
Eastern Pacific Coral Reef Provinces, Coral Community Structure and Composition: An Overview

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Abstract

Advances in our knowledge of eastern tropical Pacific (ETP) coral reef biogeography and ecology during the past two decades are briefly reviewed. Fifteen ETP subregions are recognized, including mainland and island localities from the Gulf of California (Mexico) to Rapa Nui (Easter Island, Chile). Updated species lists reveal a mean increase of 4.2 new species records per locality or an overall increase of 19.2 % in species richness during the past decade. The largest increases occurred in tropical mainland Mexico, and in equatorial Costa Rica and Colombia, due mainly to continuing surveys of these under-studied areas. Newly discovered coral communities are also now known from the southern Nicaraguan coastline. To date 47 zooxanthellate scleractinian species have been recorded in the ETP, of which 33 also occur in the central/south Pacific, and 8 are presumed to be ETP endemics. Usually no more than 20–25 zooxanthellate coral species are present at any given locality, with the principal reef-building genera being *Pocillopora*, *Porites*, *Pavona*, and *Gardineroseris*. This compares with 62–163 species at four of the nearest central/south Pacific localities. Hydrocorals in the genus *Millepora* also occur in the ETP and are reviewed in the context of their global distributions. Coral community associates engaged in corallivory, bioerosion, and competition for space are noted for several localities. Reef framework construction in the ETP typically occurs at shallow depths (2–8 m) in sheltered habitats or at greater depths (10–30 m) in more exposed areas such as oceanic island settings with high water column light penetration. Generally, eastern Pacific reefs do not reach sea level with the development of drying reef flats, and instead experience brief periods of exposure during extreme low tides or drops in sea level during La Niña events. High rates of mortality during El Niño disturbances have occurred in many ETP equatorial areas, especially in Panama and the Galápagos Islands during the 1980s and 1990s. Remarkably, however, no loss of resident, zooxanthellate scleractinian species has occurred at these sites, and many ETP coral reefs have demonstrated significant recovery from these disturbances during the past two decades.

Keywords

Species distributions • Biogeography • Eastern Pacific • Coral occurrences • Species richness

5.1 Introduction

Geographic isolation, variable oceanographic conditions, and a peninsula-like shallow shelf spanning a relatively narrow latitudinal corridor, have all figured prominently in the formation and development of the eastern tropical Pacific coral reef region. The vast majority of eastern Pacific corals belong to species occurring in the central and south Pacific (e.g., Wells 1983; Cortés 1986; Grigg and Hey 1992; Veron 1995; Glynn and Ault 2000). Dana (1975) recognized this faunal affinity, and suggested the existence of genetic connectivity between the central Pacific Line Islands and the equatorial eastern Pacific by larval dispersal via the North Equatorial Counter Current (NECC). A few species are eastern Pacific endemics, and there is a near absence of Caribbean and western Atlantic species along western American shores. The narrowing and eventual closing of the Central American isthmus, 3.5–3.0 Ma (Coates and Obando

1996), brought about extinctions of once-shared transoceanic species, and the invasion of coral species from the west by long-distance dispersal (see Chap. 2, López-Pérez, and Chap. 16, Lessios and Baums; López-Pérez and Budd 2009).

The occurrence of reef-building corals along coastal western America is highly skewed toward the northern hemisphere, with a strong presence of corals from the northern Gulf of California, Mexico (~30°N) to just a few degrees south of the equator along coastal Ecuador (2°S). Modern zooxanthellate coral communities have not been reported on continental shores south of the Gulf of Guayaquil, Ecuador, which are under the influence of the cool, north-flowing Peru Coastal Current (Fig. 5.1). Five relatively isolated oceanic island groups, from the northern-most Revillagigedo Islands off Mexico to the Galápagos Islands lying astride the equator, also belong to the eastern Pacific coral reef region. Vibrant coral communities and incipient

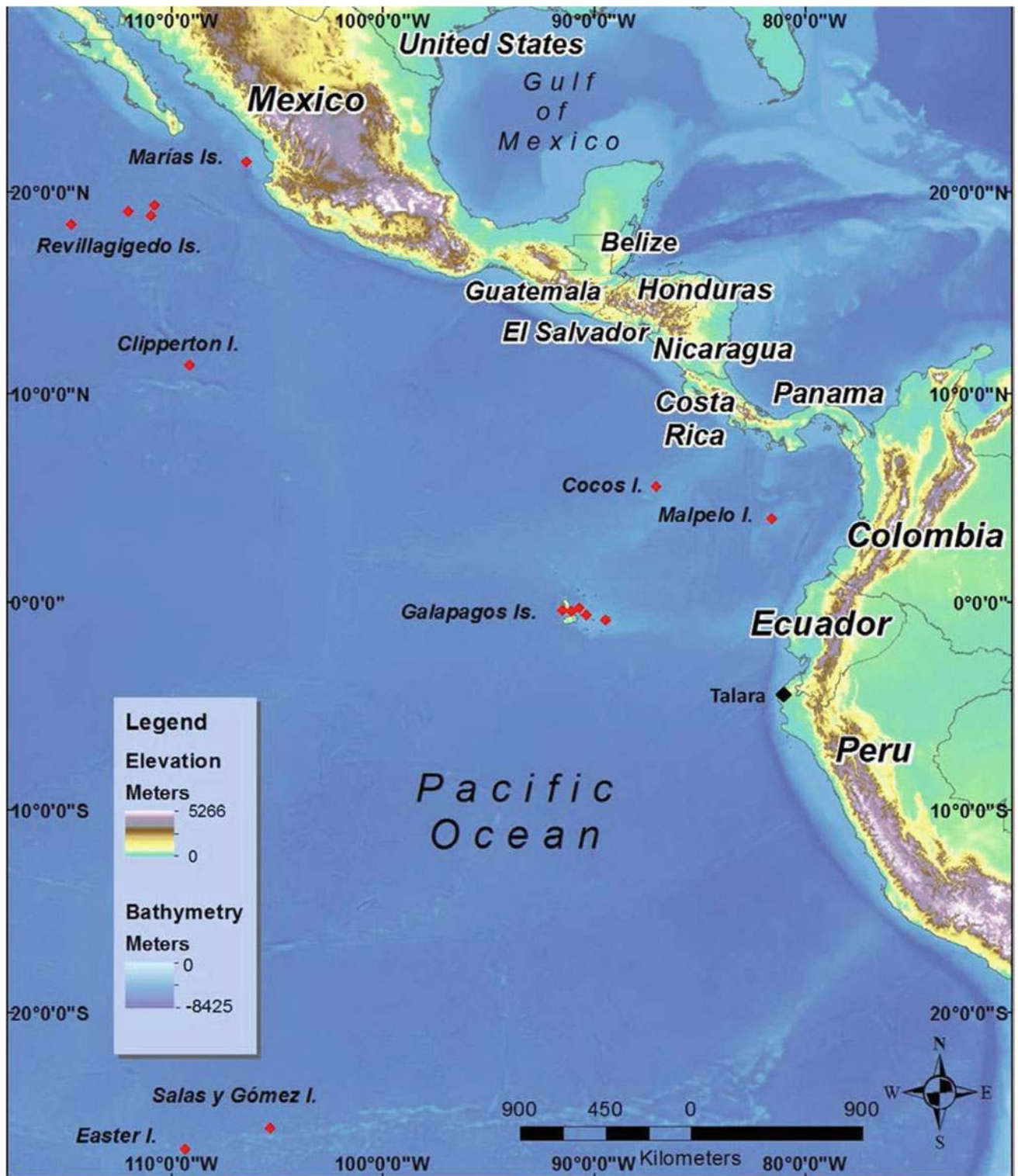


Fig. 5.1 Eastern Pacific coral and coral reef localities, Gulf of California, Mexico to Ecuador, and remote Rapa Nui (Easter) and Salas y Gómez Islands, Chile. Red diamonds identify oceanic localities supporting coral communities and coral reefs

reef development also appear far to the south and west, about 4000 km from the Galápagos Islands and coastal Ecuador. Rapa Nui (Easter) and Salas y Gómez Islands, at 27°S and

over 1000 km west of Chile, represent the most isolated ETP coral outpost (Glynn et al. 2003, 2007; Hubbard and Garcia 2003; see Chap. 6, Toth et al.).

The most recent overview of the eastern tropical Pacific (ETP) coral reef region was published over 10 years ago, in *Latin American Coral Reefs* edited by Jorge Cortés (2003). This work comprised seven chapters dedicated to the ETP, describing coral communities and coral reefs from mainland Mexico to Ecuador, including offshore islands from the Revillagigedo Islands, Mexico to Rapa Nui, Chile. These summaries included historical sketches of coral reef research, site descriptions, species lists, natural and anthropogenic disturbances, and issues pertaining to protection and management. The present chapter updates Cortés (2003), and several earlier reviews of eastern Pacific coral reef biology (e.g., Durham and Barnard 1952; Squires 1959; Durham 1966; Glynn and Wellington 1983; Cortés 1986, 1997, 2011; Glynn 2001).

An updated biogeographic analysis of zooxanthellate Scleractinia is examined first to review species relationships within the eastern Pacific region, and secondly extra-regional faunal affinities. This analysis will serve as a framework for brief introductions to the main coral reef localities from Mexico to Ecuador, including the Galápagos Islands. An overview of the more distant eastern tropical Pacific coral outposts of Rapa Nui and Salas y Gómez Islands will conclude the chapter. Eastern Pacific *Millepora* records are also reviewed, and compared to hydrocoral occurrences in the central and southeastern Pacific. Each eastern Pacific coral reef locality is examined in terms of its community structure, species richness, physical environmental conditions, and chief ecological interactions. The treatments are admittedly unbalanced, with more information provided for newly discovered or less well-known areas. It is hoped that this overview will serve to stimulate further research, and provide information for comparisons with other coral reef regions. A comprehensive review of ETP Holocene reef development—including coral framework structures, time of reef growth initiation, thicknesses, and accumulation rates—can be found in Toth et al., Chap. 6.

5.2 Eastern Pacific Coral Reef Region

The eastern tropical Pacific region consists of a distinct biotic assemblage of marine species isolated from the Caribbean Sea by the Central American land bridge. The west American marine biota is also separated from the central/western Pacific region by a wide expanse (5000–8000 km) of open ocean waters, namely the eastern Pacific Barrier or Ekman's Barrier (e.g., Darwin 1880; Ekman 1953; Briggs 1974; Grigg and Hey 1992; Veron 1995). Abundant recent evidence indicates that this barrier, which Darwin considered impassable, is best viewed as a filter bridge since numerous shallow-living species from several major taxa—

scleractinian corals (Glynn and Ault 2000), bryozoans (Banta 1991), crustaceans (Garth 1991), molluscs (Scheltema 1988; Emerson 1991), echinoderms (Lessios et al. 1998), and fishes (Robertson and Allen 1996; Robertson et al. 2004; Lessios and Robertson 2006; see Chap. 16, Lessios and Baums)—have crossed this deep water stretch by larval dispersal or rafting. In particular, many marine species shared between the eastern Pacific and central/western Pacific are common members of coral communities and coral reefs. Indeed, the majority of ETP reef-building coral taxa—species in the genera *Pocillopora*, *Porites*, *Pavona*, *Gardineroseris*—are conspecific with those found west of the ETP filter bridge.

Within the eastern Pacific region, four subtropical/tropical biogeographic provinces were early recognized: (1) Cortez Province (middle to upper Gulf of California), (2) Mexican Province (lower Gulf of California to the Gulf of Tehuantepec), (3) Panamanian Province (Tangola-Tangola Bay, Gulf of Tehuantepec to the Gulf of Guayaquil), and (4) the Galápagos Islands (Briggs 1974). More recent analyses of coral distributions have recognized an eastern Pacific oceanic islands fauna, with coral affinities shared among Clipperton Atoll, Malpelo Island, Easter Island, and Tabuaeran (=Fanning Atoll) in the Line Islands group (Glynn and Ault 2000; Glynn et al. 2007). Another recent biogeographic analysis, based on the distributions of 1135 resident shore fishes, also recognized the oceanic islands as a distinct faunal subdivision, but concluded that the continental coast is best represented by two provinces, namely (1) the Cortez, comprising the Gulf of California and lower Pacific Baja California areas, and (2) the Panamic, southward to the Gulf of Guayaquil, Ecuador (Robertson and Cramer 2009).

With an increasing knowledge of zooxanthellate scleractinian distributions, particularly during the last two decades, several workers have examined patterns of coral faunal distributions along the eastern tropical Pacific (Glynn 1997; Glynn and Ault 2000; Glynn et al. 2007; Alvarado et al. 2011), and within specific eastern Pacific countries and locations (Reyes-Bonilla 1993a; chapters in Cortés 2003; Reyes-Bonilla and López-Pérez 1998, 2009). The existence of a dispersal barrier within the eastern Pacific, the long-recognized Pacific Central American faunal gap (PCAFG), has been supported mainly by the absence of corals (Glynn and Ault 2000) and reef-associated fishes (Springer 1958; Briggs 1974) between Tangola Tangola (southern Mexico) south to the Gulf of Fonseca (El Salvador/Honduras/Nicaragua). However, the faunal similarity of newly discovered coral communities and reef-associated fishes in Nicaragua and El Salvador with central Mexican coastal areas (Michoacán, Colima) is very high, suggesting some degree of genetic connectivity across the PCAFG (Robertson and Cramer 2009; Alvarado et al. 2011).

5.2.1 Zooxanthellate Scleractinian Fauna

Here we present an updated biogeographic analysis of zooxanthellate scleractinian coral faunas from 15 ETP localities and four central/south Pacific localities. In the present data-base, the listed ETP fauna consists of 47 species in 11 genera, and the central/south Pacific localities 225 species in 40 genera (Appendix). The central/south Pacific localities include the Hawaiian Islands, Johnston Atoll, Pitcairn Islands, and Palmyra Atoll, and will be employed for analysis of and comparison with the ETP fauna. The species in this analysis were delineated chiefly from gross morphometric characteristics; the sources of this information are listed in the caption to Fig. 5.7. In light of current molecular genetic studies (Flot et al. 2010; Pinzón and LaJeunesse 2011; Pinzón et al. 2013; Boulay et al. 2014; see Chap. 14, Pinzón), and evidence from a combination of genetic and morphological analyses—including both macro- and micromorphometric characteristics (Schmidt-Roach et al. 2014)—it is evident that species delineations across the Indo-Pacific region are still unresolved. On the one hand, genetic studies offer evidence of fewer species of *Pocillopora* in the ETP than recognized morphometrically, three instead of seven species (Pinzón et al. 2013; see Chap. 14, Pinzón). On the other hand, new previously unrecognized cryptic species have come to light in the *Pocillopora damicornis* complex on the Great Barrier Reef (Schmidt-Roach et al. 2014).

A major three-way grouping of coral faunas is evident from an agglomerative hierarchical cluster analysis: (a) dominantly equatorial eastern Pacific (EEP) localities (PAN-GLN), (b) ETP oceanic islands (MAL-REV), and (c) central/southern Pacific localities (Fig. 5.2). [Locality acronyms, e.g. PAN = Panama, follow those employed in Fig. 5.2.] A notable east to west increase in species richness is apparent with eastern Pacific localities demonstrating impoverished faunas compared with potential source areas to the west of Ekman's filter bridge, a long recognized pattern (e.g., Stehli and Wells 1971; Veron 1995). All ETP reef communities consist of fewer than 30 species, whereas central/south Pacific localities exhibit from 44 to 163 species. At Palmyra Atoll, which is in the path of the North Equatorial Counter Current (Maragos et al. 2008a, b), 163 coral species have been recorded. The North Equatorial Counter Current has been considered a likely means of transport across the filter bridge (e.g., Dana 1975; Glynn and Ault 2000), and species-rich Palmyra is a potential source locality. In the EEP cluster, the low species occurrences in the southern Galápagos Islands (GLS) and Gulf of California (GOC) are likely due, in large part, to their subtropical locations. Two species-poor mainland sites (NIC, ELS) cluster with the ETP oceanic islands. The recently-discovered coral communities

in Nicaragua may reveal additional species upon further study; El Salvador, however, is a marginal coral area with highly turbid waters and is not expected to show a significant increase in species richness. Additional comments on ETP faunal relationships are noted below, in site-specific sections.

Eight species in this updated inventory are likely ETP endemics, suggesting a moderate level of speciation in this region (Table 5.1). This number represents 19.5 % of the total ETP fauna reported to date. Latitudinally, all eight coral endemics are concentrated in Mexico, in the northern-most eastern Pacific region. Three *Porites* endemics, *P. arnaudi*, *P. sverdrupi*, and *P. baueri*, demonstrate restricted distributions, known from only one or two localities. All eastern Pacific endemic species are absent from Rapa Nui (Easter Island). Three endemics, *Pavona gigantea*, *Pocillopora effusus*, and *Porites panamensis*, are widely distributed throughout the eastern Pacific, presently reported from eight to 14 localities.

