



Differential roles of NHERF1, NHERF2, and PDZK1 in regulating CFTR-mediated intestinal anion secretion in mice

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The epithelial anion channel CFTR interacts with multiple PDZ domain-containing proteins. Heterologous expression studies have demonstrated that the Na⁺/H⁺ exchanger regulatory factors, NHERF1, NHERF2, and PDZK1 (NHERF3), modulate CFTR membrane retention, conductivity, and interactions with other transporters. To study their biological roles in vivo, we investigated CFTR-dependent duodenal HCO₃⁻ secretion in mouse models of *Nherf1*, *Nherf2*, and *Pdzk1* loss of function. We found that *Nherf1* ablation strongly reduced basal as well as forskolin-stimulated (FSK-stimulated) HCO₃⁻ secretory rates and blocked β₂-adrenergic receptor (β₂-AR) stimulation. Conversely, *Nherf2*^{-/-} mice displayed augmented FSK-stimulated HCO₃⁻ secretion. Furthermore, although lysophosphatidic acid (LPA) inhibited FSK-stimulated HCO₃⁻ secretion in WT mice, this effect was lost in *Nherf2*^{-/-} mice. *Pdzk1* ablation reduced basal, but not FSK-stimulated, HCO₃⁻ secretion. In addition, laser microdissection and quantitative PCR revealed that the β₂-AR and the type 2 LPA receptor were expressed together with CFTR in duodenal crypts and that colocalization of the β₂-AR and CFTR was reduced in the *Nherf1*^{-/-} mice. These data suggest that the NHERF proteins differentially modulate duodenal HCO₃⁻ secretion: while NHERF1 is an obligatory linker for β₂-AR stimulation of CFTR, NHERF2 confers inhibitory signals by coupling the LPA receptor to CFTR.

Introduction

The epithelial anion channel CFTR has been recognized as a hub protein, connecting to many other proteins and thus able to form protein-protein networks and affecting an astonishingly large number of cellular functions in addition to its ability to transport Cl⁻ and HCO₃⁻ (1–4). Important for this property as a connector hub is its C-terminal postsynaptic density protein PSD95-*Drosophila* homolog discs-large and tight junction protein ZO-1-binding (PDZ-binding) motif, which enables CFTR to bind to the PDZ domains of a variety of PDZ adapter proteins, including Na⁺/H⁺ exchanger regulatory factor 1 (NHERF1; also known as NHERF, EBP50, or SLC9A3R1), NHE3 kinase A regulatory protein (NHERF2; also known as E3KARP or SLC9A3R2), and PDZ domain-containing protein in kidney 1 (PDZK1; also known as CAP70 or NHERF3) of the NHERF family of PDZ adapters (2, 3). Heterologous expression studies have provided evidence for the importance of these interactions for CFTR trafficking and membrane retention (5–7), dimerization (8, 9), membrane mobility (10, 11), interaction with other transporters (12–14), and, perhaps most interestingly, the forma-

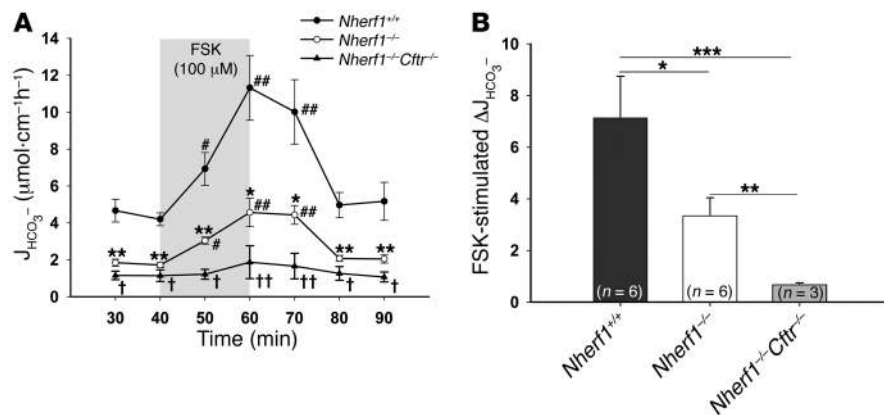
tion of multiprotein signaling complexes in which receptor-mediated signals are directly conveyed to the transporter (15, 16). Highly exciting to basic scientists as well as physicians treating CF patients were the recent reports that in a cell line expressing a CFTR trafficking mutant (ΔF508), either overexpression of NHERF1 (17) or siRNA-mediated knockdown of CAL (18), a Golgi-located PDZ protein that targets CFTR to the lysosomal pathway, resulted in redistribution of the mutated CFTR to the plasma membrane and an increased Cl⁻ efflux. These and other findings make NHERF- and other PDZ domain-mediated protein-protein interactions both a desirable drug target and a likely point of origin for pathophysiological changes (19, 20). On the other hand, the high sequence homology among the NHERF proteins (21–25), their overlapping tissue expression (26–28), a seemingly similar effect on CFTR function in heterologous expression systems (3), and the relative health of NHERF-deficient mice (12, 29, 30) have raised questions about redundancy and the importance of these proteins' role in native tissue and in the living organism.

Broere et al. have recently reported a decreased cAMP-activated short circuit current (I_{sc}) and HCO₃⁻ secretory rate in *Nherf1*^{-/-} mouse isolated small intestinal mucosa and a less intense CFTR staining pattern in the brush border membrane (BBM) of jejunal crypt cells, whereas no change in these parameters was seen in *Nherf2*^{-/-} mouse small intestine (31). Hillesheim et al. observed a mild decrease in forskolin-stimulated (FSK-stimulated) I_{sc} and HCO₃⁻ secretion in *Pdzk1*^{-/-} mouse small intestine (32). To our knowledge, no in vivo studies have previously been performed in these mice.

Conflict of interest: The authors have declared that no conflict of interest exists.

Nonstandard abbreviations used: β₂-AR, β₂-adrenergic receptor; BBM, brush border membrane; FSK, forskolin; I_{sc}, short circuit current; LPA, lysophosphatidic acid; LPA₂R, type 2 LPA receptor; NHERF1, Na⁺/H⁺ exchanger regulatory factor 1; NHERF2, NHE3 kinase A regulatory protein; PDZ, postsynaptic density protein PSD95-*Drosophila* homolog discs-large and tight junction protein ZO-1; PDZK1, PDZ domain-containing protein in kidney 1.

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**Figure 1**

Basal and FSK-stimulated HCO₃⁻ secretion in *Nherf1*^{-/-}, *Nherf1*^{-/-}*Cftr*^{-/-}, and WT murine duodenum in vivo. (A) Duodenal HCO₃⁻ secretion was measured in anesthetized *Nherf1*^{-/-}, *Nherf1*^{-/-}*Cftr*^{-/-}, and WT littermates in the basal state, and during and after 20 min luminal perfusion with 10⁻⁴ M FSK. Basal as well as FSK-stimulated duodenal HCO₃⁻ secretion was dramatically reduced in the *Nherf1*^{-/-} and *Nherf1*^{-/-}*Cftr*^{-/-} mice compared with WT littermates (*n* = 6 pairs of KO and WT mice, *P* < 0.001). The 20-min application time is denoted by shading. **P* < 0.01, ***P* < 0.001, ****P* < 0.0001 versus WT; #*P* < 0.01, ##*P* < 0.001 versus basal value; †*P* < 0.01, ††*P* < 0.001 versus *Nherf1*^{-/-}. (B) The net peak FSK-stimulated HCO₃⁻ secretory rate was significantly lower in *Nherf1*^{-/-} and *Nherf1*^{-/-}*Cftr*^{-/-} mice than in WT littermates. The net peak was calculated for each experiment in this and subsequent experiments by taking the peak value and subtracting the average of the 2 basal values before the application of 10⁻⁴ M FSK. **P* < 0.01; ***P* < 0.001; ****P* < 0.0001.

