Diverse novel functions of neutrophils in immunity, inflammation, and beyond

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Neutrophils have long been considered simple suicide killers at the bottom of the hierarchy of the immune response. That view began to change 10-20 yr ago, when the sophisticated mechanisms behind how neutrophils locate and eliminate pathogens and regulate immunity and inflammation were discovered. The last few years witnessed a new wave of discoveries about additional novel and unexpected functions of these cells. Neutrophils have been proposed to participate in protection against intracellular pathogens such as viruses and mycobacteria. They have been shown to intimately shape the adaptive immune response at various levels, including marginal zone B cells, plasmacytoid dendritic cells and T cell populations, and even to control NK cell homeostasis. Neutrophils have been shown to mediate an alternative pathway of systemic anaphylaxis and to participate in allergic skin reactions. Finally, neutrophils were found to be involved in physiological and pathological processes beyond the immune system, such as diabetes, atherosclerosis, and thrombus formation. Many of those functions appear to be related to their unique ability to release neutrophil extracellular traps even in the absence of pathogens. This review summarizes those novel findings on versatile functions of neutrophils and how they change our view of neutrophil biology in health and disease.

Neutrophils (also known as neutrophilic granulocytes or polymorphonuclear leukocytes [PMNs]) are the most abundant white blood cells in the human circulation. They play a crucial role in the immune defense against bacterial and fungal pathogens, and they also participate in the development of the inflammatory reaction (Nathan, 2006). Although neutrophils are mostly viewed as playing a beneficial role to the host, their improper activation may also lead to tissue damage during an autoimmune or exaggerated inflammatory reaction (Nathan, 2006; Németh and Mócsai, 2012).

Neutrophils participate in antimicrobial host defense both as the first line of innate immune defense and as effectors of adaptive immunity. They are short-lived cells that usually die while performing their antimicrobial function. Because their primary role is the localization and elimination of invading microorganisms at any expense, a simplistic view of neutrophils being not more than "dumb" suicide killers has prevailed for a long time.

A major wave of discoveries during the 1990s and early 2000s made immunologists begin to appreciate the amazing complexity and sophistication of neutrophil functions. It became evident that neutrophils release cytokines and contribute to orchestrating the immune/inflammatory response (Bazzoni et al., 1991a,b; Cassatella, 1995). A highly sophisticated machinery directing neutrophil migration (Ley et al., 2007; Nourshargh et al., 2010) and a surprising complexity of neutrophil granules (Borregaard et al., 2007) also begun to emerge. Novel but controversial concepts about how neutrophils may fight microbes, including possible regulation of granule enzyme function by ion fluxes (Reeves et al., 2002) and formation of neutrophil extracellular traps (NETs; Brinkmann et al., 2004) were also proposed during that period. Both mechanisms have been subject to intense debate (see next section). Nevertheless, all those studies indicated that neutrophils use highly sophisticated and complex mechanisms to perform their role in immune defense and inflammation and led to improved and refined models of neutrophil biology (Witko-Sarsat

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et al., 2000; Nathan, 2006; Mantovani et al., 2011; Amulic et al., 2012; Fig. 1).

The last few years have borne witness to another wave of discoveries of novel and unexpected roles of neutrophils in diverse aspects of immunity and inflammation, as well as beyond the traditional scope of the immunological sciences. Those include novel roles of neutrophils in immunity against intracellular pathogens such as viruses and intracellular bacteria; shaping of adaptive immunity at different levels; and roles in disease states previously not associated with neutrophils, such as allergy and anaphylaxis, metabolic diseases, atherosclerosis, or thrombus formation. Those cases also revealed novel potential roles for NETs beyond antimicrobial functions. Many of those studies exploited approaches allowing the antibody-mediated depletion of neutrophils by anti-Gr1 or the more neutrophil-specific anti-Ly6G antibodies in mice (Daley et al., 2008), or genetic manipulations leading to the partial or complete genetic deletion of the neutrophil lineage (Liu et al., 1996; Karsunky et al., 2002; Hock et al., 2003; Jonsson et al., 2005; Ordoñez-Rueda et al., 2012). Though those novel studies have yet to pass the scrutiny of the scientific community, they suggest that neutrophils may have a much more widespread role in immunity, inflammation, and beyond than previously anticipated.

After a brief summary of established and well-known (though sometimes debated) neutrophil functions, this review will summarize the novel biological functions of neutrophils revealed in the last few years, within and beyond immunity and inflammation. I will primarily focus on what neutrophils do rather than how they do it. Readers will be referred to additional recent reviews on related subjects such as certain aspects of the molecular mechanisms of neutrophil functions.

Neutrophil functions: state of the art in the early 2000s

A tremendous amount of knowledge on neutrophils had been generated by the end of the 20th century and during the first years of the 2000s. The most crucial aspects of that knowledge, which forms the basis of our understanding of neutrophil biology, are briefly summarized in the following section and in Fig. 1.

Neutrophils are terminally differentiated cells that develop in the bone marrow where they are also stored for release into the circulation (Borregaard, 2010). During their maturation, neutrophils develop several intracellular compartments (secretory granules and vesicles) that store proteins critical for the antimicrobial and proinflammatory missions of the cell (Borregaard et al., 2007). Once released into the circulation, neutrophils begin to seek signs of infection and inflammation which lead to a series of events culminating in the migration of neutrophils through the vessel wall and inside the tissue to the site of microbial invasion (Ley et al., 2007; Nourshargh et al., 2010; Fig. 1).

When encountering infectious particles, neutrophils begin a professional antimicrobial killing program. The infectious particle will be phagocytosed (Nordenfelt and Tapper, 2011) and various noxious agents, including reactive oxygen species generated through the activity of the NADPH oxidase (Leto and Geiszt, 2006), and the contents of intracellular granules (Borregaard et al., 2007; Soehnlein, 2009) are then released into the phagosome, resulting in killing and digestion of the microorganisms (Fig. 1).

Besides the well-established role of phagocytosis, two additional mechanisms contributing to neutrophil-mediated antimicrobial responses have been proposed during the early

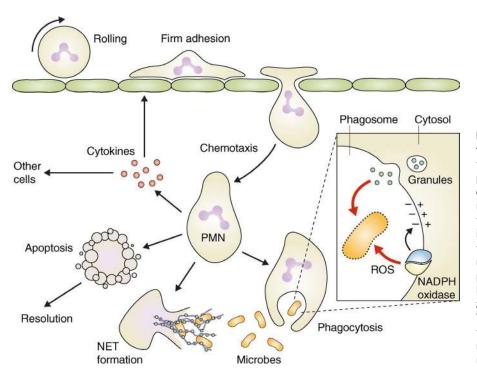


Figure 1. Neutrophil functions: state of the art in the early 2000s. After migrating to the site of inflammation, neutrophils (PMN) phagocytose and digest the invading microbes; release NETs, which likely trap bacteria; and produce cytokines, which contribute to the inflammatory reaction. Once infection is cleared, neutrophils die by apoptosis and trigger an active program to resolve inflammation. Inset, pathogen killing inside the phagosome occurs by ROS generated by the NADPH oxidase, as well as by granule enzymes released from intracellular granules. The NADPH oxidase also induces depolarization of the phagosomal membrane, which may be required for providing optimal environment inside the phagosome.

2000s, both of them triggering intensive scrutiny and debate from the scientific community. Anthony Segal's group proposed an interesting molecular interplay whereby ion fluxes triggered by the NADPH oxidase would promote the antimicrobial function of granule proteins (Reeves et al., 2002). Though many components of the proposed mechanism (such as the electrogenicity of the NADPH oxidase [Fig. 1, inset] providing driving force for other ion currents; Geiszt et al., 1997; Schrenzel et al., 1998; Rada et al., 2005) are generally accepted, it is still debated whether and to what extent ion fluxes that compensate for the activity of the NADPH oxidase contribute to the antibacterial effects of granule enzymes (Geiszt et al., 2001; Rada et al., 2004; Rada et al., 2005; Femling et al., 2006; Essin et al., 2009). Another interesting and similarly provocative idea was that, in addition to intraphagosomal killing, neutrophils may also be able to expel their nuclear content (including DNA) complexed with granule proteins into the extracellular space and that this complex (NETs) may inhibit microbial growth without phagocytosis of the pathogen (Brinkmann et al., 2004; Fig. 1). Though neutrophil-derived extracellular DNA has long been known to be present at the site of infection (in part as a component of pus formation) and NET formation has recently been directly visualized during bacterial infection in vivo (Yipp et al., 2012; McDonald et al., 2012), the contribution of NET formation to antibacterial host defense and killing is still under intensive debate (Brinkmann and Zychlinsky, 2012; Menegazzi et al., 2012; Nauseef, 2012; Parker et al., 2012a).

Neutrophils are short-lived cells that die within a limited time after entering the circulation (Geering and Simon, 2011). In the absence of infection or inflammation, they die by a spontaneous apoptosis program (Geering and Simon, 2011; Witko-Sarsat et al., 2011), likely within 1 d (although some investigators propose this time to be up to 5 d; Pillay et al., 2010). Inflammatory signals are capable of prolonging the lifespan of the cells by several days, during which they release inflammatory mediators and contribute to the orchestration of the inflammatory response (Mantovani et al., 2011; Fig. 1). However, even under those conditions, neutrophils will soon die by apoptosis (or, possibly, NETosis), contributing to pus formation inside infected tissues. Once neutrophils die, another program for removal of dead neutrophils by macrophages is initiated (Bratton and Henson, 2011). Parallel to the inflammatory response, biochemical and transcriptional changes leading to the resolution of the inflammation begin (Serhan, 2007; Serhan et al., 2008; Soehnlein and Lindbom, 2010), and this is further promoted by engulfment of the corpses of dead neutrophils by macrophages (El Kebir and Filep, 2010).

The role of neutrophils in antiviral host defense

Whereas neutrophils are critical for immunity against bacterial and fungal pathogens, their role in antiviral host defense is much less appreciated. In a recent study, Saitoh et al. (2012) proposed a role for NETs in the control of HIV-1 infection. HIV-1 triggered NET formation by human neutrophils and neutrophil-derived NETs were able to capture HIV-1 particles and reduce their efficiency to infect T cells. Most importantly, when neutrophils and T cells were co-cultured, neutrophils were able to reduce the infection efficiency of HIV-1. Pharmacological studies indicated that NET formation was dependent on TLR7 and TLR8 in endocytic compartments and that the effect of neutrophil-derived NETs on T cells could be inhibited by DNase treatment, underscoring the role of extracellular DNA. In agreement with those findings, the neutrophil-mediated inhibition of HIV-1 infectivity on T cells required TLR7 and TLR8, and it was suspended by DNase treatment.

Another recent study provided additional evidence for the role of neutrophils and NETs in antiviral host defense (Jenne et al., 2013). Systemic injection of viral TLR ligands or poxvirus infection led to dramatic accumulation of neutrophils in liver sinusoids that formed aggregates with platelets and released NETs inside the vessels. Importantly, DNase treatment increased the percentage of liver cells infected with poxviruses, suggesting that extracellular DNA exerts a protective effect against viral infection (Jenne et al., 2013).

Although the aforementioned studies suggest a role for neutrophils and NETs in antiviral host defense, they also raise several questions. Most studies on NET formation by neutrophils indicate a critical role for NADPH oxidase activity in that process (including the original NET study [Brinkmann et al., 2004] where NET formation was triggered by phorbol ester-induced activation of neutrophils) and it is difficult to envisage how viral particles would be able to trigger such a strong stimulation of neutrophils. However, because NET formation in the absence of NADPH oxidase activity has recently been shown to occur under milder and more physiological conditions (Chen et al., 2012; Parker et al., 2012b), it may be possible that even viral particles trigger NET formation. In addition, Jenne et al. (2013) used systemic administration of LPS to induce protection from viral infection. It is unclear whether that response induces specific activation of neutrophils and whether the DNase treatment used only affects neutrophil-derived NETs. Nevertheless, though additional studies in the field are warranted, these two studies (together with additional ones showing priming of virus-specific T cells by neutrophils; Duffy et al., 2012) open up new avenues on the role of neutrophils in the host defense against viral pathogens.

Neutrophils in infection with intracellular pathogens

The traditional view is that neutrophils eradicate extracellular bacteria and fungi, whereas intracellular pathogens such as mycobacteria are primarily dealt with by macrophages. Although prior studies indicated possible roles for neutrophils during mycobacterial infection (Appelberg, 2007; Lowe et al., 2012), recent studies provided important novel insights into the role of neutrophils in the fight against intracellular pathogens. Analysis of healthy human individuals who have been in contact with tuberculosis patients revealed strong inverse correlation between the risk of tuberculosis infection and

peripheral blood neutrophil counts (Martineau et al., 2007). The authors also showed that ex vivo growth of mycobacteria in whole blood was dramatically increased upon neutrophil depletion, whereas neutrophil-derived antimicrobial peptides inhibited mycobacterial growth from in vitro bacterial cultures (Martineau et al., 2007). Another recent study identified a unique blood transcriptional signature in human tuberculosis patients that closely matched a putative type I and type II interferon-induced gene expression signature in neutrophils (Berry et al., 2010). This signature was clearly distinguishable from those in other diseases characterized by a major neutrophil component (such as Gram-positive bacterial infections) or by a type I IFN signature (such as SLE). Those two studies suggested that neutrophils actively fight against mycobacteria and may provide protection from infection.