5.2.2 *Millepora*

Zooxanthellate hydrocorals in the genus *Millepora* can be important contributors to reef frameworks and coral reef growth. *Millepora* has a very restricted distribution in the ETP. Three species of *Millepora* were present on reefs in the Gulf of Chiriquí (Panama) before the 1982–83 El Niño event: *Millepora intricata* Milne-Edwards 1857, *Millepora boschmai* Weerdt and Glynn 1991, and *Millepora platyphylla* Hemprich and Ehrenberg 1834. All three species bleached and experienced high mortality, during and immediately following the period of elevated sea temperatures (Glynn and de Weerdt 1991). Living colonies of *M. platyphylla* could no longer be found in Panama after 1983 (as of 2015), and are now apparently absent from the ETP (Glynn 2011). *Millepora boschmai* also disappeared from survey sites at Uva Island after 1983, but a small population was found nearby in 1992 (Glynn and Feingold 1992). All colonies from this remnant population disappeared in early 2000, as well as a few known colonies at Coiba Island (Brenes et al. 1993), and none have been observed in the eastern Pacific since that time (Maté 2003). This species was recognized in collections made from two localities in Indonesia by Razak and Hoeksema (2003), including South Sulawesi in 1994 and Sumba in 1984. In Panama, *Millepora intricata* re-appeared a few years after 1983, ostensibly by sexual recruitment from deep refuge populations (Smith et al. 2014; see Chap. 17, Smith et al.); it is again an important member of coral communities in the Gulf of Chiriquí, Panama (Fig. 5.3). Unlike *M. platyphylla* and *M. boschmai*, with populations restricted to shallow habitats (usually ≤ 6 –8 m depth), *M. intricata* occurs in deep

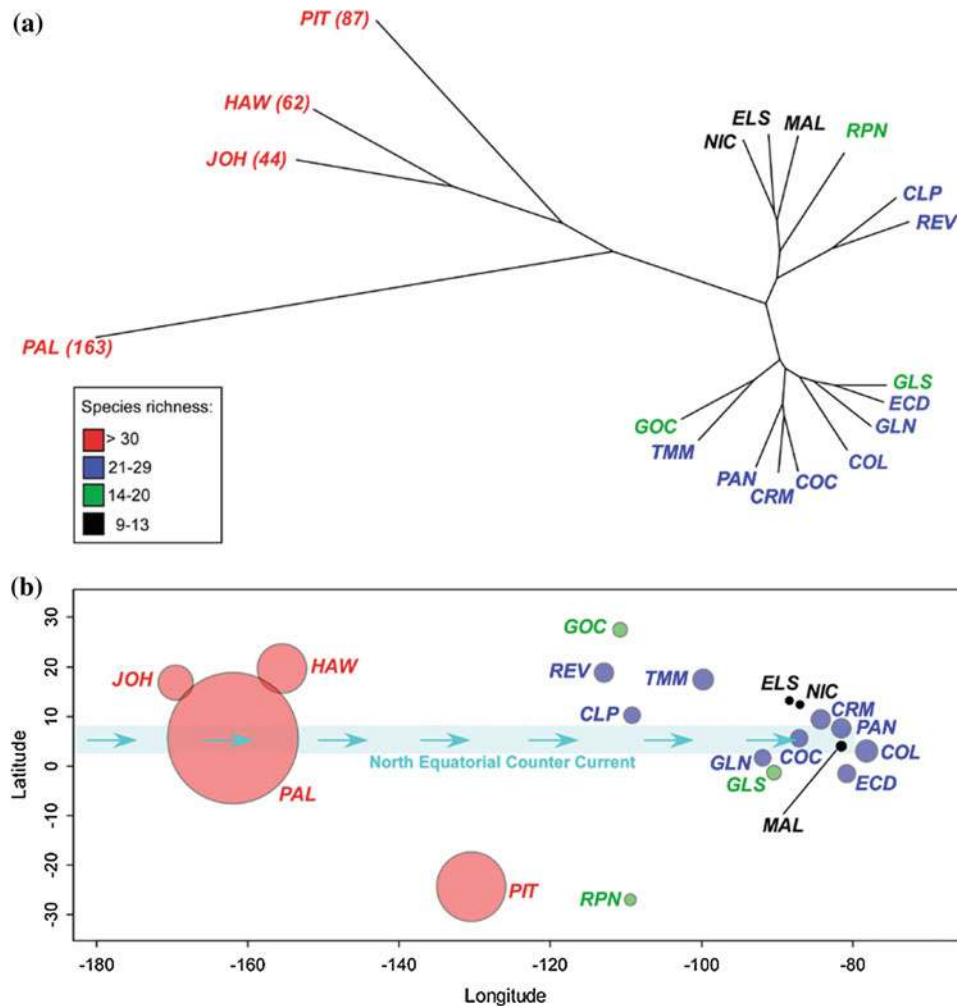


Fig. 5.2 **a** Relationships of zooxanthellate scleractinian coral faunas at 15 eastern Pacific and four central and south Pacific localities. “Spatial patterns” of an agglomerative, hierarchical cluster analysis were portrayed using a Euclidean distance measure and group-average method for cluster formation. Constructed from presence/absence data in Appendix. **b** Geographic pattern of coral species richness across EP filter bridge in relation to NECC eastward flow. Circle areas are proportional to species richness. GOC, Gulf of California; TMM,

tropical Mexican mainland; REV, Revillagigedo Islands; CLP, Clipperton Atoll; ELS, El Salvador; NIC, Nicaragua; CRM, Costa Rican mainland; COC, Cocos Island; PAN, Panama; COL, Colombia; MAL, Malpelo Island; ECD, Ecuador; GLS, southern Galápagos Islands; RPN, Rapa Nui (Easter) and Salas y Gómez Islands; HAW, Hawaiian Islands; JOH, Johnston Island; PIT, Pitcairn (Henderson, Oeno, Ducie Islands); PAL Palmyra Atoll

(>11 m depth) as well as shallow reef zones. This allowed *M. intricata* in deep-water habitats to survive two of the strongest El Niño events on record (1982–83, 1997–98), and then to re-populate shallow reef sites (Smith et al. 2014).

Millepora exaesa is known only from Clipperton Atoll in the eastern Pacific, and colonies are uncommon there (Glynn et al. 1996a, b; Carricart-Ganivet and Reyes-Bonilla 1999; Flot and Adjeroud 2009; Glynn, unpublished observation). The nearest reported central Pacific record of *M. exaesa* is at Wake Island (167°E), over 5500 km west of Clipperton (Table 5.2).

Early surveys showed that *Millepora* spp. are widespread throughout the Indo-Pacific region, and this has been confirmed by recent surveys. However, their relatively

infrequent reproduction and short-lived dispersive phase (gonochoric medusae) appear inconsistent with their widespread global occurrence (Lewis 1989; see also Chap. 15, Glynn et al.). Veron (2000) shows *Millepora* present in all parts of the Indian Ocean and the western and southern Pacific (Fig. 5.4). Some early and more recent surveys show that various *Millepora* species also occur widely across the central Pacific region, e.g. Johnston Atoll (Maragos and Jokiel 1986), the Phoenix Islands (Maragos et al. 2008a, b), Tabuaeran (formerly Fanning, Maragos 1974) and other Line Islands (Maragos et al. 2008a, b); Marquesas (Chevalier 1978), Tuamotus (Ricard 1985; Harmelin-Vivien 1985; Montaggioni 1985), Society Islands (Galzin and Pointier 1985), and the Pitcairn Islands (Irving and Dawson 2012;

Table 5.1 Regional occurrence of eastern tropical Pacific endemic zooxanthellate Scleractinia at 14 localities

Species	Localities	
	Present (no. localities)	Absent
<i>Pavona gigantea</i>	MEX EEP (14)	RPN
<i>Pocillopora effusus</i>	MEX EEP (8)	RPN
<i>Pocillopora inflata</i>	MEX EEP (6)	RPN, CLP
<i>Porites panamensis</i>	MEX EEP (9)	RPN, CLP
<i>Porites arnaudi</i>	MEX CLP (2)	RPN, EEP
<i>Porites sverdrupi</i>	MEX (2)	EEP, RPN, CLP
<i>Porites baueri</i>	MEX (1)	EEP, RPN, CLP
<i>Psammocora brighami</i>	MEX (2) EEP (5)	RPN

MEX (Mexico)—Gulf of California, Revillagigedo Is., tropical Mexican mainland; EEP (equatorial eastern Pacific)—El Salvador, Nicaragua, Costa Rica, Cocos Is, Panama, Colombia, Malpelo, Ecuador, Galápagos Iss; RPN—Easter Is; CLP—Clipperton Atoll

USNM collection records) (Fig. 5.4; Table 5.2). Subfossil fragments of *Millepora intricata* recovered from reef cores revealed radiocarbon dates of ~5000 cal year BP, indicating the presence of this hydrocoral species in Panama for much of the Holocene Epoch (Smith et al. 2014). The highly restricted occurrence of *Millepora* in the ETP is mirrored in the eastern Atlantic where it is known from only two island groups, the Canary and Cape Verde Islands (Clemente et al. 2011). It is hypothesized to be a recent introduction to the eastern Atlantic region (Clemente et al. 2011).

5.3 Intra-regional Précis

Coral faunas and coral communities within the eastern tropical Pacific region are reviewed below, from northern Mexico to the southern-most mainland assemblages of Ecuador. The eastern Pacific oceanic islands—Revillagigedo Islands, Clipperton Atoll, Isla del Coco, Galápagos Islands, and Rapa Nui—are also considered along a N–S latitudinal



Fig. 5.3 *Millepora intricata* prominence in a coral community at Brincanco Island, Gulf of Chiriquí, Panama (5.4 m depth, 4 April 2012, courtesy Irving Bethencourt). *Pocillopora* sp. in foreground ~30 cm in diameter

Table 5.2 Documented collection records of *Millepora* species in the central and south eastern Pacific Ocean

Location	Longitude	Taxa	Authority
Wake Is.	167°E	<i>Millepora</i> 2 spp.	Lobel and Lobel (2008)
		<i>Millepora tuberosa (exaesa)</i> ^a	Maragos, unpublished record, 1979–2005
Phoenix Iss.	172°E	<i>Millepora</i> sp.	Maragos (1974)
Johnson Atoll	169°E	<i>Millepora dichotoma</i>	Maragos et al. (2008a, b)
		<i>Millepora tenera</i>	Maragos and Jokiel (1986)
Cook Iss.	160–162° W	<i>Millepora</i>	Maragos (1974)
Line Iss.	157–169° W	<i>Millepora</i>	Maragos et al. (2008a, b)
Kingman reef	162°W	<i>Millepora platyphylla</i>	Maragos, personal communication
Tabuaeran	159°W	<i>Millepora platyphylla</i>	Maragos (1973)
Moorea Is.	150°W	<i>Millepora platyphylla</i>	Galzin and Pointier (1985)
Tuamotus Iss.	135–150° W		
Makatea Is.	148°W	<i>Millepora</i> sp.	Montaggioni (1985)
Mataiva Atoll	148°W	<i>Millepora platyphylla</i>	Delasalle (1985)
Tikahau Atoll	148°W	<i>Millepora platyphylla</i>	Harmelin-Viven (1985)
Rangiroa Atoll	147–148° W	<i>Millepora platyphylla</i>	Riccard (1985)
Takapoto Atoll	145°W	<i>Millepora platyphylla</i>	Salvat and Richard (1985)
Marquesas Iss.	138–141° W	<i>Millepora platyphylla</i>	Chevalier (1978)
Pitcairn Iss.	124–131° W	<i>Millepora</i> sp.	Irving and Dawson (2012)
Pitcairn Is.	130°W	<i>Millepora platyphylla</i>	D. Devaney USNM No. 79568
Henderson Is.	128°W	<i>Millepora platyphylla</i>	G. Paulay USNM No. 91571, 91572

^a*Millepora tuberosa* is a junior synonym of *M. exaesa* (see Razak and Hoeksema 2003)

gradient. This overview will hopefully provide basic coral community-level information to help evaluate intra-regional similarities and differences. We recognize that these brief intra- and inter-regional treatments would reveal only approximate comparisons at best due to the often great differences in available habitat area and habitat diversity. Regional Holocene reef development, including the coral taxa that construct frameworks, times of reef initiation, framework thickness, and accumulation rates are reviewed by Toth et al. in Chap. 6.

5.3.1 Gulf of California (Mexico)

The northern-most limit of zooxanthellate corals is in the upper reaches of the 1000 km-long Gulf of California (Fig. 5.5). Corals occur commonly over most of the length of the gulf, with increasing species richness in the southerly warmer waters. The higher thermal conditions in the southern gulf are exemplified by satellite imagery of sea surface temperatures (SSTs) in January 2006, with upper gulf SSTs of 14.5–18.3 °C and lower gulf SSTs of 20.1–22.8 °C (Ledesma-Vázquez et al. 2009). Corals are virtually absent from the western shores of the Baja California peninsula due to the southerly-flowing, cool California Current (see Chap. 3, Fiedler and Lavín). Distributional analyses based on presence/absence data have revealed two coral faunas within the Gulf of California, one in the northern and one in the southern gulf (Reyes-Bonilla and López-Pérez 2009). The northern fauna ranges from Punta Peñasco (31.3°N) to Punta Prieta (27°N), and consists predominantly of encrusting, monospecific *Porites panamensis* populations. Occasional colonies of *Porites sverdrupi* are also present on rocky substrates, and *Cycloseris curvata*, the latter in deeper rhodolith communities. The southern fauna ranges from Loreto to Cabo San Lucas. This fauna is more diverse, consisting of 20 species with a close affinity with tropical mainland Mexico (Fig. 5.2). According to recent assessments, the gulf coral fauna has demonstrated an increase from 16 to 20 species (e.g., Reyes-Bonilla et al. 2005, 2010; Reyes-Bonilla and López-Pérez 2009) (Fig. 5.6). Sixteen of these species belong to the Indo-Pacific fauna, three to the eastern Pacific fauna, and one (*P. sverdrupi*) is endemic to the gulf and mainland Mexico.

The coral assemblages and associated biota in the gulf were classified into four biotopes by Reyes-Bonilla and López-Pérez (2009): (1) isolated colonies or patches, (2) corals in rhodolith communities and other soft bottom habitats, (3) coral communities, and (4) coral reefs. Biotopes 1 and 2 occur in the northern gulf, and coral communities and coral reefs in the southern gulf. Corals, predominantly *Porites panamensis*, are highly dispersed in type 1 assemblages, usually amounting to <1 % cover where present. Even at low abundance, however, Halfar et al. (2005) demonstrated a potentially high carbonate sediment production rate for *P. panamensis* at Bahía de Los Ángeles (Fig. 5.5). Only a single rhodolith community, with associated fungiid corals, was reported in the northern gulf, while five were reported in the southern gulf. Three of these communities occurred at 25–30 m depth between La Paz and Monserrat Island near Loreto. *Diaseris distorta* is predominant with the occasional presence of *Cycloseris curvata*

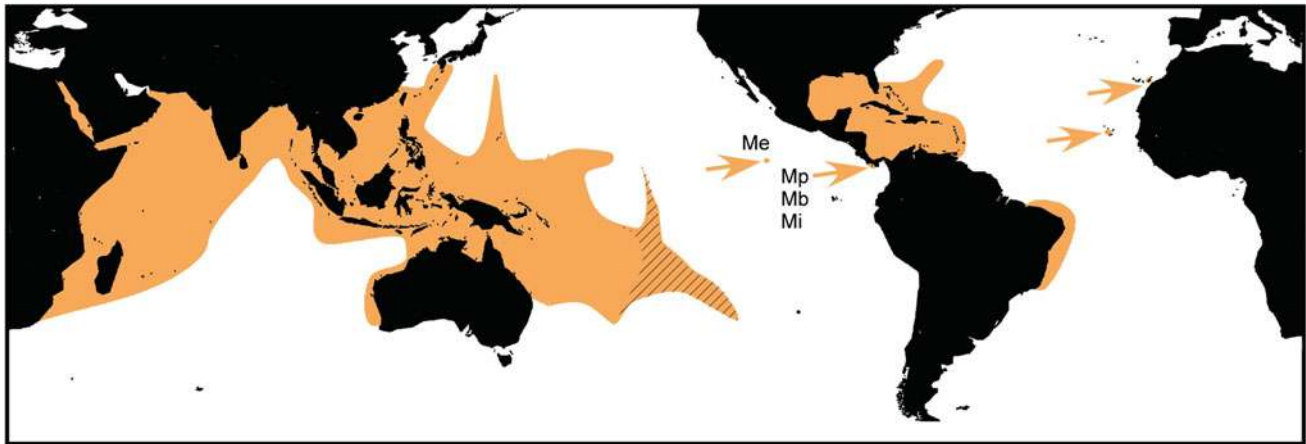


Fig. 5.4 Known global distribution of *Millepora* spp. as of 2014. In the eastern Pacific: Me, *M. exaesa*; Mp, *M. platyphylla*; Mb, *M. boschmai*; Mi, *M. intricata*. Mp and Mb now regionally extinct.

Eastern Atlantic, Canary and Cape Verde Islands, *Millepora* sp., possibly *M. alcicornis*. Cross hatching in south-central Pacific denotes records of *Millepora* occurrences in Line Islands and Pitcairn Islands

and mobile species of *Psammodora*, *Porites*, and rarely *Pavona* (Table 5.3). Present taxonomic evidence indicates that the coralline flora of rhodolith communities consists of four species in four genera (Steller et al. 2009). This biotope provides shelter and trophic resources for a diverse assemblage of non-coral invertebrates (bryozoans, molluscs, crustaceans, and echinoderms), fishes, and cryptic biota (Steller et al. 2003, 2009).

