

Diversity and geographic distribution of dog tick species in Sri Lanka and the life cycle of brown dog tick, *Rhipicephalus sanguineus* under laboratory conditions

Kalpani Omantha Bandaranayaka

University of Peradeniya Faculty of Science

Upekha Indunil Dissanayake

University of Peradeniya Faculty of Science

Rupika Subashini Rajakaruna (✉ rupikar@pdn.ac.lk)

University of Peradeniya <https://orcid.org/0000-0001-7939-947X>

Research Article

Keywords: Diversity, Geographic Distribution, Life Cycle, Dog Ticks, Sri Lanka

Posted Date: February 15th, 2022

DOI: <https://doi.org/10.21203/rs.3.rs-721544/v2>

License: © ⓘ This work is licensed under a Creative Commons Attribution 4.0 International License. [Read Full License](#)

Abstract

Background: Tick infestations and tick-borne diseases have become a major emerging health concern of dogs in Sri Lanka. Information about tick species infesting dogs and their geographic distribution in Sri Lanka is largely unknown. Here we determined the tick species infesting the dogs and their distribution, and described the life cycle parameters of the dominant dog tick species under laboratory conditions.

Methods: An island-wide, cross-sectional survey of tick species infesting the domestic dog was carried out, and the life cycle of the major dog tick, *Rhipicephalus sanguineus* was studied under laboratory conditions.

Results: A total of 3,026 ticks were collected from 1,219 dogs of different breeds in all 25 districts in the three climatic zones: Wet, Dry, and Intermediate zones. Eight species in five genera were identified: *Rh. sanguineus* (63.4%), *Rhipicephalus haemaphysaloides* (22.0%), *Haemaphysalis bispinosa* (12.5%), *Haemaphysalis intermedia* (0.9%), *Haemaphysalis turturis* (0.6%), *Amblyomma integrum* (0.4%), *Dermacentor auratus* (0.2%) and *Hyalomma* sp (0.06%). The brown dog tick, *Rh. sanguineus* was the dominant species in the Dry and Wet zones, while *Rh. haemaphysaloides* was the dominant species in the Intermediate zone. Species diversity (presented as Shannon diversity index H) in the three was 1.135, 1.021 and 0.849 in Intermediate, Dry and Wet zones, respectively. Adults formed 94.7% with a significantly higher number of females, and the rest were nymphs. *Rhipicephalus sanguineus* preferred the anterior side of the host body, specifically the inner and outer side of the ear. In contrast, *Rh. haemaphysaloides* preferred the posterior side, mainly the fore and hind limbs. The three-host life cycle of *Rh. sanguineus* was completed within 70-126 days, all three stages successfully fed on the New Zealand white rabbits under laboratory conditions. The mean Reproductive Efficiency Index (REI) and Reproductive Fitness Index (RFI) were 50.8 ± 9.69 and 9.1 ± 5.01 , respectively. Larger females had higher reproductive success.

Conclusion: The dominant dog tick species and the species diversity varied in different climatic regions of Sri Lanka. The three-host life cycle of *Rh. sanguineus* was successfully completed on the New Zealand white rabbits under laboratory conditions. Information on diversity, distribution and life cycle parameters is fundamental for studies of canine tick-borne infections, zoonoses, and their epidemiology.

Background

Dog ticks are particularly important because dogs are companion animals with worldwide distribution, and their unrestricted movement may carry ticks and tick-borne pathogens to new habitats. Ticks transmit a wide variety of infectious agents to dogs. These diseases are of great concern because of costs for owners, suffering in dogs, and the risk of transmission of pathological agents from dogs to humans. Information on tick species infesting dogs is vital for the studies on tick-borne diseases and their control and prevention. Tick-borne diseases of dogs have recently become a significant focus of interest globally, especially in areas of the world in which they have traditionally been considered non-endemic. The concern that these diseases might become established in new geographical locations arose from the increased international mobility of dogs and increased contact of these animals with non-urban environments and wildlife disease reservoirs [1]. Ticks, especially the larval stage, can travel globally undetected with pets and other animal trade. Therefore, the potential impacts of increased movement between countries of people and global pet trade on the distribution of ticks is currently an area of increasing public health concern.

Historical records and recent studies in the British Isles show that the distribution of *Ixodes ricinus* in Great Britain has expanded by 17%, and abundances of ticks have increased in most of the locations surveyed [2, 3, 4]. Moreover, in Central Europe, *I. ricinus* has expanded its geographic range northwards and upwards to altitudes of up to 1100 m [5] wherein a study carried out in 1965, *I. ricinus* was not recorded above 700 m [6]. Together with the ticks, the distribution of tick-borne infections has also been expanded. For example, tick-borne encephalitis in mountainous regions of the Czech Republic [5] and the increase in human cases of Lyme disease by 30-fold between 1999 and 2008 in Scotland [7]. The expansion of the tick-borne infections among animals vector of canine babesiosis in Europe by the European meadow tick, *Dermacentor reticulatus* is extended northwards into Poland, Belgium, Germany, the Netherlands, and southern England [8, 9, 10, 11].

Among the tick species that infest dogs, *Rhipicephalus sanguineus*, the brown dog tick is the most commonly known species with a three-host life cycle. It feeds primarily on dogs and occasionally on other hosts, including humans. It has a wide distribution worldwide and is a known vector of pathogens such as *Babesia canis*, *Ehrlichia canis*, *Rickettsia conorii*, and *Leishmania* spp [12, 13]. Other tick species of dogs recorded include: *Ixodes ricinus*, *I. hexagonus*, *I. canisuga* from Great Britain [14], *Haemaphysalis longiconis*, *H. falva*, *I. avatus*, *H. hystricis*, *I. persulcatus*, *I. nipponensis*, *A. testudinarium* from Japan [15], *I. dammini* and *Dermacentor variabilis* from the USA [16]. All three stages in the life cycle, the larva, nymph and adults, feed primarily on dogs and each stage drops off after feeding and moulting occur off-host. This tick occasionally infests other hosts, including humans. The life cycle takes up to two years to complete in nature [17, 18]. However, under laboratory conditions, *Rh. sanguineus* completes its entire life cycle within 17 weeks [19].

Few studies have reported the tick species of wild and domesticated animals in Sri Lanka [20, 21, 22, 23]. Liyanaarachchi *et al.*, [24] surveyed the ticks infesting dogs together with other wild and domesticated animals and reported eight tick species: *D. auratus*, *Ha. turturis*, *Ha. cuspidata*, *A. testudinarium*, *A. clypeolatum*, *Ha. spinigera*, *Hy. isaaci*, and *Ha. aculeata* on dogs. They detected spotted fever group rickettsia in *Rh. sanguineus* collected from a dog [24]. However, all the above studies are restricted to one or a few locations in the country. Surveillance data on tick species' diversity and geographic distribution is important because it will provide valuable baseline information to detect future changes in tick prevalence and distribution patterns.

The objectives of the study are to determine the life stages and diversity of the ticks infesting the dogs; the tick distribution infesting dogs in different climatic zones in Sri Lanka and among different districts of Sri Lanka and to describe the life cycle parameters of the dominant dog tick species under laboratory conditions.

Materials And Methods

Study design

First, a descriptive, island-wide survey was conducted to assess the diversity and distribution of dog ticks. Then, the life cycle of the major tick species encountered during the survey was carried out using New Zealand white rabbits (*Oryctolagus cuniculus*) under controlled laboratory conditions.