CFTR is highly expressed in murine duodenum and is essential for the HCO₃⁻ secretion in this tissue (33–36). Measuring duodenal HCO₃⁻ secretion is one of the few parameters to assess CFTR function quantitatively in a living organism. To learn more about the biological significance of the NHERF proteins in CFTR regulation, we measured basal and FSK-stimulated duodenal HCO₃⁻ secretion in anesthetized mice lacking one or more of the NHERF proteins. The importance of NHERF1 in mediating receptor-dependent activation and of NHERF2 in receptor-dependent inhibition of CFTR was studied, and colocalization of the receptors, the NHERF proteins, and CFTR was demonstrated in the apical membranes of crypt duodenocytes. In this way, for the first time to our knowledge, the concept of G protein-coupled receptor signaling to CFTR via a specific NHERF protein was validated in the living animal.

Results

Reduced basal and FSK-stimulated duodenal HCO₃⁻ secretory rates in *Nherf1*^{-/-} mice in vivo. The basal HCO₃⁻ secretory rate in the duodenum of anesthetized *Nherf1*^{-/-} as well as *Nherf1*^{-/-}*Cftr*^{-/-} double-deficient mice was dramatically reduced compared with WT littermates, and the response to a maximally stimulatory concentration of FSK was strongly reduced, but still significant, in the *Nherf1*^{-/-} duodenum and virtually abolished in the *Nherf1*^{-/-}*Cftr*^{-/-} mice (Figure 1, A and B). This suggests that the presence of NHERF1 is essential for maximal stimulation of CFTR by an increase in intracellular cAMP in the murine duodenum. To investigate whether the NHERF1-dependent reduction of duodenal HCO₃⁻ secretion was caused by reduced CFTR activation, we next studied *Nherf1*^{-/-}*Cftr*^{-/-} mice, which displayed no significant stimulation of duodenal HCO₃⁻ secretion upon FSK treatment (Figure 1, A and B). These results were similar to those in *Cftr*^{-/-} mice (data not shown).

NHERF1 is essential for β₂-adrenergic stimulation of duodenal HCO₃⁻ secretion.

One of the most astonishing results of the previously described experiments was the strong reduction in the basal HCO₃⁻ secretory rate in the absence of NHERF1. These changes were not observed in isolated epithelium (31). We therefore investigated whether a tonic stimulation of CFTR that only occurs in vivo might explain these effects, possibly via NHERF1-mediated formation of a CFTR-receptor signaling complex. Of course, multiple possibilities for such endogenous stimulators are feasible. The only NHERF1-mediated CFTR-receptor interaction with a stimulatory action that has been studied in molecular detail in a cell line is the formation of a complex containing the β₂-adrenergic receptor (β₂-AR, encoded by *Adrb2*), NHERF1, PKA, and CFTR (16). Whether β₂-ARs are present in intestinal epithelium and whether their activation stimulates duodenal HCO₃⁻ secretion is presently unknown. We therefore amplified and sequenced an *Adrb2* transcript from isolated murine

duodenal epithelium. Interestingly, the mRNA expression of *Adrb2* was significantly upregulated in the duodenum, but not in the lungs, of *Nherf1*^{-/-} mice (Figure 2A).

We next investigated whether the β₂-AR is colocalized with CFTR and NHERF1 in the apical membrane of duodenal enterocytes. To this end, we isolated BBM from murine duodenum and performed Western blot analysis with antibodies against CFTR, β₂-AR (Figure 2B), and NHERF1 (data not shown), as has been published before (31). A band in the appropriate size for CFTR, β₂-AR, and NHERF1 was clearly enriched in the respective BBM fractions. The specificity of the antibody was checked either by blocking peptide (for β₂-AR) or by using gene-deficient mice (for CFTR).

Immunohistochemistry was used next to confirm these results and to more precisely define the colocalization of CFTR and the β₂-AR. Anti-CFTR antibody localized CFTR to the apical membranes in the duodenocytes predominantly in the cryptal region and Brunner's glands, as described previously (37). Anti-β₂-AR antibody localized the β₂-AR to the apical membrane of the villi as well as crypt duodenocytes and the epithelial lining of the Brunner's glands, and there was staining in the muscle layer as well. The apical localization of NHERF1 in the crypt and villous region has been published previously (31) and was identical in the murine duodenum (data not shown). The yellow staining in Figure 3B indicates colocalization of CFTR and β₂-AR in the WT duodenum. In the absence of *Nherf1*, the staining appeared more diffuse, although a substantial part of both proteins was still localized in the BBM.

To test whether one component of the basal HCO₃⁻ secretion is NHERF1 dependent, we applied the specific β₂-AR antagonist ICI-118551. In WT mice, but not in *Nherf1*^{-/-} mice, 100 μM ICI-118551 significantly inhibited basal HCO₃⁻ secretion (Figure 4A). This is indicative of an involvement of β₂-AR activation in the basal HCO₃⁻ secretory tone in anesthetized mice and suggests that this

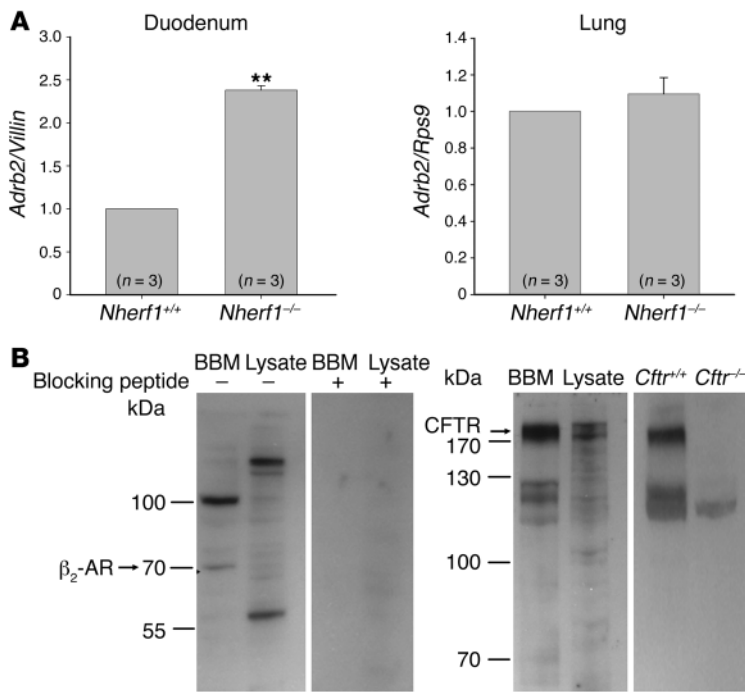


Figure 2

Duodenal epithelial *Adrb2* mRNA expression and apical membrane localization of CFTR and the β_2 -AR. **(A)** mRNA encoding the β_2 -AR was expressed in murine duodenal epithelium and was significantly higher in the duodenum of *Nherf1*^{-/-} than in WT mice. Conversely, no difference in *Adrb2* mRNA expression level was observed in the lungs of *Nherf1*^{-/-} mice. Experiments were performed in triplicate for each RNA extract from 3 pairs of mice. ***P* < 0.01 versus WT. **(B)** Western blot analysis of CFTR and the β_2 -AR in the BBM fraction and total cell lysate from duodenal epithelial scrapings showed that both proteins were enriched in the BBM. Both bands were blocked in the β_2 -AR Western blot using the blocking peptide, showing the specificity of the antibody. The higher-molecular weight band is likely a heterodimeric form of the receptor, as has been reported (62). The specificity of the CFTR band was checked using protein lysate and BBM preparation from *Cftr*^{-/-} mice. The expected molecular weight of the protein is indicated.

