

Discovery of I-BRD9, a Selective Cell Active Chemical Probe for Bromodomain Containing Protein 9 Inhibition

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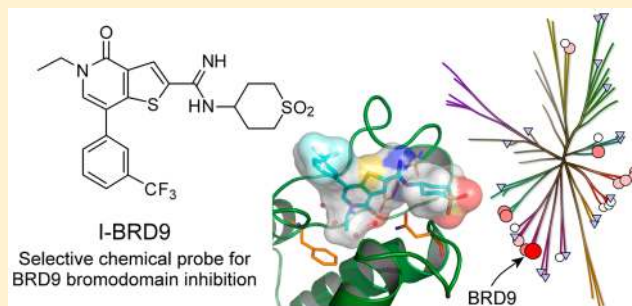
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Supporting Information

ABSTRACT: Acetylation of histone lysine residues is one of the most well-studied post-translational modifications of chromatin, selectively recognized by bromodomain “reader” modules. Inhibitors of the bromodomain and extra terminal domain (BET) family of bromodomains have shown profound anticancer and anti-inflammatory properties, generating much interest in targeting other bromodomain-containing proteins for disease treatment. Herein, we report the discovery of I-BRD9, the first selective cellular chemical probe for bromodomain-containing protein 9 (BRD9). I-BRD9 was identified through structure-based design, leading to greater than 700-fold selectivity over the BET family and 200-fold over the highly homologous bromodomain-containing protein 7 (BRD7). I-BRD9 was used to identify genes regulated by BRD9 in Kasumi-1 cells involved in oncology and immune response pathways and to the best of our knowledge, represents the first selective tool compound available to elucidate the cellular phenotype of BRD9 bromodomain inhibition.



INTRODUCTION

Chromatin has a highly complex structure composed of nucleosomes, the fundamental packaging elements of genetic information. These structural subunits consist of DNA wrapped around an octameric core of histones, which facilitate the arrangement of DNA in the nucleus of the cell.¹ Histone tails are subject to a range of post-translational modifications, aiding the regulation of chromatin accessibility.² Acetylation of lysine residues is an example of one such modification and is regulated by the dynamic action of histone acetyltransferases (HATs) and histone deacetylases (HDACs), which “write” and “erase” acetylation marks respectively.³ Bromodomain “reader” modules recognize specific acetyl-lysine (Kac) residues located on histone tails and other proteins. These recognition motifs act together with other chromatin factors to modulate regulation of gene

transcription.⁴ Bromodomains are highly conserved protein modules, comprising approximately 110 amino acids. The majority are characterized by four alpha-helices, linked by two flexible loop regions, which form the basis of the Kac binding pocket.⁵

There are at least 56 human bromodomains in 42 bromodomain containing proteins (BCPs) identified to date, neglecting splice variants, each of which has been classified into a distinct subgroup based on sequence homology.^{6–8} The bromodomain and extra terminal domain (BET) family of BCPs (BRD2, BRD3, BRD4, and BRDT) contain two

Special Issue: Epigenetics

Received: February 13, 2015

Published: April 9, 2015

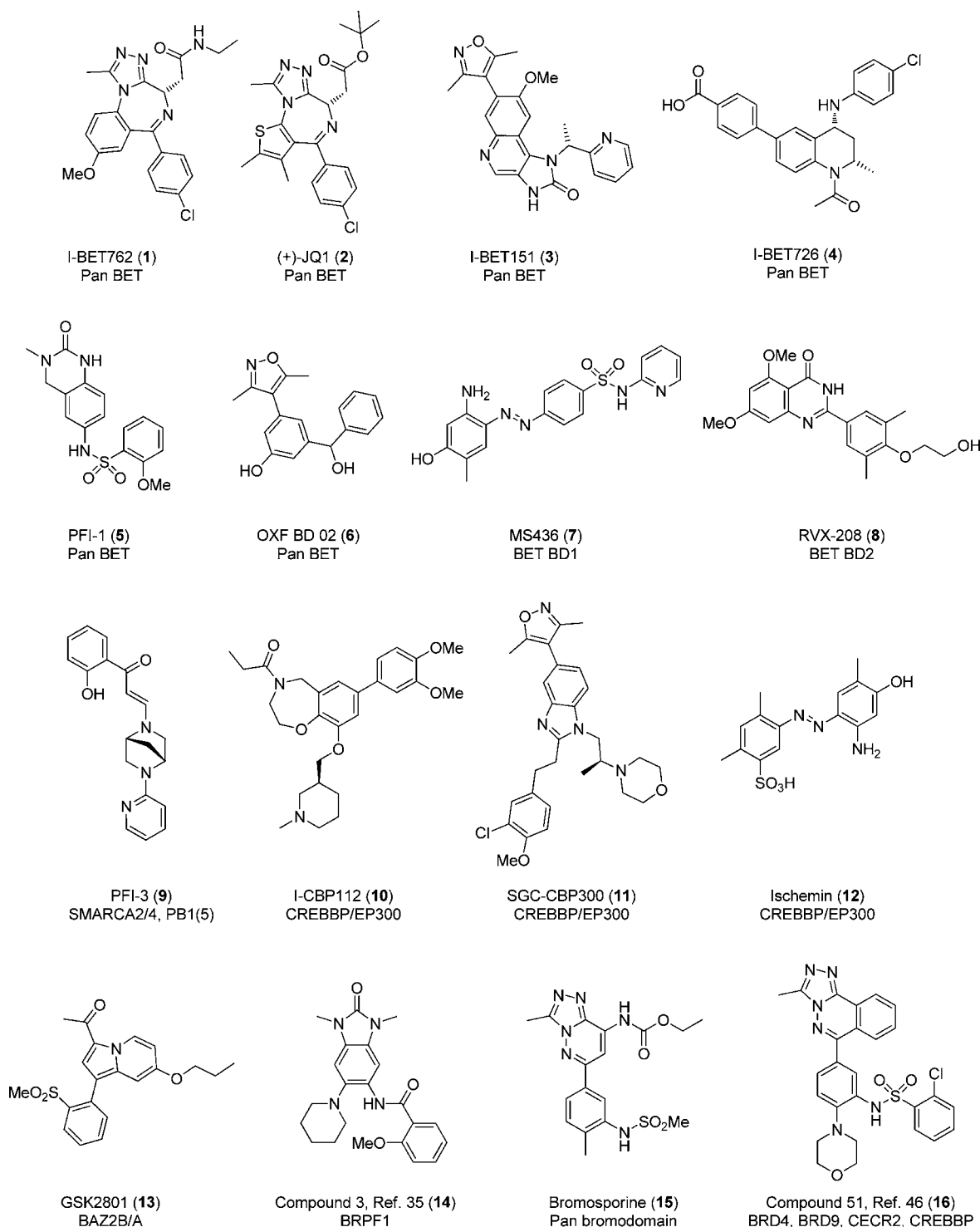


Figure 1. Chemical structures for selected bromodomain inhibitors.

bromodomains each, both capable of binding to Kac.³ The most potent and selective bromodomain inhibitors reported to date target all eight bromodomains of the BET family.^{4,9,10} Examples include I-BET762 (1),^{11–13} (+)-JQ1 (2),^{14,15} I-BET151 (3),^{16–18} I-BET726 (4),¹⁹ PFI-1 (5),²⁰ and OXF BD 02 (6)^{21,22} (Figure 1). There are also examples of compounds biased toward binding at either the N-terminal bromodomain (BD1) or C-terminal bromodomain (BD2) of the BET family such as MS436 (7)²³ and RVX-208 (8).^{24,25}

The profound biology associated with the BET family of BCPs suggests there may be therapeutic potential in targeting other members of the bromodomain phylogenetic tree for the treatment of disease. Protein knock-down studies, which block the action of an entire complex, offer a method for probing protein function and linking a BCP to a disease.³ However, as BCPs are multidomain proteins and often exist as part of multibody complexes, a chemical probe, which only inhibits the action of the bromodomain, may show different effects.²⁶ The therapeutic potential of bromodomain inhibition remains less

well explored outside of the BET family,³ in part, due to the limited availability of suitable chemical probes for each member of the human bromodomain phylogenetic tree.²⁷ Therefore, the development of high quality, selective small molecule probes for bromodomains will be a key step in facilitating validation of these reader modules as therapeutic targets. Toward this goal, several groups have generated high quality non-BET bromodomain inhibitors from a diverse set of chemotypes: PFI-3 (**9**)²⁸ for SMARCA2/4 and PB1(5) domains; I-CBP112 (**10**),²⁹ SGC-CBP300 (**11**),^{30,31} and ischemin (**12**)³² for CREBBP and EP300;³³ GSK2801 (**13**)³⁴ for BAZ2B/A, and compound **14** for BRPF1 (Figure 1).³⁵ NMR-based screening has been used to identify fragment-sized small molecule inhibitors of PCAF, BAZ2B, and ATAD2 providing further evidence for the tractability of non-BET bromodomains.^{36–38} Additionally, a fluorouracil-tagged multicomponent reaction biased around the 3,5-dimethylisoxazole Kac motif was used to generate the first compounds capable of binding to TAF1.³⁹ There are also several probes available from the Structural Genomics Consortium for the following bromodomains that have not yet had their structures disclosed: BAZ2B/A, BRPF1/2/3, and BRPF1.⁴⁰

