

University of Dundee

E3 Ligase Ligands for PROTACs

Ishida, Tasuku ; Ciulli, Alessio

Published in:
SLAS Discovery

DOI:
[10.1177/2472555220965528](https://doi.org/10.1177/2472555220965528)

Publication date:
2021

Licence:
CC BY

Document Version
Publisher's PDF, also known as Version of record

[Link to publication in Discovery Research Portal](#)

Citation for published version (APA):

Ishida, T., & Ciulli, A. (2021). E3 Ligase Ligands for PROTACs: How They Were Found and How to Discover New Ones . *SLAS Discovery*, 26(4), 484-502. <https://doi.org/10.1177/2472555220965528>

General rights

Copyright and moral rights for the publications made accessible in Discovery Research Portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

- Users may download and print one copy of any publication from Discovery Research Portal for the purpose of private study or research.
- You may not further distribute the material or use it for any profit-making activity or commercial gain.
- You may freely distribute the URL identifying the publication in the public portal.

Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

E3 Ligase Ligands for PROTACs: How They Were Found and How to Discover New Ones

SLAS Discovery

1–19

© The Author(s) 2020



DOI: 10.1177/2472555220965528

journals.sagepub.com/home/jbx

Tasuku Ishida¹ and Alessio Ciulli¹

Abstract

Bifunctional degrader molecules, also called proteolysis-targeting chimeras (PROTACs), are a new modality of chemical tools and potential therapeutics to understand and treat human disease. A required PROTAC component is a ligand binding to an E3 ubiquitin ligase, which is then joined to another ligand binding to a protein to be degraded via the ubiquitin–proteasome system. The advent of nonpeptidic small-molecule E3 ligase ligands, notably for von Hippel–Lindau (VHL) and cereblon (CRBN), revolutionized the field and ushered in the design of drug-like PROTACs with potent and selective degradation activity. A first wave of PROTAC drugs are now undergoing clinical development in cancer, and the field is seeking to extend the repertoire of chemistries that allow hijacking new E3 ligases to improve the scope of targeted protein degradation.

Here, we briefly review how traditional E3 ligase ligands were discovered, and then outline approaches and ligands that have been recently used to discover new E3 ligases for PROTACs. We will then take an outlook at current and future strategies undertaken that invoke either target-based screening or phenotypic-based approaches, including the use of DNA-encoded libraries (DELs), display technologies and cyclic peptides, smaller molecular glue degraders, and covalent warhead ligands. These approaches are ripe for expanding the chemical space of PROTACs and usher in the advent of other emerging bifunctional modalities of proximity-based pharmacology.

Keywords

PROTACs, E3 ubiquitin ligase, targeted protein degradation, binding ligands, molecular glues

Introduction

Selective modulation of targeted proteins with small molecules is a major strategy to treat disease. In the early 2000s, almost all pharmaceutical companies invested heavily in efforts to develop small-molecule protein modulators, mainly inhibitors, with desirable properties in terms of efficacy and safety. Although a lot of new molecular entities (NMEs) were launched as a result, numerous proteins remain poorly tractable and so challenging to tackle by small molecules. New modalities, such as antibody and oligonucleotides, opened a door to address some of those more challenging targets, but face other limitations such as poor cell permeability and/or chemical instability. Alternative small-molecule modalities are therefore required to expand the range of proteins being targeted for drug discovery.

Inducing degradation of target proteins by bifunctional small molecules, so-called proteolysis-targeting chimeras (PROTACs), is one of the most exciting such new modalities. PROTACs consist of a ligand for recruiting a target protein of interest (POI) and a ligand for an E3 ubiquitin

ligase, joined with an appropriate linker. PROTACs induce proximity between an E3 ligase and POI in the form of a ternary complex that results in POI ubiquitination and subsequent degradation by the proteasome (**Fig. 1**). Compared with classical inhibition by small molecules, PROTACs offer several potential advantages: (1) PROTACs are expected to exert similar phenotypes to those observed via knockdowns using genetic tools, such as small interfering RNA (siRNA), short hairpin RNA (shRNA), or clustered regularly interspaced short palindromic repeats (CRISPR),

¹Division of Biological Chemistry and Drug Discovery, School of Life Sciences, University of Dundee, Dundee, UK

Received Aug 12, 2020, and in revised form Sep 14, 2020. Accepted for publication Sep 21, 2020.

Corresponding Author:

Alessio Ciulli, Division of Biological Chemistry and Drug Discovery, School of Life Sciences, University of Dundee, Dow Street, Dundee, DDI 5EH, UK.

Email: a.ciulli@dundee.ac.uk

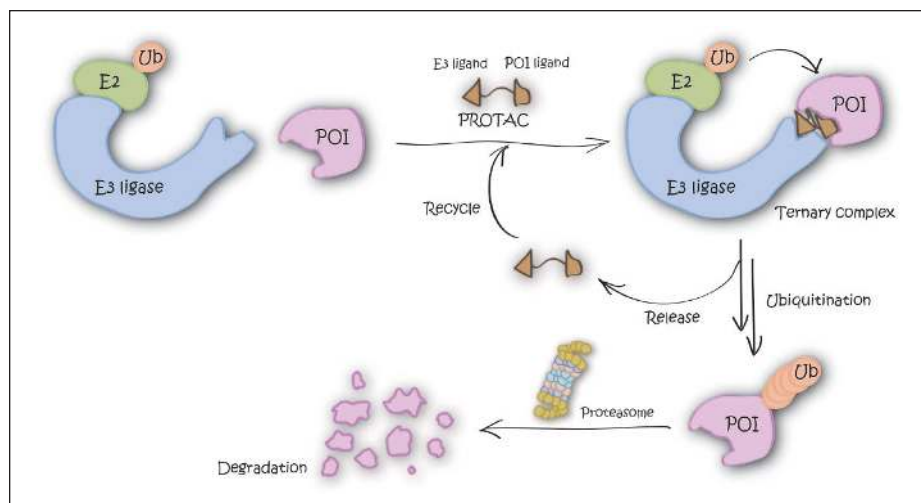


Figure 1. Graphical representation of the degradation mechanism of proteolysis-targeting chimeras (PROTACs).

because the downstream result is the same in all those cases (i.e., depletion of intracellular protein levels). Elimination of POI could give additional effect by disrupting formation of biologically functional complexes. (2) PROTACs can work catalytically (i.e., can be recycled so that one PROTAC molecule can turn over multiple molecules of POI) and so can act “sub-stoichiometrically” (i.e., at fractional occupancy of the POI). As a result of this, PROTACs often show higher POI degradation than expected based on their binding affinity to the POI alone. (3) Target protein degradation by PROTACs can suppress resistant mutation and/or upregulation of POI.

In 2001, Sakamoto and coworkers reported a first PROTAC to degrade methionine aminopeptidase (MetAP),¹ and in 2004, Schneekloth et al. described peptidic von Hippel–Lindau tumor suppressor (pVHL) protein-based PROTACs, which showed cellular permeability and degradation activity against FK506 binding protein 12 (FKBP12) and androgen receptor (AR).² These first-generation PROTACs consisted of a peptidic ligand for an E3 ligase. Peptide moieties caused limited cell permeability, synthetic tractability, and biological instability, which motivated efforts to develop more drug-like, nonpeptidic E3 ligase ligands. During the past few years, these efforts have resulted in improved small-molecule-based PROTACs recruiting cereblon (CRBN),³ von Hippel–Lindau (VHL),⁴ and inhibitors of apoptosis proteins (IAPs).⁵ By leveraging these small-molecule E3 ligase ligands, the field has since extensively demonstrated that PROTACs can induce degradation of a variety of proteins, even at sub-nanomolar concentrations, and has validated their applications not only as biological tools but also as a new chemical modality for treatment of diseases in the clinic. Moreover, modern medicinal chemistry efforts have enabled the development of PROTACs with acceptable drug-like properties. In 2020,

Arvinas presented interim results of a Phase 1/2 clinical trial of their front-line PROTAC, ARV-110, in men with metastatic castration-resistant prostate cancer (mCRPC) and showed two patients achieving responses in prostate-specific antigen (PSA) levels.^{6,7} This landmark result showed pharmacological efficacy of PROTACs in the clinic. The number of E3 ligases currently being explored by the most advanced PROTAC molecules remains small (typically, VHL, CRBN, and IAPs), however, limiting scope. Expansion of the E3 ligase toolbox will therefore be important not only to facilitate degrading a broader range of proteins but also to potentially induce less systemic and more selective (e.g., tissue- or organ-specific) degradation of target proteins by exploiting differential biology and expression levels of E3 ligases.

Here, we first briefly recount how ligands of the typical E3 ligases for PROTACs were discovered, and then review approaches that have been recently used to discover new ligands and new E3 ligases for PROTACs. We then outline the diverse technologies that, in our opinion, provide the most compelling and suited strategies to find new E3 ligase ligands for PROTACs, including applications of structure-based ligand design, DNA-encoded libraries (DELs), display technologies for macrocyclic peptides, molecular glues, and covalent warhead ligands.

Discovery of the Traditional E3 Ligase Ligands for PROTACs

VHL Ligands and Applications to PROTACs. In 1992, the hypoxia-inducible factor-1 α (HIF1 α) was found in Hep3B cells as a factor inducing erythropoietin under hypoxic conditions.⁸ After this finding, it was discovered that HIF1 α also regulates a variety of biological responses (e.g., metabolic reprogramming,⁹ suppression of reactive oxygen species

[ROS] production,¹⁰ angiogenesis,¹¹ epithelial-mesenchymal transition,¹² and cancer metastasis¹³). Under normoxic conditions, intracellular concentrations of HIF1 α are kept at low levels by hydroxylation of specific proline residues and subsequent ubiquitination and proteasomal degradation. The protein responsible for this process was later found to be the pVHL protein, a tumor suppressor protein mutated in VHL disease.^{14,15} pVHL is a subunit of the E3 ubiquitin ligase CUL2–RBX1–ElonginB–ElonginC–VHL (CRL2^{VHL}) complex.^{16–18} The cocrystal structure of the pVHL complex and C-terminal oxygen-dependent degradation (CODD) motif of HIF1 α revealed the structural recognition of a key hydroxyproline residue in hydroxylated HIF1 α by pVHL.^{19,20}

Early PROTACs consisted of epitope peptide CODD motif (ALAPYIP) conjugated to a poly-D-arginine tag at the C-terminus, and to a target binding ligand at the N-terminus. These peptidic PROTACs induced degradation of intracellular FKBP12 and AR without need of microinjection, albeit at fairly high concentrations.² The moiety of the peptidic pVHL ligand, however, limited further application as mentioned before; therefore, development of non-peptidic pVHL ligands was required to overcome this issue.

In 2012, novel peptidomimetic pVHL ligands were developed by the laboratories of Crews and Ciulli, by leveraging the hydroxyproline core fragment that forms critical binding interactions with Ser111 and His115 in the hydroxyproline binding pocket on pVHL. Computational approaches to design plausible ligands and subsequent cocrystal structures guided rational medicinal chemistry optimization that yielded a first small-molecule pVHL–HIF1 α protein–protein interaction inhibitor with dissociation constants of single-digit micromolars.^{21–23} Galdeano et al. later developed second-generation ligands via structure-based drug design using cocrystal structures and isothermal titration calorimetry (ITC) to optimize, in a stepwise rational fashion, interactions at the left-hand side of the VHL ligand. As a result of their optimization study, strong VHL binders with nanomolar binding affinity to VHL were finally obtained, disclosing VHL ligand VH032 (**Fig. 2A**).²⁴ Subsequent medicinal chemistry work by the Ciulli group optimizing the “capping group” at the left-hand side of VH032 found that this position is tolerant for and can accommodate a variety of substituents. They identified fluoro- and cyano-cyclopropyl capping groups, which led to novel compounds (VH101 and VH298, respectively; **Fig. 2A**) with sub-100 nM affinity (more potent affinity than a 10-mer HIF1 α peptide).²⁵ Biophysical and cellular studies qualified VHL inhibitor VH298 as a potent, selective, and cell-active chemical probe of the hypoxic signaling pathway.²⁶

The left-hand side of the VHL ligands provided an early suitable exit vector to develop PROTACs. Zengerle et al. leveraged the crystal structure of VH032 bound to VHL to design and develop one of the first VHL-based PROTACs, MZ1 (**Fig. 2A**), which degraded Bromodomain- and

Extraterminal domain (BET) proteins BRD2, BRD3, and BRD4 in cancer cells, with unexpectedly selective degradation for BRD4 at single- to double-digit nanomolar levels.⁴ At the same time, Bondeson et al. independently reported PROTACs bearing the same VHL ligand that degraded estrogen-related receptor- α (ERR α) and Receptor-interacting serine/threonine-protein kinase-2 (RIPK2) efficiently both in cells and in vivo (PROTAC_RIPK2 and PROTAC_ERR α ; **Fig. 2A**).²⁷ These two studies disclosed the first high-quality all-small-molecule VHL-based PROTACs, changing the game in the field and opening future avenues for VHL-based and other E3-recruiting PROTACs.