Geographic distribution and diversity of ticks infesting domestic dogs

Tick infesting the domestic dog (*Canis lupus familiaris*) of different breeds, sex and age were collected from all of the districts covering the Dry, Wet, and Intermediate zones of Sri Lanka [25] using a non-probability convenient sampling technique. At least five towns/ villages were selected from each district, and each site was visited and sampled only once. However, in certain districts where the dogs were fewer in number, less than ten dogs were sampled, where there were more dogs, more than ten were sampled. All the ticks in one-half of the host were added to the collection.

Only the ticks that were attached to the host and feeding were collected. Ticks were located by visual appraisal and by running the hand over the dog's body. Ticks were removed using a pair of tweezers or/and fingers to grasp the tick as close to the skin surface as possible ensuring the mouthparts remained attached to the tick and intact. Specimens were preserved in absolute alcohol. The preferred attachment site was determined by dividing the host body into ten different regions, ear (dorsal, inner sides), eyelids, neck, body, face, tail region, snout, forelimb pads, and hind limb pads. Information about the breed, age, sex of the dogs and the site of attachment of the tick, the intensity of infection were noted. Tick collections were done in both privately-owned dogs and free-roaming community dogs. A thorough examination of free-roaming was not possible as most of them did not have vaccination details. For those dogs who had heavy infestation only up to five ticks per animal was collected. A year-round collection was carried out by two research assistants. Ticks were brought to the laboratory, observed under dissecting microscope, and identified using available standard keys and literature. The number/percentage of different life cycle stages, species diversity according to the district, climatic zone, and diversity index (Shannon diversity index) was calculated. Descriptive statistics (such as percentages and ratios) were calculated and a bivariate analysis chi-squared test was utilized to see the independence of categories among gender etc.

Life cycle of *Rhipicephalus sanguineus* under laboratory conditions

Rhipicephalus sanguineus was the most abundant and widely distributed dog tick species; thus, its life cycle parameters were studied in a colony established by collecting engorged females in the Kandy district from four localities. They were kept under laboratory conditions (Temperature $27\pm 1^\circ\text{C}$; Relative humidity 70%-80%) and were allowed to lay eggs. Life stages were fed on the New Zealand white rabbits (*Oryctolagus cuniculus*). After hatching, 100 larvae were added into a plastic feeding vial glued onto the rabbit's clean-shaved skin. Then, the larvae were allowed to feed, and once they dropped off, they were transferred to sampling vials and allowed to moult. When larvae moulted, the nymphs (20) were introduced to the feeding chamber and allowed to feed on the rabbit. Once the engorged nymphs dropped off, they were also kept to moult under laboratory conditions. Adults (n=15) were also introduced to the feeding chamber and allowed to feed. Fully engorged females and males also were kept in laboratory conditions to determine their survival. Biological parameters such as the weight of the wild-caught females, pre-oviposition period, oviposition period, incubation period of eggs, the parasitic period of larva, nymph, and adult were recorded. Then the Reproductive Efficiency Index (REI; the weight of eggs laid/weight of the engorged females X 100%), percentage of eclosion (percentage of hatched eggs with respect to the total number of eggs laid by each female) and Reproductive Fitness Index (RFI; number of eggs that hatched into larvae/weight of the engorged females in mg) were calculated. Pearson correlation of weight of the female and the reproductive parameters were computed using Minitab 17 version. The study protocol for using lab animals was approved by the Ethical Review Committee at the Postgraduate Institute of Science, University of Peradeniya, Sri Lanka.

Results

Geographic distribution and diversity of ticks infesting domestic dogs

A total of 3,026 ticks were collected from 1,219 dogs of different breeds from the 25 districts in Sri Lanka. The collection consisted of five genera of hard ticks belonging to eight species: *Rhipicephalus sanguineus*, *Rhipicephalus haemaphysaloides*, *Haemaphysalis intermedia*, *Haemaphysalis bispinosa*, *Haemaphysalis turturis*, *Amblyomma integrum*, *Hyalomma* sp. and *Dermacentor auratus* (Family Ixodidae; Figure 1). Adults and nymphs of four species were recorded; *Rh. sanguineus*, *Rh. haemaphysaloides*, *Haemaphysalis intermedia* and *H. bispinosa*. Moreover, nymphs of *D. auratus* and *A. integrum* and only the adults of *Hyalomma* sp. and *H. turturis* were recorded (Table 1). Adults formed 94.7% of the collection with a significantly higher number of females (54.8%) was in the collection than males (40.0%; $\chi^2 = 40.3787$, $p < 0.0001$; Table 1).

Table 1
Species and the life stage (females, males and nymphs) of ticks infesting dogs in the three climatic zones of Sri Lanka (n=3,026)

Tick species	Adults (%)			Nymphs (%)	Difference of males and females		Difference of adults and nymphs	
	Females	Males	Total		χ^2/ F^*	P value	χ^2/ F^*	P value
<i>Rhipicephalus sanguineus</i>	1040 (54.2)	774 (40.4)	1,817 (94.6)	104 (5.4)	39.006	<0.05	1525.73	<0.05
<i>Rhipicephalus haemaphysaloides</i>	299 (44.8)	343 (51.4)	642 (96.3)	25 (3.7)	3.016	0.082	568.90	<0.05
<i>Haemaphysalis intermedia</i>	288 (76.2)	80 (21.2)	368 (97.4)	10 (2.6)	117.56	<0.05	339.058	<0.05
<i>Haemaphysalis bispinosa</i>	10 (58.8)	5 (29.4)	15 (88.2)	2 (11.8)	0.7104*	-	0.057*	-
<i>Haemaphysalis turturis</i>	19 (73.1)	7 (26.9)	26 (100.0)	-	5.538	0.0186	-	-
<i>Amblyomma integrum</i>	-	-	-	13 (100.0)	-	-	-	-
<i>Hyalomma</i> sp.	1 (50.0)	1 (50.0)	2 (100.0)	-	-	-	-	-
<i>Dermacentor auratus</i>	-	-	-	5 (100.0)	-	-	-	-

In Sri Lanka, *Rhipicephalus sanguineus* was the dominant dog tick species (n=1,918, 63.4%; Table 2). Species diversity (presented as Shannon diversity index H) in the three was 1.135, 1.021 and 0.849 in Intermediate, Dry and Wet zones, respectively. It was the most common dog tick in all the districts except in Polonnaruwa, Nuwara-Eliya, Monaragala, Badulla and Mannar. In these exceptions, *Rh. haemaphysaloides* was the dominant species, which was the second most common tick (n=667, 22.0%) infesting dogs in Sri Lanka. The least common species was *Hyalomma* spp. (0.06%) found only in the Vavuniya District (Table 3; Figure 2). In the three climatic zones, *R. sanguineus* was the dominant species encompassing 73.8% in the Wet Zone and 58.6% in the Dry Zone, whereas *Rhipicephalus haemaphysaloides* was the dominant species (42.0%) and *Rh. sanguineus* (34.3%) was the second most dominant species in the Intermediate Zone. *Haemaphysalis bispinosa* was recorded from all three zones, but the distribution was limited to some districts such as Anuradhapura, Polonnaruwa, Matale, Ampara, and Matara. *Haemaphysalis turturis* were only found from Daraniyagala, Trincomalee, and Kilinochchi, while *Hyalomma* sp. was found only from the Vavuniya District (Figure 2). *Amblyomma integrum* was restricted to the Wet and Intermediate zones.