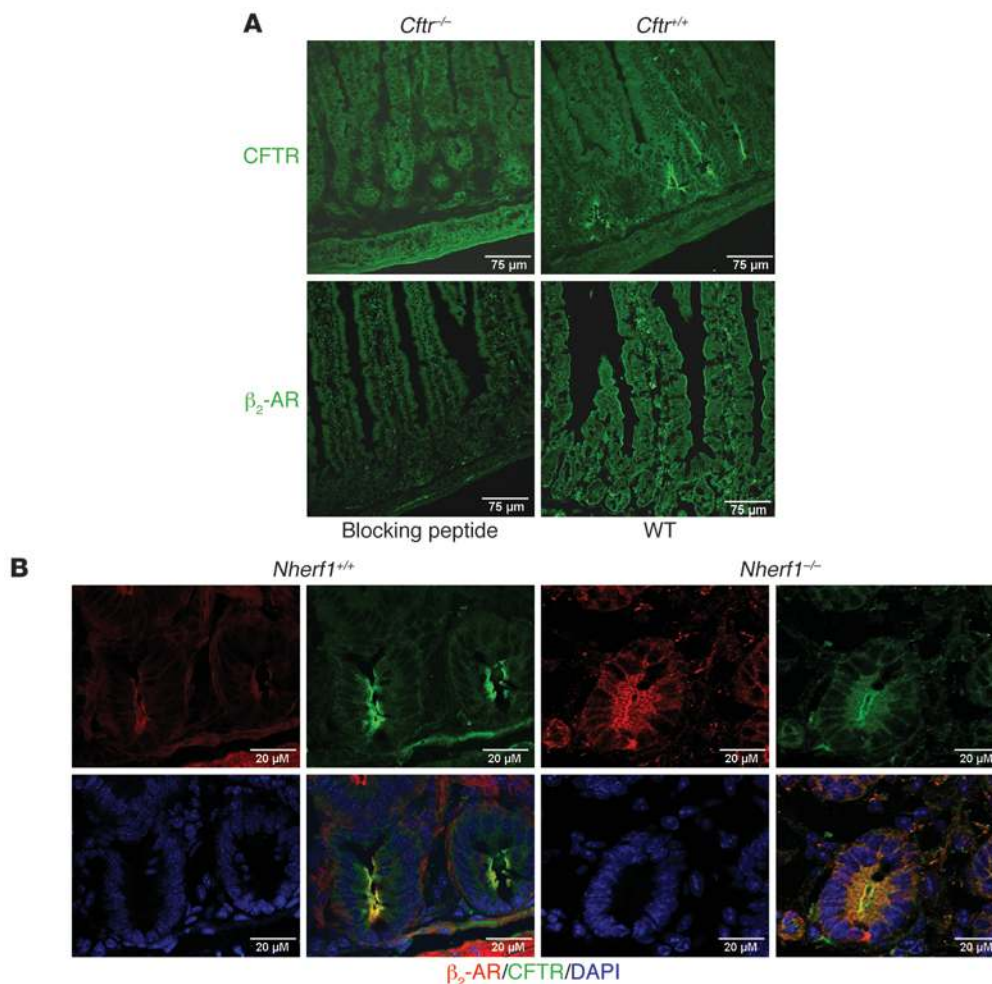
stimulation is NHERF1 dependent. To further study this hypothesis, we applied the selective β_2 -AR agonist clenbuterol to the duodenal lumen of *Nherf1*^{-/-} and WT mice and found that 50 μ M clenbuterol in the luminal perfusate caused a significant stimulation of duodenal HCO₃⁻ secretion in the WT mice that was completely absent in the *Nherf1*^{-/-} mice (Figure 4B). The total lack of response to clenbuterol in the absence of NHERF1 demonstrates the necessity of this interaction for the β_2 -AR-mediated stimulation of CFTR and strengthens the concept of NHERF1-mediated complex formation between the β_2 -AR and CFTR. To test whether this effect is specific for *Nherf1* ablation, we repeated the experiments in *Nherf2*^{-/-} and *Pdzk1*^{-/-} mice. Basal HCO₃⁻ secretory rates were different in the *Nherf2* and *Pdzk1* WT and KO mice, as discussed below, but the effects of both ICI-118551 and clenbuterol were preserved (Figure 4C).

*FSK-stimulated duodenal HCO₃⁻ secretion is increased in *Nherf2*^{-/-} mice.* *Nherf2*^{-/-} mice did not display significant differences in basal HCO₃⁻ secretion compared with WT controls, but had a significantly stronger secretory response to 10⁻⁴ M luminal FSK (Figure 5, A and B). This was highly surprising, because it was not previously observed in isolated duodenal mucosa (31), and any potential effect of NHERF2 on FSK-mediated HCO₃⁻ secretion was expected to be stimulatory instead of inhibitory based on experiments in airway cell lines (38) as well as in vitro studies (39). However, the results clearly suggested an inhibitory effect of NHERF2 on intestinal CFTR activity. We then asked 2 questions: (a) Can the reduced in vivo CFTR activity in *Nherf1*^{-/-} duodenum be rescued by additional KO of *Nherf2*?; and (b) Can we find evidence for NHERF2-mediated inhibitory receptor-CFTR signaling complex formation in murine duodenum in vivo?

To answer the first question, *Nherf1*^{-/-}, *Nherf2*^{-/-}, *Nherf1*^{+/+} *Nherf2*^{+/+}, and *Nherf1*^{-/-} *Nherf2*^{-/-} mice were bred from the same founders, and the basal as well as FSK-stimulated HCO₃⁻ secretory rates were measured in all 3 KO mouse lines as well as the

respective WT mice (Figure 5, C and D). The additional deletion of *Nherf2* significantly increased the basal HCO₃⁻ secretory rate observed in *Nherf1*^{-/-} mice (Figure 5C), suggesting that the inhibitory NHERF2-mediated event did not require NHERF1 and that it was active under basal conditions. The FSK-stimulated maximal secretory rate, on the other hand, was only mildly increased in *Nherf1*^{-/-} *Nherf2*^{-/-} mice compared with *Nherf1*^{-/-} mice, demonstrating the absolute requirement of NHERF1 for maximal duodenal CFTR activation in vivo (Figure 5D). The experiments also demonstrate that NHERF1 and NHERF2 are likely to act independently from each other on CFTR.

NHERF2 is essential for lysophosphatidic acid inhibition of duodenal HCO₃⁻ secretion. To answer the second question, we searched for potential inhibitory effects of NHERF2 in the literature. A recent report provided evidence for an inhibitory effect of lysophosphatidic acid (LPA) on ³⁶Cl⁻ efflux in a CFTR-expressing cell line, as well as on I_{sc} in intestinal tissue (15). In addition, the type 2 LPA receptor (LPA₂R, encoded by *Lpar2*), which was shown to mediate this inhibitory effect, was shown to interact with NHERF2 (15), suggesting that NHERF2 mediated the link between CFTR and the LPA₂R in the BBM. We sought to test this hypothesis in intact murine duodenum in vivo by searching for LPA₂R, CFTR, and NHERF2 colocalization in the murine duodenum using 2 different approaches. First, we performed Western blot analysis of isolated duodenal BBM and found LPA₂R as well as NHERF2 to be enriched in the apical membrane (Figure 6A). Immunohistochemical analysis was not possible because no specific (i.e. LPA₂R peptide-blocked) staining was detected. We therefore laser dissected the epithelial cells lining the crypt region and those lining the upper villous region (not the tip, but the upper half), and performed quantitative PCR to check for expression of LPA₂R, NHERF2, and CFTR. Interestingly, all 3 mentioned mRNAs were strongly enriched in the crypt region (Figure 6, B-D), whereas proteins typically predominant in the

**Figure 3**

Colocalization of CFTR and β_2 -AR in the apical membrane of duodenal crypts. **(A)** Top: Immunohistochemical staining with an anti-CFTR antibody in *Cftr*^{-/-} and *Cftr*^{+/+} duodenum showed apical crypt-predominant staining only in the *Cftr*^{+/+} tissue. Bottom: Apical staining with an antibody against β_2 -AR was blocked by preincubation with peptide used to raise the β_2 -AR antibody. β_2 -AR antibody stained the apical membrane of crypt as well as villous enterocytes. **(B)** CFTR and the β_2 -AR colocalized in the apical membrane of epithelial cells. In *Nherf1*^{-/-} mice, the distribution of both proteins appeared more diffuse. Scale bars: 75 μ m (**A**); 20 μ m (**B**).

villous, such as NHE3, were appropriately enriched in the villous region (data not shown; see Methods for details).