Ligands for CRBN and Application to PROTACs. In 1957, thalidomide (**Fig. 2B**) was launched by Chemie Grünenthal as a drug for insomnia. Shortly following administration and use as a sedative, however, it was found that thalidomide had strong teratogenicity, causing approximately 8000 to 12,000 children to be born with deformities.²⁸ After those tragic adverse events, thalidomide was finally withdrawn from the market. Fifty years later, thalidomide was found to have therapeutic effect in patients with multiple myeloma,²⁹ but the mechanism of action remained unclear. In 2010, Ito et al. found that thalidomide binds to CRBN, a subunit of the E3 ubiquitin ligase CUL4–RBX1–DDB1–CRBN (CRL4^{CRBN}), and CRBN engagement induced at least part of the teratogenic effect in in vivo models.³⁰ Two independent groups then separately reported that lenalidomide, a derivative of thalidomide and a marketed drug used for the treatment of hematological malignancies, induced degradation of transcription factor Ikaros family zinc finger protein-1 (IKZF1) and Ikaros family zinc finger protein-3 (IKZF3).^{31,32} In 2014, two X-ray cocrystal structures of DDB1–CRBN–thalidomide complexes were disclosed by the laboratories of Thomä and Celgene. The structures revealed that a portion of the phthalimide ring on thalidomide is exposed to solvent and provided room for extending a substituent or linker.^{33,34} In a manner similar to that pursued with VHL-based PROTACs, Winter et al. and Lu et al. leveraged these findings and crystal structures to develop PROTACs dBET1, ARV825, and dFKBP (**Figure 2B**), which potently and rapidly degraded BET proteins or FKBP12 and demonstrated tumor growth inhibitory activity in cells and in mouse models.^{3,35}

CRBN and VHL: Key Players in the PROTAC Field. Currently, the VHL and CRBN ligands are the most popular and most frequently used E3 ligands to design PROTAC degraders. These ligands are the frontrunners in this field because of several favorable features: (1) strong, specific, and biophysically validated binding affinities to their targeted E3 ligase; (2) acceptable physicochemical profile, for example molecular weight, lipophilicity, solubility, lack of obvious reactive groups or metabolic hotspots, and lack of pan-assay

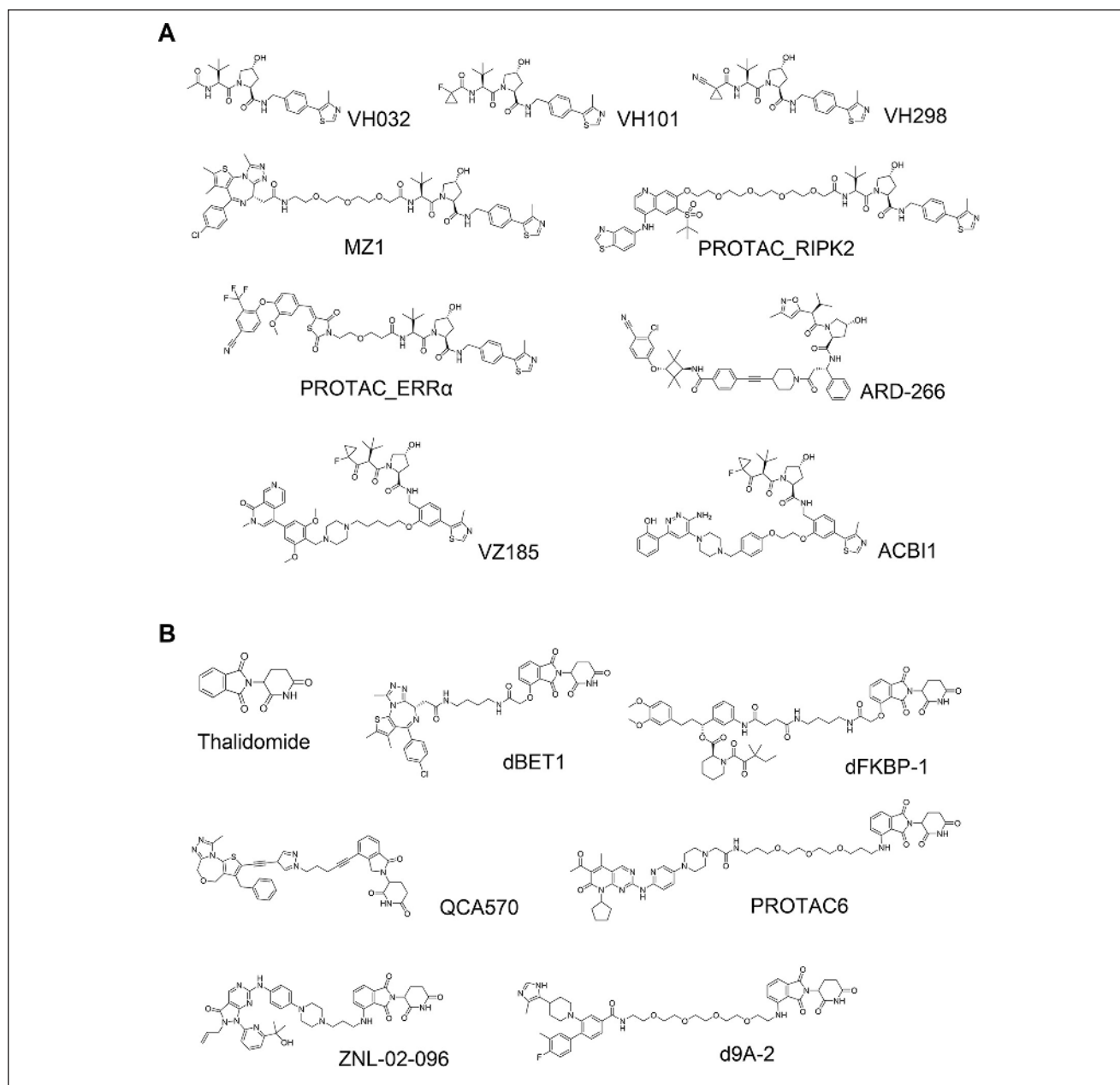


Figure 2. Von Hippel–Lindau (VHL)-based and cereblon (CRBN)-based small-molecule proteolysis-targeting chimeras (PROTACs). **(A)** Exemplified structures of VHL ligands and the PROTACs composed of them: VH032,^{24,26} VH101,²⁵ VH298,^{25,26} MZ1,⁴ PROTAC_RIPK2,²⁷ PROTAC_ERR α ,²⁷ ARD-266,¹³⁷ VZ185,⁷⁷ and ACBI1.¹³⁸ **(B)** Exemplified structures of thalidomide and PROTACs composed of CRBN ligands: thalidomide,²⁸ dBET1,³ dFKBP12,³ QCA570,¹³⁹ PROTAC6,¹⁴⁰ ZNL-02-096,¹⁴¹ and d9A-2.¹⁴²

interference compounds (PAINS)³⁶ alerts; and (3) well-characterized structural information of their binding modes. Because of these properties, numerous PROTACs that are based on CRBN or VHL have been reported that have shown excellent *in vitro* and *in vivo* degradation activities for the targeted POI (**Fig. 2**). As a result, PROTACs composed of CRBN or VHL ligand are currently considered one of the most powerful alternative modalities of chemical

intervention into biology and promising therapeutics. Therefore, not only Big Pharma but also institutes and biotech firms have filed many patents and are engaged in developing PROTACs all the way into the clinic (**Fig. 3**).

PROTACs Based on IAP Ligands. The family of IAPs is characterized by three baculovirus IAP repeat (BIR) domains at their N-termini, and has important functions in inhibiting

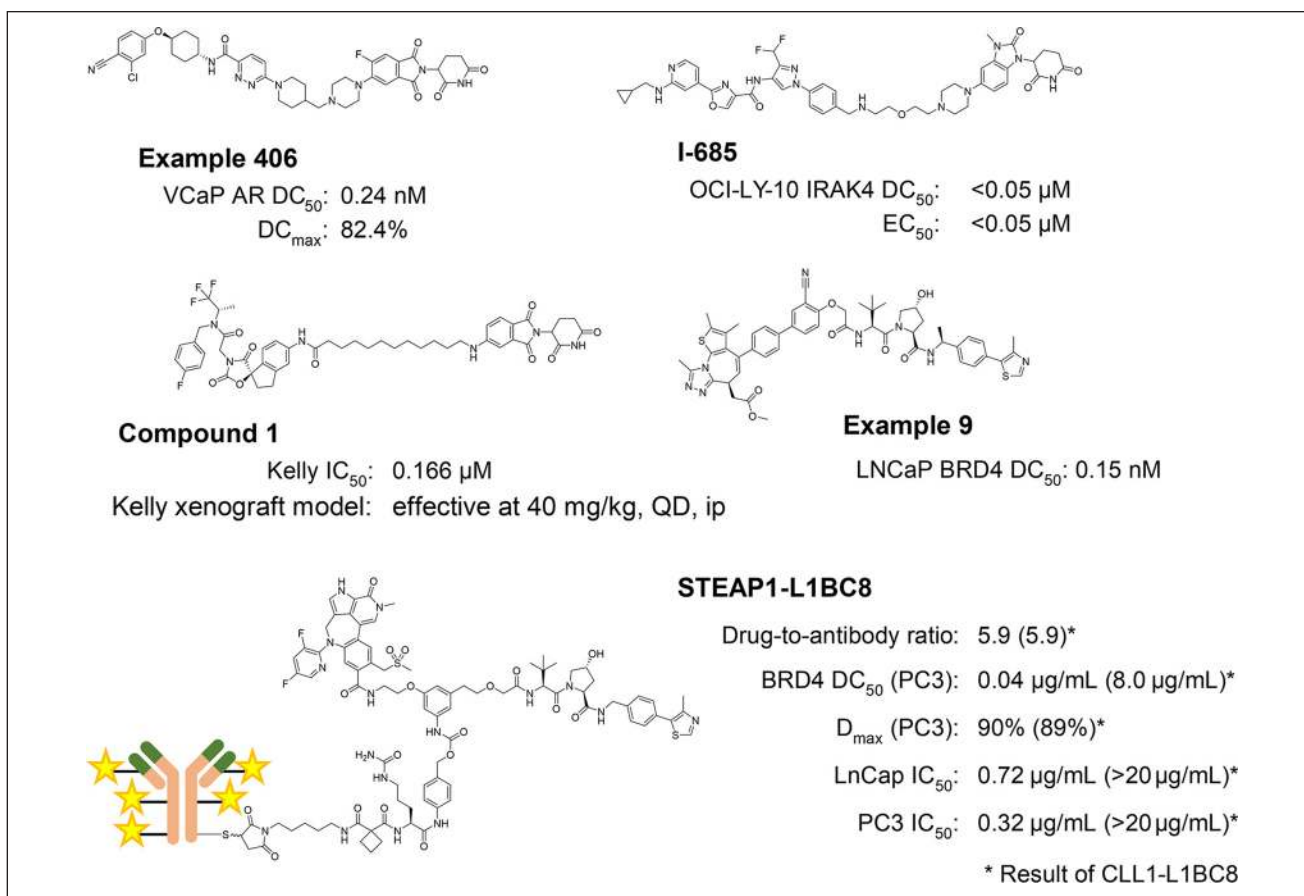


Figure 3. Representative structures and biological data of recently disclosed patents: 406,¹⁴³ I-685,¹⁴⁴ Compound 1,¹⁴⁵ Example 9,¹⁴⁶ and STEAP1-L1BC8.¹⁴⁷

cell apoptosis. Five out of eight IAPs, including cellular inhibitor of apoptosis-1 (cIAP1) and X-linked inhibitor of apoptosis protein (XIAP), also contain Really Interesting New Gene (RING) domains that bind to E2 conjugating enzymes, and so act as E3 ligases. Sekine et al. reported that methyl bestatin (MetBS; **Fig. 4A**) binds to the BIR3 domain of cIAP1 and induces self-ubiquitination following proteasomal degradation of cIAP1.³⁷ Structure-activity relationship studies of MetBS suggested that the bestatin portion binds to cIAP1, while the methyl ester is solvent exposed. Based on this observation, bifunctional small molecules were developed that comprise bestatin and all-trans retinoic acid (ATRA) linked by a polyethylene glycol (PEG) linker. These compounds were found to induce degradation of cellular retinoic acid binding protein-2 (CRABP2) (SNIPER2 and SNIPER4; **Fig. 4A**).^{5,38} The authors named this technology Specific and Nongenetic Inhibitor of Apoptosis Protein (IAP)-Dependent Protein Eraser (SNIPER) and later expanded it to other target proteins, such as estrogen receptor (ER),³⁹ BRD4,⁴⁰ and BCR-ABL⁴¹ [SNIPER(ER)-87, SNIPER(BRD4)-1, and SNIPER(ABL)-62; **Fig. 4A**]. One

characteristic feature of SNIPERs is that they can retain self-ubiquitination activity toward cIAP1. This effect may show synergistic activity to types of cancer cells in which cIAP1-induced degradation is beneficial. The self-degradation effect, however, reduces intracellular levels of cIAP1 and consequently causes suppression of degradation activity toward the target protein.⁴²

PROTACs Based on MDM2 Ligands. Tumor protein p53 is a transcription factor that regulates DNA repair, cell cycle, and apoptosis. p53 is mutated in more than 50% of cancers.⁴³ Mouse double minute 2 homolog (MDM2) protein is an E3 ubiquitin ligase that induces ubiquitination of p53 and subsequent proteasomal degradation of p53. Nutlin (**Fig. 4B**) is a well-known compound that binds to MDM2 and disrupts MDM2-p53 interaction, and it is among the first small molecules that were discovered to bind to E3 ligase proteins.⁴⁴ In 2008, Schneekloth and coworkers developed a nonpeptidic PROTAC that consisted of an AR ligand and the nutlin motif (PROTAC 14; **Fig. 4B**).⁴⁵ Some more PROTACs based on nutlin or idasanutlin⁴⁶ were later