A possible explanation for those findings is that neutrophils fight mycobacteria during and immediately after their entry into the organism. However, this may only be true for the initial contact with mycobacteria when neutrophils are able to prevent infection of the host, while macrophages "take over" when the bacilli reach intracellular compartments (Lowe et al., 2012). Indeed, neutrophils likely play an opposite, pathogenic role during later stages of the disease, through at least two different mechanisms. First, neutrophils may serve as vectors for the systemic dissemination of the pathogens. Mouse neutrophils are able to acquire live mycobacteria at the site of infection and carry those microbes to the draining lymph nodes by migration within the lymphatic vasculature (similar to the migration of DCs in a general immune response; Abadie et al., 2005). Although this may promote antimicrobial host defense, an alternative scenario is that mycobacteria that escape killing and survive inside neutrophils use them as "Trojan horses" to help disseminate the bacteria to distant organs. This would be similar to the situation in leishmaniasis when neutrophils engulf parasites at the site of infection and deliver them to macrophages, which take up live Leishmania parasites during phagocytosis of apoptotic neutrophils (van Zandbergen et al., 2004; Peters et al., 2008; John and Hunter, 2008). The second mechanism for the contribution of neutrophils to the chronic phase of tuberculosis may be related to their role in infection-induced tissue damage. Cruz et al. (2010) suggested that excessive release of IL-17 during repeated mycobacterial exposure leads to neutrophil recruitment and neutrophil-mediated tissue damage in mice. This response appears to be attenuated by IFN- γ production during a normal antimycobacterial immune response (Nandi and Behar, 2011), indicating that neutrophil accumulation and the resulting tissue damage may be caused by the failure of the immune system to contain the infectious agents.

The dual role of neutrophils in tuberculosis is also reflected in another recent study, suggesting that neutrophils down-regulate inflammation at early stages of mycobacterial infection but contribute to inflammation and mycobacterial clearance during chronic tuberculosis in experimental mice (Zhang et al., 2009). The importance of those findings in human pathology has yet to be assessed, in part because it is uncertain whether IL-10, the cytokine proposed to be responsible for most of the reported findings in mice (Zhang et al., 2009), is also released by human neutrophils (Davey et al., 2011).

The latest twist in the role of neutrophils in tuberculosis comes from a recent study on zebrafish (Yang et al., 2012). By monitoring mycobacterial infection, Yang et al. (2012) found that neutrophils did not interact with the microbes during the very first phase of infection but engulfed dying infected macrophages during early granuloma formation and killed the surviving intracellular pathogens. That study suggests that efferocytosis (the engulfment of dying cells) not only serves to clean up the site of infection but also contributes to containing the infection, at least in the case of mycobacterial challenge.

Another unexpected role for neutrophils in infectious diseases has been revealed by a recent study on malaria pathogenesis (Porcherie et al., 2011). The authors showed that genetic deficiency of IgE- or FcERI-protected mice from cerebral malaria and death, but not parasitemia after infection with Plasmodium falciparum parasites. Interestingly, FcERI was dramatically up-regulated on circulating neutrophils after parasitic infection and FcERI-positive neutrophils became the dominant FcERI-positive leukocyte population in the brain. Importantly, depletion of neutrophils using the NIMP-R14 anti-Gr1 antibody protected mice both from neurological signs of cerebral malaria and from malaria-induced death, but not from parasitemia, suggesting an important role for neutrophils in malaria pathogenesis (Porcherie et al., 2011). Although the mechanism whereby neutrophils contribute to cerebral manifestations of malaria is incompletely understood, the authors suggested that production of proinflammatory cytokines (e.g., IFN- γ) by neutrophils accumulating in the brain of infected animals leads to excessive inflammation and subsequent neurological signs that are characteristic of cerebral malaria.

Collectively, the aforementioned studies suggest that neutrophils may have a much wider (protective or sometimes pathogenic) role during infectious diseases than previously appreciated.

Neutrophils in the regulation of B and T cells

Neutrophils have long been seen as effector cells that have little influence on shaping the adaptive immune response. That view is being challenged by several recent reports.

Marginal zone B cells are unique B cells of the spleen that are located outside the germinal centers and close to the marginal sinus, thus closely positioned to the blood circulation. Their primary function is the rapid initiation of T cellindependent antibody responses to blood-borne microbes. Interestingly, a prior study revealed that neutrophils are one of the major cell types responsible for capturing and transporting circulating bacteria to the splenic marginal zone in mice (Balázs et al., 2002). Additionally, prior human studies showed that activated (but not resting) neutrophils express the B cell–activating cytokine BAFF (also known as BLyS; Scapini et al., 2003) and that neutrophils are a major source of the BAFF-related, B cell-stimulating cytokine APRIL in B cell lymphomas (Schwaller et al., 2007) and mucosal plasma cell niches (Huard et al., 2008; Scapini et al., 2008). A recent work by Puga et al. (2011) revealed an intricate relationship between neutrophils and marginal zone B cells and proposed that this interaction mediates T cell-independent antibody responses through BAFF and APRIL (Fig. 2 A). The authors found a large number of neutrophils around the splenic marginal zone in humans, macaques, and mice. In mice, splenic but not circulating neutrophils were able to strongly activate marginal zone B cells, whereas they had limited stimulatory effect on follicular B cells and even inhibited CD4⁺ T cells. Importantly, splenic neutrophils promoted B cell survival, antibody production, class switching to IgG and IgA, and somatic hypermutation, indicating that they qualify to be bona fide B helper cells. Splenic, but not circulating, neutrophils express large amounts of BAFF, APRIL, and IL-21, and all those cytokines were required for the stimulatory effect on marginal zone B cells. Circulating neutrophils acquired the ability of activating marginal zone B cells upon exposure to splenic sinusoidal endothelial cells in the presence of microbial signals in vitro. The colonization of the splenic marginal zones by neutrophils primarily occurred soon after birth in humans, and marginal zone neutrophils were significantly less abundant in germ-free

mice than in normal specific pathogen-free animals, indicating that microbial products, likely originating from the commensal flora, play an important role in the accumulation of splenic marginal zone neutrophils. Importantly, human patients suffering from various congenital defects of neutrophil development or function had normal total circulating B cell numbers but strongly reduced numbers of circulating marginal zone B cells, and their serum levels of antibodies against T cell-independent antigens were significantly reduced despite normal levels of antibodies against T cell-dependent antigens. Collectively, the elegant study of Puga et al. (2011) revealed a potentially important immunological and immunehomeostatic mechanism whereby splenic neutrophils may facilitate the antibody response of marginal zone B cells to T cell-independent microbial antigens. This mechanism requires the reprogramming of circulating neutrophils by microbial signals, as well as the neutrophil-derived BAFF, APRIL, and IL-21 cytokines. Further studies will be required to determine the full importance of this mechanism in immune defense and to more closely define the nature and developmental stage of the proposed B cell helper neutrophil population relative to their circulating counterparts.

The role of neutrophils in the regulation of T cell function appears to be more complex and less well understood. Regulation of T cells by myeloid cells primarily occurs via

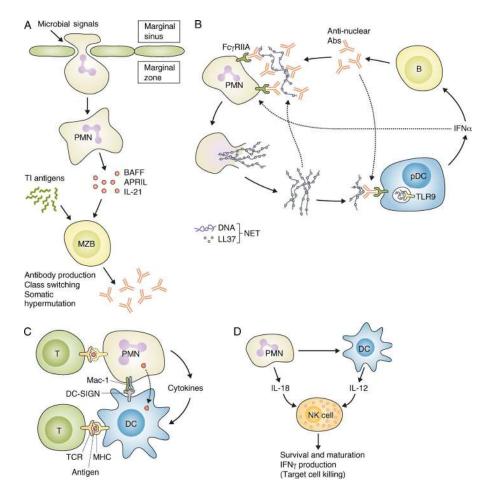


Figure 2. Novel interactions of neutrophils with other immune cells. (A) Neutrophils (PMN) accumulate in the splenic marginal zone upon microbial challenge and facilitate the antibody production and maturation of marginal zone B cells (MZB) in response to T cell-independent (TI) antigens. (B) Neutrophils participate in a vicious cycle during autoimmune disease (e.g., SLE) pathogenesis by releasing NETs that trigger pDCs to release IFN- α , which then promotes antibody production by B cells (B). (C) Neutrophils may directly present antigens to T cells (T) and they also activate DCs and deliver antigens to them. (D) Neutrophils and DCs jointly activate NK cells (NK).

antigen presentation by DCs or macrophages, supported by the migration of antigen-loaded DCs to the draining lymph nodes. Interestingly, human and mouse neutrophils also appear to be able to carry antigens to the lymph nodes by migrating in lymphatic vessels in a CCR7-dependent manner (Abadie et al., 2005; Maletto et al., 2006; Beauvillain et al., 2011). A recent study also showed that vaccinia virus-specific CD8⁺T cells emerge in the bone marrow and, based on fluorescence tracking and Ly6G antibody-mediated depletion approaches, identified neutrophils as the cells transporting the antigen from the dermis to the bone marrow (Duffy et al., 2012). Neutrophils also express molecules required for antigen processing and presentation to T cells (Ashtekar and Saha, 2003; Beauvillain et al., 2007). Late precursors committed to the neutrophil lineage are also important precursors of DC development (Oehler et al., 1998). Furthermore, mouse neutrophils are able to cross-prime naive T cells, i.e., present extracellular antigens on MHC class II molecules to CD8+ T cells (Beauvillain et al., 2007), and also prime antigen-specific Th1 and Th17 cells (Abi Abdallah et al., 2011). The transport of antigens from the periphery to the site of T cell priming and the direct priming/activation of antigen-specific T cells in an MHC-restricted manner raise the possibility that neutrophils may possibly even perform some level of classical antigen-presenting cell function (Fig. 2 C).

Although the aforementioned studies suggest that neutrophils positively regulate antigen-specific T cell responses, several works indicate a negative role for the neutrophil lineage in T cell function. As mentioned above, splenic neutrophils negatively regulate CD4⁺ T cell function in mice (Puga et al., 2011). A subset of human neutrophils also inhibits T cell responses during acute systemic inflammatory reactions such as bacterial sepsis (Pillay et al., 2012). Using Ly6G antibodymediated depletion and G-CSF receptor-deficient mice, a recent study showed that neutrophils prevent the spread of the T cell response to distal lymph nodes during immunization (Yang and Unanue, 2013). In addition, a large number of immature myeloid cells with inhibitory functions on T cells and other immune function, termed myeloid-derived suppressor cells (MDSCs), can be observed in cancer patients and in various other infectious or inflammatory conditions (Gabrilovich and Nagaraj, 2009). A subset of those cells (so-called granulocytic MDSCs) share many characteristics with (immature) neutrophils, highlighting the possibility of negative regulation of T cell function by the neutrophil lineage.

Regulation of DCs by neutrophils

The effect of neutrophils on adaptive immunity is not only mediated by direct effects on B and T cells, but also by affecting the function of DCs. Indeed, mouse neutrophils are capable of, and required for, the activation of DCs during microbial infection (Bennouna et al., 2003; Charmoy et al., 2010). Human and mouse studies suggest that neutrophils directly interact with DCs, which may lead to various responses including DC maturation and activation, access to antigens captured by neutrophils, or DC-mediated proliferation and $T_{\rm H}1$ polarization of T cells (Bennouna and Denkers, 2005; van Gisbergen et al., 2005b; Megiovanni et al., 2006). The interaction between human neutrophils and DCs has been proposed to be at least in part mediated by a unique interaction between DC-SIGN and Mac-1 on the neutrophil surface (van Gisbergen et al., 2005a,b; Boudaly, 2009; Yang et al., 2009; Fig. 2 C).

Recent studies have also revealed another unique role of neutrophils in DC activation during the pathogenesis of autoimmune diseases such as systemic lupus erythematosus (SLE) or type I diabetes (T1D). Characteristic features of SLE, an immune complex-mediated systemic autoimmune disease characterized by autoantibodies against nuclear antigens, include the chronic activation of plasmacytoid DCs (pDCs) leading to constitutive secretion of type I interferons such as IFN- α (Obermoser and Pascual, 2010) and the presence of circulating low-density granulocytes reminiscent of immature neutrophils (Bennett et al., 2003; Kaplan, 2011). Two recent studies on human patients shed novel light on the role of pDCs, type I interferons, and neutrophils in the core pathogenesis of SLE (Lande et al., 2011; Garcia-Romo et al., 2011; Fig. 2 B). SLE neutrophils were more prone to cell death by NETosis and to release NETs into the extracellular environment. Those NETs contained self-DNA in complex with antimicrobial proteins including LL37 (the C-terminal peptide derived from the human cathelicidin hCAP-18) and were able to activate pDCs through the TLR9 pathway and to trigger type I interferon production. This led to activation of B cells that produced autoantibodies against both self-DNA and the antimicrobial peptides. Type I interferons and immune complexes triggered further activation of neutrophils releasing more NETs (Lande et al., 2011), establishing a vicious cycle at the core of SLE pathogenesis (Fig. 2 B). Low-affinity Fcyreceptors (such as the human FcyRIIA) and TLR7 were required for immune complex-induced neutrophil activation (Garcia-Romo et al., 2011). FcyRIIA may also participate in the uptake of DNA-containing immune complexes by pDCs (Båve et al., 2003), followed by recognition of DNAantimicrobial peptide complexes by the TLR9 pathway (Garcia-Romo et al., 2011; Lande et al., 2011). It should be noted that pDCs do not recognize self-DNA under nonpathological conditions. However, the presence of LL37 in the DNA complexes in NETs prevented degradation of DNA, leading to delivery of DNA to early endosomes of pDCs, where it could activate the TLR9 pathway (Lande et al., 2007; Lande et al., 2011). Therefore, the unique composition of NETs makes them a suitable candidate to trigger pDC activation in the core of SLE pathogenesis. These results are also in line with a proposed contribution of NET degradation defects to SLE pathogenesis (Yasutomo et al., 2001; Hakkim et al., 2010), as well as a previous study on the presence of NET formation in autoimmune small vessel vasculitis (Kessenbrock et al., 2009).

The role of neutrophils in pDC activation and development of autoimmune diseases has also recently been proposed in the pathogenesis of T1D, which is also thought to involve type I interferon production by pDCs (Diana et al., 2013). Diana et al. (2013) reported early infiltration of the pancreas of young nonobese diabetic (NOD) mice with neutrophils, B-1a B cells, and pDCs. Pancreatic β-cell death triggered production and release of anti-DNA IgG antibodies from B-1a cells, which formed DNA-anti-DNA immune complexes and triggered, supposedly in an Fc-receptor-dependent manner, neutrophil NET formation and the release of the murine cathelicidin peptide CRAMP. The DNA-anti-DNA immune complexes and CRAMP then activated pDCs through TLR9 to release IFN- α , which is proposed to trigger further T cell-mediated autoimmune processes. In agreement with the proposed role of neutrophils in the development of T1D, injection of 2-wk-old NOD mice with the NIMP-R14 anti-Gr1 antibody reduced later development of diabetes in those animals (Diana et al., 2013).

