H, Masuzaki H, Nakao K, Kawamura M, Takemura M, Kakui K, Ogawa Y, Fujii S 2005 Role of premature leptin surge in obesity resulting from intrauterine undernutrition. *Cell Metab* 1:371–378
302. Casabiell X, Piñeiro V, Tomé MA, Peinó R, Diéguez C, Casanueva FF 1997 Presence of leptin in colostrum and/or breast milk from lactating mothers: a potential role in the regulation of neonatal food intake. *J Clin Endocrinol Metab* 82:4270–4273
303. Morash BA, Imran A, Wilkinson D, Ur E, Wilkinson M 2003 Leptin receptors are developmentally regulated in rat pituitary and hypothalamus. *Mol Cell Endocrinol* 210:1–8
304. Kraeft S, Schwarzer K, Eiden S, Nuesslein-Hildesheim B, Preibisch G, Schmidt I 1999 Leptin responsiveness and gene dosage for leptin receptor mutation (fa) in newborn rats. *Am J Physiol* 276:E836–E842
305. Proulx K, Richard D, Walker CD 2002 Leptin regulates appetite-related neuropeptides in the hypothalamus of developing rats without affecting food intake. *Endocrinology* 143:4683–4692
306. Stehling O, Döring H, Ertl J, Preibisch G, Schmidt I 1996 Leptin reduces juvenile fat stores by altering the circadian cycle of energy expenditure. *Am J Physiol* 271:R1770–R1774
307. Blumberg MS, Deaver K, Kirby RF 1999 Leptin disinhibits nonshivering thermogenesis in infants after maternal separation. *Am J Physiol* 276:R606–R610
308. Zhang Y, Olbort M, Schwarzer K, Nuesslein-Hildesheim B, Nicolson M, Murphy E, Kowalski TJ, Schmidt I, Leibel RL 1997 The leptin receptor mediates apparent autocrine regulation of leptin gene expression. *Biochem Biophys Res Commun* 240:492–495
309. Mistry AM, Swick A, Romsos DR 1999 Leptin alters metabolic rates before acquisition of its anorectic effect in developing neonatal mice. *Am J Physiol* 277:R742–R747
310. Ahima RS, Hileman SM 2000 Postnatal regulation of hypothalamic neuropeptide expression by leptin: implications for energy balance and body weight regulation. *Regul Pept* 92:1–7
311. Ahima RS, Bjorbaek C, Osei S, Flier JS 1999 Regulation of neuronal and glial proteins by leptin: implications for brain development. *Endocrinology* 140:2755–2762
312. Bouret SG, Draper SJ, Simerly RB 2004 Trophic action of leptin on hypothalamic neurons that regulate feeding. *Science* 304:108–110
313. Heidenreich KA, Toledo SP 1989 Insulin receptors mediate growth effects in cultured fetal neurons. I. Rapid stimulation of protein synthesis. *Endocrinology* 125:1451–1457
314. Huck S 1983 Serum-free medium for cultures of the postnatal mouse cerebellum: only insulin is essential. *Brain Res Bull* 10:667–674
315. Schechter R, Yanovitch T, Abboud M, Johnson 3rd G, Gaskins J 1998 Effects of brain endogenous insulin on neurofilament and MAPK in fetal rat neuron cell cultures. *Brain Res* 808:270–278
316. Pinto S, Roseberry AG, Liu H, Diano S, Shanabrough M, Cai X, Friedman JM, Horvath TL 2004 Rapid rewiring of arcuate nucleus feeding circuits by leptin. *Science* 304:110–115
317. Kokoeva MV, Yin H, Flier JS 2005 Neurogenesis in the hypothalamus of adult mice: potential role in energy balance. *Science* 310:679–683
318. Xu Y, Tamamaki N, Noda T, Kimura K, Itokazu Y, Matsumoto N, Dezawa M, Ide C 2005 Neurogenesis in the ependymal layer of the adult rat 3rd ventricle. *Exp Neurol* 192:251–264
319. Fowler CD, Liu Y, Wang Z 2008 Estrogen and adult neurogenesis in the amygdala and hypothalamus. *Brain Res Rev* 57:342–351
320. Bouret SG, Draper SJ, Simerly RB 2004 Formation of projection pathways from the arcuate nucleus of the hypothalamus to hypothalamic regions implicated in the neural control of feeding behavior in mice. *J Neurosci* 24:2797–2805
321. Foster GA, Woodhams PL 1992 Neuropeptide Y. In: Björklund A, Hokfelt T, Tohyama M, eds. *Handbook of chemical neuroanatomy*. Vol 10. Ontogeny of transmitters and peptides in the CNS. Amsterdam: Elsevier; 521–546
322. Kagotani Y, Hashimoto T, Tsuruo Y, Kawano H, Daikoku S, Chihara K 1989 Development of the neuronal system containing neuropeptide Y in the rat hypothalamus. *Int J Dev Neurosci* 7:359–374
323. Woodhams PL, Allen YS, McGovern J, Allen JM, Bloom SR, Balazs R, Polak JM 1985 Immunohistochemical analysis of the early ontogeny of the neuropeptide Y system in rat brain. *Neuroscience* 15:173–202

324. Singer LK, Kuper J, Brogan RS, Smith MS, Grove KL 2000 Novel expression of hypothalamic neuropeptide Y during postnatal development in the rat. *Neuroreport* 11:1075–1080
325. Nowak FV, Gore AC 1999 Perinatal developmental changes in expression of the neuropeptide genes preoptic regulatory factor-1 and factor-2, neuropeptide Y and GnRH in rat hypothalamus. *J Neuroendocrinol* 11:951–958
326. Grove KL, Brogan RS, Smith MS 2001 Novel expression of neuropeptide Y (NPY) mRNA in hypothalamic regions during development: region-specific effects of maternal deprivation on NPY and agouti-related protein mRNA. *Endocrinology* 142:4771–4776
327. Allen JM, McGregor GP, Woodhams PL, Polak JM, Bloom SR 1984 Ontogeny of a novel peptide, neuropeptide Y (NPY) in rat brain. *Brain Res* 303:197–200
328. Leibowitz SF, Sepiashvili K, Akabayashi A, Karatayev O, Davydova Z, Alexander JT, Wang J, Chang GQ 2005 Function of neuropeptide Y and agouti-related protein at weaning: relation to corticosterone, dietary carbohydrate and body weight. *Brain Res* 1036:180–191
329. Grove KL, Allen S, Grayson BE, Smith MS 2003 Postnatal development of the hypothalamic neuropeptide Y system. *Neuroscience* 116:393–406
330. Nilsson I, Johansen JE, Schalling M, Hökfelt T, Fetissov SO 2005 Maturation of the hypothalamic arcuate agouti-related protein system during postnatal development in the mouse. *Brain Res Dev Brain Res* 155:147–154
331. Kowalski TJ, Houpt TA, Jahng J, Okada N, Chua Jr SC, Smith GP 1998 Ontogeny of neuropeptide Y expression in response to deprivation in lean Zucker rat pups. *Am J Physiol* 275:R466–R470
332. Capuano CA, Leibowitz SF, Barr GA 1993 Effect of paraventricular injection of neuropeptide Y on milk and water intake of preweanling rats. *Neuropeptides* 24:177–182
333. Hindelang C, Félix JM, Laurent FM, Klein MJ, Stoeckel ME 1990 Ontogenesis of proopiomelanocortin gene expression and regulation in the rat pituitary intermediate lobe. *Mol Cell Endocrinol* 70:225–235
334. Angelogianni P, Li HL, Gianoulakis C 2000 Ontogenesis of proopiomelanocortin and its processing to  $\beta$ -endorphin by the fetal and neonatal rat brain. *Neuroendocrinology* 72:231–241
335. Kerrigan JR, Martha Jr PM, Krieg Jr RJ, Queen TA, Monahan PE, Rogol AD 1991 Augmented hypothalamic proopiomelanocortin gene expression with pubertal development in the male rat: evidence for an androgen receptor-independent action. *Endocrinology* 128:1029–1035
336. Wiemann JN, Clifton DK, Steiner RA 1989 Pubertal changes in gonadotropin-releasing hormone and proopiomelanocortin gene expression in the brain of the male rat. *Endocrinology* 124:1760–1767
337. Khachaturian H, Alessi NE, Munfakh N, Watson SJ 1983 Ontogeny of opioid and related peptides in the rat CNS and pituitary: an immunocytochemical study. *Life Sci* 33(Suppl 1):61–64
338. Schwartzberg DG, Nakane PK 1982 Ontogenesis of adrenocorticotropin-related peptide determinants in the hypothalamus and pituitary gland of the rat. *Endocrinology* 110:855–864
339. Daikoku S, Chikamori M, Adachi T, Okamura Y, Nishiyama T, Tsuruo Y 1983 Ontogenesis of hypothalamic immunoreactive ACTH cells in vivo and in vitro: role of Rathke's pouch. *Dev Biol* 97:81–88
