

## Drinkable Preparation of Theracurmin Exhibits High Absorption Efficiency—A Single-Dose, Double-Blind, 4-Way Crossover Study

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**Curcumin has various biological activities including antioxidant and antiinflammatory actions, and alcohol detoxification. However, because of its poor absorption efficiency, it is difficult for orally administered curcumin to reach blood levels sufficient to realize its bioactivities. We have generated capsules and tablets containing Theracurmin, a highly absorptive curcumin. In addition, we recently created a drinkable preparation of Theracurmin. To evaluate the absorption efficiency of this type of curcumin, we performed a single-dose, double-blind, 4-way crossover study. We compared plasma curcumin levels after the administration of Theracurmin beverage and 3 other drinkable types of curcumin sold in Japan. Twenty-four healthy subjects (male/female=13/11, age: 23–32) were administered with these 4 drinkable preparations of curcumin. The area under the blood concentration–time curve at 0–8 h was found to be 1.5 to 4.0-fold higher with Theracurmin than with the other 3 kinds of curcumin beverage. Moreover, maximal plasma curcumin concentrations (0–8 h) of Theracurmin were 1.8 to 3.8 times higher than those of the other 3 curcumin beverages. These data indicate that our newly prepared Theracurmin beverage exhibits a much better absorption efficiency than other kinds of curcumin beverage sold in Japan.**

**Key words** curcumin; drinkable preparation; Theracurmin; human; plasma level

Curcumin is a yellow-colored substance widely known as turmeric, which is prepared from the root of the *Curcuma longa* plant, a member of the ginger family (Zingiberaceae) and native to India and Southeast Asia.<sup>1)</sup> Curcumin is well-known to have a broad spectrum of biological and pharmacological activities,<sup>2)</sup> such as antioxidant,<sup>3–6)</sup> antiinflammatory,<sup>3,6,7)</sup> antibacterial,<sup>8)</sup> antifungal,<sup>9)</sup> and anticarcinogenic activities,<sup>10,11)</sup> as described in many reports. Moreover, curcumin also possesses cardio- and neuroprotective effects.<sup>12–14)</sup> Curcumin has been used to treat a broad range of common ailments in Indian Ayurvedic medicine for at least 4000 years, as well as in Chinese, Arabic, and other traditional medicines. Curcumin is in modern use worldwide as a cooking spice, flavouring agent, and colorant. One of the reasons why curcumin has a broad and long history of use is its safety. No studies in either animals or humans have demonstrated significant toxicity associated with the use of curcumin, even at high doses.<sup>15,16)</sup> However, a problem with the use of curcumin is its poor water solubility and short biological half-life.<sup>17,18)</sup>

Several approaches have been tested to increase the oral bioavailability of curcumin, including adjuvant, nanoparticles, micelles, phospholipid delivery systems, and liposomes.<sup>18–21)</sup> However, no suitable delivery options, such as a soluble formulation of curcumin, have been found so far. Here, we have generated Theracurmin, a highly absorptive form of curcumin, using a technique of a micro-particle and surface-controlled colloidal dispersion. Theracurmin consisted of 10 w/w% of curcumin, 2% of other curcuminoids such as de-

methoxycurcumin and isdemethoxycurcumin, 46% glycerin, 4% gum ghatti, and 38% water. Theracurmin demonstrated oral bioavailability nearly 30-times higher than that of curcumin powder in both rats and humans.<sup>22)</sup> A minimal dose of Theracurmin was sufficient to improve the left ventricular systolic function in post-myocardial infarction rats, suggesting the clinical usefulness of Theracurmin for heart failure treatment.<sup>23)</sup>

Several curcumin beverages are sold as health foods in many countries including Japan. However, it has yet to be determined whether plasma curcumin levels sufficient to demonstrate bioactivities can be obtained by taking these beverages. Without improving the absorption efficiency of these beverages containing curcumin, their bioactivities might not be significant. We created the drinkable preparation of Theracurmin and performed a clinical study in humans using Theracurmin beverage and other curcumin beverages sold in Japan to compare the plasma levels of curcumin.

### MATERIALS AND METHODS

**Drinkable Preparations** Theracurmin was obtained from Theravalues Corporation (Tokyo, Japan).<sup>22)</sup> A drinkable preparation containing Theracurmin was prepared. There other 3 commercial drinkable preparations containing curcumin were obtained from a store. These 4 drinkable preparations were re-bottled in new, unified bottles and labeled A to D, respectively. The contents of these health beverages are indicated in

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Table 1.

**Subjects** The Institutional Review Board at Shizuoka General Hospital approved this study. All subjects provided written informed consent prior to participation. Twenty-four healthy subjects consented to be in the study. Screening procedures included a medical history, physical exam, hematologic profile, and blood chemistries. Subjects were not taking any medications nor were they taking any dietary or herbal supplements. Women could not be pregnant or breast-feeding.

**Study Design and Procedures** Subjects participated in a single-dose, double-blind, 4-way crossover study. Subjects were divided into four groups as shown in Table 2. In each group, the four types of drinkable preparations of curcumin were administered every 7 d. Subjects did not take curcumin containing food for more than 7 d before this study and fasted overnight except for water. In the morning, blood specimens were obtained immediately prior to the drinkable preparations and at 0.5, 1, 2, 4, and 8 h after taking the drinkable preparations. Subjects received a light lunch 6 h after taking the preparations. All blood specimens were drawn in 5-mL blood-collecting vessels containing heparin. They were immediately placed in an ice bath and protected from the light. Vessels were centrifuged at 3000 RPM for 20 min at 4°C to separate the plasma. The plasma samples were then frozen at -70°C.