The aforementioned studies suggest an unexpected role for neutrophils in very early phases of autoimmune disease pathogenesis through triggering of type I interferon production by pDCs. There are, however, several open questions that need to be addressed. First, it is widely believed that NET formation requires the NADPH oxidase (Fuchs et al., 2007); therefore SLE pathogenesis would be expected to rely on the presence of this enzyme. However, deficiency of the NADPH oxidase component Nox2 exacerbated, rather than inhibited disease pathogenesis in lupus-prone mice (Campbell et al., 2012), arguing against the role of NETs in SLE pathogenesis. This issue is further complicated by NADPH oxidaseindependent NET formation proposed by other recent studies (Chen et al., 2012; Parker et al., 2012b). Second, although Fc-receptor polymorphisms have been shown to be associated with human SLE (Takai, 2005), no such associations have been reported for T1D (even though NOD mice lacking activating Fc-receptors are partially protected from diabetes development; Inoue et al., 2007). Therefore, the role of the aforementioned mechanism in diabetes development may not be entirely relevant to human patients. Third, because NETs are likely often formed during bacterial infections (Yipp et al., 2012; McDonald et al., 2012), it is unclear how autoimmunity is avoided under normal healthy conditions. Nevertheless, the aforementioned mechanism provides an attractive model for the role of neutrophils in autoimmune disease pathogenesis that will merit further investigation. A similar mechanism has also been proposed during development of atherosclerosis (see The role of neutrophils in atherosclerosis), and NETs have also been suggested to participate in the pathogenesis of rheumatoid arthritis by displaying citrullinated autoantigens in an inflammatory environment (Khandpur et al., 2013), further extending the importance of such a pathway in human diseases.

Neutrophils in the regulation of NK cell homeostasis

NK cells are innate lymphocytes with critical roles in the elimination of virus-infected and tumor cells. In vitro interactions between neutrophils and NK cells were described before and a functional cooperation between the two cell types had been proposed (Costantini and Cassatella, 2011). The most detailed study tested the role of neutrophils in *Legionella pneumophila* infection in mice, aiming to understand why neutrophils, which are not capable of killing *Legionella* species themselves, still play a role in clearing *L. pneumophila* infection (Spörri et al., 2008). The authors confirmed that depletion of neutrophils by anti-Gr1 antibodies attenuated host defense against Legionella but it also blocked IFN- γ production by NK cells. Using elegant genetic approaches and mixed bone marrow chimeric studies, they showed that IFN- γ production by NK cells was directly stimulated by IL-18 released from neutrophils in response to Legionella infection (Spörri et al., 2008), suggesting that interactions between neutrophils and NK cells may be of functional importance (Fig. 2 D).

A recent study has provided novel and unexpected genetic evidence for the role of neutrophils in regulating NK cell homeostasis and function (Jaeger et al., 2012). The authors set up a random chemical mutagenesis screen to identify novel mutant mouse strains with defective NK cell development and function. They identified a mutation called Genista, which led to hyporesponsive NK cells as indicated by reduced in vitro and in vivo responses to target cells and ligation of NK cell-activating receptors (Jaeger et al., 2012). Interestingly, adoptive transfer studies revealed hyporesponsiveness of wild-type NK cells transferred to Genista recipients, indicating that the NK cell phenotype was, at least in part, caused by an environmental, non-cell autonomous defect. In parallel studies, the Genista mutation was identified to be a hypomorphic missense mutation of the Gfi-1 transcription factor, leading to defective terminal differentiation of neutrophils (as indicated by the absence of fully mature neutrophils in the periphery) without negatively affecting T or B cell lineages or other myeloid (monocyte, macrophage, DC) compartments (Ordoñez-Rueda et al., 2012; Jaeger et al., 2012). This was in line with the similar, though more severe, defect in neutrophil maturation in mice with complete disruption of the Gfi-1-encoding gene (Karsunky et al., 2002; Hock et al., 2003) and congenital neutropenia in human patients carrying a dominant-negative Gfi-1 mutation (Person et al., 2003). To further confirm the role of neutrophils in NK cell function, the authors showed that depletion of neutrophils by an anti-Ly6G antibody also reduced the functional responsiveness of NK cells (Jaeger et al., 2012). Further studies revealed that both the Genista mutation and the Ly6G antibody-mediated depletion led to defective terminal maturation and poor survival of NK cells. The authors also reported that patients suffering from various forms of congenital or acquired (autoimmune) neutropenia had higher percentages of NK cells showing an immature phenotype and reduced responsiveness to target cells (Jaeger et al., 2012). Interestingly, one of the patients with autoimmune neutropenia showed a remission during the course of the study, during which normalization of peripheral neutrophil count and NK cell function occurred in parallel. Collectively, those results suggest that neutrophils likely play a role in regulating the

terminal differentiation and functional responsiveness of NK cells (Fig. 2 D). The colocalization and direct physical interaction of the two cell types in mouse lymphoid organs (Jaeger et al., 2012) and inflamed human tissues (Costantini et al., 2011) suggest that neutrophils may directly regulate the NK cell compartment.

Although the aforementioned studies indicate an important role for neutrophils in the regulation of NK cell function, the contribution of a third cell type (such as DCs) in this interaction cannot be excluded. Indeed, besides neutrophilderived IL-18, IFN- γ production by NK cells upon *Legionella* infection also required IL-12 derived from DCs as a result of their reciprocal regulation with NK cells (Spörri et al., 2008; Fig. 2 D). A tripartite complex formed by neutrophils, NK cells and a minor subset of DCs (slanDCs) has also been observed in human samples (including inflammatory lesions in Crohn's disease and psoriasis) and was proposed to promote IFN- γ production by NK cells (Costantini et al., 2011).

Collectively, neutrophils have recently emerged as potentially important regulators of NK cell homeostasis and activation, possibly in close relationship with DCs.

Neutrophils in allergy and anaphylaxis

Allergic and anaphylactic reactions are adaptive immune responses triggered by repeated exposure to allergens, and are typically characterized by vascular reactions such as vasodilation and edema formation. Several recent studies suggest unexpected roles for neutrophils in those reactions.

The prevailing paradigm of anaphylaxis development is that allergen-induced cross-linking of IgE molecules bound to Fce-receptors on mast cells and basophils triggers histamine release and subsequent systemic vasodilation (Khan and Kemp, 2011). Although this mechanism is likely responsible for the majority of anaphylactic reactions, several observations indicate that alternative mechanisms, possibly mediated by IgG and cellular lineages other than mast cells and basophils, may also be at play (Finkelman, 2007; Tsujimura et al., 2008; Ohnmacht et al., 2010). Jönsson et al. (2011) identified neutrophils as important players of active and IgG-induced passive systemic anaphylaxis in experimental mice (Fig. 3 A). Passive systemic anaphylaxis triggered by IgG antibodies was blocked by neutrophil depletion using the RB6-8C5 anti-Gr1 antibody. Active systemic anaphylaxis was partially reduced by anti-Gr1 antibodies alone, and it was nearly completely blocked by simultaneously depleting both neutrophils and basophils (Jönsson et al., 2011). Interestingly, mice lacking all activating Fcy- and Fce-receptors except FcyRIV were able to mount normal active anaphylactic reactions which were, however, completely blocked by anti-Gr1 antibodies. Adoptive transfer of wild-type neutrophils rescued anaphylaxis development in FcR γ -chain-deficient animals (which lack all activating $Fc\gamma$ - and $Fc\varepsilon$ -receptors including FcyRIV, and fail to mount active systemic anaphylaxis). Those results suggest that neutrophils are responsible for the FcyRIVmediated component of active systemic anaphylaxis in mice

and that this component is sufficient to trigger a full-blown anaphylaxis response. Additional experiments indicated that the neutrophil-dependent anaphylactic reaction was mediated primarily by PAF rather than histamine (Jönsson et al., 2011). Importantly, active systemic anaphylaxis could be restored by adoptive transfer of human neutrophils to FcR $\gamma^{-/-}$ animals (Jönsson et al., 2011) or by transgenic expression of human Fc γ RIIA in FcR $\gamma^{-/-}$ mice (Jönsson et al., 2012), suggesting that human neutrophils may also be able to mediate systemic anaphylactic reactions.

The various forms of skin allergies comprise a major group of localized allergic diseases. As with other allergic diseases, their mechanism can also be divided into two conceptually different phases, the sensitization phase (leading to priming of the adaptive immune system) and the elicitation phase (causing local tissue damage through activation of the primed adaptive immune cells). They are also often complicated by additional inflammatory insults, such as chemically induced tissue damage in allergic contact dermatitis or mechanical damage caused by itching-induced scratching in atopic dermatitis. Several recent studies indicate an important role for neutrophils in skin allergies. Anti-Gr1 antibody treatment has been known to prevent ear swelling and T cell infiltration during contact hypersensitivity, a mouse model of allergic contact dermatitis (Dilulio et al., 1999). In a recent study, neutrophils expressing the LTB₄-receptor BLT1 were found to be required for inflammation and recruitment of allergen-primed T cells at the allergen challenge site in a novel mouse model of atopic dermatitis combining ovalbumin immunization and challenge with mechanical skin injury (reminiscent of itching-induced scratching; Oyoshi et al., 2012).

Both of the aforementioned studies revealed important roles for neutrophils in the elicitation phase of allergic skin inflammation. However, although innate immune mechanisms likely also play important roles in DC activation and T cell priming during the earlier and conceptually different sensitization phase of allergic skin diseases (Kaplan et al., 2012), no studies on the role on neutrophils in that phase have been completed yet. Interestingly, our own recent studies (unpublished data; published in abstract form in Weber et al., 2012) suggested an important role for neutrophils in the sensitization phase of contact hypersensitivity. In that study, we found that Ly6G antibody-mediated depletion of neutrophils, selectively during the sensitization phase, abrogated the contact hypersensitivity reaction, and diminished DC activation and priming of allergen-specific T cells. In addition, lymph node cells from allergen-sensitized mice that were previously treated with neutrophil-depleting anti-Ly6G antibodies or were genetically deficient of neutrophils because of a myeloid-specific deletion of the Mcl-1 protein (Dzhagalov et al., 2007) were unable to transfer sensitivity toward contact allergens to normal naive recipients (Weber et al., 2012). Therefore, neutrophils appear to be critical players in both the sensitization and elicitation phases of allergic skin inflammatory reactions.

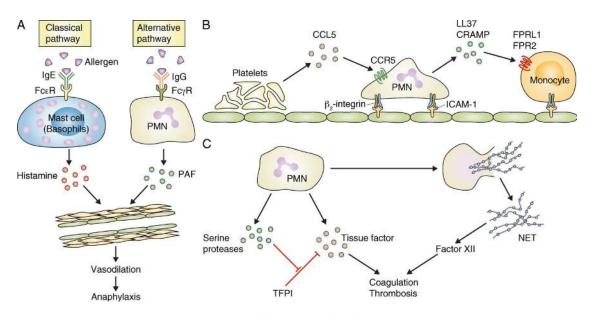


Figure 3. Novel functions of neutrophils in vascular diseases. (A) Neutrophils (PMN) mediate an alternative pathway of anaphylactic reaction which is distinct from the classical pathway mediated by mast cells or basophils. (B) Sequential recruitment of platelets, neutrophils, and monocytes (Mono) participates in the pathogenesis of atherosclerosis. (C) Neutrophils contribute to coagulation and thrombosis by releasing tissue factor and inhibitor (TFPI), as well as the procoagulant NETs.

The role of neutrophils in metabolic diseases

Metabolic diseases such as diabetes and obesity are among the largest health challenges of the 21st century. Several recent studies suggest unexpected roles for neutrophils in those diseases.

Obesity and the resulting insulin resistance is one of the major causes of metabolic syndrome and type 2 diabetes. It has long been known that obesity has an important inflammatory component and that this contributes to the resulting insulin resistance (Gregor and Hotamisligil, 2011). The primary cell type responsible for inflammation in obesity has been considered to be the inflammatory macrophage in adipose tissues (Olefsky and Glass, 2010). However, neutrophils have also been shown to be recruited to adipose tissues early during the course of diet-induced obesity in mice (Elgazar-Carmon et al., 2008) and to be present in increased numbers in the fatty liver of obese human patients (Rensen et al., 2009). A recent report has confirmed the dramatic accumulation of adipose tissue neutrophils, as well as the increased presence and activity of the neutrophil-specific protease neutrophil elastase, during high-fat diet in experimental mice (Talukdar et al., 2012). Importantly, the pharmacological inhibition or genetic deletion of neutrophil elastase led to improved in vivo glucose tolerance and insulin sensitivity, likely due to improved responsiveness of liver and adipose tissue cells to insulin. The authors proposed that neutrophil elastase gains access to the intracellular space (endocytic compartments) of hepatocytes and degrades insulin receptor substrate 1 (IRS-1), leading to attenuation of insulin signal transduction pathways (Talukdar et al., 2012; similar to a mechanism previously proposed to occur in lung cancer cells; Houghton et al., 2010). Although there is precedent for translocation of proteases through cell membranes (such as

perforin-mediated translocation of granzymes), it is unclear how this would happen in case of neutrophil elastase and how elastase would selectively cleave IRS-1 without wreaking havoc inside the target cell. The problem of protease-mediated protein degradation during lysis of neutrophil-containing biological samples is also well known in the scientific community; therefore more careful studies to exclude the possibility of IRS-1 degradation during sample preparation are warranted.

Another study mentioned earlier suggests a role for neutrophils in the pathogenesis of T1D, an autoimmune disease caused by degradation of pancreatic β -cells (Diana et al., 2013). In that study, neutrophils were shown to participate in the activation of type I cytokine production by pDCs and in subsequent activation of the adaptive immune response (Diana et al., 2013). That study places neutrophils high in the hierarchy of autoimmune disease pathogenesis, participating in the shaping of the adaptive immune response.