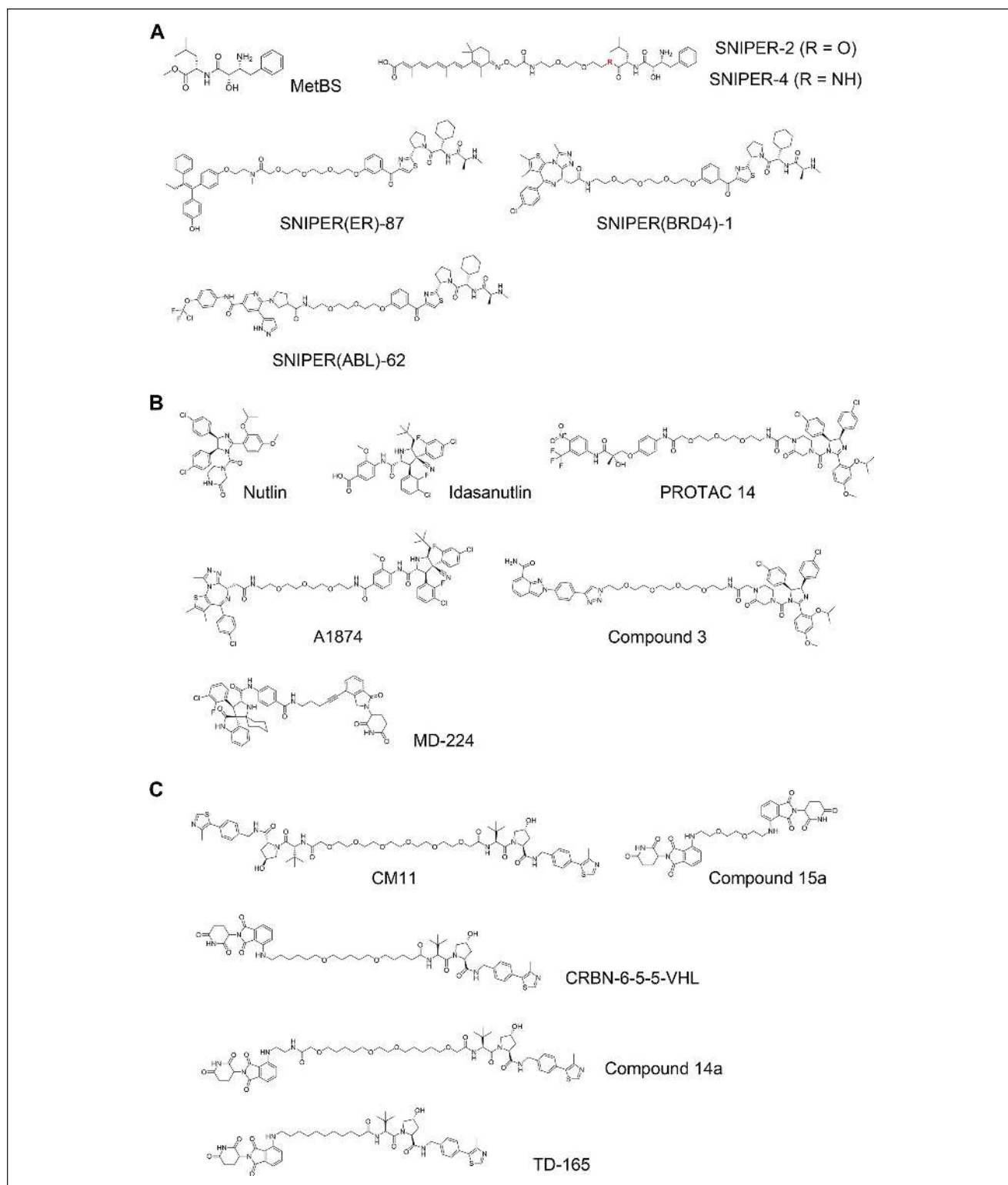


Figure 4. Inhibitor of apoptosis protein (IAP)-based and mouse double minute 2 homolog (MDM2)-based proteolysis-targeting chimeras (PROTACs), and von Hippel–Lindau (VHL)-based and cereblon (CRBN)-based homo-PROTACs. **(A)** Exemplified structures of methyl bestatin (MetBS) and Specific and Nongenetic Inhibitors of Apoptosis Protein (IAP)-Dependent Protein Eraser (SNIPERs): MetBS,³⁷ SNIPER-2 (R = O)⁵ and SNIPER-4 (R = NH),³⁸ SNIPER(ER)-87,³⁹ SNIPER(BRD4)-1,⁴⁰ and SNIPER(ABL)-62.⁴¹ **(B)** Structures of MDM2 inhibitors and PROTACs incorporating with them: nutlin,⁴⁴ idasanutlin,⁴⁶ PROTAC 14,⁴⁵ A1874,¹⁴⁸ Compound 3,¹⁴⁹ and MD-224.⁴⁷ **(C)** Structures of homo-PROTACs and related PROTACs composed with CRBN and VHL ligands: CM11,⁴⁸ Compound 15a,⁴⁹ CRBN-6-5-5-VHL,⁵⁰ Compound 14a,⁵¹ and TD-165.⁵²

developed (**Fig. 4B**), but the number of MDM2 PROTACs has remained limited, due to their challenging physico-chemical profile and limited degradation activity. PROTACs composed of a ligand for CRBN linked to a ligand for MDM2 were found to potently degrade MDM2 protein selectively (MD-224; **Fig. 4B**).⁴⁷ That implies that the degradation efficiency and/or activity of MDM2 is supposed to be limited compared to the Cullin RING ligase complexes formed by CRBN or VHL.

Homo-PROTACs. The mode of action of PROTACs toward POIs was expanded by the development of unusual homobifunctional molecules that comprised two ligands for the same E3 ligase. Maniaci et al. developed a novel type of PROTACs that were dimers of two VHL ligands joined by PEG linkers, so-called homo-PROTACs.⁴⁸ The best compound, CM11 (**Fig. 4C**), induced dimerization of CRL2^{VHL} with high cooperativity (~20) in vitro and induced intracellular VHL degradation at >100-fold lower concentration of the binary K_d value. CM11 also induced degradation of CUL2, suggesting a mechanism of bystander ubiquitination and degradation of a subunit different from that being recruited by the homo-PROTAC within the context of the whole CRL complex. Later, Steinebach et al. extended this approach to CRBN and developed homo-PROTACs that degrade CRL4^{CRBN} ligase (Compound 15a; **Fig. 4C**).⁴⁹ Two groups, Steinebach et al. and Girardini et al., then independently reported hetero-bifunctional compounds that are composed with a ligand for CRBN at one end, and a ligand for VHL at the other end (CRBN-6-5-5-VHL and Compound 14a; **Fig. 4C**). Both groups demonstrated that their PROTACs preferably induced the degradation of CRL4^{CRBN} rather than CRL2^{VHL}.^{50,51} The same finding was subsequently reported by a third group (TD-165; **Fig. 4C**).⁵² All three groups developed PROTAC degraders of slightly different chemical structures, yet in all cases preferential degradation of CRBN was observed, suggesting that the homo-PROTAC approach could be used to hijack E3 ligases against each other for inducing selective E3 ligase degradation.

Expansion of the E3 Ligase Toolbox for PROTACs

Currently, PROTAC design and development center on the use of primarily VHL and CRBN ligands, and to some extent IAP ligands. There are, however, more than 600 E3 ligases known to function in human cells, suggesting a large untapped pool of E3 ligases that are potentially hijackable for targeted protein degradation. To expand the scope of the PROTAC modality, efforts to expand the E3 ligase toolbox with novel chemistries have received significant efforts. Next, we briefly highlight the main progress from the past 4–5 years at finding ligands for new E3 ligases.

KEAP1. Kelch-like ECH-associated protein-1 (KEAP1) is known to interact with nuclear factor erythroid 2-related factor-2 (Nrf2). The main function of KEAP1 is to regulate cellular responses for chemical and oxidative stress by its role as a substrate recognition subunit of a cullin-3 (CUL3) E3 ligase to bind to induce ubiquitination and degradation of substrate Nrf2.⁵³ Academic and industrial groups have throughout the years developed a variety of small molecules that disrupt the KEAP1–Nrf2 interaction based on natural bioactive compounds,^{54–57} high-throughput screening (HTS),^{58–60} virtual screening,^{61,62} or fragment-based drug discovery (FBDD).^{63,64} In 2018, the first PROTACs recruiting the KEAP1–CUL3 E3 ligase were reported that were shown to induce degradation of intracellular Tau at micromolar concentration.⁶⁵ The peptidic nature of the PROTACs, however, which used an Nrf2-based epitope peptide as KEAP1 ligand, limits their potential for expanding their use as in vivo tools for clinical application. Recently, Tong et al. developed the first nonpeptidic PROTACs that consist of JQ1 and bardoxolone methyl (CDDO-Me) as ligands of BET bromodomains and KEAP1, respectively (**Fig. 5A**).⁶⁶ They found that the degradation activity of BRD4 was diminished by removing the Michael acceptor motif of CDDO-Me, indicating that formation of covalent bonds between cysteines on KEAP1 and the Michael acceptor moiety on CDDO-Me is essential to recruit BRD4 to the KEAP1 E3 ligase.

DCAF15. Sulfonamide derivatives indisulam (E7070),⁶⁷ E7820,⁶⁸ and chloroquinoxaline sulfonamide (CQS)⁶⁹ were found to have antitumor activity and were tested in the clinic for the treatment of cancers (**Fig. 5B**). Yokoi et al. reported that a cancer cell line resistant to indisulam was also resistant to E7820 and CQS, but not to doxorubicin and paclitaxel;⁷⁰ however, the mechanism of action of those sulfonamide derivatives remained unclear for a long time. In 2017, two groups, Uehara et al.⁷¹ and Ting et al.,⁷² independently reported that these sulfonamides induce ubiquitination and subsequently proteasomal degradation of coactivator of activating protein-1 and estrogen receptors (CAPER α) via formation of a CAPER α –sulfonamide–DCAF15–DDB1–CUL4 complex. In 2019, three independent groups released cocrystal structures of CAPER α –sulfonamide–DCAF15–DDB1–DDA1 multimeric complexes and confirmed that sulfonamides induce formation of the complexes as a molecular glue.^{73–75} Their structural information could help to further optimize sulfonamides using structure-based drug design and leverage them as DCAF15 ligands for PROTAC design. Until recently, however, it has remained challenging to develop indisulam-based DCAF15-recruiting PROTACs,^{76,77} most likely because of the weak binding affinities of these compounds to DCAF15 alone, or perhaps due to unsuitable orientation between the binding pocket of sulfonamides and the RING domain of CUL4.⁷⁴ In 2020, Li

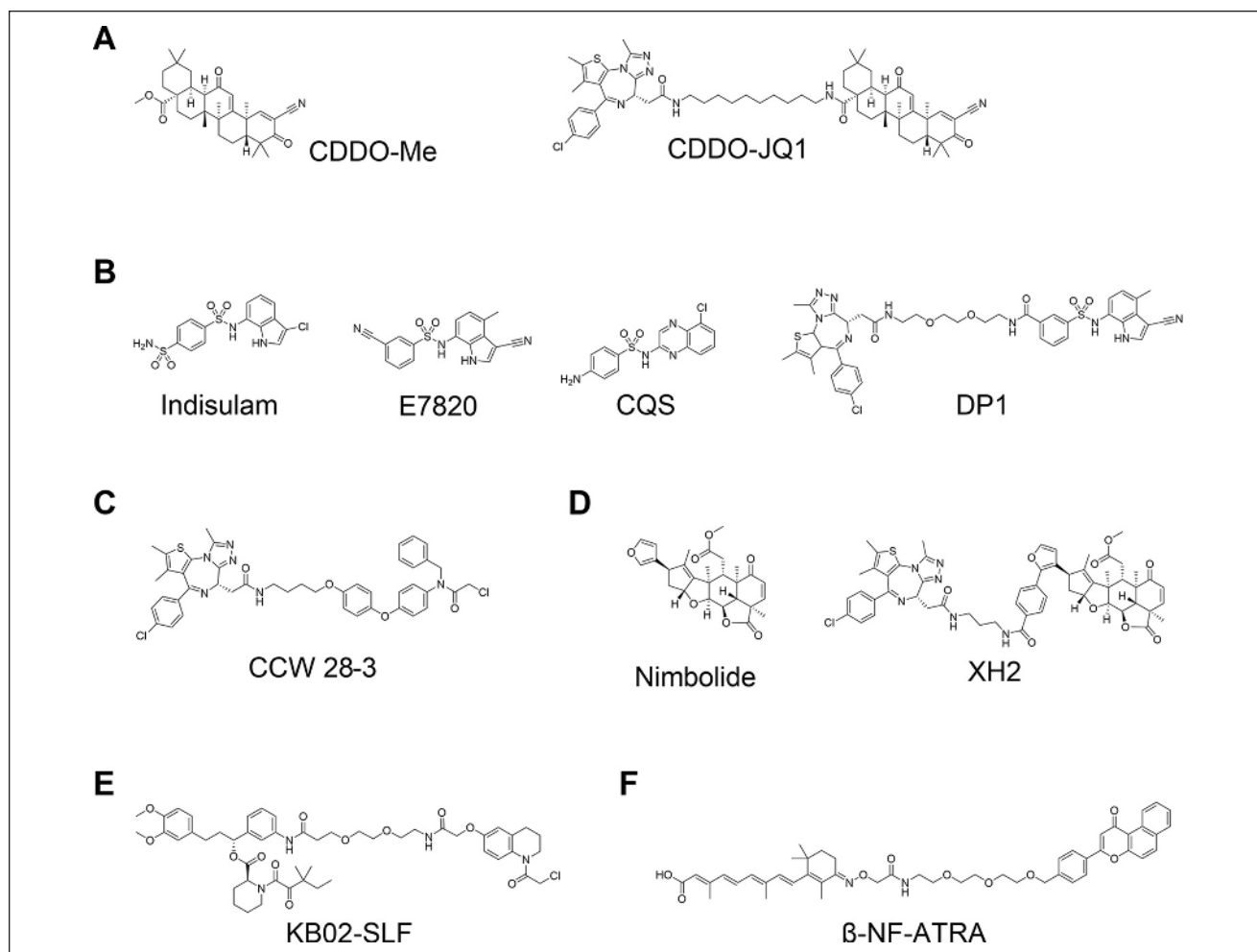


Figure 5. Proteolysis-targeting chimeras (PROTACs) using nonconventional E3 ligase ligands. **(A)** Structures of CDDO-Me^{56,57} and CDDO-JQ1.⁶⁶ **(B)** Structures of sulfonamide derivatives, indisulam,⁶⁷ E7820,⁶⁸ CQS,⁶⁹ and DP1.⁷⁸ **(C)** Structure of CCW 28-3.⁷⁹ **(D)** Structures of nimbolide and XH2.⁸⁰ **(E)** Structure of KB02-SLF.⁸¹ **(F)** Structure of β -NF-ATRA.⁸²

et al. reported a first aryl sulfonamide-based PROTAC, DP1 (**Fig. 5B**), that induces degradation of BRD4 and shows tumor growth inhibitory activity in mouse models *in vivo*. The biological activity of DP1 is limited, but the results demonstrate that DCAF15 can be applied for development of PROTACs.⁷⁸ Aryl sulfonamide derivatives have a good physicochemical profile (i.e., lower logD values and lower molecular weight compared to most E3 ligase ligands available); therefore, DCAF15 is likely to gain increased attention, and more DCAF15-mediated PROTACs are expected to be developed in the future.