Although BRD9 has been reported as a component of the chromatin remodeling SWI/SNF BAF complex,^{41–43} further information regarding its role within disease is somewhat limited.⁴⁴ As such, a chemical probe for the BRD9 bromodomain would be invaluable in better understanding the biological role and therapeutic potential of this reader module. The Structural Genomics Consortium have disclosed bromosporine (**15**),⁴⁵ a broad spectrum bromodomain inhibitor, which shows a thermal shift of +3.9 °C at BRD9, suggesting bromodomain binding. In addition, structurally similar compound **16** was reported as the first nonselective inhibitor of BRD9.^{46,47} This compound shows mixed bromodomain pharmacology, with submicromolar activity against BRD9, BRD4, CECR2, and CREBBP. To the best of our knowledge, there are no BRD9 selective chemical probes described in the literature to date.⁴⁸ Herein, we report the discovery of I-BRD9, the first selective cellular chemical probe for BRD9. The development of I-BRD9 was driven by iterative medicinal chemistry, utilizing structure based design to deliver nanomolar potency at BRD9, >700-fold selectivity over the BET family and >70-fold against a panel of 34 bromodomains. I-BRD9 meets and exceeds the chemical probe criteria we defined at the outset of this project, inspired by literature from Bunnage and co-workers,⁴⁹ who highlighted the importance of high quality chemical probes for target validation: 100 nM or greater potency against the bromodomain of BRD9 as determined by a biochemical assay; 100-fold selectivity over the BET family of bromodomains; 30-fold selectivity over other bromodomain families; cellular activity; and broader selectivity over a range of receptor sites, ion channels, transporters, and enzymes.

RESULTS AND DISCUSSION

Throughout the course of the investigations to identify a chemical probe for BRD9, selectivity over the BET family of bromodomains was considered of utmost importance. It was felt that the profound biology associated with BET bromodomain inhibition^{9–12} required at least 100-fold selectivity (as determined using TR-FRET analysis, see Supporting Information), to ensure that any phenotype observed through the use of a BRD9 chemical probe was driven by BRD9 inhibition alone. BRD4 BD1 (*N*-terminal, bromodomain 1) was used as a representative member of the BET family, driving investigations into selectivity. Analysis of the X-ray crystal structures of apo

BRD9 and BRD4 BD1 in complex with I-BET762 (**1**) revealed a distinct contrast in the architecture of their ZA channels, due to differences in their constituent amino acids. In BRD9, the Tyr106 gate keeper residue blocks access to the lipophilic “WPF” shelf, providing a key difference to the Ile146 gate keeper residue present at the equivalent position of BRD4 BD1 (Figure 2, Table

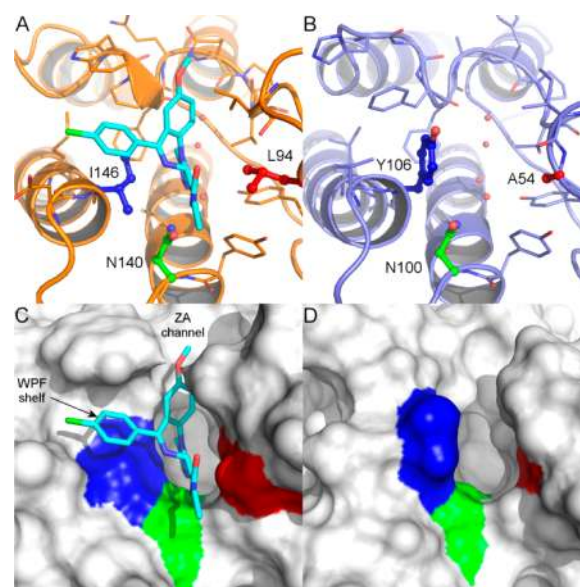


Figure 2. X-ray crystal structures of (A) BRD4 BD1 bromodomain (orange) with I-BET762 (**1**) (cyan); (B) BRD9 bromodomain (blue); (C) BRD4 BD1 surface, and (D) BRD9 surface. In each, the conserved asparagine residue is colored green (BRD4 Asn140/BRD9 Asn100), and the gatekeeper residue is colored blue (BRD4 Ile146/BRD9 Tyr106). BRD4 Leu94 and BRD9 Ala54 are colored red.

1). At this stage, it was unclear whether or not selectivity could be gained over the highly homologous BRD7, given its proximity to BRD9 on the phylogenetic tree and sequence similarity within the Kac binding pocket (Figure 3, Table 1).^{50,51}

Nonetheless, in order to identify inhibitors of the BRD9 bromodomain, a cross-screening strategy of GSK internal compounds was conducted. Thienopyridone **17** was identified from this effort as a potent binder to BRD9 (TR-FRET assay), with pIC_{50} values of 6.7 and 4.7 against BRD9 and BRD4 BD1, respectively (Figure 4).⁵²

To explain the BRD9 potency of **17** and its selectivity over BRD4, 1.3 and 1.2 Å resolution X-ray crystal structures were solved in BRD9 and BRD4 BD1, respectively. The interactions of the thienopyridone ring are similar in both bromodomains (Figure 4B–D). In BRD9, the carbonyl moiety of the pyridone ring forms a direct hydrogen-bond to the side chain NH_2 group of Asn100, the conserved asparagine residue important for Kac recognition. The pyridone carbonyl also makes a shorter hydrogen-bond to a conserved water molecule bound by Tyr57. The pyridone *N*-methyl group sits in a small cavity surrounded by four crystallographically conserved water molecules, Phe45 and Val49. The *N*-methyl pyridone moiety acts as a direct structural analogue of Kac (Figure 4D), whose interactions with BRD9 and BRD4 are typical of those reported for fragments in the corresponding site of BRD4.^{50,51} The thienopyridone rings fill a tight hydrophobic groove formed by Val49 and Ala54 of the ZA loop on one side, and Tyr106 (the gatekeeper residue) on the other. The 7-dimethoxyphenyl substituent extends outward through the ZA channel, in close

Table 1. Sequence Similarity between the Residues of the Acetyl-Lysine Binding Sites of BRD4 BD1, BRD9, and BRD7^a

	BRD4 BD1	BRD9	BRD7	
1	TRP81	GLY43	ALA154	WPF motif
2	PRO82	PHE44	PHE155	WPF motif
3	PHE83	PHE45	PHE156	WPF motif
4	GLN84	ALA46	SER157	ZA channel
5	GLN85	PHE47	PHE158	ZA channel
6	PRO86	PRO48	PRO159	ZA channel
7	VAL87	VAL49	VAL160	ZA channel
8	ASP88	THR50	THR161	ZA loop
9	ALA89	ASP51	ASP162	ZA loop
10	LYS91	INS	INS	ZA loop
11	LEU92	INS	INS	ZA loop
12	ASN93	ILE53	ILE164	ZA loop
13	LEU94	ALA54	ALA165	ZA loop
14	TYR97	TYR57	TYR168	Water-binding Tyr
15	TYR98	SER58	SER169	
16	ILE101	ILE61	ILE172	
17	MET132	MET92	MET203	
18	ASN135	ASN95	ASN206	
19	CYS136	ALA96	ALA207	Conserved Ala
20	TYR139	TYR99	TYR210	Conserved Tyr
21	ASN140	ASN100	ASN211	Conserved Asn
22	ASP144	THR104	THR215	
23	ASP145	VAL105	ILE216	
24	ILE146	TYR106	TYR217	Gatekeeper
25	VAL147	TYR107	TYR218	
26	MET149	LEU109	ALA220	
27	ALA150	ALA110	ALA221	

^aColors represent residue properties: dark green = aromatic, light green = hydrophobic, red = acidic, blue = basic, orange = polar, purple = proline, white = small. INS = indel.

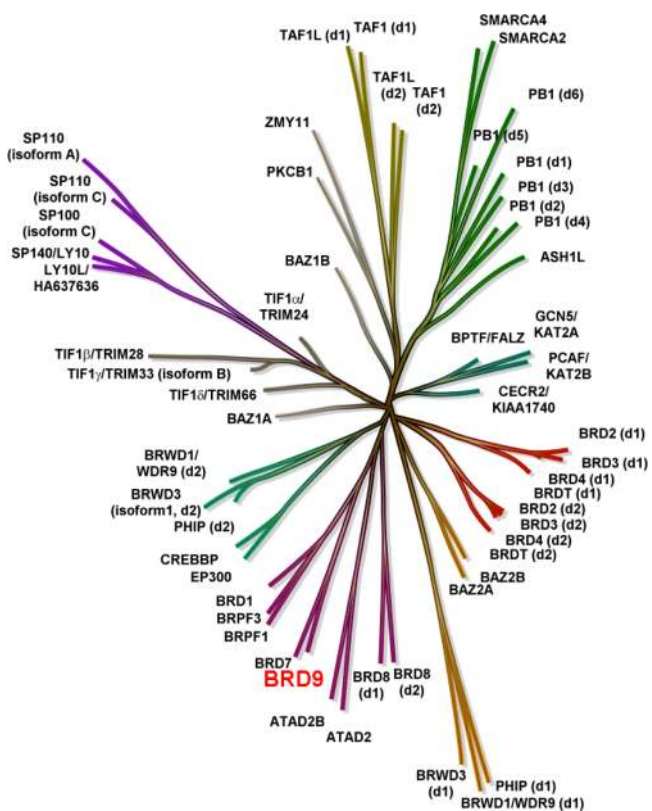


Figure 3. Bromodomain phylogenetic tree.

contact with Phe44 and Ile53 on either face. The dimethoxy groups participate in a network of hydrogen-bonded water molecules, the 4-methoxy in particular, in a water-mediated interaction with the carbonyl of His42.