We then assessed the effect of different concentrations of LPA 20:4 in the luminal perfusate on FSK-stimulated duodenal HCO_3^- secretion. Based on our findings, 50 μ M LPA 20:4 was used for all subsequent experiments (see Methods). In WT mice, the simultaneous application of 50 μ M LPA 20:4 with 10^{-4} M FSK resulted in a significantly reduced secretory response compared with FSK alone (Figure 7, A and B). In *Nherf2*^{-/-} mice, no significant difference was seen in the HCO_3^- stimulatory response to FSK with and without coapplication of 50 μ M LPA 20:4 (Figure 7, C and D). The loss of *Nherf1* or *Pdzk1*, on the other hand, did not abolish the inhibitory effect of LPA 20:4 on duodenal HCO_3^- secretion in vivo (Figure 7E). These results demonstrate an absolute requirement of NHERF2 for LPA₂R-mediated inhibition of CFTR.

Pdzk1 deletion suppresses overall HCO_3^- secretory rate, but does not interfere with the stimulatory effect of FSK. *Pdzk1* deletion resulted in a significant reduction of basal HCO_3^- secretion, and although

this decrease in HCO_3^- secretory rate remained during FSK stimulation, the relative stimulatory effect of FSK was preserved (Figure 8, A and B). In *Pdzk1*^{-/-}*Cftr*^{-/-} double-deficient mice, very low basal bicarbonate secretion and no significant stimulation of the bicarbonate secretion after FSK perfusion was observed. Surprisingly, the combined deficiency of PDZK1 and NHERF1 (*Nherf1*^{-/-}*Pdzk1*^{-/-} mice) did not result in a significant further reduction in basal and FSK-stimulated secretion compared with that in *Nherf1*^{-/-} mice, and the absence of all 3 NHERF proteins (*Nherf1*^{-/-}*Nherf2*^{-/-}*Pdzk1*^{-/-} mice) resulted in a slight but significant increase in basal and FSK-stimulated HCO_3^- secretory rate compared with *Nherf1*^{-/-}*Pdzk1*^{-/-} mice (Figure 8, C and D).

Discussion

The results of the present study revealed important and differential roles of the NHERF1–NHERF3 proteins in the regulation of CFTR-mediated HCO_3^- secretion in vivo. The absence of NHERF1 caused a strong reduction in basal HCO_3^- secretory rate in vivo,

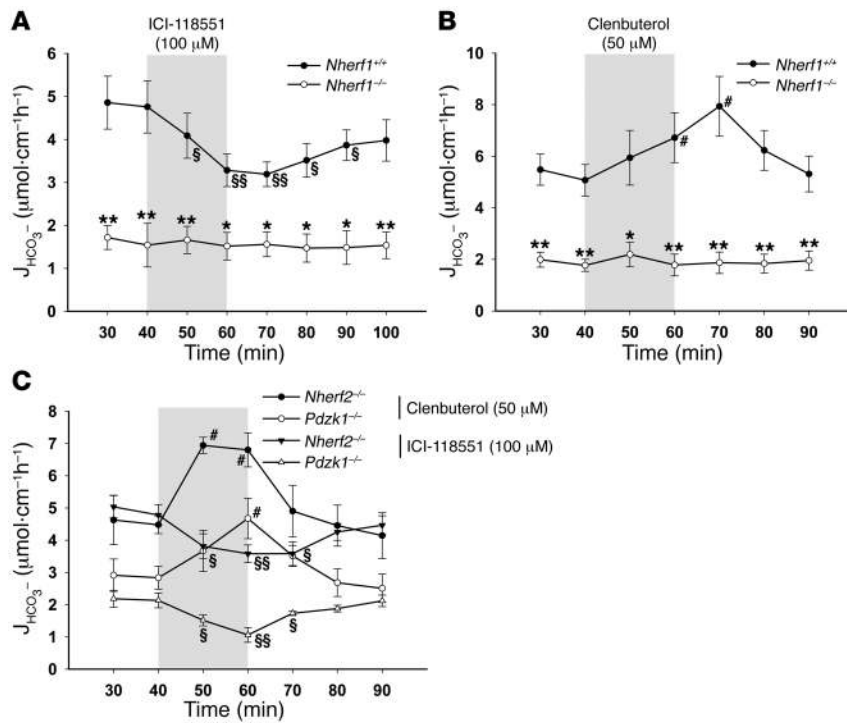


Figure 4
 NHERF1 is essential for β_2 -AR-regulated duodenal HCO_3^- secretion. **(A and B)** The luminal application of 100 μ M ICI-118551 (see Methods) significantly reduced the basal HCO_3^- secretory rate in WT but not *Nherf1^{-/-}* mice ($n = 5$). **(B)** Conversely, the luminal application of 50 μ M clenbuterol (see Methods) significantly stimulated the duodenal HCO_3^- secretory rate in WT but not *Nherf1^{-/-}* mice ($n = 5$). **(C)** Both ICI-118551 and clenbuterol had inhibitory and activating effects in *Nherf2^{-/-}* and *Pdzk1^{-/-}* mice, which indicates that the β_2 -AR-dependent regulation of CFTR is specific for NHERF1. The 20-min application time is denoted by shading. * $P < 0.01$, ** $P < 0.001$ versus WT; § $P < 0.01$, §§ $P < 0.001$ versus basal value (decrease); # $P < 0.01$ versus basal value (increase).

which was not observed in a previous in vitro study (31). Furthermore, the maximal HCO_3^- secretory rate after FSK addition was strongly reduced in *Nherf1^{-/-}* mice, consistent with the previous in vitro findings (31). In addition, a total abrogation of the HCO_3^- secretory response to the β_2 -AR agonist clenbuterol was observed in *Nherf1^{-/-}* duodenum, suggesting NHERF1-dependent regulation of this G protein-coupled receptor in vivo.

The deletion of the structurally similar NHERF2 resulted in a completely different phenotype. The basal duodenal HCO_3^- secretory rate was slightly, but not significantly, higher than in WT mice, but the FSK-stimulated HCO_3^- secretory rate was significantly increased. This suggested that NHERF2 exerts an inhibitory effect on intestinal CFTR function. To validate this concept, we studied LPA-mediated inhibition of FSK-stimulated HCO_3^- secretion and found this inhibition to be absent in *Nherf2^{-/-}* mice. Also, the additional *Nherf2* deletion partly rescued the depressed basal HCO_3^- secretory rate in the *Nherf1^{-/-}* mice, but augmented the maximal FSK-stimulated rate relatively little, which suggests that NHERF1 and NHERF2 act independently of one another.