Collectively, although the studies on the role of neutrophils in diabetes and related metabolic diseases may open up new directions of research that may significantly improve our understanding of the inflammatory origin of metabolic diseases, they also raise several additional questions and concerns that have yet to be addressed.

The role of neutrophils in atherosclerosis

Cardiovascular diseases are among the most prevalent human diseases and are the leading causes of death in developed countries. Several recent studies suggest that neutrophils play an important role in those diseases, in particular the development and progression of atherosclerosis and the pathogenesis of thrombosis.

It has long been appreciated that atherosclerosis results from chronic inflammation of the vascular wall (Hansson, 2005). Recent reports indicate that neutrophils are present in unique areas of atherosclerotic plaques in both humans and mice (Naruko et al., 2002; Ionita et al., 2010; Drechsler et al., 2010; Rotzius et al., 2010) and that their presence correlates with poor prognosis and signs of plaque destabilization in human patients (Naruko et al., 2002; Ionita et al., 2010). Importantly, depletion of neutrophils by antibodies against Ly6G strongly reduced the area of atherosclerotic lesions in apolipoprotein E-deficient (ApoE^{-/-}) mice fed a high-fat diet for 4 wk, suggesting an important role for neutrophils in atherosclerotic plaque formation (Drechsler et al., 2010). A recent study provided additional support for the role of neutrophils in atherosclerosis, showing that the genetic deficiency of CRAMP, a member of the cathelicidin family of antimicrobial peptides expressed in neutrophils, leads to reduced atherosclerotic lesion size in ApoE^{-/-} mice fed a highfat diet (Döring et al., 2012a). CRAMP and the homologous human cathelicidin peptide LL37 (the C-terminal portion of its precursor hCAP-18) promoted the adhesion of classical (inflammatory) monocytes to the vessel wall in mice, and this was inhibited by antagonists of formyl-peptide receptors, the receptors thought to be responsible for cathelicidin-mediated leukocyte migration (Döring et al., 2012a). In addition, activation of pDCs by complexes of cathelicidins with extracellular DNA from neutrophils or other dying cells may also contribute to atherosclerosis development in mice (Döring et al., 2012b) in a manner similar to that proposed for the pathogenesis of SLE and T1D (Lande et al., 2011; Garcia-Romo et al., 2011; Talukdar et al., 2012). The recruitment of neutrophils to the inflamed mouse arterial wall could also be reduced by platelet depletion or by the genetic deletion of CCR5, the receptor for the platelet-derived CCL5 (RANTES) chemokine (Drechsler et al., 2010) which is also required for atherosclerotic plaque formation (Braunersreuther et al., 2007). The release and deposition of CCL5 appears to be dependent on P-selectin on platelets (Huo et al., 2003; Drechsler et al., 2010). Platelets were previously proposed to participate in atherosclerotic plaque formation (Massberg et al., 2002; Huo et al., 2003) and platelet-derived CCL5 was shown to promote monocyte adhesion to the luminal surface of early atherosclerotic lesions in the inflamed arterial wall in mice (von Hundelshausen et al., 2001; Huo et al., 2003). Mice lacking P-selectin, CD18, or ICAM-1 were previously shown to have reduced atherosclerotic plaque formation (Johnson et al., 1997; Nageh et al., 1997; Collins et al., 2000). Collectively, atherosclerotic plaque formation likely involves the sequential recruitment of platelets, neutrophils, and monocytes, whereby platelet-derived CCL5, acting on CCR5, is at least partially responsible for neutrophil recruitment and neutrophil promotion of monocyte recruitment by cathelicidin-induced activation of formyl-peptide receptors (Fig. 3 B). Platelet P-selectin is likely involved in platelet deposition (although an additional role of endothelial P-selectin in adhesion of leukocyte rolling cannot be excluded) and neutrophil/monocyte

adhesion is likely mediated by interaction between β_2 -integrins and ICAM-1.

It should be mentioned that another recent study on vascular healing after wire injury, a mouse model of the vascular injury triggered by insertion of an arterial stent, placed the role of neutrophils in a different perspective (Soehnlein et al., 2011). Ly6G antibody-mediated depletion of neutrophils or genetic deficiency of CRAMP inhibited endothelial regeneration and promoted neointima formation (the major cause of arterial restenosis) after vessel injury, whereas LL37 promoted endothelial recovery in an in vitro wound-healing assay (Soehnlein et al., 2011). Most importantly, covalent coating of arterial stents with LL37 significantly reduced instent stenosis (Soehnlein et al., 2011). Those results suggest that neutrophils and neutrophil-derived cathelicidins may play a protective role during vascular healing after stent-mediated vascular damage and raise the possibility that LL37 treatment may reduce in-stent stenosis after stent insertion.

The aforementioned studies suggest potentially important roles for neutrophils in atherosclerosis and related vascular diseases. However, additional work will be required to delineate the exact mechanism of how and in which phase and spatiotemporal order neutrophils participate in plaque formation. For example, do neutrophils get activated inside or outside the vasculature? How do intravascular events promote plaque formation under the endothelium? Are the roles of neutrophils in initial monocyte adhesion and eventual plaque destabilization related or independent events? Although plaque formation and stent restenosis likely occur through different mechanisms, the opposing effect of neutrophils and CRAMP/ LL37 on those two endothelial damage-associated processes may also require additional mechanistic explanation. The role of CCR5 in neutrophil activation also raises questions, as this receptor is not expressed on neutrophils at high levels (although increased expression was shown upon apoptosis and at the site of inflammation; Ariel et al., 2006; Hartl et al., 2008).

Neutrophils in thrombus formation

Arterial or venous thrombosis is another major complication of vascular diseases, and causes significant morbidity and mortality worldwide. Several recent studies suggest that neutrophils may play a major role in triggering thrombosis in experimental mice.

Neutrophils were found to be the first cell type to adhere to the site of endothelial injury in both arterial and venous thrombosis models (Darbousset et al., 2012; von Brühl et al., 2012), and their accumulation at the site of vessel injury (along with that of monocytes) preceded the activation of platelets (Darbousset et al., 2012). Importantly, thrombus formation was dramatically reduced upon depletion of neutrophils by anti-Ly6G antibodies in a flow restriction-induced venous thrombosis model (von Brühl et al., 2012) or by anti-Gr1 antibodies in laser-induced microvascular injury (Darbousset et al., 2012). Whereas contact of plasma proteins with extravascular tissue factor is the primary trigger of thrombus formation upon vessel injury, the source of prothrombotic agents during arterial or venous thrombosis with an intact endothelium has been poorly understood. After some debate about the presence of tissue factor in neutrophils (Maugeri et al., 2005) it now appears that activated (but not resting) neutrophils express and release a significant amount of tissue factor (Maugeri et al., 2006; Fig. 3 C). Importantly, neutrophilmediated thrombus formation required tissue factor in a myeloid compartment (von Brühl et al., 2012; Darbousset et al., 2012), and adoptive transfer studies suggested that neutrophils themselves may be an important source of tissue factor in a microvascular thrombosis model in mice (Darbousset et al., 2012).

The coagulation system is an important host defense system in lower organisms, and there is also significant interaction between innate immunity and coagulation in higher vertebrates (Esmon, 2004). A recent study suggested that neutrophil serine proteases play an important role in the reciprocal coupling between innate immunity and coagulation in mice (Massberg et al., 2010; Fig. 3 C). The authors showed that the neutrophil-derived proteases neutrophil elastase and cathepsin G were required for thrombus formation after chemically induced vessel injury, as well as for activation of factor X upon co-incubation of neutrophils and platelets. The two enzymes inactivated the tissue factor pathway inhibitor protein, thereby promoting the coagulation cascade. Importantly, triggering of coagulation also prevented systemic dissemination of infectious Escherichia coli bacteria, suggesting the existence of a poorly appreciated level of innate host defense in mammals.

The aforementioned study also suggested that neutrophilderived externalized nucleosomal structures (likely corresponding to NETs) also play a procoagulant role in thrombosis (Massberg et al., 2010; Fig. 3 C). Indeed, NETs were shown to be able to trigger platelet activation and thrombus formation in vitro (Fuchs et al., 2010). A significant amount of extracellular DNA trap formation inside the thrombus could be observed upon flow restriction-induced venous thrombosis in mice (Brill et al., 2012; von Brühl et al., 2012) and obstruction-induced venous thrombosis in baboons (Fuchs et al., 2010). Importantly, blocking NET formation by DNase treatment strongly reduced venous thrombus formation (Brill et al., 2012; von Brühl et al., 2012). Interestingly, various malignant states, including chronic myelogenous leukemia and various solid tumors, promote NET formation, which may be an important pathogenic mechanism of cancerassociated thrombosis (Demers et al., 2012). The proposed mechanism of NET-induced thrombus formation may involve the activation of Factor XII by the negatively charged NET surface, with a direct effect on the stability of the fibrin network (i.e., independent of the activation of Factor XI; von Brühl et al., 2012).

Collectively, neutrophils may contribute to the thrombosis and coagulation process by releasing tissue factor, even in the absence of vascular injury, by releasing serine proteases that inactivate the tissue factor pathway inhibitor, and/or by the formation of pro-coagulant NETs (Fig. 3 C).

Additional subjects not discussed in detail

Several topics of major importance could not be covered in detail in this review. A brief discussion below points to some of those topics, with key references for further reading.

The role of neutrophils in tumor biology has long been appreciated (Pekarek et al., 1995). There has been several recent developments in the field such as the polarization of tumor-associated neutrophils by TGF- β (Fridlender et al., 2009), a proposed role of neutrophil elastase in tumor growth (Houghton et al., 2010), the inhibition of tumor metastasis by neutrophils (Granot et al., 2011), neutrophils in cancerinduced thrombosis (Demers et al., 2012), the role of neutrophil-derived factors in maintaining B cell lymphoma (Schwaller et al., 2007), the potential role of neutrophils in tumor angiogenesis (Jablonska et al., 2010), and the role of histamine in preventing inflammation-induced carcinogenesis (Yang et al., 2011). Neutrophil-related immature myeloid cells called myeloid-derived suppressor cells also appear to play an immunosuppressive role in cancer (Gabrilovich and Nagaraj, 2009). The readers are referred to recent reviews for further information on this very interesting but diverse field of study (Gregory and Houghton, 2011; Fridlender and Albelda, 2012).

Besides the mostly positive roles of neutrophils in various physiological and pathological processes, neutrophils also play inhibitory or regulatory roles under certain conditions. Apoptosis of neutrophils triggers an active process promoting resolution of inflammation (Serhan, 2007; Serhan et al., 2008; Soehnlein and Lindbom, 2010). A subset of neutrophilrelated immature myeloid cells called myeloid-derived suppressor cells play a critical role in regulation of inflammation in cancer and potentially other conditions (Gabrilovich and Nagaraj, 2009). Neutrophils may also be able to produce the antiinflammatory cytokine IL-10 (Zhang et al., 2009; De Santo et al., 2010), though this is disputed in case of human neutrophils (Davey et al., 2011). Further information can be found in recent studies (Serhan et al., 2008; Gabrilovich and Nagaraj, 2009; Soehnlein and Lindborn, 2010; Mantovani et al., 2011).

Whereas many other leukocyte lineages have been subdivided into various subsets, neutrophils have so far been mostly considered as a homogeneous population. That view is beginning to change with novel functions ascribed to subpopulations of neutrophils (Fridlender et al., 2009; Puga et al., 2011; Pillay et al., 2012).

This review has focused on what neutrophils do rather than how they do it; therefore, most information regarding the mechanism of neutrophil function has been omitted. There are, however, exciting new discoveries in those fields, as well, such as various new mechanisms and cues guiding the migration of neutrophils (McDonald et al., 2010;Woodfin et al., 2011; Proebstl et al., 2012; Stark et al., 2013). Two brand new and possibly related studies also propose exciting and provocative ideas about how neutrophils may fight microbes: Timár et al. (2013) have recently proposed an antimicrobial function of neutrophil-derived microvesicles, whereas Yipp et al. (2012) suggested that anuclear neutrophils (formed after releasing the nuclear material to NETs) preserve at least part of their antimicrobial activities. In addition, the role of neutrophils in host defense and tissue damage is mediated by diverse intracellular signal transduction processes (Futosi et al., 2013).

Potential caveats

After several years of being somewhat overlooked by immunologists, the last few years have witnessed an explosion of major studies on the diverse roles of neutrophils in health and disease. However, as with any trendy and fashionable subjects, it is reasonable to exercise healthy skepticism toward novel results until they are confirmed and widely accepted by the scientific community. As the aim of this review was to discuss novel findings that have not yet been subjected to detailed examination and scrutiny, this particularly applies to the reports discussed in this review.

Indeed, one has the feeling that some of the closely related papers discussed here followed each other at a pace that did not allow detailed reexamination of the original concepts and methodological approaches. As an example, the study proposing cleavage of intracellular IRS-1 by neutrophil elastase during high fat diet-induced insulin resistance (Talukdar et al., 2012) apparently relied on a previous similar report on lung cancer growth (Houghton et al., 2010) without carefully reexamining the approaches used to distinguish true intracellular cleavage from degradation during sample processing. That has left room for skepticism about the conclusion drawn, especially because the very large amount of proteolytic activity in neutrophil lysates.

A large number of studies also propose interesting novel roles for NETs in autoimmunity, inflammation, and vascular diseases. However, there is ongoing debate about when and how NETs are formed in vivo, and whether they can indeed be generated in sufficient quantities under sterile conditions to support the proposed processes.

We should also admit that we do not have perfect tools to study the role of neutrophils in any given disease process. Typical examples are the use of various antibodies to deplete neutrophils, of which antibodies against Gr1 (RB6-8C5 and, likely, NIMP-R14) may also deplete leukocyte lineages other than neutrophils. We also miss appropriate genetic tools to specifically delete neutrophils; indeed, none of the various approaches (Liu et al., 1996; Karsunky et al., 2002; Hock et al., 2003; Jonsson et al., 2005; Dzhagalov et al., 2007; Ordoñez-Rueda et al., 2012) have been tested in sufficient detail to allow exclusion of an effect on other leukocyte lineages. Several proposed functions of neutrophils are also based on evidence gained from knockout mice lacking granule proteins (e.g., elastase or CRAMP) expressed in neutrophils. However, those proteins are also expressed in other myeloid cells such as macrophages, casting doubt about the lineage specificity of those experiments.