Other E3 Ligases for PROTACs: RNF4, RNF114, DCAF16, and AhR. Unconventional approaches have shown promising applications to find new chemistry to expand the repertoire of E3 ligases for developing PROTACs. Ward et al. found novel covalent RNF4 binders by activity-based protein profiling (ABPP)-based screening, and then their optimized

ligand was applied to develop a novel type of PROTAC (CCW 28-3; **Fig. 5C**) to target BRD4 for degradation.⁷⁹ Around the same time, Spradlin and coworkers reported that nimbolide (**Figure 5D**), a natural compound that exhibits biological activities in cancer cells, was identified as a covalent binder for the E3 ligase RNF114.⁸⁰ In addition, they also developed a PROTAC that comprised nimbolide and JQ1 (XH2; **Fig. 5D**), and demonstrated that it induced some degradation activity of endogenous BRD4. Zhang et al. also synthesized hetero-bifunctional molecules consisting of a ligand for FKBP12 and three different electrophilic warheads, joined by a PEG linker.⁸¹ They treated HEK293T cells with those compounds and found that KB02-SLF (**Fig. 5E**) showed moderate degradation activity on FKBP12. Proteomics analysis and a following confirmatory pulldown SILAC (stable isotope labeling by amino acids in cell culture) experiment revealed that KB02-SLF degrades FKBP12 via recruitment of CRL4^{DCAF16}. Ohoka et al. also reported a

PROTAC that uses a novel E3 ligase.⁸² They chose to focus on the arylhydrocarbon receptor (AhR) E3 ligase and developed PROTACs based on ligands of AhR. β -NF-ATRA (**Fig. 5F**), a PROTAC that directed CRABPs, induced degradation of CRABP1 with AhR dependency, albeit with the fastidious effect of inducing self-degradation of AhR, akin to the mode of action of some SNIPERS.^{39–41}

Target-Based Screening versus Phenotypic Screening Approaches

The exciting progress during the past few years in targeting E3 ubiquitin ligases with more drug-like small molecules, and in using them for developing cell-active PROTAC degraders, motivates the deployment of known, and the development of new, hit- and ligand-finding approaches to screen E3 ligases to discover new chemical matter against them.⁸³ HTS against large compound collections is a well-established approach to find hit compounds for “druggable” targets [e.g., kinases, G protein-coupled receptors (GPCRs), and ion channels]. HTS campaigns for finding ligands of E3 ligases are not, however, straightforward. Among the reasons are difficulties to select a proper readout to detect compound activity against a targeted E3 ligase, because the ubiquitination activity of E3 ligases does not provide straightforward readouts that can be monitored in bioassays. Targeting of the substrate binding site of E3 ligases has shown much success for developing inhibitors and PROTACs (e.g., for VHL, CRBN, and IAPs), but that requires the disruption of protein–protein interactions, which are challenging targets for drug discovery.⁸⁴ Moreover, many substrate-binding pockets on E3 ligases and E3 substrate receptor subunits are shallow or highly charged, features that do not make them tractable targets with classical compound libraries. Because of the reasons mentioned above, and because for PROTAC development only binding ligands are needed, regardless of their inhibition activity on their own, other approaches are being developed and applied to find new binders of E3 ligases.

Fragment-Based Drug Design (FBDD) Approach. E3 ligands for PROTACs are not required to have functional activity in their own right against their targeted ligase. In principle, it suffices that they “bind” to a pocket or surface of the E3 ligase, to provide a suitable starting point for the design of the bifunctional PROTACs. To this end, fragment-based screening is a gold standard method to find new binding pockets and new ligands for proteins. Lucas et al. screened more than 1200 small compounds that complied with the “rule of three”⁸⁵ using differential scanning fluorometry (DSF) against the VHL–ElonginB–ElonginC complex of E3 ligase.⁸⁶ Primary hit compounds were validated by proton nuclear magnetic resonance (¹H NMR) spectroscopy and

X-ray crystal structure analysis, and then they identified three hit compounds that have weak but measurable binding affinity at two pockets distinct from the HIF1 α recognition site: one involving the EloC–CUL2 interface, and the other one involving a previously unknown cryptic pocket in VHL. Fragment-based approaches were also applied to find nonpeptidic ligands for substrate recognition sites on E3 ligases. Researchers at Astex Pharmaceuticals conducted fragment library screening over IAPs and identified nonpeptidic hit compounds, even though their binding affinity to IAPs was in the weak millimolar range of dissociation constants.⁸⁷ They next elaborated hit compounds by structure-guided optimization and medicinal chemistry effort, and finally obtained ASTX660, which inhibits cIAP1 and XIAP at nanomolar levels (**Fig. 6A**).^{87–89} ASTX660 is currently in a Phase II trial being investigated for the treatment of advanced solid tumors and lymphomas.

Screening of noncovalent fragment libraries will continue to play a major role in our efforts to find suitable starting points for E3 ligase ligand design. Fragment-based screening with small molecular covalent binders is emerging as an attractive approach too, however, and therefore should provide an alternative and powerful strategy to identify covalent binders at targetable hotspots of E3 ligases. Resnick and coworkers constructed a library of about 1000 small fragments that have acrylamide or chloroacetamide as covalent-binding warheads.⁹⁰ After validating their nonselective reactivity to thiols, they screened them against 10 proteins that have reactive cysteines by liquid chromatography–mass spectrometry (LC-MS) analysis and identified two fragments that bind OTUB2 or NUDT7. In addition, a variety of covalent warheads for reactive amino acid residues were developed (**Fig. 6B–6G**), allowing us to expand the arsenal of covalent-binding small-molecule fragments.⁹¹ As mentioned before, there are initial examples that PROTACs based on ligands that covalently bind to E3 ligases were shown to have degradation activity for targeted proteins,^{65,78–80} and that means that weak covalent-binding fragments could be applied to develop novel PROTAC degraders. Therefore, these recent updates could help us to identify new covalent E3 ligase binders in the future.

Structure-Guided Drug Design Approach. Solving cocrystal structures of E3 ligases with fragment peptides of substrate proteins is also a major strategy to guide the design of small molecular binders of the ligase. A notable success of this approach has been the development of nonpeptidic ligands of VHL, as described in the section titled “VHL Ligands and Applications to PROTACs,” which combined information provided by the binding of the substrate HIF peptide with fragment-based ligand design. Solving more examples of cocrystal structures of other E3 ligases with corresponding fragment substrate peptides will therefore be important.

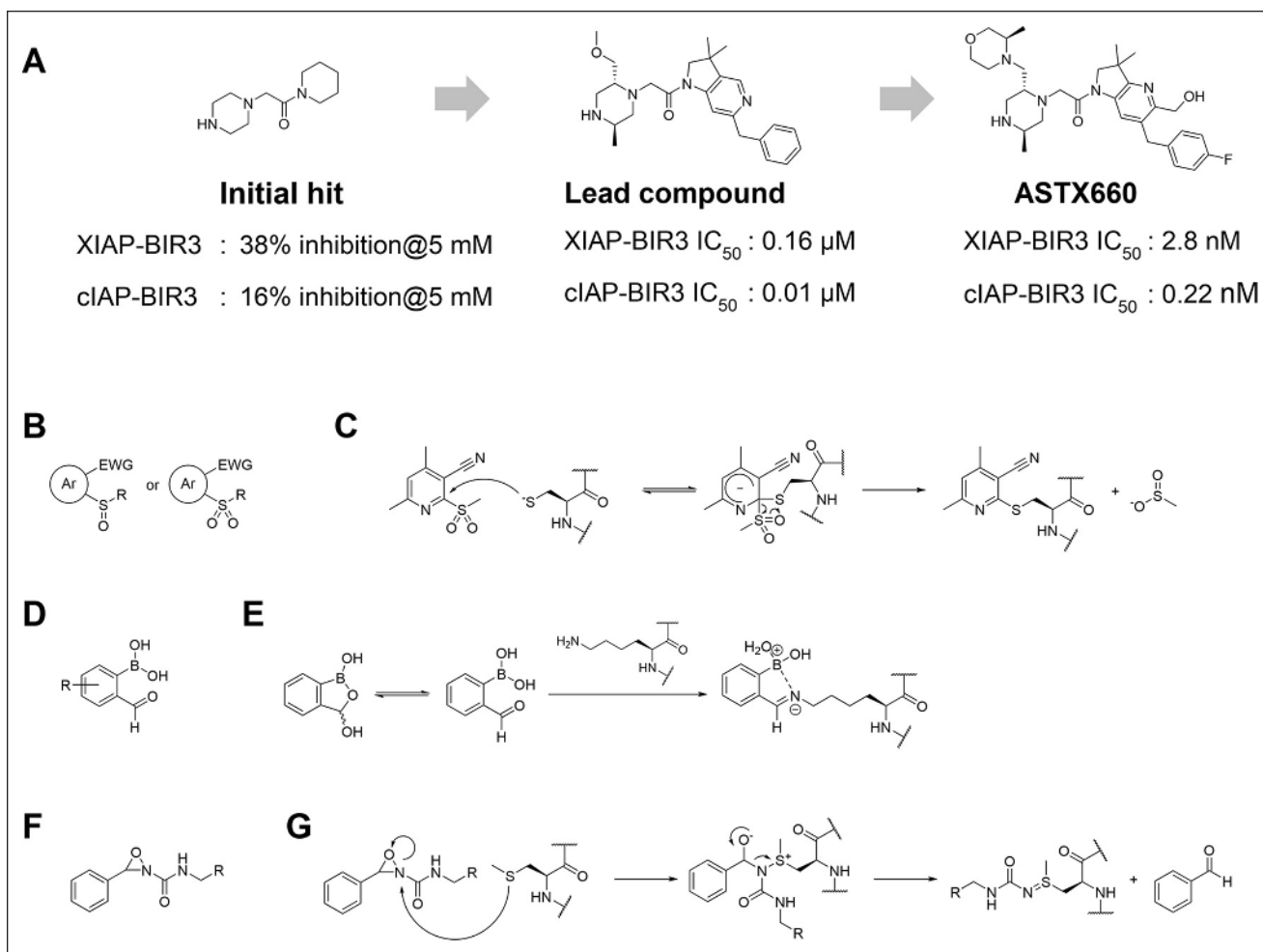


Figure 6. Noncovalent and covalent fragment-based approaches for hit finding and ligand development for E3 ligases. **(A)** Initial hit compound to the final clinical compound, ASTX660.^{87–89} **(B–F)** Examples of specific residue-selective covalent warheads: **(B)** cysteine,¹⁵⁰ **(D)** lysine,¹⁵¹ and **(F)** methionine;¹⁵² and **(C,E,G)** the reaction mechanisms.

SOCS2

Kung et al. reported cocrystal structures of suppressor of cytokine signaling-2 (SOCS2)–ElonginB–ElonginC complexes with 11-residue phosphopeptides of erythropoietin receptor or growth hormone receptor that center their binding on the Tyr426 and Tyr595 regions, respectively.⁹² Superposition of both cocrystal structures revealed essential key interactions of both substrate peptides to SOCS2 and now provide guiding features for designing nonpeptidic small ligands. Phosphotyrosine was found to have a respectable binding affinity for SOCS2 as a zwitterionic amino acid (in the 10^{−4} M range of dissociation constant), suggesting that it could provide a suitable starting point for ligand design.⁹³ The poor permeability of the negatively charged phosphotyrosine group will, however, impose some hurdles to be circumvented in their development as cell-active compounds. In addition, a cocrystal structure of suppressor of

cytokine signaling-6 (SOCS6) with a substrate peptide was reported.⁹⁴

KLHL12

Kelch-like protein-12 (KLHL12) forms the CUL3–E3 ubiquitin ligase complex, and it is known to negatively regulate the Wnt-signaling pathway by ubiquitination and subsequent proteasomal degradation against segment polarity protein Dishevelled homolog-3 (Dvl3). Zhao et al. measured binding affinity of several peptide fragments of C-terminal of Dvl3 to KLHL12 by ¹H–¹⁵N heteronuclear single quantum coherence (HSQC) NMR analysis and identified the key peptide motif for binding to KLHL12.⁹⁵ They then solved a cocrystal structure of KLHL12 with a truncated peptide of Dvl3 and found important hydrophobic interaction of prolines with KLHL12.

Table 1. List of E3 Ligases for Which 3D Crystal Structures Are Reported with Bound Ligands or Partner Substrate Proteins, but That Are Yet to Be Explored for PROTAC Applications.

E3 Ligase	Class	Ligand	Pocket	Reference
UBR2	UBR	Peptide	Positive/shallow	153
SPOP	BTB	Peptide	Shallow	154,155
KLHL3	BTB	Peptide	Negative/shallow	156
KLHL12	BTB	Peptide	Shallow	95
KLHL20	BTB	Peptide	Negative/shallow	157
KLHDC2	BTB	Peptide	Negative/deep	96
SPSB1	SPRY	Peptide	Shallow	158
SPSB2	SPRY	Peptide	Shallow	158–160
SPSB4	SPRY	Peptide	Shallow	160
SOCS2	SOCS	Peptide	Negative/shallow	92
SOCS6	SOCS	Peptide	Negative	94
FBXO4	FBXO	Protein	Shallow	161
FBXO31	FBXO	Peptide	Negative	162
BTRC	FBXW	Peptide	Negative	163
FBW7	FBXW	Peptide	Negative	164,165
CDC20	APC/C	Peptide	Shallow	166
CDC20	APC/C	Small molecule	Hydrophobic	167
ITCH	HECT	Peptide	Shallow	168,169
PML	TRIM	Peptide	Shallow	170
TRIM21	TRIM	Peptide	Shallow	171
TRIM24	TRIM	Peptide	Shallow	172
TRIM24	TRIM	Small molecule	Deep	173,174
TRIM33	TRIM	Peptide	Shallow	175
GID4	CTLH	Peptide	Positive/deep	98

3D: Three-dimensional; PROTAC: proteolysis-targeting chimera.

KLHDC2

Rusnac et al. reported cocrystal structures of Kelch domain-containing protein-2 (KLHDC2), which is one of the BTB E3 ligases and acts as a substrate receptor of the CRL2 E3 ligase complex, with an eight-amino-acid C-end degron peptide of substrate proteins.⁹⁶ Those structures showed that these peptides were bound to the KLHDC2 pocket in a twisted conformation, and the C-terminal diglycine degron motif was fitted in a tight groove with strong coordination of the terminal carboxyl group via three highly conserved residues of KHDLC2.

GID4

Glucose-induced degradation protein-4 homolog (GID4) is one of the gastric inhibitory polypeptide (GIP) family and part of a GID family that composes the CTLH E3 ligase complex. GID4 recognizes N-terminal proline residues selectively in substrates to mark their ubiquitination and proteasomal degradation,⁹⁷ but the mechanism of molecular recognition remained elusive. Dong et al. reported the cocrystal structures of the globular domain of GID4 with N-end proline fragments and observed that the terminal

proline forms key interactions with the residues at the bottom of a deep substrate binding pocket.⁹⁸ In addition, the binding of degrons induces dynamic conformational change around the binding pocket to form hydrophobic interaction with the residues in the pocket.