Coral community biotopes occur commonly at shallow depths (≤ 10 m) along the southwestern peninsular gulf coast. The predominant coral genera in these communities are *Pocillopora* and *Pavona*. While buildups can attain several meters in thickness (height), framework structures per se are not evident. Particularly robust coral communities, e.g. at Cabo Pulmo and La Paz, demonstrated mean coral cover values of 15–20 % with 4–6 species present. Both *Pocillopora damicornis* and *Pocillopora verrucosa* contribute substantially to these communities. The structural complexity of these coral communities offers numerous shelter sites that greatly increase the diversity of macroinvertebrates, fishes, and cryptic biota. In addition, metazoan coral symbionts and diverse coral bioeroding organisms inhabit these communities. Gastropod, echinoderm, and fish corallivores are ubiquitous in the gulf and actively prey on all coral species (e.g., Reyes-Bonilla 2003; Hendrickx et al. 2005; Álvarez-Filip et al. 2006; Reyes-Bonilla and López-Pérez 2009). *Acanthaster planci* is widespread, especially in the lower half of the gulf where corals are in high abundance (Dana and Wolfson 1970; Barham et al. 1973). Reyes-Bonilla (1993a, b, 2003) reported relatively high population densities of *A. planci* at Cabo Pulmo, 16 to >40 ind ha⁻¹ over a 13-year period, but with no indication of local coral depletion.

True coral reefs, i.e. those with wave-resistant framework structures, were reported by Reyes-Bonilla and López-Pérez

(2009) to be present at five sites in the extreme southern sector of the Baja California peninsula. These reefs are built predominantly by *Pocillopora* spp., with coral frameworks reaching 2 m in height. The largest and most thoroughly studied reef is located at Cabo Pulmo (Brusca and Thomson 1975; Reyes-Bonilla et al. 1997a, b). It is often cited as the northern-most eastern Pacific coral reef. It covers 150 ha and contributes to its surroundings an estimated 13,500–31,000 tons of CaCO₃ annually (Reyes-Bonilla and López-Pérez 2009). This reef has formed on intrusive dikes and rocky hardgrounds. Even though the carbonate production rate of this reef is high, the coarser, reef-derived sediments are transported off-reef, especially during the passage of tropical cyclones, with no appreciable sediment retention. This loss greatly limits vertical reef accretion. Riegl et al. (2007) concluded that the Cabo Pulmo formation, sedimentologically, functions only in a limited way like a coral reef.

Compared to natural biotic effects, such as predation and bioerosion, extreme physical conditions have demonstrated the greatest negative influence on coral communities in the Gulf of California. Hurricanes and tropical storms regularly frequent the southern-most sector of the gulf (see Chap. 3, Fiedler and Lavín), and can limit reef development as noted above at Cabo Pulmo. Storm-generated turbulence and sediment transport are responsible for local coral breakage and the burial of fungiid populations (Reyes-Bonilla and López-Pérez 2009). During the past few decades, ENSO-related high and low seawater temperature extremes have caused significant coral bleaching and mortality in the lower gulf (Reyes-Bonilla 2001; Iglesias-Prieto et al. 2004; Paz-García et al. 2012). These thermal-bleaching events can lead to widespread and spatially uniform mortality of *Pocillopora* spp., approaching 20 %, over 10s–100s of km of coastline. While noteworthy, this level of mortality is significantly lower than that reported in other ETP localities. Reyes-Bonilla (2001) hypothesized

Fig. 5.5 Gulf of California (GOC) and Revillagigedo Islands (REV), Mexico. Red diamonds identify surveyed sites



that the relatively low level of thermal bleaching/mortality is possibly a result of local upwelling during the summer season, and proximity of lower gulf waters to the California Current system that contributes to the formation of a cool front at the entrance to the gulf.

5.3.2 Tropical Mainland Mexico

Beyond the Gulf of California, the tropical mainland coast of Mexico extends toward the southeast from southern Sinaloa to Chiapas states, a distance of nearly 2000 km (Fig. 5.7). Only two poritid species have been reported from Sinaloa (*Porites*

panamensis, *Porites sverdrupi*) and, excluding extensive Miocene-Pliocene fossil reefs, no zooxanthellate corals are known from Chiapas. Both of these regions are dominated by coastal lagoons, sandy beaches, estuaries and mangrove ecosystems (Flores-Verdugo et al. 2001). The greatest reef development occurs in Nayarit and Oaxaca, each with 20 and 16 coral species, respectively (Reyes-Bonilla 2003; Reyes-Bonilla et al. 2005). Both of these areas are near upwelling centers; the northern center is located at Banderas Bay, Nayarit (López-Sandoval et al. 2009), and the southern upwelling area occurs along the Huatulco coast, Oaxaca (Steenburgh et al. 1998), which borders the Gulf of Tehuantepec (see Chap. 3, Fiedler and Lavín). Pocilloporid fringing

Table 5.3 Occurrence of zooxanthellate scleractinian corals in three Pacific Mexican regions

Species	Location		
	Gulf of California	Tropical Mainland Mexico	Revillagigedo Iss
<i>Cycloseris curvata</i> (Hoeksema, 1989)	R	R	R
<i>Fungia vaughani</i> Boschma, 1923	R		
<i>Diaseris distorta</i> (Michelin, 1843)	R	R	R
<i>Gardineroseris planulata</i> (Dana, 1846)		R	
<i>Leptoseris papyracea</i> (Dana, 1846)	R	R	
<i>Pavona clavus</i> (Dana, 1846)	C	C	C
<i>Pavona duerdeni</i> Vaughan, 1907	R		R
<i>Pavona gigantea</i> Verrill, 1869	A	C	C
<i>Pavona maldivensis</i> (Gardiner, 1905)			R
<i>Pavona minuta</i> Wells, 1954			R
<i>Pavona varians</i> Verrill, 1864	R	R	R
<i>Pocillopora damicornis</i> (Linnaeus, 1758)	C	A	R
<i>Pocillopora effusus</i> Veron, 2002		R	
<i>Pocillopora elegans</i> Dana, 1846	C	A	C
<i>Pocillopora eydouxi</i> Milne Edwards and Haime, 1860	R	R	C
<i>Pocillopora inflata</i> Glynn, 1999	R	R	
<i>Pocillopora meandrina</i> Dana, 1846	A	C	C
<i>Pocillopora capitata</i> Verrill, 1864	A	C	R
<i>Pocillopora verrucosa</i> (Ellis and Solander, 1786)	A	A	A
<i>Pocillopora woodjonesi</i> Vaughan, 1918			R
<i>Porites arnaudi</i> Reyes-Bonilla and Carricart-Ganivet, 2000			R
<i>Porites australiensis</i> Vaughan, 1918			R
<i>Porites baueri</i> Squires, 1959		R	
<i>Porites evermanni</i> Vaughan, 1907		R	
<i>Porites lichen</i> Dana, 1846			C
<i>Porites lobata</i> Dana, 1846		R	A
<i>Porites lutea</i> Milne Edwards and Haime, 1851			R
<i>Porites panamensis</i> Verrill, 1866	A	R	R
<i>Porites sverdrupi</i> Durham, 1947	R		
<i>Psammocora brighami</i> (Vaughan, 1907)	R		
<i>Psammocora haimeana</i> Milne Edwards and Haime, 1851	R		
<i>Psammocora profundacella</i> Gardiner, 1898		R	R
<i>Psammocora stellata</i> Verrill, 1864	C	C	C
<i>Psammocora superficialis</i> Gardiner, 1898		R	R
Total number species	20	22	24

A abundant (>50 % presence at surveyed sites); C common (20–50 % presence); R rare (<20 % presence). Records are from Reyes-Bonilla et al. (2010). Species names in this and following tables from Wells (1983), Reyes-Bonilla et al. (2005) and Veron (2000)

reefs occur in both of these areas, usually at sheltered sites; reef frameworks attain vertical heights of up to 6 m, live coral cover commonly ranges between 20 and 50 %, and extends over several ha at several sites (Carriquiry and Reyes-Bonilla 1997; Reyes-Bonilla 2003). Coral communities and small patch reefs are also present in other west Mexican areas,

particularly along coastal Guerrero and Michoacan states. Recent exploration has dramatically increased the species richness of zooxanthellate corals over a relatively brief period, from 19 (Reyes-Bonilla 2003) to 26 (Reyes-Bonilla et al. 2005, 2010; Glynn et al. 2007; Boulay et al. 2014), due mainly to continued exploration (Fig. 5.6; Appendix).

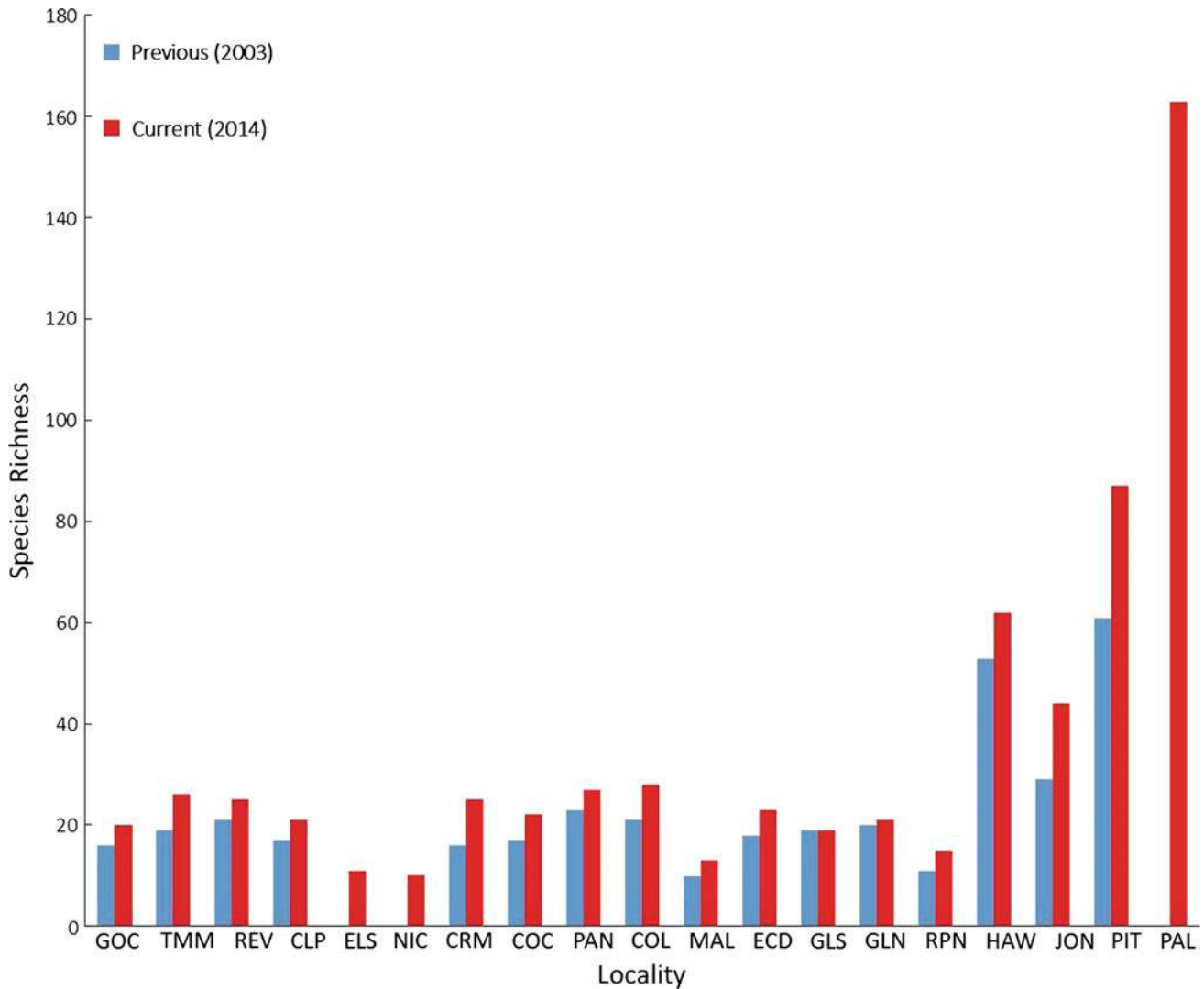


Fig. 5.6 Differences in zooxanthellate scleractinian coral species richness at eastern and central Pacific localities before (blue) and after (red) major surveys. Most 'previous' records were published in 2003 (Cortés 2003), but some are from major works as early as the latter half of the 1980s. Single bars (ELS, NIC, PAL) denote a lack of data from before the time periods examined. *Sources Previous/Current* GOC 1/2; TMM 1/2,3; REV 1/2; CLP 4/3,4; ELS -/5; NIC -/6; CRM 7,8,9/3,10; COC 8/3,10; PAN 11, 12/3,12; COL 13/14; MAL 10,13/12,14; ECD 15/16; GLS 15/17; GLN 15/3,4,18; RPN 19/4,20; HAW 21/22,23; JOH 24/25,26; PIT 27/28; PAL -/29. (1) Reyes-Bonilla (2003); (2) Reyes-Bonilla et al. (2010); (3) Boulay et al. (2013); (4) Glynn et al. (2007); (5) Reyes-Bonilla and Barraza (2003);

(6) Alvarado et al. (2010), (2011); (7) Cortés and Guzmán (1998); (8) Cortés and Jiménez (2003); (9) Jiménez (2007a, b); (10) Cortés (updated checklist); (11) Holst and Guzmán (1993), Maté (2003); (12) Maté (updated checklist); (13) Zapata and Vargas-Ángel (2003); (14) Zapata (updated checklist); (15) Glynn (2003); (16) Rivera and Martínez (2011); (17) Glynn (updated checklist); (18) Hickman (2008); (19) Glynn et al. (2003); (20) Wieters and Navarrete (updated checklist); (21) Maragos (1995); (22) Maragos (updated checklist); (23) Maragos et al. (2004); (24) Maragos and Jokiel (1986); (25) Lobel and Lobel (2008); (26) Maragos (updated checklist); (27) Paulay (1989); (28) Irving and Dawson (2012); (29) Williams et al. (2008), Maragos (updated checklist)

The coral faunas of tropical mainland Mexico (TMM) and the Gulf of California (GOC) are very similar (Fig. 5.2a), however, mainland Mexico is more speciose (26 vs. 20 species), with several reef-building species (in the genera *Pocillopora*, *Porites*, *Gardineroseris*) that have not been reported from the Gulf.

Information on coral community development at the Marías Islands, Nayarit, was lacking until relatively recently, with the delayed publication of field studies conducted in

1997 (Pérez-Vivar et al. 2006). All 10 species of zooxanthellate corals collected there are known from other Mexican mainland areas. *Porites baueri*, long considered an endemic species restricted to the Marías Islands and elsewhere on the Nayarit coast (Reyes-Bonilla 1993b), was not found. This species may be synonymous with *Porites lobata*, which was abundant in the islands in 1997. Including museum collections, the presently known coral fauna of the Marías Islands consists of 16 species. Coral zonation was characterized by



Fig. 5.7 Tropical mainland Mexico (TMM), Sonora to Chiapas. Red diamonds identify surveyed sites

shallow populations of *Pocillopora*, and deeper occurrences, to 10 m depth and deeper, of species of *Porites*, *Pavona*, and *Psammocora*. This is a frequent pattern observed throughout Mexico (Reyes-Bonilla and López Pérez 1998) and elsewhere in the eastern Pacific (Glynn 1976; Wellington 1982). The community of algal endosymbionts (*Symbiodinium*) inhabiting particular coral host species was found recently to be a critical factor in defining this depth zonation. From a combination of genetic and photo-physiological analyses of *Pocillopora verrucosa* and *Pavona gigantea*, Iglesias-Prieto et al. (2004) demonstrated that the physiological performance of host-specific zooxanthellae were adapted to different depth-related light regimes. Thus, pocilloporid holobionts exhibited optimal growth at shallow depths, whereas the growth rates of massive colonies were highest in deeper waters.

It is now recognized that the notable coral development of the Huatulco reef tract is subject to a variety of relatively frequent and severe natural disturbances with high

inter-annual variability (Glynn and Leyte-Morales 1997; López-Pérez et al. 2002; López-Pérez and Hernández-Ballesteros 2004). These disturbances range from seasonal upwelling events to plankton blooms and high turbidity conditions, hurricanes, ENSO high and low temperature extremes, and sea urchin outbreaks leading to intense grazing and bioerosion (e.g., Leyte-Morales 2000; Lirman et al. 2001; Herrera-Escalante et al. 2005; López-Pérez et al. 2007; Benítez-Villalobos et al. 2008). López-Pérez and Hernández-Ballesteros (2004) have proposed that coral mortality caused by such random spatio-temporal disturbances are likely to produce a mosaic of patches at different stages of recovery, thus contributing to dynamic and diverse coral community structures. It would be of interest to study the coral-rich Nayarit area, also subject to upwelling and severe ENSO activity, to determine if disturbance regimes there similarly influence coral community structure and dynamics.