Finally, most of the studies discussed were performed on experimental mice. Although the physiology of mice and humans appears to be similar in many sense, there are also significant differences, such as the different expression of Fcreceptors (Bruhns, 2012) the apparent lack of IL-10 production by human but not murine neutrophils (Davey et al., 2011), as well as more widespread differences in the overall immune response (Mestas and Hughes, 2004; Seok et al., 2013). Therefore, care should be taken when extrapolating mouse studies to human physiology and pathophysiology.

Concluding remarks and future directions

Neutrophils are the most abundant circulating leukocytes, yet their full spectrum of biological functions in immunity and beyond is just beginning to emerge, likely because of the difficulties in manipulating this terminally differentiated, shortlived cell type. Studies published in the last few years propose that neutrophils play a much more diverse role than previously appreciated. Their putative novel roles described in this review include functions in viral infections and infections with intracellular pathogens; intricate interaction with various other components of the immune system; and roles in diseases such as autoimmune diseases, anaphylaxis, diabetes, atherosclerosis, and thrombosis. Although those studies have yet to be closely examined, scrutinized, and, hopefully, confirmed by the scientific community, they propose exciting novel avenues for the understanding of how our immune system contributes to health and disease.

The function of neutrophils in immune defense and their role in various immune-mediated (e.g., autoimmune; Németh and Mócsai, 2012) diseases result in significant interest in this cell type in clinical medicine. The emerging information on what neutrophils do and how they do it may hopefully be translated to important novel approaches of the diagnosis and treatment of diseases with a substantial neutrophil component.

l apologize for the omission of many seminal papers due to space limitation. The author thanks Erzsébet Ligeti, Marco Cassatella, Oliver Söhnlein, Zoltán Jakus, and Tamás Németh for helpful suggestions and careful reading of the manuscript.

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REFERENCES

- Abadie, V., E. Badell, P. Douillard, D. Ensergueix, P.J. Leenen, M. Tanguy, L. Fiette, S. Saeland, B. Gicquel, and N. Winter. 2005. Neutrophils rapidly migrate via lymphatics after Mycobacterium bovis BCG intradermal vaccination and shuttle live bacilli to the draining lymph nodes. *Blood*. 106:1843–1850. http://dx.doi.org/10.1182/blood-2005-03-1281
- Abi Abdallah, D.S., C.E. Egan, B.A. Butcher, and E.Y. Denkers. 2011. Mouse neutrophils are professional antigen-presenting cells programmed to instruct Th1 and Th17 T-cell differentiation. *Int. Immunol.* 23:317–326. http://dx.doi.org/10.1093/intimm/dxr007
- Amulic, B., C. Cazalet, G.L. Hayes, K.D. Metzler, and A.Zychlinsky. 2012. Neutrophil function: from mechanisms to disease. *Annu. Rev. Immunol.* 30:459–489. http://dx.doi.org/10.1146/annurev-immunol-020711-074942
- Appelberg, R. 2007. Neutrophils and intracellular pathogens: beyond phagocytosis and killing. *Trends Microbiol.* 15:87–92. http://dx.doi.org/ 10.1016/j.tim.2006.11.009

- Ariel, A., G. Fredman, Y.P. Sun, A. Kantarci, T.E. Van Dyke, A.D. Luster, and C.N. Serhan. 2006. Apoptotic neutrophils and T cells sequester chemokines during immune response resolution through modulation of CCR5 expression. *Nat. Immunol.* 7:1209–1216. http://dx.doi.org/10.1038/ni1392
- Ashtekar, A.R., and B. Saha. 2003. Poly's plea: membership to the club of APCs. *Trends Immunol*. 24:485–490. http://dx.doi.org/10.1016/S1471-4906(03)00235-7
- Balázs, M., F. Martin, T. Zhou, and J. Kearney. 2002. Blood dendritic cells interact with splenic marginal zone B cells to initiate T-independent immune responses. *Immunity*. 17:341–352. http://dx.doi.org/10.1016/S1074-7613(02)00389-8
- Båve, U., M. Magnusson, M.L. Eloranta, A. Perers, G.V. Alm, and L. Rönnblom. 2003. Fc γ RIIa is expressed on natural IFN-α-producing cells (plasmacytoid dendritic cells) and is required for the IFN-α production induced by apoptotic cells combined with lupus IgG. J. Immunol. 171:3296–3302.
- Bazzoni, F., M.A. Cassatella, C. Laudanna, and F. Rossi. 1991a. Phagocytosis of opsonized yeast induces tumor necrosis factor-α mRNA accumulation and protein release by human polymorphonuclear leukocytes. J. Leukoc. Biol. 50:223–228.
- Bazzoni, F., M.A. Cassatella, F. Rossi, M. Ceska, B. Dewald, and M. Baggiolini. 1991b. Phagocytosing neutrophils produce and release high amounts of the neutrophil-activating peptide 1/interleukin 8. J. Exp. Med. 173:771– 774. http://dx.doi.org/10.1084/jem.173.3.771
- Beauvillain, C., Y. Delneste, M. Scotet, A. Peres, H. Gascan, P. Guermonprez, V. Barnaba, and P. Jeannin. 2007. Neutrophils efficiently cross-prime naive T cells in vivo. *Blood*. 110:2965–2973. http://dx.doi.org/10.1182/ blood-2006-12-063826
- Beauvillain, C., P. Cunin, A. Doni, M. Scotet, S. Jaillon, M.L. Loiry, G. Magistrelli, K. Masternak, A. Chevailler, Y. Delneste, and P. Jeannin. 2011. CCR7 is involved in the migration of neutrophils to lymph nodes. *Blood*. 117:1196–1204. http://dx.doi.org/10.1182/blood-2009-11-254490
- Bennett, L., A.K. Palucka, E. Arce, V. Cantrell, J. Borvak, J. Banchereau, and V. Pascual. 2003. Interferon and granulopoiesis signatures in systemic lupus erythematosus blood. J. Exp. Med. 197:711–723. http://dx.doi.org/ 10.1084/jem.20021553
- Bennouna, S., and E.Y. Denkers. 2005. Microbial antigen triggers rapid mobilization of TNF-alpha to the surface of mouse neutrophils transforming them into inducers of high-level dendritic cell TNF-alpha production. *J. Immunol.* 174:4845–4851.
- Bennouna, S., S.K. Bliss, T.J. Curiel, and E.Y. Denkers. 2003. Cross-talk in the innate immune system: neutrophils instruct recruitment and activation of dendritic cells during microbial infection. J. Immunol. 171:6052–6058.
- Berry, M.P., C.M. Graham, F.W. McNab, Z. Xu, S.A. Bloch, T. Oni, K.A. Wilkinson, R. Banchereau, J. Skinner, R.J. Wilkinson, et al. 2010. An interferon-inducible neutrophil-driven blood transcriptional signature in human tuberculosis. *Nature*. 466:973–977. http://dx.doi.org/10.1038/ nature09247
- Borregaard, N. 2010. Neutrophils, from marrow to microbes. *Immunity*. 33:657–670. http://dx.doi.org/10.1016/j.immuni.2010.11.011
- Borregaard, N., O.E. Sørensen, and K. Theilgaard-Mönch. 2007. Neutrophil granules: a library of innate immunity proteins. *Trends Immunol.* 28:340– 345. http://dx.doi.org/10.1016/j.it.2007.06.002
- Boudaly, S. 2009. Activation of dendritic cells by polymorphonuclear neutrophils. Front. Biosci. 14:1589–1595. http://dx.doi.org/10.2741/3326
- Bratton, D.L., and P.M. Henson. 2011. Neutrophil clearance: when the party is over, clean-up begins. *Trends Immunol.* 32:350–357. http://dx.doi.org/ 10.1016/j.it.2011.04.009
- Braunersreuther, V., A. Zernecke, C. Arnaud, E.A. Liehn, S. Steffens, E. Shagdarsuren, K. Bidzhekov, F. Burger, G. Pelli, B. Luckow, et al. 2007. Ccr5 but not Ccr1 deficiency reduces development of diet-induced atherosclerosis in mice. *Arterioscler. Thromb. Vasc. Biol.* 27:373–379. http://dx.doi.org/10.1161/01.ATV.0000253886.44609.ae
- Brill, A., T.A. Fuchs, A.S. Savchenko, G.M. Thomas, K. Martinod, S.F. De Meyer, A.A. Bhandari, and D.D. Wagner. 2012. Neutrophil extracellular traps promote deep vein thrombosis in mice. *J. Thromb. Haemost.* 10:136– 144. http://dx.doi.org/10.1111/j.1538-7836.2011.04544.x

- Brinkmann, V., and A. Zychlinsky. 2012. Neutrophil extracellular traps: is immunity the second function of chromatin? J. Cell Biol. 198:773–783. http://dx.doi.org/10.1083/jcb.201203170
- Brinkmann, V., U. Reichard, C. Goosmann, B. Fauler, Y. Uhlemann, D.S. Weiss, Y. Weinrauch, and A. Zychlinsky. 2004. Neutrophil extracellular traps kill bacteria. *Science*. 303:1532–1535. http://dx.doi.org/10 .1126/science.1092385
- Bruhns, P. 2012. Properties of mouse and human IgG receptors and their contribution to disease models. *Blood.* 119:5640–5649. http://dx.doi.org/ 10.1182/blood-2012-01-380121
- Campbell, A.M., M. Kashgarian, and M.J. Shlomchik. 2012. NADPH oxidase inhibits the pathogenesis of systemic lupus erythematosus. *Sci. Transl. Med.* 4:157ra141. http://dx.doi.org/10.1126/scitranslmed.3004801
- Cassatella, M.A. 1995. The production of cytokines by polymorphonuclear neutrophils. *Immunol. Today.* 16:21–26. http://dx.doi.org/10 .1016/0167-5699(95)80066-2
- Charmoy, M., S. Brunner-Agten, D. Aebischer, F. Auderset, P. Launois, G. Milon, A.E. Proudfoot, and F. Tacchini-Cottier. 2010. Neutrophilderived CCL3 is essential for the rapid recruitment of dendritic cells to the site of *Leishmania major* inoculation in resistant mice. *PLoS Pathog.* 6:e1000755. http://dx.doi.org/10.1371/journal.ppat.1000755
- Chen, K., H. Nishi, R. Travers, N. Tsuboi, K. Martinod, D.D. Wagner, R. Stan, K. Croce, and T.N. Mayadas. 2012. Endocytosis of soluble immune complexes leads to their clearance by FcγRIIIB but induces neutrophil extracellular traps via FcγRIIA in vivo. *Blood*. 120:4421–4431. http://dx.doi.org/10.1182/blood-2011-12-401133
- Collins, R.G., R. Velji, N.V. Guevara, M.J. Hicks, L. Chan, and A.L. Beaudet. 2000. P-Selectin or intercellular adhesion molecule (ICAM)-1 deficiency substantially protects against atherosclerosis in apolipoprotein E-deficient mice. J. Exp. Med. 191:189–194. http://dx.doi.org/ 10.1084/jem.191.1.189
- Costantini, C., and M.A. Cassatella. 2011. The defensive alliance between neutrophils and NK cells as a novel arm of innate immunity. J. Leukoc. Biol. 89:221–233. http://dx.doi.org/10.1189/jlb.0510250
- Costantini, C., F. Calzetti, O. Perbellini, A. Micheletti, C. Scarponi, S. Lonardi, M. Pelletier, K. Schakel, G. Pizzolo, F. Facchetti, et al. 2011. Human neutrophils interact with both 6-sulfo LacNAc+ DC and NK cells to amplify NK-derived IFNγ: role of CD18, ICAM-1, and ICAM-3. *Blood*. 117:1677–1686. http://dx.doi.org/10.1182/blood-2010-06-287243
- Cruz, A., A.G. Fraga, J.J. Fountain, J. Rangel-Moreno, E. Torrado, M. Saraiva, D.R. Pereira, T.D. Randall, J. Pedrosa, A.M. Cooper, and A.G. Castro. 2010. Pathological role of interleukin 17 in mice subjected to repeated BCG vaccination after infection with *Mycobacterium tuberculosis. J. Exp. Med.* 207:1609–1616. http://dx.doi.org/10.1084/jem.20100265
- Daley, J.M., A.A. Thomay, M.D. Connolly, J.S. Reichner, and J.E. Albina. 2008. Use of Ly6G-specific monoclonal antibody to deplete neutrophils in mice. J. Leukoc. Biol. 83:64–70. http://dx.doi.org/10.1189/jlb.0407247
- Darbousset, R., G.M. Thomas, S. Mezouar, C. Frère, R. Bonier, N. Mackman, T. Renné, F. Dignat-George, C. Dubois, and L. Panicot-Dubois. 2012. Tissue factor-positive neutrophils bind to injured endothelial wall and initiate thrombus formation. *Blood.* 120:2133–2143. http://dx.doi.org/ 10.1182/blood-2012-06-437772
- Davey, M.S., N. Tamassia, M. Rossato, F. Bazzoni, F. Calzetti, K. Bruderek, M. Sironi, L. Zimmer, B. Bottazzi, A. Mantovani, et al. 2011. Failure to detect production of IL-10 by activated human neutrophils. *Nat. Immunol.* 12:1017–1018, author reply :1018–1020. http://dx.doi.org/ 10.1038/ni.2111
- De Santo, C., R. Arscott, S. Booth, I. Karydis, M. Jones, R. Asher, M. Salio, M. Middleton, and V. Cerundolo. 2010. Invariant NKT cells modulate the suppressive activity of IL-10-secreting neutrophils differentiated with serum amyloid A. *Nat. Immunol.* 11:1039–1046. http://dx.doi.org/ 10.1038/ni.1942
- Demers, M., D.S. Krause, D. Schatzberg, K. Martinod, J.R. Voorhees, T.A. Fuchs, D.T. Scadden, and D.D. Wagner. 2012. Cancers predispose neutrophils to release extracellular DNA traps that contribute to cancerassociated thrombosis. *Proc. Natl. Acad. Sci. USA*. 109:13076–13081. http://dx.doi.org/10.1073/pnas.1200419109