There are other E3 ligases with reported 3D cocrystal structures containing ligands and proteins that have yet to be exploited for the development of PROTACs (**Table 1**). More structures of E3 ligases with or without bound substrates or small-molecule ligands are expected to be made available in the near future, and they could serve as a starting point for ligand development. Although some ligand-binding pockets seem unsuitable for designing peptidomimetic ligands for PROTACs (e.g., they have a wide and shallow binding area, or a highly charged pocket), a structure-based approach using structural information is one of the most attractive ways to design peptidomimetic ligands for disease-specific E3 ligases and apply them to PROTACs. Toward this goal, our laboratory collaborates as part of a large consortium named EUbOPEN (Enabling & Unlocking Biology in the OPEN), an Innovative Medicines Initiative (IMI)-funded project whose goal is to enable and unlock biology by developing chemical probes and ligands for novel protein

classes, including E3 ubiquitin ligases (<https://www.eubopen.org>). The consortium comprises 22 different partner organizations, including universities, research institutes, European Federation of Pharmaceutical Industries and Associations (EFPIA) members, and small and medium-sized enterprises (SMEs). EUBOPEN will operate under the auspices of a longer-term initiative called Target 2035.^{99,100}

Other Target-Based Approaches

HTS is a well-known strategy to identify compounds that bind to target proteins; however, even large compound libraries (e.g., composed of millions of compounds, as available to large pharmaceutical companies) can only cover a tiny area of chemical space. To address this issue, DNA-encoded libraries (DELs) are an attractive combinatorial approach that allows extension of chemical space coverage by small molecules and so is increasingly being applied to drug discovery in academia and pharmaceutical companies.¹⁰¹ One of the most widely applied methods to construct DELs is a split-and-pool approach. This type of DEL usually consists of two to four building blocks, enabling a theoretical single DEL that can have more than a billion compounds in some cases. One of the limitations of DELs is that the libraries can be screened with only cell-free assay systems (e.g., against recombinant purified target proteins); therefore, it is challenging to measure the biological activity of hit compounds directly in a cellular environment. Screening target proteins *in vitro* means that DELs are a method of choice for the identification of binding ligands, irrespective of their functionality in bioassays or in cells. To this end, DELs are potentially highly suited as an approach to screen for new E3 ligase binders for developing PROTACs. Indeed, several pharmaceutical and protein degradation biotech companies, including Arvinas, Nurix, and Kymera, have disclosed that they are pursuing applications of DEL technologies (either developed as proprietary in house, or via partnerships) to find new E3 ligase ligands. Perhaps curiously, however, no peer-reviewed publication has yet emerged describing the application of DEL technology to identify novel E3 ligase ligands for PROTACs, potentially suggesting specific challenges with targeting the protein surface and protein–protein interaction binding sites that are required to be targeted on E3 ligases. In contrast, novel DELs designed for targeting E3 ligases, for instance covalent DELs,^{102,103} should provide an attractive approach to discover selective E3 binders and develop novel PROTACs. In fact, some pharmaceutical companies have announced increasing activity and investment to expand the repertoire of E3 ligases for PROTACs by DEL technology.

Phage display technology is one of the greatest ways to obtain peptide ligands for target proteins for which natural substrates and ligands have not been identified.¹⁰⁴ In this

technology, a randomized synthetic DNA library is inserted into a phage coat protein gene in bacteriophages to express peptides of desired length onto their coat proteins. The obtained “phage library” is incubated with immobilized target (protein, cDNA, antibody, enzyme, small molecule, etc.) and then washed out to remove unbound phages. The remaining phages are eluted and then used to infect bacterial cells to amplify phages for the next cycle. After a few rounds of selection, phages that express high-affinity peptides to the target are accumulated and could be sequenced to identify these peptides. This approach is also suitable to isolate appropriate peptide ligands for developing PROTACs for the same reason as with DELs. It requires additional medicinal chemistry to develop peptidomimetic ligands and improve drug-like properties; however, peptidic PROTACs could be developed rapidly based on peptides identified by phage display as chemical tools to study the recruitment of specific E3 ligases for targeted protein degradation.

In addition to allowing identification of linear peptides, phage display, yeast display, and other display technologies can be used to identify and develop cyclic and macrocyclic peptides too. Cyclic peptides are structurally constrained macrocycles that have emerged as a promising class of molecules to bind to and so target proteins, including targeting protein surfaces and protein–protein interactions in targets once considered unligandable and so undruggable.^{105,106} Due to their constrained “locked” nature and potential to form intramolecular hydrogen bonds, cyclic peptides can exhibit greater permeability and higher metabolic stability than their linear counterparts, making them attractive as chemical tools to probe novel biology.^{107,108}

Several approaches are being pursued to develop macrocyclic peptides as bioactive or binding/affinity agents. These include rational design,¹⁰⁹ DNA-templated synthesis and selection,¹¹⁰ phage-encoded combinatorial display of bicyclic peptides,^{111,112} and messenger RNA (mRNA) display followed by cyclization.¹¹³

Among the mRNA display methods available, the Random Nonstandard Peptides Integrated Discovery (RaPID) system involves peptide spontaneous cyclization and selection, and allows flexible incorporation of nonproteinogenic amino acids. RaPID has demonstrated success at generating cyclic peptide ligands of high binding affinity and specificity for a variety of protein targets.^{114,115} In an early related application, cyclic peptides were identified targeting E6AP, a HECT E3 ligase target involved in p53-dependent apoptosis.¹¹⁶

Phenotypic Screening Approach

Molecular glues are attractive starting points for identifying applicable E3 ligases for PROTACs; however, the previously identified molecular glues, immunomodulatory drugs

(IMiDs) and sulfonamides, were found serendipitously. For instance, indisulam is a representative compound in the sulfonamide-type molecular glues that was discovered by cell growth inhibitory assay and cell cycle analysis without understanding of its mode of action at that time.¹¹⁷ The major challenge for the discovery of new molecular glues is a limited availability of rational strategies for their identification or design that would induce proteasomal degradation of target proteins. Recently, strategies and approaches have been developed to begin to address these shortcomings. Słabicki et al. analyzed cytotoxicity data of 4518 clinical and preclinical small molecules against 578 cancer cell lines to show correlation between cytotoxicity and expression level of 499 E3 ligase mRNA.¹¹⁸ They found that pan cyclin-dependent kinase (CDK) inhibitor CR8 shows a level of correlation between cytotoxicity of each cell line and the mRNA levels in the same cell line of CDK12 and DNA damage-binding protein-1 (DDB1), which is a component of DDB1–CUL4 E3 ligase complexes. Treatment of CR8 was found to induce degradation of cyclin K without changing the cyclin K mRNA level, suggesting that cyclin K degradation might occur as a result of its ubiquitination via DDB1-associated ubiquitin–proteasome machineries. The cocrystal structure of the cyclin K–CDK12–CR8–DDB1 complex revealed that CDK12 formed extensive protein–protein interactions to DDB1 directly, essentially mimicking interactions formed by DDB1-associated substrate recognition subunits (also known as DCAFs), and that CR8 acted as a molecular glue to enhance this weak-affinity interaction, bringing it to sufficient stability to induce efficient ubiquitination and proteasomal degradation of the CDK12-associated subunit cyclin K. In another study, Mayor-Ruiz et al. performed focused screens in neddylation-impaired cell lines that had the neddylation E2 conjugating enzyme Ube2M knocked out, to bias identification of cytotoxic compounds that were preferentially active and so required neddylation-competent CRL machineries. These screens enabled the authors to find novel chemotypes as hits that were shown to act as molecular glue degraders via DCAF15–CAPER α and DDB1–CDK12–CyclinK, akin to the mode of action of indisulam-like sulfonamides and CR8.¹¹⁹ These reports not only provide a first step toward more systematic approaches to identify small molecules that act as molecular glues, but also expand opportunities to find molecules that hit new E3 ligases in an unbiased manner, in those cases allowing the discovery of molecules that do not interact with the substrate recognition subunit proteins of Cullin RING ligases directly.

Phenotypic screens both provide the opportunity to find molecular glues and can be applied to discover bifunctional PROTAC compounds. For example, anchoring a covalent warhead to a ligand of a target protein provides another way to fish for novel E3 ligases that could be applied to PROTACs, as demonstrated in the case of the studies

identifying DCAF16 and RNF4 as described in the section titled “Other E3 Ligases for PROTACs: RNF4, RNF114, DCAF16, and AhR.”^{79,81} Envisaged extensions of these approaches will involve synthesizing bifunctional molecules that consist of a ligand for a disease-relevant protein, linked to different covalent warheads. Cell lines treated with these compounds and subsequently assessed for degradation of POI represent a strategy to find novel covalently targeted E3 ligases.¹²⁰

Future Perspective

Enabled by the discoveries of high-quality, crystallographically defined ligands for the E3 ligases VHL and CRBN, PROTACs have had a meteoric resurgence in the limelight, a remarkable development that has culminated in the first PROTAC degraders being demonstrated as safe and efficacious in the clinic.^{6,7} These advances have marked a significant step forward and provided added confidence to drug hunters to embark on campaigns to develop PROTAC-based drugs for unmet medical needs, including cancers. It remains challenging, however, to efficiently degrade many proteins using PROTACs. This is because PROTACs need to form stable ternary complexes between the E3 ligase and POI, and PROTAC-induced protein–protein interactions are important to facilitate efficient and selective target ubiquitination and degradation.^{121,122} It follows that for given target–E3 ligase combinations, it may be challenging to achieve ternary complexes of sufficient stability and suitable properties that make them productive to downstream target ubiquitination and degradation.

The lack of suitable chemical matters for most E3 ligases and undruggable disease targets represents a major stumbling block to our ability to efficiently and effectively navigate degrader drug space. Therefore, there is no doubt that enabling more E3 ligases with new chemistry and new binding ligands becomes an important objective that, when met, will augment the applications and potential of PROTACs to expand in scope to more disease-relevant target proteins. In forecasting novel E3 ligase discovery, we believe that the variety of approaches we have highlighted in this review should help to enrich the armory of E3 ligases that can become amenable to PROTAC drug development. The tremendous progress seen in recent years in technologies such as atomic-level cryogenic electron microscopy (cryo-EM) analysis,^{123,124} refined protein modeling by next-generation computer-aided design and artificial intelligence,¹²⁵ and others are expected to accelerate this area of research dramatically. Using disease-, tissue-, and/or organ-specific E3 ligases to develop PROTACs should allow for more cell-type-selective degraders, to aid better understanding of targets' biology and usher in safer drugs with reduced dose-limiting toxicities. To evaluate protein degradation activities more accurately, however, it is essential to better

understand not only the localization of target proteins and E3 ligases but also their dynamics in cells under disease-oriented microenvironments. In addition, there is still a long way to go to develop rational, predictive, a priori strategies to design PROTACs that form stable and cooperative ternary complexes between a target protein and E3 ligase. Nonetheless, comprehensive in silico approaches, for instance molecular dynamics simulation and prediction models by artificial intelligence, aided by enormous computational capability (supercomputers and on-demand cloud computing), are ripe for supporting PROTAC design in the future.

PROTACs represent an emerging modality in drug discovery, as a prominent example of a new class of multispecific drugs that form proximity-induced protein–protein interactions between targeted proteins.^{126,127} Novel degrader molecules are now being developed hot on the heels of PROTACs, and they differentiate by the mechanism of action or the targets being recruited.¹²⁸ These approaches include monomeric protein degraders,^{129,130} autophagy-targeting chimeras (AUTACs),¹³¹ biological PROTACs (bioPROTACs),¹³² lysosome-targeting chimeras (LYTACs),¹³³ autophagosome-tethering compound (ATTEC),¹³⁴ and others. PROTACs remain frontrunners in the targeted protein degradation space, and their rise to popularity has affected the emergence of other modalities of induced proximity pharmacology, including bifunctional molecules recruiting enzymatic activities beyond E3 ubiquitin ligases, such as protein phosphatases¹³⁵ and protein kinases.¹³⁶ It could not be a more exciting time to start finding novel binders for E3 ligases and other unliganded target proteins.

Declaration of Conflicting Interests

The authors declared the following potential conflicts of interest with respect to the research, authorship, and/or publication of this article: The Ciulli laboratory receives or has received sponsored research support from Amphista Therapeutics, Boehringer Ingelheim, Eisai, Nurix Therapeutics, and Ono Pharmaceutical. A.C. is a scientific founder, shareholder, director, and consultant of Amphista Therapeutics, a company that is developing targeted protein degradation therapeutic platforms. T.I. is an employee of Eisai Co., Ltd.

Funding

The authors disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: The Ciulli laboratory research on targeting E3 ligases and PROTACs receives or has received funding from the European Research Council (ERC, Starting Grant ERC-2012-StG-311460 DrugE3CRLs) and the Innovative Medicines Initiative 2 (IMI2) Joint Undertaking under grant agreement No. 875510 (EUbOPEN project). The IMI2 Joint Undertaking receives support from the European Union's Horizon 2020 research and innovation program, European Federation of Pharmaceutical Industries and

Associations (EFPIA) companies, and associated partners KTH, OICR, Diamond, and McGill.

