

# Differential activities of the RET tyrosine kinase receptor isoforms during mammalian embryogenesis

Esther de Graaff,<sup>1,4</sup> Shankar Srinivas,<sup>2,5</sup> Carol Kilkenny,<sup>1,7</sup> Vivette D'Agati,<sup>3</sup> Baljinder S. Mankoo,<sup>1,7</sup> Frank Costantini,<sup>2</sup> and Vassilis Pachnis<sup>1,6,8</sup>

<sup>1</sup>Division of Developmental Neurobiology, Medical Research Council (MRC) National Institute for Medical Research, The Ridgeway, Mill Hill, London NW7 1AA, UK; <sup>2</sup>Department of Genetics and Development and <sup>3</sup>Department of Pathology, College of Physicians and Surgeons, Columbia University, New York, New York 10032, USA

The RET receptor tyrosine kinase has a critical role in kidney organogenesis and the development of the enteric nervous system. Two major isoforms, RET9 and RET51, differ in the amino acid sequence of the C-terminal tail as a result of alternative splicing. To determine the roles of these isoforms *in vivo*, we used targeted mutagenesis to generate mice that express either RET9 or RET51. Monoisoformic RET9 mice, which lack RET51, are viable and appear normal. In contrast, monoisoformic RET51 animals, which lack RET9, have kidney hypodysplasia and lack enteric ganglia from the colon. To study the differential activities of the two RET isoforms further, we generated transgenic mice expressing ligand-dependent and constitutively active forms of RET9 or RET51 under the control of the *Hoxb7* regulatory sequences. Such RET9 transgenes are capable of rescuing the kidney agenesis in RET-deficient mice or causing kidney hypodysplasia in wild-type animals. In contrast, similar RET51 transgenes fail to rescue the kidney agenesis or cause hypodysplasia. Our findings show that RET9 and RET51 have different signaling properties *in vivo* and define specific temporal and spatial requirements of *c-Ret* function during renal development and histogenesis of the enteric nervous system.

[Key Words: c-Ret; signaling; Hirschsprung's disease; nephrogenesis]

Received April 9, 2001; revised version accepted July 19, 2001.

The *c-Ret* proto-oncogene encodes a receptor tyrosine kinase (RTK) that is expressed widely in mammalian embryos and has diverse roles in development and disease (Takahashi et al. 1985, 1988; Taraviras and Pachnis 1999; Baloh et al. 2000; Jhiang 2000; Parisi and Kapur, 2000; Schedl and Hastie 2000). During embryogenesis, the main sites of *c-Ret* expression are the excretory and the nervous systems (Pachnis et al. 1993; Tsuzuki et al. 1995). In the nervous system, *c-Ret* is expressed in the progenitors of the enteric nervous system (ENS), in the enteric, autonomic, and sensory neurons of the peripheral nervous system (PNS), and in the motor and catecholaminergic neurons of the central nervous system (CNS).

Mice homozygous for a targeted mutation of *c-Ret* (*Ret.k-*) have severe hypodysplasia or aplasia of the kidneys (Schuchardt et al. 1994, 1996; Durbec et al. 1996; Srinivas et al. 1999). The mammalian kidney is generated by reciprocal inductive interactions between the ureteric bud (UB) and the undifferentiated metanephric mesenchyme. Whereas the mesenchyme is important for the growth and branching of the UB, which gives rise to the renal collecting system, the tips of the UB branches induce the surrounding mesenchymal cells to condense into epithelial vesicles, which differentiate into the various segments of the nephrons (Saxen 1987). *c-Ret* is expressed in the UB and its function is necessary for the evagination, growth, and branching of this structure (Pachnis et al. 1993; Schuchardt et al. 1994, 1996; Sainio et al. 1997; Ehrenfels et al. 1999; Schedl and Hastie 2000).

*c-Ret* also has a critical role in the development of the ENS. The majority of enteric neurons and glia are derived from a subset of neural crest (NC) cells that emigrate from the neural tube at the level of somites 1–7. On invading the foregut mesenchyme, enteric NC (ENC) cells migrate in a rostrocaudal direction and colonize the wall of the gastrointestinal tract (Yntema and Hammond

Present addresses: <sup>4</sup>Department of Clinical Genetics, Erasmus University, Dr. Molewaterplein 50, 3015 GE Rotterdam, The Netherlands; <sup>5</sup>Division of Mammalian Development and <sup>6</sup>Division of Molecular Neurobiology, MRC National Institute for Medical Research, The Ridgeway, Mill Hill, London NW7 1AA, UK; <sup>7</sup>Randall Centre for Molecular Mechanisms of Cell Function, GKT School of Biomedical Sciences, New Hunt's House, King's College London, Guy's Campus, London SE1 1UL, UK. <sup>8</sup>Corresponding author.

E-MAIL [vpachni@nimr.mrc.ac.uk](mailto:vpachni@nimr.mrc.ac.uk); FAX 44-20-89138536.

Article and publication are at <http://www.genesdev.org/cgi/doi/10.1101/gad.205001>.

1954; Le Douarin and Teillet 1973; Kapur et al. 1992; Durbec et al. 1996). Mutations of *c-RET* in humans lead to absence of enteric ganglia from the distal colon and congenital megacolon (Hirschsprung's disease, HSCR; Parisi and Kapur 2000), whereas *Ret.k-/-Ret.k-* mice lack all enteric ganglia posterior to the stomach (intestinal aganglionosis; Schuchardt et al. 1994; Durbec et al. 1996). Although some of the cellular processes controlled by RET (such as survival, migration, and differentiation of ENC) have been identified, the mechanisms that lead to the localized absence of enteric ganglia in HSCR patients remain unclear.

Gain-of-function mutations of *c-Ret* have also been described in humans and reproduced in mice by targeted mutagenesis. These mutations result in the constitutive activation of RET and are associated with multiple endocrine neoplasia (MEN) types 2A and 2B and familial medullary thyroid carcinoma (FMTTC), inherited cancer syndromes characterized by tumors of neuroendocrine origin (Jhiang 2000; Smith-Hicks et al. 2000).

RET is the signaling component of multisubunit receptor complexes for glial cell line-derived neurotrophic factor (GDNF) family ligands (GFLs), which belong to the TGF- $\beta$  superfamily (Baloh et al. 2000; Saarma 2000). The interaction between the GFLs and RET is mediated by glycosylphosphatidylinositol (GPI)-linked cell-surface glycoproteins, called GFR $\alpha$ 1–4 (Airaksinen et al. 1999). Among the signaling pathways activated by RET are the MAP kinase (MAPK) and the PI3-K pathway (van Weering and Bos 1998). As is the case for other RTKs, phosphorylated tyrosine (Tyr) residues in RET are critical for the formation of docking sites for intracellular adaptor and effector signaling molecules, such as phospholipase C- $\gamma$ , Shc, Enigma, SNT/FRS2, Gab1, Nck, Crk, Grb2, and p62<sup>DOC</sup> (van Weering and Bos 1998; Airaksinen et al. 1999; Hansford and Mulligan 2000). Among these Tyr residues, Tyr 1062 has a critical role in signal initiation during embryogenesis and tumorigenesis, serving as a

docking site for protein complexes that can activate both the MAPK and the PI3-K pathways (Besset et al. 2000; Hayashi et al. 2000).

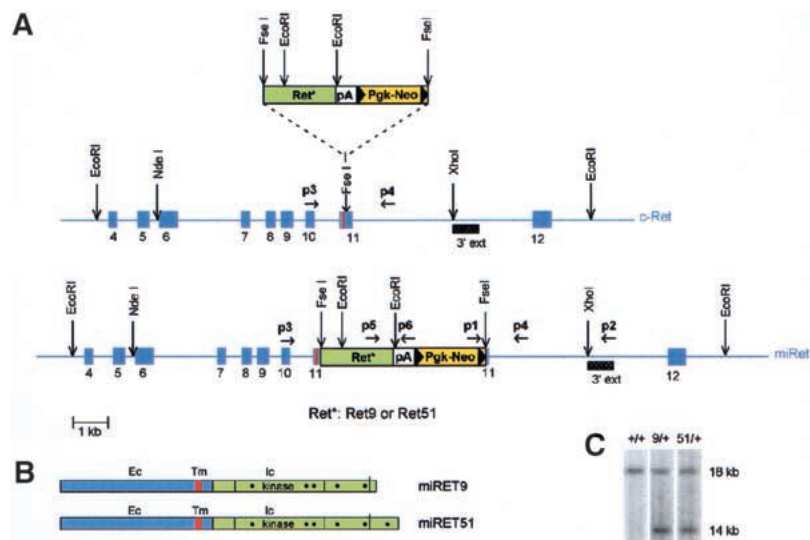
In mammals, *c-Ret* encodes two major isoforms, RET9 and RET51, which are generated by alternative splicing and differ only at their C-terminal tails; RET9 has a nine-amino-acid tail, which in RET51, is replaced by 51 unrelated amino acids (Tahira et al. 1990). Although the two isoforms behave similarly in a number of in vitro assays, several observations have suggested that they have different and possibly tissue-specific effects on embryogenesis and tumorigenesis. To address this issue, we used targeted mutagenesis in embryonic stem (ES) cells and pronuclear DNA microinjection to generate mice that express single RET isoforms. Our experiments reveal that RET9 is sufficient to support normal embryogenesis and postnatal life. Mice expressing only RET51, however, have severe defects in the innervation of the gut and renal development. Furthermore, activated forms of RET9 and RET51 differ in their ability to induce dominant renal malformations in transgenic mice. We discuss the implications of these findings in ENS histogenesis and nephrogenesis.