340. Monnet-Tschudi F, Eberle AN, Honegger P 1986 In vivo and in vitro development of  $\alpha$ -MSH and ACTH in the embryonic and postnatal rat brain. *Dev Brain Res* 26:125–132
341. Melnick I, Pronchuk N, Cowley MA, Grove KL, Colmers WF 2007 Developmental switch in neuropeptide Y and melanocortin effects in the paraventricular nucleus of the hypothalamus. *Neuron* 56:1103–1115
342. Baram TZ, Lerner SP 1991 Ontogeny of corticotropin-releasing hormone gene expression in rat hypothalamus—comparison with somatostatin. *Int J Dev Neurosci* 9:473–478
343. Brischoux F, Fellmann D, Risold PY 2001 Ontogenetic development of the diencephalic MCH neurons: a hypothalamic 'MCH area' hypothesis. *Eur J Neurosci* 13:1733–1744
344. Burgunder JM, Taylor T 1989 Ontogeny of thyrotropin-releasing hormone gene expression in the rat diencephalon. *Neuroendocrinology* 49:631–640
345. Grino M, Young 3rd WS, Burgunder JM 1989 Ontogeny of expression of the corticotropin-releasing factor gene in the hypothalamic paraventricular nucleus and of the proopiomelanocortin gene in rat pituitary. *Endocrinology* 124:60–68
346. Steining TL, Kilduff TS, Behan M, Benca RM, Landry CF 2004 Comparison of hypocretin/orexin and melanin-concentrating hormone neurons and axonal projections in the embryonic and postnatal rat brain. *J Chem Neuroanat* 27:165–181
347. Fuse Y, Polk DH, Lam RW, Fisher DA 1991 Ontogeny of thyrotropin releasing hormone and precursor peptide in the rat. *Pediatr Res* 30:28–33
348. Okamura Y, Kawano H, Daikoku S 1991 Spatial-temporal appearance of developing immunoreactive TRH neurons in the neuroepithelial wall of the diencephalon. *Brain Res Dev Brain Res* 63:21–31
349. Covarrubias L, Uribe RM, Méndez M, Charli JL, Joseph-Bravo P 1988 Neuronal TRH synthesis: developmental and circadian TRH mRNA levels. *Biochem Biophys Res Commun* 151:615–622
350. Emanuel RL, Thull DL, Girard DM, Majzoub JA 1989 Developmental expression of corticotropin releasing hormone messenger RNA and peptide in rat hypothalamus. *Peptides* 10:1165–1169
351. Taylor T, Gyves P, Burgunder JM 1990 Thyroid hormone regulation of TRH mRNA levels in rat paraventricular nucleus of the hypothalamus changes during ontogeny. *Neuroendocrinology* 52:262–267
352. Van Den Pol AN, Patrylo PR, Ghosh PK, Gao XB 2001 Lateral hypothalamus: early developmental expression and response to hypocretin (orexin). *J Comp Neurol* 433:349–363
353. Yamamoto Y, Ueta Y, Hara Y, Serino R, Nomura M, Shibuya I, Shirahata A, Yamashita H 2000 Postnatal development of orexin/hypocretin in rats. *Brain Res Mol Brain Res* 78:108–119
354. Sawai N, Ueta Y, Nakazato M, Ozawa H 2010 Developmental and aging change of orexin-A and -B immunore-



- active neurons in the male rat hypothalamus. *Neurosci Lett* 468:51–55
355. Clancy B, Darlington RB, Finlay BL 2001 Translating developmental time across mammalian species. *Neuroscience* 105:7–17
  356. Dobbing J, Sands J 1979 Comparative aspects of the brain growth spurt. *Early Hum Dev* 3:79–83
  357. McGee EA, Hsueh AJ 2000 Initial and cyclic recruitment of ovarian follicles. *Endocr Rev* 21:200–214
  358. Grayson BE, Allen SE, Billes SK, Williams SM, Smith MS, Grove KL 2006 Prenatal development of hypothalamic neuropeptide systems in the nonhuman primate. *Neuroscience* 143:975–986
  359. Wollmann HA 1998 Intrauterine growth restriction: definition and etiology. *Horm Res* 49(Suppl 2):1–6
  360. Gale CR, Martyn CN, Kellingray S, Eastell R, Cooper C 2001 Intrauterine programming of adult body composition. *J Clin Endocrinol Metab* 86:267–272
  361. Li H, Stein AD, Barnhart HX, Ramakrishnan U, Martorell R 2003 Associations between prenatal and postnatal growth and adult body size and composition. *Am J Clin Nutr* 77:1498–1505
  362. Singhal A, Wells J, Cole TJ, Fewtrell M, Lucas A 2003 Programming of lean body mass: a link between birth weight, obesity, and cardiovascular disease? *Am J Clin Nutr* 77:726–730
  363. Walker SP, Gaskin PS, Powell CA, Bennett FI 2002 The effects of birth weight and postnatal linear growth retardation on body mass index, fatness and fat distribution in mid and late childhood. *Public Health Nutr* 5:391–396
  364. Wells JC, Hallal PC, Wright A, Singhal A, Victora CG 2005 Fetal, infant and childhood growth: relationships with body composition in Brazilian boys aged 9 years. *Int J Obes (Lond)* 29:1192–1198
  365. Yliharsilä H, Kajantie E, Osmond C, Forsén T, Barker DJ, Eriksson JG 2007 Birth size, adult body composition and muscle strength in later life. *Int J Obes (Lond)* 31:1392–1399
  366. Hediger ML, Overpeck MD, Kuczmarski RJ, McGlynn A, Maurer KR, Davis WW 1998 Muscularity and fatness of infants and young children born small- or large-for-gestational-age. *Pediatrics* 102:E60
  367. Ibáñez L, Suárez L, Lopez-Bermejo A, Díaz M, Valls C, de Zegher F 2008 Early development of visceral fat excess after spontaneous catch-up growth in children with low birth weight. *J Clin Endocrinol Metab* 93:925–928
  368. Jaquet D, Gaboriau A, Czernichow P, Levy-Marchal C 2000 Insulin resistance early in adulthood in subjects born with intrauterine growth retardation. *J Clin Endocrinol Metab* 85:1401–1406
  369. Stein AD, Kahn HS, Rundle A, Zybert PA, van der Pal-de Bruin K, Lumey LH 2007 Anthropometric measures in middle age after exposure to famine during gestation: evidence from the Dutch famine. *Am J Clin Nutr* 85:869–876
  370. Barker M, Robinson S, Osmond C, Barker DJ 1997 Birth weight and body fat distribution in adolescent girls. *Arch Dis Child* 77:381–383
  371. Fall CH, Osmond C, Barker DJ, Clark PM, Hales CN, Stirling Y, Meade TW 1995 Fetal and infant growth and cardiovascular risk factors in women. *BMJ* 310:428–432
  372. Ibáñez L, Ong K, Dunger DB, de Zegher F 2006 Early development of adiposity and insulin resistance after catch-up weight gain in small-for-gestational-age children. *J Clin Endocrinol Metab* 91:2153–2158
  373. Ibáñez L, Lopez-Bermejo A, Suárez L, Marcos MV, Díaz M, de Zegher F 2008 Visceral adiposity without overweight in children born small-for-gestational-age. *J Clin Endocrinol Metab* 93:2079–2083
  374. Jaquet D, Deghmoun S, Chevenne D, Collin D, Czernichow P, Lévy-Marchal C 2005 Dynamic change in adiposity from fetal to postnatal life is involved in the metabolic syndrome associated with reduced fetal growth. *Diabetologia* 48:849–855
  375. Laitinen J, Pietiläinen K, Wadsworth M, Sovio U, Järvelin MR 2004 Predictors of abdominal obesity among 31-y-old men and women born in Northern Finland in 1966. *Eur J Clin Nutr* 58:180–190
  376. Law CM, Barker DJ, Osmond C, Fall CH, Simmonds SJ 1992 Early growth and abdominal fatness in adult life. *J Epidemiol Community Health* 46:184–186
  377. Martorell R, Stein AD, Schroeder DG 2001 Early nutrition and later adiposity. *J Nutr* 131:874S–880S
  378. Malina RM, Katzmarzyk PT, Beunen G 1996 Birth weight and its relationship to size attained and relative fat distribution at 7 to 12 years of age. *Obes Res* 4:385–390
  379. Te Velde SJ, Twisk JW, Van Mechelen W, Kemper HC 2003 Birth weight, adult body composition, and subcutaneous fat distribution. *Obes Res* 11:202–208
  380. Valdez R, Athens MA, Thompson GH, Bradshaw BS, Stern MP 1994 Birthweight and adult health outcomes in a biethnic population in the USA. *Diabetologia* 37:624–631