Several recent molecular genetic studies have helped to elucidate the connectivity between coral populations and

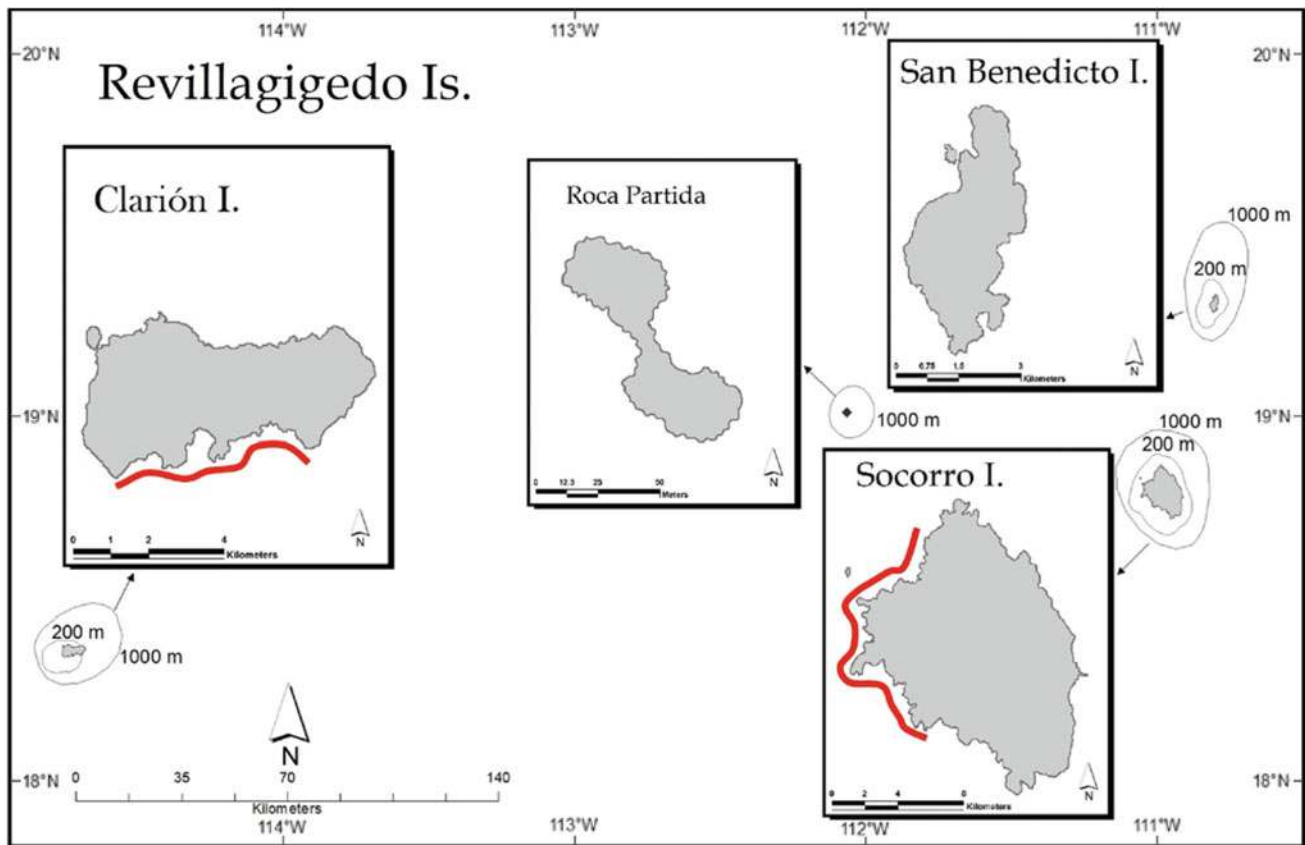


Fig. 5.8 Revillagigedo Islands (REV), Mexico. Red lines skirting the Clarión and Socorro coastlines denote abundant coral growth

their biogeographic relationships in Mexican waters and beyond. Most of these studies indicate a high degree of genetic structure in local populations and, presumably, limited gene flow. For example, significant geographic differentiation was reported for *Pocillopora damicornis*, a broadcast spawning species (Chavez-Romo et al. 2008), and *Porites panamensis*, a brooder with a brief larval life (Paz-García et al. 2008). Limited gene flow among most sampled localities was also reported for *Pavona gigantea*, a wide-ranging broadcast spawning species (Saavedra-Sotelo et al. (2011). However, the largest estimated migration rate indicated moderate and unidirectional gene flow from Huatulco to the Marietas Islands in Banderas Bay (Nayarit) and to La Paz. Blancas-López (2009) also observed weak genetic differentiation (i.e. connectivity) among populations of *Pocillopora verrucosa*, suggesting some level of gene flow among populations of this broadcast spawning species. Among the various physical and biotic factors invoked to explain these patterns, the surface circulation of west Mexico is critical. There is a general northerly set of surface currents, involving the Costa Rica Coastal Current (CRCC) moving along the Huatulco coast and the West Mexican Current that flows from Oaxaca to the entrance of the Gulf of California (see Chap. 3, Fiedler and Lavín). This would potentially

allow certain source populations in the south to supply larvae that could recruit to downstream Mexican mainland areas and eventually as far north as the Gulf of California. Since the CRCC originates in the equatorial eastern Pacific, it could conceivably serve as a vehicle of transport for coral larvae to reach Mexico from Costa Rica and Panama.

5.3.3 Revillagigedo Islands (Mexico)

The volcanic islands making up the Revillagigedo Islands group are dispersed between 19° to 20°N latitude, and range from 750 km (San Benedicto and Socorro Islands) to 1200 km (Clarión Island) to the west of the Mexican mainland coast (Fig. 5.8). Roca Partida, consisting of two small rock peaks with a very limited shallow shelf, is located to the NE of Clarión Island. The number of coral records in the Revillagigedo Islands increased dramatically in the early 1990s, thanks to scuba surveys conducted by Mexican workers. This increase in species richness, from 12 to 21 then to 25, was due mostly to the discovery of Indo-Pacific corals not present on the west Mexican coast or elsewhere in the eastern tropical Pacific (Ketchum and Reyes-Bonilla 1997, 2001). Several of these species are shared with

Clipperton Atoll, located nearly 1000 km to the south (at 10° N), which is occasionally in the path of the North Equatorial Counter Current (Glynn et al. 1996a, b). The current species list for the Revillagigedos totals 25 zooxanthellate scleractinians (Appendix). The strong coral faunal similarities between the Revillagigedos and Clipperton, also involving brachyuran crabs (Garth 1992) and reef fishes (Robertson and Allen 1996), prompted Ketchum and Reyes-Bonilla (1997, 2001) to suggest that this oceanic region should be recognized as a unique biogeographic subregion of the eastern tropical Pacific. The latter workers noted that surface currents would allow dispersal from Clipperton to the Revillagigedos, especially during the summer season, and that both island areas could serve as stepping stones for the migration of inshore marine species from the central to the more northern eastern tropical Pacific region. Traditionally, the W to E biotic migration route was assumed to be closer to the equator, from the Line Islands to Isla del Coco and Central America (Costa Rica, Panama) via the NECC (Dana 1975; Cortés 1997; Glynn and Ault 2000). Based on distributional evidence, it appears that there may be two dispersal routes, one just north of the equator and one leading to northern Mexican waters via oceanic island stepping stones.

The principal fringing reefs of Socorro and Clarión Islands are generally present in bays on western and southern shores in the lee of westerly-moving cyclones, which frequent this region (Fig. 5.8; Reyes-Bonilla 2003; see Chap. 3, Fiedler and Lavín). *Pocillopora* spp. predominate at shallow depths and massive corals (*Porites*, *Pavona*) increase in abundance from 5 to 30 m depth; coral rubble also increases with depth. Relatively few coral species were observed at Roca Partida and San Benedicto Island, likely due to the small size of these islands and restricted shelf space. In addition, the Bárcena volcano on San Benedicto Island was active as recently as 1952, and the resulting lava flows, debris, and fine sediments probably devastated near-shore communities and interfered with recovery. The six coral species observed there in 1994 were found principally on large basalt blocks and not on unstable volcanic sediments (Glynn et al. 1996a, b).

Moderate numbers of *Acanthaster planci* were observed at Clarión and San Benedicto Islands in 1994 (Glynn, unpublished observation). On 29 and 30 April, 36 individuals were counted on each of two occasions by a single diver between 6 and 17 m near Roca del Cuervo (18° 21' 13.7"; 114° 41' 06.2"), off the southeast shore of Clarión Island. The sea stars were feeding mostly on crustose coralline algae, but also on massive *Porites lobata* colonies, and large and small *Pocillopora eydouxi* colonies as well as broken branches of *Pocillopora* spp. Several *A. planci* also were observed feeding on *P. eydouxi* in coral communities on the northwestern end of San Benedicto Island on 3 May, 1994.

5.3.4 Clipperton Atoll (France)

Clipperton Atoll, the only atoll present in the eastern Pacific region, is located about 1300 km SW of the Mexican mainland and 950 km SSE of the Revillagigedo Islands (Fig. 5.1). Clipperton is volcanic in origin, part of a chain of guyots on the Clipperton fault, bordering the Cocos (south) and Pacific (north) plates. It is an 'almost atoll', with a 29 m high trachyte rock on the SE shore (Rocher Clipperton, Fig. 5.9), and an enclosed meromictic lagoon (Charpy et al. 2009; Trichet 2009). Present-day shallow, stratified brackish waters and H₂S-enriched bottom waters prevent coral growth inside the lagoon. However, before the mid-1800s at least two channels communicated with the open ocean, thus allowing for coral framework construction; fossilized coral deposits are still evident today inside the lagoon (Carricart-Ganivet and Reyes-Bonilla 1999). It is likely that a tectonic uplift event (Trichet 2009) closed off the lagoon between 1839 and 1849 (Charpy et al. 2010). Bottom profiling to 200 m depth around the perimeter of the atoll revealed a prominent 20 m terrace that is generally present at all seaward exposures (Glynn et al. 1996a, b). The 200 m isobath is located relatively close to shore, from 200 to 500 m seaward of the reef flat.

With recent coral studies supported by scuba, the known coral fauna has increased from 16 or 17 species (Carricart-Ganivet and Reyes-Bonilla 1999; Flot and Adjerdoud 2009) to 21 species (Fig. 5.7; Appendix). Contributing to this increase is the recognition of additional species of *Pocillopora* (e.g., *P. elegans* and *P. verrucosa*), the naming of a new poritid species (*Porites arnaudi*), and a molecular genetic study revealing the presence of *Porites evermanni* (Boulay et al. 2014). These new species' records are not considered recent introductions, but resident species present for some undetermined time. *Pavona gigantea* was listed from Clipperton by Durham and Barnard (1952), but has not been observed at the atoll for several decades since. The zooxanthellate scleractinian fauna of Clipperton, consisting of mainly (~90 %) Indo-Pacific species is most closely allied with the Revillagigedo Islands in the EP (Fig. 5.2).

In 1994, live coral cover was exceptionally high (10–100 %) at all insular shelf depths from 8 to 60 m, and crustose coralline algae were abundant (5–40 % cover) between 0.5 and 7 m depth (Glynn et al. 1996a, b). *Porites lobata* and *Pavona varians* contributed prominently to live cover, the former up to 50 % of cover between 10 and 30 m, and the latter commonly as high as 100 % between 25 and 60 m and deeper. Monospecific patches of *Pavona varians* were present at 70 m. *Porites arnaudi*, noted as *Porites* sp., was commonly observed, but *P. lobata* predominated in most areas. *Porites arnaudi* was often more common on the near-vertical faces of the 20 m terrace (Fig. 5.10). *Pocillopora* sp. patches, 10s to 100s m² in area, were highly

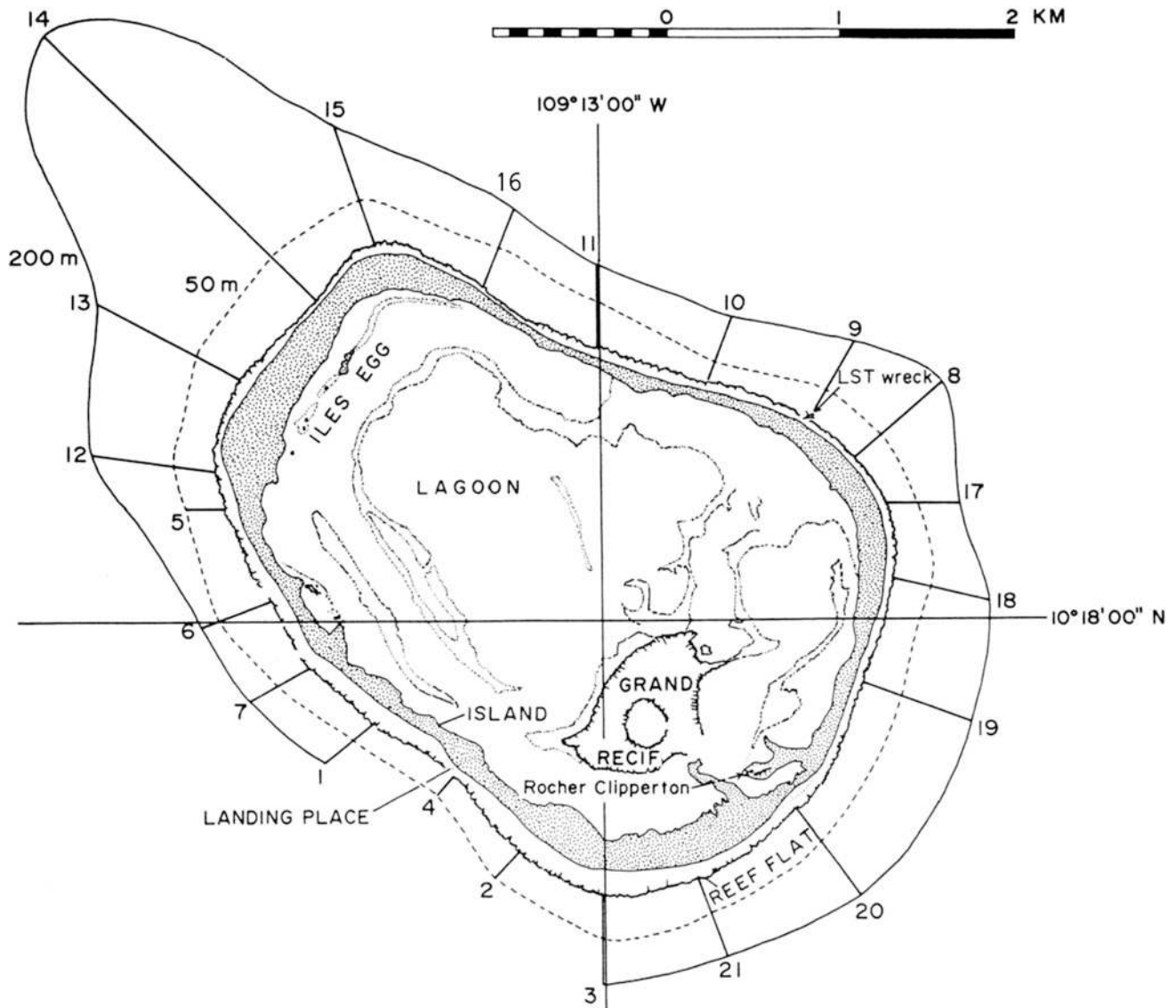


Fig. 5.9 Clipperton Atoll (CLP), modified after Glynn et al. 1996a. 50 and 200 m isobaths are based on the 21 bottom-profiling transects shown

dispersed and usually present between 5 and 20 m depth. The corals in most of these patches were dead and encrusted by the coralline alga *Porolithon onkodes*. The pocilloporid species making up these patches, later identified by Carricart-Ganivet and Reyes-Bonilla (1999), was *Pocillopora meandrina*. More recent workers have not commented on whether the *Pocillopora* patches were dead or recovering. *Pavona minuta*, *Pavona maldivensis*, and *Leptoseris scabra* were generally minor components of the coral communities. Surveys conducted 10 years later indicated no significant change in the overall robust state of Clipperton's coral communities (Flot and Adjeroud 2009). Two permanently-marked monitoring sites were established in 2005 at 10–12 m depth on the north and south reef slopes, thus allowing an opportunity to track coral community condition over time (Salvat et al. 2008).

Like most ETP localities, the coral reef biota of Clipperton exhibits a strong affinity with the central/south Pacific region. In addition to the zooxanthellate coral fauna, high proportions of reef algae (Payri et al. 2009), crustaceans (Poupin et al. 2009), molluscs (Kaiser 2009), echinoderms (Solís-Marín and Laguarda-Figueras 2009), and fishes at Clipperton (Robertson and Allen 1996; Robertson et al. 2004; Béarez and Séret 2009) exhibit transpacific distributions. For coral reef fishes, Robertson et al. (2004) noted that pelagic spawners with strong dispersal ability were highly represented among transpacific species. Also, many reef-associated Indo-Pacific invertebrate taxa inhabiting eastern Pacific reefs possess teleplanic larvae (Scheltema 1988; Glynn and Ault 2000). Molecular studies demonstrate gene flow and recent invasions between the Central Pacific and Clipperton Atoll of coral, echinoderm, and fish species

Fig. 5.10 *Porites arnaudi*, cascading skirt-like growth form, 18 m, NE exposure, Clipperton Atoll, 23 April 1994 (photo credit M. Kazmers, Shark Song)



(Lessios et al. 1998; Lessios and Robertson 2006; Baums et al. 2012). Some of these studies have shown east to west gene flow, and connectivity of reef species among eastern Pacific oceanic islands (see Chap. 16, Lessios and Baums). Thus, it is necessary to know the flow regimes and transit times of the North Equatorial Counter Current (NECC), the primary eastward-flowing current in the central and eastern tropical Pacific.