ORCID iD

Alessio Ciulli  <https://orcid.org/0000-0002-8654-1670>

References

1. Sakamoto, K. M.; Kim, K. B.; Kumagai, A.; et al. Protacs: Chimeric Molecules That Target Proteins to the Skp1-Cullin-F Box Complex for Ubiquitination and Degradation. *Proc. Natl. Acad. Sci. U.S.A.* **2001**, *98*, 8554–8559.
2. Schneekloth, J. S., Jr.; Fonseca, F. N.; Koldobskiy, M.; et al. Chemical Genetic Control of Protein Levels: Selective In Vivo Targeted Degradation. *J. Am. Chem. Soc.* **2004**, *126*, 3748–3754.
3. Winter, G. E.; Buckley, D. L.; Paulk, J.; et al. Drug Development. Phthalimide Conjugation as a Strategy for In Vivo Target Protein Degradation. *Science*. **2015**, *348*, 1376–1381.
4. Zengerle, M.; Chan, K.-H.; Ciulli, A. Selective Small Molecule Induced Degradation of the BET Bromodomain Protein BRD4. *ACS Chem. Biol.* **2015**, *10*, 1770–1777.
5. Itoh, Y.; Ishikawa, M.; Naito, M.; et al. Protein Knockdown Using Methyl Bestatin-Ligand Hybrid Molecules: Design and Synthesis of Inducers of Ubiquitination-Mediated Degradation of Cellular Retinoic Acid-Binding Proteins. *J. Am. Chem. Soc.* **2010**, *132*, 5820–5826.
6. Petrylak, D. P.; Gao, X.; Vogelzang, N. J.; et al. First-in-Human Phase I Study of ARV-110, an Androgen Receptor (AR) PROTAC Degradator in Patients (pts) with Metastatic Castrate-Resistant Prostate Cancer. *J. Clin. Oncol.* **2020**, *38* (15_suppl.), 3500.
7. Proof-of-Concept with PROTACs in Prostate Cancer. *Cancer Discov.* **2020**, *10*, 1084.
8. Semenza, G. L.; Wang, G. L. A Nuclear Factor Induced by Hypoxia via De Novo Protein Synthesis Binds to the Human Erythropoietin Gene Enhancer at a Site Required for Transcriptional Activation. *Mol. Cell. Biol.* **1992**, *12*, 5447–5454.
9. Gordan, J. D.; Thompson, C. B.; Simon, M. C. HIF and c-Myc: Sibling Rivals for Control of Cancer Cell Metabolism and Proliferation. *Cancer Cell*. **2007**, *12*, 108–113.
10. Tello, D.; Balsa, E.; Acosta-Iborra, B.; et al. Induction of the Mitochondrial NDUFA4L2 Protein by HIF-1 α Decreases Oxygen Consumption by Inhibiting Complex I Activity. *Cell Metab.* **2011**, *14*, 768–779.
11. Forsythe, J. A.; Jiang, B.-H.; Iyer, N. V.; et al. Activation of Vascular Endothelial Growth Factor Gene Transcription by Hypoxia-Inducible Factor 1. *Mol. Cell. Biol.* **1996**, *16*, 4604–4613.
12. Jiang, J.; Tang, Y.-L.; Liang, X.-H.; EMT: A New Vision of Hypoxia Promoting Cancer Progression. *Cancer Biol. Ther.* **2011**, *11*, 714–723.
13. Semenza, G. L. Molecular Mechanisms Mediating Metastasis of Hypoxic Breast Cancer Cells. *Trends Mol. Med.* **2012**, *18*, 534–543.
14. Maxwell, P. H.; Wiesener, M. S.; Chang, G.-W.; et al. The Tumour Suppressor Protein VHL Targets Hypoxia-Inducible

- Factors for Oxygen-Dependent Proteolysis. *Nature*. **1999**, 399, 271–275.
15. Maher, E. R.; Kaelin, W. G., Jr. Von Hippel-Lindau Disease. *Medicine (Baltimore)*. **1997**, 76, 381–391.
 16. Kamura, T.; Koepp, D. M.; Conrad, M. N.; et al. Rbx1, a Component of the VHL Tumor Suppressor Complex and SCF Ubiquitin Ligase. *Science*. **1999**, 284, 657–661.
 17. Stebbins, C. E.; Kaelin, W. G., Jr.; Pavletich, N. P. Structure of the VHL-ElonginC-ElonginB Complex: Implications for VHL Tumor Suppressor Function. *Science*. **1999**, 284, 455–461.
 18. Cardote, T. A. F.; Gadd, M. S.; Ciulli, A. Crystal Structure of the Cul2-Rbx1-EloBC-VHL Ubiquitin Ligase Complex. *Structure*. **2017**, 25, 901–911.e3.
 19. Hon, W.-C.; Wilson, M. I.; Harlos, K.; et al. Structural Basis for the Recognition of Hydroxyproline in HIF-1 Alpha by pVHL. *Nature*. **2002**, 417, 975–978.
 20. Min, J.-H.; Yang, H.; Ivan, M.; et al. Structure of an HIF-1alpha-pVHL Complex: Hydroxyproline Recognition in Signaling. *Science*. **2002**, 296, 1886–1889.
 21. Buckley, D. L.; Van Molle, I.; Gareiss, P. C.; et al. Targeting the von Hippel-Lindau E3 Ubiquitin Ligase Using Small Molecules to Disrupt the VHL/HIF-1 α Interaction. *J. Am. Chem. Soc.* **2012**, 134, 4465–4468.
 22. Buckley, D. L.; Gustafson, J. L.; Van Molle, I.; et al. Small-Molecule Inhibitors of the Interaction between the E3 Ligase VHL and HIF1 α . *Angew. Chem. Int. Ed.* **2012**, 51, 11463–11467.
 23. Van Molle, I.; Thomann, A.; Buckley, D. L.; et al. Dissecting Fragment-Based Lead Discovery at the von Hippel-Lindau Protein:Hypoxia Inducible Factor 1 α Protein-Protein Interface. *Chem. Biol.* **2012**, 19, 1300–1312.
 24. Galdeano, C.; Gadd, M. S.; Soares, P.; et al. Structure-Guided Design and Optimization of Small Molecules Targeting the Protein-Protein Interaction Between the von Hippel-Lindau (VHL) E3 Ubiquitin Ligase and the Hypoxia Inducible Factor (HIF) Alpha Subunit with In Vitro Nanomolar Affinities. *J. Med. Chem.* **2014**, 57, 8657–8663.
 25. Soares, P.; Gadd, M. S.; Frost, J.; et al. Group-Based Optimization of Potent and Cell-Active Inhibitors of the von Hippel-Lindau (VHL) E3 Ubiquitin Ligase: Structure-Activity Relationships Leading to the Chemical Probe (2S,4R)-1-((S)-2-(1-Cyanocyclopropanecarboxamido)-3,3-dimethylbutanoyl)-4-hydroxy-N-(4-(4-methylthiazol-5-yl)benzyl)pyrrolidine-2-carboxamide (VH298). *J. Med. Chem.* **2018**, 61, 599–618.
 26. Frost, J.; Galdeano, C.; Soares, P.; et al. Potent and Selective Chemical Probe of Hypoxic Signalling Downstream of HIF- α Hydroxylation via VHL Inhibition. *Nat. Commun.* **2016**, 7, 13312.
 27. Bondeson, D. P.; Mares, A.; Smith, I. E.; et al. Catalytic In Vivo Protein Knockdown by Small-Molecule PROTACs. *Nat. Chem. Biol.* **2015**, 11, 611–617.
 28. Vijay, V. M.; Ujjwala, K.; Urvashi, I. P. Thalidomide. *Bombay Hosp. J.* **2008**, 50, 472–476.
 29. Singhal, S.; Mehta, J.; Desikan, R.; et al. Antitumor Activity of Thalidomide in Refractory Multiple Myeloma. *N. Engl. J. Med.* **1999**, 341, 1565–1571.
 30. Ito, T.; Ando, H.; Suzuki, T.; et al. Identification of a Primary Target of Thalidomide Teratogenicity. *Science*. **2010**, 327, 1345–1350.
 31. Krönke, J.; Udeshi, N. D.; Narla, A.; et al. Lenalidomide Causes Selective Degradation of IKZF1 and IKZF3 in Multiple Myeloma Cells. *Science*. **2014**, 343, 301–305.
 32. Lu, G.; Middleton, R. E.; Sun, H.; et al. The Myeloma Drug Lenalidomide Promotes the Cereblon-Dependent Destruction of Ikaros Proteins. *Science*. **2014**, 343, 305–309.
 33. Fischer, E. S.; Böhm, K.; Lydeard, J. R.; et al. Structure of the DDB1-CRBN E3 Ubiquitin Ligase in Complex with Thalidomide. *Nature*. **2014**, 512, 49–53.
 34. Chamberlain, P. P.; Lopez-Girona, A.; Miller, K.; et al. Structure of the Human Cereblon-DDB1-Lenalidomide Complex Reveals Basis for Responsiveness to Thalidomide Analogs. *Nat. Struct. Mol. Biol.* **2014**, 21, 803–809.
 35. Lu, J.; Qian, Y.; Altieri, M.; et al. Hijacking the E3 Ubiquitin Ligase Cereblon to Efficiently Target BRD4. *Chem. Biol.* **2015**, 22, 755–763.
 36. Baell, J. B.; Holloway, G. A. New Substructure Filters for Removal of Pan Assay Interference Compounds (PAINS) from Screening Libraries and for Their Exclusion in Bioassays. *J. Med. Chem.* **2010**, 53, 2719–2740.
 37. Sekine, K.; Takubo, K.; Kikuchi, R.; et al. Small Molecules Destabilize cIAP11 by Activating Auto-Ubiquitylation. *J. Biol. Chem.* **2008**, 283, 8961–8968.
 38. Okuhira, K.; Ohoka, N.; Sai, K.; et al. Specific Degradation of CRABP-II via cIAP11-Mediated Ubiquitylation Induced by Hybrid Molecules That Crosslink cIAP11 and the Target Protein. *FEBS Lett.* **2011**, 585, 1147–1152.
 39. Ohoka, N.; Okuhira, K.; Ito, M.; et al. In Vivo Knockdown of Pathogenic Proteins via Specific and Nongenetic Inhibitor of Apoptosis Protein (IAP)-Dependent Protein Erasers (SNIPERs). *J. Biol. Chem.* **2017**, 292, 4556–4570.
 40. Ohoka, N.; Ujikawa, O.; Shimokawa, K.; et al. Different Degradation Mechanisms of Inhibitor of Apoptosis Proteins (IAPs) by the Specific and Nongenetic IAP1-Dependent Protein Eraser (SNIPER). *Chem. Pharm. Bull. (Tokyo)*. **2019**, 67, 203–209.
 41. Shimokawa, K.; Shibata, N.; Sameshima, T.; et al. Targeting the Allosteric Site of Oncoprotein BCR-ABL as an Alternative Strategy for Effective Target Protein Degradation. *ACS Med. Chem. Lett.* **2017**, 8, 1042–1047.
 42. Naito, M.; Ohoka, N.; Shibata, N. SNIPERs-Hijacking IAP Activity to Induce Protein Degradation. *Drug Discov. Today. Technol.* **2019**, 31, 35–42.
 43. Piette, J.; Neel, H.; Maréchal, V. Mdm2: Keeping p53 under Control. *Oncogene*. **1997**, 15, 1001–1010.
 44. Vassilev, L. T.; Vu, B. T.; Graves, B.; et al. In Vivo Activation of the p53 Pathway by Small-Molecule Antagonists of MDM2. *Science*. **2004**, 303, 844–848.
 45. Schneekloth, A. R.; Pucheault, M.; Tae, H. S.; et al. Targeted Intracellular Protein Degradation Induced by a Small Molecule: En Route to Chemical Proteomics. *Bioorg. Med. Chem. Lett.* **2008**, 18, 5904–5908.
 46. Ding, Q.; Zhang, Z.; Liu, J.-J.; et al. Discovery of RG7388, a Potent and Selective p53-MDM2 Inhibitor in Clinical Development. *J. Med. Chem.* **2013**, 56, 5979–5983.
 47. Li, Y.; Yang, J.; Aguilar, A.; et al. Discovery of MD-224 as a First-in-Class, Highly Potent, and Efficacious Proteolysis Targeting Chimera Murine Double Minute 2 Degradator