  381. Ekelund U, Ong K, Linné Y, Neovius M, Brage S, Dunger DB, Wareham NJ, Rössner S 2006 Upward weight percentile crossing in infancy and early childhood independently predicts fat mass in young adults: the Stockholm Weight Development Study (SWEDES). *Am J Clin Nutr* 83:324–330
  382. Ong KK, Ahmed ML, Emmett PM, Preece MA, Dunger DB 2000 Association between postnatal catch-up growth and obesity in childhood: prospective cohort study. *BMJ* 320:967–971
  383. Ong KK, Petry CJ, Emmett PM, Sandhu MS, Kiess W, Hales CN, Ness AR, Dunger DB 2004 Insulin sensitivity and secretion in normal children related to size at birth, postnatal growth, and plasma insulin-like growth factor-I levels. *Diabetologia* 47:1064–1070
  384. Ong KK, Emmett PM, Noble S, Ness A, Dunger DB 2006 Dietary energy intake at the age of 4 months predicts postnatal weight gain and childhood body mass index. *Pediatrics* 117:e503–e508
  385. Stettler N, Stallings VA, Troxel AB, Zhao J, Schinnar R, Nelson SE, Ziegler EE, Strom BL 2005 Weight gain in the first week of life and overweight in adulthood: a cohort study of European American subjects fed infant formula. *Circulation* 111:1897–1903
  386. Fetita LS, Sobngwi E, Serradas P, Calvo F, Gautier JF 2006 Consequences of fetal exposure to maternal diabetes in offspring. *J Clin Endocrinol Metab* 91:3718–3724
  387. Taylor PD, Poston L 2007 Developmental programming of obesity in mammals. *Exp Physiol* 92:287–298
  388. Catalano PM, Drago NM, Amini SB 1995 Maternal carbohydrate metabolism and its relationship to fetal growth

- and body composition. *Am J Obstet Gynecol* 172:1464–1470
389. Kliegman RM, Gross T 1985 Perinatal problems of the obese mother and her infant. *Obstet Gynecol* 66:299–306
  390. Silverman BL, Rizzo TA, Cho NH, Metzger BE 1998 Long-term effects of the intrauterine environment. The Northwestern University Diabetes in Pregnancy Center. *Diabetes Care* 21(Suppl 2):B142–B149
  391. Surkan PJ, Hsieh CC, Johansson AL, Dickman PW, Cnattingius S 2004 Reasons for increasing trends in large for gestational age births. *Obstet Gynecol* 104:720–726
  392. Villamor E, Cnattingius S 2006 Interpregnancy weight change and risk of adverse pregnancy outcomes: a population-based study. *Lancet* 368:1164–1170
  393. Oken E, Taveras EM, Kleinman KP, Rich-Edwards JW, Gillman MW 2007 Gestational weight gain and child adiposity at age 3 years. *Am J Obstet Gynecol* 196:322–328
  394. Chen Z, Du J, Shao L, Zheng L, Wu M, Ai M, Zhang Y 2010 Prepregnancy body mass index, gestational weight gain, and pregnancy outcomes in China. *Int J Gynaecol Obstet* 109:41–44
  395. Catalano PM, Thomas A, Huston-Presley L, Amini SB 2007 Phenotype of infants of mothers with gestational diabetes. *Diabetes Care* 30(Suppl 2):S156–S160
  396. Dabelea D, Hanson RL, Lindsay RS, Pettitt DJ, Imperatore G, Gabir MM, Roumain J, Bennett PH, Knowler WC 2000 Intrauterine exposure to diabetes conveys risks for type 2 diabetes and obesity: a study of discordant sibships. *Diabetes* 49:2208–2211
  397. Gale CR, Javaid MK, Robinson SM, Law CM, Godfrey KM, Cooper C 2007 Maternal size in pregnancy and body composition in children. *J Clin Endocrinol Metab* 92:3904–3911
  398. Hillier TA, Pedula KL, Schmidt MM, Mullen JA, Charles MA, Pettitt DJ 2007 Childhood obesity and metabolic imprinting: the ongoing effects of maternal hyperglycemia. *Diabetes Care* 30:2287–2292
  399. Hunter WA, Cundy T, Rabone D, Hofman PL, Harris M, Regan F, Robinson E, Cutfield WS 2004 Insulin sensitivity in the offspring of women with type 1 and type 2 diabetes. *Diabetes Care* 27:1148–1152
  400. Salsberry PJ, Reagan PB 2005 Dynamics of early childhood overweight. *Pediatrics* 116:1329–1338
  401. Whitaker RC 2004 Predicting preschooler obesity at birth: the role of maternal obesity in early pregnancy. *Pediatrics* 114:e29–e36
  402. Smith J, Cianflone K, Biron S, Hould FS, Lebel S, Marceau S, Lescelleur O, Biertho L, Simard S, Kral JG, Marceau P 2009 Effects of maternal surgical weight loss in mothers on intergenerational transmission of obesity. *J Clin Endocrinol Metab* 94:4275–4283
  403. Böhler T, Krämer T, Janecke AR, Hoffmann GF, Linderkamp O 1999 Increased energy expenditure and fecal fat excretion do not impair weight gain in small-for-gestational-age preterm infants. *Early Hum Dev* 54:223–234
  404. Brooke OG 1982 Energy requirements and utilization of the low birthweight infant. *Acta Paediatr Scand Suppl* 296: 67–70
  405. Cauderay M, Schutz Y, Micheli JL, Calame A, Jéquier E 1988 Energy-nitrogen balances and protein turnover in small and appropriate for gestational age low birthweight infants. *Eur J Clin Nutr* 42:125–136
  406. Chessex P, Reichman B, Verellen G, Putet G, Smith JM, Heim T, Swyer PR 1984 Metabolic consequences of intrauterine growth retardation in very low birthweight infants. *Pediatr Res* 18:709–713
  407. Davies PS, Clough H, Bishop NJ, Lucas A, Cole JJ, Cole TJ 1996 Total energy expenditure in small for gestational age infants. *Arch Dis Child Fetal Neonatal Ed* 75:F46–F48
  408. Jackson JA, Wailoo MP, Thompson JR, Petersen SA 2004 Early physiological development of infants with intrauterine growth retardation. *Arch Dis Child Fetal Neonatal Ed* 89:F46–F50
  409. Mericq V, Iñiguez G, Bazaes R, Bouwman C, Avila A, Salazar T, Carrasco F 2009 Differences in body composition and energy expenditure in prepubertal children born term or preterm appropriate or small for gestational age. *J Pediatr Endocrinol Metab* 22:1041–1050
  410. Ounsted M, Sleight G 1975 The infant's self-regulation of food intake and weight gain. Difference in metabolic balance after growth constraint or acceleration in utero. *Lancet* 1:1393–1397
  411. Boonstra VH, Arends NJ, Stijnen T, Blum WF, Akkerman O, Hokken-Koelega AC 2006 Food intake of children with short stature born small for gestational age before and during a randomized GH trial. *Horm Res* 65:23–30
  412. Lussana F, Painter RC, Ocke MC, Buller HR, Bossuyt PM, Roseboom TJ 2008 Prenatal exposure to the Dutch famine is associated with a preference for fatty foods and a more atherogenic lipid profile. *Am J Clin Nutr* 88:1648–1652
  413. Stein AD, Rundle A, Wada N, Goldbohm RA, Lumey LH 2009 Associations of gestational exposure to famine with energy balance and macronutrient density of the diet at age 58 years differ according to the reference population used. *J Nutr* 139:1555–1561
  414. Gómez L, Carrascosa A, Yeste D, Potau N, Riqué S, Ruiz-Cuevas P, Almar J 1999 Leptin values in placental cord blood of human newborns with normal intrauterine growth after 30–42 weeks of gestation. *Horm Res* 51: 10–14
  415. Jaquet D, Leger J, Levy-Marchal C, Oury JF, Czernichow P 1998 Ontogeny of leptin in human fetuses and newborns: effect of intrauterine growth retardation on serum leptin concentrations. *J Clin Endocrinol Metab* 83:1243–1246
  416. Marchini G, Fried G, Ostlund E, Hagenäs L 1998 Plasma leptin in infants: relations to birth weight and weight loss. *Pediatrics* 101:429–432
  417. Martos-Moreno GA, Barrios V, Sáenz de Pipaón M, Pozo J, Dorransoro I, Martínez-Biarge M, Quero J, Argente J 2009 Influence of prematurity and growth restriction on the adipokine profile, IGF1, and ghrelin levels in cord blood: relationship with glucose metabolism. *Eur J Endocrinol* 161:381–389