Satellite-tracked drifting buoy data analyzed over a 15 year period (1979–1993) revealed a mostly eastward flow of the NECC across the E Pacific ‘barrier’ between 6° and 10°N, occasionally reaching 12–14°N (Glynn et al. 1996a, b). Travel times from the Line Islands in the Central Pacific to Clipperton Atoll were commonly 200 days, although one record during an

ENSO event showed the NECC travelling across 46° of longitude in about 60 days. A comparison of west to east distances travelled during non-El Niño and El Niño years indicated a significant increase in NECC velocity during the latter periods. Accordingly, the NECC could offer an effective means of larval transport to Clipperton, which in turn could serve as a stepping stone to other ETP localities, e.g. the Revillagigedo Islands as suggested by Ketchum and Reyes-Bonilla (1997).

5.3.5 El Salvador

The recently described coral assemblages of El Salvador and Nicaragua are populated by a combined total of 14

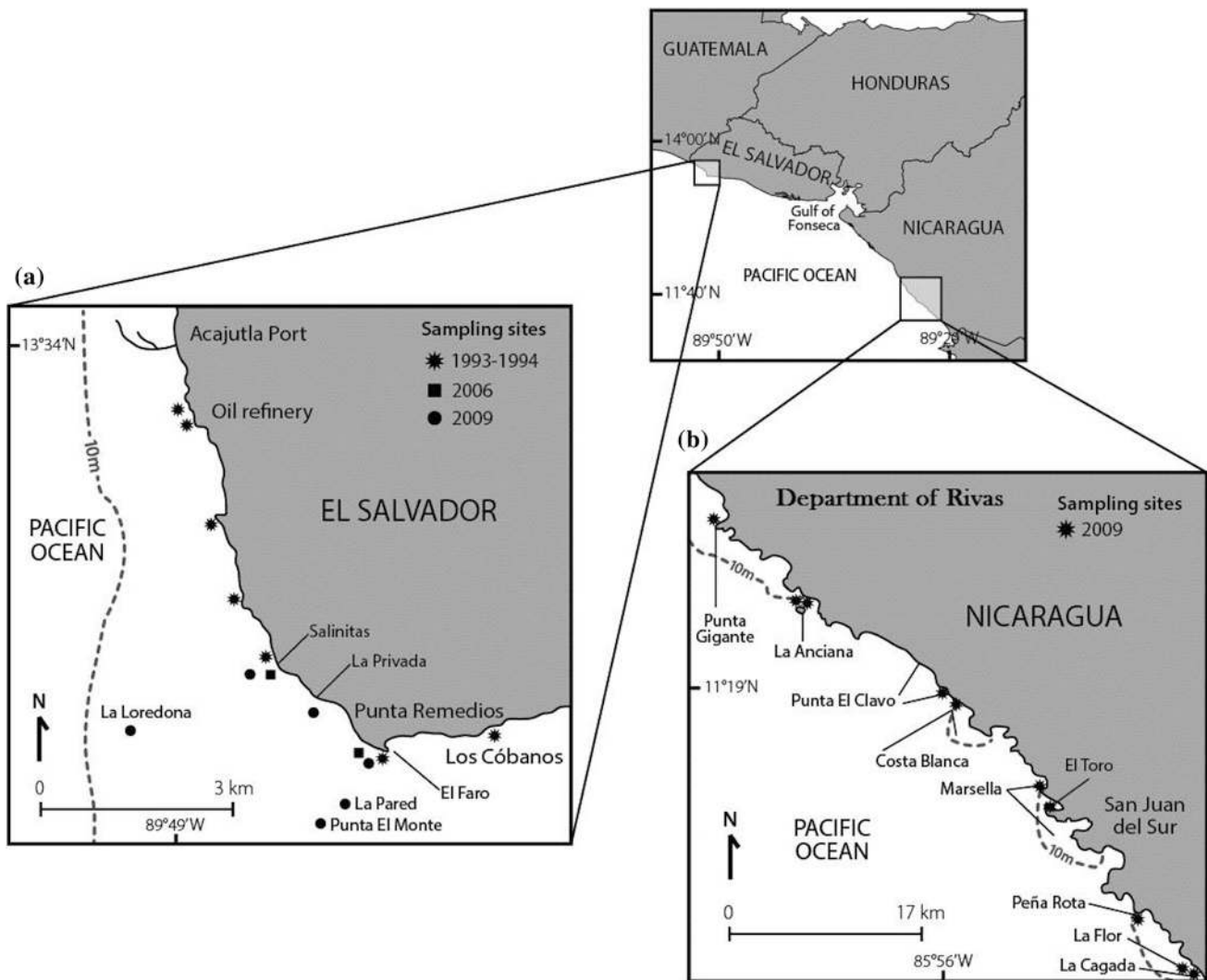


Fig. 5.11 a Northwestern El Salvador (ELS); b southern Nicaragua (NIC), Central America. Coral development on Nicaraguan coast is within the Department of Rivas. Ten meter isobaths from US Navy Hydrographic chart no. 21026

zooxanthellate species. Notable differences, however, are the occurrences of *Porites lobata* at El Salvador only (Reyes-Bonilla and Barraza 2003), and *Pavona clavus*, *Pavona varians*, and *Gardineroseris planulata* in Nicaragua but not at El Salvador (Alvarado et al. 2011). No structural coral reef development has been reported from these areas (see Chap. 6, Toth et al.), although coral community patches are present.

The coastal stretch between Guatemala and northwestern Nicaragua (Gulf of Fonseca) is known as the “Pacific Central American Faunal Gap (PCAFG)”, originally named due to the absence of rocky shore fishes (Springer 1958). The Guatemalan coast and the northern sector of El Salvador consist of sandy beaches and mangrove forests. Until recently the PCAFG was also understood to separate the coral faunas of southern Mexico and Costa Rica (Glynn and

Ault 2000). At Los Cóbanos (El Salvador), there is an abrupt change in the orientation of the coastline with the appearance of low rocky outcrops and platforms (Fig. 5.11a). The remainder of the Salvadoran coast consists of high energy sandy beaches or muddy estuaries. The littoral fringe of the Gulf of Fonseca consists of rocky outcrops and one of the most extensive mangrove ecosystems of Central America. The southern Nicaraguan coast consists of a combination of rocky headlands and sandy beaches (Cortés 2007). The occurrence of coral populations within the PCAFG could serve as a dispersal corridor, and explain the close association of southern and northern ETP coral faunas (Fig. 5.2).

The rocky outcrops of Los Cóbanos support the best developed and most diverse coral communities in El Salvador (Reyes-Bonilla and Barraza 2003). Small pocilloporid coral communities occur on basalt outcrops, most no larger

than 30 m², that occupy a total of 159 km² of a large terrace (8000 ha) that borders the coast. Thirteen coral species (eight zooxanthellate and five azooxanthellate) were reported at this site. Lemus et al. (1994) performed the first quantitative coral surveys at Los Cóbános in 1993 and 1994; these workers sampled *Porites lobata* and reported highly divergent live cover values between 0.41 and 62 % for this species. The mean diameter of the sampled *Porites* colonies was ~50 cm. At Punta Remedios (Fig. 5.11a), they reported 0.42 % live coral cover, 34.6 % algae, and 44.8 % sand and rock. Only dead *Pocillopora* fragments were noted at this site.

New sites at Los Cóbános (Playa El Faro, Playa Salinitas or Decameron Beach) were sampled in 2006 by Segovia-Prado and Navarrete-Calero (2007). They reported high live cover of large *Porites* colonies: El Faro—33.7 % cover, 161 colonies, mean diameter = 121 cm; Salinitas—65.4 % cover, 75 colonies, mean diameter = 146 cm. Algal cover also was high: El Faro—41.7 %; Salinitas—29.5 %. These mixed coral/algal communities were classified into five distinct assemblages, based on generic prevalence: (1) *Padina-Halimeda-Codium*, (2) *Porites-Padina-Halimeda*, (3) *Porites-Acanthophora-Padina*, (4) *Porites-Padina-Hypnea*, and (5) *Porites-Dictyota-Galaxaura*. Concerns were raised over possible threats to corals by the invasive alga *Acanthophora spicifera*, and a high incidence of bioeroders (*Lithophaga*, *Cliona*) and potential space competitors (vermetid gastropods). Reyes-Bonilla and Barraza (2003) noted other macroalgae, invertebrates (e.g., octocorals, molluscs) and reef-associated fishes observed at Los Cóbános. A potentially significant physical threat noted by Reyes-Bonilla and Barraza (2003) is the seasonally high rainfall period from May to October with associated severe coastal erosion and transport of suspended sediments.

At Playa el Zope (Los Cóbános), López and Jiménez (2008) found healthy colonies of *Psammocora obtusangula* and *Psammocora stellata* interspersed with *Porites lobata*. They reported 81 colonies of *P. obtusangula* (mean length × width = 10.8 × 7.2 cm) and two small colonies of *P. stellata* (<6 cm²). Several *P. obtusangula* colonies bore small wounds resembling fish bite marks.

The most recent near-shore surveys at Los Cóbános, in 2009 and at the same sites sampled by Lemus et al. (1994), and Segovia-Prado and Navarrete-Calero (2007), failed to reveal living or dead *Pocillopora* and reported only low *Porites lobata* cover of 0.1 and 5.5 % (Alvarado, unpublished data). A live *Pocillopora* community was reported in the early literature at Los Cóbános, the Punta Remedios site (Orellana-Amador 1985; Gotuzzo 1996), but has not been re-located in recent surveys (Reyes-Bonilla and Barraza 2003). All *Pocillopora* spp. collected in this study were identified from dead beach fragments (Appendix). At relatively deep (10–15 m) offshore sites, only azooxanthellate

corals were observed, with *Tubastraea coccinea* predominant. Considering the high variability of live coral cover reported over the years by different workers, and the uncertain location of sampling sites, this coastal sector is in need of a systematic and technologically advanced survey protocol.

5.3.6 Nicaragua

Before 2010, coral surveys along the Pacific coast of Nicaragua indicated the possibility of isolated colonies of *Pocillopora* close to the Costa Rican border in the area of San Juan del Sur (Durham and Barnard 1952; Glynn and Ault 2000; Spalding et al. 2001; Ryan and Zapata 2003; Cortés 2007). The presumed meagre occurrence of corals in this area may be related to severe and prolonged upwelling (Glynn et al. 1983).

In July 2009, Alvarado et al. (2010, 2011) quantitatively assessed the composition of ten coral communities along the southern coast of Nicaragua, including sites in the Department of Rivas, Municipalities of San Juan del Sur, and Tola (11° 23' N; 86° 02' W and 11° 07' N; 85° 47' W; Fig. 5.11b). They reported the presence of 39 species of macroalgae, 13 scleractinian corals (nine zooxanthellate and four azooxanthellate species), two crustaceans, five molluscs, 11 echinoderms, and 52 reef fishes. Mean coral cover across all sites was 9.0 ± 1.9 (SD)%, with the highest cover at Guacalito-La Anciana (18.5 ± 8.8 %) and Punta Pie de Gigante (16.7 ± 5.3 %). The most common species were *Gardineroseris planulata* and *Pavona gigantea* with notably high cover at Punta Gigante and La Anciana (Fig. 5.11b). A relatively large and vibrant *Pocillopora* incipient reef was present in the Punta El Toro embayment with mean live cover of 14.5 ± 7.9 %.

The Rivas coral biotopes, such as those present at Punta Gigante, La Anciana, and Punta El Toro, were small, patchy, and confined to a series of small coves and islets protected from wave assault. These communities were dominated by *Pavona gigantea*. The coral reef fish assemblages were less diverse than those reported for more developed eastern Pacific reef areas elsewhere, but were similar to those in marginal environments, such as upwelling centers (Alvarado et al. 2011). The coral-rich area between Punta Gigante and La Anciana, a 13-km coastal stretch, was touted by Alvarado and co-workers as a hotspot—despite the lack of reef development—and in urgent need of protection and management.

5.3.7 Mainland Costa Rica

A dramatic increase in coral species richness, from 10 to 26 species, is evident in faunal inventories when passing from Nicaragua to Costa Rica (Table 5.4; Appendix). This

Table 5.4 Reef building corals from Pacific Costa Rica

Location	1	2	3	4	5	6	7	8	9	10	11
Suborder ASTROCOENIINA, Family Pocilloporidae											
1. <i>Pocillopora capitata</i> Verrill, 1864				CA	CA	CA				CA	CA
2. <i>Pocillopora damicornis</i> (Linnaeus, 1758)		RB	RB	CA	RB	RB	RB	RB	RB	RB	RB
3. <i>Pocillopora elegans</i> Dana, 1846		RB	RB	RB	RB	RB	RB	RB	RB	RB	RB
4. <i>Pocillopora eydouxi</i> Milne Edwards and Haime, 1860		RB	CA	CA	CA			CA		CA	RB
5. <i>Pocillopora inflata</i> Glynn, 1999		RB	RB	CA	CA					CA	CA
6. <i>Pocillopora meandrina</i> Dana, 1846		CA	CA	CA	CA						CA
7. <i>Pocillopora verrucosa</i> (Ellis and Solander, 1786) ^a				RB	CA					RB	
Suborder Fungiina, Family Poritidae											
8. <i>Porites evermanni</i> Vaughan, 1907					RB		RB	RB		RB	CA
9. <i>Porites lobata</i> Dana, 1846		CA	CA	CA	RB	RB	RB	RB	RB	RB	RB
10. <i>Porites panamensis</i> Verrill, 1866	CA	CA	CA	CA	CA	CA	CA	CA		CA	
11. <i>Porites rus</i> (Forskål, 1775)					CA						
Family SIDERASTREIDAE											
12. <i>Psammocora obtusangula</i> (Lamarck, 1816)					CA	CA	CA	CA	RB	CA	
13. <i>Psammocora stellata</i> Verrill, 1864		CA	RB	RB	CA	CA	CA	CA	CA	RB	CA
14. <i>Psammocora superficialis</i> Gardiner, 1898		CA	CA	CA	CA	CA	CA	CA	CA	CA	CA
Family Agariciidae											
15. <i>Gardineroseris planulata</i> (Dana, 1846)		RB	RB	CA			CA	CA		RB	RB
16. <i>Leptoseris papyracea</i> (Dana, 1846)			CA	RB	CA						CA
17. <i>Leptoseris scabra</i> (Vaughan, 1907)			CA								CA
18. <i>Pavona chiriquiensis</i> Glynn et al. 2001			CA	CA		CA				CA	CA
19. <i>Pavona clavus</i> (Dana, 1846)		CA	RB	RB	CA	CA		CA	CA	RB	RB
20. <i>Pavona frondifera</i> (Lamarck, 1816)				CA			CA		RB	CA	CA
21. <i>Pavona gigantea</i> Verrill, 1869	RB	RB	CA	RB	CA	CA		CA	CA	CA	CA
22. <i>Pavona maldivensis</i> (Gardiner, 1905)		CA	CA	CA		CA	CA			CA	CA
23. <i>Pavona varians</i> Verrill, 1864	CA	CA	CA	CA		CA		CA	CA	CA	CA
24. <i>Pavona minuta</i> Wells, 1954											CA
Family Fungiidae											
25. <i>Cycloseris curvata</i> (Hoeksema, 1989)				CA					CA		CA
26. <i>Diaseris distorta</i> (Michelin, 1843)				CA							CA
Total number species	3	14	17	21	17	13	11	13	11	19	22

RB = Main reef builder, CA = coral community associate. Areas 1 = Bahía Salinas, 2 = Península de Santa Elena, 3 = Islas Murciélago, 4 = Bahía Culebra, 5 = Península de Nicoya, 6 = Pacífico Central, 7 = Parque Nacional Marino Ballena, 8 = Península de Osa, 9 = Golfo Dulce, 10 = Reserva Biológica Isla del Caño and 11 = Parque Nacional Isla del Coco. RB reef building reported; CA coral community associate

^aThis species was not recognized in the field until recently; it is probably more widespread than indicated

disparity is likely due to a combination of factors: (a) the recent discovery of coral communities in Nicaragua, (b) a concentrated research effort on coral reefs in Costa Rica since the 1980s, (c) the relatively large area of varied coastal habitats along Costa Rica's coast, and (d) the stressful conditions associated with upwelling along the southeastern Nicaraguan coast. The mainland coastline of Costa Rica consists of diverse marine ecosystems as well as islets and islands of various sizes, and differences in proximity to the

coast (Denyer and Cárdenas 2000; Cortés and Wehrmann 2009). The presently recognized tally of 26 zooxanthellate coral species from mainland Costa Rica has been relatively stable during the past decade, thanks to the sustained studies of local workers (e.g., Guzmán and Cortés 1989a; Cortés 1990; Jiménez 1997, 2001a, b; Cortés and Guzmán 1998). This inventory takes into account a single new record for Costa Rica, *Porites evermanni*, revealed in a molecular genetic study (Boulay et al. 2014), and *Porites rus*, not