- Capable of Achieving Complete and Durable Tumor Regression. *J. Med. Chem.* **2019**, *62*, 448–466.
48. Maniaci, C.; Hughes, S. J.; Testa, A.; et al. Homo-PROTACs: Bivalent Small-Molecule Dimerizers of the VHL E3 Ubiquitin Ligase to Induce Self-Degradation. *Nat. Commun.* **2017**, *8*, 830.
49. Steinebach, C.; Lindner, S.; Udeshi, N. D.; et al. Homo-PROTACs for the Chemical Knockdown of Cereblon. *ACS Chem. Biol.* **2018**, *13*, 2771–2782.
50. Steinebach, C.; Kehm, H.; Lindner, S.; et al. PROTAC-Mediated Crosstalk between E3 Ligases. *Chem. Commun. (Camb)*. **2019**, *55*, 1821–1824.
51. Girardini, M.; Maniaci, C.; Hughes, S. J.; et al. Cereblon versus VHL: Hijacking E3 Ligases against Each Other Using PROTACs. *Bioorg. Med. Chem.* **2019**, *27*, 2466–2479.
52. Kim, K.; Lee, D.-H.; Park, S.; et al. Disordered Region of Cereblon Is Required for Efficient Degradation by Proteolysis-Targeting Chimera. *Sci. Rep.* **2019**, *9*, 19654.
53. Lee, S.; Hu, L. Nrf2 Activation through the Inhibition of Keap1-Nrf2 Protein-Protein Interaction. *Med. Chem. Res.* **2020**, *29*, 846–867.
54. Honda, T.; Rounds, B. V.; Gribble, G. W.; et al. Design and Synthesis of 2-Cyano-3,12-dioxolean-1,9-dien-28-oic Acid, a Novel and Highly Active Inhibitor of Nitric Oxide Production in Mouse Macrophages. *Bioorg. Med. Chem. Lett.* **1998**, *8*, 2711–2714.
55. Cleasby, A.; Yon, J.; Day, P. J.; et al. Structure of the BTB Domain of Keap1 and Its Interaction with the Triterpenoid Antagonist CDDO. *PLoS One.* **2014**, *9*, e98896.
56. Konopleva, M.; Tsao, T.; Ruvolo, P.; et al. Novel Triterpenoid CDDO-Me is a Potent Inducer of Apoptosis and Differentiation in Acute Myelogenous Leukemia. *Blood.* **2002**, *99*, 326–335.
57. Kim, K. B.; Lotan, R.; Yue, P.; et al. Identification of a Novel Synthetic Triterpenoid, Methyl-2-cyano-3,12-dioxoleana-1,9-dien-28-oate, That Potently Induces Caspase-Mediated Apoptosis in Human Lung Cancer Cells. *Mol. Cancer Ther.* **2002**, *1*, 177–184.
58. Hu, L.; Magesh, S.; Chen, L.; et al. Discovery of a Small-Molecule Inhibitor and Cellular Probe of Keap1-Nrf2 Protein-Protein Interaction. *Bioorg. Med. Chem. Lett.* **2013**, *23*, 3039–3043.
59. Jiang, C.-S.; Zhuang, C.-L.; Zhu, K.; et al. Identification of a Novel Small-Molecule Keap1-Nrf2 PPI Inhibitor with Cytoprotective Effects on LPS-Induced Cardiomyopathy. *J. Enzyme Inhib. Med. Chem.* **2018**, *33*, 833–841.
60. Marcotte, D.; Zeng, W.; Hus, J.-C.; et al. Small Molecules Inhibit the Interaction of Nrf2 and the Keap1 Kelch Domain through a Non-Covalent Mechanism. *Bioorg. Med. Chem.* **2013**, *21*, 4011–4019.
61. Zhuang, C.; Narayanapillai, S.; Zhang, W.; et al. Rapid Identification of Keap1-Nrf2 Small-Molecule Inhibitors through Structure-Based Virtual Screening and Hit-Based Substructure Search. *J. Med. Chem.* **2014**, *57*, 1121–1126.
62. Sun, H.-P.; Jiang, Z.-Y.; Zhang, M.-Y.; et al. Novel Protein-Protein Interaction Inhibitor of Nrf2-Keap1 Discovered by Structure-Based Virtual Screening. *Med. Chem. Commun.* **2014**, *5*, 93–98.
63. Davies, T. G.; Wixted, W. E.; Coyle, J. E.; et al. Monoacidic Inhibitors of the Kelch-like ECH-Associated Protein 1: Nuclear Factor Erythroid 2-Related Factor 2 (KEAP1:NRF2) Protein-Protein Interaction with High Cell Potency Identified by Fragment-Based Discovery. *J. Med. Chem.* **2016**, *59*, 3991–4006.
64. Heightman, T. D.; Callahan, J. F.; Chiarparin, E.; et al. Structure-Activity and Structure-Conformation Relationships of Aryl Propionic Acid Inhibitors of the Kelch-like ECH-Associated Protein 1/Nuclear Factor Erythroid 2-Related Factor 2 (KEAP1/NRF2) Protein-Protein Interaction. *J. Med. Chem.* **2019**, *62*, 4683–4702.
65. Lu, M.; Liu, T.; Jiao, Q.; et al. Discovery of a Keap1-Dependent Peptide PROTAC to Knockdown Tau by Ubiquitination-Proteasome Degradation Pathway. *Eur. J. Med. Chem.* **2018**, *146*, 251–259.
66. Tong, B.; Luo, M.; Xie, Y.; et al. Targeted Protein Degradation via a Covalent Reversible Degradation Based on Bardoxolone. *ChemRxiv.* **2020**, 12055935. <https://doi.org/10.26434/chemrxiv.12055935.v1>.
67. Baur, M.; Gneist, M.; Owa, T.; et al. Clinical Complete Long-Term Remission of a Patient with Metastatic Malignant Melanoma under Therapy with Indisulam (E7070). *Melanoma Res.* **2007**, *17*, 329–331.
68. Mita, M.; Kelly, K. R.; Mita, A.; et al. Phase I Study of E7820, an Oral Inhibitor of Integrin Alpha-2 Expression with Antiangiogenic Properties, in Patients with Advanced Malignancies. *Clin. Cancer Res.* **2011**, *17*, 193–200.
69. Bekaii-Saab, T. S.; Mortazavi, A.; Hicks, L. G.; et al. A Phase II Study of Chloroquinoxaline Sulfonamide (CQS) in Patients with Metastatic Colorectal Carcinoma (MCR). *Invest. New Drugs.* **2006**, *24*, 343–346.
70. Yokoi, A.; Kuromitsu, J.; Kawai, T.; et al. Profiling Novel Sulfonamide Antitumor Agents with Cell-Based Phenotypic Screens and Array-Based Gene Expression Analysis. *Mol. Cancer Ther.* **2002**, *1*, 275–286.
71. Uehara, T.; Minoshima, Y.; Sagane, K.; et al. Selective Degradation of Splicing Factor CAPER α by Anticancer Sulfonamides. *Nat. Chem. Biol.* **2017**, *13*, 675–680.
72. Han, T.; Goralski, M.; Gaskill, N.; et al. Anticancer Sulfonamides Target Splicing by Inducing RBM39 Degradation via Recruitment to DCAF15. *Science.* **2017**, *356*, eaal3755.
73. Bussiere, D. E.; Xie, L.; Srinivas, H.; et al. Structural Basis of Indisulam-Mediated RBM39 Recruitment to DCAF15 E3 Ligase Complex. *Nat. Chem. Biol.* **2020**, *16*, 15–23.
74. Faust, T. B.; Yoon, H.; Nowak, R. P.; et al. Structural Complementarity Facilitates E7820-Mediated Degradation of RBM39 by DCAF15. *Nat. Chem. Biol.* **2020**, *16*, 7–14.
75. Du, X.; Volkov, O. A.; Czerwinski, R. M.; et al. Structural Basis and Kinetic Pathway of RBM39 Recruitment to DCAF15 by a Sulfonamide Molecular Glue E7820. *Structure.* **2019**, *27*, 1625–1633.e3.
76. Coomar, S.; Gillingham, D. G. Exploring DCAF15 for Reprogrammable Targeted Protein Degradation. *bioRxiv.* **2019**, 542506. <https://doi.org/10.1101/542506>.
77. Zoppi, V.; Hughes, S. J.; Maniaci, C.; et al. Iterative Design and Optimization of Initially Inactive Proteolysis Targeting Chimeras (PROTACs) Identify VZ185 as a Potent, Fast, and Selective von Hippel-Lindau (VHL) Based Dual Degradation Probe of BRD9 and BRD7. *J. Med. Chem.* **2019**, *62*, 699–726.

78. Li, L.; Mi, D.; Pei, H.; et al. In Vivo Target Protein Degradation Induced by PROTACs Based on E3 Ligase DCAF15. *Signal Transduct. Target Ther.* **2020**, *5*, 129.
79. Ward, C. C.; Kleinman, J. I.; Brittain, S. M.; et al. Covalent Ligand Screening Uncovers a RNF4 E3 Ligase Recruiter for Targeted Protein Degradation Applications. *ACS Chem. Biol.* **2019**, *14*, 2430–2440.
80. Spradlin, J. N.; Hu, X.; Ward, C. C.; et al. Harnessing the Anti-Cancer Natural Product Nimbolide for Targeted Protein Degradation. *Nat. Chem. Biol.* **2019**, *15*, 747–755.
81. Zhang, X.; Crowley, V. M.; Wucherpfennig, T. G.; et al. Electrophilic PROTACs That Degrade Nuclear Proteins by Engaging DCAF16. *Nat. Chem. Biol.* **2019**, *15*, 737–746.
82. Ohoka, N.; Tsuji, G.; Shoda, T.; et al. Development of Small Molecule Chimeras That Recruit AhR E3 Ligase to Target Proteins. *ACS Chem. Biol.* **2019**, *14*, 2822–2832.
83. Bulatov, E.; Ciulli, A. Targeting Cullin-RING E3 Ubiquitin Ligases for Drug Discovery: Structure, Assembly and Small-Molecule Modulation. *Biochem. J.* **2015**, *467*, 365–386.
84. Arkin, M. R.; Tang, Y.; Wells, J. A. Small-Molecule Inhibitors of Protein-Protein Interactions: Progressing toward the Reality. *Chem Biol.* **2014**, *21*, 1102–1114.
85. Congreve, M.; Carr, R.; Murray, C.; et al. A “Rule of Three” for Fragment-Based Lead Discovery? *Drug Discov. Today.* **2003**, *8*, 876–877.
86. Lucas, X.; Van Molle, I.; Ciulli, A. Surface Probing by Fragment-Based Screening and Computational Methods Identifies Ligandable Pockets on the von Hippel-Lindau (VHL) E3 Ubiquitin Ligase. *J. Med. Chem.* **2018**, *61*, 7387–7393.
87. Chessari, G.; Buck, I. M.; Day, J. E. H.; et al. Fragment-Based Drug Discovery Targeting Inhibitor of Apoptosis Proteins: Discovery of a Non-Alanine Lead Series with Dual Activity against cIAP1 and XIAP. *J. Med. Chem.* **2015**, *58*, 6574–6588.
88. Tamanini, E.; Buck, I. M.; Chessari, G.; et al. Discovery of a Potent Nonpeptidomimetic, Small-Molecule Antagonist of Cellular Inhibitor of Apoptosis Protein 1 (cIAP1) and X-Linked Inhibitor of Apoptosis Protein (XIAP). *J. Med. Chem.* **2017**, *60*, 4611–4625.
89. Johnson, C. N.; Ahn, J. S.; Buck, I. M.; et al. A Fragment-Derived Clinical Candidate for Antagonism of X-Linked and Cellular Inhibitor of Apoptosis Proteins: 1-(6-[(4-Fluorophenyl)methyl]-5-(hydroxymethyl)-3,3-dimethyl-1 H,2 H,3 H-pyrrolo[3,2-b]pyridin-1-yl)-2-[(2 R,5 R)-5-methyl-2-[(3R)-3-methylmorpholin-4-yl]methyl]piperazin-1-yl]ethan-1-one (ASTX660). *J. Med. Chem.* **2018**, *61*, 7314–7329.
90. Resnick, E.; Bradley, A.; Gan, J.; et al. Rapid Covalent-Probe Discovery by Electrophile-Fragment Screening. *J. Am. Chem. Soc.* **2019**, *141*, 8951–8968.
91. Gehring, M.; Laufer, S. A. Emerging and Re-Emerging Warheads for Targeted Covalent Inhibitors: Applications in Medicinal Chemistry and Chemical Biology. *J. Med. Chem.* **2019**, *62*, 5673–5724.
92. Kung, W.-W.; Ramachandran, S.; Makukhin, N.; et al. Structural Insights into Substrate Recognition by the SOCS2 E3 Ubiquitin Ligase. *Nat. Commun.* **2019**, *10*, 2534.
93. Bulatov, E.; Martin, E. M.; Chatterjee, S.; et al. Biophysical Studies on Interactions and Assembly of Full-Size E3 Ubiquitin Ligase: Suppressor of Cytokine Signaling 2 (SOCS2)-Elongin BC-Cullin 5-Ring Box Protein 2 (RBX2). *J. Biol. Chem.* **2015**, *290*, 4178–4191.
94. Zadjali, F.; Pike, A. C.; Vesterlund, M.; et al. Structural Basis for c-KIT Inhibition by the Suppressor of Cytokine Signaling 6 (SOCS6) Ubiquitin Ligase. *J. Biol. Chem.* **2011**, *286*, 480–490.
95. Zhao, B.; Payne, W. G.; Sai, J.; et al. Structural Elucidation of Peptide Binding to KLHL-12, a Substrate Specific Adapter Protein in a Cul3-Ring E3 Ligase Complex. *Biochemistry.* **2020**, *59*, 964–969.
96. Rusnac, D.-V.; Lin, H.-C.; Canzani, D.; et al. Recognition of the Diglycine C-End Degron by CRL2^{KLHDC2} Ubiquitin Ligase. *Mol. Cell.* **2018**, *72*, 813–822.e4.
97. Chen, S.-J.; Wu, X.; Wadas, B.; et al. An N-End Rule Pathway That Recognizes Proline and Destroys Gluconeogenic Enzymes. *Science.* **2017**, *355*, eaal3655.
98. Dong, C.; Zhang, H.; Li, L.; et al. Molecular Basis of GID4-Mediated Recognition of Degrons for the Pro/N-End Rule Pathway. *Nat. Chem. Biol.* **2018**, *14*, 466–473.
99. Carter, A. J.; Kraemer, O.; Zwick, M.; et al. Target 2035: Probing the Human Proteome. *Drug Discov. Today.* **2019**, *24*, 2111–2115.
100. Mullard, A. A Probe for Every Protein. *Nat. Rev. Drug Discov.* **2019**, *18*, 733–736.
101. Goodnow, R. A., Jr.; Dumelin, C. E.; Keefe, A. D. DNA-Encoded Chemistry: Enabling the Deeper Sampling of Chemical Space. *Nat. Rev. Drug Discov.* **2017**, *16*, 131–147.
102. Zambaldo, C.; Dagher, J.-P.; Saarbach, J.; et al. Screening for Covalent Inhibitors Using DNA-Display of Small Molecule Libraries Functionalized with Cysteine Reactive Moieties. *Med. Chem. Commun.* **2016**, *7*, 1340–1351.
103. Chan, A. I.; McGregor, L. M.; Jain, T.; et al. Discovery of a Covalent Kinase Inhibitor from a DNA-Encoded Small-Molecule Library × Protein Library Selection. *J. Am. Chem. Soc.* **2017**, *139*, 10192–10195.
104. Smith, G. P. Phage Display: Simple Evolution in a Petri Dish (Nobel Lecture). *Angew. Chem. Int. Ed.* **2019**, *58*, 14428–14437.
105. Cardote, T. A. F.; Ciulli, A. Cyclic and Macrocyclic Peptides as Chemical Tools To Recognise Protein Surfaces and Probe Protein-Protein Interactions. *ChemMedChem.* **2016**, *11*, 787–794.
106. Dougherty, P. G.; Qian, Z.; Pei, D. Macrocycles as Protein-Protein Interaction Inhibitors. *Biochem. J.* **2017**, *474*, 1109–1125.
107. Naylor, M. R.; Bockus, A. T.; Blanco, M. J.; et al. Cyclic Peptide Natural Products Chart the Frontier of Oral Bioavailability in the Pursuit of Undruggable Targets. *Curr. Opin. Chem. Biol.* **2017**, *38*, 141–147.
108. Walport, L. J.; Obexer, R.; Suga, H. Strategies for Transitioning Macrocyclic Peptides to Cell-Permeable Drug Leads. *Curr. Opin. Biotechnol.* **2017**, *48*, 242–250.
109. Boehm, M.; Beaumont, K.; Jones, R.; et al. Discovery of Potent and Orally Bioavailable Macrocyclic Peptide-Peptoid Hybrid CXCR7 Modulators. *J. Med. Chem.* **2017**, *60*, 9653–9663.
110. Maianti, J. P.; McFedries, A.; Foda, Z. H.; et al. Anti-Diabetic Activity of Insulin-Degrading Enzyme Inhibitors Mediated by Multiple Hormones. *Nature.* **2014**, *511*, 94–98.