**Sample Preparation and Measurement of Plasma Curcumin Levels** Blood sample preparation and the measurement of plasma curcumin levels were previously reported.<sup>22</sup> Briefly, each plasma sample was incubated with 0.1 M sodium acetate buffer (pH 5.0) containing 1000 U  $\beta$ -glucuronidase (Wako Pure Chemical Industries, Ltd., Osaka, Japan) at 37°C for 1 h to hydrolyze the curcumin conjugates. After extraction with chloroform, the dried extracts were reconstituted in 100  $\mu$ L of 50% methanol and injected into a chromatographic system. Plasma concentrations of curcumin were measured using the HPLC-MS/MS system comprising the Prominence

micro-LC system (Shimadzu, Kyoto, Japan) and an API 3200 tandem mass spectrometer (Applied Biosystems, CA, U.S.A.) with (+)electrospray ionization (ESI), as described previously.<sup>23</sup>

**Pharmacokinetics** The area under the curve (AUC) was calculated using the trapezoidal method. Maximum concentrations ( $C_{max}$ ) are the observed values.

**Statistical Analysis** Data are expressed as the mean  $\pm$  standard deviation (S.D.). Statistical comparisons were performed using analysis of variance with Scheffe's test. Linear regression analysis with Pearson's coefficients was performed to investigate correlations.

## RESULTS

### Subject Demographic Characteristics and Disposition

Twenty-four subjects were enrolled in this study; the cohort included 13 males (54%), with a mean age of 24 (23–32) years. The mean body weight was 60 (40–92) kg, mean height was 1.69 (1.53–1.84) m, and mean BMI was 20.8 (16.3–28.7) kg/mm<sup>2</sup>. No subjects were withdrawn from the study. No adverse effects are observed.

**Pharmacokinetics** Figure 1 shows representative HPLC chromatograms of plasma after enzymatic hydrolysis. The main pharmacokinetic data of curcumin in healthy volunteers ( $n=24$ ) administered a single oral dose of each curcumin beverage are presented in Table 3, and mean plasma concentrations of curcumin for each drinkable preparation over time are shown in Fig. 2. For all treatments, plasma curcumin levels were quantifiable 30 min after administration. Peak plasma concentrations for all drinkable preparations were not detected during 8 h. At all points, plasma levels of curcumin were higher in A than B, C, and D. This difference was apparent in males. In females, plasma levels of curcumin were significantly higher in A than C and D, and tended to be higher in A than

Table 1. Composition of Each Drink Containing Curcumin

Sample	Curcumin content (display value)	Composition
Drink A	30 mg/100 mL	Water, sugar group (high-fructose corn syrup, sugar), cinnamon extract, ginger, alanine, acidulant, turmeric colorant (Theracurmin), vitamin C, flavor, niacinamide, sweetener (licorice, sucralose), calcium pantothenate, vitamin B <sub>6</sub> , vitamin B <sub>2</sub> , vitamin B <sub>1</sub> , vitamin B <sub>12</sub>
Drink B	30 mg/100 mL	Water, sugar, turmeric extract, Korean ginseng extract, alanine, trehalose, citric acid, arginine, flavor, turmeric colorant, vitamin C, sweetener (sucralose), niacin, vitamin B <sub>2</sub> , vitamin B <sub>6</sub> , vitamin P, phenylalanine, isoleucine, threonine, monosodium glutamate
Drink C	40 mg/120 mL	Water, high-fructose corn syrup, dextrin, <i>Curcuma longa</i> extract, <i>Curcuma zedoaria</i> extract, acidulant, vitamin C, polysaccharide thickener, turmeric colorant, inositol, flavor, cyclic oligosaccharide, sweetener (sucralose, acesulfame potassium, thaumatin), niacin, vitamin B <sub>1</sub> , vitamin E, emulsifier, vitamin B <sub>6</sub>
Drink D	30 mg/100 mL	Water, high-fructose corn syrup, dextrin, <i>Curcuma longa</i> extract, salt, acidulant, vitamin C, polysaccharide thickener, inositol, turmeric colorant, flavor, cyclic oligosaccharide, niacin, sweetener (sucralose, acesulfame potassium, thaumatin), vitamin E, vitamin B <sub>6</sub> , antioxidant (catechin)

Table 2. Assignment of Volunteers in Cross-over Study

	Group 1 ( $n=6$ )	Group 2 ( $n=6$ )	Group 3 ( $n=6$ )	Group 4 ( $n=6$ )
1st week	Drink A	Drink B	Drink C	Drink D
2nd week	Drink B	Drink C	Drink D	Drink A
3rd week	Drink C	Drink D	Drink A	Drink B
4th week	Drink D	Drink A	Drink B	Drink C