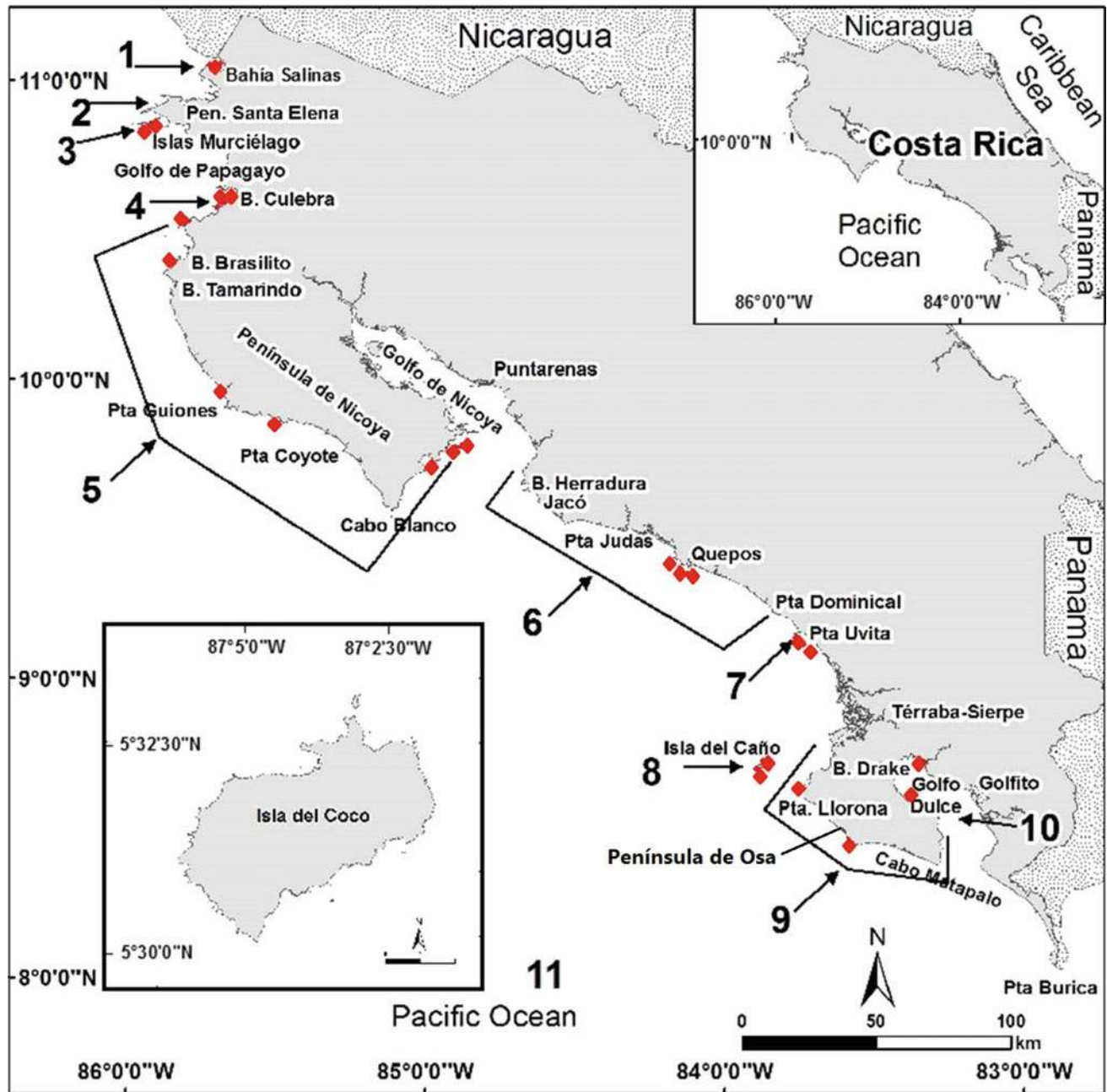


Fig. 5.12 Costa Rica mainland (CRM) partitioned into 10 coastal and one offshore (Isla del Coco) coral-bearing sectors. Red diamonds identify sites with abundant corals

observed again since its discovery at Bahía Sámara (sector 5, Fig. 5.12) in the early 1980s (Cortés and Murillo 1985). *Pavona chiriquiensis*, initially described from populations in Panama, is now known to occur at several sites along the Costa Rican coast. New distributional records for *P. chiriquiensis* in the central and south Pacific include the Tokelau Islands (~172°W), American Samoa (~170°W), the northern and southern Phoenix Islands (~170°W), and the Line Islands (155°–160°W) (Maragos, unpublished

records). A cluster analysis of presence/absence data demonstrates a close affinity with the coral faunas of Panama and Colombia (Fig. 5.2).

The northwestern Pacific sector of Costa Rica is characterized by a marked dry season (January–April), with strong easterly trade winds that promote seasonal upwelling (Cortés 1996/1997; Jiménez 2001b; Alfaro et al. 2012). Upwelling is most pronounced further north, toward the Nicaraguan border. The dry climate extends south to Golfo de Nicoya,

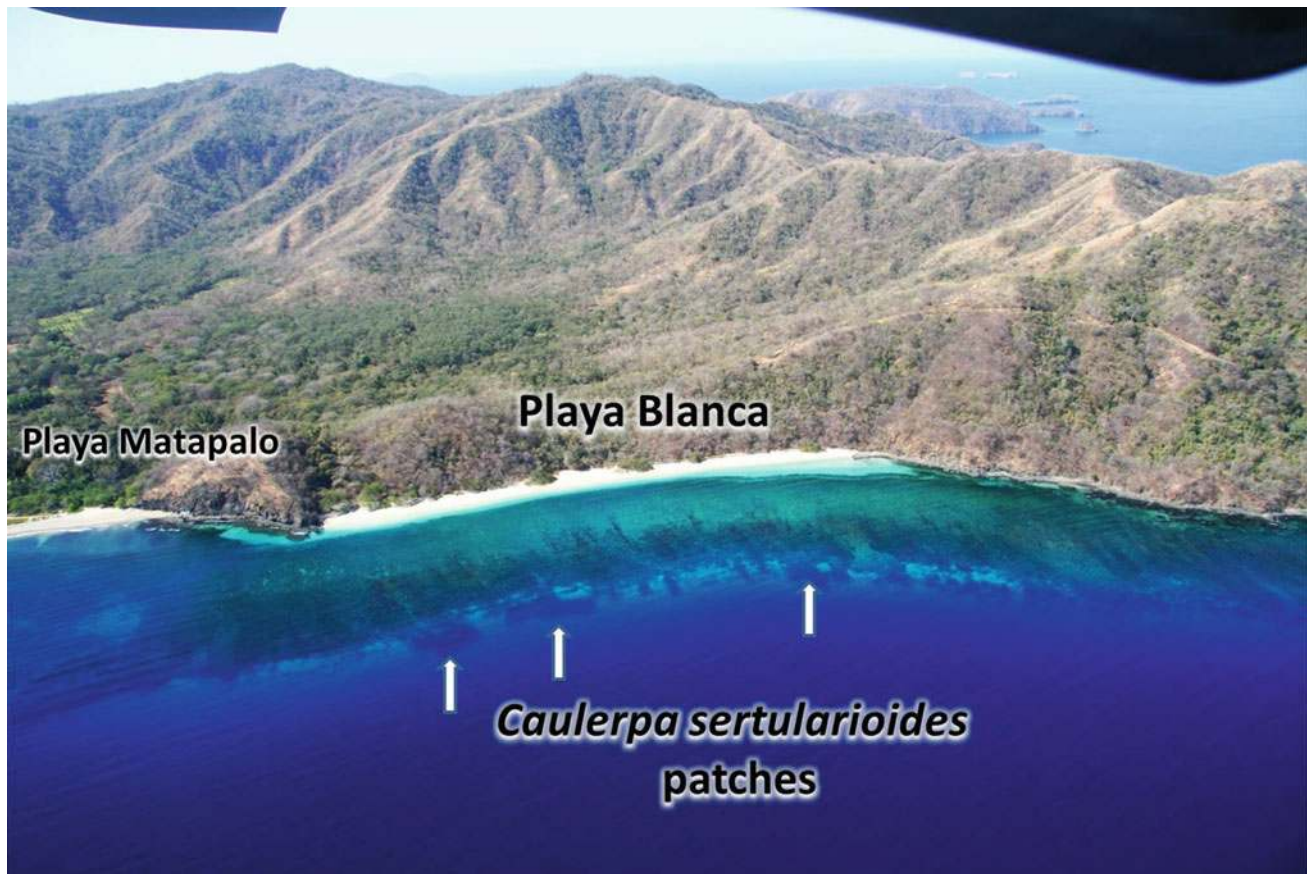


Fig. 5.13 Matapalo fringing reef, Punta Gorda, Costa Rica (10 February 2007, ~1000 m elevation, courtesy María M. Chavarría). Patches of an invasive green alga, *Caulerpa sertularioides*, seaward of live coral cover, ~10 m depth

the largest estuarine environment in Costa Rica. The central coastal section is a transitional zone with an increasing humid climate to the south. The southeastern coast is subject to year-round high rainfall, which supports a tropical rain forest ecosystem (Jiménez and Soto 1985; Herrera 1986; Kappelle et al. 2002).

For ease of discussion, mainland Costa Rica is divided into 10 sectors where coral communities and reefs occur, from the Nicaraguan border to Golfo Dulce near Panama (Fig. 5.12). Oceanic Isla del Coco (Cocos Island) is assigned to sector 11, and is considered below in Sect. 5.3.8.

The northern-most area, Bahía Salinas (sector 1), is heavily influenced by seasonal upwelling in the Gulf of Papagayo (Alfaro et al. 2012). Water temperature in this area can be as low as 12 °C with low oxygen levels and low pH (recorded by Rixen et al. 2012 in Bahía Culebra; see also Jiménez 2001a; Alfaro and Cortés 2012; Alfaro et al. 2012). Because of these extreme conditions, reef and coral community development are limited; *Pavona gigantea* is predominant with isolated colonies of *Porites panamensis*. Upwelling effects generally decrease to the south and southeast towards the Nicoya Peninsula. At Peninsula de

Santa Elena (sector 2), relatively large pocilloporid reefs are present; one of these structural reefs is built mainly by *Pocillopora eydouxi* (Cortés 1996/1997; Bassey-Fallas 2010). Immediately south of Santa Elena, lie the Islas Murciélago (sector 3). Several coral reefs occur in this area; one is constructed of massive *Gardineroseris planulata* colonies. An extensive pocilloporid reef, consisting of large areas of *Pocillopora inflata*, was killed during a phytoplankton bloom in 2007 (Jiménez et al., in preparation). Coral recovery, including sexual and asexual recruitment, is very slow in this cool upwelling zone (Cortés, unpublished data). Bahía Culebra (sector 4) is one of the most intensely studied areas in Pacific Costa Rica (Cortés 2012a). Coral reefs and coral communities have been surveyed and monitored in this large bay and surrounding areas since the early to mid-1980s (Cortés and Murillo 1985; Jiménez 1997, 2001b; Jiménez et al. 2001, 2010; Bezy et al. 2006; Cortés et al. 2010; Cortés 2012a).

The Matapalo fringing reef in the southern part of sector 4, 1.4 km in length, is possibly the largest coral reef bordering mainland Costa Rica (Jiménez 2007a; Fig. 5.13). This reef, first surveyed quantitatively in 1978, was largely

degraded at that time; it consisted of large tracts of dead pocilloporid corals, and at most 0.1 % live cover of *Psammocora stellata* (Glynn et al. 1983). [An aerial view of this reef in 1973, with the name Punta Gorda, is shown in Fig. 5.4a in Glynn et al. 1983.] The death of the reef at that time was attributed to cold-water stress due to intense upwelling near the end of the Little Ice Age (150–300 years BP). The overall live cover of pocilloporid corals on this reef amounted to >40 % when surveyed in 2007. Species richness was high, consisting of 16 living species, and 19 species total, including freshly dead coralla of three species (*Cycloseris curvata*, *Diaseris distorta*, and *Leptoseris papyracea*). Carbonate buildups or bioherms of *Leptoseris papyracea* are unique to this area. The Matapalo reef has again experienced recent high coral mortality first observed in 2007, and coincidental with extensive and prolonged harmful algal blooms (Jiménez 2007b). It is likely that recent declines in coral cover are due, at least in part, to coastal development activities and poor waste management promoting algal blooms, which are now nearly monthly occurrences.

Overall, much of the reef building in the extreme northwestern area of Costa Rica (sectors 1–4) has been achieved by massive instead of branching pocilloporid species. For example, the major reef builders north of Islas Murciélago are *Pavona gigantea* and *Gardineroseris planulata*. At Bahía Culebra, *Pavona clavus* is one of the major reef builders (Cortés and Jiménez 2003).

Extensive growths of the invasive green alga *Caulerpa sertularioides* have been reported in northwestern Costa Rica at Bahía Culebra (Fernández and Cortés 2005; Fernández-García 2007; Fernández-García et al. 2012), and at the Matapalo reef (Jiménez 2007a). Since 2001, *C. sertularioides* cover has been expanding with negative impacts on coral reefs and communities. At La Penca, Bahía Culebra, a *Psammocora* reef lost 95 % live cover at the expense of this alga during a 6 year period (Bezy et al. 2006). It is a strong competitor for space, overgrowing and smothering low-profile coral colonies. *Caulerpa racemosa* blooms are also known to cause coral mortality in Panama (see Fig. 8.31 in Chap. 8, and Fong et al., Chap. 11).

Little studied and isolated reefs occur in sector 5 along the outer coastline of Península de Nicoya (Cortés and Jiménez 2003; Bezy et al. 2006; Jiménez et al. 2010). A few small colonies of *Porites rus*, first observed in the eastern Pacific at Bahía Sámará, about 10 km SE of Punta Guiones, disappeared a few years after their discovery (after 1983) and have not been observed anywhere in the EP since. *Porites evermanni* was recently reported in coral communities on the Península de Nicoya, in other areas of Costa Rica, and elsewhere in the equatorial eastern Pacific (Boulay et al. 2014). The identity of this species was determined from a molecular genetic study; judging from its widespread

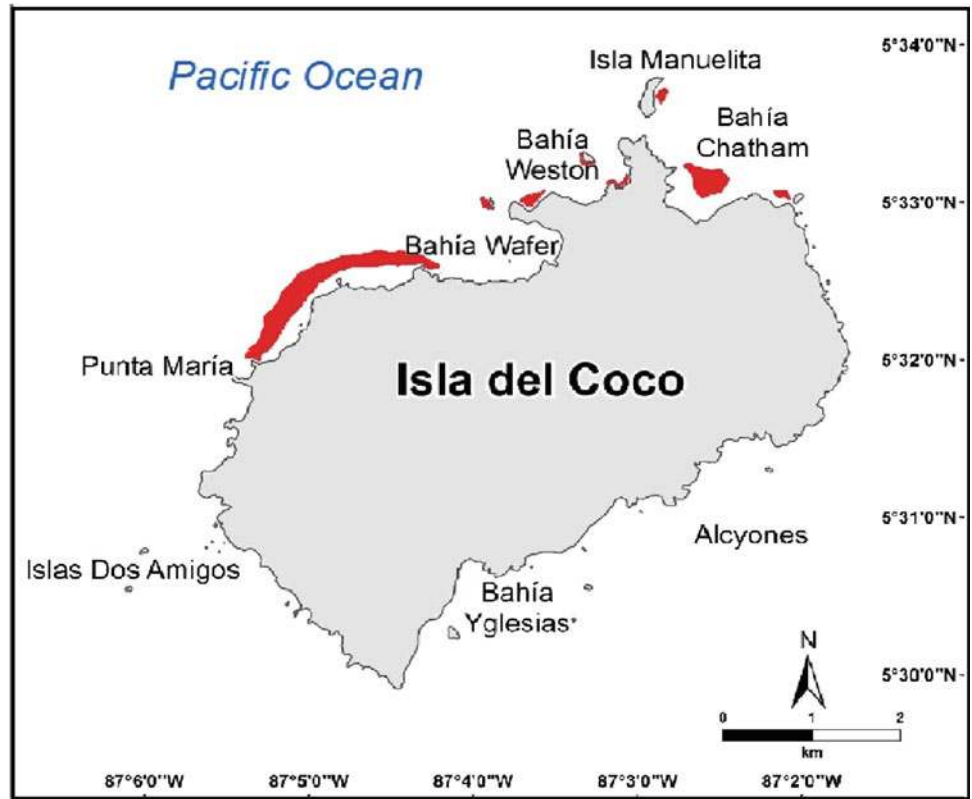
occurrence and large population sizes it is probably a long-term resident of the equatorial eastern Pacific. Reef-building no longer occurs in the Gulf of Nicoya due to high suspended sediment loading and freshwater dilution. Currently only dead pocilloporid frameworks are present in the gulf, usually in sheltered environments.

The central Pacific coastline (sector 6) extends from the east margin of the outer Gulf of Nicoya to Punta Dominical. Isolated reefs and coral communities also have been reported in this area (Cortés and Jiménez 2003). Parque Nacional Marino Ballena (sector 7) is exceptional in the central Pacific coastal area because of its well-developed reefs (Alvarado et al. 2005). These reefs have been impacted by El Niño warming events (Jiménez and Cortés 2001) and excessive terrigenous sedimentation resulting from coastal development (Alvarado et al. 2005, 2009).