  418. Matsuda J, Yokota I, Iida M, Murakami T, Naito E, Ito M, Shima K, Kuroda Y 1997 Serum leptin concentration in cord blood: relationship to birth weight and gender. *J Clin Endocrinol Metab* 82:1642–1644
  419. Ong KK, Ahmed ML, Sherriff A, Woods KA, Watts A, Golding J, Dunger DB 1999 Cord blood leptin is associated with size at birth and predicts infancy weight gain in humans. *J Clin Endocrinol Metab* 84:1145–1148
  420. Ibáñez L, Sebastiani G, Diaz M, Gómez-Roig MD, Lopez-

- Bermejo A, de Zegher F 2010 Low body adiposity and high leptinemia in breast-fed infants born small-for-gestational-age. *J Pediatr* 156:145–147
421. Jaquet D, Leger J, Tabone MD, Czernichow P, Levy-Marchal C 1999 High serum leptin concentrations during catch-up growth of children born with intrauterine growth retardation. *J Clin Endocrinol Metab* 84:1949–1953
422. Martínez-Aguayo A, Capurro T, Peña V, Iñiguez G, Hernández MI, Avila A, Salazar T, Asenjo S, Mericq V 2007 Comparison of leptin levels, body composition and insulin sensitivity and secretion by OGTT in healthy, early pubertal girls born at either appropriate- or small-for-gestational age. *Clin Endocrinol (Oxf)* 67:526–532
423. Phillips DI, Fall CH, Cooper C, Norman RJ, Robinson JS, Owens PC 1999 Size at birth and plasma leptin concentrations in adult life. *Int J Obes Relat Metab Disord* 23:1025–1029
424. Singhal A, Farooqi IS, O’Rahilly S, Cole TJ, Fewtrell M, Lucas A 2002 Early nutrition and leptin concentrations in later life. *Am J Clin Nutr* 75:993–999
425. Farquhar J, Heiman M, Wong AC, Wach R, Chessex P, Chanoine JP 2003 Elevated umbilical cord ghrelin concentrations in small for gestational age neonates. *J Clin Endocrinol Metab* 88:4324–4327
426. Onal EE, Cinaz P, Atalay Y, Türkyilmaz C, Bideci A, Aktürk A, Okumu<sup>o</sup> N, Unal S, Koç E, Ergenekon E 2004 Umbilical cord ghrelin concentrations in small- and appropriate-for-gestational age newborn infants: relationship to anthropometric markers. *J Endocrinol* 180:267–271
427. Iñiguez G, Ong K, Peña V, Avila A, Dunger D, Mericq V 2002 Fasting and post-glucose ghrelin levels in SGA infants: relationships with size and weight gain at one year of age. *J Clin Endocrinol Metab* 87:5830–5833
428. Mahajan SD, Aalinkel R, Singh S, Shah P, Gupta N, Kochupillai N 2005 Thyroid hormone dysregulation in intrauterine growth retardation associated with maternal malnutrition and/or anemia. *Horm Metab Res* 37:633–640
429. Radetti G, Renzullo L, Gottardi E, D’Addato G, Messner H 2004 Altered thyroid and adrenal function in children born at term and preterm, small for gestational age. *J Clin Endocrinol Metab* 89:6320–6324
430. McMillen IC, Robinson JS 2005 Developmental origins of the metabolic syndrome: prediction, plasticity, and programming. *Physiol Rev* 85:571–633
431. Bieswal F, Hay SM, McKinnon C, Reusens B, Cuignet M, Rees WD, Remacle C 2004 Prenatal protein restriction does not affect the proliferation and differentiation of rat preadipocytes. *J Nutr* 134:1493–1499
432. Coupé B, Grit I, Darmaun D, Parnet P 2009 The timing of “catch-up growth” affects metabolism and appetite regulation in male rats born with intrauterine growth restriction. *Am J Physiol Regul Integr Comp Physiol* 297:R813–R824
433. Lucas A, Baker BA, Desai M, Hales CN 1996 Nutrition in pregnant or lactating rats programs lipid metabolism in the offspring. *Br J Nutr* 76:605–612
434. Shepherd PR, Crowther NJ, Desai M, Hales CN, Ozanne SE 1997 Altered adipocyte properties in the offspring of protein malnourished rats. *Br J Nutr* 78:121–129
435. Tonkiss J, Shukitt-Hale B, Formica RN, Rocco FJ, Galler JR 1990 Prenatal protein malnutrition alters response to reward in adult rats. *Physiol Behav* 48:675–680
436. Aihie Sayer A, Dunn R, Langley-Evans S, Cooper C 2001 Prenatal exposure to a maternal low protein diet shortens life span in rats. *Gerontology* 47:9–14
437. Langley-Evans SC, Sculley DV 2006 The association between birthweight and longevity in the rat is complex and modulated by maternal protein intake during fetal life. *FEBS Lett* 580:4150–4153
438. McCarthy HD, Pickard CL, Speed J, Jackson AA 1994 Sexual dimorphism of macronutrient selection and regional adipose tissue accumulation following in utero exposure to maternal low-protein diet. *Proc Nutr Soc* 53:172A (Abstract)
439. Rees WD, Hay SM, Cruickshank M, Reusens B, Remacle C, Antipatis C, Grant G 2006 Maternal protein intake in the pregnant rat programs the insulin axis and body composition in the offspring. *Metabolism* 55:642–649
440. Bellinger L, Lilley C, Langley-Evans SC 2004 Prenatal exposure to a maternal low-protein diet programmes a preference for high-fat foods in the young adult rat. *Br J Nutr* 92:513–520
441. Bellinger L, Sculley DV, Langley-Evans SC 2006 Exposure to undernutrition in fetal life determines fat distribution, locomotor activity and food intake in ageing rats. *Int J Obes (Lond)* 30:729–738
442. Zambrano E, Bautista CJ, Deás M, Martínez-Samayoa PM, González-Zamorano M, Ledesma H, Morales J, Larrea F, Nathanielsz PW 2006 A low maternal protein diet during pregnancy and lactation has sex- and window of exposure-specific effects on offspring growth and food intake, glucose metabolism and serum leptin in the rat. *J Physiol* 571:221–230
443. Ozanne SE, Nicholas Hales C 2005 Poor fetal growth followed by rapid postnatal catch-up growth leads to premature death. *Mech Ageing Dev* 126:852–854
444. Bieswal F, Ahn MT, Reusens B, Holvoet P, Raes M, Rees WD, Remacle C 2006 The importance of catch-up growth after early malnutrition for the programming of obesity in male rat. *Obesity (Silver Spring)* 14:1330–1343
445. Desai M, Gayle D, Babu J, Ross MG 2005 Programmed obesity in intrauterine growth-restricted newborns: modulation by newborn nutrition. *Am J Physiol Regul Integr Comp Physiol* 288:R91–R96
446. Holemans K, Verhaeghe J, Dequeker J, Van Assche FA 1996 Insulin sensitivity in adult female rats subjected to malnutrition during the perinatal period. *J Soc Gynecol Invest* 3:71–77
447. Jimenez-Chillaron JC, Hernandez-Valencia M, Lightner A, Faucette RR, Reamer C, Przybyla R, Ruest S, Barry K, Otis JP, Patti ME 2006 Reductions in caloric intake and early postnatal growth prevent glucose intolerance and obesity associated with low birthweight. *Diabetologia* 49:1974–1984
448. Perez H, Nunez H, Ruiz S, White A, Gotteland M 2004 Hypertension induced by fetal exposure to maternal undernutrition increased plasmatic corticosterone and hypothalamic corticotropin-releasing factor mRNA expression in the rat. *FENS Abstr* 2:A148.19 (Abstract)
449. Thompson NM, Norman AM, Donkin SS, Shankar RR, Vickers MH, Miles JL, Breier BH 2007 Prenatal and postnatal pathways to obesity: different underlying mecha-



- nisms, different metabolic outcomes. *Endocrinology* 148:2345–2354
450. **Vickers MH, Reddy S, Ikenasio BA, Breier BH** 2001 Dysregulation of the adipoinular axis—a mechanism for the pathogenesis of hyperleptinemia and adipogenic diabetes induced by fetal programming. *J Endocrinol* 170:323–332
  451. **Vickers MH, Gluckman PD, Coveny AH, Hofman PL, Cutfield WS, Gertler A, Breier BH, Harris M** 2008 The effect of neonatal leptin treatment on postnatal weight gain in male rats is dependent on maternal nutritional status during pregnancy. *Endocrinology* 149:1906–1913