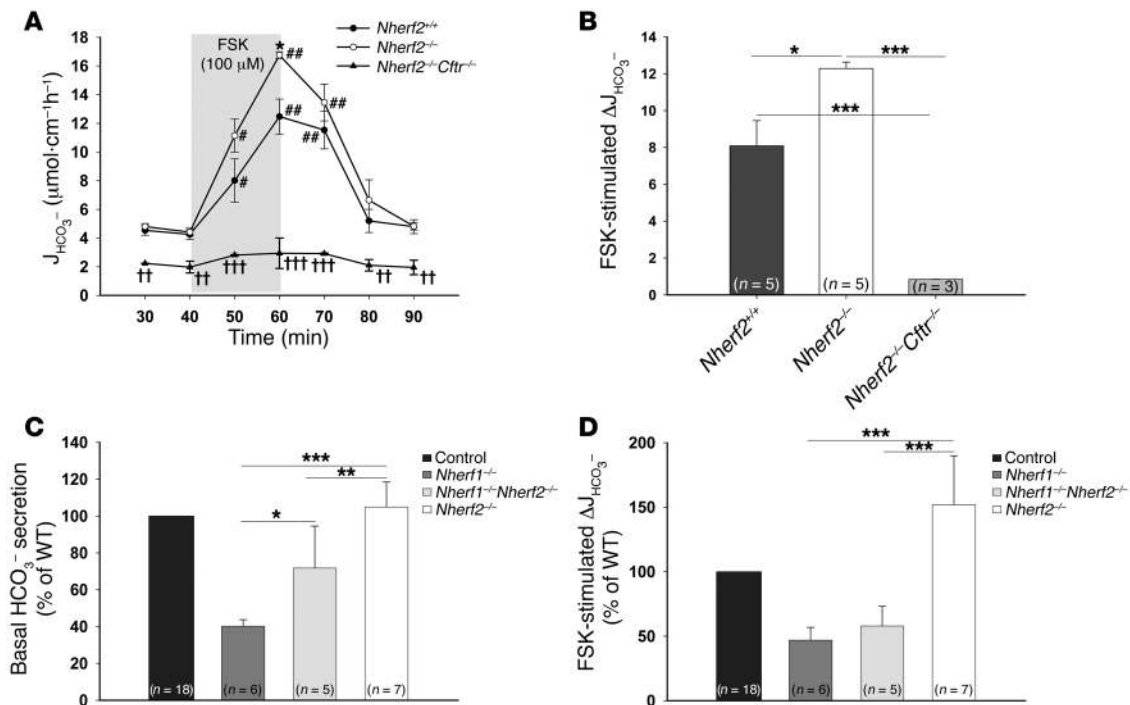
Pdzk1 ablation caused reduced basal duodenal HCO_3^- secretion in vivo, but left the relative stimulatory effect of FSK unaffected. Interestingly, *Nherf1^{-/-}Pdzk1^{-/-}* mice displayed no significant further reduction in basal or stimulated HCO_3^- secretion compared with *Nherf1^{-/-}* mice, which suggests that PDZK1 may not be able to augment CFTR activity in the absence of NHERF1.

The literature provides ample evidence for a role of the PDZ-binding motif of CFTR in the trafficking, membrane retention (5–7), dimerization (8, 9), regulation of the lateral mobility in the membrane (10, 11), and interaction with other transport proteins (12–14). Most of these studies were performed in cell culture, expressing WT and truncated CFTR and studying the above-named functions in a cell-specific setting. Other studies have coexpressed CFTR and one of the NHERF proteins and studied the effect on one of the functions mentioned (2, 3, 40, 41). Cross-

linking experiments allowed the isolation and study of multimeric CFTR-containing complexes from native apical membranes (4). The various studies have provided detailed insight into the molecular interactions of CFTR, the NHERF proteins, and other partners in the PDZ adapter-mediated protein complexes. They have, however, provided strong differences in the consequences of disrupting CFTR-NHERF interactions, from a virtually complete trafficking deficit (5) to very mild functional disturbances (6, 7, 42). Whereas some studies suggested an essential role of NHERF proteins for CFTR dimerization (8, 9, 43), others proposed that CFTR dimers form cAMP dependently, but NHERF independently (4). Thus, the studies of CFTR function in the various NHERF-deficient mice were eagerly awaited.

CFTR functional activity in vivo is assessed by nasal potential difference measurements (44), but these measurements are confounded by the very active Na^+ conductance in these epithelia (45–47). In the duodenum, CFTR expression is particularly high within the gastrointestinal tract (48), and murine duodenal HCO_3^- secretion and its agonist-mediated regulation are virtually completely dependent on CFTR expression (33–36). We therefore believe that the measurement of murine duodenal HCO_3^- secretion may currently be one of the best parameters of studying CFTR regulation in vivo.

Duodenal HCO_3^- secretion has been studied in the *Nherf1^{-/-}* mice in vitro (31). The basal HCO_3^- secretion and I_{sc} were similar to those of WT intestine, but the stimulatory response to cAMP analogs was reduced, corresponding to a reduction of CFTR staining in the apical plasma membrane by approximately 35% in jejunal crypts (31). The abundance of PDZK1, the absence of which resulted in a decrease of HCO_3^- secretion and I_{sc} in isolated duodenum as well (32), was reduced in enterocytes of *Nherf1^{-/-}* mice (31), and it was therefore unclear whether the reduction of *Pdzk1* in the *Nherf1^{-/-}* jejunum may have been partly responsible for this finding.

**Figure 5**

Basal and FSK-stimulated HCO₃⁻ secretion in *Nherf2*^{-/-}, *Nherf2*^{-/-}*Cftr*^{-/-}, *Nherf1*^{-/-}*Nherf2*^{-/-}, and WT murine duodenum in vivo. (A) Duodenal HCO₃⁻ secretion was measured as described in Figure 1. The basal HCO₃⁻ secretion in *Nherf2*^{-/-} mice was unaltered, whereas the FSK-stimulated duodenal bicarbonate secretion was significantly enhanced compared with WT littermates (*n* = 5). Both basal and FSK-stimulated response were significantly reduced in *Nherf2*^{-/-}*Cftr*^{-/-} mice (*n* = 3). The 20-min application time is denoted by shading. **P* < 0.05 versus WT; #*P* < 0.01, ##*P* < 0.001 versus basal value; †*P* < 0.01, ††*P* < 0.001, †††*P* < 0.0001 versus *Nherf2*^{-/-}. (B) The relative increase in HCO₃⁻ secretory rate was significantly higher in *Nherf2*^{-/-} mice than in WT littermates (net peak, *P* < 0.05 and *P* < 0.001). (C) The absence of both *Nherf1* and *Nherf2* resulted in a significantly higher basal HCO₃⁻ secretory rate compared with *Nherf1* deletion alone, demonstrating that NHERF2 exerts an inhibitory effect independently of NHERF1. (D) Combined *Nherf1* and *Nherf2* deletion neither worsened nor significantly improved the strongly reduced FSK peak secretory rate observed in *Nherf1*^{-/-} mice. (B–D) **P* < 0.01; ***P* < 0.001; ****P* < 0.0001.

We undertook the present in vivo study to assess the role of the 3 structurally similar NHERF proteins in CFTR regulation. In the absence of NHERF1, we found a surprisingly strong reduction of basal HCO₃⁻ secretory rate, which was not seen ex vivo in Ussing chamber experiments of isolated duodenal mucosa from the same *Nherf1*^{-/-} mouse strain (31). Most likely, the HCO₃⁻ secretory rate in vivo is under a considerable secretory tone, and this involves modes of CFTR activation that are NHERF1 dependent. Evidence has been presented that receptor-mediated signaling to CFTR, mediated via the formation of a multiprotein signaling complex, may be lost in the absence of NHERF1. One example, well documented in heterologous expression systems, is the formation of a complex among the β₂-AR, NHERF1, ezrin, PKA, and CFTR that mediates cAMP-dependent, agonist-specific activation of the CFTR current (16). We therefore searched for the presence and potential colocalization of β₂-AR with CFTR in duodenal mucosa as well as for evidence of β₂-AR stimulation of duodenal HCO₃⁻ secretion. Interestingly, the β₂-AR was colocalized with CFTR in the apical membrane of duodenocytes in the crypt region, and the luminal application of the β₂-AR agonist clenbuterol elicited a robust secretory response in WT mice, which was completely abolished in *Nherf1*^{-/-} duodenum, but present in *Nherf2*^{-/-} and *Pdzk1*^{-/-} duodenum. In addition, the β₂-AR antagonist ICI-118551 caused a significant decrease in the basal duodenal HCO₃⁻ secretory rate

in vivo in *Nherf1*^{+/+} mice, but not in *Nherf1*^{-/-} mice. This suggests (a) that an activation of β₂-ARs and a β₂-mediated stimulation of bicarbonate secretion are indeed contributing to the basal secretory rate in vivo, and (b) that receptor-mediated CFTR activation can be totally dependent on a specific NHERF isoform, despite the fact that highly homologous PDZ proteins are expressed in the same cells. Interestingly, *Adrb2* mRNA expression was significantly upregulated in the duodenum of *Nherf1*^{-/-} mice. Little is known about the physiological role of β₂-AR stimulation in the small intestine, but a body of literature exists showing that dopamine and dopamine agonists stimulate duodenal HCO₃⁻ secretion (49) and are protective for gastrointestinal mucosa, whereas dopamine antagonists cause duodenal ulceration (50). Recent data demonstrate that dopamine acts in part via binding to β₂-ARs (51), and that not only neural, but also gastrointestinal, epithelial cells can synthesize dopamine, particularly the gastric mucosa (52). Thus, a dopamine release into the luminal fluid, with subsequent access to luminal receptors, is a potential scenario for duodenal mucosal protection and deserves further study.