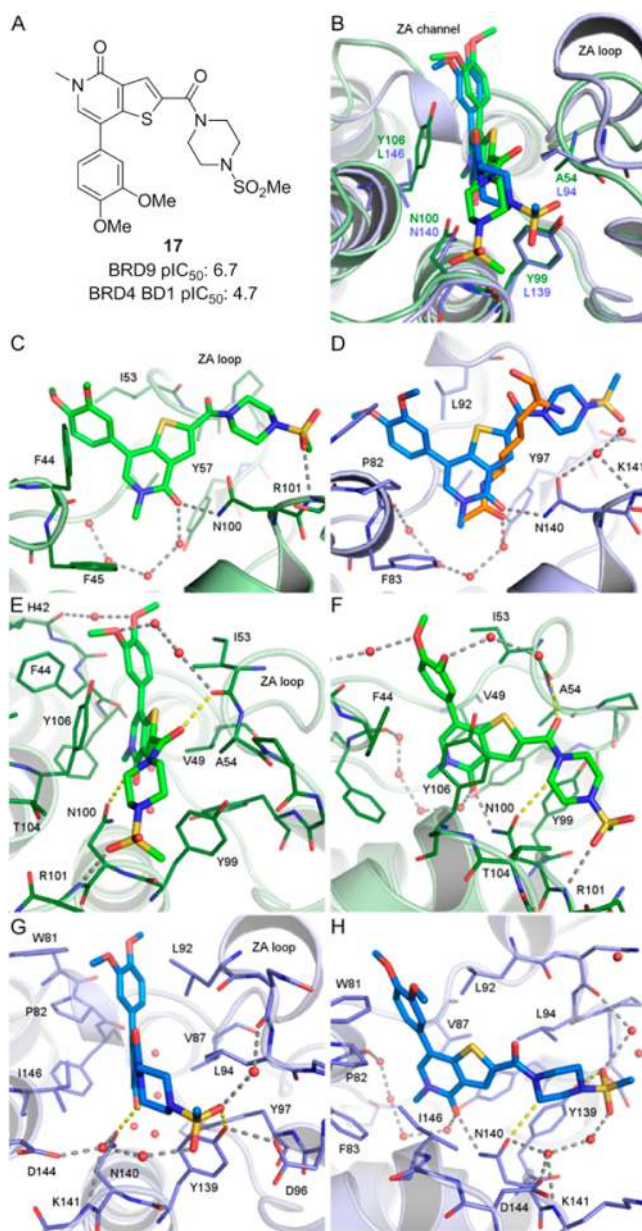
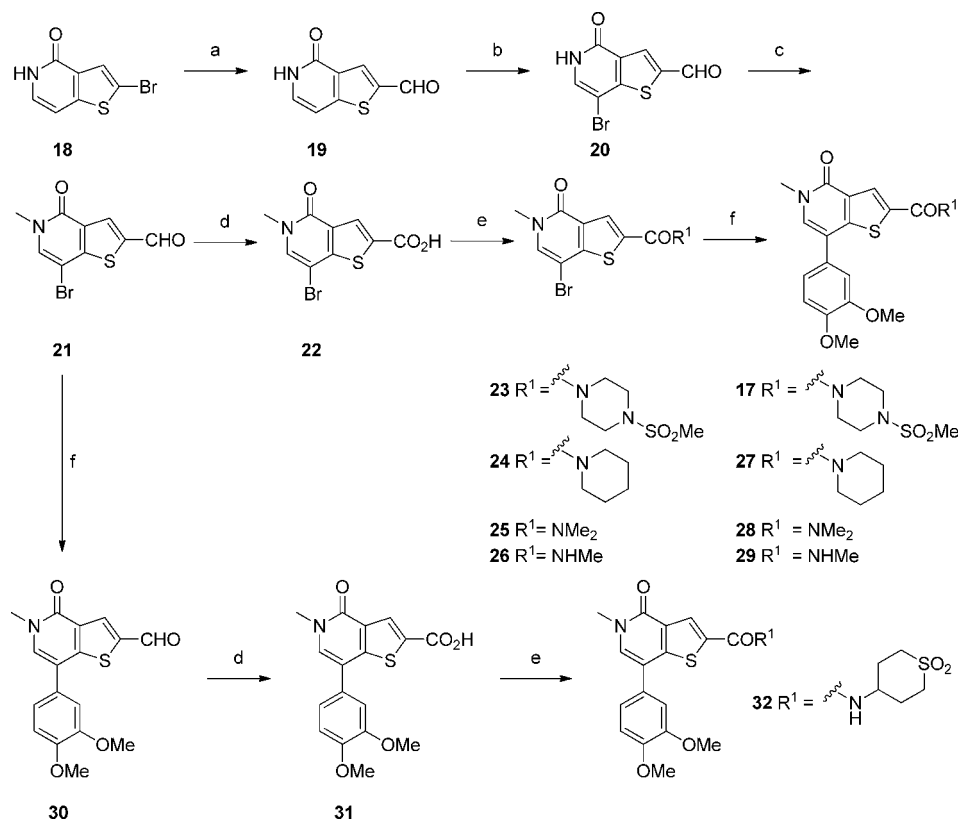


Figure 4. (A) pIC_{50} values for compound 17 in the bromodomains of BRD9 and BRD4 BD1 as determined by TR-FRET assays.⁵² (B) Differences between X-ray crystal structures of 17 in complex with the bromodomain of BRD9 (green) and BRD4 BD1 (blue). (C) Binding mode of 17 in BRD9. Hydrogen-bonds are shown as gray dashed lines and the four conserved water molecules as red spheres. Unfavorable interactions are marked as yellow dashed lines. (D) Binding mode of 17 in BRD4 BD1 (blue) with the position of acetyl-lysine (orange, bromodomains superimposed, from protein data bank entry 2wp2).⁵³ (E,F) Two different orientations of the complex of 17 with BRD9. (G,H) The complex of 17 with BRD4 BD1.

At the other end of the molecule, the piperazine amide presents a complex pattern of both desirable and undesirable interactions (Figure 4E). The first unfavorable interaction involves the amide carbonyl oxygen of the linker, which lies 2.7 Å from the carbonyl oxygen of Ile53, a close interaction for two electronegative atoms. The second involves the piperidinyl 2-methylene group, which lies 2.9 Å from the carbonyl oxygen of the conserved asparagine (Asn100) side chain, preventing its solvation by water. In compensation, the piperazine amide acts as

Scheme 1. Synthesis of Amides 17, 27–29, and 32^a

^aReagents and conditions: (a) *n*-BuLi, THF, -78°C , then DMF, THF, -60°C to rt, 65%; (b) NBS, THF, rt, 79%; (c) MeI, Cs₂CO₃, THF, rt, 71%; (d) NaClO₂, NaH₂PO₄, H₂O₂, DMSO, H₂O, rt, 42–72%; (e) HATU, amine, DIPEA, DMF, rt, 13–79%; (f) (3,4-dimethoxyphenyl)boronic acid, PdCl₂(PPh₃)₂, K₂CO₃, DME, H₂O, microwave, 120°C , 24–62%.

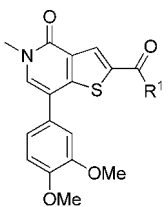
a conformationally restricted linker, directing the terminal methyl sulfonamide to a position where one of its oxygen atoms accepts a hydrogen bond from the backbone NH of Arg101.

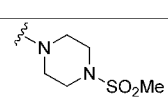
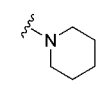
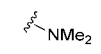
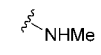
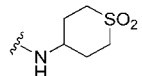
The general binding mode of the thienopyridone to BRD4 is the same as to BRD9, but more detailed interactions of this selective compound differ. One difference arises from the double substitution of BRD9 Ala54/Tyr106 for BRD4 Leu94/Ile46 on either side of the thienopyridone ring, leading to a differently shaped pocket illustrated in Figure 2. These result in a tilt of the plane of the thienopyridone ring by approximately 10° away from Leu94 in BRD4 (Figure 4B). The close proximity of Leu94 in BRD4 also forces the thiophene 2-position carbonyl bond torsion into an almost completely planar conformation, in contrast to the $\sim 30^{\circ}$ angle seen in BRD9. For a model thienopyridone with a dimethyl amide substituent, the planar BRD4-bound conformation is calculated to be slightly unfavorable (~ 0.7 kcal/mol) with respect to the energy minimum, which corresponds to the BRD9-bound conformation, being $\sim 30^{\circ}$ out-of-plane (see Supporting Information, Figure S4). Another consequence is a knock-on effect on the pendant piperidiny sulfonamide, which must adopt a different position, and as a result the sulfonamide is unable to hydrogen-bond to the backbone of Lys141 (the equivalent of Arg101 in BRD9). The electron density for this group is not well-defined (Figure S2, Supporting Information), but the best placement of the sulfonamide group is in a region predicted to be electrostatically unfavorable, close to the side chain oxygen of Tyr139. We believe that this is consistent with the lower potency

of 17 for BRD4 compared to BRD9. The hydrogen-bonding potential of Lys141 is satisfied not by the sulfonamide but instead by a water molecule, which also interacts with the side chains of Asp144 and Asp140 (Figure 4G,H).

