### University of Nebraska - Lincoln DigitalCommons@University of Nebraska - Lincoln

Publications from USDA-ARS / UNL Faculty

U.S. Department of Agriculture: Agricultural Research Service, Lincoln, Nebraska

2015

# Draft Genome Sequence of *Cercospora arachidicola*, Causal Agent of Early Leaf Spot in Peanuts

Valerie A. Orner USDA-ARS National Peanut Research Laboratory, Dawson, Georgia, valerie.orner@ars.usda.gov

Emily G. Cantowine Valdosta State University, egcantonwine@valdosta.edu

Xinye Monica Wang USDA-ARS National Peanut Research Laboratory, Dawson, Georgia

Amr Abouelleil Broad Institute of MIT and Harvard, amr@broadinstitute.org

James Bochicchio Broad Institute of MIT and Harvard, Cambridge, Massachusetts, jboch@broadinstitute.org

See next page for additional authors Follow this and additional works at: https://digitalcommons.unl.edu/usdaarsfacpub

Part of the Agricultural Science Commons, Agronomy and Crop Sciences Commons, Genetics and Genomics Commons, and the Plant Pathology Commons

Orner, Valerie A.; Cantowine, Emily G.; Wang, Xinye Monica; Abouelleil, Amr; Bochicchio, James; Nusbaum, Chad; Culbreath, Albert C.; Abdo, Zaid; and Arias, Renee S., "Draft Genome Sequence of *Cercospora arachidicola*, Causal Agent of Early Leaf Spot in Peanuts" (2015). *Publications from USDA-ARS / UNL Faculty*. 1549.

https://digitalcommons.unl.edu/usdaarsfacpub/1549

This Article is brought to you for free and open access by the U.S. Department of Agriculture: Agricultural Research Service, Lincoln, Nebraska at DigitalCommons@University of Nebraska - Lincoln. It has been accepted for inclusion in Publications from USDA-ARS / UNL Faculty by an authorized administrator of DigitalCommons@University of Nebraska - Lincoln.

### Authors

Valerie A. Orner, Emily G. Cantowine, Xinye Monica Wang, Amr Abouelleil, James Bochicchio, Chad Nusbaum, Albert C. Culbreath, Zaid Abdo, and Renee S. Arias





## Draft Genome Sequence of *Cercospora arachidicola*, Causal Agent of Early Leaf Spot in Peanuts

## Valerie A. Orner,<sup>a</sup> Emily G. Cantonwine,<sup>b</sup> Xinye Monica Wang,<sup>a</sup> Amr Abouelleil,<sup>c</sup> James Bochicchio,<sup>c</sup> Chad Nusbaum,<sup>c</sup> Albert K. Culbreath,<sup>d</sup> Zaid Abdo,<sup>e\*</sup> Renee S. Arias<sup>a</sup>

National Peanut Research Laboratory, Dawson, Georgia, USA<sup>a</sup>; Department of Biology, Valdosta State University, Valdosta, Georgia, USA<sup>b</sup>; Broad Institute of MIT and Harvard, Cambridge, Massachusetts, USA<sup>c</sup>; Department of Plant Pathology, University of Georgia, Tifton, Georgia, USA<sup>d</sup>; USDA, ARS, SEA, Athens, Georgia, USA<sup>e</sup>

\* Present address: Zaid Abdo, Department of Microbiology, Immunology and Pathology, Colorado State University, Fort Collins, Colorado, USA.

*Cercospora arachidicola*, causal agent of early leaf spot, is an economically important peanut pathogen. Lack of genetic information about this fungus prevents understanding the role that potentially diverse genotypes may have in peanut breeding programs. Here, we report for the first time a draft genome sequence of *C. arachidicola*.

Received 23 September 2015 Accepted 28 September 2015 Published 5 November 2015

Citation Orner VA, Cantonwine EG, Wang XM, Abouelleil A, Bochicchio J, Nusbaum C, Culbreath AK, Abdo Z, Arias RS. 2015. Draft genome sequence of *Cercospora arachidicola*, causal agent of early leaf spot in peanuts. Genome Announc 3(6):e01281-15. doi:10.1128/genomeA.01281-15.

Copyright © 2015 Orner et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 3.0 Unported license.

Address correspondence to Renee S. Arias, renee.arias@ars.usda.gov, or Valerie A. Orner, valerie.orner@ars.usda.gov.