## Results

### Generation of mice expressing single RET isoforms

To study the relative roles of the RET isoforms in vivo, we first targeted the mouse *c-Ret* locus and inserted a cDNA fragment encoding the intracellular portion of the human RET9 or RET51 isoforms. Our goal was to disrupt the wild-type gene and replace it with alleles encoding single chimeric receptors composed of the extracellular domain of mouse RET fused in-frame to the intracellular segment of human RET9 or RET51 (Fig. 1A,B). We named these alleles *monoisoformic Ret<sup>9</sup> (miRet<sup>9</sup>)* and *miRet<sup>51</sup>*. ES cell clones carrying the *miRet<sup>9</sup>* or *miRet<sup>51</sup>*

**Figure 1.** Targeting of the *c-Ret* locus. (A) Schematic representation of the targeting strategy. A cassette (*top*) consisting of cDNA fragments encoding the intracellular part of human RET9 or RET51 (*Ret\**), the  $\beta$ -globin polyadenylation signal (*pA*), and the Neo<sup>R</sup> gene (*Pgk-Neo*) was inserted in frame into exon 11 of the mouse locus, immediately after the segment encoding the transmembrane domain (red stripe, *middle*). The targeted locus is shown at the *bottom*. Solid arrowheads indicate loxP sites. (B) The targeted loci encode single chimeric RET receptors with identical extracellular (*Ec*) and transmembrane (*Tm*) domains and the intracellular (*Ic*) segment of human RET9 or RET51. The positions of Tyr-residues thought to be important for RET signaling are indicated by dots. (C) Southern blot analysis of DNA from ES cell clones using the probe shown in A (3' ext). On digestion with *Eco*RI, the wild-type and targeted loci generate 18-kb and 14-kb fragments, respectively.



alleles (Fig. 1C) were used to generate animals with various combinations of wild-type and *miRet* alleles.

Heterozygous (+/*miRet*<sup>9</sup> or +/*miRet*<sup>51</sup>), heteroallelic (*miRet*<sup>9</sup>/*miRet*<sup>51</sup>), and homozygous *miRet*<sup>9</sup> mice were viable and displayed no abnormalities, indicating that expression of the *miRet* alleles was not by itself detrimental. In contrast, the majority of *miRet*<sup>51</sup>/*miRet*<sup>51</sup> mice died as neonates and <5% survived to 2–3 mo of age, exhibiting severe growth retardation.

To examine the products of the *miRet* alleles, RNA from embryos or newborn brain was analyzed by RT-PCR using allele-specific primers. As expected, homozygous *miRet*<sup>9</sup> or *miRet*<sup>51</sup> mice expressed miRet9 or miRet51 mRNA, respectively, and lacked wild-type *c-Ret* transcripts, whereas +/*miRet*<sup>9</sup> and +/*miRet*<sup>51</sup> animals expressed both the wild-type and the appropriate chimeric isoforms (Fig. 2A). Semiquantitative RT-PCR showed that the levels of transcripts encoding RET9 and RET51 in monoisoformic embryos and brain were comparable with those of isoform specific transcripts in wild-type tissues (Fig. 2B; data not shown).

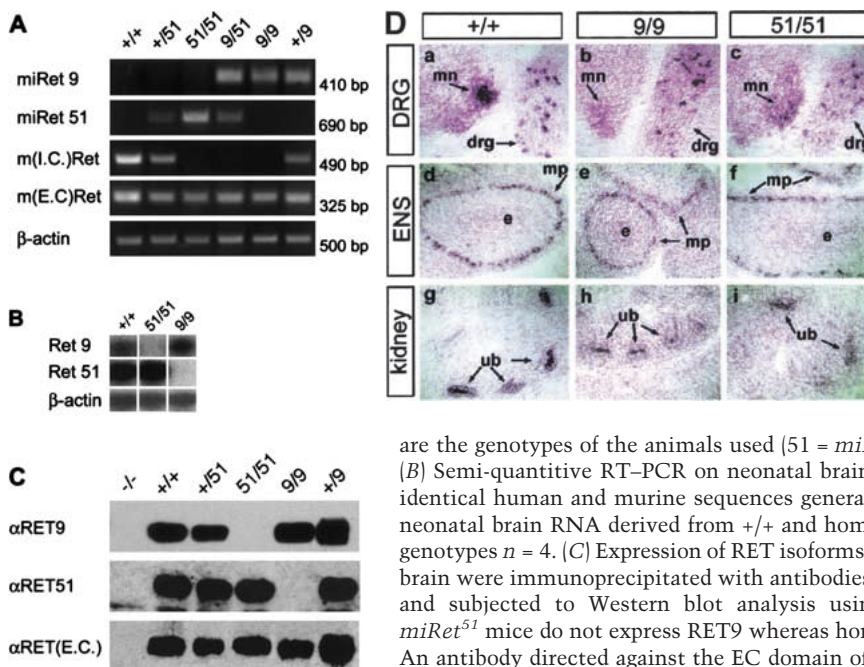
To further analyze the products of the *miRet* alleles, protein extracts from embryos and newborn brain were immunoprecipitated with antibodies specific for the extracellular domain of RET and the immunoprecipitates were characterized by Western blotting using isoform specific antibodies. Consistent with the RT-PCR analysis, animals carrying at least one wild-type *c-Ret* allele produced both RET9 and RET51 isoforms (Fig. 2C). In contrast, homozygous *miRet*<sup>9</sup> or *miRet*<sup>51</sup> animals generated RET9 or RET51 exclusively, respectively. Further-

more, the levels of RET9 and RET51 expressed in monoisoformic mice were comparable with those in wild-type animals (Fig. 2C). In summary, the novel *miRet* alleles we have generated encode single RET isoforms at levels comparable to those detected in wild-type animals.

To examine the tissue specificity of *miRet*<sup>9</sup> or *miRet*<sup>51</sup> expression during embryogenesis, we analyzed sections of monoisoformic mouse embryos by in situ hybridization using riboprobes corresponding to the extracellular domain of RET. As is the case for *c-Ret*, both *miRet*<sup>9</sup> and *miRet*<sup>51</sup> were expressed in the motoneurons of the spinal cord and in subsets of peripheral sensory neurons of the dorsal root ganglia (Fig. 2D, panels a–c), as well as in enteric (Fig. 2D, panels d–f), and autonomic ganglia (data not shown). Outside the nervous system, miRet9 and miRet51 mRNA were detected in the Wolffian duct (data not shown) and the branches of the UB throughout nephrogenesis (Fig. 2D, panels g–i). We conclude that *miRet*<sup>9</sup> and *miRet*<sup>51</sup> are expressed with a spatial and temporal pattern indistinguishable from that of wild-type *c-Ret*.

#### The role of RET9 and RET51 in the development of the excretory system

Given the critical role of RET signaling in nephrogenesis, we examined the excretory system of newborn *miRet* animals. Consistent with the normal appearance and healthy status of *miRet*<sup>9</sup> heterozygotes and homozygotes, their kidneys were morphologically and histo-



**Figure 2.** Expression of the *miRet* alleles. (A) RT-PCR analysis of total RNA from neonatal brain. Products specific for miRet51 or miRet9 mRNAs are generated only in samples derived from animals carrying at least one copy of *miRet*<sup>51</sup> or *miRet*<sup>9</sup>, respectively. Although no signal specific for sequences encoding the intracellular domain of mouse RET [m(I.C.)Ret] was detected in either homozygous or heteroallelic monoisoformic mice, sequences corresponding to the extracellular domain of mouse RET [m(E.C.)Ret] were amplified irrespective of the genotype. Amplification of  $\beta$ -actin mRNA verified the integrity of the mRNA. Shown at top

are the genotypes of the animals used (51 = *miRet*<sup>51</sup> and 9 = *miRet*<sup>9</sup>). For all genotypes  $n = 9$ . (B) Semi-quantitative RT-PCR on neonatal brain. Isoform-specific primers that correspond to identical human and murine sequences generate products of similar intensity in samples of neonatal brain RNA derived from +/+ and homozygous *miRet*<sup>51</sup> and *miRet*<sup>9</sup> animals. For all genotypes  $n = 4$ . (C) Expression of RET isoforms in *miRet* mice. Protein extracts from neonatal brain were immunoprecipitated with antibodies against the extracellular (EC) domain of RET and subjected to Western blot analysis using isoform-specific antibodies. Homozygous *miRet*<sup>51</sup> mice do not express RET9 whereas homozygous *miRet*<sup>9</sup> mice do not express RET51. An antibody directed against the EC domain of mouse RET [ $\alpha$ RET(E.C.)] detects comparable levels of the receptor in all samples. For all genotypes  $n = 6$ . (D) Cell-specific expression of RET isoforms in E13.5 embryos using a riboprobe that corresponds to the EC domain of mouse RET. Specific signal was present in motoneurons (mn), sensory neurons of dorsal root ganglia (drg), the ganglia of the myenteric plexus (mp), and the tips of the ureteric bud branches (ub). e, Endoderm of the gut. For all genotypes  $n = 12$ .

de Graaff et al.

logically normal (Fig. 3A,B, panels a and b). In contrast, the kidneys of *miRet*<sup>51</sup> homozygotes were always smaller (one-fourth to one-third the volume of +/+ controls), cystic and had fewer nephrons (Fig. 3A, panel c and B, panel d). In addition, the peripheral nephrogenic zone, in which UB branching and induction of new nephrons take place, was mostly absent. These abnormalities were similar to, although less severe than, those observed in newborn *Ret.k*- homozygotes (Schuchardt et al. 1994, 1996). No abnormalities were observed in the kidneys of +/*miRet*<sup>51</sup> or *miRet*<sup>9</sup>/*miRet*<sup>51</sup> animals (Fig. 3A, panels d,e and B, panel c).

To analyze the developmental basis of the renal abnormalities of *miRet*<sup>51</sup> homozygotes, we compared histological sections of +/+ and *miRet*<sup>51</sup>/*miRet*<sup>51</sup> embryos at various stages. Although the first stages of metanephric kidney development (i.e., evagination of the UB and its initial branching between E10.5 and E12.5), occurred normally in *miRet*<sup>51</sup> embryos (Fig. 3C, panels a,b), a reduction in the number of UB branches was first observed at E13.5 and more pronounced at later stages (Fig. 3C, panels c–f). This suggested that although signaling by RET51 is capable of supporting the early stages of UB outgrowth and branching, it is unable to sustain normal nephrogenesis at later stages of embryogenesis.