  452. **Anguita RM, Sigulem DM, Sawaya AL** 1993 Intrauterine food restriction is associated with obesity in young rats. *J Nutr* 123:1421–1428
  453. **Jones AP, Friedman MI** 1982 Obesity and adipocyte abnormalities in offspring of rats undernourished during pregnancy. *Science* 215:1518–1519
  454. **Jones AP, Assimon SA, Friedman MI** 1986 The effect of diet on food intake and adiposity in rats made obese by gestational undernutrition. *Physiol Behav* 37:381–386
  455. **Knight BS, Pennell CE, Adamson SL, Lye SJ** 2007 The impact of murine strain and gender on postnatal development after maternal dietary restriction during pregnancy. *J Physiol* 581:873–881
  456. **Krechowec SO, Vickers M, Gertler A, Breier BH** 2006 Prenatal influences on leptin sensitivity and susceptibility to diet-induced obesity. *J Endocrinol* 189:355–363
  457. **Vickers MH, Ikenasio BA, Breier BH** 2001 IGF-I treatment reduces hyperphagia, obesity, and hypertension in metabolic disorders induced by fetal programming. *Endocrinology* 142:3964–3973
  458. **Fiorotto ML, Davis TA, Schoknecht P, Mersmann HJ, Pond WG** 1995 Both maternal over- and undernutrition during gestation increase the adiposity of young adult progeny in rats. *Obes Res* 3:131–141
  459. **Ikenasio-Thorpe BA, Breier BH, Vickers MH, Fraser M** 2007 Prenatal influences on susceptibility to diet-induced obesity are mediated by altered neuroendocrine gene expression. *J Endocrinol* 193:31–37
  460. **Léonhardt M, Lesage J, Croix D, Dutriez-Casteloot I, Beauvillain JC, Dupouy JP** 2003 Effects of perinatal maternal food restriction on pituitary-gonadal axis and plasma leptin level in rat pup at birth and weaning and on timing of puberty. *Biol Reprod* 68:390–400
  461. **Miñana-Solis Mdel C, Escobar C** 2007 Increased susceptibility to metabolic alterations in young adult females exposed to early malnutrition. *Int J Biol Sci* 3:12–19
  462. **Guan H, Arany E, van Beek JP, Chamson-Reig A, Thyssen S, Hill DJ, Yang K** 2005 Adipose tissue gene expression profiling reveals distinct molecular pathways that define visceral adiposity in offspring of maternal protein-restricted rats. *Am J Physiol Endocrinol Metab* 288:E663–E673
  463. **Passos MC, da Fonte Ramos C, Dutra SC, Mouço T, de Moura EG** 2002 Long-term effects of malnutrition during lactation on the thyroid function of offspring. *Horm Metab Res* 34:40–43
  464. **Passos MC, Vicente LL, Lisboa PC, de Moura EG** 2004 Absence of anorectic effect to acute peripheral leptin treatment in adult rats whose mothers were malnourished during lactation. *Horm Metab Res* 36:625–629
  465. **Ramos CF, Lima AP, Teixeira CV, Brito PD, Moura EG** 1997 Thyroid function in post-weaning rats whose dams were fed a low-protein diet during suckling. *Braz J Med Biol Res* 30:133–137
  466. **Stocker C, O'Dowd J, Morton NM, Wargent E, Sennitt MV, Hislop D, Glund S, Seckl JR, Arch JR, Cawthorne MA** 2004 Modulation of susceptibility to weight gain and insulin resistance in low birthweight rats by treatment of their mothers with leptin during pregnancy and lactation. *Int J Obes Relat Metab Disord* 28:129–136
  467. **Teixeira C, Passos M, Ramos C, Dutra S, Moura E** 2002 Leptin serum concentration, food intake and body weight in rats whose mothers were exposed to malnutrition during lactation. *J Nutr Biochem* 13:493–498
  468. **Chen H, Morris MJ** 2009 Differential responses of orexigenic neuropeptides to fasting in offspring of obese mothers. *Obesity (Silver Spring)* 17:1356–1362
  469. **Nivoit P, Morens C, Van Assche FA, Jansen E, Poston L, Remacle C, Reusens B** 2009 Established diet-induced obesity in female rats leads to offspring hyperphagia, adiposity and insulin resistance. *Diabetologia* 52:1133–1142
  470. **Chen H, Simar D, Lambert K, Mercier J, Morris MJ** 2008 Maternal and postnatal overnutrition differentially impact appetite regulators and fuel metabolism. *Endocrinology* 149:5348–5356
  471. **Chen H, Simar D, Morris MJ** 2009 Hypothalamic neuroendocrine circuitry is programmed by maternal obesity: interaction with postnatal nutritional environment. *PLoS One* 4:e6259
  472. **Férézou-Viala J, Roy AF, Sérougne C, Grippois D, Parquet M, Baillieux V, Gertler A, Delplanque B, Djiane J, Riottot M, Taouis M** 2007 Long-term consequences of maternal high-fat feeding on hypothalamic leptin sensitivity and diet-induced obesity in the offspring. *Am J Physiol Regul Integr Comp Physiol* 293:R1056–R1062
  473. **Gupta A, Srinivasan M, Thamadilok S, Patel MS** 2009 Hypothalamic alterations in fetuses of high fat diet-fed obese female rats. *J Endocrinol* 200:293–300
  474. **Morris MJ, Chen H** 2009 Established maternal obesity in the rat reprograms hypothalamic appetite regulators and leptin signaling at birth. *Int J Obes (Lond)* 33:115–122
  475. **Shankar K, Harrell A, Liu X, Gilchrist JM, Ronis MJ, Badger TM** 2008 Maternal obesity at conception programs obesity in the offspring. *Am J Physiol Regul Integr Comp Physiol* 294:R528–R538
  476. **White CL, Purpera MN, Morrison CD** 2009 Maternal obesity is necessary for programming effect of high-fat diet on offspring. *Am J Physiol Regul Integr Comp Physiol* 296:R1464–R1472
  477. **Akyol A, Langley-Evans SC, McMullen S** 2009 Obesity induced by cafeteria feeding and pregnancy outcome in the rat. *Br J Nutr* 102:1601–1610
  478. **Bouanane S, Merzouk H, Benkalfat NB, Soulimane N, Merzouk SA, Gresti J, Tessier C, Narce M** 20 May 2010 Hepatic and very low-density lipoprotein fatty acids in obese offspring of overfed dams. *Metabolism* doi:10.1016/j.metabol.2010.04.003
  479. **Gorski JN, Dunn-Meynell AA, Levin BE** 2007 Maternal obesity increases hypothalamic leptin receptor expression and sensitivity in juvenile obesity-prone rats. *Am J Physiol Regul Integr Comp Physiol* 292:R1782–R1791
  480. **Kirk SL, Samuelsson AM, Argenton M, Dhonye H, Kalamatianos T, Poston L, Taylor PD, Coen CW** 2009 Mater-

- nal obesity induced by diet in rats permanently influences central processes regulating food intake in offspring. *PLoS One* 4:e5870
481. Levin BE, Govek E 1998 Gestational obesity accentuates obesity in obesity-prone progeny. *Am J Physiol* 275:R1374–R1379
  482. Levin BE, Dunn-Meynell AA 2002 Maternal obesity alters adiposity and monoamine function in genetically predisposed offspring. *Am J Physiol Regul Integr Comp Physiol* 283:R1087–R1093
  483. Bayol SA, Farrington SJ, Stickland NC 2007 A maternal ‘junk food’ diet in pregnancy and lactation promotes an exacerbated taste for ‘junk food’ and a greater propensity for obesity in rat offspring. *Br J Nutr* 98:843–851
  484. Samuelsson AM, Matthews PA, Argenton M, Christie MR, McConnell JM, Jansen EH, Piersma AH, Ozanne SE, Twinn DF, Remacle C, Rowleron A, Poston L, Taylor PD 2008 Diet-induced obesity in female mice leads to offspring hyperphagia, adiposity, hypertension, and insulin resistance: a novel murine model of developmental programming. *Hypertension* 51:383–392
  485. Oben JA, Patel T, Mouralidarane A, Samuelsson AM, Matthews P, Pombo J, Morgan M, McKee C, Soeda J, Novelli M, Poston L, Taylor P 2010 Maternal obesity programmes offspring development of non-alcoholic fatty pancreas disease. *Biochem Biophys Res Commun* 394:24–28
  486. Boloker J, Gertz SJ, Simmons RA 2002 Gestational diabetes leads to the development of diabetes in adulthood in the rat. *Diabetes* 51:1499–1506