With compound 17 as an encouraging hit, key learnings from the crystallography were taken forward to guide the design of compounds with greater potency and improved selectivity over BRD4. With the hypothesis that the selectivity over BRD4 was driven by the amide carbonyl of compound 17, amide modifications were explored to test our idea.

A synthetic route was developed to access the amide analogues from known bromothienopyridone 18⁵⁴ (Scheme 1). Halogen-lithium exchange of bromide 18 with *n*-BuLi at -78°C , followed by the addition of DMF afforded aldehyde 19. Subsequent electrophilic substitution with NBS, followed by pyridone *N*-methylation provided key intermediate 21. Amides 17 and 27–29 were synthesized by an initial Pinnick oxidation of aldehyde 21 to give acid 22, followed by amide formation. A subsequent Suzuki–Miyaura coupling with compounds 23–26 was utilized to install the dimethoxyphenyl motif. In contrast, amide 32 was synthesized first by a Suzuki–Miyaura coupling of intermediate 21, followed by a Pinnick oxidation to afford acid 31. Final amide formation using standard peptide coupling conditions furnished compound 32. This alternative route proved more useful, providing access to late stage intermediate 31. Functionalization at the 2-position of 31, through a single amide coupling, allowed for the introduction of structural diversity in the final step of the synthesis, providing straightforward access to a range of amides.

Table 2. SAR for BRD9 and BRD4 BD1 Activity of the Thienopyridone Amide Series^a


Cmpd	R ¹ =	BRD9 pIC ₅₀	BRD4 BD1 pIC ₅₀	Fold Selectivity
17		6.7 ± 0.16	4.7 ± 0.14 ⁵²	×100
27		5.4 ± 0.14	4.8 ± 0.04	×4
28		5.3 ± 0.10	4.9 ± 0.16	×3
29		6.9 ± 0.12	6.7 ± 0.16	×2
32		7.9 ± 0.09	7.3 ± 0.06	×4

^aAll results are $n \geq 3$ as determined by TR-FRET analysis.

Analysis of the resulting data relative to compound 17 indicated that the methyl sulfonamide moiety was important for BRD9 potency and, as a result, selectivity over BRD4 BD1 (Table 2). Consistent with the hydrogen-bonding interaction made by the sulfonamide with Arg101 in BRD9 but not Lys141 in BRD4, truncating to piperidine 27 or dimethyl amide 28 reduced BRD9 potency, with little effect on BRD4 activity (compare compounds 17 to 27 and 28). Interestingly, although not selective over BRD4, secondary methyl amide 29 regained the BRD9 potency observed for 17, suggesting that the amide NH makes positive interactions in both BRD4 and BRD9. Finally, cyclic sulfone amide 32 provided a 10-fold improvement in BRD9 activity relative to 29, albeit with little selectivity over BRD4. X-ray crystallography of 32 in the bromodomains of both BRD9 and BRD4 was used to rationalize the observed data (Figure 5).

The binding mode of the thienopyridone ring of 32 superimposes closely with that seen for 17, making identical interactions (Figure 5A). However, the secondary amide of 32 can move closer to the side chain of Asn100 than is possible for the tertiary amine of 17, forming a direct hydrogen-bond to the asparagine side chain carbonyl group. This additional interaction may explain the increased potency of 29 relative to 28 and contributes to the greater potency of 32. The electron density for the tetrahydro-2*H*-thiopyran 1,1-dioxide substituent is quite weak, but interestingly the best fit is with the amide at the axial rather than the equatorial position of the ring (Supporting Information, Figure S3). Another unexpected feature is that when complexed to 32 the ZA loop moves away from the inhibitor compared to its position in the complex with 17 (Figure 5A). This hinge-like motion, involving residues 50–56, results from the tetrahydro-2*H*-thiopyran ring pushing on the loop through its contacts with Tyr99.

Unexpectedly, the selectivity window for BRD9 over BRD4 is much reduced for 32 than for 17. This can be rationalized by the crystal structures of the compound bound to the two

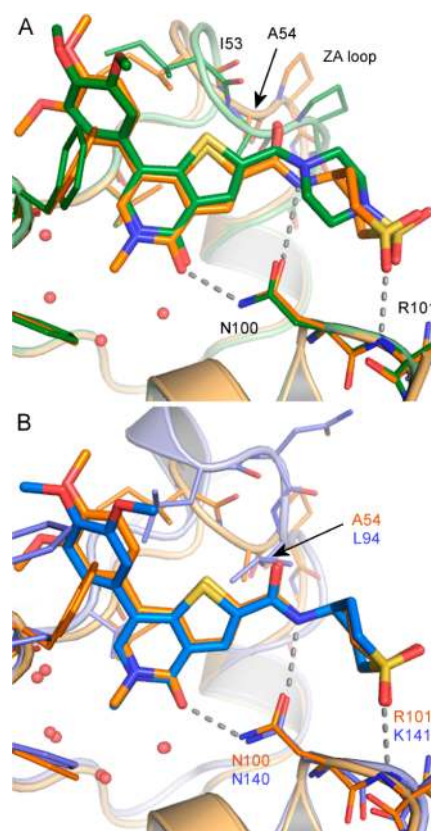
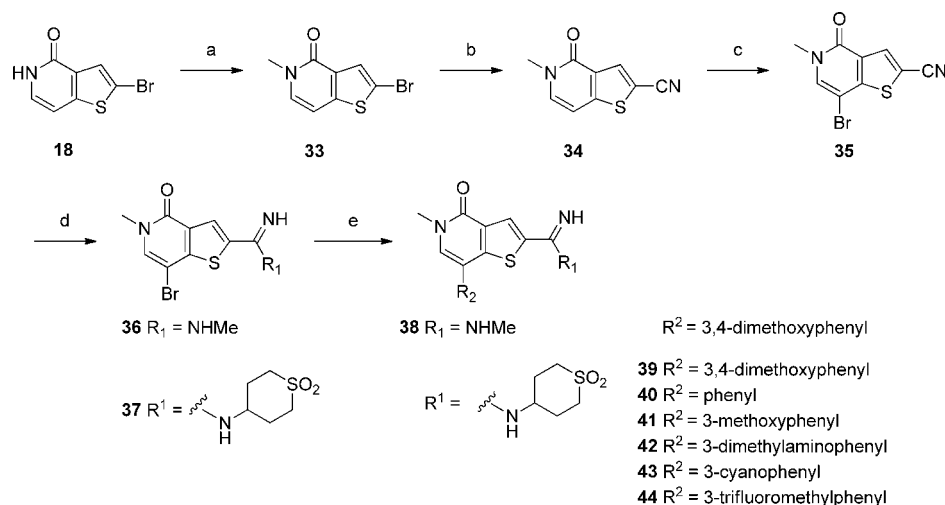
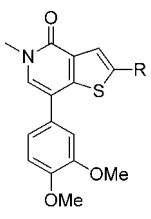


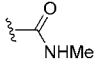
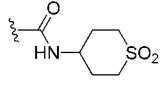
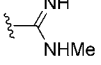
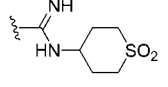
Figure 5. (A) X-ray crystal structures of BRD9 with 32 (orange) and 17 (green). (B) X-ray crystal structure of 32 in the bromodomains of BRD9 (orange) and BRD4 BD1 (blue). Hydrogen bonds are shown as gray dashed lines and water molecules as red spheres.

bromodomains (Figure 5B). In contrast to the situation for 17 described above, where different thiophene-amide bond torsion angles in each bromodomain give rise to a selectivity window, the

Scheme 2. Synthesis of Amidines 38–44^a

^aReagents and conditions: (a) MeI, Cs₂CO₃, THF, rt, quant.; (b) Zn(CN)₂, Pd(PPh₃)₄, DMF, microwave, 115 °C, 68%; (c) NBS, THF, rt, 86%; (d) NaOMe, MeOH, 75 °C, then amine hydrochloride salt, 87%–quant.; (e) R²B(OH)₂, K₂CO₃, PEPPSI-^tPr, IPA, H₂O, microwave, 120 °C, 4–52%.

Table 3. SAR for BRD9 and BRD4 BD1 Activity of Thienopyridone Amidines and Their Amide Analogues^a


Cmpd	R =	BRD9 pIC ₅₀	BRD4 BD1 pIC ₅₀	Fold Selectivity
29		6.9 ± 0.12	6.7 ± 0.16	×2
32		7.9 ± 0.09	7.3 ± 0.06	×4
38		7.1 ± 0.13	5.9 ± 0.11	×16
39		8.1 ± 0.19	6.4 ± 0.02	×50

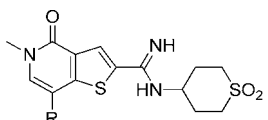
^aAll results are $n \geq 3$ as determined by TR-FRET.