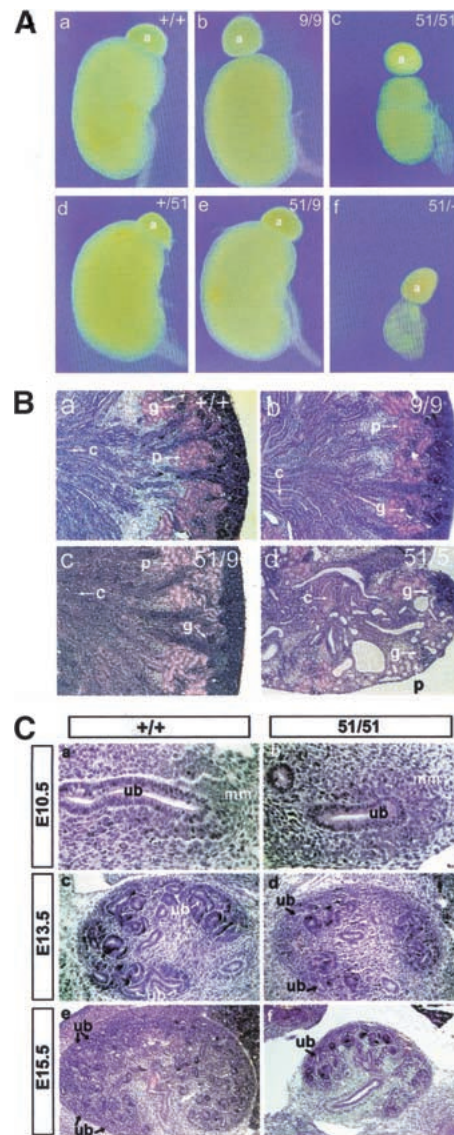
To address the possibility that the kidney defects observed in *miRet*<sup>51</sup> homozygotes were attributable to excessive RET signaling, we generated mice heteroallelic for *miRet*<sup>51</sup> and the null *Ret.k*- allele. The kidneys of these animals were even more severely affected than those of *miRet*<sup>51</sup> homozygotes (Fig. 3A, cf. panels c and f), suggesting that overactivation of the RET signaling pathway is not the cause of the renal abnormalities in *miRet*<sup>51</sup> animals.

#### Molecular analysis of kidney development in monoisoformic RET mice

We first analyzed the expression of *Gdnf*, *Gfra1*, and *Pax2* (Schedl and Hastie 2000) but we observed no differences in the expression of these genes between the metanephroi of wild-type and *miRet*<sup>51</sup>/*miRet*<sup>51</sup> embryos (Fig. 4, panels a–i).

The Wnt family of signaling molecules has an important role in renal development (Vainio et al. 1999; Schedl and Hastie 2000), and one of them, *Wnt11*, is normally coexpressed with *c-Ret* at the tips of the growing UB. We found that the levels of *Wnt11* mRNA were reduced in the metanephroi of *miRet*<sup>51</sup> homozygotes relative to *miRet*<sup>9</sup>/*miRet*<sup>9</sup> or +/+ embryos (Fig. 4j–l), indicating that the expression of *Wnt11* is differentially regulated by RET51 and RET9.

In addition to the UB and the nephrogenic mesenchyme, stromal cells also have an important role in kidney development (Bard 1996; Hatini et al. 1996; Batourina et al. 2001). Genes that are expressed specifically in stromal cells and are important for normal renal development include *BF-2*, which encodes a winged-helix transcription factor (Bard 1996; Hatini et al. 1996) and *Raldh2*, which is essential for retinoic acid (RA) synthe-



**Figure 3.** Renal hypodysplasia in *miRet*<sup>51</sup> animals. (A) Kidneys from *miRet*<sup>9</sup>/*miRet*<sup>9</sup> (b), +/*miRet*<sup>51</sup> (d) and *miRet*<sup>9</sup>/*miRet*<sup>51</sup> (e) animals appeared identical to the wild type (a). The kidneys of *miRet*<sup>51</sup>/*miRet*<sup>51</sup> (c) or *miRet*<sup>51</sup>/*Ret.k*- (f) animals, however, were hypodysplastic. +/+, *n* = 134; 9/9, *n* = 43; 51/51, *n* = 103; +/51, *n* = 197; 9/51, *n* = 35; 51/-, *n* = 17. a, Adrenal gland. (B) Histological analysis of kidneys of *miRet* neonates. The histology of *miRet*<sup>9</sup>/*miRet*<sup>9</sup> (b) and *miRet*<sup>9</sup>/*miRet*<sup>51</sup> (c) kidneys was indistinguishable from the wild type (a). However, the kidneys of *miRet*<sup>51</sup>/*miRet*<sup>51</sup> neonates (d) show many cysts and a reduced number of nephrons. g, Glomerulus; p, proximal convoluting tubule; c, collecting tubule. For all genotypes *n* = 7. (C) Reduced branching of the UB in *miRet*<sup>51</sup> homozygote embryos. Sections through the metanephros of E10.5 +/+ (a) and *miRet*<sup>51</sup> (b) embryos reveal no difference in the evagination of the UB (ub) and the invasion of the metanephric mesenchyme (mm). Note the normal condensation of the mesenchyme in the *miRet*<sup>51</sup> embryo. At E13.5, a small reduction in the number of UB branches is detected in the *miRet*<sup>51</sup> embryos (d) in comparison with +/+ (c). This difference becomes clearer at subsequent developmental stages (E15.5; cf. panels e and f). For all genotypes and stages *n* = 5.

sis (Niederreither et al. 1999). In E13.5  $+/+$  and  $miRet^{\rho}/miRet^{\rho}$  embryos, these two genes were co-expressed in a thin layer of peripheral stromal cells and in narrow strips of cells intercalated between the mesenchymal condensates and the UB tips (Fig. 4, panels m,n,p,q). In contrast, in E13.5  $miRet^{51}$  homozygotes, the layer of peripheral stromal cells expressing *BF-2* and *Raldh2* was significantly wider whereas the intercalated bands of positive stromal cells were reduced (Fig. 4o,r). Interestingly, this defect appears similar to that in kidneys from mice doubly mutant for RA receptors (RAR)  $\alpha$  and  $\beta$ 2. The renal defects in  $RAR\alpha\beta$ 2 mutant mice were attributed to an indirect effect of the mutation on the UB, which resulted in progressively reduced expression of RET (all isoforms), and could be rescued with a *Hoxb7-Ret9* transgene (Battourina et al. 2001).

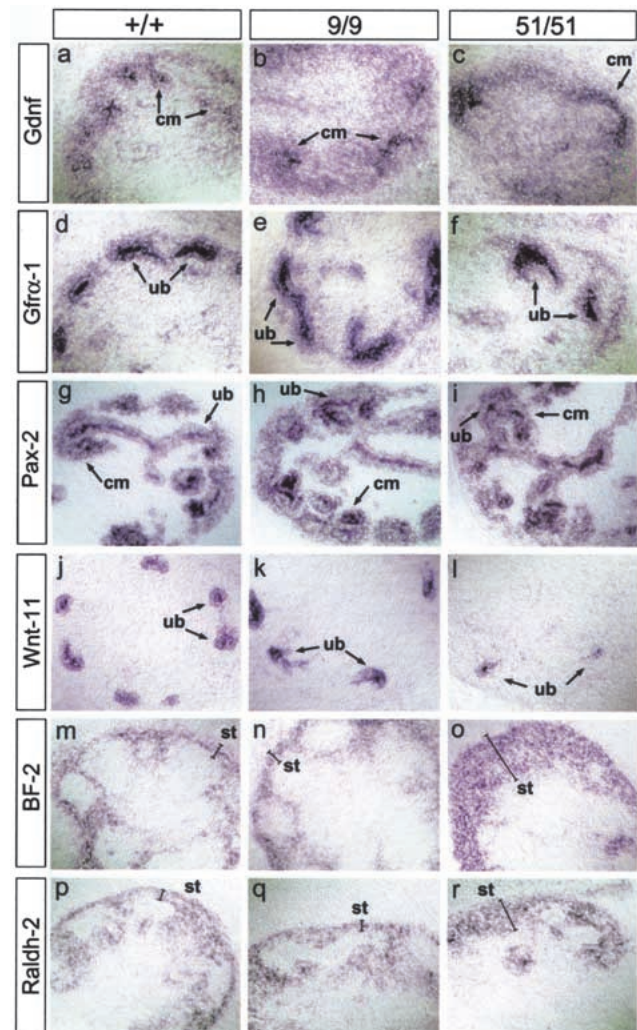
*A Hoxb7-Ret51 transgene, unlike Hoxb7-Ret9, does not rescue kidney development in Ret.k- mice*

We have described previously a series of transgenic (Tg) mice that expressed a *Ret9* cDNA under control of the *Hoxb7* promoter, which is active throughout the UB and its derivatives (the epithelium of the collecting system in the mature kidney; Srinivas et al. 1999). Consistent with the normal kidney development observed in  $miRet^{\rho}$  mice, the *Hoxb7-Ret9* transgene was capable of fully rescuing kidney development in *Ret.k-* homozygotes (Srinivas et al. 1999). To independently assess the role of RET51 in kidney development, a similar series of *Hoxb7-Ret51* Tg mice was generated. Three *Hoxb7-Ret51* Tg lines were found to express the transgene in adult kidneys by Northern and Western blotting, at a range of levels comparable with the *Hoxb7-Ret9* transgene (Fig. 5A; data not shown). When the *Hoxb7-Ret51* transgene was crossed into a *Ret.k-/Ret.k-* background, it had only a minimally beneficial effect on kidney development (Fig. 5B). As kidney size correlated inversely with the degree of dysplasia, the size distribution provides a rough quantitative measure of normal kidney development (Srinivas et al. 1999). *Hoxb7-Ret51* reduced the frequency of renal agenesis, and slightly increased the sizes of the kidneys formed, but never provided a full rescue. These experiments support the conclusion that RET51 cannot support normal kidney development in the absence of RET9.

On a wild-type background, the *Hoxb7-Ret9* transgene caused an incompletely penetrant, dominant renal hypodysplasia through an unknown mechanism thought to result from ectopic expression of RET9 throughout the UB (Srinivas et al. 1999). In contrast, no such dominant defect was seen in the *Hoxb7-Ret51* Tg mice, based on their normal distribution of kidney sizes (Fig. 5C) and histology (data not shown).