This document is a U.S. government work and is not subject to copyright in the United States.

arly leaf spot caused by Cercospora arachidicola S. Hori (teleomorph Mycosphaerella arachidis Deighton) is one of two important leaf spot diseases in peanut (Arachis hypogaea L.) responsible for significant economic loss to the industry (1, 2). Infections by C. arachidicola appear as small necrotic lesions on the leaves, petioles, or stems, which may be followed by premature defoliation, and, if left unmanaged on susceptible cultivars, can severely decrease yield (1). An effective, yet expensive, disease management strategy consists of multiple fungicide applications throughout the growing season (3). Other strategies such as strip-tillage instead of conventional tillage (4) or weather forecast models that predict disease outbreaks (5) can help minimize the number of fungicide treatments. However, the development of leaf-spotresistant cultivars that require no fungicide application would be the most desirable means of control (6). The recent completion of the peanut genome (http://www.peanutbase.org) will aid breeding programs, but the negligible amount of C. arachidicola genetic information hinders progress. Currently, C. arachidicola entries in the NCBI-GenBank database total 8,077 bp in 21 sequences, with half of these entries corresponding to rRNA and the rest only 61 bp each. The genome sequence of C. arachidicola will provide relevant information for the advancement of leaf-spot resistant cultivars, be a useful resource to aid in the selection of target genes for disease control, and contribute to the study of genetic diversity of C. arachidicola.

A single-spore isolate of *C. arachidicola* from an infected peanut plant near Tifton, Georgia, USA, was grown on potato dextrose agar (Difco, Franklin Lakes, NJ, USA) for 6 months. The fungus was removed from the agar and ground using a Kleco tissue pulverizer (Garcia Machine, Visalia, CA, USA). Genomic DNA was extracted using phenol/chloroform/isoamyl alcohol followed by isopropanol precipitation (7) and cleaned using the GeneJET gel Extraction kit (Thermo, Fisher Scientific, Waltham, MA, USA). Sequencing and assembly were performed by the Broad Institute of MIT and Harvard using an Illumina HiSeq 2500 whole-genome shotgun approach. A total of 465,511,514 reads with an estimated genome coverage of  $>100\times$  were assembled *de novo* using ALLPATHS (8) and generated 796 contigs (>400 bp each) with an average size of 40,930 bp that assembled into 491 scaffolds (>1,000 bp each). The average size of the scaffolds was 67,710 bp with a total of 33,245,410 bp and a maximum length of 1,387,526 bp. Most hits from BLAST analysis corresponded to the genera *Pseudocercospora* and *Passalora*.

**Nucleotide sequence accession numbers.** This whole-genome shotgun project has been deposited in DDBJ/ENA/GenBank under the accession number LIHB00000000. The version described in this paper is the first version, LIHB01000000.

#### ACKNOWLEDGMENTS

This project was funded by the USDA-ARS NP301 CRIS Project 6604-21000-004-00D and by the National Peanut Board Project 6044-21000-004-06R.

We thank Imana Power for assistance with this project. Mention of trade names or commercial products in this article is solely for the purpose of providing specific information and does not imply recommendation or endorsement by the U.S. Department of Agriculture.

### REFERENCES

- Nutter FW, Jr, Shokes FM. 1995. Management of foliar diseases caused by fungi, p 65–73. *In* Melouk HA, Shokes FM (ed), Peanut health management. APS Publishing, St. Paul, MN.
- 2. Leidner J. 2012. The peanut genomics initiative. Southeastern Peanut Farmer 50:15.
- Smith DH, Littrell RH. 1980. Management of peanut foliar diseases with fungicides. Plant Dis 64:356–361. http://dx.doi.org/10.1094/PD-64-356.
- Cantonwine EG, Culbreath AK, Stevenson KL. 2007. Characterization of early leaf spot suppression by strip tillage in peanut. Phytopathology 97: 187–194. http://dx.doi.org/10.1094/PHYTO-97-2-0187.
- 5. Olatinwo RO, Prabha TV, Paz JO, Hoogenboom G. 2012. Predicting favorable conditions for early leaf spot of peanut using output from the

weather research and forecasting (WRF) model. Int J Biometeorol 56: 259–268. http://dx.doi.org/10.1007/s00484-011-0425-6.

- Branch WD, Culbreath AK. 2013. Yield performance and pest resistance among peanut genotypes when grown without fungicides or insecticides. Crop Protect 52:22–25. http://dx.doi.org/10.1016/j.cropro.2013.05.005.
- 7. Sambrook J, Fritsch EF, Maniatis T. 1989. Molecular cloning: a laboratory

manual, 2nd ed. Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY.

 Butler J, MacCallum I, Kleber M, Shlyakhter IA, Belmonte MK, Lander ES, Nusbaum C, Jaffe DB. 2008. ALLPATHS: de novo assembly of wholegenome shotgun microreads. Genome Res 18:810–820. http://dx.doi.org/ 10.1101/gr.7337908.