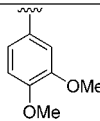

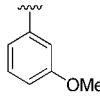
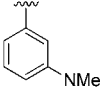
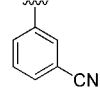
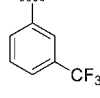
thiophene and amide of **32** are coplanar when bound to either bromodomain. As a result, the sulfone of **32** is able to form a hydrogen bond in each case, to the corresponding backbone NH atoms of Arg101 in BRD9 and Lys141 in BRD4. The similar geometry and interactions in each bromodomain account for the relatively small selectivity window between BRD9 and BRD4.

Despite the low selectivity of **32**, the BRD9 potency encouraged further effort to widen the selectivity window. One approach suggested by the X-ray crystal structure of **32** in complex with the bromodomain of BRD9 arose from the observation that the amide carbonyl lies close (within 5 Å) to the backbone carbonyl group of Ile53 (Figure 5A). Although the vectors appeared challenging, it was hypothesized that the replacement of the amide carbonyl group with a hydrogen-bond

donor moiety, such as an amidine, could increase activity, either through a direct interaction with the carbonyl of Ile53 or via a through-water hydrogen-bond. Furthermore, it was postulated that selectivity could be gained through exploitation of a key residue difference between BRD9 and BRD4 BD1. Considering the basic nature of the amidine moiety, it was proposed that when charged it would sit more favorably in the less hydrophobic environment alongside Ala54 of BRD9 than beside Leu94 of BRD4 BD1 (Figure 5B). To this end, the amidine analogues of secondary amides **29** and **32** were designed and accessed (Scheme 2).

N-Methylation of known thienopyridone **18**⁵⁴ afforded intermediate **33**. Following this, a palladium catalyzed Negishi cyanation gave nitrile **34**. Subsequent bromination at the 7-

Table 4. SAR for BRD9 and BRD4 BD1 Activity of Thienopyridone Amidines^a


Cmpd	R =	BRD9 pIC ₅₀	BRD4 BD1 pIC ₅₀	Fold Selectivity
39		8.1 ± 0.19	6.4 ± 0.02	×50
40		7.4 ± 0.13	5.8 ± 0.02	×40
41		7.9 ± 0.12	6.1 ± 0.06	×60
42		8.0 ± 0.23	6.3 ± 0.06	×50
43		7.4 ± 0.12	5.6 ± 0.04	×60
44		7.8 ± 0.21	5.6 ± 0.03	×160

^aAll results are $n \geq 3$ as determined by TR-FRET.

position of the thienopyridone core with NBS provided **35**. Conversion of the nitrile moiety to substituted amidines **36** and **37** was achieved via a Pinner reaction, using NaOMe in MeOH, followed by addition of the appropriate amine.⁵⁵ With these key intermediates in hand, a Suzuki–Miyaura coupling was utilized to install the different aryl motifs, employing the appropriate boronic acid.

Pleasingly, amidines **38** and **39** retained the BRD9 activity of their direct amide analogues **29** and **32**, respectively, with improved levels of selectivity over BRD4 BD1 (Table 3). Transformation of methyl amide **29** to amidine **38** produced a substantial increase in selectivity from 2 to 16-fold. In addition, amidine **39** was 50-fold selective over BRD4, an improvement on the four-fold seen for its amide analogue, **32**.

These exciting results provided confidence in our hypothesis that the charged amidine was favored in the less hydrophobic environment of BRD9. With this in mind, further work was conducted in order to gain additional selectivity. The 7-position of the thienopyridone core was selected as an appropriate vector for modification, given its location in the ZA channel. Reaction of advanced intermediate **37** with various boronic acids provided 7-aryl substituted compounds containing both electron-donating and electron-withdrawing substituents (Scheme 2).

Gratifyingly, all amidines (**39**–**44**) delivered ≥ 40 -fold selectivity over the first bromodomain of BRD4 (Table 4). These data indicated that both the amidine and phenyl moieties were important for selectivity, with the electronics of the phenyl ring substituents perhaps also playing a role. Unfunctionalized

phenyl substituted compound **40** and other electron-donating substituents such as **41** and **42** delivered no significant improvement in selectivity over BRD4 BD1 compared to dimethoxyphenyl derivative **39**. However, greater selectivity resulted from replacement of the electron-donating group by electron-withdrawing substituents. Nitrile substituted compound **43** delivered a 60-fold selectivity window, while the trifluoromethyl group of compound **44** provided a 160-fold window, satisfying the BET selectivity⁵⁶ and BRD9 potency probe criteria we defined at the outset of this project.

X-ray crystal structures of **44** in the bromodomains of BRD9 and BRD4 BD1 provided insights into the interactions responsible for the observed selectivity and supported our hypothesis that led to the synthesis of the amidine. Figure 6A shows the binding mode in BRD9 of **44** compared to that of **32**. Altering the substituents of the ZA-channel phenyl ring from 3,4-dimethoxy to 3-trifluoromethyl makes essentially no difference to the observed binding mode of the core of the molecule. The subtle change of the amide of **32** to the amidine of **44** maintains its hydrogen-bond to the conserved Asn100 side chain (Figure 6A) but leads to more notable changes that suggest significant electrostatic differences between the two molecules. The stronger electrostatics of the amidine NH group allow it to move closer to the backbone carbonyl of Ile53 than was possible with the carbonyl of the amide, resulting in a direct hydrogen-bond between these groups as we had hoped (Figure 6A). One result of this is a $\sim 30^\circ$ rotation of the thiophene–amidine bond of **44**. This allows the tetrahydro-2H-thiopyran 1,1-dioxide

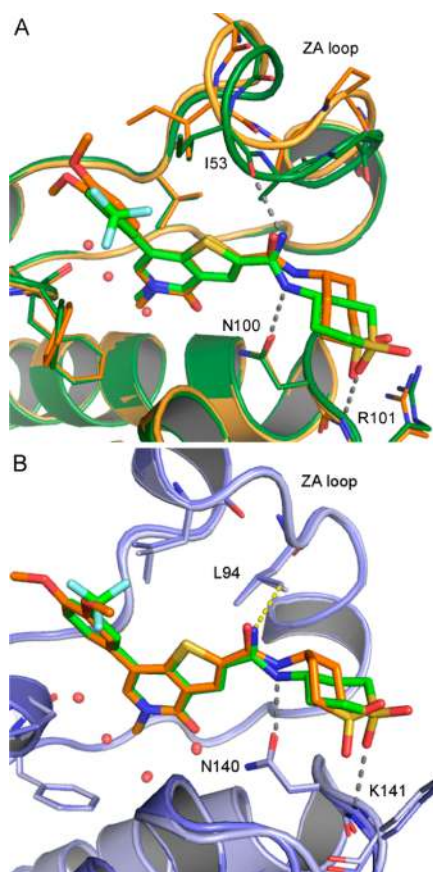


Figure 6. X-ray structure of (A) **32** (orange) and **44** (green) in BRD9, showing the rotation around the thiophene–amidine bond of **44**. The significant difference in conformation of the ZA loop that brings the carbonyl group of Ile53 close to the amidine of **44** is also visible. (B) Compounds **32** (orange) and **44** (green) in BRD4 BD1 with the 3.6 Å distance between the amidine nitrogen and Leu94 shown as a yellow dashed line.

amide substituent to resume its equatorial geometry while maintaining the hydrogen bond between the cyclic sulfone oxygen and the backbone NH of Arg101. The combined effect allows both Tyr99 and residues 50–56 of the ZA loop to relax to a position similar to that seen in the complex with **17**, rather than the conformation seen with **32**.

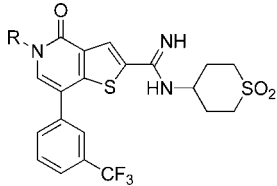
In contrast, the binding mode of **44** in BRD4 BD1 differs little from that of **32**, with no movement of the ZA loop (Figure 6B). A very slight movement of the amidine closer to Asn140 probably reflects a tighter charged hydrogen-bonding interaction. The best fit to the density of the tetrahydro-2*H*-thiopyran 1,1-dioxide amide substituent has equatorial geometry, but this maintains the hydrogen bond between cyclic sulfone and the backbone NH of Lys141 (Figure S2, Supporting Information). An explanation for the reduced BRD4 potency of **44** with respect to **32** is our original hypothesis that a charged amidine group in close proximity to the hydrophobic BRD4 Leu94 side chain is unfavorable (in the X-ray structure, the distance is 3.6 Å). The basic nature of the amidine functionality was confirmed by measurement: the pK_a of **44** was determined to be 8.4, so the group would be protonated at neutral pH.