*Constitutively active forms of RET9, but not RET51, induce abnormal growth of the UB in transgenic mice*

We reported previously that an oncogenic form of RET9 (*Ret9-PTC2*), when expressed under the *Hoxb7* pro-

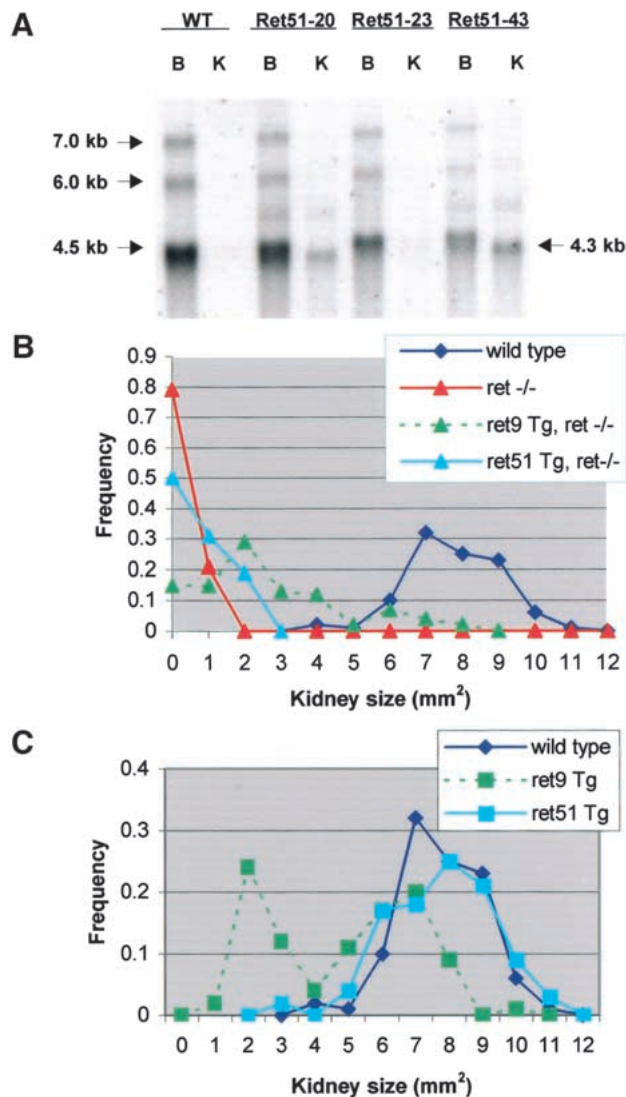


**Figure 4.** Molecular analysis of metanephric development in *miRet* mice. In situ hybridization with riboprobes for *Gdnf* (a–c); *Gfra1* (d–f); *Pax2* (g–i); *Wnt11* (j–l); *BF-2* (m–o); and *Raldh2* (p–r) on serial sections of E13.5 kidneys from  $+/+$  (a,d,g,j,m,p),  $miRet^{\rho}$  (b,e,h,k,n,q), and  $miRet^{51}$  (c,f,i,l,o,r) embryos. cm, Condensing mesenchyme; ub, ureteric bud tips; st, stroma. For each genotype and probe  $n = 4$ .

motor, induced the formation of abnormal renal nodules, apparently formed by deregulated growth and branching of the UB (Srinivas et al. 1999). To compare the effects of activated forms of RET9 and RET51, we constructed four more *Hoxb7-Ret* transgenes that expressed two additional oncogenic forms of each of the two RET isoforms. One pair of transgenes contained the C634K mutation found in the MEN2A syndrome (Donis-Keller et al. 1993; Mulligan et al. 1993). The second pair contained the M918T substitution associated with the related syndrome MEN2B (Carlson et al. 1994; Hofstra et al. 1994).

Because oncogenic *Hoxb7-Ret* transgenes may cause perinatal lethality (Srinivas et al. 1999), we sacrificed and examined the founder Tg mice at E18.5. Three out of five E18.5 *Hoxb7-Ret9-MEN2A* Tg founders had hyperplastic renal cysts, primarily in the nephrogenic zone (and

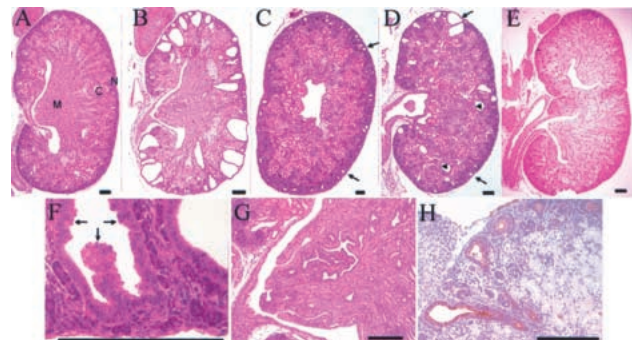
de Graaff et al.



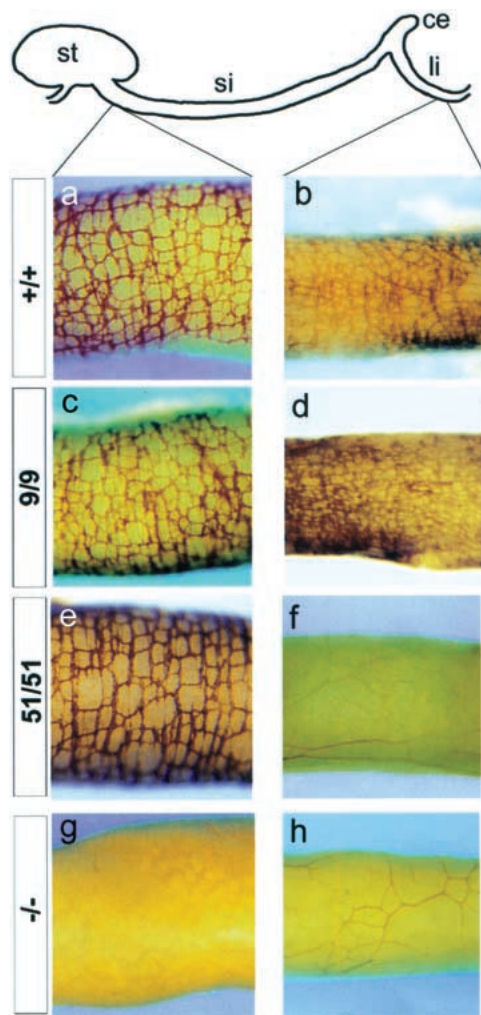
**Figure 5.** The Hoxb7–Ret51 transgene fails to rescue kidney development in *Ret.k*–/*Ret.k*– mice, or to induce dominant hypodysplasia in wild-type mice, unlike the Hoxb7–Ret9 transgene (Srinivas et al. 1999). (A) Northern blot of 5 µg of poly(A)<sup>+</sup> RNA from adult brain and kidney of wild-type and Hoxb7–Ret51 Tg mice, hybridized with a c-*Ret* probe. The major endogenous c-*Ret* transcripts are indicated at left. The transgene encodes a major 4.3-kb mRNA, and several larger bands of unknown origin. (WT) Wild-type control. RET51-20, RET51-23, and RET51-43 are three different Tg lines. Brain RNA (B) expresses endogenous c-*Ret* mRNA in wild-type and Tg mice, whereas kidney (K) does not express detectable c-*Ret* mRNA in wild-type adult, but expresses the Tg mRNA. (B,C) Kidney sizes in newborn mice (a value of zero indicates no kidney). (B) Although the Hoxb7–Ret9 transgene increased kidney size in *Ret.k*–/*Ret.k*– mice, occasionally into the normal range, the Hoxb7–Ret51 transgene had only a small beneficial effect. (C) On a wild-type background, although the Hoxb7–Ret9 transgene caused renal hypodysplasia in ~50% of Tg mice, Hoxb7–Ret51 had no significant effect. Data for *Ret.k*– homozygotes (*ret*–/–), *n* = 38; +/+, *n* = 138; Ret9 Tg, *ret*–/–, *n* = 92; Ret51 Tg, *ret*–/–, *n* = 32; Ret9 Tg, *n* = 76; Ret51 Tg, *n* = 128. Data were pooled for three Hoxb7–Ret51 and three Hoxb7–Ret9 Tg lines.

extending into the cortex), and abnormal branching of collecting ducts in the medulla (Fig. 6B,C,F–H). Similarly, two out of three E18.5 Hoxb7–Ret9–MEN2B Tg founders displayed cystic dilation of UB tips in the nephrogenic zone as well as cysts derived from more proximal maturing tubules in the cortex (Fig. 6D). Although the renal defects induced by Ret9–MEN2A and Ret9–MEN2B differed somewhat from those induced by Ret9–PTC2 (Srinivas et al. 1999), they also appeared to result from the abnormal growth, branching, and proliferation of the UB and collecting ducts.

In contrast, no abnormalities of this type were observed in the kidneys of Tg mice expressing either mutant form of RET51. None of five Hoxb7–Ret51–MEN2B or 11 Hoxb7–Ret51–MEN2A Tg founders showed any cortical cysts or abnormally branched medullary collecting ducts. Interestingly, two of the Hoxb7–Ret51–MEN2A mice had a unilateral bifid kidney, which contained a duplicated ureter, renal pelvis, and pyramid (Fig. 6E), but were otherwise normal in size and histology. Bifid kidneys, which sometimes occur in humans, are believed to result from the evagination of two UBs from the Wolffian duct (N'Guessan and Stephens 1983). This suggests that an activated form of RET51, although not perturbing the growth and branching of the UB within the developing kidney as do similar forms of RET9, can cause ectopic outgrowth of a secondary UB.