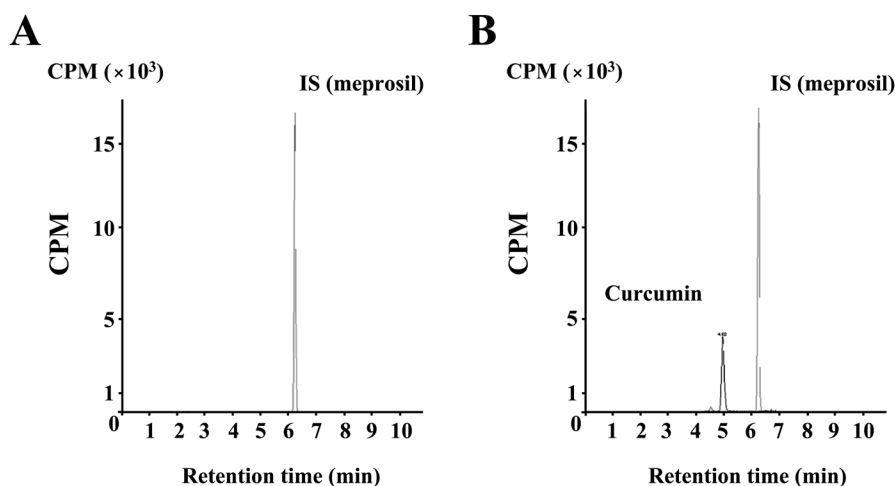


Fig. 1. Representative Examples of HPLC Chromatograms

Curcumin was identified in plasma after enzymatic hydrolysis. 0h (A) and 1h (B) after oral intake of Theracurmin beverage, IS: internal standard.

Table 3. Pharmacokinetic Parameters of Curcumin in the Plasma

	$C_{max}$ (ng/mL)	$AUC_{0-0.5h}$ (ng/mL)	$AUC_{0-1h}$ (ng/mL)	$AUC_{0-2h}$ (ng/mL)	$AUC_{0-4h}$ (ng/mL)	$AUC_{0-8h}$ (ng/mL)
Drink A	25.5±12.2 <sup>*,#,\dagger</sup>	2.0±1.7	6.8±4.9 <sup>*,\dagger</sup>	18.3±11.5 <sup>*,\dagger</sup>	46.0±27.8 <sup>*,#,\dagger</sup>	121.2±65.6 <sup>*,#,\dagger</sup>
Male	28.3±13.8 <sup>*,#,\dagger</sup>	2.0±1.1	6.8±3.9	18.4±10.8 <sup>*,\dagger</sup>	47.3±28.7 <sup>*,\dagger</sup>	124.6±72.2 <sup>*,#,\dagger</sup>
Female	22.3±9.8 <sup>*,\dagger</sup>	2.1±2.3	6.7±6.3	17.3±13.3 <sup>*,\dagger</sup>	43.3±27.8 <sup>*,\dagger</sup>	110.6±61.9 <sup>*,\dagger</sup>
Drink B	14.9±5.4 <sup>*,\dagger</sup>	1.4±0.7	4.9±2.4	14.0±6.3 <sup>*,\dagger</sup>	33.9±14.2 <sup>*,\dagger</sup>	79.5±31.4 <sup>*,\dagger</sup>
Male	14.5±5.3	1.4±0.8	5.1±2.7	13.3±6.8	29.6±12.9	68.9±31.0
Female	15.4±5.8 <sup>\dagger</sup>	1.4±0.7	4.9±2.1	14.8±6.3	36.9±15.7 <sup>\dagger</sup>	83.8±34.2 <sup>\dagger</sup>
Drink C	8.6±4.9	1.2±0.6	3.5±1.5	7.7±3.9	16.8±10.6	40.9±24.9
Male	8.8±5.2	1.2±0.4	3.5±1.5	7.8±4.1	15.3±10.2	38.0±24.5
Female	8.4±4.7	1.2±0.7	3.4±1.7	7.7±3.9	18.7±11.4	44.3±27.3
Drink D	6.7±3.0	1.3±0.5	3.7±1.3	7.5±2.8	13.9±6.6	30.1±14.4
Male	7.2±3.7	1.3±0.5	3.5±1.2	6.9±2.6	12.6±6.6	30.4±17.6
Female	6.2±2.0	1.4±0.6	3.9±1.4	8.2±3.0	15.4±6.6	29.8±11.3

The data are shown as the mean value±standard deviation.  $AUC$ : Area under the curve;  $C_{max}$ : Maximal concentration. \* $p$ <0.05 versus drink B. # $p$ <0.05 versus drink C.  $\dagger p$ <0.05 versus drink D.

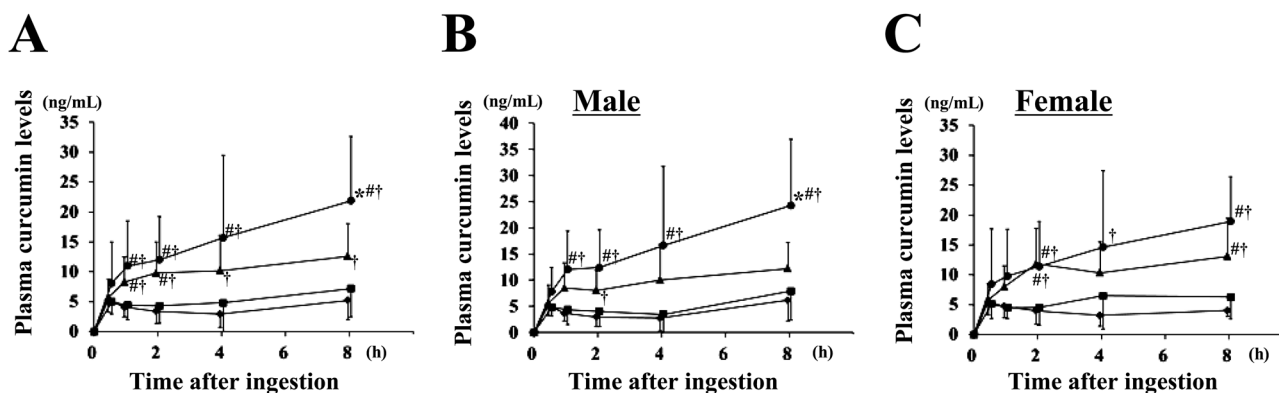


Fig. 2. Change in the Plasma Concentration of Curcumin in Healthy Volunteers

●, drink A. ▲, drink B. ■, drink C. ◆, drink D. Each point and bar represents the mean±S.D. ( $n=24$ ). \* $p$ <0.05 versus drink B. # $p$ <0.05 versus drink C.  $\dagger p$ <0.05 versus drink D.