  487. Ericsson A, Säljö K, Sjöstrand E, Jansson N, Prasad PD, Powell TL, Jansson T 2007 Brief hyperglycaemia in the early pregnant rat increases fetal weight at term by stimulating placental growth and affecting placental nutrient transport. *J Physiol* 581:1323–1332
  488. Plagemann A, Harder T, Rake A, Melchior K, Rittel F, Rohde W, Dörner G 1998 Hypothalamic insulin and neuropeptide Y in the offspring of gestational diabetic mother rats. *Neuroreport* 9:4069–4073
  489. Yamashita H, Shao J, Qiao L, Pagliassotti M, Friedman JE 2003 Effect of spontaneous gestational diabetes on fetal and postnatal hepatic insulin resistance in *Lepr(db/+)* mice. *Pediatr Res* 53:411–418
  490. Plagemann A, Harder T, Janert U, Rake A, Rittel F, Rohde W, Dörner G 1999 Malformations of hypothalamic nuclei in hyperinsulinemic offspring of rats with gestational diabetes. *Dev Neurosci* 21:58–67
  491. Fahrenkrog S, Harder T, Stolaczyk E, Melchior K, Franke K, Dudenhausen JW, Plagemann A 2004 Cross-fostering to diabetic rat dams affects early development of mediobasal hypothalamic nuclei regulating food intake, body weight, and metabolism. *J Nutr* 134:648–654
  492. Khan NA 2007 Role of lipids and fatty acids in macrosomic offspring of diabetic pregnancy. *Cell Biochem Biophys* 48:79–88
  493. Plagemann A, Harder T, Melchior K, Rake A, Rohde W, Dörner G 1999 Elevation of hypothalamic neuropeptide Y-neurons in adult offspring of diabetic mother rats. *Neuroreport* 10:3211–3216
  494. Wigglesworth JS 1964 Experimental growth retardation in the foetal rat. *J Pathol Bacteriol* 88:1–13
  495. Neitzke U, Harder T, Schellong K, Melchior K, Ziska T, Rodekamp E, Dudenhausen JW, Plagemann A 2008 Intrauterine growth restriction in a rodent model and developmental programming of the metabolic syndrome: a critical appraisal of the experimental evidence. *Placenta* 29:246–254
  496. Engelbregt MJ, van Weissenbruch MM, Lips P, van Lingen A, Roos JC, Delemarre-van de Waal HA 2004 Body composition and bone measurements in intra-uterine growth retarded and early postnatally undernourished male and female rats at the age of 6 months: comparison with puberty. *Bone* 34:180–186
  497. Lugo G, O’Neil L, Cassady G 1971 Carcass water, fat, and chloride in the fetal growth-retarded rat. *Am J Obstet Gynecol* 110:358–361
  498. Nüsken KD, Dötsch J, Rauh M, Rascher W, Schneider H 2008 Uteroplacental insufficiency after bilateral uterine artery ligation in the rat: impact on postnatal glucose and lipid metabolism and evidence for metabolic programming of the offspring by sham operation. *Endocrinology* 149:1056–1063
  499. Schreuder MF, Fodor M, van Wijk JA, Delemarre-van de Waal HA 2006 Association of birth weight with cardiovascular parameters in adult rats during baseline and stressed conditions. *Pediatr Res* 59:126–130
  500. Tashima L, Nakata M, Anno K, Sugino N, Kato H 2001 Prenatal influence of ischemia-hypoxia-induced intrauterine growth retardation on brain development and behavioral activity in rats. *Biol Neonate* 80:81–87
  501. Hohenauer L, Oh W 1969 Body composition in experimental intrauterine growth retardation in the rat. *J Nutr* 99:23–26
  502. Kollée LA, Monnens LA, Trijbels JM, Veerkamp JH, Janssen AJ 1979 Experimental intrauterine growth retardation in the rat. Evaluation of the Wigglesworth model. *Early Hum Dev* 3:295–300
  503. Cha CJ, Gelardi NL, Oh W 1987 Growth and cellular composition in rats with intrauterine growth retardation: effects of postnatal nutrition. *J Nutr* 117:1463–1468
  504. Huizinga CT, Oudejans CB, Delemarre-van de Waal HA 2001 Persistent changes in somatostatin and neuropeptide Y mRNA levels but not in growth hormone-releasing hormone mRNA levels in adult rats after intrauterine growth retardation. *J Endocrinol* 168:273–281
  505. Ogata ES, Bussey ME, LaBarbera A, Finley S 1985 Altered growth, hypoglycemia, hypoalaninemia, and ketonemia in the young rat: postnatal consequences of intrauterine growth retardation. *Pediatr Res* 19:32–37
  506. Schreuder MF, van Wijk JA, Delemarre-van de Waal HA 2006 Intrauterine growth restriction increases blood pressure and central pulse pressure measured with telemetry in aging rats. *J Hypertens* 24:1337–1343
  507. Sadiq HF, deMello DE, Devaskar SU 1998 The effect of intrauterine growth restriction upon fetal and postnatal hepatic glucose transporter and glucokinase proteins. *Pediatr Res* 43:91–100
  508. Vuguin P, Raab E, Liu B, Barzilai N, Simmons R 2004 Hepatic insulin resistance precedes the development of diabetes in a model of intrauterine growth retardation. *Diabetes* 53:2617–2622
  509. Styurd J, Eriksson UJ, Grill V, Swenne I 2005 Experimental intrauterine growth retardation in the rat causes a reduc-

- tion of pancreatic B-cell mass, which persists into adulthood. *Biol Neonate* 88:122–128
510. Engelbregt MJ, van Weissenbruch MM, Popp-Snijders C, Lips P, Delemarre-van de Waal HA 2001 Body mass index, body composition, and leptin at onset of puberty in male and female rats after intrauterine growth retardation and after early postnatal food restriction. *Pediatr Res* 50:474–478
  511. Rajakumar PA, He J, Simmons RA, Devaskar SU 1998 Effect of uteroplacental insufficiency upon brain neuropeptide Y and corticotropin-releasing factor gene expression and concentrations. *Pediatr Res* 44:168–174
  512. Simmons RA, Templeton LJ, Gertz SJ 2001 Intrauterine growth retardation leads to the development of type 2 diabetes in the rat. *Diabetes* 50:2279–2286
  513. Selak MA, Storey BT, Peterside I, Simmons RA 2003 Impaired oxidative phosphorylation in skeletal muscle of intrauterine growth-retarded rats. *Am J Physiol Endocrinol Metab* 285:E130–E137
  514. Kennedy GC 1957 The effect of age on the somatic and visceral response to overnutrition in the rat. *Proc. Soc. Endocr.* 60th. *J Endocrinol* 15:xix-xxxiv
  515. Kennedy GC 1957 The development with age of hypothalamic restraint upon the appetite of the rat. *J Endocrinol* 16:9–17
  516. Aubert R, Suquet JP, Lemonnier D 1980 Long-term morphological and metabolic effects of early under- and overnutrition in mice. *J Nutr* 110:649–661
  517. Fiorotto ML, Burrin DG, Perez M, Reeds PJ 1991 Intake and use of milk nutrients by rat pups suckled in small, medium, or large litters. *Am J Physiol* 260:R1104–R1113
  518. Hausberger FX, Volz JE 1984 Feeding in infancy, adipose tissue cellularity and obesity. *Physiol Behav* 33:81–87
  519. Lambert EV, Koeslag JH 1992 No persistent effect of preweaning nutrition on postweaning food intake, feeding efficiency, or body energy stores in Long-Evans rats. *Physiol Behav* 52:363–372
  520. López M, Seoane LM, Tovar S, García MC, Nogueiras R, Diéguez C, Señaris RM 2005 A possible role of neuropeptide Y, agouti-related protein and leptin receptor isoforms in hypothalamic programming by perinatal feeding in the rat. *Diabetologia* 48:140–148
  521. Muralidhara DV, Shetty PS 1986 Effects of preweaning nutritional deprivation on basal metabolism and thermoregulatory thermogenesis in the rat. *Br J Nutr* 56:615–623
  522. Plagemann A, Harder T, Rake A, Waas T, Melchior K, Ziska T, Rohde W, Dörner G 1999 Observations on the orexigenic hypothalamic neuropeptide Y-system in neonatally overfed weanling rats. *J Neuroendocrinol* 11:541–546