Table 2
Distribution of tick species collected from dogs by the Wet, Dry and Intermediate zones of Sri Lanka (n=3,026)

Tick species	No. of Ticks (Percentage of species by zone)			
	Wet zone	Dry zone	Intermediate zone	Total
<i>Rhipicephalussanguineus</i>	1,036 (73.8)	785 (58.6)	97 (34.3)	1,918 (63.4)
<i>Rhipicephalushaemaphysaloides</i>	192 (13.7)	356 (26.6)	119 (42.0)	667 (22.0)
<i>Haemaphysalisintermedia</i>	12 (0.9)	14 (1.0)	0 (0.0)	26 (0.9)
<i>Haemaphysalisbispinosa</i>	143 (10.2)	173 (12.9)	62 (21.9)	378 (12.5)
<i>Haemaphysalisturturis</i>	8 (0.6)	9 (0.7)	0 (0.0)	17 (0.6)
<i>Amblyommaintegrum</i>	8 (0.6)	0 (0.0)	5 (38.5)	13 (0.4)
<i>Hyalommasp.</i>	0 (0.0)	2 (0.1)	0 (0.0)	2 (0.1)
<i>Dermacentor auratus</i>	5 (0.4)	0 (0.0)	0 (0.0)	5 (0.2)
Total	1,404 (46.4)	1,339 (44.2)	283 (9.4)	3,026
Shannon Diversity Index (H)	0.849	1.021	1.135	0.991

Three agroclimatic zones of Sri Lanka includes several administrative districts. Wet zone - Nuwaraeliya, Ratnapura, Colombo, Kandy, Kegalle, Matara, Galle, Gampaha, Kalutara. Intermediate zone -Kurunegala, Badulla, Matale. Dry zone - Hambantota, Ampara, Vavuniya, Puttlam, Trincomalee, Kilinochchi, Anuradhapura, Batticaloa, Jaffna, Monaragala, Polonnaruwa, Mannar, Mullathivu. Some districts geographically fall partly in two zones

Site of attachment

Most ticks were attached to the anterior part of the body, especially the head region than the posterior side with one exception, *Rh. haemaphysaloides* which preferred the rear side of the host (Table 4). All the nymphs (100%) and most of the adults (59.9%) of *Rh. haemaphysaloides* preferred the posterior side of the host and was found attached to hind limbs mostly while the most common species, *Rh. Sanguineus*, both nymphs (88.0%) and adults (54.0%) preferred head region, specifically the inner and outer ear (Table 4). All other species, exclusively (100%) preferred the anterior of the host body, except in *H. intermedia* which the majority (57.8%) preferred the anterior sites for attachment.

Table 4
Attachment site of the tick species on the host's body

Stage	Tick species	Percentage of ticks										Total anterior (%)	Total posterior (%)
		Eye lids	Inner ear	Outer ear	Snout	Face	Body	Neck	Tail	Fore Limb	Hind limb		
Adult	<i>Rh.sanguineus</i>	7.2	25.5	21.4	0.8	1.5	4.1	5.5	0.4	21.1	12.3	54.9	44.9
	<i>Rh. haemaphysaloides</i>	10.5	14.8	20.9	1.2	2.1	5.9	8.7	0.0	28.1	15.1	47.4	59.9
	<i>H. intermedia</i>	35.5	12.1	10.2	0	1.2	0	0.2	0	28.1	13.9	57.8	42.2
	<i>H. bispinosa</i>	0	85.7	4.3	0	0	0	0	0	0	0	100	0
	<i>H. turturis</i>	0	100	0	0	0	0	0	0	0	0	100	0
	<i>Hyalomma sp.</i>	0	100	0	0	0	0	0	0	0	0	100	0
Nymphs	<i>Rh.sanguineus</i>	0	88.0	0	0	0	0	0	0	8	4	88.0	12.0
	<i>Rh. haemaphysaloides</i>	0	0	0	0	0	0	0	0	61.0	39.1	0	100
	<i>A. integrum</i>	92.1	8.0	0	0	0	0	0	0	0	0	100	0
	<i>D. auretus</i>	0	100	0	0	0	0	0	0	0	0	100	0

Abbreviations: *Rh*, *Rhipicephalus*; *H*, *Haemaphysalis*; *A*, *Amblyomma*; *D*, *Demacentor*

Life cycle parameters of *Rhipicephalus sanguineus*

The engorged, wild-caught females had a mean weight of 133.2 (54 – 187) mg and completed oviposition in 14.0±3.47 days with a pre-oviposition period of 4.9±1.02 days (Table 5 and 6). The mean number of eggs laid by a female was 1444.3±531.08 (101 – 2708) (Table 6). The egg incubation period was 29.6±3.96 days (range 20-43 days; Table 5). Larvae had a mean parasitic period of 3.1±0.56 days, and of the larvae introduced 62.7% successfully fed and dropped off for moulting (Table 5 and 6). Of those engorged larvae, 31.1% moulted into nymphs after a moulting period of 13.4±1.32 days (Tables 5 and 6). The nymphs fed for 5.0±0.0 days, dropped off and spent a moulting period of 14.3±0.48 days. The parasitic period of adult females was 12.6±2.94 days (Table 5).

Table 5
Life cycle parameters of *Rhipicephalus sanguineus* fed on New Zealand white rabbits under controlled laboratory conditions (Temperature 27±1°C; Relative humidity 70%-80%)

Life stage	Parameter	Number of days			
		Mean	SD	Minimum	Maximum
Female[#]	Pre-oviposition	4.9	1.02	4	7
	Oviposition period	14.0	3.47	3	19
	Post-oviposition survival	1.1	0.99	1	5
Eggs	Incubation period	29.6	3.96	20	43
Larva	Parasitic period	3.1	0.56	2	3
	Moulting period	13.4	1.32	13	14
Nymph	Parasitic period	5.0	0.00	5	5
	Moulting period	14.3	0.48	13	15
Female	Parasitic period	12.6	2.94	9	15

[#]Wild caught females; SD =Standard Deviation

Table 6

Biological parameters of *Rhipicephalussanguineus* fed on New Zealand White rabbits under controlled laboratory conditions (Temperature 27±1°C; Relative humidity 70%-80%)

Parameter	Mean	SD	Minimum	Maximum
#Engorged females weight (mg)	109.7	35.09	25.0	200.0
#Number of eggs laid by engorged females	1444.3	531.08	101	2708
#Reproductive Fitness Index (RFI)	9.1	5.01	0.0	22.6
#Reproductive Efficiency Index (REI)	50.8	9.69	22.0	70.0
#Percent eclosion	63.0	22.72	0.0	95.4
Parameter	Percentage			
Feeding success of larvae	62.7			
Moulting success of larvae to nymph	31.7			
Feeding success of nymphs	77.0			
Moulting success of nymph to adult	86.9			
Feeding success of Adult	50.0			
#Wild caught females; SD = Standard Deviation				

Table 7

Correlation (Pearson Correlation) between reproductive parameters and the female weight of wild caught females of *Rhipicephalussanguineus* measured at the laboratory after and during oviposition (Temperature 27±1°C; Relative humidity 70%-80%)