**Figure 6.** Constitutively active forms of RET9 but not RET51 induce cystic kidneys. Histological sections of kidneys from a +/+ mouse (A), two Hoxb7–Ret9–MEN2A Tg mice (B,C), a Hoxb7–Ret9–MEN2B mouse (D) and a Hoxb7–Ret51–MEN2A mouse, all at E18.5. The medulla (M), cortex (C), and nephrogenic zone (N) are labeled in A. (B) A severe case of cystic dysplasia caused by Hoxb7–Ret9–MEN2A. The cysts, which appear to derive from UB tips, replace much of the nephrogenic zone, and display hyperplastic features, such as stratification of the epithelium, as shown at higher magnification in F (arrows). In addition, the medulla displays abnormal branching of the collecting ducts, as shown at higher magnification in G. (C) A less severe case of cystic UB tips (arrows) in a different Hoxb7–Ret9–MEN2A founder. (H) A section for the same kidney as in C, stained with the UB-specific lectin dolichos biflorus (brown stain) (D'Agati and Trudel 1992), confirming the origin of the epithelial cysts. (D) a kidney from a Hoxb7–Ret9–MEN2B founder displays cystic UB tips in the peripheral nephrogenic zone (arrows) as well as cysts deeper in the cortex (arrowheads). (E) A bifid kidney from a Hoxb7–Ret51–MEN2A Tg mouse, with duplication of the ureter, pelvis, and renal pyramid.



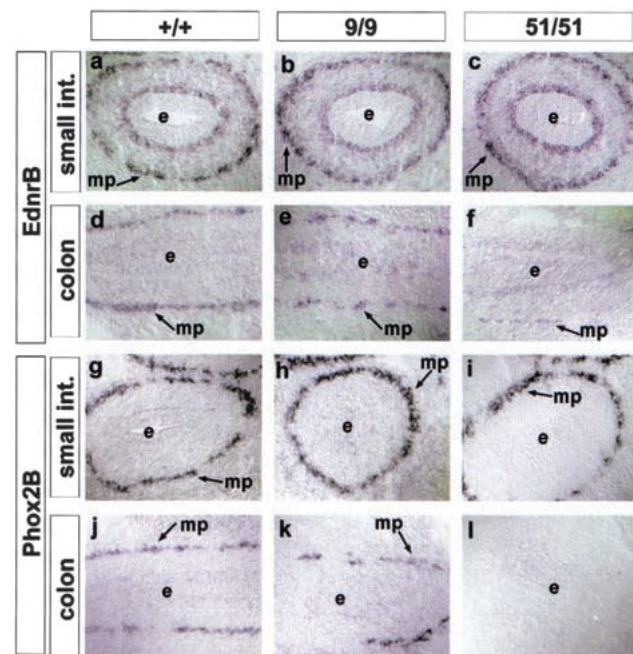
**Figure 7.** Aganglionosis of the colon of *miRet*<sup>51</sup>/*miRet*<sup>51</sup> animals. Whole-mount acetylcholinesterase histochemistry of neonatal duodenum (a,c,e,g) and colon (b,d,f,h) of +/+ (a,b), *miRet*<sup>0</sup>/*miRet*<sup>0</sup> (c,d), *miRet*<sup>51</sup>/*miRet*<sup>51</sup> (e,f) and *Ret.k*<sup>-</sup>/*Ret.k*<sup>-</sup> (g,h) neonates. In *miRet*<sup>51</sup> homozygotes, although an apparently normal myenteric plexus forms in the small intestine, no enteric ganglia were present in the colon. RET-deficient animals fail to develop the myenteric plexus throughout the length of the intestine. (Top) Diagrammatic representation of the various gut segments. st, Stomach; si, small intestine; ce, cecum; li, large intestine. +/+, n = 37; 9/9, n = 18; 51/51, n = 23; -/-, n = 3.

#### The role of RET9 and RET51 in the development of the ENS of the mouse

Genetic studies in humans and mice have shown that signaling by RET has a pivotal role in the development of the ENS (Taraviras and Pachnis 1999). To examine the individual roles of RET9 and RET51 during ENS histogenesis, we first analyzed the myenteric plexus of +/+, *miRet*<sup>0</sup>/*miRet*<sup>0</sup>, and *miRet*<sup>51</sup>/*miRet*<sup>51</sup> newborn mice by whole-mount acetylcholinesterase (AChE) histochemistry (Enomoto et al. 1998). We observed that the number of enteric ganglia and their distribution along the length

of the gut were indistinguishable between +/+ and *miRet*<sup>0</sup>/*miRet*<sup>0</sup> animals (Fig. 7a–d). Similarly, an apparently normal myenteric plexus was present in the stomach and small intestine of *miRet*<sup>51</sup> homozygotes (Fig. 7e). A severe reduction or complete absence of enteric ganglia, however, was reproducibly observed in the terminal three-fourths of the colon of *miRet*<sup>51</sup> mice at birth (Fig. 7f). Careful examination of gut preparations from these animals using molecular markers of enteric neurons, such as PGP9.5, indicated that a reduced number of neurons was also detectable in the proximal quarter of the colon (data not shown).

Aganglionosis in *miRet*<sup>51</sup> mice could result from failure of the ENS progenitors to enter the colon during embryogenesis or from their inability to survive within its microenvironment. As a first step toward distinguishing between these possibilities, we analyzed the distribution of ENC cells and their derivatives in the gut at E11.5–E13.5, a period during which ENS progenitors are colonizing the gut (Kapur et al. 1992; Durbec et al. 1996). In situ hybridization for markers of ENS progenitors, such as *EdnrB*, *Phox2b*, and *Gfra1* revealed no obvious differences in the number or distribution of ENC-derived cells throughout the gastrointestinal (GI) tract of +/+ and *miRet*<sup>0</sup> homozygotes (Fig. 8a,b,d,e,g,h,j,k). Similarly, normal numbers of *EdnrB*<sup>+</sup>, *Phox2B*<sup>+</sup>, and *Gfra1*<sup>+</sup> cells were present in the stomach and the small intestine of *miRet*<sup>51</sup> embryos (Fig. 8c,f,i). However, no ENC-derived cells were present in the distal three-fourths of the colon at any stage analyzed (Fig. 8f,l). These findings suggest



**Figure 8.** The progenitors of the ENS fail to colonize the colon of *miRet*<sup>51</sup>/*miRet*<sup>51</sup> embryos. In situ hybridization with riboprobes specific for *EdnrB* (a–f) and *Phox2B* (g–l) mRNAs in serial sections of small intestine (a–c,g–i) and colon (d–f,j–l) of +/+ (a,d,g,i), *miRet*<sup>0</sup>/*miRet*<sup>0</sup> (b,e,h,k) and *miRet*<sup>51</sup>/*miRet*<sup>51</sup> (c,f,j,l) E13.5 embryos. mp, Myenteric plexus. For all genotypes n = 3.

de Graaff et al.

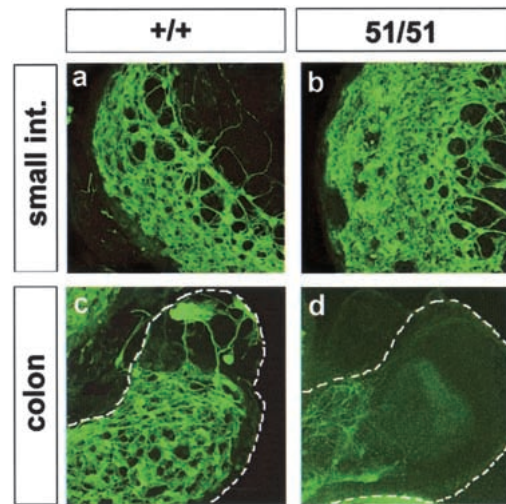
that the absence of enteric ganglia from *miRet*<sup>51</sup>/*miRet*<sup>51</sup> neonates results from failure of ENC cells to enter the colon during embryogenesis.

#### *The colonic aganglionosis of miRet*<sup>51</sup> mice is reproduced in vitro

We previously developed a fetal gut culture system in which key steps of ENS histogenesis, such as ENC cell migration and differentiation, can be reproduced in vitro (Natarajan et al. 1999). For this system, gut is isolated from E11.0–E11.5 mouse embryos (a stage at which NC cells have migrated up to the cecum) and cultured in defined medium. After 1–3 wk, ENC cells have colonized the entire gut and a large fraction have differentiated into neuronal and glial cells. Although the identity of the morphologically distinct segments of the gut (esophagus, stomach, small intestine, cecum, and large intestine) is maintained, the overall growth and elongation of the organ in culture is limited in comparison with its in vivo counterpart (Natarajan et al. 1999). To determine whether the colonic aganglionosis in *miRet*<sup>51</sup> homozygotes can be reproduced under conditions in which the gut undergoes limited growth, gut from *+/+* and *miRet*<sup>51</sup>/*miRet*<sup>51</sup> E11.0 embryos was cultured for 4 d. The extent of colonization by ENS progenitors and their differentiation was assayed by immunostaining with the neuronal markers PGP9.5 and TuJ1. A large number of PGP9.5 positive cells were detected throughout the length of the gut derived from *+/+* and *miRet*<sup>51</sup>/*miRet*<sup>51</sup> embryos (Fig. 9a,c). Similarly, an apparently normal complement of neurons was present in the stomach and small intestine of parallel cultures derived from *miRet*<sup>51</sup>/*miRet*<sup>51</sup> embryos (Fig. 9b). However, the colon of these explants was almost completely devoid of neurons and contained only PGP9.5-positive axonal projections that originated in the more proximal small intestine (Fig. 9d). These data indicate that the aganglionic phenotype of *miRet*<sup>51</sup> animals is reproduced in the fetal gut culture system.