  523. Plagemann A, Harder T, Rake A, Voits M, Fink H, Rohde W, Dörner G 1999 Perinatal elevation of hypothalamic insulin, acquired malformation of hypothalamic galaninergic neurons, and syndrome X-like alterations in adulthood of neonatally overfed rats. *Brain Res* 836:146–155
  524. Schmidt I, Schoelch C, Ziska T, Schneider D, Simon E, Plagemann A 2000 Interaction of genetic and environmental programming of the leptin system and of obesity disposition. *Physiol Genomics* 3:113–120
  525. Schmidt I, Fritz A, Schölch C, Schneider D, Simon E, Plagemann A 2001 The effect of leptin treatment on the development of obesity in overfed suckling Wistar rats. *Int J Obes Relat Metab Disord* 25:1168–1174
  526. Bassett DR, Craig BW 1988 Influence of early nutrition on growth and adipose tissue characteristics in male and female rats. *J Appl Physiol* 64:1249–1256
  527. Mozes S, Sefčíková Z, Lenhardt L, Racek L 2004 Obesity and changes of alkaline phosphatase activity in the small intestine of 40- and 80-day-old rats subjected to early postnatal overfeeding or monosodium glutamate. *Physiol Res* 53:177–186
  528. Velkoska E, Cole TJ, Morris MJ 2005 Early dietary intervention: long-term effects on blood pressure, brain neuropeptide Y, and adiposity markers. *Am J Physiol Endocrinol Metab* 288:E1236–E1243
  529. Boullu-Ciocca S, Dutour A, Guillaume V, Achard V, Oliver C, Grino M 2005 Postnatal diet-induced obesity in rats upregulates systemic and adipose tissue glucocorticoid metabolism during development and in adulthood: its relationship with the metabolic syndrome. *Diabetes* 54:197–203
  530. Davidowa H, Plagemann A 2001 Inhibition by insulin of hypothalamic VMN neurons in rats overweight due to postnatal overfeeding. *Neuroreport* 12:3201–3204
  531. Davidowa H, Li Y, Plagemann A 2002 Hypothalamic ventromedial and arcuate neurons of normal and postnatally overnourished rats differ in their responses to melanin-concentrating hormone. *Regul Pept* 108:103–111
  532. Davidowa H, Li Y, Plagemann A 2002 Differential response to NPY of PVH and dopamine-responsive VMH neurons in overweight rats. *Neuroreport* 13:1523–1527
  533. Faust IM, Johnson PR, Hirsch J 1980 Long-term effects of early nutritional experience on the development of obesity in the rat. *J Nutr* 110:2027–2034
  534. Heidel E, Plagemann A, Davidowa H 1999 Increased response to NPY of hypothalamic VMN neurons in postnatally overfed juvenile rats. *Neuroreport* 10:1827–1831
  535. Knittle JL, Hirsch J 1968 Effect of early nutrition on the development of rat epididymal fat pads: cellularity and metabolism. *J Clin Invest* 47:2091–2098
  536. Li Y, Plagemann A, Davidowa H 2002 Increased inhibition by agouti-related peptide of ventromedial hypothalamic neurons in rats overweight due to early postnatal overfeeding. *Neurosci Lett* 330:33–36
  537. López M, Tovar S, Vázquez MJ, Nogueiras R, Seoane LM, García M, Señaris RM, Diéguez C 2007 Perinatal overfeeding in rats results in increased levels of plasma leptin but unchanged cerebrospinal leptin in adulthood. *Int J Obes Lond* 31:371–377
  538. Miller DS, Parsonage SR 1972 The effect of litter size on subsequent energy utilization. *Proc Nutr Soc* 31:30A–31A (Abstract)
  539. Oscai LB, McGarr JA 1978 Evidence that the amount of food consumed in early life fixes appetite in the rat. *Am J Physiol* 235:R141–R144
  540. Oscai LB 1982 Dietary-induced severe obesity: a rat model. *Am J Physiol* 242:R212–R215
  541. Plagemann A, Heidrich I, Götz F, Rohde W, Dörner G 1992 Obesity and enhanced diabetes and cardiovascular risk in adult rats due to early postnatal overfeeding. *Exp Clin Endocrinol* 99:154–158
  542. Widdowson EM, McCance RA 1960 Some effects of ac-



- celerating growth. I. General somatic development. *Proc R Soc Lond B Biol Sci* 152:188–206
543. Wiedmer P, Klaus S, Ortman S 2002 Energy metabolism of young rats after early postnatal overnutrition. *Br J Nutr* 88:301–306
544. Houdijk ME, Engelbregt MT, Popp-Snijders C, Delemarre van der Waal HA 2003 Long-term effects of early postnatal food restriction on growth hormone secretion in rats. *JPEN J Parenter Enteral Nutr* 27:260–267
545. Remmers F, Fodor M, Delemarre-van de Waal HA 2008 Neonatal food restriction permanently alters rat body dimensions and energy intake. *Physiol Behav* 95:208–215
546. Williams JP, Tanner JM, Hughes PC 1974 Catch-up growth in female rats after growth retardation during the suckling period: comparison with males. *Pediatr Res* 8:157–162
547. Williams JP, Tanner JM, Hughes PC 1974 Catch-up growth in male rats after growth retardation during the suckling period. *Pediatr Res* 8:149–156
548. Erhuma A, Bellinger L, Langley-Evans SC, Bennett AJ 2007 Prenatal exposure to undernutrition and programming of responses to high-fat feeding in the rat. *Br J Nutr* 98:517–524
549. Sutton GM, Centanni AV, Butler AA 2010 Protein malnutrition during pregnancy in C57BL/6J mice results in offspring with altered circadian physiology before obesity. *Endocrinology* 151:1570–1580
550. Bol VV, Delattre AI, Reusens B, Raes M, Remacle C 2009 Forced catch-up growth after fetal protein restriction alters the adipose tissue gene expression program leading to obesity in adult mice. *Am J Physiol Regul Integr Comp Physiol* 297:R291–R299
551. Jones AP, Simson EL, Friedman MI 1984 Gestational undernutrition and the development of obesity in rats. *J Nutr* 114:1484–1492
552. Vickers MH, Gluckman PD, Coveny AH, Hofman PL, Cutfield WS, Gertler A, Breier BH, Harris M 2005 Neonatal leptin treatment reverses developmental programming. *Endocrinology* 146:4211–4216
553. Vickers MH, Breier BH, Cutfield WS, Hofman PL, Gluckman PD 2000 Fetal origins of hyperphagia, obesity, and hypertension and postnatal amplification by hypercaloric nutrition. *Am J Physiol Endocrinol Metab* 279:E83–E87
554. Toth MJ 2001 Comparing energy expenditure data among individuals differing in body size and composition: statistical and physiological considerations. *Curr Opin Clin Nutr Metab Care* 4:391–397
555. Vickers MH, Breier BH, McCarthy D, Gluckman PD 2003 Sedentary behavior during postnatal life is determined by the prenatal environment and exacerbated by postnatal hypercaloric nutrition. *Am J Physiol Regul Integr Comp Physiol* 285:R271–R273
556. Peterside IE, Selak MA, Simmons RA 2003 Impaired oxidative phosphorylation in hepatic mitochondria in growth-retarded rats. *Am J Physiol Endocrinol Metab* 285: E1258–E1266
557. Simmons RA, Saponitsky-Kroyter I, Selak MA 2005 Progressive accumulation of mitochondrial DNA mutations and decline in mitochondrial function lead to  $\beta$ -cell failure. *J Biol Chem* 280:28785–28791
558. Remmers F, Schreuder MF, Gemke RJ, Delemarre-van de Waal HA 2008 Energy intake and resting energy expenditure in adult male rats after early postnatal food restriction. *Br J Nutr* 99:1149–1156
559. Desai M, Gayle D, Han G, Ross MG 2007 Programmed hyperphagia due to reduced anorexigenic mechanisms in intrauterine growth-restricted offspring. *Reprod Sci* 14: 329–337
560. Morgan BL, Naismith DJ 1982 The effect of early postnatal undernutrition on the growth and development of the rat brain. *Br J Nutr* 48:15–23
561. Plagemann A, Harder T, Rake A, Melchior K, Rohde W, Dörner G 2000 Hypothalamic nuclei are malformed in weanling offspring of low protein malnourished rat dams. *J Nutr* 130:2582–2589
562. Plagemann A, Waas T, Harder T, Rittel F, Ziska T, Rohde W 2000 Hypothalamic neuropeptide Y levels in weaning offspring of low-protein malnourished mother rats. *Neuropeptides* 34:1–6