B. As shown in Table 3, the  $AUC_{0-8h}$  values of A became about 1.5 to 4.0-fold higher than those of the other 3 kinds of curcumin beverage. Plasma  $C_{max}$  (0–8h) of Theracurmin were 1.8 to 3.8 times higher than those of the other 3 curcumin beverages (Table 3). No significant differences were observed between genders. The results of this study indicate that the bioavailability as measured by the  $AUC$  is significantly higher

with Theracurmin beverage than the other 3 curcumin beverages.

To compare the absorption rates, we evaluated  $AUC_{0-2h}$ ,  $AUC_{0-4h}$ , and  $AUC_{0-8h}$ . The  $AUC_{0-2h}$  of A was 1.3-, 2.4-, and 2.4-fold higher than that of B, C, and D, respectively, the  $AUC_{0-4h}$  was 1.4-, 2.7-, and 3.3-fold higher, and the  $AUC_{0-8h}$  was 1.5-, 3.0-, and 4.0-fold higher in A than B, C, and D. The

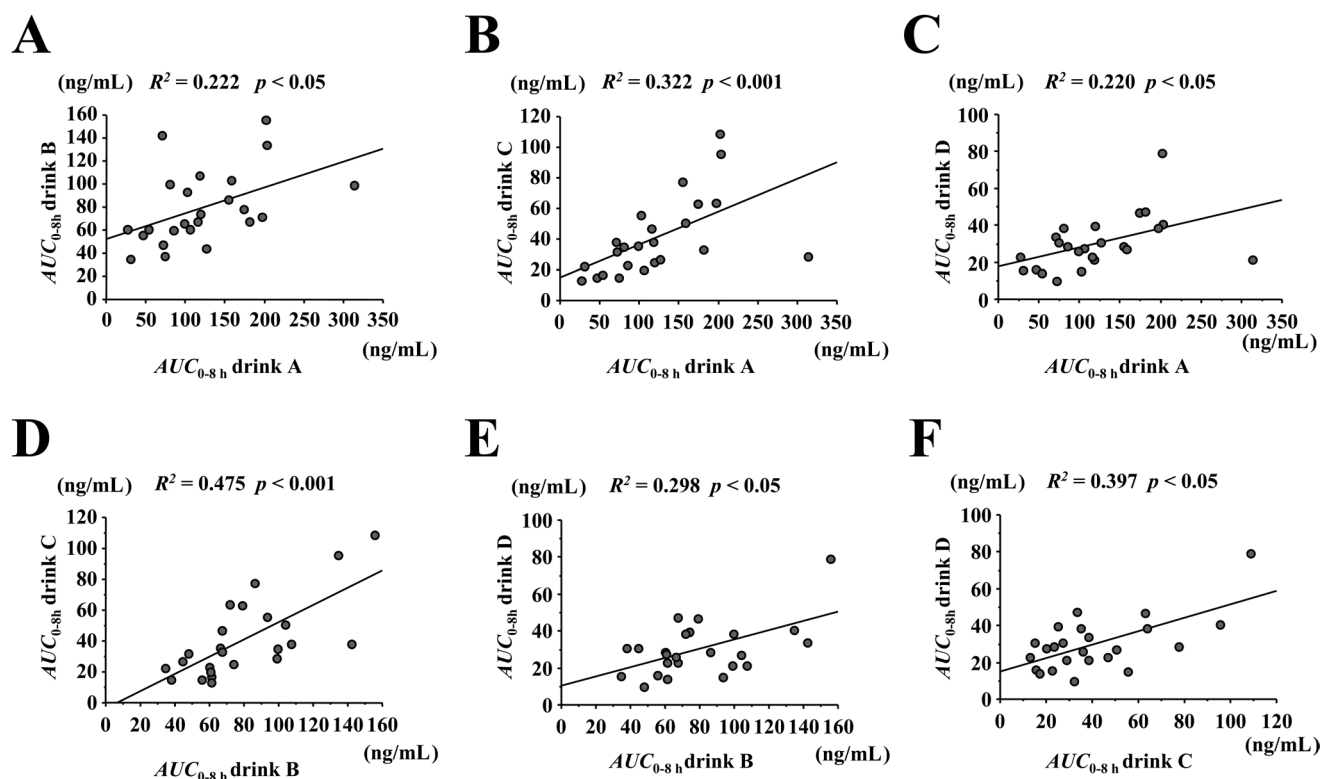


Fig. 3. Correlations of  $AUC_{0-8h}$  Values of Curcumin among A, B, C, and D

differences became larger as the duration was longer.

The differences of  $AUC_{0-8h}$  among volunteers were marked. To determine whether low or high absorption efficiency in each individual is shared by all 4 types of curcumin beverage, we examined  $AUC_{0-8h}$  correlations among these beverages in each individual. As shown in Fig. 3, significant good correlations were observed between any 2 types of curcumin beverage. These findings suggest that individual differences in the pharmacokinetics of curcumin are shared by all 4 types of curcumin beverage.