#### Discussion

To gain insight into the differential roles of the RET isoforms, we generated monoisoformic mouse strains that express only RET9 or RET51 in place of the normal complement of *c-Ret* gene products. Our genetic and embryological studies demonstrated that signaling by RET9 is sufficient for normal embryonic development and postnatal life. In contrast, signaling by RET51, in the absence of RET9, resulted in characteristic defects in the development of the excretory and enteric nervous systems. Further evidence for differential signaling between the RET isoforms was provided by transgenic experiments in which RET9, but not RET51, when expressed in the UB and its branches was capable of rescuing the renal defects of *Ret.k*<sup>-</sup> mice (this study; Srinivas et al. 1999). These findings indicate that the RET isoforms have distinct signaling properties and suggest that



**Figure 9.** The colonic aganglionosis phenotype of *miRet*<sup>51</sup>/*miRet*<sup>51</sup> is reproduced in organ culture. Guts were removed from *+/+* and *miRet*<sup>51</sup>/*miRet*<sup>51</sup> (51/51) E11.5 embryos, cultured for 4 d and processed for immunostaining with PGP9.5. (a,b) Fluorescent images of segments of the small intestine of *+/+* and *miRet*<sup>51</sup>/*miRet*<sup>51</sup> gut. (c,d) The corresponding colonic segments. Note the absence of a ganglionic plexus from the colon of explants derived from *miRet*<sup>51</sup>/*miRet*<sup>51</sup> embryos. *+/+*, *n* = 5; 51/51, *n* = 4.

RET51 is dispensable during embryogenesis, whereas RET9 is necessary and sufficient for normal development of the ENS and the excretory system. We cannot exclude the possibility that the *miRet*<sup>51</sup> animals have subtle developmental abnormalities, however, an issue that is currently being addressed by further experimentation.

Several lines of evidence suggest that the phenotypic differences between the monoisoformic RET strains reflect different signaling properties of the two isoforms, rather than differences in the expression levels of the *miRet*<sup>9</sup> and *miRet*<sup>51</sup> alleles. First, we introduced equivalent cDNA fragments encoding intracellular portions of RET9 and RET51 into an identical position of the *c-Ret* locus, thereby generating alleles that were very similar in sequence and organization. Second, similar levels of expression of the isoform-specific transcripts were detected in RET9 and RET51 homozygous animals. Third, using the in vitro survival of DRG neurons (in the presence of GDNF or neurturin) as a measure of RET activation, we have observed that RET9 and RET51 can activate the anti-apoptotic machinery of peripheral sensory neurons to the same extent (E. de Graaf, S. Taraviras, and V. Pachnis, unpubl.). Finally, strong support for the differential signaling of RET9 and RET51 is derived from transgenic experiments showing that both ligand-dependent and -independent forms of the two RET isoforms have distinct biological effects on the developing kidney.

The phenotype of *miRet*<sup>51</sup> mice is similar, albeit milder, to that of RET-deficient animals. Therefore, whereas *Ret.k*<sup>-</sup> animals have either renal agenesis or very small and highly dysplastic kidney rudiments, all



*miRet*<sup>51</sup> mice develop somewhat larger kidney rudiments. The reproducible formation of a kidney rudiment is consistent with the observation that the first stages of metanephric development (i.e., evagination of the UB and its initial branching at E10.5–E12.5) are apparently normal in these animals, and that defects become evident only during later stages of nephrogenesis. Analysis of Hoxb7–Ret51 Tg mice also showed that although this transgene failed to support normal kidney development in *Ret.k*– homozygotes, it reduced the frequency of renal agenesis and slightly increased the kidney size, consistent with a limited role for RET51 in early UB growth. A milder phenotype was also observed in the ENS. Whereas *Ret.k*– animals have total intestinal aganglionosis, *miRet*<sup>51</sup> mice have an apparently normal complement of enteric ganglia up to the cecum, but fail to develop an ENS plexus in the post-cecal colon. The apparently normal status of heteroallelic (*miRet*<sup>9</sup>/*miRet*<sup>51</sup>) animals strongly suggests that the intracellular part of the chimeric receptors encoded by the *miRet* alleles are capable of normal signaling in mouse cells. Also, the failure to detect novel phenotypes in *miRet* animals (in comparison with *Ret.k*– homozygotes) indicates that these alleles do not interfere with other unrelated signaling pathways. Finally, the normal status of +/*miRet*<sup>9</sup> and +/*miRet*<sup>51</sup> mice indicates that neither *miRet*<sup>9</sup> nor *miRet*<sup>51</sup> have dominant-negative effects. Based on these findings, we suggest that *miRet*<sup>51</sup> represents a hypomorphic allele of *c-Ret*.

The failure of RET51 to support normal embryogenesis is surprising. In addition to all the Tyr-based signaling motifs it shares with RET9, RET51 contains two additional Tyr residues (corresponding to Tyr 1090 and Tyr 1096 of human RET). At least one of these (Tyr 1096) forms a docking site for the adaptor protein Grb2 (Liu et al. 1996; Besset et al. 2000), and biochemical studies have shown that it contributes to the activation of the PI3-K and the MAP-kinase signaling pathways (Besset et al. 2000). Nevertheless, our in vivo analysis implies that Tyr 1096, and the entire RET51-specific C-terminal tail, is not necessary for normal embryogenesis and that RET9 is sufficient for activation of all intracellular signaling pathways required for normal ENS histogenesis and renal development. In light of this, it is interesting to note that the RET9-specific C-terminal tail is identical in all vertebrates examined so far, whereas the RET51-specific sequence is less well conserved (Tahira et al. 1990; GenBank accession nos.: mouse, AF209436; rat, AJ299016 and AJ299017; chicken, Z49898; zebrafish, AF007949; pufferfish, U58673; and *Xenopus*, AF286643).

One hypothesis to explain the differential activities of RET9 and RET51 is that their specific carboxy-terminal tails may modulate the efficiency with which active signaling complexes are assembled. In this regard, it is interesting that the two isoforms diverge in sequence one amino acid after Tyr 1062, a residue essential for binding of the adaptor protein Shc and the assembly of signaling complexes (Besset et al. 2000; De Vita et al. 2000; Hayaishi et al. 2000). Because amino acids flanking Tyr-residues can determine the efficiency of active complex for-

mation and intracellular signaling, it is possible that in the context of RET51, Tyr 1062 forms a less efficient docking site. Consistent with this hypothesis, the C-terminal tail of RET isoforms can influence the strength of binding to the SH2 and PTB domains of Shc (Lorenzo et al. 1997; Ohiwa et al. 1997; Ishiguro et al. 1999). Furthermore, it is possible that other more distantly located amino acids of the RET51-specific tail have an inhibitory effect on active complex formation on the intracellular part of the receptor. Alternatively, the RET9-specific sequences that are absent from RET51 may have previously unrecognized signaling properties. Although at present we cannot distinguish between these possibilities, the strategy we have employed here allows us to test efficiently in vivo recombinant cDNAs encoding mutant forms of the intracellular segments of RET9 or RET51 isoforms.

The differential signaling by the RET isoforms could also result from the segregation of RET9 and RET51 in distinct cell membrane subdomains. This could influence their interactions with downstream signaling components, coreceptors or ligands. The availability of mouse strains that express single RET isoforms could facilitate studies on the differential localization of these molecules in polarized cells (such as the epithelial cells of the UB) or nonpolarized cells (such as the progenitors of the ENS).

#### *The role of RET isoforms in nephrogenesis*

In contrast to the frequent failure of UB outgrowth observed in *Ret.k*– mouse embryos (Schuchardt et al. 1996), the early stages of metanephric development, including the formation of the UB, invasion of the metanephric mesenchyme and the first branching events, occurred normally in *miRet*<sup>51</sup> homozygous embryos. As development progressed, however, the ability of the tips of the UB to branch further was reduced, and the kidney failed to achieve a normal size and histoarchitecture. Therefore, either RET51 or RET9 can transduce the signals for the outgrowth of the UB from the mesonephric duct and its initial branching, whereas only RET9 is able to support the further development of the metanephros. Although some studies have suggested that GDNF/RET signaling is required primarily for UB initiation (Sainio et al. 1997), our results indicate that RET signaling (specifically RET9) is also required for later stages of UB growth and branching. Our studies also provide the first suggestion that the intracellular signaling events downstream of RET during the early stages of UB outgrowth and branching may be distinct from those required at later stages of metanephric development.

These conclusions were supported further by the analysis of Hoxb7–Ret51 Tg embryos on a *Ret.k*–/*Ret.k*– background, where the transgene partially rescued the renal agenesis phenotype (reflecting a partial rescue of UB outgrowth) but failed to support normal kidney development. This contrasts with the complete rescue of kidney development achieved by a Hoxb7–Ret9 transgene (Srinivas et al. 1999). We also found that mutant,

de Graaff et al.

ligand-independent forms of the two RET isoforms had distinct dominant effects on the development of UB derivatives in the kidney. Hoxb7–Ret9 (but not Hoxb7–Ret51) containing the activating mutations associated with human MEN2A or MEN2B frequently caused cystic dilation, hyperplasia, or abnormal branching of UB-derived collecting duct epithelia. Therefore, constitutive RET9 but not RET51 signaling seems able to perturb the normal growth and development of the collecting ducts. In contrast, Hoxb7–Ret51–MEN2A mice sometimes displayed a duplicated ureter, which most likely results from evagination of duplicate UBs. This finding appears consistent with the specific ability of RET51 to support the early stages of UB growth.

#### *The role of RET isoforms in the development of ENS*

Although the ENS of *miRet*<sup>9</sup> mice appeared to develop normally, the gut of *miRet*<sup>51</sup> animals was only partially colonized by the progenitors of the ENS; enteric ganglia were present in the esophagus, stomach, and small intestine but were absent from the colon. The aganglionic phenotype of *miRet*<sup>51</sup> mice reproduces that of human congenital megacolon (HSCR), which is usually characterized by partial aganglionosis of the colon. In this respect, the *miRet*<sup>51</sup> strain provides an excellent model for studies on the pathogenetic mechanisms underlying this neural deficiency. For example, an unresolved issue in the field of HSCR is the state of the ENS in the apparently normoganglionic proximal segments of the gastrointestinal tract. This is of utmost medical importance, given that a large percentage of HSCR patients continue to have abnormal gut motility, even following a successful surgical resection of the aganglionic segment and anastomosis. Experiments are currently in progress to determine whether subtle histological defects of the ENS are present in the stomach and small intestine of *miRet*<sup>51</sup> animals.