NHERF2 is highly homologous to NHERF1 (21–25) and has been previously shown to provide the link among CFTR, ezrin, and PKA in airway cells as well as in the A6 kidney cell line (38, 53), thereby allowing cAMP stimulation of CFTR. Surprisingly, we found a significantly increased rate of FSK-

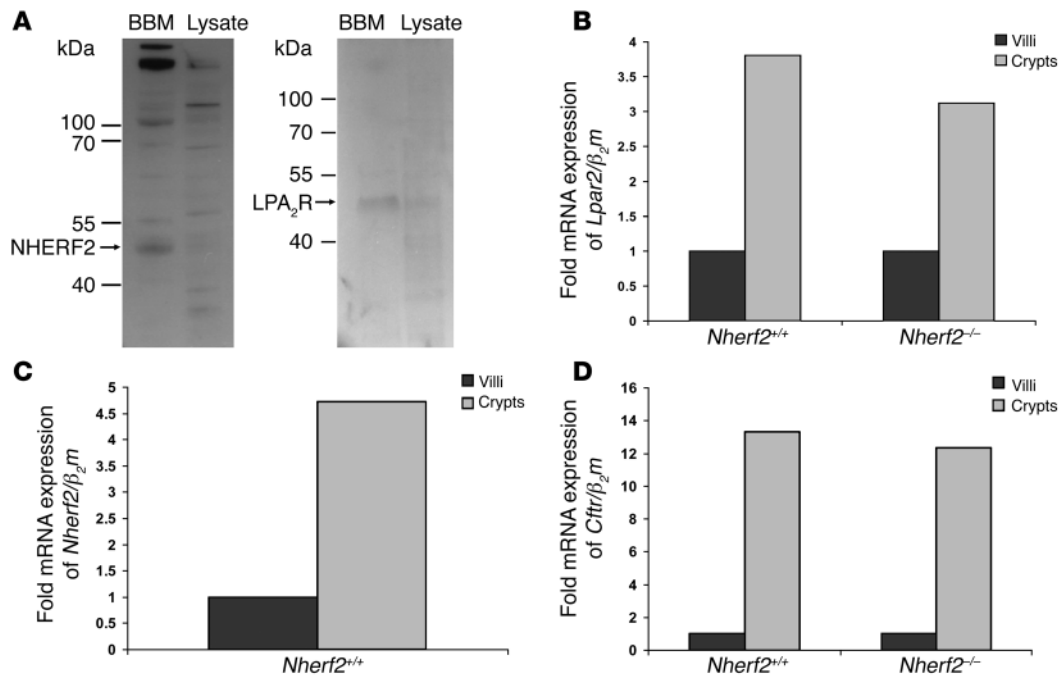


Figure 6

Duodenal epithelial apical membrane localization of NHERF2 and the LPA₂R and mRNA expression of *Lpar2*, *Nherf2*, and *Ctrr*. (A) Western blot analysis of enriched BBM and duodenal epithelial lysate using antibodies against NHERF2 and LPA₂R. Both proteins were enriched in the duodenocyte BBM fraction. The expected molecular weight of the protein is indicated. (B–D) The mRNA expression levels of *Lpar2* (B), *Nherf2* (C), and *Ctrr* (D) were all markedly higher in the crypt region than in the villous region, indicating coexpression of these 3 proteins in the cryptal region of the duodenum. The *Ctrr* and *Lpar2* mRNA expression levels were not significantly different in the *Nherf2*^{-/-} mice compared with WT.

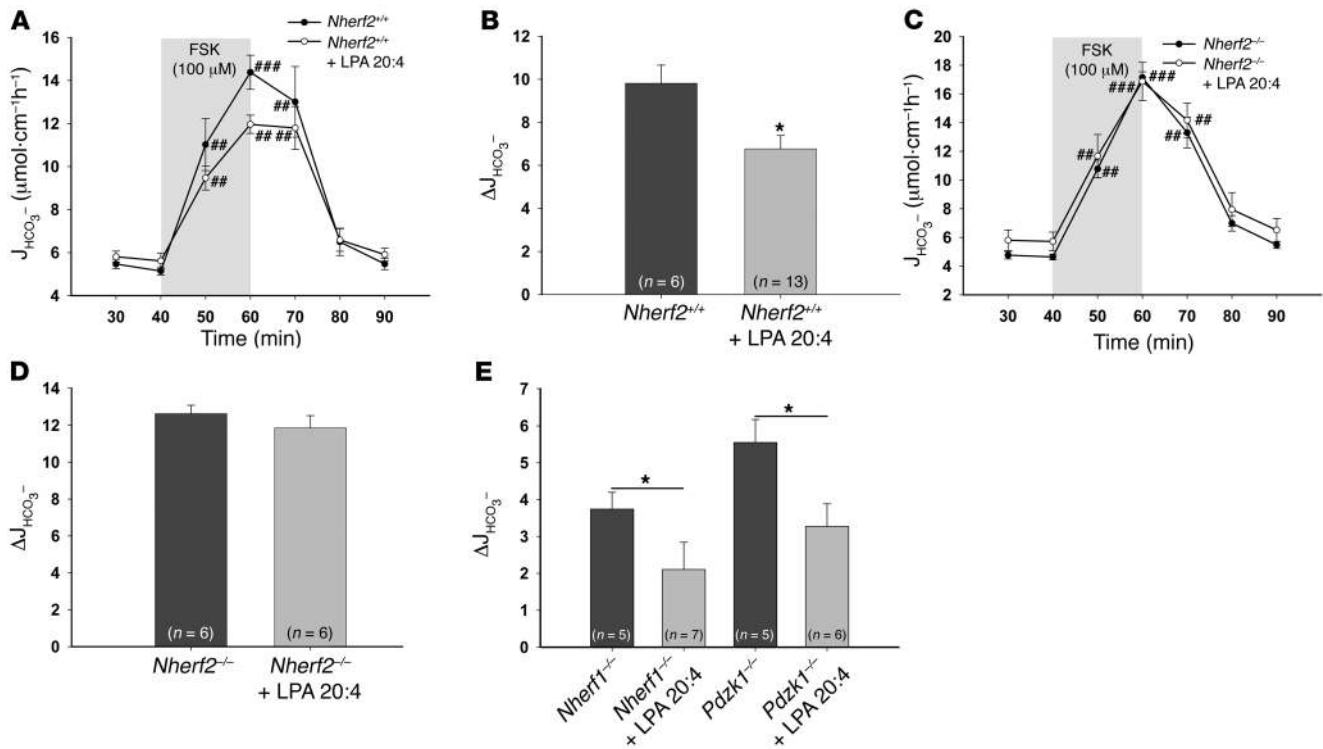
stimulated secretory response in *Nherf2*^{-/-} compared with WT mice. Obviously, the presence of NHERF2 had some inhibitory effect on CFTR activation by FSK in vivo. To further explore this surprising finding, we generated *Nherf1*^{-/-}*Nherf2*^{-/-} mice and compared them with *Nherf1*^{-/-} mice from the same generation and *Nherf1*^{+/+}*Nherf2*^{+/+} cousins. The additional *Nherf2* deletion increased the basal HCO₃⁻ secretory rate, in line with an inhibitory tone on CFTR conveyed by NHERF2, independent of the presence or absence of NHERF1. Li, Naren, and colleagues provided evidence for the formation of a complex between the apical LPA₂R and CFTR via NHERF2 in Calu-3 cells and further demonstrated that the administration of LPA 20:4 reduced adenosine-amiliorated I_{sc} in chambered mucosa as well as the fluid secretory response to cholera toxin in isolated intestinal loops in vivo (15). We tested various LPA types and concentrations and found them to indeed inhibit maximal FSK-stimulated HCO₃⁻ secretory response when administered at the luminal side concomitantly with FSK. Interestingly, this inhibition was completely absent in *Nherf2*^{-/-} mice, but present in *Nherf1*^{-/-} and *Pdzk1*^{-/-} duodenum, demonstrating the absolute requirement of NHERF2 for LPA-mediated CFTR inhibition. Additional NHERF-mediated regulation of CFTR by G protein-coupled receptors may exist, and may be responsible for the augmented FSK-stimulated HCO₃⁻ response in the absence of NHERF2, as well as the increase in basal duodenal HCO₃⁻ secretory rate in *Nherf1*^{-/-}*Nherf2*^{-/-} mice compared with *Nherf1*^{-/-} mice.