- Diana, J., Y. Simoni, L. Furio, L. Beaudoin, B. Agerberth, F. Barrat, and A. Lehuen. 2013. Crosstalk between neutrophils, B-1a cells and plasmacy-toid dendritic cells initiates autoimmune diabetes. *Nat. Med.* 19:65–73. http://dx.doi.org/10.1038/nm.3042
- Dilulio, N.A., T. Engeman, D. Armstrong, C. Tannenbaum, T.A. Hamilton, and R.L. Fairchild. 1999. Groalpha-mediated recruitment of neutrophils is required for elicitation of contact hypersensitivity. *Eur. J. Immunol.* 29:3485–3495.
- Döring, Y., M. Drechsler, S. Wantha, K. Kemmerich, D. Lievens, S. Vijayan, R.L. Gallo, C. Weber, and O. Soehnlein. 2012a. Lack of neutrophilderived CRAMP reduces atherosclerosis in mice. *Circ. Res.* 110:1052– 1056. http://dx.doi.org/10.1161/CIRCRESAHA.112.265868
- Döring, Y., H.D. Manthey, M. Drechsler, D. Lievens, R.T. Megens, O. Soehnlein, M. Busch, M. Manca, R.R. Koenen, J. Pelisek, et al. 2012b. Auto-antigenic protein-DNA complexes stimulate plasmacytoid dendritic cells to promote atherosclerosis. *Circulation*. 125:1673–1683. http:// dx.doi.org/10.1161/CIRCULATIONAHA.111.046755
- Drechsler, M., R.T. Megens, M. van Zandvoort, C. Weber, and O. Soehnlein. 2010. Hyperlipidemia-triggered neutrophilia promotes early atherosclerosis. *Circulation*. 122:1837–1845. http://dx.doi.org/10.1161/ CIRCULATIONAHA.110.961714
- Duffy, D., H. Perrin, V. Abadie, N. Benhabiles, A. Boissonnas, C. Liard, B. Descours, D. Reboulleau, O. Bonduelle, B. Verrier, et al. 2012. Neutrophils transport antigen from the dermis to the bone marrow, initiating a source of memory CD8+ T cells. *Immunity*. 37:917–929. http:// dx.doi.org/10.1016/j.immuni.2012.07.015
- Dzhagalov, I., A. St John, and Y.W. He. 2007. The antiapoptotic protein Mcl-1 is essential for the survival of neutrophils but not macrophages. *Blood*. 109:1620–1626. http://dx.doi.org/10.1182/blood-2006-03-013771
- El Kebir, D., and J.G. Filep. 2010. Role of neutrophil apoptosis in the resolution of inflammation. *ScientificWorldJournal*. 10:1731–1748. http://dx.doi.org/ 10.1100/tsw.2010.169
- Elgazar-Carmon, V., A. Rudich, N. Hadad, and R. Levy. 2008. Neutrophils transiently infiltrate intra-abdominal fat early in the course of highfat feeding. J. Lipid Res. 49:1894–1903. http://dx.doi.org/10.1194/jlr .M800132-JLR.200
- Esmon, C.T. 2004. Interactions between the innate immune and blood coagulation systems. *Trends Immunol.* 25:536–542. http://dx.doi.org/ 10.1016/j.it.2004.08.003
- Essin, K., M. Gollasch, S. Rolle, P. Weissgerber, M. Sausbier, E. Bohn, I.B. Autenrieth, P. Ruth, F.C. Luft, W.M. Nauseef, and R. Kettritz. 2009. BK channels in innate immune functions of neutrophils and macrophages. *Blood*. 113:1326–1331. http://dx.doi.org/10.1182/blood-2008-07-166660
- Femling, J.K., V.V. Cherny, D. Morgan, B. Rada, A.P. Davis, G. Czirják, P. Enyedi, S.K. England, J.G. Moreland, E. Ligeti, et al. 2006. The antibacterial activity of human neutrophils and eosinophils requires proton channels but not BK channels. J. Gen. Physiol. 127:659–672. http://dx.doi.org/ 10.1085/jgp.200609504
- Finkelman, F.D. 2007. Anaphylaxis: lessons from mouse models. J. Allergy Clin. Immunol. 120:506–515.
- Fridlender, Z.G., and S.M. Albelda. 2012. Tumor-associated neutrophils: friend or foe? *Carcinogenesis*. 33:949–955. http://dx.doi.org/ 10.1093/carcin/bgs123
- Fridlender, Z.G., J. Sun, S. Kim, V. Kapoor, G. Cheng, L. Ling, G.S. Worthen, and S.M. Albelda. 2009. Polarization of tumor-associated neutrophil phenotype by TGF-β: "N1" versus "N2" TAN. *Cancer Cell*. 16:183–194. http://dx.doi.org/10.1016/j.ccr.2009.06.017
- Fuchs, T.A., U. Abed, C. Goosmann, R. Hurwitz, I. Schulze, V. Wahn, Y. Weinrauch, V. Brinkmann, and A. Zychlinsky. 2007. Novel cell death program leads to neutrophil extracellular traps. J. Cell Biol. 176:231– 241. http://dx.doi.org/10.1083/jcb.200606027
- Fuchs, T.A., A. Brill, D. Duerschmied, D. Schatzberg, M. Monestier, D.D. Myers Jr., S.K. Wrobleski, T.W. Wakefield, J.H. Hartwig, and D.D. Wagner. 2010. Extracellular DNA traps promote thrombosis. *Proc. Natl. Acad. Sci. USA.* 107:15880–15885. http://dx.doi.org/ 10.1073/pnas.1005743107
- Futosi, K., S. Fodor, and A. Mócsai. 2013. Neutrophil cell surface receptors and their intracellular signal transduction pathways. *Int. Immunopharmacol.* In press.

- Gabrilovich, D.I., and S. Nagaraj. 2009. Myeloid-derived suppressor cells as regulators of the immune system. *Nat. Rev. Immunol.* 9:162–174. http://dx.doi.org/10.1038/nri2506
- Garcia-Romo, G.S., S. Caielli, B. Vega, J. Connolly, F. Allantaz, Z. Xu, M. Punaro, J. Baisch, C. Guiducci, R.L. Coffman, et al. 2011. Netting neutrophils are major inducers of type I IFN production in pediatric systemic lupus erythematosus. *Sci. Transl. Med.* 3:73ra20. http://dx.doi.org/ 10.1126/scitranslmed.3001201
- Geering, B., and H.U. Simon. 2011. Peculiarities of cell death mechanisms in neutrophils. Cell Death Differ. 18:1457–1469. http://dx.doi.org/ 10.1038/cdd.2011.75
- Geiszt, M., A. Kapus, K. Német, L. Farkas, and E. Ligeti. 1997. Regulation of capacitative Ca²⁺ influx in human neutrophil granulocytes. Alterations in chronic granulomatous disease. J. Biol. Chem. 272:26471–26478. http://dx.doi.org/10.1074/jbc.272.42.26471
- Geiszt, M., A. Kapus, and E. Ligeti. 2001. Chronic granulomatous disease: more than the lack of superoxide? J. Leukoc. Biol. 69:191–196.
- Granot, Z., E. Henke, E.A. Comen, T.A. King, L. Norton, and R. Benezra. 2011. Tumor entrained neutrophils inhibit seeding in the premetastatic lung. *Cancer Cell*. 20:300–314. http://dx.doi.org/10 .1016/j.ccr.2011.08.012
- Gregor, M.F., and G.S. Hotamisligil. 2011. Inflammatory mechanisms in obesity. Annu. Rev. Immunol. 29:415–445. http://dx.doi.org/10.1146/ annurev-immunol-031210-101322
- Gregory, A.D., and A.M. Houghton. 2011. Tumor-associated neutrophils: new targets for cancer therapy. *Cancer Res.* 71:2411–2416. http://dx.doi .org/10.1158/0008-5472.CAN-10-2583
- Hakkim, A., B.G. Fürnrohr, K. Amann, B. Laube, U.A. Abed, V. Brinkmann, M. Herrmann, R.E. Voll, and A. Zychlinsky. 2010. Impairment of neutrophil extracellular trap degradation is associated with lupus nephritis. *Proc. Natl. Acad. Sci. USA*. 107:9813–9818. http:// dx.doi.org/10.1073/pnas.0909927107
- Hansson, G.K. 2005. Inflammation, atherosclerosis, and coronary artery disease. N. Engl. J. Med. 352:1685–1695. http://dx.doi.org/10.1056/ NEJMra043430
- Hartl, D., S. Krauss-Etschmann, B. Koller, P.L. Hordijk, T.W. Kuijpers, F. Hoffmann, A. Hector, E. Eber, V. Marcos, I. Bittmann, et al. 2008. Infiltrated neutrophils acquire novel chemokine receptor expression and chemokine responsiveness in chronic inflammatory lung diseases. J. Immunol. 181:8053–8067.
- Hock, H., M.J. Hamblen, H.M. Rooke, D. Traver, R.T. Bronson, S. Cameron, and S.H. Orkin. 2003. Intrinsic requirement for zinc finger transcription factor Gfi-1 in neutrophil differentiation. *Immunity*. 18: 109–120. http://dx.doi.org/10.1016/S1074-7613(02)00501-0
- Houghton, A.M., D.M. Rzymkiewicz, H. Ji, A.D. Gregory, E.E. Egea, H.E. Metz, D.B. Stolz, S.R. Land, L.A. Marconcini, C.R. Kliment, et al. 2010. Neutrophil elastase-mediated degradation of IRS-1 accelerates lung tumor growth. *Nat. Med.* 16:219–223. http://dx.doi.org/ 10.1038/nm.2084
- Huard, B., T. McKee, C. Bosshard, S. Durual, T. Matthes, S. Myit, O. Donze, C. Frossard, C. Chizzolini, C. Favre, et al. 2008. APRIL secreted by neutrophils binds to heparan sulfate proteoglycans to create plasma cell niches in human mucosa. J. Clin. Invest. 118:2887–2895.
- Huo, Y., A. Schober, S.B. Forlow, D.F. Smith, M.C. Hyman, S. Jung, D.R. Littman, C. Weber, and K. Ley. 2003. Circulating activated platelets exacerbate atherosclerosis in mice deficient in apolipoprotein E. *Nat. Med.* 9:61–67. http://dx.doi.org/10.1038/nm810
- Inoue, Y., T. Kaifu, A. Sugahara-Tobinai, A. Nakamura, J. Miyazaki, and T. Takai. 2007. Activating Fc γ receptors participate in the development of autoimmune diabetes in NOD mice. *J. Immunol.* 179:764–774.
- Ionita, M.G., P. van den Borne, L.M. Catanzariti, F.L. Moll, J.P. de Vries, G. Pasterkamp, A. Vink, and D.P. de Kleijn. 2010. High neutrophil numbers in human carotid atherosclerotic plaques are associated with characteristics of rupture-prone lesions. *Arterioscler. Thromb. Vasc. Biol.* 30:1842–1848. http://dx.doi.org/10.1161/ATVBAHA.110.209296
- Jablonska, J., S. Leschner, K. Westphal, S. Lienenklaus, and S. Weiss. 2010. Neutrophils responsive to endogenous IFN-β regulate tumor angiogenesis and growth in a mouse tumor model. *J. Clin. Invest.* 120:1151– 1164. http://dx.doi.org/10.1172/JCI37223