Reproductive parameter	R	p
Total laid eggs per female	0.468	0.001*
Total hatched eggs per female	0.355	0.014*
Weight of egg mass per female	0.896	<0.001**
Reproductive Fitness Index (RFI)	0.181	0.224
Reproductive Efficiency Index (REI)	0.162	0.277
Percent eclosion	0.223	0.132
*indicate the significance p<0.05 ** indicates the significance p < 0.001		

The REI and RFI values were 50.8±9.69 (22.0 – 70.0) and 9.1±5.01 (0 – 22.6) and 8.8 (1.8 – 20), respectively (Table 6). Male: female sex ratio was 5:2 in laboratory-raised adults. The number of eggs laid increased rapidly within the first three days and gradually decreased. There were positive correlations found between the female weight and three reproductive parameters tested; the total number of eggs laid, the number of eggs hatched, and the weight of the egg mass (Pearson correlation: $r = 0.468, 0.355, 0.896$ respectively; $p < 0.05$: Table 6, Figure 3). The egg-laying potential of an engorged female is directly related to her capacity to feed; therefore, heavier females showed the highest values in the number of eggs laid as well as the weight of the egg mass, and after that, a higher hatch rate. However, no significant correlations were found between the three calculated reproductive parameters (REI, RFI and percent eclosion) with the female weight ($p > 0.05$; Table 6 and Figure 4). The regression fit could not fully explain REI, RFI or percent eclosion since the R^2 values were <5.0% (Figure 4).

All the life cycle stages, including larvae, nymphs, and adults successfully fed on the New Zealand white rabbits under controlled laboratory conditions and completed their life cycle within 70 – 126 (mean 98±14.74) days (Table 5).

Discussion

The survey results showed brown dog tick, *Rh. sanguineus* was the major tick species that infest domestic dogs island-wide. More than fifty years ago, Seneviratne in 1965 [20] reported *Rh. sanguineus* as the most frequently found tick on dogs in Ceylon (now Sri Lanka). It is also the most widespread dog tick worldwide [26]. This tick can be found on dogs living in both urban and rural areas, being highly adapted to live within human dwellings and being active throughout the year not only in tropical and subtropical regions but also in some temperate areas

[26]. Although the dog is the main host, it has also been recorded on other animals like wild canids [27] and a wide range of wild and domestic animals of Sri Lanka including ox, horse, buffalo, cat, goat, sheep, and also humans [20, 22, 23, 28]. This indicates that free-ranging wild animals might be involved in its maintenance and dispersion through different regions. This could have implications in tick control and the epidemiology of tick-borne diseases, particularly in areas where dogs live in close contact with wildlife [26]. Studies have shown that some dog breeds appear to be more resistant than others to infestations by *Rh. sanguineus* [29]. The local mongrels crossbred with different breeds or crossbred with the local Sinhala hound or Sinhalese hound belonging to the species *Sinhala sunakaya* which is found in Sri Lanka and parts of India seem to be more resistant to tick bites, but further studies are needed to assess the dog breed susceptibility to ticks. This tick was attached everywhere on the dog but the preferred site was the head region, which is consistent with the description in the review of Dantas-Torres, 2010 [26] and therefore makes it difficult for dogs to remove them, whereas immatures feed on lower areas of the dog's body (e.g., belly, rump, and hind legs), probably because of their more limited mobility [12]. *Rhipicephalus sanguineus* is the most important species from the veterinary standpoint as it is a vector of many pathogens affecting dogs and occasionally humans [26, 30, 31, 32]. It is known to carry many pathogens, including *Babesia vogeli*, *Ehrlichia canis*, *Hepatozoon canis*, *Rickettsia conorii* and *Rickettsia rickettsii*, the last two also been responsible for disease in humans [13]. Liyanaarachchi *et al.* (2015b) [24] detected DNA of spotted fever group rickettsiae in *Rh. sanguineus* collected from a domestic dog in Sri Lanka.

In addition to *Rh. sanguineus*, seven other species were recorded in dogs: *Rhipicephalus haemaphysaloides*, *Haemaphysalis bispinosa*, *Haemaphysalis intermedia*, *Haemaphysalis turturis*, *Amblyomma integrum*, *Dermacentor auratus* and *Hyalomma* sp. Previously, Seneviratne (1965) [20] reported six species of ticks, *Rh. sanguineus*, *Rh. haemaphysaloides*, *H. intermedia*, *H. bispinosa*, *Boophilus* sp, *A. integrum* (immature stages) collected from dogs in Sri Lanka. Later, Liyanaarachchi *et al.* (2015a) [22] recorded 14 species of ticks including, *Rh. sanguineus*, *Rh. haemaphysaloides*, *H. intermedia*, *H. bispinosa*, *Rh. microplus*, *A. integrum*, *H. cuspidata*, *H. spinigera*, *H. aculeata*, *A. testudinarium*, *A. clypeolatum*, *Hy. isaaci* and *D. auratus* from dogs. Out of the previously recorded six species, *Rh. microplus*, *H. cuspidata*, *H. spinigera*, *H. aculeata*, *A. testudinarium* and *A. clypeolatum* were not reported in the present study. Some of these rare species could be accidental occurrences due to close contact of dogs with livestock and wildlife.

Rhipicephalus haemaphysaloides was the second most common tick on dogs recorded from all over the island with a wide geographical distribution but is less common at elevations above 3000 ft. It was the dominant dog tick species in the Intermediate zone especially in Polonnaruwa, Nuwara-Eliya, Monaragala, Badulla and Mannar districts. Although rare in urban areas, it was common in rural areas and areas closer to the forests. It has a limited distribution in the world occurring over the Oriental and parts of the Palaearctic and Australasian zoogeographic regions from Afghanistan in the west to the Philippines and Indonesia in the east and the Indian subcontinent [20, 33, 34]. It is known to infest all domestic animals, including ox, water buffalo, and horse but more commonly seen on sheep and goats [20, 23]. This tick species has a great potential for the transmission of Kaysanur forest disease virus in India [35] and *Rickettsia rhipicephali* in Taiwan [36], and as the principal vector of bovine babesiosis (e.g. *Babesia orientalis*) in China [37].

Three species of *Haemaphysalis* were recorded in the present study: *H. intermedia*, *H. bispinosa* and *H. turturis*, among which *H. intermedia* was widely distributed in all the districts in all three agro-climatic zones but the abundance was low. In 1965, Seneviratne [20] also reported that *H. intermedia* was a widespread tick species in Sri Lanka and collected from all domestic animals throughout the island. These two species, *H. bispinosa*, and *H. intermedia* are the major goat tick in Sri Lanka [23] and it also infests cattle (unpublished observations). Globally, *H. bispinosa* has a limited geographical distribution, being restricted to certain parts of Southern India and Sri Lanka. *Haemaphysalis turturis* was a rare species found only from Kagalle, Kilinochchi and Trincomalee Districts.

Dermacentor auratus and *Hyalomma* sp. were recorded from only one location in the Kandy and Vavuniya Districts, respectively. The primary host of *D. auratus* in wild boar and the tick has an island-wide distribution and a rapid increase in the population has been observed due to restricting the killing of wild boars. Nymphs of *D. auratus* have been identified as the major tick species responsible for human otoacariasis in Sri Lanka [28, 38, 39]. Dogs also carry this tick species, and it is possible when the wild boars visit the human habitats, the engorged females, drop-off and lay eggs and larvae can be easily picked by dogs as well as humans. *Hyalomma* sp was only found from one location in the Dry Zone. According to the Seneviratne in 1965 [20], they collected ticks from buffalo, neat cattle and goats but not from the dogs. This result of the present study may be an accidental occurrence and may be due to their contact with cattle.