PDZK1 has previously been shown to augment CFTR dimerization, resulting in an increase in the maximal CFTR current

(9). It has also been implicated in forming a complex containing CFTR and the ABC transporter MRP4 in intestinal cell lines, promoting the efflux of cAMP, thereby downregulating CFTR activity (14). If this system were active in the murine duodenum, a lack of PDZK1 could also result in increased CFTR activity, or an enhancement of the response to a cAMP-dependent stimulus, which we did not observe. MRP4 expression in the duodenum, however, is very low (54).

In fact, the in vivo duodenal HCO₃⁻ secretory rate in *Pdzk1*^{-/-} mice was significantly decreased compared with WT mucosa, and while this decreased rate was maintained during stimulation by FSK, the secretory response to FSK was completely normal. One explanation may be that among the NHERF proteins, PDZK1 is the preferential binding partner for DRA (Slc26a3) in rabbit BBM (55) and for PAT1 (Slc26a6) in murine proximal tubule (56). Both Slc26 members are highly expressed in the duodenal BBM and involved in electroneutral HCO₃⁻ secretion (57, 58). One may speculate that DRA and/or PAT1 membrane abundance and/or the coupling to CFTR is PDZK1 dependent. We cannot test this attractive hypothesis presently because of a lack of suitable antibodies.

In summary, the results of the present study demonstrate that the absence of each of the 3 CFTR-interacting NHERF proteins resulted in distinct alterations in the regulation of CFTR-dependent murine duodenal HCO₃⁻ secretion. Despite their structural similarities, each NHERF protein enabled a unique subset of regulatory CFTR channel interactions. Thus, genetic changes in the different NHERF proteins are likely to result in pathological

**Figure 7**

The inhibitory effect of LPA on CFTR-mediated duodenal HCO_3^- secretion is absent in $Nherf2^{-/-}$ mice. (A–D) LPA 20:4 (50 μM), when added simultaneously with 100 μM FSK to the luminal perfusate, resulted in significant inhibition of the FSK-stimulated HCO_3^- secretory response in WT mice (A and B), but not in the $Nherf2^{-/-}$ mice (C and D), demonstrating that NHERF2 confers inhibitory signals to CFTR-mediated anion secretion in vivo. In A and C, the 20-min application time is denoted by shading. (E) In contrast, both $Nherf1^{-/-}$ and $Pdzk1^{-/-}$ mice showed a significant decrease in the FSK-stimulated duodenal bicarbonate secretion upon coproduction with 50 μM LPA 20:4, indicating that interaction between CFTR and LPA₂R is NHERF2 specific. * $P < 0.05$ versus respective control; # $P < 0.01$, ### $P < 0.001$, #### $P < 0.001$ versus basal value.

alterations of electrolyte transport, and therapeutic interventions targeting individual NHERF proteins may be more effective and specific than previously recognized.

Methods

Chemicals and solutions. Unless otherwise indicated, reagents were purchased from Calbiochem and Sigma-Aldrich. FSK was diluted in DMSO (10⁻² M), stored at -20°C, and added to the NaCl perfusate to a final concentration of 10⁻⁴ M. LPA 20:4 (Avanti Polar Lipids) was diluted to 0.1% (w/v) in PBS (pH 7.2) and added to the NaCl perfusate to a final concentration of 50 μM just before delivery to the duodenum. Clenbuterol and ICI-118551 were purchased from Alexis Biochemicals and diluted in water. Final concentrations in the perfusate were 50 μM of clenbuterol and 100 μM of ICI-118551.

Animals. All studies were approved by the Hannover Medical School Committee on investigations involving animals and the Regierungspräsidentium. All *Nherf* mutations were congenic for the FVB/N background for at least 15 generations. Age- and sex-matched littermates or cousins were used as controls, and, in the case of double- or triple-KO mice, controls were derived from the same founders. The genotype was determined by PCR. The primer sequences for genotyping *Nherf1* mice were as follows: NHERF1 WT forward, TCGGGGTTGTTGGCTGGAGAC; NHERF1 common reverse, AGCCCAACCCGCACTTACCA; NHERF1 mutant forward, AGGGCTGGCACTCTGTCG. Amplicons generated by PCR were 294 bp for the WT gene and 242 bp for the KO gene. For the *Nherf2*^{-/-} mice (originally generated from Lexicon genetics clone OST2298; ref. 31), the 3 oligonucleotide primers

were as follows: E3KARP WT reverse, CCCACCCCATCGCTGCTC; E3KARP common forward, TTCTATAAGCCTCCATTTCTCT; and E3KARP mutant reverse, GCGCCAGTCCTCCGATTGA. Amplicons generated by PCR were 303 bp for the WT gene and 229 bp for the KO gene. *Pdzk1* mice were genotyped as described previously (29). The mice were housed in a standard animal care room with a 12-h light/12-h dark cycle and were allowed free access to food and water until the start of the experiments.

Surgical procedure and experimental protocol for the measurement of duodenal bicarbonate secretion in vivo. The procedure for the measurement of duodenal HCO_3^- secretion in vivo has been recently described in detail (36). The concentration of FSK (100 μM) was based on the previous report by Hogan et al. (35). The concentration of LPA (50 μM) was based on the paper by Li et al. (15), and different concentrations were tested prior to the use of this concentration. The concentrations for specific β_2 -AR agonist clenbuterol (50 μM) and specific β_2 -AR antagonist ICI-118551 (100 μM) were taken from the literature (59) and verified experimentally.

Measurement of luminal alkalization. The rate of luminal alkalization was determined by back-titration (PHM82 STANDARD pH meter; Radiometer), as previously described (36).

Quantitative real-time PCR. For reverse transcription of RNA, the SuperScript III RNaseH Reverse Transcriptase (Invitrogen) was used. Primers for *Adrb2*, *Villin*, and *Rps9* as the reference genes were designed with Primer Express (version 2.0; Applied Biosystems) or PRIME (Husar, STZ Genominformatik) software. The sequences for the *Adrb2* (encoding β_2 -AR; GenBank accession no. NM_007420) were as follows: *Adrb2* forward, CTTCATTGATGTGTTGTGCG; *Adrb2* reverse, ATTCTTGGT-