## DISCUSSION

We have newly created a drinkable preparation of Theracurmin, a highly absorptive curcumin. The present double-blind, 4-way crossover study in healthy volunteers demonstrated that the Theracurmin beverage yielded higher  $C_{max}$  and  $AUC$  than the other curcumin beverages sold in Japan. This indicates that Theracurmin beverage possesses a higher absorption efficiency compared with other curcumin beverages.

The problem of curcumin after oral administration is its poor systemic bioavailability, due to its low solubility in water as many other natural polyphenols, and its rapid metabolism. Recently, drug delivery systems accompanied by nanoparticle technology have emerged as prominent solutions to the bioavailability as therapeutic agents. Although nanoparticle-based delivery systems might be suitable for highly hydrophobic agents like curcumin to circumvent the pitfalls of poor aqueous solubility, very few studies have been reported regarding curcumin nanoparticles. A polymer-based nanoparticle of curcumin, "nanocurcumin," with a particle size of less than 100 nm size was synthesized. While nanocurcumin works well as curcumin *in vitro*, no efficacy of nanaocurcumin over free

curcumin *in vivo* has been reported.<sup>24)</sup> Curcuminoid-loaded solid lipid nanoparticles were developed for long-term stability at room temperature and reduced light and oxygen sensitivity.<sup>25)</sup> To increase the bioavailability of curcumin, nanoparticles encapsulating curcumin have been prepared by the emulsion technique.<sup>26)</sup> These include an optimized poly(lactic-co-glycolic acid) nano-formulation,<sup>27)</sup> dextran sulfate-chitosan nanoparticles with curcumin,<sup>28)</sup> polymeric nanoparticle-encapsulated curcumin,<sup>29,30)</sup> and water-dispersible hybrid nanogels.<sup>31)</sup> However, these nanoparticle-based systems for curcumin delivery are still in their infancy, and much progress is warranted in this area. Here, we have generated highly absorptive curcumin, Theracurmin, using a micro-particle and surface-controlled colloidal dispersion method, which markedly improves oral bioavailability. The  $AUC$  after the oral administration of Theracurmin was more than 40- and 27-fold higher than that of curcumin powder in rats and humans, respectively.<sup>22)</sup> Thus, Theracurmin may be useful to exert clinical benefits in humans at lower dosages.

We reported that the plasma curcumin level reaches a maximum at 1 h after the oral administration of 30 mg of Theracurmin in healthy volunteers.<sup>22)</sup> Many reports indicate that oral administration of curcumin in humans and rodents results in peak plasma levels at around 1 h after the intake.<sup>32-35)</sup> However, in this study, drinkable types of curcumin including the Theracurmin beverage yielded maximum plasma curcumin levels more than 8 h after drinking. In all experiments of pharmacokinetics, subjects were in completely fasting states before and after taking samples. Thus, the influence of meals could be discounted. A number of clinical studies demonstrated that concomitant food and drug intake reduces  $C_{max}$  but increases  $AUC$ . These findings suggest that food intake increases the overall bioavailability but reduces the peak systemic exposure

relative to fasting conditions.<sup>36,37)</sup> Therefore, one of the reasons for the delayed peak is that drinkable types of curcumin contain many components other than curcumin, which may affect its absorption speed.<sup>21)</sup> In normal life, it is expected that curcumin beverages are taken before and after meals, and their absorption speeds should be slower than those indicated by pharmacokinetic experiments. Another reason for this delay in the curcumin peak is enterohepatic circulation. When curcumin was given orally to rats, most of it was excreted in the feces and negligible amounts were found in the urine.<sup>38)</sup> Intravenous and intraperitoneal administrations of curcumin resulted in biliary excretion in rats.<sup>39)</sup> These data indicate that curcumin is re-absorbed from feces and that the plasma concentration of curcumin is maintained at high levels for a long time. This may be useful to exert and sustain the physiological effects of the drinkable types of curcumin, especially Theracurmin beverage.

In this study, we compared plasma levels of curcumin in Theracurmin beverage and those of other curcumin beverages sold in Japan. We reported that the plasma level of curcumin is significantly higher after the oral administration of Theracurmin than that of curcumin powder in humans.<sup>22)</sup> Since the compositions differ between Theracurmin beverage and other curcumin beverages (Table 1), such differences may affect curcumin levels. However, as the precise composition, origin, extraction method, and curcumin modification of each beverage are unclear, the substitution of curcumin beverages with Theracurmin was impossible. Further pharmacokinetic studies are needed to clarify possible effects of the beverage components on blood curcumin levels.

Curcumin is the active ingredient of the dietary spice turmeric and has been consumed for medicinal purposes for thousands of years.<sup>40,41)</sup> Modern science has shown that curcumin modulates various signaling molecules, including inflammatory molecules, transcription factors, enzymes, protein kinases, and protein reductases. Moreover, curcumin has been reported to possess bioactivity, such as anti-inflammatory, anti-oxidant, pro-apoptotic, chemopreventive, chemotherapeutic, anti-proliferative, wound healing, anti-nociceptive, anti-parasitic, and anti-malarial properties. Animal studies have suggested that curcumin may be active against a wide range of human diseases, including diabetes, obesity, neurologic and psychiatric disorders, and cancer, as well as chronic illnesses affecting the eyes, lungs, liver, kidneys, and gastrointestinal and cardiovascular systems. Curcumin has been shown to have the potential to improve many diseases in human research as well as experimental studies including cultured cells and animal models. In addition, more than 90 clinical trials using supplemental curcumin are on-going in many countries. Furthermore, curcumin is used as a supplement in several countries, including India, Japan, the United States, Thailand, China, Korea, Turkey, South Africa, Nepal, and Pakistan. However, this inexpensive, apparently well-tolerated, and potentially active curcumin has not yet been approved for the treatment of any human disease. Many clinical trials evaluating curcumin's safety and efficacy against human ailments have already been completed.<sup>42)</sup> In the near future, curcumin may be used not only in health foods, but also medical agents.