Immature stages of *Amblyomma integrum* were recorded from several locations in Rathnapura, Matara, Monaragala and Badulla districts. This species is very widespread in Sri Lanka and is more common in jungle areas [22]. According to Seneviratne 1965 [20], the incidence of *A. integrum* was lower in the higher elevations around Nuwara-Eliya which is similar to the present study and the more thickly populated parts of the Western Province which is not similar to the present study.

All the life stages except larvae of *Rh. sanguineus* and *Rh. haemaphysaloides* were found on dogs. Among adults, more females were present than males. The presence of more females has been reported in Shimada *et al.* 2003 [15] and the presence of more males has been

reported in Dantas-Torres and Otranto 2011 [40] and Diyes & Rajakaruna 2015 [23]. Only nymphs of *D. auratus* and *A. integrum* and only adults of *H. turturis* and *Hyalomma* sp were recorded.

The three-host life cycle of *Rh. sanguineus* was successfully completed within an average of 98 days (70-126 days) and all three stages fed on the New Zealand white rabbits. The duration of the life cycle was shorter compared to the previous studies where it has taken 162-177 days to complete the life cycle [19, 41]. This might be due to the diverse conditions of temperature and relative humidity in which ticks were maintained, as well as the climatic conditions of different geographic regions. However, there might be also some strain-related differences [41]. The weight of the wild-caught females of this study was 133.2 (54-187) mg which is lower than previously recorded values (289.5 mg; [41]). Wild-caught females were removed deliberately from the host while in the study done by Dantas-Torres in 2008 [41] females were allowed to naturally drop off after feeding. The number of eggs laid, during this study was consistent with the previous records, however, with a lower mean number [19, 41]. Similarly, recorded pre-oviposition period, incubation period and parasitic periods of all three life stages were lower than previously recorded durations [19, 41]. The REI and RFI were 50.8 ± 9.69 (22.0-70.0) and 9.1 ± 5.01 (0-22.6) respectively. Dantas-Torres [41], reported an REI of 13 and an RFI of 13.2 for the same species in Italy. The egg-laying potential of an engorged female is directly related to her capacity to feed; therefore heavier females show the highest values in the number of eggs laid as well as the weight of the egg mass and after that a higher hatch rate. This would explain higher REI, Reproductive Affinity Index (RAI) and RFI in Italy population [41, 42].

The distribution and abundance of dog tick species may depend on many factors such as climatic factors such as temperature, humidity, rainfall, presence of other domestic and wild animals and management practices including usage of acaricides and the life cycle can be different from other geographic regions according to the different climatic factors present. Moreover, Dantas-Torres (2010) [26] highlights that in the era of globalization and climate changes; *Rh. sanguineus* is becoming increasingly relevant from a public health perspective. This tick has also been implicated in the transmission of pathogens of zoonotic concern (e.g., *R. rickettsii*) and recent studies have shown that *Rh. sanguineus* ticks exposed to high temperatures are more prone to bite humans [43]. This scenario highlights that the climate change could affect *Rh. sanguineus* populations of around the world and, consequently, the epidemiology of certain tick-borne infections [43].

Conclusion

The dominant dog tick species and the species diversity varied in different climatic regions of Sri Lanka. The brown dog tick, *Rh. sanguineus* was the dominant species in the Dry and Wet zones, while *Rh. haemaphysaloides* was the dominant species in the Intermediate zone. Species diversity varied as follows in the three zones: 1.135, 1.021 and 0.849 in Intermediate, Dry and Wet zones, respectively. New Zealand white rabbit can be used to determine the life cycle parameters of the brown dog tick *Rh. Sanguineus* as all three stages successfully fed and moulted. Information on diversity, distribution and life cycle parameters is fundamental for studies of canine tick-borne infections, zoonoses, and their epidemiology.

Abbreviations

A.	<i>Amblyomma</i>
D.	<i>Dermacentor</i>
DNA	Deoxyribonucleic acid
H.	<i>Haemaphysalis</i>
Ha.	<i>Hyalomma</i>
I.	<i>Ixodes</i>
R.	Rickettsia
RAI	Reproductive Affinity Index
REI	Reproductive Efficiency Index
RFI	and Reproductive Fitness Index
Rh	<i>Rhipicephalus</i>

Declarations

Acknowledgements - Financial assistance from National Research Council (Grant 11-44) is acknowledged.

Funding – Financial assistance from the National Research Council (Grant 11-44)

Availability of Data and Material – All data are available in hard copies and soft copies with the principal investigator stored securely releasable upon any reasonable request

Ethics Approval - All the study protocols and objectives were approved by the Institutional Ethical Clearance Committee of the postgraduate Institute of Peradeniya, Sri Lanka

Consent to Participate – Not Applicable.

Consent for publication – Not Applicable.

Conflicts of Interest – Authors declare that there is no conflict of interest

Author Information

Affiliations - Department of Zoology, University of Peradeniya, Peradeniya, 20400, Sri Lanka, Postgraduate Institute of Science, University of Peradeniya, Peradeniya, 20400, Sri Lanka

Kalpani Omantha Bandaranayaka, Upekha Indunil Dissanayake and Rupika Subashini Rajakaruna

Contributions - SUR designed the study, KOB and UID performed the research, KOB and UID analyzed the data, SUR and KOB wrote the manuscript. all authors read and approved the final manuscript.

Corresponding author - Correspondence to Rupika Subashini Rajakaruna.