563. Cripps RL, Martin-Gronert MS, Archer ZA, Hales CN, Mercer JG, Ozanne SE 2009 Programming of hypothalamic neuropeptide gene expression in rats by maternal dietary protein content during pregnancy and lactation. *Clin Sci (Lond)* 117:85–93
564. García AP, Palou M, Priego T, Sánchez J, Palou A, Picó C 2010 Moderate caloric restriction during gestation results in lower arcuate nucleus NPY- and  $\alpha$ MSH-neurons and impairs hypothalamic response to fed/fasting conditions in weaned rats. *Diabetes Obes Metab* 12:403–413
565. Jezová D, Skultétová I, Makatsori A, Moncek F, Duncko R 2002 Hypothalamo-pituitary-adrenocortical axis function and hedonic behavior in adult male and female rats prenatally stressed by maternal food restriction. *Stress* 5:177–183
566. Pérez H, Ruiz S, Núñez H, White A, Gotteland M, Hernández A 2006 Paraventricular-coerulear interactions: role in hypertension induced by prenatal undernutrition in the rat. *Eur J Neurosci* 24:1209–1219
567. Delahaye F, Breton C, Risold PY, Enache M, Dutriez-Casteloot I, Laborie C, Lesage J, Vieau D 2008 Maternal perinatal undernutrition drastically reduces postnatal leptin surge and affects the development of arcuate nucleus POMC neurons in neonatal male rat pups. *Endocrinology* 149:470–475
568. Irani BG, Le Foll C, Dunn-Meynell AA, Levin BE 2009 Ventromedial nucleus neurons are less sensitive to leptin excitation in rats bred to develop diet-induced obesity. *Am J Physiol Regul Integr Comp Physiol* 296:R521–R527
569. Remmers F, Verhagen LA, Adan RA, Delemarre-van de Waal HA 2008 Hypothalamic neuropeptide expression of juvenile and middle-aged rats after early postnatal food restriction. *Endocrinology* 149:3617–3625
570. Feng P, Vurbic D, Wu Z, Strohl KP 2007 Brain orexins and wake regulation in rats exposed to maternal deprivation. *Brain Res* 1154:163–172
571. Kim HJ, Lee JH, Choi SH, Lee YS, Jahng JW 2005 Fasting-induced increases of arcuate NPY mRNA and plasma corticosterone are blunted in the rat experienced neonatal maternal separation. *Neuropeptides* 39:587–594
572. Bouret SG, Simerly RB 2007 Development of leptin-sensitive circuits. *J Neuroendocrinol* 19:575–582
573. Bouret SG, Simerly RB 2006 Developmental programming of hypothalamic feeding circuits. *Clin Genet* 70:295–301
574. Horvath TL, Bruning JC 2006 Developmental program-

- ming of the hypothalamus: a matter of fat. *Nat Med* 12:52–53
575. **Louis GW, Myers Jr MG** 2007 The role of leptin in the regulation of neuroendocrine function and CNS development. *Rev Endocr Metab Disord* 8:85–94
576. **McMillen IC, Edwards LJ, Duffield J, Muhlhausler BS** 2006 Regulation of leptin synthesis and secretion before birth: implications for the early programming of adult obesity. *Reproduction* 131:415–427
577. **Remacle C, Bieswal F, Reusens B** 2004 Programming of obesity and cardiovascular disease. *Int J Obes Relat Metab Disord* 28(Suppl 3):S46–S53
578. **Vickers MH** 2007 Developmental programming and adult obesity: the role of leptin. *Curr Opin Endocrinol Diabetes Obes* 14:17–22
579. **Nilsson C, Swolin-Eide D, Ohlsson C, Eriksson E, Ho HP, Björntorp P, Holmång A** 2003 Reductions in adipose tissue and skeletal growth in rat adult offspring after prenatal leptin exposure. *J Endocrinol* 176:13–21
580. **Picó C, Oliver P, Sánchez J, Miralles O, Caimari A, Priego T, Palou A** 2007 The intake of physiological doses of leptin during lactation in rats prevents obesity in later life. *Int J Obes (Lond)* 31:1199–1209
581. **Attig L, Solomon G, Ferezou J, Abdennebi-Najar L, Taouis M, Gertler A, Djiane J** 2008 Early postnatal leptin blockade leads to a long-term leptin resistance and susceptibility to diet-induced obesity in rats. *Int J Obes (Lond)* 32:1153–1160
582. **de Oliveira Cravo C, Teixeira CV, Passos MC, Dutra SC, de Moura EG, Ramos C** 2002 Leptin treatment during the neonatal period is associated with higher food intake and adult body weight in rats. *Horm Metab Res* 34:400–405
583. **Lins MC, de Moura EG, Lisboa PC, Bonomo IT, Passos MC** 2005 Effects of maternal leptin treatment during lactation on the body weight and leptin resistance of adult offspring. *Regul Pept* 127:197–202
584. **Toste FP, de Moura EG, Lisboa PC, Fagundes AT, de Oliveira E, Passos MC** 2006 Neonatal leptin treatment programmes leptin hypothalamic resistance and intermediary metabolic parameters in adult rats. *Br J Nutr* 95:830–837
585. **Stocker CJ, Wargent E, O'Dowd J, Cornick C, Speakman JR, Arch JR, Cawthorne MA** 2007 Prevention of diet-induced obesity and impaired glucose tolerance in rats following administration of leptin to their mothers. *Am J Physiol Regul Integr Comp Physiol* 292:R1810–R1818
586. **Stocker CJ, Cawthorne MA** 2008 The influence of leptin on early life programming of obesity. *Trends Biotechnol* 26:545–551
587. **de Moura EG, Passos MC** 2005 Neonatal programming of body weight regulation and energetic metabolism. *Biosci Rep* 25:251–269
588. **Fowden AL, Giussani DA, Forhead AJ** 2005 Endocrine and metabolic programming during intrauterine development. *Early Hum Dev* 81:723–734
589. **Plagemann A** 2008 A matter of insulin: developmental programming of body weight regulation. *J Matern Fetal Neonatal Med* 21:143–148
590. **Plagemann A, Harder T, Rake A, Janert U, Melchior K, Rohde W, Dörner G** 1999 Morphological alterations of hypothalamic nuclei due to intrahypothalamic hyperinsulinism in newborn rats. *Int J Dev Neurosci* 17:37–44
591. **Bouret SG** 2009 Early life origins of obesity: role of hypothalamic programming. *J Pediatr Gastroenterol Nutr* 48(Suppl 1):S31–S38
592. **Levin BE** 2006 Metabolic imprinting: critical impact of the perinatal environment on the regulation of energy homeostasis. *Philos Trans R Soc Lond B Biol Sci* 361:1107–1121
593. **Sullivan EL, Grove KL** 2010 Metabolic imprinting in obesity. *Forum Nutr* 63:186–194
594. **Langley-Evans SC** 2009 Nutritional programming of disease: unravelling the mechanism. *J Anat* 215:36–51
595. **Seckl JR, Holmes MC** 2007 Mechanisms of disease: glucocorticoids, their placental metabolism and fetal ‘programming’ of adult pathophysiology. *Nat Clin Pract Endocrinol Metab* 3:479–488
596. **Neel JV** 1962 Diabetes mellitus: a “thrifty” genotype rendered detrimental by “progress”? *Am J Hum Genet* 14:353–362
597. **Freathy RM, Bennett AJ, Ring SM, Shields B, Groves CJ, Timpson NJ, Weedon MN, Zeggini E, Lindgren CM, Lango H, Perry JR, Pouta A, Ruokonen A, Hyppönen E, Power C, Elliott P, Strachan DP, Jarvelin MR, Smith GD, McCarthy MI, Frayling TM, Hattersley AT** 2009 Type 2 diabetes risk alleles are associated with reduced size at birth. *Diabetes* 58:1428–1433
598. **Freathy RM, Mook-Kanamori DO, Sovio U, Prokopenko I, Timpson NJ, Berry DJ, Warrington NM, Widen E, Hottenga JJ, Kaakinen M, Lange LA, Bradfield JP, Kerkhof M, Marsh JA, Mägi R, Chen CM, Lyon HN, Kirin M, Adair LS, Aulchenko YS, Bennett AJ, Borja JB, Bouatia-Naji N, Charoen P, Coin LJ, Cousminer DL, de Geus EJ, Deloukas P, Elliott P, Evans DM, Froguel P, Glaser B, Groves CJ, Hartikainen AL, Hassanali N, et al.** 2010 Variants in *ADCY5* and near *CCNL1* are associated with fetal growth and birth weight. *Nat Genet* 42:430–435
599. **Paxinos G, Watson C** 1997 The rat brain in stereotaxic coordinates. 3rd ed. New York: Academic Press