111. Heinis, C.; Rutherford, T.; Freund, S.; et al. Phage-Encoded Combinatorial Chemical Libraries Based on Bicyclic Peptides. *Nat. Chem. Biol.* **2009**, *5*, 502–507.
112. Zorzi, A.; Deyle, K.; Heinis, C. Cyclic Peptide Therapeutics: Past, Present and Future. *Curr. Opin. Chem. Biol.* **2017**, *38*, 24–29.
113. Huang, Y.; Wiedmann, M. M.; Suga, H. RNA Display Methods for the Discovery of Bioactive Macrocycles. *Chem. Rev.* **2019**, *119*, 10360–10391.
114. McAllister, T. E.; Yeh, T.-L.; Abboud, M. I.; et al. Non-Competitive Cyclic Peptides for Targeting Enzyme-Substrate Complexes. *Chem. Sci.* **2018**, *9*, 4569–4578.
115. Nawatha, M.; Rogers, J. M.; Bonn, S. M.; et al. De Novo Macrocyclic Peptides That Specifically Modulate Lys48-Linked Ubiquitin Chains. *Nat. Chem.* **2019**, *11*, 644–652.
116. Yamagishi, Y.; Shoji, I.; Miyagawa, S.; et al. Natural product-Like Macrocyclic N-Methyl-Peptide Inhibitors against a Ubiquitin Ligase Uncovered from a Ribosome-Expressed De Novo Library. *Chem. Biol.* **2011**, *18*, 1562–1570.
117. Owa, T.; Okauchi, T.; Yoshimatsu, K.; et al. A Focused Compound Library of Novel N-(7-Indolyl)benzenesulfonamides for the Discovery of Potent Cell Cycle Inhibitors. *Bioorg. Med. Chem. Lett.* **2000**, *10*, 1223–1226.
118. Ślabicki, M.; Kozicka, Z.; Petzold, G.; et al. The CDK Inhibitor CR8 Acts as a Molecular Glue Degradator That Depletes Cyclin K. *Nature.* **2020**, *585*, 293–297.
119. Mayor-Ruiz, C.; Bauer, S.; Brand, M.; et al. Rational Discovery of Molecular Glue Degradators via Scalable Chemical Profiling. *Nat. Chem. Biol.* **2020**, *16*, 1199–1207.
120. Brand, M.; Winter, G. E. Stick It to E3s. *Nat. Chem. Biol.* **2019**, *15*, 655–656.
121. Gadd, M. S.; Testa, A.; Lucas, X.; et al. Structural Basis of PROTAC Cooperative Recognition for Selective Protein Degradation. *Nat. Chem. Biol.* **2017**, *13*, 514–521.
122. Bondeson, D. P.; Smith, B. E.; Burslem, G. M.; et al. Lessons in PROTAC Design from Selective Degradation with a Promiscuous Warhead. *Cell Chem. Biol.* **2018**, *25*, 78–87.e5.
123. Yip, K. M.; Fischern, N.; Paknia, E.; et al. Breaking the Next Cryo-EM Resolution Barrier – Atomic Resolution Determination of Proteins! *bioRxiv.* **2020**, 106740. <https://doi.org/10.1101/2020.05.21.106740>.
124. Nakane, T.; Kotecha, A.; Sente, A.; et al. Single-Particle Cryo-EM at Atomic Resolution. *bioRxiv.* **2020**, 110189. <https://doi.org/10.1101/2020.05.22.110189>.
125. Senior, A. W.; Evans, R.; Jumper, J.; et al. Improved Protein Structure Prediction Using Potentials from Deep Learning. *Nature.* **2020**, *577*, 706–710.
126. Deshaies, R. J. Multispecific Drugs Herald a New Era of Biopharmaceutical Innovation. *Nature.* **2020**, *580*, 329–338.
127. Maniaci, C.; Ciulli, A. Bifunctional Chemical Probes Inducing Protein-Protein Interactions. *Curr. Opin. Chem. Biol.* **2019**, *52*, 145–156.
128. Whitworth, C.; Ciulli, A. New Class of Molecule Targets Proteins outside Cells for Degradation. *Nature.* **2020**, *584*, 193–194.
129. Cornella-Taracido, I.; Garcia-Echeverria, C. Monovalent Protein-Degradors – Insights and Future Perspectives. *Bioorg. Med. Chem. Lett.* **2020**, *30*, 127202.
130. Hanan, E. J.; Liang, J.; Wang, X.; et al. Monomeric Targeted Protein Degradors. *J. Med. Chem.* doi:10.1021/acs.jmedchem.0c00093. <https://pubs.acs.org/doi/10.1021/acs.jmedchem.0c00093>.
131. Takahashi, D.; Moriyama, J.; Nakamura, T.; et al. AUTACs: Cargo-Specific Degradors Using Selective Autophagy. *Mol. Cell.* **2019**, *76*, 797–810.e10.
132. Lim, S.; Khoo, R.; Peh, K. M.; et al. BioPROTACs as Versatile Modulators of Intracellular Therapeutic Targets Including Proliferating Cell Nuclear Antigen (PCNA). *Proc. Natl. Acad. Sci. U.S.A.* **2020**, *117*, 5791–5800.
133. Banik, S. M.; Pedram, K.; Wisnovsky, S.; et al. Lysosome-Targeting Chimaeras for Degradation of Extracellular Proteins. *Nature.* **2020**, *584*, 291–297.
134. Li, Z.; Wang, C.; Wang, Z.; et al. Allele-Selective Lowering of Mutant HTT Protein by HTT-LC3 Linker Compounds. *Nature.* **2019**, *575*, 203–209.
135. Yamazoe, S.; Tom, J.; Fu, Y.; et al. Heterobifunctional Molecules Induce Dephosphorylation of Kinases – A Proof of Concept Study. *J. Med. Chem.* **2020**, *63*, 2807–2813.
136. Siriwardena, S. U.; Munkanatta Godage, D. N. P.; Shoba, V. M.; et al. Phosphorylation-Inducing Chimeric Small Molecules. *J. Am. Chem. Soc.* **2020**, *142*, 14052–14057.
137. Han, X.; Zhao, L.; Xiang, W.; et al. Discovery of Highly Potent and Efficient PROTAC Degradors of Androgen Receptor (AR) by Employing Weak Binding Affinity VHL E3 Ligase Ligands. *J. Med. Chem.* **2019**, *62*, 11218–11231.
138. Farnaby, W.; Koegl, M.; Roy, M. J.; et al. BAF Complex Vulnerabilities in Cancer Demonstrated via Structure-Based PROTAC Design. *Nat. Chem. Biol.* **2019**, *15*, 672–680.
139. Qin, C.; Hu, Y.; Zhou, B.; et al. Discovery of QCA570 as an Exceptionally Potent and Efficacious Proteolysis Targeting Chimera (PROTAC) Degradator of the Bromodomain and Extra-Terminal (BET) Proteins Capable of Inducing Complete and Durable Tumor Regression. *J. Med. Chem.* **2018**, *61*, 6685–6704.
140. Rana, S.; Bendjennat, M.; Kour, S.; et al. Selective Degradation of CDK6 by a Palbociclib Based PROTAC. *Bioorg. Med. Chem. Lett.* **2019**, *29*, 1375–1379.
141. Li, Z.; Pinch, B. J.; Olson, C. M.; et al. Development and Characterization of a Wee1 Kinase Degradator. *Cell Chem. Biol.* **2020**, *27*, 57–65.e9.
142. Bensimon, A.; Pizzagalli, M. D.; Kartnig, F.; et al. Targeted Degradation of SLC Transporters Reveals Amenability of Multi-Pass Transmembrane Proteins to Ligand-Induced Proteolysis. *Cell Chem. Biol.* **2020**, *27*, 728–739.e9.
143. Crew, A. P.; Snyder, L. B.; Wang, J. Compounds and Methods for the Targeted Degradation of Androgen Receptor. U.S. Patent 10,584,101, March 10, 2020.
144. Mainolfi, N.; Ji, N.; Kluge, A. F.; et al. IRAK Degradors and Uses Thereof. World Intellectual Property Organization 2019/133531, July 4, 2019.
145. Qi, J. Acetylation Writer Inhibitor Development and Uses Thereof. World Intellectual Property Organization 2020/092907, May 7, 2019.

146. Yamazaki, R.; Ibuki, T.; Sawaguchi, Y.; et al. Amide Compound Having BET Proteolysis-Inducing Action and Medicinal Application Thereof. World Intellectual Property Organization 2020/009176, January 9, 2020.
147. Dragovich, P.; Pillow, T.; Blake, R. A.; et al. Conjugated Chemical Inducers of Degradation and Methods of Use. World Intellectual Property Organization 2020/086858, April 30, 2020.
148. Hines, J.; Lartigue, S.; Dong, H.; et al. MDM2-Recruiting PROTAC Offers Superior, Synergistic Antiproliferative Activity via Simultaneous Degradation of BRD4 and Stabilization of p53. *Cancer Res.* **2019**, *79*, 251–262.
149. Zhao, Q.; Lan, T.; Su, S.; et al. Induction of Apoptosis in MDA-MB-231 Breast Cancer Cells by a PARP1-Targeting PROTAC Small Molecule. *Chem. Commun. (Camb)*. **2019**, *55*, 369–372.
150. Zambaldo, C.; Vinogradova, E. V.; Qi, X.; et al. 2-Sulfonylpyridines as Tunable, Cysteine-Reactive Electrophiles. *J. Am. Chem. Soc.* **2020**, *142*, 8972–8979.
151. Cal, P. M. S. D.; Vicente, J. B.; Pires, E.; et al. Iminoboronates: A New Strategy for Reversible Protein Modification. *J. Am. Chem. Soc.* **2012**, *134*, 10299–10305.
152. Lin, S.; Yang, X.; Jia, S.; et al. Redox-Based Reagents for Chemoselective Methionine Bioconjugation. *Science*. **2017**, *355*, 597–602.
153. Muñoz-Escobar, J.; Matta-Camacho, E.; Cho, C.; et al. Bound Waters Mediate Binding of Diverse Substrates to a Ubiquitin Ligase. *Structure*. **2017**, *25*, 719–729.e3.
154. Ostertag, M. S.; Hutwelker, W.; Plettenburg, O.; et al. Structural Insights into BET Client Recognition of Endometrial and Prostate Cancer-Associated SPOP Mutants. *J. Mol. Biol.* **2019**, *431*, 2213–2221.
155. Ostertag, M. S.; Messias, A. C.; Sattler, M.; et al. The Structure of the SPOP-Pdx1 Interface Reveals Insights into the Phosphorylation-Dependent Binding Regulation. *Structure*. **2019**, *27*, 327–334.e3.
156. Schumacher, F.-R.; Sorrell, F. J.; Alessi, D. R.; et al. Structural and Biochemical Characterization of the KLHL3-WNK Kinase Interaction Important in Blood Pressure Regulation. *Biochem. J.* **2014**, *460*, 237–246.
157. Chen, Z.; Picaud, S.; Filippakopoulos, P.; et al. Structural Basis for Recruitment of DAPK1 to the KLHL20 E3 Ligase. *Structure*. **2019**, *27*, 1395–1404.e4.
158. Filippakopoulos, P.; Low, A.; Sharpe, T. D.; et al. Structural Basis for Par-4 Recognition by the SPRY Domain- and SOCS Box-Containing Proteins SPSB1, SPSB2, and SPSB4. *J. Mol. Biol.* **2010**, *401*, 389–402.
159. You, T.; Wang, Y.; Li, K.; et al. Crystal Structure of SPSB2 in Complex with a Rational Designed RGD-Containing Cyclic Peptide Inhibitor of SPSB2-iNOS Interaction. *Biochem. Biophys. Res. Commun.* **2017**, *489*, 346–352.
160. Sadek, M. M.; Barlow, N.; Leung, E. W. W.; et al. A Cyclic Peptide Inhibitor of the iNOS-SPSB Protein-Protein Interaction as a Potential Anti-Infective Agent. *ACS Chem. Biol.* **2018**, *13*, 2930–2938.
161. Zeng, Z.; Wang, W.; Yang, Y.; et al. Structural Basis of Selective Ubiquitination of TRF1 by SCFFbx4. *Dev. Cell.* **2010**, *18*, 214–225.
162. Li, Y.; Jin, K.; Bunker, E.; et al. Structural Basis of the Phosphorylation-Independent Recognition of Cyclin D1 by the SCF^{FBXO31} Ubiquitin Ligase. *Proc. Natl. Acad. Sci. U.S.A.* **2018**, *115*, 319–324.
163. Wu, G.; Xu, G.; Schulman, B. A.; et al. Structure of a Beta-TrCP1-Skp1-Beta-Catenin Complex: Destruction Motif Binding and Lysine Specificity of the SCF(beta-TrCP1) Ubiquitin Ligase. *Mol. Cell.* **2003**, *11*, 1445–1456.
164. Hao, B.; Oehlmann, S.; Sowa, M. E.; et al. Structure of a Fbw7-Skp1-Cyclin E Complex: Multisite-Phosphorylated Substrate Recognition by SCF Ubiquitin Ligases. *Mol. Cell.* **2007**, *26*, 131–143.
165. Yalla, K.; Elliott, C.; Day, J. P.; et al. FBXW7 Regulates DISC1 Stability via the Ubiquitin-Proteasome System. *Mol. Psychiatry*. **2018**, *23*, 1278–1286.
166. Tian, W.; Li, B.; Warrington, R.; et al. Structural Analysis of Human Cdc20 Supports Multisite Degron Recognition by APC/C. *Proc. Natl. Acad. Sci. U.S.A.* **2012**, *109*, 18419–18424.
167. Sackton, K. L.; Dimova, N.; Zeng, X.; et al. Synergistic Blockade of Mitotic Exit by Two Chemical Inhibitors of the APC/C. *Nature*. **2014**, *514*, 646–649.
168. Janz, J. M.; Sakmar, T. P.; Min, K. C. A Novel Interaction between Atrophin-Interacting Protein 4 and Beta-p21-Activated Kinase-Interactive Exchange Factor Is Mediated by an SH3 Domain. *J. Biol. Chem.* **2007**, *282*, 28893–28903.
169. Liu, Y.; Lau, J.; Li, W.; et al. Structural Basis for the Regulatory Role of the PPxY Motifs in the Thioredoxin-Interacting Protein TXNIP. *Biochem. J.* **2016**, *473*, 179–187.
170. Cappadocia, L.; Mascle, X. H.; Bourdeau, V.; et al. Structural and Functional Characterization of the Phosphorylation-Dependent Interaction between PML and SUMO1. *Structure*. **2015**, *23*, 126–138.
171. James, L. C.; Keeble, A. H.; Khan, Z.; et al. Structural Basis for PRYSPRY-Mediated Tripartite Motif (TRIM) Protein Function. *Proc. Natl. Acad. Sci. U.S.A.* **2007**, *104*, 6200–6205.
172. Tsai, W.-W.; Wang, Z.; You, T. T.; et al. TRIM24 Links a Non-Canonical Histone Signature to Breast Cancer. *Nature*. **2010**, *468*, 927–932.
173. Palmer, W. S.; Poncet-Montange, G.; Liu, G.; et al. Structure-Guided Design of IACS-9571, a Selective High-Affinity Dual TRIM24-BRPF1 Bromodomain Inhibitor. *J. Med. Chem.* **2016**, *59*, 1440–1454.
174. Liu, J.; Li, F.; Bao, H.; et al. The Polar Warhead of a TRIM24 Bromodomain Inhibitor Rearranges a Water-Mediated Interaction Network. *FEBS J.* **2017**, *284*, 1082–1095.
175. Xi, Q.; Wang, Z.; Zaromytidou, A.-I.; et al. A Poised Chromatin Platform for TGF- β Access to Master Regulators. *Cell*. **2011**, *147*, 1511–1524.