References

1. Shaw S, Day M, Birtles R, Breitschwerdt E. Tick-borne diseases of dogs. *Trends Parasitol.* 2001;17:74–80. 10.1016/S1471-4922(00)01856-0.
2. Kirby AD, Smith AA, Benton TG, Hudson PJ. Rising burden of immature sheep ticks (*Ixodes ricinus*) on red grouse (*Lagopus lagopus scoticus*) chicks in the Scottish uplands. *Med. Vet. Entomol.* 2004;18(1):67-70; doi: 10.1111/j.0269-283x.2004.0479.x. PMID: 15009449.
3. Pietzsch ME, Medlock JM, Jones L, Avenell D, Abbott J, Harding P, Leach S. Distribution of *Ixodes ricinus* in the British Isles: investigation of historical records. *Med Vet Entomol.* 2005;19:306–14 (11) (PDF) British ticks; https://www.researchgate.net/publication/289959651_British_ticks [accessed Oct 05 2020].
4. Scharlemann J, Johnson P, Smith AA, Macdonald D, Randolph S. Trends in Ixodid tick abundance and distribution in Great Britain. *Med Vet Entomol.* 2008;22:238–47. 10.1111/j.1365-2915.2008.00734.x.
5. Daniel M, Materna J, Honig V, Metelka L, Danielová V, Harcarik J, Grubhoffer L. Vertical Distribution of the Tick *Ixodes ricinus* and Tick-borne Pathogens in the Northern Moravian Mountains Correlated with Climate Warming (Jeseníky Mts., Czech Republic). *Cent. Eur J Public Health.* 2009;17:139–45. 10.21101/cejph.a3550.
6. Černý V, Rosický B, Ašmera J, Kadlčík K, Kobík V, Kratochvílová E, et al. Results of investigations of phenology of the common tick *Ixodes ricinus* (L.) in the Czech lands in the years 1960 - 1962. *Folia Parasitologia.* 1965;12(2):125–31.
7. Health Protection Scotland. Glasgow. United Kingdom. 2009. <http://www.documents.hps.scot.nhs.uk/giz/10-year-tables/lyme.pdf>. [Accessed March 2020].
8. Matjila TP, Nijhof AM, Taoufik A, Houwers D, Teske E, Penzhorn BL, et al. Autochthonous canine babesiosis in the Netherlands. *Vet Parasitol.* 2005;131:23–9. <https://doi.org/10.1016/j.vetpar.2005.04.020>.
9. Dautel H, Dippel C, Oehme R, Hartelt K, Schettler E. Evidence for an increased geographical distribution of *Dermacentor reticulatus* in Germany and detection of *Rickettsia* sp. RpA4. *Int. J. Med. Microbiol. Suppl.* 2006;296(S1)(2006):149-156.
10. Jameson LJ, Medlock JM. Results of HPA tick surveillance in Great Britain. *Vet Rec Open.* 2009;165:154.
11. Zygner W, Górski P, Wedrychowicz H. Detection of the DNA of *Borrelia afzelii*, *Anaplasma phagocytophilum* and *Babesia canis* in blood samples from dogs in Warsaw. *Vet. Rec.* 2009;164:465-7; 10.1136/vr.164.15.465.
12. Dantas-Torres F. Ticks as vectors of *Leishmania* parasites. *Trends Parasitol.* 2011;27(4):155–9.
13. Dantas-Torres F, Chomel B, Otranto D. Ticks and tick-borne diseases: A One Health perspective. *Trends parasitol.* 2012;28:437–46. 10.1016/j.pt.2012.07.003.

14. Smith FD, Ballantyne R, Morgan ER, Wall R. Prevalence, distribution and risk associated with tick infestation of dogs in Great Britain. *Med Vet Entomol.* 2011. doi:10.1111/j.1365-2915.2011.00954.x.
15. Shimada Y, Beppu T, Inokuma H, Okuda M, Onishi T. Ixodid tick species recovered from domestic dogs in Japan. *Med Vet Entomol.* 2003;17:38–45.
16. Anderson JF, Magnarelli LA. Vertebrate Host Relationships and Distribution of Ixodid Ticks (Acari: Ixodidae) in Connecticut, USA. *J Med Entomol.* 1980;17:314–23. <https://doi.org/10.1093/jmedent/17.4.314>.
17. Sonenshine DE. *Biology of Ticks.* Vol. I: Oxford University Press; 1991.
18. Sonenshine DE. *Biology of Ticks.* Vol. II: Oxford University Press; 1991.
19. Troughton D, Levin M. Life Cycles of Seven Ixodid Tick Species (Acari: Ixodidae) Under Standardized Laboratory Conditions. *J. Med. Entomol.* 2007;44:732-40. 10.1603/0022-2585(2007)44[732:LCOSIT]2.0.CO;2.
20. Seneviratne P. The Ixodidae (Ticks) of Ceylon. Part II and III. *Sri Lanka Vet. J.* 1965;XIII(2):28–45.
21. Liyanaarachchi DR, Jinadasa HRN, Dilrukshi PRMP, Rajapakse RPVJ. Epidemiological Study on ticks in farm animals in selected areas of Sri Lanka. *Trop Agric Res.* 2013;24:336–46.
22. Liyanaarachchi DR, Rajakaruna RS, Dikkumbura AW, Rajapakse RPVJ. Ticks infesting wild and domestic animals and humans of Sri Lanka with new host records. *Acta Trop.* 2015a;142:64–70.
23. Diyes GCP, Rajakaruna RS. Diversity and distribution of tick species infesting goats with two new host records from Sri Lanka. *JNSF.* 2015;43(3):225–34.
24. Liyanaarachchi DR, Rajakaruna RS, Rajapakse RPVJ. Spotted fever group rickettsia in ticks infesting humans, wild and domesticated animals of Sri Lanka: one health approach. *CJS (Biological Sciences).* 2015b;44(2):67–74.
25. Punyawardena BVR. Climate. In: Mapa R, editor. *The Soils of Sri Lanka.* World Soils Book Series. 2020. Springer, Cham. https://doi.org/10.1007/978-3-030-44144-9_2.
26. Dantas-Torres F. Biology and ecology of the brown dog tick, *Rhipicephalus sanguineus*. *Parasit Vectors.* 2010;3:26.
27. Labruna MB, Jorge RS, Sana DA, Jácomo AT, Kashivakura CK, Furtado MM, et al. Ticks (Acari: Ixodida) on wild carnivores in Brazil. *Exp Appl Acarol.* 2005;36:149–63.
28. Ariyaratne S, Apanaskevich DA, Amarasinghe PH, Rajakaruna RS. Occurrence and distribution of tick species associated with human otoacariasis in selected districts of Sri Lanka. *Exp Appl Acarol.* 2016;70(1):99–123. doi:10.1007/s10493-016-0056-z. Epub 2016 Jul 6.
29. Louly CCB, Soares S, Silveira D, Neto O, Silva A, Borges L. Differences in the susceptibility of two breeds of dogs, English cocker spaniel and beagle, to *Rhipicephalus sanguineus* (Acari: Ixodidae). *Int J Acarology.* 2009;35:25–32.
30. Regendanz P, Muniz J. *Rhipicephalus sanguineus* como transmissor da piroplasmose canina no Brasil. *Mem Inst Oswaldo Cruz.* 1936;31:81–4.
31. Ewing SA, Mathew JS, Panciera RJ. Transmission of Hepatozoon americanum (Apicomplexa: Adeleorina) by ixodids (Acari: Ixodidae). *J Med Entomol.* 2002;39:631–4.
32. Forlano M, Scofield A, Elisei C, Fernandes KR, Ewing SA, Massard CL. Diagnosis of Hepatozoon spp. in *Amblyomma ovale* and its experimental transmission in domestic dogs in Brazil. *Vet Parasitol.* 2005;134:1–7.
33. Walker A. The Genus *Rhipicephalus* (Acari, Ixodidae): A Guide to the Brown Ticks of the World; Jane B. Walker, James E. Keirans and Ivan G. Horak *Trop Anim Health Prod.* 2000;32:417–8; <https://doi.org/10.1023/A:1005237804239>.
34. Guglielmone AA, Robbins RG, Apanaskevich D, Petney TN, Estada-Pena A, Horak IG, Shao R, Barker SC. The Argasidae, Ixodidae and Nuttalliellidae (Acari: Ixodida) of the world: a list of valid species names. *Zootaxa.* 2010;2528:1–28.
35. Bhat HR, Naik SV, Ilkal MA, Banerjee K. Transmission of Kyasanur Forest disease virus by *Rhipicephalus haemaphysaloides* ticks. *Acta Virologia.* 1978;22(3):241-4. PMID: 27975.
36. Hsu YM, Lin CC, Chomel BB, Tsai KH, Wu WJ, Huang CG, Chang CC. Identification of *Rickettsia felis* in fleas but not ticks on stray cats and dogs and the evidence of *Rickettsia rhipicephali* only in adult stage of *Rhipicephalus sanguineus* and *Rhipicephalus haemaphysaloides*. *Comp Immunol Microbiol Infect Dis.* 2011;34(6):513–8. doi:10.1016/j.cimid.2011.09.005. Epub 2011 Oct 13. PMID: 22000945.
37. Liu Q, Zhou YQ, Zhou DN, Liu EY, Du K, Chen SG, et al. Semi-nested PCR detection of *Babesia orientalis* in its natural hosts *Rhipicephalus haemaphysaloides* and buffalo. *Vet Parasitol.* 2007;143:260–6. 10.1016/j.vetpar.2006.08.016.
38. Edussuriya BD, Weilgama DJ. Case reports: intra-aural tick infestations in humans in Sri Lanka. *Trans R Soc Trop Med Hyg.* 2003;97:412–3.