We have shown that curcumin exhibits biological activity to prevent the deterioration of systolic functions in rat heart failure models at  $C_{\max}$  with  $10.7 \pm 1.7$  and  $5.0 \pm 2.4$  ng/mL.<sup>23)</sup> In this

study,  $C_{\max}$  of Theracurmin beverage was  $25.5 \pm 12.2$  ng/mL, which is higher than those with protective effects on the heart. We previously examined whether Theracurmin exerts effects on alcohol metabolism after drinking ethanol in healthy volunteers. We demonstrated that Theracurmin could reduce plasma levels of acetaldehyde, a product of ethanol.<sup>22)</sup> These data indicate that Theracurmin directly affects the metabolism of acetaldehyde and accelerates the detoxification of ethanol. The  $AUC_{0-6h}$  value which reduces the plasma concentration of acetaldehyde after ethanol consumption in human is  $113 \pm 61$  ng/mL·h and almost comparable to  $AUC_{0-8h}$  ( $121.2 \pm 65.6$  ng/mL·h) after taking Theracurmin beverage in this study.<sup>22)</sup> Therefore, Theracurmin beverage may be useful to obtain plasma curcumin levels sufficient to exert beneficial effects such as alcohol detoxification. Moreover, many studies indicate that curcumin shows powerful hepatoprotective effects against oxidative damage caused by several hepatotoxins including ethanol.<sup>43)</sup> In those studies, curcumin, not only attenuated lipid peroxidation but also recovered the activity of endogenous antioxidative defense system.<sup>44-49)</sup> Therefore, Theracurmin might provide protection against alcoholic liver damage.

These findings demonstrate that Theracurmin beverage shows the highest bioavailability among currently available preparations of curcumin. Thus, it may be useful to exert its physiological benefits in humans at lower dosages.

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**Conflict of Interest** Contributing authors Y. Otsuka, T. Hamada and A. Imaizumi are full-time employees of Theravalues Corporation. Y. Nonaka, T. Fuwa and T. Teramoto are full-time employees of Suntory Beverage and Food Limited. The other authors declare that no competing interests exist.

## REFERENCES

- 1) Epstein J, Sanderson IR, Macdonald TT. Curcumin as a therapeutic agent: the evidence from *in vitro*, animal and human studies. *Br. J. Nutr.*, **103**, 1545–1557 (2010).
- 2) Shimatsu A, Kakeya H, Imaizumi A, Morimoto T, Kanai M, Maeda S. Clinical application of “curcumin,” a multi-functional substance. *Anti-Aging Med.*, **9**, 75–83 (2012).
- 3) Esatbeyoglu T, Huebbe P, Ernst IM, Chin D, Wagner AE, Rimbach G. Curcumin—From molecule to biological function. *Angew. Chem. Int. Ed. Engl.*, **51**, 5308–5332 (2012).
- 4) Selvam R, Subramanian L, Gayathri R, Angayarkanni N. The anti-oxidant activity of turmeric (*Curcuma longa*). *J. Ethnopharmacol.*, **47**, 59–67 (1995).
- 5) Ruby AJ, Kuttan G, Babu KD, Rajasekharan KN, Kuttan R. Antitumour and antioxidant activity of natural curcuminoids. *Cancer Lett.*, **94**, 79–83 (1995).
- 6) Menon VP, Sudheer AR. Antioxidant and anti-inflammatory properties of curcumin. *Adv. Exp. Med. Biol.*, **595**, 105–125 (2007).
- 7) Rao TS, Basu N, Siddiqui HH. Anti-inflammatory activity of curcumin analogues. *Indian J. Med. Res.*, **75**, 574–578 (1982).
- 8) Negi PS, Jayaprakasha GK, Jagan Mohan Rao L, Sakariah KK.