Given the promising activity and selectivity profile displayed by compound **44**, this molecule was taken forward for further structure–activity relationship (SAR) exploration. It has previously been reported that Kac mimetics containing alkyl

groups larger than a methyl can be accommodated in the bromodomain binding pocket.^{29,57} With this in mind, we sought to investigate the effect of alternative Kac mimetic alkyl groups. To this end, *N*-ethyl analogue **45** and *N*-isopropyl analogue **46** were synthesized via a similar route to that described in Scheme 2.⁵⁸

Increasing the Kac mimetic alkyl chain length from *N*-methyl to *N*-ethyl maintained ≥ 100 -fold selectivity over BRD4 BD1 (Table 5). However, the introduction of the more sterically

Table 5. SAR for BRD9 and BRD4 BD1 Activity of 3-Trifluoromethylphenyl Substituted Thienopyridone Amidines^a



44, R = Me
45, R = Et
46, R = *i*-Pr

Cmpd	BRD9 pIC ₅₀	BRD4 BD1 pIC ₅₀	Fold Selectivity
44	7.8 ± 0.21	5.6 ± 0.03	×160
45	7.3 ± 0.18	5.3 ± 0.07	×100
46	5.5 ± 0.26	<4.3	≥×15

^aAll results are $n \geq 3$ as determined by TR-FRET analysis.

demanding isopropyl substituent, as in compound **46**, reduced BRD9 and BRD4 activities, suggesting that extending beyond *N*-ethyl is not well tolerated in either of these bromodomains (compare compounds **44** and **46**). X-ray crystallography of compound **45** in complex with BRD9 was obtained in order to confirm its binding mode. As expected, **45** adopts a similar conformation to compound **44** (Figure 7A), making identical interactions to Asn100, Ile53, and Arg101. The *N*-ethyl Kac mimetic is comfortably accommodated in the hydrophobic binding pocket with a slight movement of Phe45, having little effect on the conformation of the remainder of the site.

In order to assess selectivity over non-BET bromodomains, compounds **44** and **45** were profiled in a selection of internal TR-FRET assays, the results of which indicated that compound **45** displayed a superior selectivity profile. Considering this result, compound **45** was progressed for BROMOscan³⁹ profiling against 34 bromodomains to evaluate the effect of the *N*-ethyl Kac mimetic (Figure 7B). Pleasingly, the results from this broader bromodomain screen indicated nanomolar affinity binding at BRD9, with $pK_d = 8.7$ (for full data, see Supporting Information). Compound **45** showed >700-fold selectivity over the BET family of bromodomains and 200-fold over the highly homologous BRD7 bromodomain ($pK_d = 6.4$). Furthermore, **45** displayed >70-fold selectivity against every other bromodomain tested.

The selectivity of compound **45** over BRD7 is particularly surprising considering the 85% sequence homology between the bromodomains of BRD9 and BRD7 (Table 1). Although an NMR structure of BRD7 has been published,⁶⁰ there is no crystal structure reported to date. Therefore, it was challenging to rationalize the observed selectivity over this bromodomain. Presumably, selectivity arises from the differences in the constituent amino acids of the WPF shelf (BRD4 nomenclature, see Table 1) and ZA channels of these bromodomains. In BRD9, Gly43 is replaced by Ala154 in the WPF region of BRD7, and Ala46, by Ser157 in the ZA channel. These key changes result in a

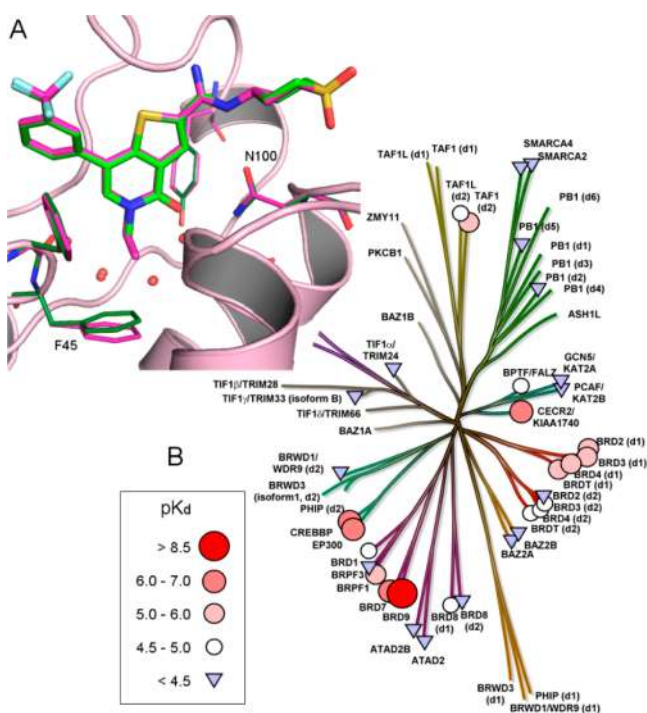


Figure 7. (A) Superimposed X-ray crystal structures of the bromodomain of BRD9 complexed with compounds 45 (magenta) and 44 (green), showing the subtle movement of Phe45 in response to the N1-methyl/ethyl difference between the two compounds. (B) BROMOScan data for compound 45 (generated at DiscoverX Corp.).

difference in the architecture of these binding regions, potentially altering the conformation of other residues in the ZA channel.

The observed potency was higher in the BROMOScan format than in house TR-FRET assays. This may be due to use of the alternative detection system, protein construct, and sample preparation methods. As the rank order is maintained between formats, numerical differences in the reported affinities did not affect decision making.

Based on the high BRD9 affinity and excellent broader bromodomain selectivity results, compound 45 (I-BRD9), was chosen as the chemical probe for BRD9.

As all measurements of binding affinity of I-BRD9 to date had been carried out with truncated bromodomain proteins, we were keen to confirm these findings were consistent with full-length targets in their native context. Using a chemoproteomic competition binding assay in HUT-78 cell lysate, the binding of I-BRD9 to endogenous BRD9 displayed >625-fold selectivity against BET family member BRD3 (Figure 8A).⁵⁸ These data confirm potency at BRD9, and selectivity over the BET family is maintained with endogenous proteins.

Critically for a bromodomain probe, cellular target engagement of BRD9 and disruption of chromatin binding was demonstrated through a NanoBRET assay measuring displacement of NanoLuc-tagged BRD9 bromodomain from Halo-tagged histone H3.3 (Figure 8B).⁵⁸

Having established endogenous protein binding, cell and nuclear permeability, and excellent selectivity over the BET family and other non-BET bromodomains, further profiling of I-BRD9 was conducted. These experiments aimed to evaluate the selectivity of the compound over a broad range of pharmaco-

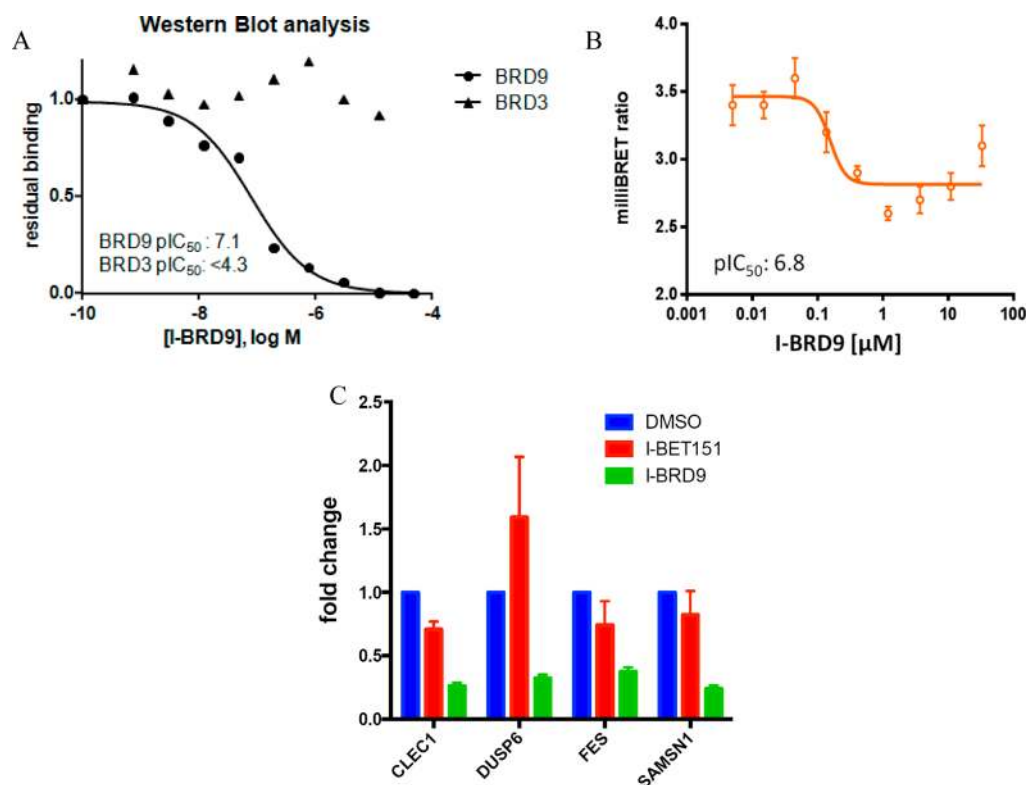
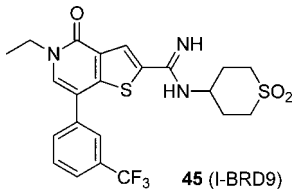


Figure 8. (A) Dose–response binding of compound 45 (I-BRD9) for endogenous BRD9 and BRD3 from HuT-78 cell lysates, measured in a chemoproteomic competition binding assay followed by Western blot analysis. (B) BRD9 bromodomain cellular NanoBRET dose–response curve of compound 45 (I-BRD9). (C) qPCR validation of CLEC1, DUSP6, FES, and SAMS1 genes selectively regulated by compound 45 (I-BRD9) (10 μM), but not by I-BET151 (1 μM), mean ± SD; *n* = 3. Genes were previously identified by full gene transcriptomics in Kasumi-1 cells.

logical targets including various receptors, transporters, ion channels, kinases, and other enzymes. Pleasingly, I-BRD9 showed no activity at less than 5 μM against a panel of 49 targets.⁵⁸ A summary of the properties of I-BRD9 is given in Table 6.