- Jaeger, B.N., J. Donadieu, C. Cognet, C. Bernat, D. Ordoñez-Rueda, V. Barlogis, N. Mahlaoui, A. Fenis, E. Narni-Mancinelli, B. Beaupain, et al. 2012. Neutrophil depletion impairs natural killer cell maturation, function, and homeostasis. *J. Exp. Med.* 209:565–580. http://dx.doi .org/10.1084/jem.20111908
- Jenne, C.N., C.H. Wong, F.J. Zemp, B. McDonald, M.M. Rahman, P.A. Forsyth, G. McFadden, and P. Kubes. 2013. Neutrophils recruited to sites of infection protect from virus challenge by releasing neutrophil extracellular traps. *Cell Host Microbe*. 13:169–180. http://dx.doi.org/ 10.1016/j.chom.2013.01.005
- John, B., and C.A. Hunter. 2008. Immunology. Neutrophil soldiers or Trojan Horses? Science. 321:917–918. http://dx.doi.org/10.1126/science .1162914
- Johnson, R.C., S.M. Chapman, Z.M. Dong, J.M. Ordovas, T.N. Mayadas, J. Herz, R.O. Hynes, E.J. Schaefer, and D.D. Wagner. 1997. Absence of P-selectin delays fatty streak formation in mice. J. Clin. Invest. 99:1037–1043. http://dx.doi.org/10.1172/JCI119231
- Jonsson, H., P. Allen, and S.L. Peng. 2005. Inflammatory arthritis requires Foxo3a to prevent Fas ligand-induced neutrophil apoptosis. *Nat. Med.* 11:666–671. http://dx.doi.org/10.1038/nm1248
- Jönsson, F., D.A. Mancardi, Y. Kita, H. Karasuyama, B. Iannascoli, N. Van Rooijen, T. Shimizu, M. Daëron, and P. Bruhns. 2011. Mouse and human neutrophils induce anaphylaxis. J. Clin. Invest. 121:1484–1496. http://dx.doi.org/10.1172/JCI45232
- Jönsson, F., D.A. Mancardi, W. Zhao, Y. Kita, B. Iannascoli, H. Khun, N. van Rooijen, T. Shimizu, L.B. Schwartz, M. Daëron, and P. Bruhns. 2012. Human FcγRIIA induces anaphylactic and allergic reactions. *Blood.* 119:2533–2544. http://dx.doi.org/10.1182/blood-2011-07-367334
- Kaplan, M.J. 2011. Neutrophils in the pathogenesis and manifestations of SLE. Nat. Rev. Rheumatol. 7:691–699. http://dx.doi.org/10.1038/ nrrheum.2011.132
- Kaplan, D.H., B.Z. Igyártó, and A.A. Gaspari. 2012. Early immune events in the induction of allergic contact dermatitis. *Nat. Rev. Immunol.* 12:114–124.
- Karsunky, H., H. Zeng, T. Schmidt, B. Zevnik, R. Kluge, K.W. Schmid, U. Dührsen, and T. Möröy. 2002. Inflammatory reactions and severe neutropenia in mice lacking the transcriptional repressor Gfi1. *Nat. Genet.* 30:295–300. http://dx.doi.org/10.1038/ng831
- Kessenbrock, K., M. Krumbholz, U. Schönermarck, W. Back, W.L. Gross, Z. Werb, H.J. Gröne, V. Brinkmann, and D.E. Jenne. 2009. Netting neutrophils in autoimmune small-vessel vasculitis. *Nat. Med.* 15:623– 625. http://dx.doi.org/10.1038/nm.1959
- Khan, B.Q., and S.F. Kemp. 2011. Pathophysiology of anaphylaxis. Curr. Opin. Allergy Clin. Immunol. 11:319–325. http://dx.doi.org/10 .1097/ACI.0b013e3283481ab6
- Khandpur, R., C. Carmona-Rivera, A. Vivekanandan-Giri, A. Gizinski, S. Yalavarthi, J.S. Knight, S. Friday, S. Li, R.M. Patel, V. Subramanian, et al. 2013. NETs are a source of citrullinated autoantigens and stimulate inflammatory responses in rheumatoid arthritis. *Sci. Transl. Med.* 5:178ra40. http://dx.doi.org/10.1126/scitranslmed.3005580
- Lande, R., J. Gregorio, V. Facchinetti, B. Chatterjee, Y.H. Wang, B. Homey, W. Cao, Y.H. Wang, B. Su, F.O. Nestle, et al. 2007. Plasmacytoid dendritic cells sense self-DNA coupled with antimicrobial peptide. *Nature*. 449:564–569. http://dx.doi.org/10.1038/nature06116
- Lande, R., D. Ganguly, V. Facchinetti, L. Frasca, C. Conrad, J. Gregorio, S. Meller, G. Chamilos, R. Sebasigari, V. Riccieri, et al. 2011. Neutrophils activate plasmacytoid dendritic cells by releasing self-DNA-peptide complexes in systemic lupus erythematosus. *Sci. Transl. Med.* 3:73ra19. http://dx.doi.org/10.1126/scitranslmed.3001180
- Leto, T.L., and M. Geiszt. 2006. Role of Nox family NADPH oxidases in host defense. Antioxid. Redox Signal. 8:1549–1561. http://dx.doi.org/ 10.1089/ars.2006.8.1549
- Ley, K., C. Laudanna, M.I. Cybulsky, and S. Nourshargh. 2007. Getting to the site of inflammation: the leukocyte adhesion cascade updated. *Nat. Rev. Immunol.* 7:678–689. http://dx.doi.org/10.1038/nri2156
- Liu, F., H.Y.Wu, R. Wesselschmidt, T. Kornaga, and D.C. Link. 1996. Impaired production and increased apoptosis of neutrophils in granulocyte colonystimulating factor receptor-deficient mice. *Immunity*. 5:491–501. http:// dx.doi.org/10.1016/S1074-7613(00)80504-X

- Lowe, D.M., P.S. Redford, R.J. Wilkinson, A. O'Garra, and A.R. Martineau. 2012. Neutrophils in tuberculosis: friend or foe? *Trends Immunol*. 33:14–25. http://dx.doi.org/10.1016/j.it.2011.10.003
- Maletto, B.A., A.S. Ropolo, D.O. Alignani, M.V. Liscovsky, R.P. Ranocchia, V.G. Moron, and M.C. Pistoresi-Palencia. 2006. Presence of neutrophil-bearing antigen in lymphoid organs of immune mice. *Blood*. 108:3094–3102. http://dx.doi.org/10.1182/blood-2006-04-016659
- Mantovani, A., M.A. Cassatella, C. Costantini, and S. Jaillon. 2011. Neutrophils in the activation and regulation of innate and adaptive immunity. *Nat. Rev. Immunol.* 11:519–531. http://dx.doi.org/10.1038/ nri3024
- Martineau, A.R., S.M. Newton, K.A. Wilkinson, B. Kampmann, B.M. Hall, N. Nawroly, G.E. Packe, R.N. Davidson, C.J. Griffiths, and R.J. Wilkinson. 2007. Neutrophil-mediated innate immune resistance to mycobacteria. J. Clin. Invest. 117:1988–1994. http://dx.doi.org/ 10.1172/JCI31097
- Massberg, S., K. Brand, S. Grüner, S. Page, E. Müller, I. Müller, W. Bergmeier, T. Richter, M. Lorenz, I. Konrad, et al. 2002. A critical role of platelet adhesion in the initiation of atherosclerotic lesion formation. J. Exp. Med. 196:887–896. http://dx.doi.org/10.1084/jem .20012044
- Massberg, S., L. Grahl, M.L. von Bruehl, D. Manukyan, S. Pfeiler, C. Goosmann, V. Brinkmann, M. Lorenz, K. Bidzhekov, A.B. Khandagale, et al. 2010. Reciprocal coupling of coagulation and innate immunity via neutrophil serine proteases. *Nat. Med.* 16:887–896. http://dx.doi .org/10.1038/nm.2184
- Maugeri, N., G. de Gaetano, A. Carbone, M.B. Donati, and C. Cerletti. 2005. More on: tissue factor in neutrophils. J. Thromb. Haemost. 3:1114. http://dx.doi.org/10.1111/j.1538-7836.2005.01320.x
- Maugeri, N., M. Brambilla, M. Camera, A. Carbone, E. Tremoli, M.B. Donati, G. de Gaetano, and C. Cerletti. 2006. Human polymorphonuclear leukocytes produce and express functional tissue factor upon stimulation. J. Thromb. Haemost. 4:1323–1330. http://dx.doi.org/ 10.1111/j.1538-7836.2006.01968.x
- McDonald, B., K. Pittman, G.B. Menezes, S.A. Hirota, I. Slaba, C.C. Waterhouse, P.L. Beck, D.A. Muruve, and P. Kubes. 2010. Intravascular danger signals guide neutrophils to sites of sterile inflammation. *Science*. 330:362–366. http://dx.doi.org/10.1126/science.1195491
- McDonald, B., R. Urrutia, B.G. Yipp, C.N. Jenne, and P. Kubes. 2012. Intravascular neutrophil extracellular traps capture bacteria from the bloodstream during sepsis. *Cell Host Microbe*. 12:324–333. http://dx.doi .org/10.1016/j.chom.2012.06.011
- Megiovanni, A.M., F. Sanchez, M. Robledo-Sarmiento, C. Morel, J.C. Gluckman, and S. Boudaly. 2006. Polymorphonuclear neutrophils deliver activation signals and antigenic molecules to dendritic cells: a new link between leukocytes upstream of T lymphocytes. J. Leukoc. Biol. 79:977–988. http://dx.doi.org/10.1189/jlb.0905526
- Menegazzi, R., E. Decleva, and P. Dri. 2012. Killing by neutrophil extracellular traps: fact or folklore? *Blood*. 119:1214–1216. http://dx.doi .org/10.1182/blood-2011-07-364604
- Mestas, J., and C.C. Hughes. 2004. Of mice and not men: differences between mouse and human immunology. J. Immunol. 172:2731–2738.
- Nageh, M.F., E.T. Sandberg, K.R. Marotti, A.H. Lin, E.P. Melchior, D.C. Bullard, and A.L. Beaudet. 1997. Deficiency of inflammatory cell adhesion molecules protects against atherosclerosis in mice. *Arterioscler. Thromb. Vasc. Biol.* 17:1517–1520. http://dx.doi.org/10.1161/01 .ATV.17.8.1517
- Nandi, B., and S.M. Behar. 2011. Regulation of neutrophils by interferon-γ limits lung inflammation during tuberculosis infection. J. Exp. Med. 208:2251–2262. http://dx.doi.org/10.1084/jem.20110919
- Naruko, T., M. Ueda, K. Haze, A.C. van der Wal, C.M. van der Loos, A. Itoh, R. Komatsu, Y. Ikura, M. Ogami, Y. Shimada, et al. 2002. Neutrophil infiltration of culprit lesions in acute coronary syndromes. *Circulation*. 106:2894–2900. http://dx.doi.org/10.1161/01 .CIR.0000042674.89762.20
- Nathan, C. 2006. Neutrophils and immunity: challenges and opportunities. Nat. Rev. Immunol. 6:173–182. http://dx.doi.org/10.1038/nri1785
- Nauseef, W.M. 2012. Editorial: Nyet to NETs? A pause for healthy skepticism. J. Leukoc. Biol. 91:353–355. http://dx.doi.org/10.1189/jlb.1011495

- Németh, T., and A. Mócsai. 2012. The role of neutrophils in autoimmune diseases. *Immunol. Lett.* 143:9–19. http://dx.doi.org/10.1016/ j.imlet.2012.01.013
- Nordenfelt, P., and H. Tapper. 2011. Phagosome dynamics during phagocytosis by neutrophils. J. Leukoc. Biol. 90:271–284. http://dx.doi.org/ 10.1189/jlb.0810457
- Nourshargh, S., P.L. Hordijk, and M. Sixt. 2010. Breaching multiple barriers: leukocyte motility through venular walls and the interstitium. *Nat. Rev. Mol. Cell Biol.* 11:366–378. http://dx.doi.org/10.1038/nrm2889
- Obermoser, G., and V. Pascual. 2010. The interferon-α signature of systemic lupus erythematosus. *Lupus*. 19:1012–1019. http://dx.doi.org/ 10.1177/0961203310371161
- Oehler, L., O. Majdic, W.F. Pickl, J. Stöckl, E. Riedl, J. Drach, K. Rappersberger, K. Geissler, and W. Knapp. 1998. Neutrophil granulocyte-committed cells can be driven to acquire dendritic cell characteristics. J. Exp. Med. 187:1019–1028. http://dx.doi.org/10.1084/jem .187.7.1019
- Ohnmacht, C., C. Schwartz, M. Panzer, I. Schiedewitz, R. Naumann, and D. Voehringer. 2010. Basophils orchestrate chronic allergic dermatitis and protective immunity against helminths. *Immunity*. 33:364–374. http://dx.doi.org/10.1016/j.immuni.2010.08.011
- Olefsky, J.M., and C.K. Glass. 2010. Macrophages, inflammation, and insulin resistance. Annu. Rev. Physiol. 72:219–246. http://dx.doi.org/ 10.1146/annurev-physiol-021909-135846
- Ordoñez-Rueda, D., F. Jönsson, D.A. Mancardi, W. Zhao, A. Malzac, Y. Liang, E. Bertosio, P. Grenot, V. Blanquet, S. Sabrautzki, et al. 2012. A hypomorphic mutation in the Gfi1 transcriptional repressor results in a novel form of neutropenia. *Eur. J. Immunol.* 42:2395–2408. http://dx.doi.org/10.1002/eji.201242589
- Oyoshi, M.K., R. He,Y. Li, S. Mondal, J.Yoon, R. Afshar, M. Chen, D.M. Lee, H.R. Luo, A.D. Luster, et al. 2012. Leukotriene B₄-driven neutrophil recruitment to the skin is essential for allergic skin inflammation. *Immunity*. 37:747–758. http://dx.doi.org/10.1016/j.immuni.2012.06.018
- Parker, H., A.M. Albrett, A.J. Kettle, and C.C. Winterbourn. 2012a. Myeloperoxidase associated with neutrophil extracellular traps is active and mediates bacterial killing in the presence of hydrogen peroxide. J. Leukoc. Biol. 91:369–376. http://dx.doi.org/10.1189/jlb.0711387
- Parker, H., M. Dragunow, M.B. Hampton, A.J. Kettle, and C.C. Winterbourn. 2012b. Requirements for NADPH oxidase and myeloperoxidase in neutrophil extracellular trap formation differ depending on the stimulus. *J. Leukoc. Biol.* 92:841–849. http://dx.doi.org/10 .1189/jlb.1211601
- Pekarek, L.A., B.A. Starr, A.Y. Toledano, and H. Schreiber. 1995. Inhibition of tumor growth by elimination of granulocytes. J. Exp. Med. 181:435– 440. http://dx.doi.org/10.1084/jem.181.1.435
- Person, R.E., F.Q. Li, Z. Duan, K.F. Benson, J. Wechsler, H.A. Papadaki, G. Eliopoulos, C. Kaufman, S.J. Bertolone, B. Nakamoto, et al. 2003. Mutations in proto-oncogene GFI1 cause human neutropenia and target ELA2. *Nat. Genet.* 34:308–312. http://dx.doi.org/10.1038/ng1170
- Peters, N.C., J.G. Egen, N. Secundino, A. Debrabant, N. Kimblin, S. Kamhawi, P. Lawyer, M.P. Fay, R.N. Germain, and D. Sacks. 2008. In vivo imaging reveals an essential role for neutrophils in leishmaniasis transmitted by sand flies. *Science*. 321:970–974. http://dx.doi.org/10.1126/science.1159194
- Pillay, J., I. den Braber, N. Vrisekoop, L.M. Kwast, R.J. de Boer, J.A. Borghans, K. Tesselaar, and L. Koenderman. 2010. In vivo labeling with ²H₂O reveals a human neutrophil lifespan of 5.4 days. *Blood.* 116:625– 627. http://dx.doi.org/10.1182/blood-2010-01-259028
- Pillay, J., V.M. Kamp, E. van Hoffen, T. Visser, T. Tak, J.W. Lammers, L.H. Ulfman, L.P. Leenen, P. Pickkers, and L. Koenderman. 2012. A subset of neutrophils in human systemic inflammation inhibits T cell responses through Mac-1. J. Clin. Invest. 122:327–336. http://dx.doi.org/10.1172/ JCI57990
- Porcherie, A., C. Mathieu, R. Peronet, E. Schneider, J. Claver, P.H. Commere, H. Kiefer-Biasizzo, H. Karasuyama, G. Milon, M. Dy, et al. 2011. Critical role of the neutrophil-associated high-affinity receptor for IgE in the pathogenesis of experimental cerebral malaria. J. Exp. Med. 208:2225–2236. http://dx.doi.org/10.1084/jem.20110845
- Proebstl, D., M.B. Voisin, A. Woodfin, J. Whiteford, F. D'Acquisto, G.E. Jones, D. Rowe, and S. Nourshargh. 2012. Pericytes support neutrophil

subendothelial cell crawling and breaching of venular walls in vivo. J. Exp. Med. 209:1219–1234. http://dx.doi.org/10.1084/jem.20111622