The Reserva Biológica Isla del Caño (Caño Island Biological Reserve, sector 8) is located about 15 km west of Península de Osa. Well-developed coral reefs are present at Isla del Caño, from drying reef flats (unusual in the ETP) with abundant *Pocillopora* spp., crustose coralline algae and *Porites* microatolls to deep patch reefs constructed of *Porites lobata* and *Pavona clavus* (Guzmán and Cortés 1989a; Macintyre et al. 1993; Cortés and Jiménez 2003). These coral communities have been intensely studied, including corallivore activity (Guzmán 1988a, b), coral growth rates (Guzmán and Cortés 1989b), zooplankton ecology (Guzmán and Obando 1988), and meiofaunal populations of reef sediments (Guzmán et al. 1987a). Also, coral mortality associated with algal blooms (“red tides”) was documented at Isla del Caño following the 1982–83 El Niño warming event (Guzmán et al. 1987b, 1990). Finally, Glynn et al. (1991, 1994, 1996a, b, 2000, 2008, 2011, 2012; see Chap. 15, Glynn et al.) have completed long-term studies on the reproductive activities of eleven reef-building coral species and one abundant azooxanthellate coral at Isla del Caño. This work has demonstrated a potentially important role of sexual reproduction in recovery processes in Costa Rica (Guzmán and Cortés 2001), and at several other areas in the equatorial ETP (Glynn and Colley 2008; Glynn et al. 2012).

Coral communities and reefs occur on the seaward side of the Península de Osa (sector 9), but are generally only present in sheltered locations due to high wave assault (Cortés and Jiménez 1996, 2003). Golfo Dulce (sector 10), a fjord-like embayment (Hebbeln and Cortés 2001), is protected from high seas; numerous coral communities and a large fringing reef are present in this gulf (Cortés 1990; Macintyre et al. 1993; Cortés and Jiménez 2003). One of the upper gulf reefs (Punta Islotes), constructed primarily of *Porites lobata*, initiated growth over 5000 years ago (Cortés et al. 1994). Highest accretion occurred between 2500 and 500 years BP, and then slowed due to changes in the local

Fig. 5.14 Isla del Coco (COC), Costa Rica. Red patches denote location of coral reefs (modified after Cortés and Jiménez 2003)



geological setting that increased freshwater input. Reef growth ceased during the last 60 years due to high terrigenous sediment loads.

5.3.8 Isla del Coco (Costa Rica)

Isla del Coco (sector 11), is located over 500 km offshore of Costa Rica, about half way between the Pacific coast of Central America and the Galápagos Islands (Cortés 2008). It is a volcanic island situated on the northeastern-trending Coco Volcanic Cordillera (Rojas and Alvarado 2012). The island and surrounding waters are a marine protected area—Parque Nacional Isla del Coco—the largest MPA in Costa Rica, and is now imbedded in the Área Marina de Manejo de Montes Submarinos (Sea Mounts Marine Management Area).

Twenty-two zooxanthellate scleractinians make up the Isla del Coco coral fauna, compared to 25 species total for all 10 Costa Rican mainland sectors (Table 5.4; Appendix). Of all eastern tropical Pacific coral localities, the Isla del Coco coral fauna demonstrates its closest affinity with the Costa Rican mainland (Fig. 5.2; Cortés 2012b). *Pavona xarifae*, once considered a unique Isla del Coco record in the eastern tropical Pacific, has been synonymized with *Pavona minuta* by Veron (2000). Veron's synonymy is recognized here; therefore *P. minuta* is now more broadly distributed,

considered present at Clipperton Atoll, in the Revillagigedo Islands, and at Palmyra Atoll (Appendix). Verification is necessary, however, ideally from specimens collected at their type localities and subjected to the same genomic and morphological analyses.

Coral reefs are best developed on the northern island shelf (from Punta María to Wafer Bay, and at Weston and Chatham bays), which is protected from southern swells (Guzmán and Cortés 1992; Cortés 2014). The largest Costa Rican coral reefs occur at Isla del Coco, with one reef exceeding 2 km in length (Fig. 5.14). *Porites lobata* is the chief framework species, with colonies of *Pavona clavus*, *Pavona gigantea*, and *Gardineroseris planulata* also contributing to reef building; some reef areas have extensive cover of *Pocillopora* spp. as well (i.e., *P. elegans*, *P. damicornis*, and *P. eydouxi*). Severe coral bleaching and high levels of coral mortality occurred at Isla del Coco during the 1982–83 El Niño warming event (Guzmán and Cortés 1992; Macintyre et al. 1993). Pre-El Niño coral cover of 80–90 % on some reefs declined to 3–26 % after the event. However, by 2002 recovery had been impressive and continues to the present (as of 2015). Since the mid- to late-1990s, *Leptoseris scabra*, a wide-ranging Indo-Pacific species, appeared for the first time on Isla del Coco reefs and now plays a role there in reef consolidation. The appearance of other species not reported from Isla del Coco before the mid-1990s include *Pavona maldivensis*, *Pavona frondifera*, *Pavona chiriquiensis*, and

Pocillopora inflata. All three *Pavona* species are known to occur at Palmyra Atoll, Line Islands (Appendix), and *Pocillopora inflata* may be present in the Phoenix Islands (Obura 2002). The sudden appearance of these species at Isla del Coco, in the downstream path of the North Equatorial Counter Current (NECC), prompted Guzmán and Cortés (1992, 2007) to suggest that long-distance dispersal may be an important factor in the recovery of these eastern Pacific reefs following ENSO disturbances.

Summarizing for all Costa Rican reef sites subject to disturbance and long-term monitoring, a gradient of coral recovery is apparent that is related to increasing anthropogenic impacts and levels of protection (Cortés et al. 2010). The emerging pattern, from coral reefs exhibiting rapid to slow recovery, is as follows: Isla del Coco, Isla del Caño, Parque Nacional Marino Ballena, and Bahía Culebra. Coral cover at oceanic Isla del Coco, the most remote of the monitored sites, exhibited higher levels of recovery than the reefs at Isla del Caño, a moderate distance off-shore and relatively un-disturbed by humans. The lowest levels of coral recovery occurred at Parque Nacional (PN) Marino Ballena and Bahía Culebra, both on-shore areas subject to diverse coastal stresses. The PN Marino Ballena is protected

whereas the coral reefs at Bahía Culebra are not. The degree of protection of these four areas is greatest at Isla del Coco and least at Bahía Culebra. A similar pattern was reported by Edgar et al. (2010) who found that eastern Pacific MPAs with higher controls and regulation had greater fish biomass and more coral cover than areas with limited or no protection.

5.3.9 Panama

Panama has been the focus of coral reef ecological studies since the early 1970s. The initial research in Panama during the 1970s was directed toward documenting the presence of structural coral reefs in the eastern Pacific region (see Chap. 1 for a historical perspective). With the discovery of numerous reefs, interests then turned to investigation of the physical and biotic controls of their distributions, particularly in relation to the different oceanographic conditions in the Gulfs of Panama and Chiriquí (Fig. 5.15). As in southern Mexico (Gulf of Tehuantepec), and the border region between Nicaragua and Costa Rica (Gulf of Papagayo), the low elevation of the Central American Cordillera in Panama,

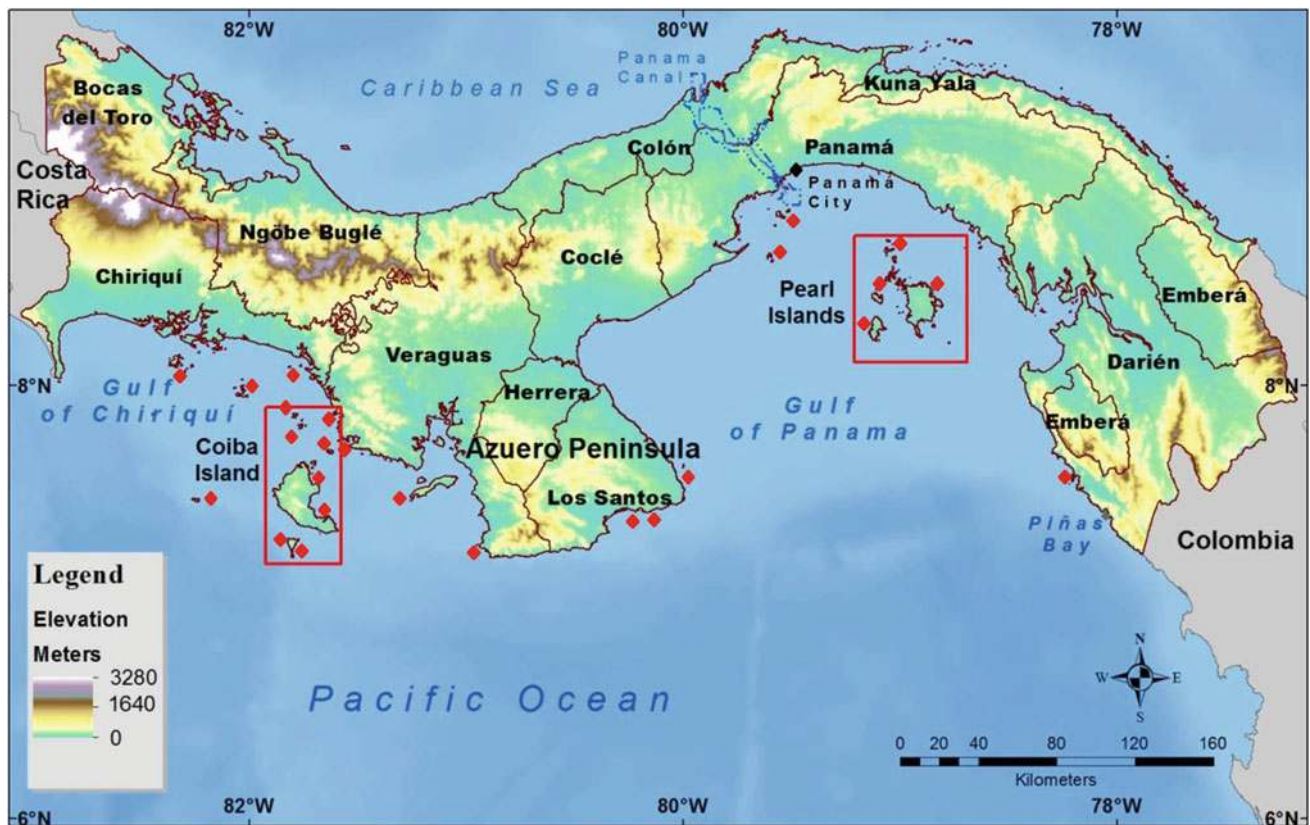


Fig. 5.15 Isthmus of Panama (PAN), framed are Las Perlas MPA, Gulf of Panama, and the Coiba National Park, Gulf of Chiriquí. Red diamonds identify sites with high coral cover. See Figs. 5.16 and 5.17 for actual MPA boundaries

at 80°W, allows seasonal (January–April) NE Trade Wind flow across the isthmus, thus causing upwelling in the Gulf of Panama and further south across the Panama Bight and along coastal Colombia. Upwelling, with accompanying low water temperatures and high nutrient conditions, promotes phytoplankton blooms that can interfere with coral growth. The high Tabasará mountain range in western Panama, reaching 3000 m elevation and more in some areas, reduces by more than two-thirds the Trade wind flow, thus preventing upwelling in the Gulf of Chiriquí, but allowing brief SST shoaling and cooling of the upper surface layer more commonly than previously suspected (D’Croz and O’Dea 2007). The extent of reef building is notable in this warm and more thermally stable enclave. Structural coral reefs are also present in the seasonally variable Gulf of Panama, but are not so well developed as in the Gulf of Chiriquí (see Chap. 6, Toth et al.). Coral community development, and the presence of coral reefs, is greatest on the numerous islands present on the isthmian shelf.

With the exception of the hydrocoral *Millepora intricata*, which often contributes importantly to coral communities and reef frameworks in the Gulf of Chiriquí (Fig. 5.3), the chief frame-building scleractinian species in both gulfs are *Pocillopora damicornis*, *Pocillopora elegans*, *Porites lobata*, *Pavona clavus*, *Pavona gigantea*, and *Gardineroseris planulata*. Zooxanthellate scleractinian species richness is virtually the same in both gulfs (Maté 2003). Three uncommon transitory species, *Leptoseris papyracea*, *Diaseris distorta* and *Cycloseris curvata*, are sometimes found in deeper (12–25 m), offreef, sedimentary habitats. A single, small, living population of *C. curvata* is presently known in the Gulf of Chiriquí; only dead skeletons of the other two species have been reported from Panama.

Like Costa Rica, the Panamanian coral fauna has remained relatively stable during the past decade, with 25 presently recognized zooxanthellate scleractinian species. Recent additions are *Porites evermanni*, *Pavona minuta*, *Pavona duerdeni*, and *Siderastrea glynni* (Fig. 5.7; Appendix). The validity of *S. glynni* is in question; possibly it is not a resident endemic, but a human-introduced Caribbean species (Forsman et al. 2005). Panama’s coral species richness is equal to that of Costa Rica, with both faunas demonstrating a strong affinity (Fig. 5.2). It is not inconceivable that the high species richness of Indo-Pacific corals in Panama, Costa Rica, and Colombia is at least in part associated with the inflow of the NECC into this equatorial region (see Chap. 3, Fiedler and Lavín). The NECC may also be responsible for the sporadic occurrence of Indo-Pacific cnidarian vagrants in equatorial eastern Pacific waters, e.g., *Porites rus* (Costa Rica), *Millepora platyphylla* (Panama), and *Acropora valida* (Colombia) as well as ephemeral occurrences of non-cnidarian invertebrate and fish

species (Glynn and Ault 2000; see Chap. 16, Lessios and Baums).

Perhaps the most influential environmental driver affecting Panama’s coral populations is ENSO. The ENSO warm phase (El Niño) in particular, but also the cool phase (La Niña) can cause coral bleaching and mortality. Mean overall coral mortality in Panama following the 1982–83 El Niño disturbance ranged from 76 to 90 % (Wellington and Glynn 2007). Coral mortality during the 1997–98 El Niño was much lower, amounting to only ~13 % in the Gulf of Chiriquí, and no detectable mortality in the Gulf of Panama. The low coral mortality in the Gulf of Chiriquí was attributed to endosymbiont resistance (see Chap. 13, Baker et al.), and the absence of coral mortality in the Gulf of Panama to upwelling, which prevented high stressful temperatures (Glynn et al. 2001).

Biotic disturbances may follow thermally-induced coral mortality. For example, bioerosion of dead corals can accelerate with increasing sea urchin population densities, which could lead to coral framework loss and the cessation of reef growth (Glynn 1988; Eakin 1992, 1996, 2001). *Acanthaster planci*, where abundant in the Gulf of Chiriquí in the 1970s and early 1980s, posed a threat to some uncommon corals that survived El Niño bleaching. However, this corallivore’s numbers decreased dramatically on reefs after the mid-1990s and is now mostly present in coral communities where it is no longer a threat (see Chap. 10, Enochs and Glynn). Past ENSO impacts can be detected from changes in coral colony morphology, increases in non-coral benthic cover, eroded reef structures, and increases in coral rubble accumulations. ENSO-related coral mortality has not been observed in Panama during the last 18 years, and significant recovery of live coral cover has occurred on reefs in the Gulfs of Panama and Chiriquí (Baker et al. 2008).

The demonstrated capacity for sustained high levels of sexual reproduction as well as asexual regeneration (Glynn and Fong 2006) has likely figured importantly in the recovery of coral communities. Continued study of coral sexual reproduction has demonstrated that the reproductive modes and spawning patterns of *Pavona clavus*, *Psammocora stellata* and *Psammocora profundacella* (Glynn et al. 2011, 2012) are similar to those of seven previously investigated species (*Pocillopora*, two spp; *Porites lobata*; agariciids, 4 spp), namely (a) broadcast spawning of minute ova at or a few days following full moon, (b) high annual fecundity due to frequent and prolonged spawning, and (c) reproduction most active during periods of high but not stressful sea water temperature (see Chap. 15, Glynn et al.). Year-round gametogenesis occurred in non-upwelling Gulf of Chiriquí, but ceased during the 3-month upwelling season in the Gulf of Panama. Gametogenesis continued during

mild El Niño warming events (Colley et al. 2006). Glynn and Colley (2008) hypothesized that broadcast spawning coral species can better tolerate and survive unstable environmental conditions in the eastern Pacific than brooding species. *Tubastraea coccinea*, a ubiquitous azooxanthellate brooder in Panama (and elsewhere in the eastern Pacific), releases planulae during new and full lunar phases, mostly during high thermal periods (Glynn et al. 2008). This species also is highly fecund, and is reproductively mature when colonies possess as few as 2 to <10 polyps.