Table 6. Summary of Properties of I-BRD9

		45 (I-BRD9)
BRD9 pIC ₅₀ (TR-FRET)		7.3
BRD4 BD1 pIC ₅₀ (TR-FRET)		5.3
BRD9 NanoBRET pIC ₅₀		6.8
BROMOscan selectivity over other bromodomains ⁵⁸	BETs: > \times 700 BRD7: \times 200 all others: > \times 70	
ChromLogD _{pH7.4} ⁶¹		3.7
aqueous solubility (CLND)		359 μM (high)
artificial membrane permeability		210 nm/s (high)

With a high quality soluble and permeable bromodomain probe in hand, we sought to investigate genes selectively modulated by BRD9 bromodomain inhibition. Accordingly, Kasumi-1 cells were treated with I-BET151^{16–18} and I-BRD9 for 6 h, the concentrations of which were chosen to pharmacologically match the BET activity of the two probes. RNA was harvested, reverse transcribed, and hybridized on an Illumina expression array. While there was an expected overlap, the majority of genes were selectively regulated by I-BRD9.⁶² Analysis of four representative genes (CLEC1,⁶³ DUSP6,⁶⁴ FES,⁶⁵ and SAMSN1⁶⁶) that were strongly down-regulated by I-BRD9 and not by I-BET151, by qPCR further validated the microarray data and gave insights into the effect of BRD9 inhibition (Figure 8C). These four genes are implicated in cancer and immunology pathways, suggesting for the first time a role for the BRD9 bromodomain in their regulation. This down regulation also implies potential utility for synergistic disease modulation by BRD9 inhibition: for example, DUSP6 expression has been implicated in modulating drug sensitivity in cancers, which could be important for chemotherapy.⁶⁴

CONCLUSION

Through several iterations of structure-based design, starting from tertiary amide 17, we have identified I-BRD9 as the first selective cellular chemical probe for the non-BET bromodomain BRD9. This key contribution provides for the first time the ability to selectively target BRD9 and further enhances the tool set available for targeting human bromodomains. Critical to the success of this endeavor was the selectivity enhancing amidine, which drove unfavorable interactions between I-BRD9 and the BET family. This optimized compound is highly potent at BRD9 and displays >700-fold selectivity over the BET family and >70-fold against the panel of 34 bromodomains tested. It is of note that I-BRD9 is selective for a single bromodomain, whereas the vast majority of bromodomain probes bind to ≥ 2 . With proof of bromodomain cellular target engagement, binding to endogenous BRD9, and broader selectivity against a variety of pharmacological targets, we believe this molecule satisfies the

requirements necessary for a high quality chemical probe. I-BRD9 was used in the identification of a number of BRD9 inhibitor sensitive genes, many of which are involved in immune function and cancer, which might indicate the biological role of the BRD9 bromodomain, and provides insight and direction for further investigation.

EXPERIMENTAL SECTION

All solvents were purchased from Sigma-Aldrich (Hy-Dry anhydrous solvents), and commercially available reagents were used as received. All reactions were followed by TLC analysis (TLC plates GF254, Merck) or liquid chromatography mass spectrometry (LCMS) using a Waters ZQ instrument. IR spectra were obtained on a PerkinElmer Spectrum 1 machine. Melting points were measured with a Stuart SMP10, 230 V apparatus and were uncorrected.

NMR spectra were recorded at ambient temperature using standard pulse methods on any of the following spectrometers and signal frequencies: Bruker AV-400 (¹H = 400 MHz, ¹³C = 101 MHz) and Bruker AV-500 (¹H = 500 MHz, ¹³C = 125 MHz). Chemical shifts are reported in ppm and are referenced to tetramethylsilane (TMS) or the following solvent peaks: CDCl₃ (¹H = 7.27 ppm, ¹³C = 77.00 ppm), DMSO-*d*₆ (¹H = 2.50 ppm, ¹³C = 39.51 ppm), and MeOH-*d*₄ (¹H = 3.31 ppm, ¹³C = 49.15 ppm). Coupling constants are quoted to the nearest 0.1 Hz, and multiplicities are given by the following abbreviations and combinations thereof: s (singlet), d (doublet), t (triplet), q (quartet), m (multiplet), br (broad). Column chromatography was performed on prepacked silica gel columns (30–90 mesh, IST) using a biotage SP4. For detailed LCMS/MDAP/HRMS methodology, see Supplementary Methods.

The purity of all compounds tested was determined by LCMS and ¹H NMR to be >95%.

Selected Experimental Procedures. *2-Bromo-5-ethylthieno[3,2-*c*]pyridin-4(5*H*)-one (47)*. A single portion of ethyl iodide (1.40 mL, 17.39 mmol) was added to a solution of bromothieno[3,2-*c*]pyridin-4(5*H*)-one (18)⁵⁴ (2.00 g, 8.69 mmol) and Cs₂CO₃ (8.50 g, 26.10 mmol) in THF (35 mL) at room temperature. The reaction mixture was heated at 60 °C for 18 h, then allowed to cool to room temperature and diluted with EtOAc (50 mL) and H₂O (50 mL). The separated aqueous phase was extracted with EtOAc (3 \times 50 mL), and the combined organic layers were passed through a hydrophobic frit and concentrated *in vacuo* to give 47 (2.30 g, quant.) as a light brown solid. Fifty milligrams of 47 was taken forward for purification by mass directed autopreparation (formic acid), providing suitably pure material (44 mg, 0.17 mmol) for full characterization; mp 124–125 °C; ν_{max} (solid)/cm⁻¹ 1632, 1577, 1082, 846, 761; ¹H NMR (500 MHz, (CD₃)₂SO) δ ppm 7.62 (s, 1H), 7.56 (d, *J* = 7.2, 1H), 6.89 (d, *J* = 7.2, 1H), 3.99 (q, *J* = 7.2, 2H), 1.24 (t, *J* = 7.2, 3H); ¹³C NMR (125 MHz, (CD₃)₂SO) δ ppm 156.4, 148.4, 133.8, 130.2, 127.0, 111.7, 100.8, 43.3, 14.6; HRMS (M + H)⁺ calculated for C₉H₉BrNOS 257.9583; found 257.9585. LCMS (formic acid) (M + H)⁺ = 258.0, 260.0, R_t = 0.89 min (98%).

*5-Ethyl-4-oxo-4,5-dihydrothieno[3,2-*c*]pyridine-2-carbonitrile (48)*. A mixture of 47 (1.10 g, 4.26 mmol), zinc cyanide (1.00 g, 8.52 mmol), and Pd(PPh₃)₄ (0.44 g, 0.3 mmol) in DMF (10 mL) was heated to 115 °C in a microwave reactor for 3.5 h. This process was repeated to provide a second identical batch. The two batches were allowed to cool to room temperature, diluted with EtOAc (50 mL each), and combined. The solution was filtered through Celite and concentrated *in vacuo*. The resulting residue was purified by silica gel chromatography (0–3% MeOH in CH₂Cl₂). The appropriate fractions were combined and concentrated *in vacuo* to give 48 (1.70 g, 67%) as a light brown solid. Fifty milligrams of 48 was taken forward for purification by mass directed autopreparation (formic acid), providing suitably pure material (33 mg, 0.16 mmol) for full characterization; mp 157–158 °C; ν_{max} (solid)/cm⁻¹ 2211, 1637, 1595, 1247, 1078, 879, 780, 769; ¹H NMR (400 MHz, (CD₃)₂SO) δ ppm 8.32 (s, 1H), 7.82 (d, *J* = 7.2, 1H), 7.03 (d, *J* = 7.2, 1H), 4.00 (q, *J* = 7.2, 2H), 1.25 (t, *J* = 7.2, 3H); ¹³C NMR (100 MHz, (CD₃)₂SO) δ ppm 157.1, 151.1, 137.1, 136.7, 128.9, 114.2, 106.4, 100.9, 43.4, 14.5; HRMS (M + H)⁺ calculated for C₁₀H₉N₂OS

205.0430; found 205.0429. LCMS (formic acid) (M + H)⁺ = 205.0, R_t = 0.73 min (98%).