39. Bandaranayaka KO, Kularatne SAM, Rajapakse RPVJ, Abeyundara UB, Rajapaksha RMMA, Rajakaruna RS. Human Otoacariasis in Two Climatically Diverse Districts in Sri Lanka: Seasonality, Risk Factors, and Case Notes. *Acta Parasitol.* 2021 Dec;66(4):1326–40. doi:10.1007/s11686-021-00372-w. Epub 2021 May 16. PMID: 33993424.
40. Dantas-Torres F, Otranto D. *Rhipicephalus sanguineus* on dogs: relationships between attachment sites and tick developmental stages. *Exp Appl Acarol.* 2011;52:289–397.
41. Dantas-Torres F. The brown dog tick, *Rhipicephalus sanguineus* (Latreille, 1806) (Acari: Ixodidae): from taxonomy to control. *Vet Parasitol.* 2008;152(3-4):173–85.
42. Gaxiola-Camacho S, García-Vázquez Z, Cruz-Vázquez C, Portillo-Loera J, Vázquez-Peláez C, Quintero-Martínez MT, Rosario-Cruz R. Comparison of efficiency and reproductive aptitude indexes between a reference and field strains of the cattle tick *Rhipicephalus (Boophilus) microplus*, in Sinaloa, Mexico *Rev Bras Parasitol Vet.* 2009;18(4):9–13.
43. Parola P, Socolovschi C, Jeanjean L, Bitam I, Fournier PE, Sotto A, Labauge P, Raoult D. Warmer weather linked to tick attack and emergence of severe rickettsioses. *PLoS Negl Trop Dis.* 2008;2:e338.

Table

Table 3 is only available as a download in the Supplemental Files section.

Figures

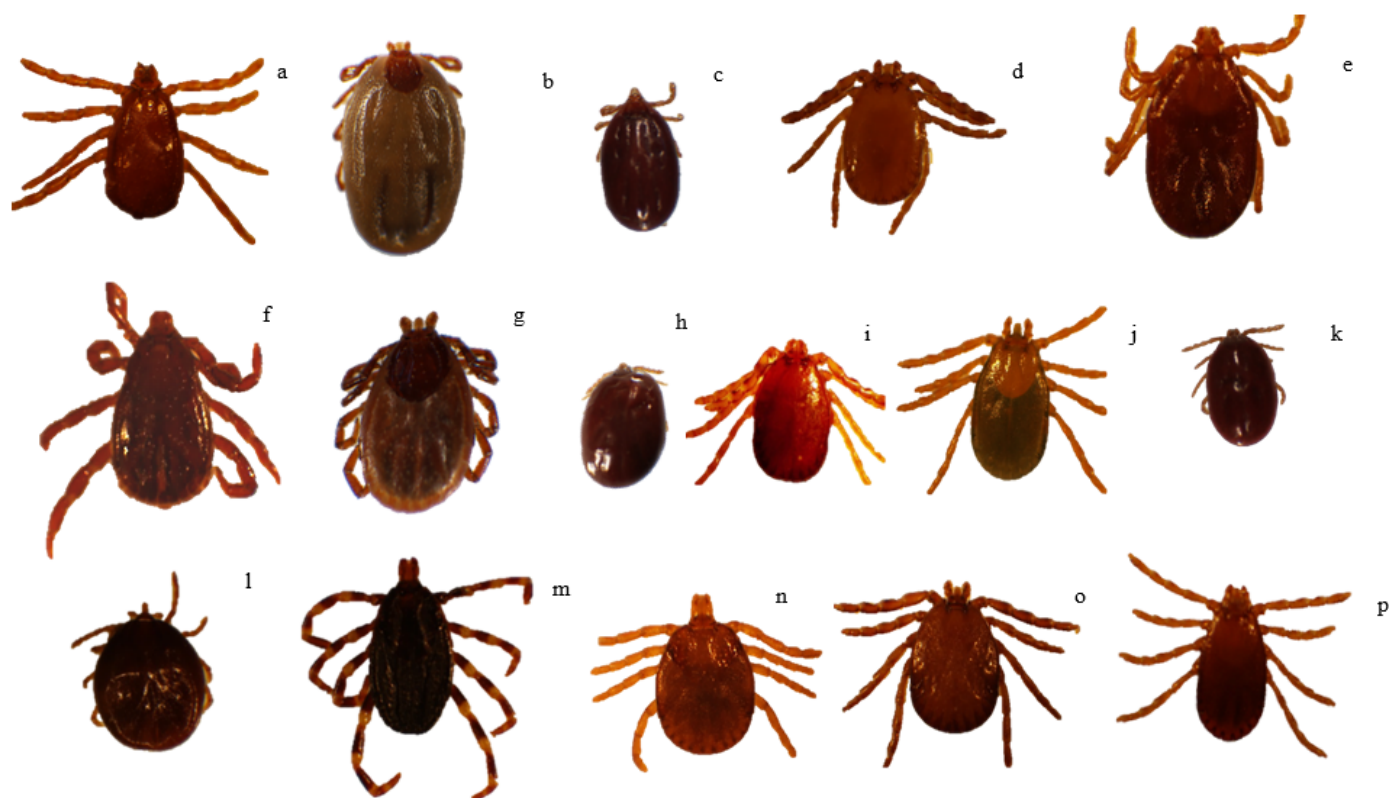


Figure 1

Tick species infesting domestic dog in the three climatic zones of Sri Lanka. a-f) *Rhipicephalus* species a) *Rh. sanguineus* Adult male, b) *Rhipicephalus sanguineus* Adult female, c) *Rhipicephalus sanguineus* Nymph, d) *Rhipicephalus haemaphysaloides* Adult male, e) *Rhipicephalus haemaphysaloides* Adult female, f) *Rhipicephalus haemaphysaloides* Nymph g–m) *Haemaphysalis* species g) *Haemaphysali intermedia* Adult male, h) *Haemaphysali intermedia* Adult female, i) *Haemaphysali intermedia* Nymph, j) *Haemaphysali bispinosa* Adult male, k) *Haemaphysali bispinosa* Adult female, l) *Haemaphysali turturis* Adult male, m) *Haemaphysali turturis* Adult female. n–p) Other species n) *Dermacenter auratus* Nymph, o) *Hyalomma* sp Adult female, p) *Amblyomma integrum* Nymph