- Antibacterial activity of turmeric oil: a byproduct from curcumin manufacture. *J. Agric. Food Chem.*, **47**, 4297–4300 (1999).
- 9) Wuthi-udomlert M, Grisanapan W, Luanratana O, Caichompoo W. Antifungal activity of *Curcuma longa* grown in Thailand. *Southeast Asian J. Trop. Med. Public Health*, **31** (Suppl. 1), 178–182 (2000).
  - 10) López-Lázaro M. Anticancer and carcinogenic properties of curcumin: considerations for its clinical development as a cancer chemopreventive and chemotherapeutic agent. *Mol. Nutr. Food Res.*, **52** (Suppl 1), S103–S127 (2008).
  - 11) Das L, Vinayak M. Anti-carcinogenic action of curcumin by activation of antioxidant defence system and inhibition of NF-kappaB signalling in lymphoma-bearing mice. *Biosci. Rep.*, **32**, 161–170 (2012).
  - 12) Morimoto T, Sunagawa Y, Kawamura T, Takaya T, Wada H, Nagasawa A, Komeda M, Fujita M, Shimatsu A, Kita T, Hasegawa K. The dietary compound curcumin inhibits p300 histone acetyltransferase activity and prevents heart failure in rats. *J. Clin. Invest.*, **118**, 868–878 (2008).
  - 13) Sunagawa Y, Morimoto T, Wada H, Takaya T, Katanasaka Y, Kawamura T, Yanagi S, Marui A, Sakata R, Shimatsu A, Kimura T, Takeya H, Fujita M, Hasegawa K. A natural p300-specific histone acetyltransferase inhibitor, curcumin, in addition to angiotensin converting enzyme inhibitor exerts beneficial effects on left ventricular systolic function after myocardial infarction in rats. *Circ. J.*, **75**, 2151–2159 (2011).
  - 14) Thiyyagarajan M, Sharma SS. Neuroprotective effect of curcumin in middle cerebral artery occlusion induced focal cerebral ischemia in rats. *Life Sci.*, **74**, 969–985 (2004).
  - 15) Shankar TN, Shantha NV, Ramesh HP, Murthy IA, Murthy VS. Toxicity studies on turmeric (*Curcuma longa*): acute toxicity studies in rats, guineapigs and monkeys. *Indian J. Exp. Biol.*, **18**, 73–75 (1980).
  - 16) Vareed SK, Kakarala M, Ruffin MT, Crowell JA, Normolle DP, Djuric Z, Brenner DE. Pharmacokinetics of curcumin conjugate metabolites in healthy human subjects. *Cancer Epidemiol. Biomarkers Prev.*, **17**, 1411–1417 (2008).
  - 17) Sharma RA, Steward WP, Gescher AJ. Pharmacokinetics and pharmacodynamics of curcumin. *Adv. Exp. Med. Biol.*, **595**, 453–470 (2007).
  - 18) Anand P, Kunnumakkara AB, Newman RA, Aggarwal BB. Bioavailability of curcumin: problems and promises. *Mol. Pharm.*, **4**, 807–818 (2007).
  - 19) Fang M, Jin Y, Bao W, Gao H, Xu M, Wang D, Wang X, Yao P, Liu L. *In vitro* characterization and *in vivo* evaluation of nanostructured lipid curcumin carriers for intragastric administration. *Int. J. Nanomedicine*, **7**, 5395–5404 (2012).
  - 20) Grynkiewicz G, Ślifierki P. Curcumin and curcuminoids in quest for medicinal status. *Acta Biochim. Pol.*, **59**, 201–212 (2012).
  - 21) Shoba G, Joy D, Joseph T, Majeed M, Rajendran R, Srinivas PS. Influence of piperine on the pharmacokinetics of curcumin in animals and human volunteers. *Planta Med.*, **64**, 353–356 (1998).
  - 22) Sasaki H, Sunagawa Y, Takahashi K, Imaizumi A, Fukuda H, Hashimoto T, Wada H, Katanasaka Y, Takeya H, Fujita M, Hasegawa K, Morimoto T. Innovative preparation of curcumin for improved oral bioavailability. *Biol. Pharm. Bull.*, **34**, 660–665 (2011).
  - 23) Sunagawa Y, Wada H, Suzuki H, Sasaki H, Imaizumi A, Fukuda H, Hashimoto T, Katanasaka Y, Shimatsu A, Kimura T, Takeya H, Fujita M, Hasegawa K, Morimoto T. A novel drug delivery system of oral curcumin markedly improves efficacy of treatment for heart failure after myocardial infarction in rat. *Biol. Pharm. Bull.*, **35**, 139–144 (2012).
  - 24) Bisht S, Feldmann G, Soni S, Ravi R, Karikar C, Maitra A, Maitra A. Polymeric nanoparticle-encapsulated curcumin (“nanocurcumin”): a novel strategy for human cancer therapy. *J. Nanobiotechnology*, **5**, 3 (2007).
  - 25) Tiyaboonchai W, Tungpradit W, Plianbangchang P. Formulation and characterization of curcuminoids loaded solid lipid nanoparticles. *Int. J. Pharm.*, **337**, 299–306 (2007).
  - 26) Shaikh J, Ankola DD, Beniwal V, Singh D, Kumar MN. Nanoparticle encapsulation improves oral bioavailability of curcumin by at least 9-fold when compared to curcumin administered with piperine as absorption enhancer. *Eur. J. Pharm. Sci.*, **37**, 223–230 (2009).
  - 27) Tsai YM, Jan WC, Chien CF, Lee WC, Lin LC, Tsai TH. Optimized nano-formulation on the bioavailability of hydrophobic polyphenol, curcumin, in freely-moving rats. *Food Chem.*, **127**, 918–925 (2011).
  - 28) Anitha A, Deepagan VG, Divya Rani VV, Menon D, Nair SV, Jayakumar R. Preparation, characterization, *in vitro* drug release and biological studies of curcumin loaded dextran sulphate-chitosan nanoparticles. *Carbohydr. Polym.*, **84**, 1158–1164 (2011).
  - 29) Bhawana, Basniwal RK, Buttar HS, Jain VK, Jain N. Curcumin nanoparticles: preparation, characterization, and antimicrobial study. *J. Agric. Food Chem.*, **59**, 2056–2061 (2011).
  - 30) Yallapu MM, Jaggi M, Chauhan SC. Curcumin nanoformulations: a future nanomedicine for cancer. *Drug Discov. Today*, **17**, 71–80 (2012).
  - 31) Wu W, Shen J, Banerjee P, Zhou S. Water-dispersible multifunctional hybrid nanogels for combined curcumin and photothermal therapy. *Biomaterials*, **32**, 598–609 (2011).
  - 32) Shoba G, Joy D, Joseph T, Majeed M, Rajendran R, Srinivas PS. Influence of piperine on the pharmacokinetics of curcumin in animals and human volunteers. *Planta Med.*, **64**, 353–356 (1998).
  - 33) Pan MH, Huang TM, Lin JK. Biotransformation of curcumin through reduction and glucuronidation in mice. *Drug Metab. Dispos.*, **27**, 486–494 (1999).
  - 34) Cheng AL, Hsu CH, Lin JK, Hsu MM, Ho YF, Shen TS, Ko JY, Lin JT, Lin BR, Ming-Shiang W, Yu HS, Jee SH, Chen GS, Chen TM, Chen CA, Lai MK, Pu YS, Pan MH, Wang YJ, Tsai CC, Hsieh CY. Phase I clinical trial of curcumin, a chemopreventive agent, in patients with high-risk or pre-malignant lesions. *Anticancer Res.*, **21** (4B), 2895–2900 (2001).
  - 35) Sharma RA, Euden SA, Platton SL, Cooke DN, Shafayat A, Hewitt HR, Marczylo TH, Morgan B, Hemingway D, Plummer SM, Pirmohamed M, Gescher AJ, Steward WP. Phase I clinical trial of oral curcumin: biomarkers of systemic activity and compliance. *Clin. Cancer Res.*, **10**, 6847–6854 (2004).
  - 36) Bass A, Stark JG, Pixton GC, Sommerville KW, Zamora CA, Leibowitz M, Rolleri R. Dose proportionality and the effects of food on bioavailability of an immediate-release oxycodone hydrochloride tablet designed to discourage tampering and its relative bioavailability compared with a marketed oxycodone tablet under fed conditions: a single-dose, randomized, open-label, 5-way crossover study in healthy volunteers. *Clin. Ther.*, **34**, 1601–1612 (2012).
  - 37) Benziger DP, Kaiko RF, Miotto JB, Fitzmartin RD, Reder RF, Chasin M. Differential effects of food on the bioavailability of controlled-release oxycodone tablets and immediate-release oxycodone solution. *J. Pharm. Sci.*, **85**, 407–410 (1996).
  - 38) Wahlström B, Blennow G. A study on the fate of curcumin in the rat. *Acta Pharmacol. Toxicol. (Copenh.)*, **43**, 86–92 (1978).
  - 39) Holder GM, Plummer JL, Ryan AJ. The metabolism and excretion of curcumin (1,7-bis-(4-hydroxy-3-methoxyphenyl)-1,6-heptadiene-3,5-dione) in the rat. *Xenobiotica*, **8**, 761–768 (1978).
  - 40) Graham A. Curcumin adds spice to the debate: lipid metabolism in liver disease. *Br. J. Pharmacol.*, **157**, 1352–1353 (2009).
  - 41) Gupta SC, Patchva S, Koh W, Aggarwal BB. Discovery of curcumin, a component of golden spice, and its miraculous biological activities. *Clin. Exp. Pharmacol. Physiol.*, **39**, 283–299 (2012).
  - 42) Gupta SC, Sung B, Kim JH, Prasad S, Li S, Aggarwal BB. Multi-targeting by turmeric, the golden spice: From kitchen to clinic. *Mol. Nutr. Food Res.*, **57**, 1510–1528 (2013).
  - 43) Goel A, Kunnumakkara AB, Aggarwal BB. Curcumin as “Curcumin”: From kitchen to clinic. *Biochem. Pharmacol.*, **75**, 787–809 (2008).
  - 44) Rong S, Zhao Y, Bao W, Xiao X, Wang D, Nussler AK, Yan H, Yao