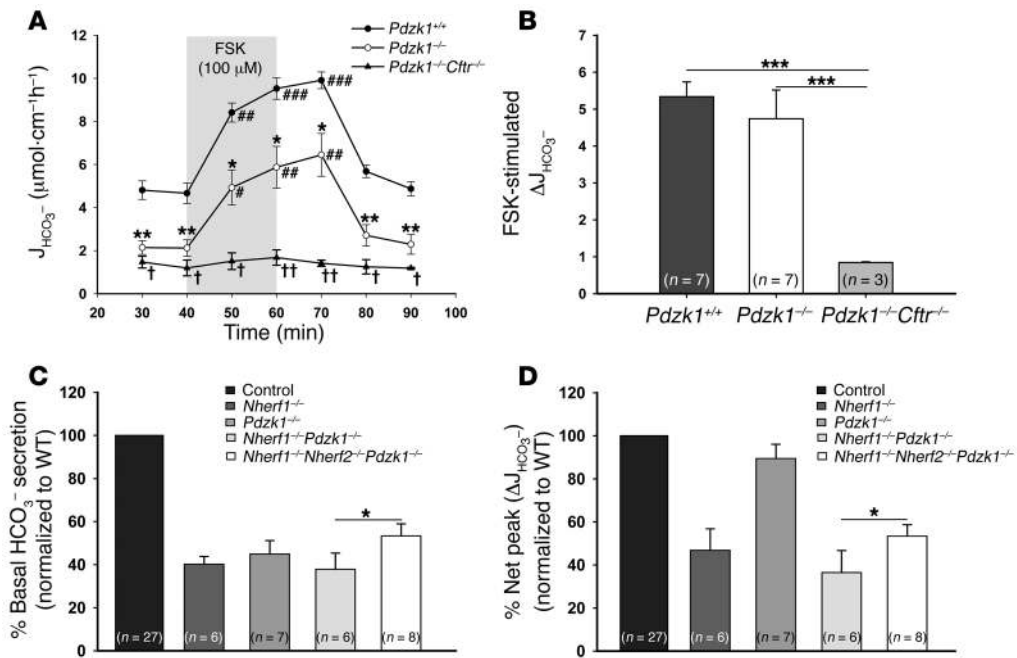


Figure 8

Basal bicarbonate secretion and FSK-stimulated peak secretory response in *Pdzk1*^{-/-}, *Pdzk1*^{-/-}*Cftr*^{-/-}, *Nherf1*^{-/-}*Pdzk1*^{-/-}, and *Nherf1*^{-/-}*Nherf2*^{-/-}*Pdzk1*^{-/-} duodenum. (A and B) Basal duodenal HCO₃⁻ secretion was significantly reduced in *Pdzk1*^{-/-} mice compared with WT littermates (n = 7). In A, the 20-min application time is denoted by shading. *P < 0.01, **P < 0.001 versus WT; #P < 0.01, ###P < 0.001, ####P < 0.001 versus basal value; †P < 0.01, ††P < 0.001 versus *Pdzk1*^{-/-}. (B) The response to FSK was not altered in *Pdzk1*^{-/-} mice, but was virtually abolished in *Pdzk1*^{-/-}*Cftr*^{-/-} mice. (C) The additional deletion of *Nherf2* resulted in a higher basal HCO₃⁻ secretory rate than in *Nherf1*^{-/-} and *Nherf1*^{-/-}*Pdzk1*^{-/-} mice, congruent with the results in *Nherf2*^{-/-} and *Nherf1*^{-/-}*Nherf2*^{-/-} mice. (D) The defect in FSK stimulation resulting from absence of *Nherf1* was the dominant disturbance in the *Nherf1*^{-/-}*Pdzk1*^{-/-} and *Nherf1*^{-/-}*Nherf2*^{-/-}*Pdzk1*^{-/-} mice. (B–D) *P < 0.05; ***P < 0.001.

CAGCAGGCTCTG; PCR product, 116 bp. The other primers and the PCR procedure have been described previously (60).

Preparation of BBMs. Mouse BBM vesicles from scrapings of duodenal epithelium were prepared as previously described by Hillesheim et al. for the jejunum (32). The duodenum was defined as the first 3 cm of the small intestine. Duodenal scrapings from 2 mice were pooled per preparation, and the experiments were repeated 3 times. The enriched BBM fraction was assayed for its enrichment factor by performing sucrase and alkaline phosphatase assays as described previously (32).

Western blot analysis. Western blots were prepared as described previously (32, 61), with the following modifications. Blots were probed with rat anti-CFTR antibody (3G11 rat monoclonal against mouse CFTR, diluted 1:500; provided by W.E. Balch, Scripps Research Institute, La Jolla, California, USA), rabbit anti-NHERF1 antibody (Ab5199, diluted 1:15,000; provided by C. Yun, Emory University, Atlanta, Georgia, USA), rabbit anti-NHERF2 antibody (Ab2170, diluted 1:5,000; provided by C. Yun), rabbit anti-β₂-AR (diluted 1:200; Santa Cruz Biotechnology Inc.) with and without blocking peptide in TBS-Tween, goat anti-LPA₂ antibody (diluted 1:200; Santa Cruz Biotechnology Inc.) in background reducing buffer (DAKO) overnight at 4°C. To check the specificity of the staining, the β₂-AR and LPA₂R blocking peptides were used. For CFTR, NHERF1, and NHERF2, tissue from KO mice was used.

Immunofluorescence staining. Duodenum from *Nherf1*^{-/-} and WT mice was pretreated as described by Cinar et al. (61). Cryosections were treated sequentially with PBS for 5 min, 6 M guanidine (pH 7.0) for 10 min, and washing buffer (PBS with 0.4 M NaCl, 0.1% BSA, and 0.01% Triton ×100) 3 times for 5 min each. Blocking in 10% goat serum for 1 h was followed by incubation overnight with primary antibodies CFTR (as described above, dilut-

ed 1:300; provided by W.E. Balch) and affinity-purified rabbit polyclonal β₂-AR (diluted 1:100; Santa Cruz Biotechnology Inc.) at 1:100 in incubation medium (PBS with 3% goat serum, 3% BSA, and 0.3% Triton ×100). Washing 4 times for 5 min in washing buffer was followed by incubation with secondary antibodies (Alexa Fluor 488-labeled goat anti-rat IgG or Alexa Fluor 546-labeled goat anti-rabbit IgG; Invitrogen) for 1 h at room temperature at a concentration of 2 μg/ml in incubation medium. After 4 washes for 5 min, each cover slide was mounted with SlowFade Gold antifade reagent with DAPI (Invitrogen), and slides were imaged on a confocal microscope (TCS SP2; Leica). Excitation wavelengths used were 405, 488, and 543 nm, and emission was quantitated at 415–450, 490–540, and 560–650 nm for detection of DAPI, Alexa Fluor 488, and Alexa Fluor 546, respectively.

Laser microdissection and mRNA quantification. For microdissection of crypt and villous duodenocytes from *Nherf2*^{-/-} and WT mice, cryostat sections were mounted onto MembraneSlides 1.0 PEN (Carl Zeiss) and fixed with 70% ethanol for 2 min, stained for 3 min in hematoxylin, immersed in 100% ethanol, soaked in xylene for 3 min, and air dried. The Laser-Microdissection Microscope (P.A.L.M.) was used to microdissect duodenocytes from the cryptal and the upper villous region (not the tip). Approximately 3,000 microdissected cells per region (crypt versus villi from each duodenum) were picked into RLT buffer, RNA was isolated with RNeasy MicroKit (Qiagen), and reverse transcription was performed as described previously (32). The PCR amplification was performed as described previously (31), but using MESA GREEN qRT-PCR Master-Mix Plus instead of SYBR Assay I dTTP (Eurogentec). Primer sequences were as follows: CFTR forward, 5'-TTCTTCACGCCCCCTATGTCGA-3'; CFTR reverse, 5'-GCTCCAATACAATGAACACCA-3'; CFTR PCR product length, 145 bp (GenBank accession no. NM_021050); NHERF2



forward, 5'-TAGTCGATCCTGAGACTGATG-3', NHERF2 reverse, 5'-ATTGTCCTTCTCTGAGCCTG-3'; NHERF2 PCR product length, 173 bp (GenBank accession no. NM_023449), β_2m forward, 5'-TGGTGCTTGTCTACTGACC-3'; β_2m reverse, 5'-CCGTTCTTACGATTTGGAT-3'; β_2m PCR product length, 160 bp (GenBank accession no. NM_009735); LPA₂ forward, 5'-AGCCTAGTCAAGACGGTTGTCA-3'; LPA₂ reverse, 5'-CGTTGCAGGACTTACAGTCCA-3'; LPA₂ PCR product length, 106 bp (GenBank accession no. NM_020028).

Statistics. Descriptive statistics are expressed as mean \pm SEM, with the number of mouse pairs (KO and WT) or individual mice, if applicable, given in parenthesis. Statistical analyses were performed using 2-tailed Student's *t* test for unpaired data and analysis of variance and Fisher's protected least significant difference for paired data. Results were considered significant at $P < 0.05$.

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