- Puga, I., M. Cols, C.M. Barra, B. He, L. Cassis, M. Gentile, L. Comerma, A. Chorny, M. Shan, W. Xu, et al. 2011. B cell-helper neutrophils stimulate the diversification and production of immunoglobulin in the marginal zone of the spleen. *Nat. Immunol.* 13:170–180. http://dx.doi.org/ 10.1038/ni.2194
- Rada, B.K., M. Geiszt, K. Káldi, C. Timár, and E. Ligeti. 2004. Dual role of phagocytic NADPH oxidase in bacterial killing. *Blood*. 104:2947–2953. http://dx.doi.org/10.1182/blood-2004-03-1005
- Rada, B.K., M. Geiszt, C. Hably, and E. Ligeti. 2005. Consequences of the electrogenic function of the phagocytic NADPH oxidase. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* 360:2293–2300. http://dx.doi.org/10 .1098/rstb.2005.1768
- Reeves, E.P., H. Lu, H.L. Jacobs, C.G. Messina, S. Bolsover, G. Gabella, E.O. Potma, A. Warley, J. Roes, and A.W. Segal. 2002. Killing activity of neutrophils is mediated through activation of proteases by K⁺ flux. *Nature*. 416:291–297. http://dx.doi.org/10.1038/416291a
- Rensen, S.S., Y. Slaats, J. Nijhuis, A. Jans, V. Bieghs, A. Driessen, E. Malle, J.W. Greve, and W.A. Buurman. 2009. Increased hepatic myeloperoxidase activity in obese subjects with nonalcoholic steatohepatitis. *Am. J. Pathol.* 175:1473–1482. http://dx.doi.org/10.2353/ajpath.2009.080999
- Rotzius, P., S. Thams, O. Soehnlein, E. Kenne, C.N. Tseng, N.K. Björkström, K.J. Malmberg, L. Lindbom, and E.E. Eriksson. 2010. Distinct infiltration of neutrophils in lesion shoulders in ApoE^{-/-} mice. Am. J. Pathol. 177:493–500. http://dx.doi.org/10.2353/ajpath.2010.090480
- Saitoh, T., J. Komano, Y. Saitoh, T. Misawa, M. Takahama, T. Kozaki, T. Uehata, H. Iwasaki, H. Omori, S. Yamaoka, et al. 2012. Neutrophil extracellular traps mediate a host defense response to human immunodeficiency virus-1. *Cell Host Microbe.* 12:109–116. http://dx.doi.org/ 10.1016/j.chom.2012.05.015
- Scapini, P., B. Nardelli, G. Nadali, F. Calzetti, G. Pizzolo, C. Montecucco, and M.A. Cassatella. 2003. G-CSF-stimulated neutrophils are a prominent source of functional BLyS. J. Exp. Med. 197:297–302. http:// dx.doi.org/10.1084/jem.20021343
- Scapini, P., F. Bazzoni, and M.A. Cassatella. 2008. Regulation of B-cellactivating factor (BAFF)/B lymphocyte stimulator (BLyS) expression in human neutrophils. *Immunol. Lett.* 116:1–6. http://dx.doi.org/10.1016/ j.imlet.2007.11.009
- Schrenzel, J., L. Serrander, B. Bánfi, O. Nüsse, R. Fouyouzi, D.P. Lew, N. Demaurex, and K.H. Krause. 1998. Electron currents generated by the human phagocyte NADPH oxidase. *Nature*. 392:734–737. http:// dx.doi.org/10.1038/33725
- Schwaller, J., P. Schneider, P. Mhawech-Fauceglia, T. McKee, S. Myit, T. Matthes, J. Tschopp, O. Donze, F.A. Le Gal, and B. Huard. 2007. Neutrophil-derived APRIL concentrated in tumor lesions by proteoglycans correlates with human B-cell lymphoma aggressiveness. *Blood*. 109:331–338. http://dx.doi.org/10.1182/blood-2006-02-001800
- Seok, J., H.S. Warren, A.G. Cuenca, M.N. Mindrinos, H.V. Baker, W. Xu, D.R. Richards, G.P. McDonald-Smith, H. Gao, L. Hennessy, et al; Inflammation and Host Response to Injury, Large Scale Collaborative Research Program. 2013. Genomic responses in mouse models poorly mimic human inflammatory diseases. *Proc. Natl. Acad. Sci. USA*. 110:3507–3512. http://dx.doi.org/10.1073/pnas.1222878110
- Serhan, C.N. 2007. Resolution phase of inflammation: novel endogenous anti-inflammatory and proresolving lipid mediators and pathways. *Annu. Rev. Immunol.* 25:101–137. http://dx.doi.org/10.1146/annurev .immunol.25.022106.141647
- Serhan, C.N., N. Chiang, and T.E. Van Dyke. 2008. Resolving inflammation: dual anti-inflammatory and pro-resolution lipid mediators. Nat. Rev. Immunol. 8:349–361. http://dx.doi.org/10.1038/nri2294
- Soehnlein, O. 2009. Direct and alternative antimicrobial mechanisms of neutrophil-derived granule proteins. J. Mol. Med. 87:1157–1164. http://dx.doi.org/10.1007/s00109-009-0508-6
- Soehnlein, O., and L. Lindbom. 2010. Phagocyte partnership during the onset and resolution of inflammation. *Nat. Rev. Immunol.* 10:427–439. http://dx.doi.org/10.1038/nri2779
- Soehnlein, O., S. Wantha, S. Simsekyilmaz, Y. Döring, R.T. Megens, S.F. Mause, M. Drechsler, R. Smeets, S. Weinandy, F. Schreiber, et al. 2011.

Neutrophil-derived cathelicidin protects from neointimal hyperplasia. *Sci. Transl. Med.* 3:103ra98. http://dx.doi.org/10.1126/ scitranslmed.3002531

- Spörri, R., N. Joller, H. Hilbi, and A. Oxenius. 2008. A novel role for neutrophils as critical activators of NK cells. J. Immunol. 181:7121–7130.
- Stark, K., A. Eckart, S. Haidari, A. Tirniceriu, M. Lorenz, M.L. von Brühl, F. Gärtner, A.G. Khandoga, K.R. Legate, R. Pless, et al. 2013. Capillary and arteriolar pericytes attract innate leukocytes exiting through venules and 'instruct' them with pattern-recognition and motility programs. *Nat. Immunol.* 14:41–51. http://dx.doi.org/10.1038/ni.2477
- Takai, T. 2005. Fc receptors and their role in immune regulation and autoimmunity. J. Clin. Immunol. 25:1–18. http://dx.doi.org/10.1007/ s10875-005-0353-8
- Talukdar, S., Y. Oh, G. Bandyopadhyay, D. Li, J. Xu, J. McNelis, M. Lu, P. Li, Q. Yan, Y. Zhu, et al. 2012. Neutrophils mediate insulin resistance in mice fed a high-fat diet through secreted elastase. *Nat. Med.* 18:1407–1412. http://dx.doi.org/10.1038/nm.2885
- Timár, C.I., A.M. Lőrincz, R. Csépányi-Kömi, A. Vályi-Nagy, G. Nagy, E.I. Buzás, Z. Iványi, A. Kittel, D.W. Powell, K.R. McLeish, and E. Ligeti. 2013. Antibacterial effect of microvesicles released from human neutrophilic granulocytes. *Blood.* 121:510–518. http://dx.doi.org/10.1182/ blood-2012-05-431114
- Tsujimura, Y., K. Obata, K. Mukai, H. Shindou, M. Yoshida, H. Nishikado, Y. Kawano, Y. Minegishi, T. Shimizu, and H. Karasuyama. 2008. Basophils play a pivotal role in immunoglobulin-G-mediated but not immunoglobulin-E-mediated systemic anaphylaxis. *Immunity*. 28:581– 589. http://dx.doi.org/10.1016/j.immuni.2008.02.008
- van Gisbergen, K.P., T.B. Geijtenbeek, and Y. van Kooyk. 2005a. Close encounters of neutrophils and DCs. *Trends Immunol.* 26:626–631. http://dx.doi.org/10.1016/j.it.2005.09.007
- van Gisbergen, K.P., M. Sanchez-Hernandez, T.B. Geijtenbeek, and Y. van Kooyk. 2005b. Neutrophils mediate immune modulation of dendritic cells through glycosylation-dependent interactions between Mac-1 and DC-SIGN. J. Exp. Med. 201:1281–1292. http://dx.doi.org/10 .1084/jem.20041276
- van Zandbergen, G., M. Klinger, A. Mueller, S. Dannenberg, A. Gebert, W. Solbach, and T. Laskay. 2004. Cutting edge: neutrophil granulocyte serves as a vector for Leishmania entry into macrophages. J. Immunol. 173:6521–6525.
- von Brühl, M.L., K. Stark, A. Steinhart, S. Chandraratne, I. Konrad, M. Lorenz, A. Khandoga, A. Tirniceriu, R. Coletti, M. Köllnberger, et al. 2012. Monocytes, neutrophils, and platelets cooperate to initiate and propagate venous thrombosis in mice in vivo. *J. Exp. Med.* 209:819–835. http://dx .doi.org/10.1084/jem.20112322
- von Hundelshausen, P., K.S. Weber, Y. Huo, A.E. Proudfoot, P.J. Nelson, K. Ley, and C. Weber. 2001. RANTES deposition by platelets triggers monocyte arrest on inflamed and atherosclerotic endothelium. *Circulation*. 103:1772–1777. http://dx.doi.org/10.1161/01.CIR.103.13.1772

- Weber, F.C., T. Nemeth, B. Ozsvari, E. Oswald, K. Futosi, L. Puskas, T. Jakob, S.F. Martin, and A. Mocsai. 2012. Neutrophils are crucial innate effector cells in contact hypersensitivity. *Eur. J. Clin. Invest.* 42:17. http://dx.doi.org/10.1111/j.1365-2362.2011.02549.x
- Witko-Sarsat, V., P. Rieu, B. Descamps-Latscha, P. Lesavre, and L. Halbwachs-Mecarelli. 2000. Neutrophils: molecules, functions and pathophysiological aspects. *Lab. Invest.* 80:617–653. http://dx.doi.org/ 10.1038/labinvest.3780067
- Witko-Sarsat, V., M. Pederzoli-Ribeil, E. Hirsch, S. Sozzani, and M.A. Cassatella. 2011. Regulating neutrophil apoptosis: new players enter the game. *Trends Immunol.* 32:117–124. http://dx.doi.org/10.1016/j.it .2011.01.001
- Woodfin, A., M.B. Voisin, M. Beyrau, B. Colom, D. Caille, F.M. Diapouli, G.B. Nash, T. Chavakis, S.M. Albelda, G.E. Rainger, et al. 2011. The junctional adhesion molecule JAM-C regulates polarized transendothelial migration of neutrophils in vivo. *Nat. Immunol.* 12:761–769. http:// dx.doi.org/10.1038/ni.2062
- Yang, C.W., and E.R. Unanue. 2013. Neutrophils control the magnitude and spread of the immune response in a thromboxane A2-mediated process. J. Exp. Med. 210:375–387. http://dx.doi.org/10.1084/jem .20122183
- Yang, D., G. de la Rosa, P. Tewary, and J.J. Oppenheim. 2009. Alarmins link neutrophils and dendritic cells. *Trends Immunol.* 30:531–537. http:// dx.doi.org/10.1016/j.it.2009.07.004
- Yang, X.D., W. Ai, S. Asfaha, G. Bhagat, R.A. Friedman, G. Jin, H. Park, B. Shykind, T.G. Diacovo, A. Falus, and T.C. Wang. 2011. Histamine deficiency promotes inflammation-associated carcinogenesis through reduced myeloid maturation and accumulation of CD11b⁺Ly6G⁺ immature myeloid cells. *Nat. Med.* 17:87–95. http:// dx.doi.org/10.1038/nm.2278
- Yang, C.T., C.J. Cambier, J.M. Davis, C.J. Hall, P.S. Crosier, and L. Ramakrishnan. 2012. Neutrophils exert protection in the early tuberculous granuloma by oxidative killing of mycobacteria phagocytosed from infected macrophages. *Cell Host Microbe*. 12:301–312. http://dx.doi .org/10.1016/j.chom.2012.07.009
- Yasutomo, K., T. Horiuchi, S. Kagami, H. Tsukamoto, C. Hashimura, M. Urushihara, and Y. Kuroda. 2001. Mutation of DNASE1 in people with systemic lupus erythematosus. *Nat. Genet.* 28:313–314. http://dx.doi .org/10.1038/91070
- Yipp, B.G., B. Petri, D. Salina, C.N. Jenne, B.N. Scott, L.D. Zbytnuik, K. Pittman, M. Asaduzzaman, K. Wu, H.C. Meijndert, et al. 2012. Infection-induced NETosis is a dynamic process involving neutrophil multitasking in vivo. *Nat. Med.* 18:1386–1393. http://dx.doi .org/10.1038/nm.2847
- Zhang, X., L. Majlessi, E. Deriaud, C. Leclerc, and R. Lo-Man. 2009. Coactivation of Syk kinase and MyD88 adaptor protein pathways by bacteria promotes regulatory properties of neutrophils. *Immunity*. 31:761–771. http://dx.doi.org/10.1016/j.immuni.2009.09.016