- P, Liu L. Curcumin prevents chronic alcohol-induced liver disease involving decreasing ROS generation and enhancing antioxidative capacity. *Phytomedicine*, **19**, 545–550 (2012).
- 45) Eybl V, Kotyzova D, Koutensky J. Comparative study of natural antioxidants—curcumin, resveratrol and melatonin—in cadmium-induced oxidative damage in mice. *Toxicology*, **225**, 150–156 (2006).
- 46) Fu Y, Zheng S, Lin J, Ryerse J, Chen A. Curcumin protects the rat liver from CCl<sub>4</sub>-caused injury and fibrogenesis by attenuating oxidative stress and suppressing inflammation. *Mol. Pharmacol.*, **73**, 399–409 (2008).
- 47) Kamalakkannan N, Rukkumani R, Varma PS, Viswanathan P, Rajasekharan KN, Menon VP. Comparative effects of curcumin and an analogue of curcumin in carbon tetrachloride-induced hepatotoxicity in rats. *Basic Clin. Pharmacol. Toxicol.*, **97**, 15–21 (2005).
- 48) Reyes-Gordillo K, Segovia J, Shibayama M, Vergara P, Moreno MG, Muriel P. Curcumin protects against acute liver damage in the rat by inhibiting NFkappaB, proinflammatory cytokines production and oxidative stress. *Biochim. Biophys. Acta*, **1770**, 989–996 (2007).
- 49) Rukkumani R, Aruna K, Varma PS, Rajasekaran KN, Menon VP. Comparative effects of curcumin and an analog of curcumin on alcohol and PUFA induced oxidative stress. *J. Pharm. Pharm. Sci.*, **7**, 274–283 (2004).