7-Bromo-5-ethyl-4-oxo-4,5-dihydrothieno[3,2-c]pyridine-2-carbonitrile (49). To a solution of **48** (1.17 g, 5.73 mmol) in THF (20 mL), NBS (1.53 g, 8.59 mmol) was added portion-wise. The reaction mixture was left to stir at room temperature for 48 h. The volatile components were removed *in vacuo*. The resulting solid was suspended in Et₂O (50 mL), filtered under reduced pressure, washed with Et₂O (50 mL), collected, and dried under vacuum at 40 °C to give **49** (1.69 g, quant.) as an off-white solid; mp 197–198 °C; ν_{\max} (solid)/cm⁻¹ 3041, 2220, 1654, 1584, 1250, 1090, 766; ¹H NMR (500 MHz, (CD₃)₂SO) δ ppm 8.50 (s, 1H), 8.26 (s, 1H), 4.02 (q, J = 7.2, 2H), 1.26 (t, J = 7.2, 3H); ¹³C NMR (125 MHz, (CD₃)₂SO) δ ppm 156.1, 152.3, 138.2, 137.6, 128.3, 113.7, 107.0, 90.4, 43.9, 14.4; HRMS (M + H)⁺ calculated for C₁₀H₈BrN₂OS 282.9535; found 282.9534. LCMS (formic acid) (M + H)⁺ = 283.0, 285.0, R_t = 0.93 min (100%).

7-Bromo-N-(1,1-dioxidotetrahydro-2H-thiopyran-4-yl)-5-ethyl-4-oxo-4,5-dihydrothieno[3,2-c]pyridine-2-carboximidamide (50). To a suspension of **49** (1.00 g, 3.53 mmol) in MeOH (30 mL) at room temperature, sodium methoxide (25% weight in MeOH, 0.81 mL, 3.53 mmol) was added in a single portion. The reaction mixture was heated at 75 °C for 5 h. 4-Aminotetrahydro-2H-thiopyran-1,1-dioxide hydrochloride (1.98 g, 10.60 mmol) was added, and the solution was heated for a further 18 h. The volatile components were removed *in vacuo*, and the resulting solid was suspended in Et₂O (50 mL), filtered under reduced pressure, washed with Et₂O (50 mL), collected, and dried under vacuum at 40 °C to give **50** (1.55 g, quant.) as a white solid. Fifty milligrams of **50** was taken forward for purification by mass directed autopreparation (high pH), providing suitably pure material (25 mg, 0.06 mmol) for full characterization; mp 284–285 °C; ν_{\max} (solid)/cm⁻¹ 1646, 1578, 1284, 1124, 853, 769; ¹H NMR (500 MHz, (CD₃)₂SO) δ ppm 8.18 (s, 1H), 8.01 (s, 1H), 6.80 (br.s, 2H), 4.01 (q, J = 7.2, 2H), 3.68–3.61 (m, 1H), 3.27–3.20 (m, 2H), 3.10–3.03 (m, 2H), 2.09–2.00 (m, 2H), 1.99–1.90 (m, 2H), 1.26 (t, J = 7.2, 3H); ¹³C NMR (125 MHz, (CD₃)₂SO) δ ppm 156.9, 149.5, 134.6, 129.7, 123.2, 109.7, 92.0, 48.6, 43.7, 30.2, 14.8; HRMS (M + H)⁺ calculated for C₁₅H₁₉BrN₃O₃S₂ 432.0046; found 432.0046. LCMS (formic acid) (M + H)⁺ = 432.1, 434.1, R_t = 0.53 min (100%).

N-(1,1-Dioxidotetrahydro-2H-thiopyran-4-yl)-5-ethyl-4-oxo-7-(3-(trifluoromethyl)phenyl)-4,5-dihydrothieno[3,2-c]pyridine-2-carboximidamide (45, I-BRD9). A mixture of **50** (150 mg, 0.35 mmol), (3-(trifluoromethyl)phenyl)boronic acid (79 mg, 0.42 mmol), PEPPSI-^tPr (21 mg, 0.03 mmol), and K₂CO₃ (115 mg, 0.83 mmol) in H₂O (1 mL) and IPA (3 mL) was heated at 120 °C in a microwave reactor for 0.5 h. The reaction mixture was allowed to cool to room temperature, diluted with EtOAc (40 mL), filtered through Celite, and concentrated *in vacuo*. The resulting residue was purified by mass directed autopreparation (formic acid). The appropriate fractions were combined and concentrated *in vacuo*. The resulting residue was dissolved in MeOH (5 mL) and passed through a preconditioned (MeOH, 20 mL) amino propyl column (10 g). The appropriate fractions were combined and concentrated *in vacuo* to give **45** (50 mg, 29%) as a white solid; mp 228–229 °C; ν_{\max} (solid)/cm⁻¹ 1640, 1587, 1340, 1303, 1117; ¹H NMR (500 MHz, (CD₃)₂SO) δ ppm 8.15 (s, 1H), 8.03–7.98 (m, 1H), 7.98–7.94 (m, 1H), 7.91 (s, 1H), 7.82–7.76 (m, 2H), 6.76 (br.s, 2H), 4.10 (q, J = 7.0, 2H), 3.64–3.57 (m, 1H), 3.21–3.12 (m, 2H), 3.10–3.00 (m, 2H), 2.06–1.96 (m, 2H), 1.96–1.85 (m, 2H), 1.32 (t, J = 7.0, 3H); ¹³C NMR (125 MHz, (CD₃)₂SO) δ ppm 157.6, 149.7, 147.7, 142.2, 137.5, 134.1, 131.9, 130.7, 130.5, 130.3 (q, J_{C-F} = 31.7), 125.0 (q, J_{C-F} = 3.7), 124.60 (q, J_{C-F} = 3.7), 124.59 (q, J_{C-F} = 272.6), 123.1, 114.4, 49.9, 49.0, 44.0, 30.6, 15.2; ¹⁹F NMR (376 MHz, (CD₃)₂SO) δ ppm –61.0; HRMS (M + H)⁺ calculated for C₂₂H₂₃F₃N₃O₃S₂ 498.1128; found 498.1110. LCMS (formic acid) (M + H)⁺ = 498.1, R_t = 0.81 min (100%).

■ ASSOCIATED CONTENT

Supporting Information

Additional text describing all biological methods and results, chemistry experimental procedures, full BROMOScan and selectivity data for I-BRD9, X-ray data collection and refinement

statistics, and computed thienopyridone amide torsional energy profiles. The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acs.jmedchem.5b00256.

Accession Codes

Atomic coordinates of BD4 BD1 bound to compounds **17** (4uiz), **32** (4uix), **44** (4uiy) and BRD9 bound to compounds **17** (4uit), **32** (4uiu), **44** (4uiv) and **45** (4uiw) have been deposited with the Protein Data Bank.

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Notes

The authors declare the following competing financial interest(s): All authors except N.H.T., N.C.O.T., H.K.C., A.B., S.C.R. and T.K. were GlaxoSmithKline and Cellzome full-time employees at the time when this study was performed.

■ ACKNOWLEDGMENTS

N.H.T. is grateful to GlaxoSmithKline R&D, Stevenage for Ph.D. studentship funding. We thank Jacqui Méndez and Danette Daniels of Promega Corporation for facilitating the NanoBRET assay. We also thank Robert Watson and Darren Mitchell for the synthesis of useful building blocks; Matthew Crane for the synthesis of compound **32**; Iain Reid for pK_a determination; Sean Lynn for assistance with NMR; Marco Smith for high resolution mass spectrometry, and Paul White, Keir Sims, and Abigail Lucas for crystallization support. We appreciate helpful discussions with Emmanuel Demont during the course of this work.

■ ABBREVIATIONS USED

BAF, BRG1/brm-associated factor; BAZ2A, bromodomain adjacent to zinc finger domain, 2A; BAZ2B, bromodomain adjacent to zinc finger domain, 2B; BCP, bromodomain containing protein; BD, bromodomain; BET, bromodomain and extra terminal domain; BRD2, bromodomain containing protein 2; BRD3, bromodomain containing protein 3; BRD4, bromodomain containing protein 4; BRD7, bromodomain containing protein 7; BRD9, bromodomain containing protein 9; BRDT, bromodomain containing protein, testis-specific; CECR2, cat eye syndrome chromosome region, candidate 2; CREBBP, cAMP response element-binding protein binding protein; DIPEA, N,N-diisopropylethylamine; EP300, E1A binding protein p300; HAT, histone acetyltransferase; HATU, O-(7-azabenzotriazol-1-yl)-N,N,N',N'-tetramethyluronium hexafluorophosphate; HDAC, histone deacetylase; IPA, isopropanol; Kac, acetyl lysine; LCMS, liquid column mass spectrometry; LPS, lipopolysaccharide; PB1(5), PolyBromo 1 (5); PCAF, P300/CREBBP-associated factor; PEPPSI-^tPr, pyridine-enhanced precatalyst preparation stabilization and initiation; PK, pharmacokinetics; SMARCA2, SWI/SNF related, matrix associated, actin dependent regulator of chromatin, subfamily A, member 2; SMARCA4, SWI/SNF related, matrix associated, actin-dependent regulator of chromatin, subfamily A, member 4; SWI/SNF, SWI/SNF/sucrose nonfermentable; TR-FRET, time-resolved Förster (fluorescence) resonance energy transfer

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