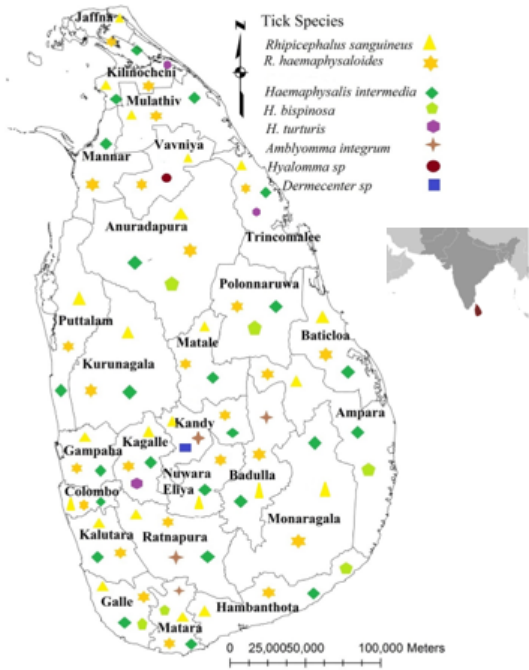


Figure 2

Distribution of dog tick species in each district in Sri Lanka (n=3026)

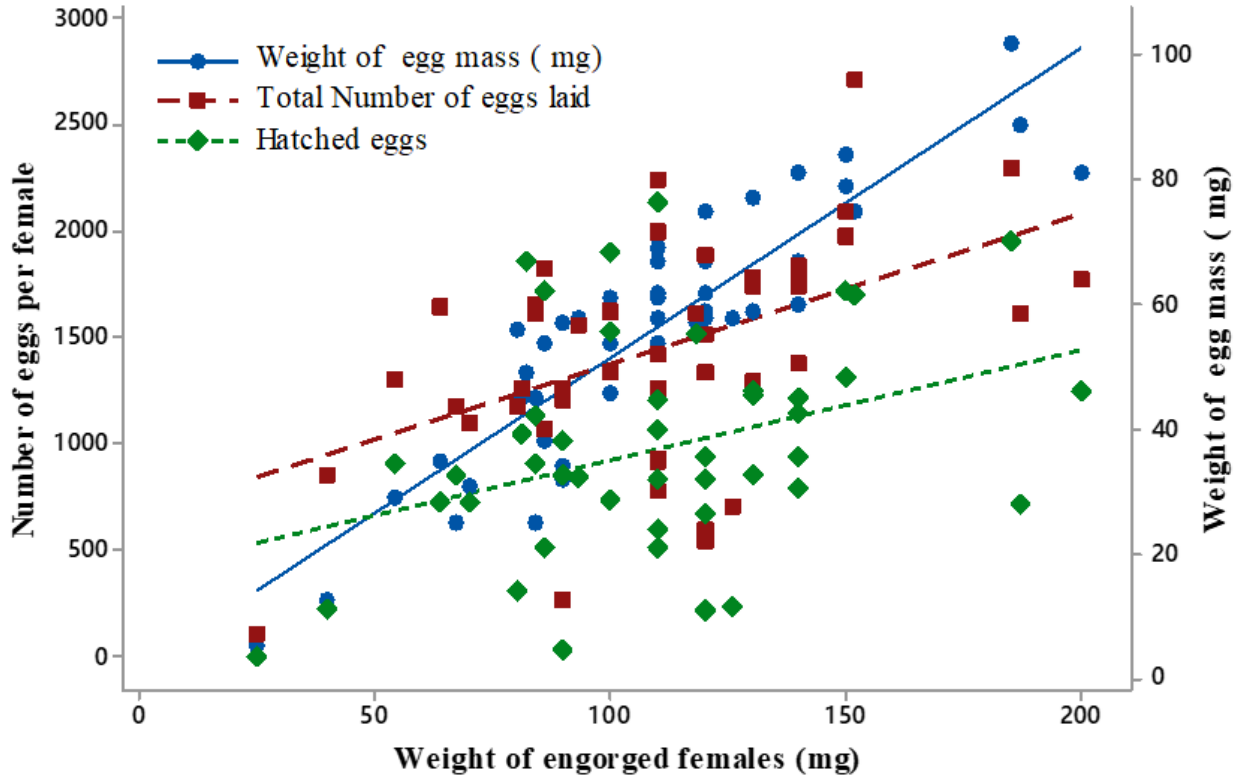


Figure 3

Total number of eggs, hatched eggs and weight of egg clutch per female in wild caught brown dog tick, *Rhipicephalus sanguineus* ($R^2=21.9\%$, 12.6% and 80.4% , respectively)

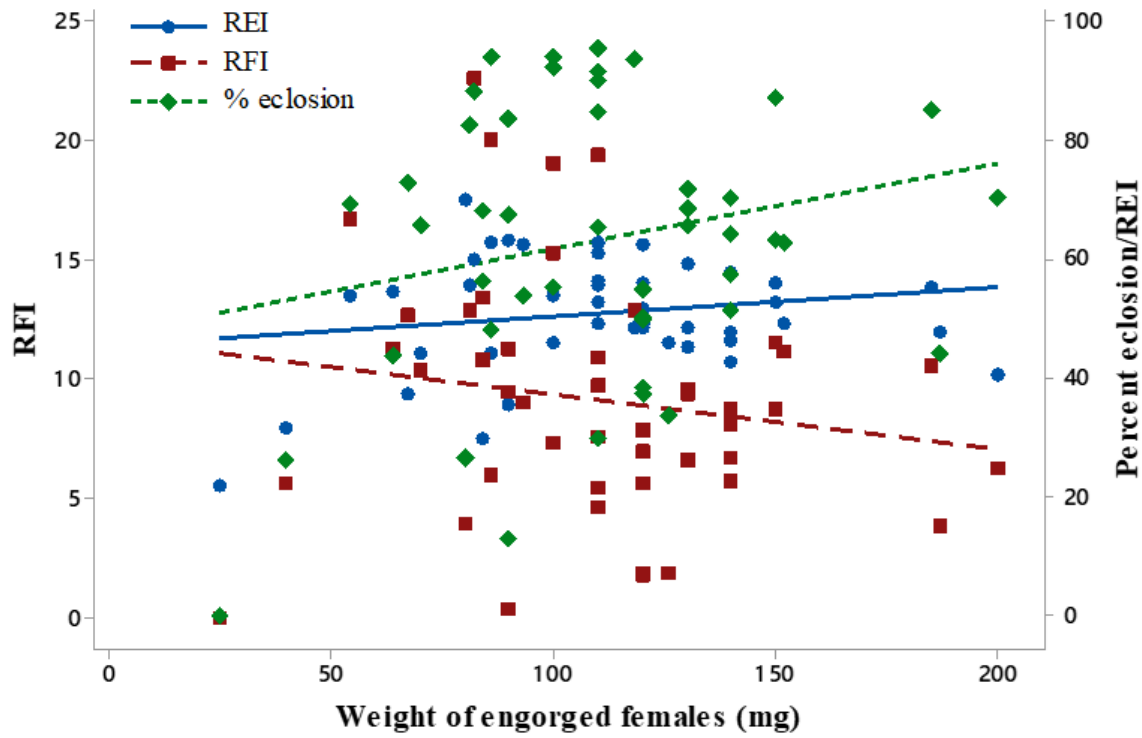


Figure 4

Percent eclosion, Reproductive Efficiency Index (REI) and Reproductive Fitness Index (RFI) per female in wild caught brown dog tick, *Rhipicephalus sanguineus* ($R^2=5.0\%$, 3.3% and 2.6% , respectively)

Supplementary Files

This is a list of supplementary files associated with this preprint. Click to download.

- [Graphicalabstract.pptx](#)
- [Table3.docx](#)