With regard to the mechanisms of aganglionosis in *miRet*<sup>51</sup> homozygous mice, our studies suggest that it results from failure of ENS progenitors to colonize efficiently the post-cecal part of the gastrointestinal tract. Therefore, whereas a normal number of ENS progenitors is present in the stomach and small intestine at E13.5, a drastically reduced number of ENC cells is found in the post-cecal colon. But, why can loss-of-function *Ret* alleles (such as *miRet*<sup>51</sup> or other alleles described in patients with HSCR) support normal colonization of the gut by ENS progenitors up to the cecum, but not in the post-cecal gut? One possible explanation is based on the view that the developing gut is a relatively homogeneous structure whose colonization is determined mainly by the number of ENC cells, their migratory efficiency, and the rate of gut elongation. Taking into account the role of *c-Ret* in ENC cell proliferation and migration (Chalazonitis et al. 1998; Hearn et al. 1998; Heuckeroth et al. 1998; Taraviras et al. 1999; Young et al. 2001), one hypothesis is that, under normal circumstances, a constantly expanding population of migrating ENS progenitors is able to keep up with a rapidly growing organ and

fully colonize it. Therefore, any mutations that affect RET signaling would impair the proliferation and migration of ENC cells, resulting in incomplete colonization of the gut. In this case, the colon would be aganglionic only because it is the most distal, and therefore hardest to reach, segment of the gut. An alternative hypothesis is that the ENS progenitors, as they migrate along the gut, are exposed to region-specific microenvironmental signals that modify the ability of RET to control cell proliferation and migration. Therefore, in *miRet*<sup>51</sup> embryos, signaling by RET51 would be sufficient for normal development of the ENS up to the cecum, but not in the colon. According to this view, it is the unique properties of the cecum and the colon that are critical and not their distance from the entry point of the ENC cells in the foregut. The reproduction of the aganglionic phenotype of *miRet*<sup>51</sup> animals in an organ culture system, which allows the proliferation, migration, and differentiation of ENC cells but fails to support extensive growth of the gut, supports the second model. We suggest that local signals produced by the microenvironment of the colon further reduce the ability of the already weakened RET51 receptor to signal efficiently, resulting in failure of its colonization by ENC cells. The identification of such signals is a subject of our current experimentation.

#### **Materials and methods**

##### *Generation of targeted or transgenic mice expressing either RET9 or RET51*

A 10-kb *NdeI*–*XhoI* genomic fragment encompassing exons 6–11 of *c-Ret* was used as the backbone for the targeting construct (Fig. 1A). The genomic region encoding the extracellular domain of murine RET was fused in-frame to cDNA fragments encoding the intracellular segments of human RET9 or RET51 (which are 95% identical to the corresponding segments of the mouse isoforms), a human  $\beta$ -globin polyadenylation signal and a floxed PGK–*Neo*<sup>R</sup> fragment (Fig. 1A). Human cDNA sequences were used to allow us to distinguish the expression of the monoisoformic from the endogenous alleles. Targeted clones of E14.2 ES cells were identified by PCR using primers p1, 5′-gccttctatcgcttcttgacgag-3′ and p2, 5′-ctagcaggctcttgatgattctgg-3′ (Fig. 1A) and confirmed by Southern blot analysis (Fig. 1C; data not shown). Construction of the Hoxb7–Ret9 transgenes has been described previously (Srinivas et al. 1999). To generate the other transgenes, the RET9 cDNA was replaced by equivalent cDNAs encoding RET51, or human RET9 and RET51 bearing the MEN2A (Cys634Lys) and MEN2B (Met918Thr) mutations.

##### *Expression analysis*

Expression of endogenous *c-Ret* was assayed by RT–PCR using mouse-specific primers from exons 11 (5′-ggagaaccaggttctgtgactct-3′) and 14 (5′-ttctgccaagtactgcatacccctc-3′). For detection of both wild-type and targeted *c-Ret*, cDNA was amplified with two primers (forward, 5′-ctgtgtgatgcgctgtgccga-3′; reverse, 5′-gtcacaggaacctgttctccg-3′) in the part of exon 11 that was not affected by the targeting strategy. Specific expression of the targeted allele was determined using the p5 and p6 primers (Fig. 1A). Primers for actin (Stratagene) were used as an internal control.

For semiquantitative RT-PCR, we used primers specific for RET<sup>9</sup> or RET51 that are identical between human and mouse. The forward primer for both isoforms was in exon 19 (5'-gatgagaggccagacaactgca-3'); the Ret9 mRNA-specific primer was in exon 19 (5'-ctagaatctagtaaatgcatg-3'), whereas the Ret51 mRNA-specific primer was in exon 20 (5'-aggactctctccaggcagttctg-3').

For Western blots, neonatal brains and E13.5 embryos were homogenized in 2% NP40, 40 mM Tris-HCl (pH 7.5), 20 mM EDTA supplemented with protease inhibitors (Complete, Roche), incubated on ice for 30 min and centrifuged at 6500 rpm at 4°C. Immunoprecipitation and Western blotting was performed according to standard procedures using protein A sepharose beads. For immunoprecipitation, we used a cocktail of monoclonal antibodies directed against the extracellular domain of RET (Lo and Anderson 1995). Immunoblotting was done using polyclonal antibodies specific for RET9 (Immunobiological Laboratories), RET51 (Santa Cruz Biotechnologies) or the common extracellular domain (R&D Systems) at a 1/250 dilution.

Histological analysis and in situ hybridization was performed according to standard protocols. Organotypic culture of fetal gut was performed as described previously (Natarajan et al. 1999). Details of the knock-in constructs and strategy and our protocols are available on request.

## Acknowledgments

The authors are grateful to Amanda Hewett and Mary Dawson for animal husbandry. We thank David Anderson for the RET monoclonal antibodies and Jacqueline Deschamps for the Hoxb7 promoter. We are indebted to Richard Maas, Andy McMahon, Cathy Mendelsohn, Pierre Chambon, Michael Gershon, and Christo Goridis for probes. We also thank members of the Pachnis and Costantini laboratories and Iris Salecker, Bruce Ponder, and Aaron Cranston for valuable suggestions and discussions. EdG is a recipient of an EMBO fellowship (ALTF 260-1996). This work was supported by the Medical Research Council (UK), the National Institutes of Health, and the Cancer Research Campaign (UK).

The publication costs of this article were defrayed in part by payment of page charges. This article must therefore be hereby marked "advertisement" in accordance with 18 USC section 1734 solely to indicate this fact.

## References

- Airaksinen, M.S., Titievsky, A., and Saarma, M. 1999. GDNF family neurotrophic factor signaling: Four masters, one servant? *Mol. Cell. Neurosci.* **13**: 313–325.
- Baloh, R.H., Enomoto, H., Johnson, E.M. Jr., and Milbrandt, J. 2000. The GDNF family ligands and receptors—implications for neural development. *Curr. Opin. Neurobiol.* **10**: 103–110.
- Bard, J. 1996. A new role for the stromal cells in kidney development. *Bioessays* **18**: 705–707.
- Batourina, E., Gim, S., Bello, N., Shy, M., Clagett-Dame, M., Srinivas, S., Costantini, F., and Mendelsohn, C. 2001. Vitamin A controls epithelial/mesenchymal interactions through ret expression. *Nat. Genet.* **27**: 74–78.
- Besset, V., Scott, R.P., and Ibanez, C.F. 2000. Signaling complexes and protein-protein interactions involved in the activation of the ras and phosphatidylinositol 3-kinase pathways by the c-Ret receptor tyrosine kinase. *J. Biol. Chem.* **275**: 39159–39166.
- Carlson, K.M., Dou, S., Chi, D., Scavarda, N., Toshima, K., Jackson, C.E., Wells, Jr., S.A., Goodfellow, P.J., and Donis-Keller, H. 1994. Single missense mutation in the tyrosine kinase catalytic domain of the RET protooncogene is associated with multiple endocrine neoplasia type 2B. *Proc. Natl. Acad. Sci.* **91**: 1579–1583.
- Chalazonitis, A., Rothman, T.P., Chen, J., and Gershon, M.D. 1998. Age-dependent differences in the effects of GDNF and NT-3 on the development of neurons and glia from neural crest-derived precursors immunoselected from the fetal rat gut: Expression of GFR $\alpha$ -1 in vitro and in vivo. *Dev. Biol.* **204**: 385–406.
- D'Agati, V. and Trudel, M. 1992. Lectin characterization of cystogenesis in the SBM transgenic model of polycystic kidney disease. *J. Am. Soc. Nephrol.* **3**: 975–983.
- De Vita, G., Melillo, R.M., Carlomagno, F., Visconti, R., Castellone, M.D., Bellacosa, A., Billaud, M., Fusco, A., Tschlis, P.N., and Santoro, M. 2000. Tyrosine 1062 of RET-MEN2A mediates activation of Akt (protein kinase B) and mitogen-activated protein kinase pathways leading to PC12 cell survival. *Cancer Res.* **60**: 3727–3731.
- Donis-Keller, H., Dou, S., Chi, D., Carlson, K.M., Toshima, K., Lairmore, T.C., Howe, J.R., Moley, J.F., Goodfellow, P., and Wells, Jr., S.A. 1993. Mutations in the RET proto-oncogene are associated with MEN 2A and FMTC. *Hum. Mol. Genet.* **2**: 851–856.
- Durbec, P.L., Larsson-Blomberg, L.B., Schuchardt, A., Costantini, F., and Pachnis, V. 1996. Common origin and developmental dependence on c-ret of subsets of enteric and sympathetic neuroblasts. *Development* **122**: 349–358.
- Ehrenfels, C. W., Carmillo, P. J., Orozco, O., Cate, R. L., and Sanicola, M. 1999. Perturbation of RET signaling in the embryonic kidney. *Dev. Genet.* **24**: 263–272.
- Enomoto, H., Araki, T., Jackman, A., Heuckeroth, R.O., Snider, W.D., Johnson, Jr., E.M., and Milbrandt, J. 1998. GFR  $\alpha$ -1-deficient mice have deficits in the enteric nervous system and kidneys. *Neuron* **21**: 317–324.
- Hansford, J.R. and Mulligan, L.M. 2000. Multiple endocrine neoplasia type 2 and RET: From neoplasia to neurogenesis. *J. Med. Genet.* **37**: 817–827.
- Hatini, V., Huh, S.O., Herzlinger, D., Soares, V.C., and Lai, E. 1996. Essential role of stromal mesenchyme in kidney morphogenesis revealed by targeted disruption of Winged Helix transcription factor BF-2. *Genes & Dev.* **10**: 1467–1478.
- Hayashi, H., Ichihara, M., Iwashita, T., Murakami, H., Shimon, Y., Kawai, K., Kurokawa, K., Murakumo, Y., Imai, T., Funahashi, H., et al. 2000. Characterization of intracellular signals via tyrosine 1062 in RET activated by glial cell line-derived neurotrophic factor. *Oncogene* **19**: 4469–4475.
- Hearn, C.J., Murphy, M., and Newgreen, D. 1998. GDNF and ET-3 differentially modulate the numbers of avian enteric neural crest cells and enteric neurons in vitro. *Dev. Biol.* **197**: 93–105.
- Heuckeroth, R.O., Lampe, P.A., Johnson, E.M., and Milbrandt, J. 1998. Neurturin and GDNF promote proliferation and survival of enteric neuron and glial progenitors in vitro. *Dev. Biol.* **200**: 116–129.
- Hofstra, R.M., Landsvater, R.M., Ceccherini, I., Stulp, R.P., Stelwagen, T., Luo, Y., Pasini, B., Hoppener, J.W., van Amstel, H. K., Romeo, G., et al. 1994. A mutation in the RET protooncogene associated with multiple endocrine neoplasia type 2B and sporadic medullary thyroid carcinoma. *Nature* **367**: 375–376.
- Ishiguro, Y., Iwashita, T., Murakami, H., Asai, N., Iida, K., Goto, H., Hayakawa, T., and Takahashi, M. 1999. The role of amino acids surrounding tyrosine 1062 in ret in specific

de Graaff et al.

- binding of the shc phosphotyrosine-binding domain. *Endocrinology* **140**: 3992–3998.
- Jhiang, S.M. 2000. The RET proto-oncogene in human cancers. *Oncogene* **19**: 5590–5597.
- Kapur, R.P., Yost, C., and Palmiter, R.D. 1992. A transgenic model for studying development of the enteric nervous system in normal and aganglionic mice. *Development* **116**: 167–175.
- Le Douarin, N.M. and Teillet, M. A. 1973. The migration of neural crest cells to the wall of the digestive tract in avian embryo. *J. Embryol. Exp. Morphol.* **30**: 31–48.
- Liu, X., Vega, Q.C., Decker, R.A., Pandey, A., Worby, C.A., and Dixon, J.E. 1996. Oncogenic RET receptors display different autophosphorylation sites and substrate binding specificities. *J. Biol. Chem.* **271**: 5309–5312.
- Lo, L. and Anderson, D.J. 1995. Postmigratory neural crest cells expressing c-RET display restricted developmental and proliferative capacities. *Neuron* **15**: 527–539.
- Lorenzo, M.J., Gish, G.D., Houghton, C., Stonehouse, T.J., Pawson, T., Ponder, B.A., and Smith, D.P. 1997. RET alternate splicing influences the interaction of activated RET with the SH2 and PTB domains of Shc, and the SH2 domain of Grb2. *Oncogene* **14**: 763–771.
- Mulligan, L.M., Kwok, J.B., Healey, C.S., Elsdon, M.J., Eng, C., Gardner, E., Love, D.R., Mole, S.E., Moore, J.K., Papi, L., et al. 1993. Germ-line mutations of the RET proto-oncogene in multiple endocrine neoplasia type 2A. *Nature* **363**: 458–460.
- Natarajan, D., Grigoriou, M., Marcos-Gutierrez, C.V., Atkins, C., and Pachnis, V. 1999. Multipotential progenitors of the mammalian enteric nervous system capable of colonising aganglionic bowel in organ culture. *Development* **126**: 157–168.
- N'Guessan, G. and Stephens, F.D. 1983. Supernumerary kidney. *J. Urol.* **130**: 649–653.
- Niederreither, K., Subbarayan, V., Dolle, P., and Chambon, P. 1999. Embryonic retinoic acid synthesis is essential for early mouse post-implantation development. *Nat. Genet.* **21**: 444–448.
- Ohiwa, M., Murakami, H., Iwashita, T., Asai, N., Iwata, Y., Imai, T., Funahashi, H., Takagi, H., and Takahashi, M. 1997. Characterization of Ret–Shc–Grb2 complex induced by GDNF, MEN 2A, and MEN 2B mutations. *Biochem. Biophys. Res. Commun.* **237**: 747–751.
- Pachnis, V., Mankoo, B., and Costantini, F. 1993. Expression of the c-ret proto-oncogene during mouse embryogenesis. *Development* **119**: 1005–1017.
- Parisi, M.A., and Kapur, R.P. 2000. Genetics of Hirschsprung disease. *Curr. Opin. Pediatr.* **12**: 610–617.
- Sarma, M. 2000. GDNF—A stranger in the TGF- $\beta$  superfamily? *Eur. J. Biochem.* **267**: 6968–6971.
- Sainio, K., Suvanto, P., Davies, J., Wartiovaara, J., Wartiovaara, K., Sarma, M., Arumae, U., Meng, X., Lindahl, M., Pachnis, V., et al. 1997. Glial-cell-line-derived neurotrophic factor is required for bud initiation from ureteric epithelium. *Development* **124**: 4077–4087.
- Saxen, L. 1987. *Organogenesis of the kidney*. Cambridge University Press, Cambridge, UK.
- Schedl, A. and Hastie, N.D. 2000. Cross-talk in kidney development. *Curr. Opin. Genet. Dev.* **10**: 543–549.
- Schuchardt, A., D'Agati, V., Larsson-Blomberg, L., Costantini, F., and Pachnis, V. 1994. Defects in the kidney and enteric nervous system of mice lacking the tyrosine kinase receptor Ret. *Nature* **367**: 380–383.
- Schuchardt, A., D'Agati, V., Pachnis, V., and Costantini, F. 1996. Renal agenesis and hypodysplasia in ret-k-mutant mice result from defects in ureteric bud development. *Development* **122**: 1919–1929.
- Smith-Hicks, C.L., Sizer, K.C., Powers, J.F., Tischler, A.S., and Costantini, F. 2000. C-cell hyperplasia, pheochromocytoma and sympathoadrenal malformation in a mouse model of multiple endocrine neoplasia type 2B. *EMBO J.* **19**: 612–622.
- Srinivas, S., Wu, Z., Chen, C.M., D'Agati, V., and Costantini, F. 1999. Dominant effects of RET receptor misexpression and ligand-independent RET signaling on ureteric bud development. *Development* **126**: 1375–1386.
- Tahira, T., Ishizaka, Y., Itoh, F., Sugimura, T., and Nagao, M. 1990. Characterization of ret proto-oncogene mRNAs encoding two isoforms of the protein product in a human neuroblastoma cell line. *Oncogene* **5**: 97–102.
- Takahashi, M., Ritz, J., and Cooper, G.M. 1985. Activation of a novel human transforming gene, ret, by DNA rearrangement. *Cell* **42**: 581–588.
- Takahashi, M., Buma, Y., Iwamoto, T., Inaguma, Y., Ikeda, H., and Hiai, H. 1988. Cloning and expression of the ret proto-oncogene encoding a tyrosine kinase with two potential transmembrane domains. *Oncogene* **3**: 571–578.
- Taraviras, S. and Pachnis, V. 1999. Development of the mammalian enteric nervous system. *Curr. Opin. Genet. Dev.* **9**: 321–327.
- Taraviras, S., Marcos-Gutierrez, C.V., Durbec, P., Jani, H., Grigoriou, M., Sukumaran, M., Wang, L.C., Hynes, M., Raisman, G., and Pachnis, V. 1999. Signalling by the RET receptor tyrosine kinase and its role in the development of the mammalian enteric nervous system. *Development* **126**: 2785–2797.
- Tsuzuki, T., Takahashi, M., Asai, N., Iwashita, T., Matsuyama, M., and Asai, J. 1995. Spatial and temporal expression of the ret proto-oncogene product in embryonic, infant and adult rat tissues. *Oncogene* **10**: 191–198.
- Vainio, S.J., Itaranta, P.V., Perasaari, J.P., and Uusitalo, M.S. 1999. Wnts as kidney tubule inducing factors. *Int. J. Dev. Biol.* **43**: 419–423.
- van Weering, D.H. and Bos, J.L. 1998. Signal transduction by the receptor tyrosine kinase Ret. *Recent Results Cancer Res.* **154**: 271–281.
- Yntema, C.L. and Hammond, W.S. 1954. The origin of intrinsic ganglia of trunk viscera from vagal neural crest in the chick embryo. *J. Comp. Neurol.* **101**: 515–542.
- Young, H.M., Hearn, C.J., Farlie, P.G., Canty, A.J., Thomas, P.Q., and Newgreen, D.F. 2001. GDNF is a chemoattractant for enteric neural cells. *Dev. Biol.* **229**: 503–516.



## Differential activities of the RET tyrosine kinase receptor isoforms during mammalian embryogenesis

Esther de Graaff, Shankar Srinivas, Carol Kilkenny, et al.

*Genes Dev.* 2001, **15**:

Access the most recent version at doi:[10.1101/gad.205001](https://doi.org/10.1101/gad.205001)

---

### References

This article cites 49 articles, 16 of which can be accessed free at:  
<http://genesdev.cshlp.org/content/15/18/2433.full.html#ref-list-1>

### License

### Email Alerting Service

Receive free email alerts when new articles cite this article - sign up in the box at the top right corner of the article or [click here](#).

---

An advertisement banner with a dark background. On the left, it says 'Dharmacon™ Reagents' with the tagline 'Custom synthesis, RNAi, and CRISPR solutions' below it. In the center, the words 'Infinite Reliability' are written in large white font. To the right of this text is a 'More' button. On the far right, the 'horizon' logo is displayed in white, with 'a PerkinElmer company' written in smaller text below it. The background features a colorful, abstract graphic of intertwined DNA strands in shades of purple, blue, and green.