During the past decade (early to mid 2000s) detailed mapping of the presence and abundances of coral and octocoral communities has been undertaken in two of

Panama's largest MPAs, the Pearl Islands (Gulf of Panama), and Coiba Island (Gulf of Chiriquí) and surroundings (Guzmán et al. 2004, 2008). [As a cautionary note, the live coral cover on these maps (1) includes corals and octocorals, (2) none of the islands supports a continuous fringing reef, and (3) coral cover on the relatively small structural reefs (not shown) often exceeds 40 %. The reader is referred to Guzmán et al. (2004, 2008) for details on the sampling methodology.] Moderate to high coral cover, 20 to >40 %, was reported in both of these archipelagos (Figs. 5.16 and 5.17) with the highest coral cover recorded on the smaller islands (Table 5.5). The high coral cover category in the Coiba Islands (18.4 %) was slightly over twice as high as

Fig. 5.16 Live coral cover in Pearl Islands MPA (168,800 ha), Gulf of Panama, Panama. The ~7 km wide Trollope Bank is shown to the SE of El Rey Island. MPA boundaries in blue. Modified after Guzmán et al. (2008)

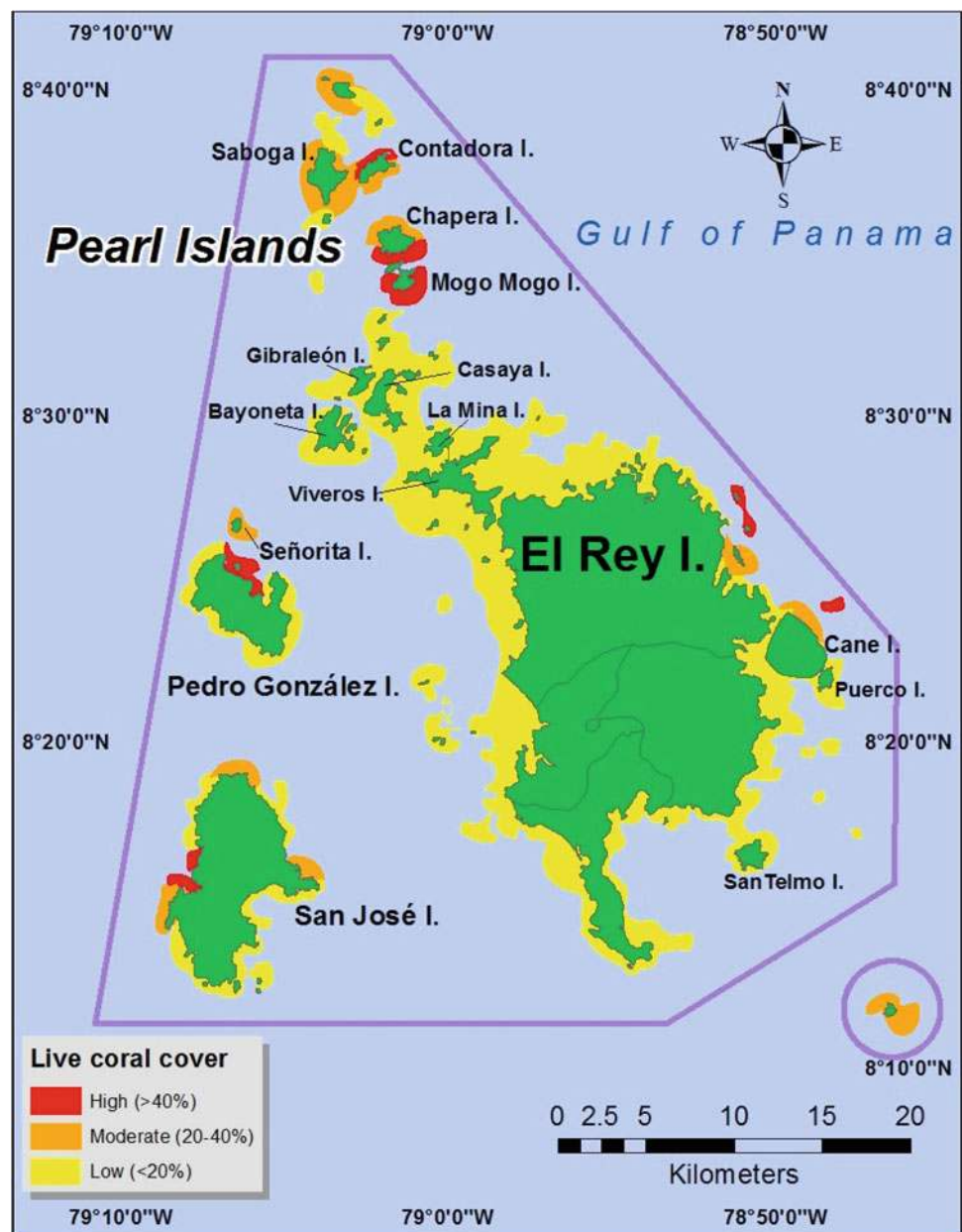
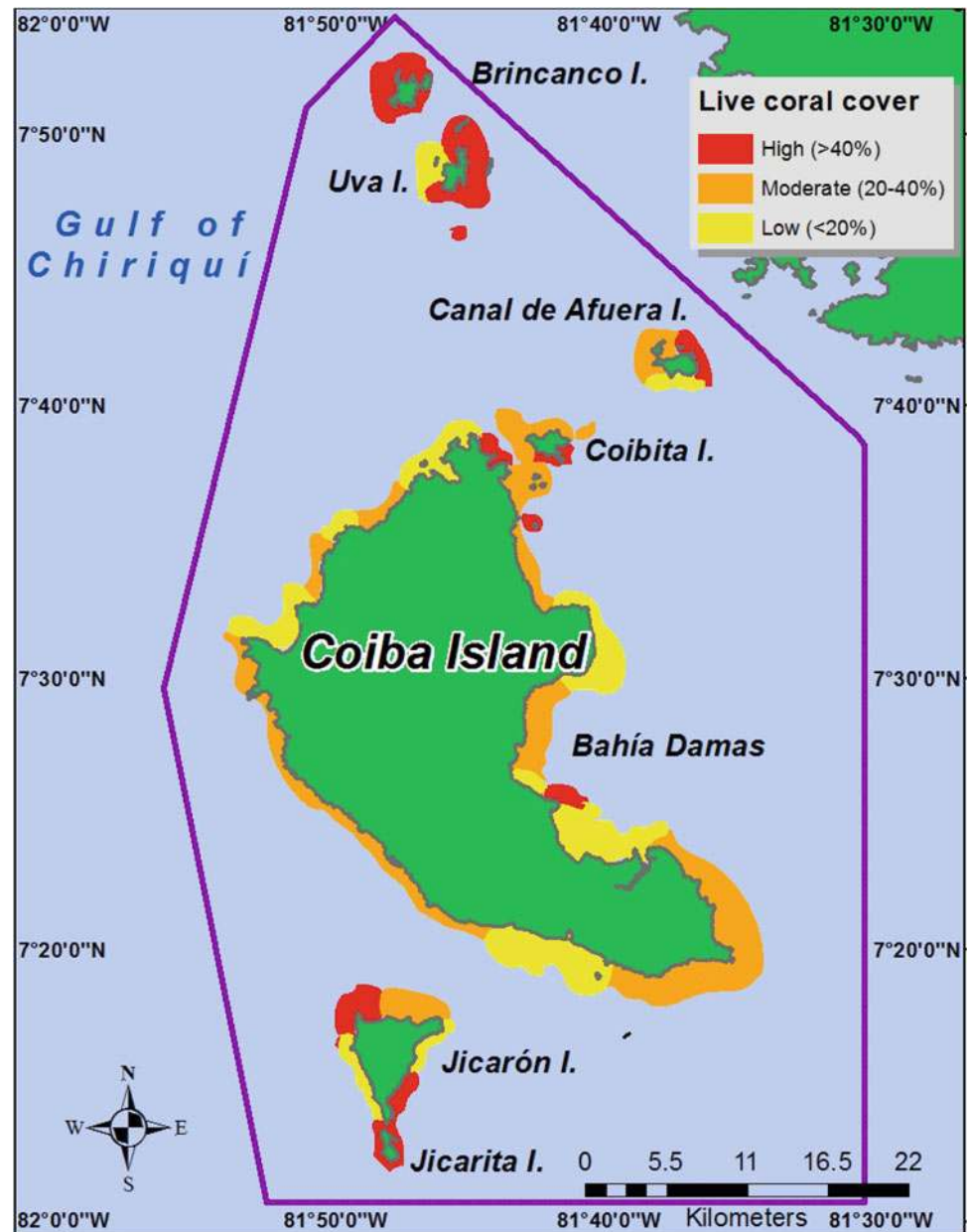


Fig. 5.17 Live coral cover in Coiba MPA (270,125 ha), Gulf of Chiriquí, Panama. MPA boundary in *blue*. Modified after Guzmán et al. (2004)



that recorded in the Pearl Islands (8.1 %). Nearly 80% of the surveyed areas in the Pearl Islands supported low coral cover (<20 %) compared with ~35 % in the Coiba Islands. It is necessary to consider at least three factors contributing to this difference: (a) deforestation, land use and sedimentation, all of which have been greatest in the Pearl Islands, (b) upwelling versus non-upwelling environments, the latter generally promoting more robust coral growth, and (c) phytoplankton blooms causing copious mucus production that can smother corals (Guzmán et al. 1990).

In these island-wide surveys a distinction was made between coral communities (CC) and coral reefs (CR), which revealed some intriguing differences: (a) scleractinian

corals and octocorals demonstrated higher species richness in CC compared to CR, (b) coral cover was higher in CC than CR in the Coiba Islands preserve, but higher in CR than CC in the Pearl Islands, (c) uncommon, rare or endangered cnidarians were encountered more frequently in CC than CR, and (d) a negative relationship between live coral cover and species richness emerged in the Coiba Islands, with CC consisting of a greater variety of cnidarian species than CR. A similar trend to that in 'a' above was reported by Benfield et al. (2008) for reef fish assemblages in the Pearl Islands, which were significantly more diverse and species rich in CC than CR. These results prompted Guzmán et al. (2004, 2008) to hypothesize that CC may serve as refugia and

Table 5.5 Live coral cover at various islands surveyed at Coiba and Pearl Islands MPAs

Location	Area of live coral cover (km ²)		
	High (>40 %)	Moderate (20–40 %)	Low (<20 %)
Coiba Islands			
Brincanco	12.237	–	–
Uva	14.587	–	4.747
Canal de Afuera	6.377	7.822	4.951
Coibita	2.384	7.829	–
Coiba	9.531	135.693	93.053
Jicarón	13.532	7.156	13.707
Jicarita	3.227	–	–
Total = 336.84	61.88 (18.4 %)	158.50 (47.1 %)	116.46 (34.6 %)
Pearl Islands			
Pacheca	–	3.261	1.260
Saboga	–	5.037	1.359
Contadora	1.387	2.133	–
Chapera	2.418	1.432	–
Mogo Mogo	4.500	–	–
Gibrleon	–	–	5.898
Bayoneta	–	–	6.947
Casaya	–	–	12.248
La Mina	–	–	7.120
Viveros	–	–	37.279
Señorita	–	2.168	–
Pedro González	3.477	–	9.725
San José	2.920	5.110	13.100
San Telmo	–	–	3.131
Puerco	–	–	3.336
Cane	–	1.560	3.600
El Rey	–	1.800	38.320
Total = 180.52	14.70 (8.1 %)	22.50 (12.5 %)	143.32 (79.4 %)

Estimated areas digitized using Arc Map 10.1 software from Guzmán et al. (2004, 2008)

source areas for larvae that could repopulate CR that become degraded during large-scale disturbances such as El Niño bleaching/mortality events.

Building on Abele's (1976) demonstration of a species-rich community of decapod crustaceans associated with living pocilloporid corals (55 species), several recent studies have revealed an impressive number of motile, cryptic invertebrates and fishes that seek refuge in living and dead coral frameworks and rubble. For example, the biomass and abundances of various invertebrate taxa (Polychaeta,

Gastropoda, Crustacea, Holothuroidea, Ophiuroidea) associated with live corals are generally higher than those sheltering among dead corals and coral rubble (Enochs and Hockensmith 2009; Enochs 2012). Species richness, however, is significantly higher in dead and degraded coral rubble (261–370 species) compared with live coral habitats (112–219). Among some of the several key reasons for this are (a) the inimical defenses of live corals (nematocysts, mucus secretions), (b) the agonistic and competitive nature of obligate coral metazoan symbionts, and (c) the greater habitat heterogeneity and niche diversity of rubble substrates. These results show that an abundant and speciose cryptic fauna is still present on highly degraded reefs that can continue to function in various reef processes, e.g. providing prey for reef fishes, scavenging, biodegradation, remineralization, bioerosion, interalia (Enochs et al. 2011; Enochs 2012; Enochs and Manzello 2012a, b).

Recent studies of the diversity, community structure and trophic relationships of coral-associated fishes have increased our understanding of their diverse and pivotal ecological roles in the ETP (Dominici-Arosemena and Wolff 2006; Benfield et al. 2008; Glynn 2008; Robertson and Allen 2008; Glynn et al. 2014). A large number of facultative corallivores, and cryptic fishes residing in coral rubble, many of the latter invertivore and piscivore predators, have been identified (Glynn 2006, 2008). Some of these taxa may play critical roles in limiting the settlement and survival of recruiting species.

5.3.10 Mainland Colombia

In spite of the adverse climatic and oceanographic conditions (Glynn and Ault 2000; Zapata and Vargas-Ángel 2003), isolated colonies of scleractinian corals occur along most of the 1450 km of the Colombian Pacific coast. Corals are most common in the northern half of the coast from the Panama-Colombia border south to Cabo Corrientes, a stretch dominated by igneous rocky shores. South of Cabo Corrientes to the Ecuador-Colombia border the coast is composed of alluvial sediments and dominated by soft bottoms and mangroves intermixed with some stretches of soft sedimentary rock from the Tertiary. The Colombian Pacific coastal region is one of the rainiest regions of the world, with annual precipitation exceeding 10,000 mm at some localities. High freshwater discharge from the western slopes of the Andes and the adjacent lowlands into the Pacific Ocean significantly reduces salinity and increases turbidity and sedimentation, particularly in the southern half of the region (Restrepo and Kjerfve 2000; Alory et al. 2012). Yet, the best developed coral reefs and coral communities of the Colombian Pacific are located in this southern portion at Gorgona Island, which is adjacent to this stretch of coastline,

but located sufficiently far offshore to reduce the limiting influence of the coastal environment. Accordingly, Kleypas et al. (1999) listed Gorgona Island and Ensenada de Utría, also in Colombia (see below), as the two lowest salinity localities with coral reefs in the eastern Pacific. Therefore, coral communities and true reefs (carbonate frameworks) are few, small, patchily distributed and poorly developed. Consequently, coral communities and reefs in this area of the eastern Pacific are species-poor (≤ 12 scleractinian coral species per site) and the entire coral fauna is composed of about 28 species, although taxonomic difficulties, particularly within the genus *Pocillopora*, make this number somewhat uncertain (Table 5.6).

Four localities along Colombian Pacific shores have been reported to have significant coral communities or coral reefs: Cabo Marzo and Punta Cruces, Punta Tebada, Ensenada de Utría, and Gorgona Island (Fig. 5.18). The northernmost area is a prominent rocky point located at Cabo Marzo (Fig. 5.19) where a significant coral formation occurs. Known locally as El Acuario, the shallow (≤ 3 m) substrate here is composed of a solid rock bed with boulders dominated by filamentous algae (47 % mean cover), scleractinian corals (33 % mean cover) and octocorals. Branching corals of the genus *Pocillopora* are the most common (27 % mean cover) followed by massive colonies of *Pavona gigantea*. There is, however, little indication of significant reef framework development at this site. Six species of zooxanthellate scleractinian corals occur at El Acuario (Zapata et al. 2008; Table 5.6). A small coral patch consisting of sparse *Pocillopora damicornis* colonies and associated rubble is also located in neighboring Bahía Aguacate. Further south at Punta Cruces, in a cove in front of the hamlet known as Piñas (Fig. 5.19), there also is a coral community dominated by massive and encrusting corals covering nearly 38 % of the rocky substrate.

Coral assemblages characterized by high colony densities, but lacking a framework buildup are common in the Utría and Tebada region. Largely, these communities develop on rocky banks, shoals, and vertical walls, particularly in small embayments and coves, ranging in depth from about 1 to 10 m; moderately strong swells and vigorous water circulation are prevalent in these habitats, however. Despite the high energy of the environment, the dominant taxa are branching *Pocillopora* species, particularly *P. elegans*, and massive colonies of *Porites lobata*, *Pavona clavus*, and *Pavona gigantea*; less abundant to rare are *Gardineroseris planulata*, *Pocillopora damicornis*, *Psammocora stellata*, and *Pavona* sp. aff. *frondifera* (Vargas-Ángel 2001, unpublished data). Together with the scleractinian corals, coralline and turf algae, and other sessile invertebrates including non-zooxanthellate corals, barnacles, sea fans, and clams, inhabit these shallow basalt substrates. Generally high levels of grazing from sea urchins and herbivorous fishes are in part responsible for the limited

macroalgal development in these communities. Coral recruitment from sexual propagules or fragmentation is also noticeably low on both the grazed basalt surfaces or the rubble and sand-covered bottom surrounding corals.

Of the coastal coral reefs, La Chola (also known as La Aguada) fringing reef, located at Ensenada de Utría (Fig. 5.18), is the largest, best developed, and probably the most thoroughly studied in this part of Colombia. The reef is located on the central, eastern shoreline of a narrow embayment, characterized by calm and relatively clear waters. Elevated turbidity and runoff events following heavy rainfall, which is common in this region, can reduce visibility to about 1.0–1.5 m, demonstrating that these corals are able to tolerate periods of elevated turbidity and reduced salinity. Thinly-branched ecomorphs of *Pocillopora* are prevalent on the reef, which contrast with the more stout and stubby congeneric colonies developing in the non-framework building communities of outer wave-exposed habitats. When the reef was surveyed in 1988–89 by Vargas-Ángel (1996), the predominant coral species were *Pocillopora damicornis* (80 %) and *Psammocora stellata* (16 %), which formed a terrace-like framework, covering approximately 10.5 ha; uncommon to rare species included *Pocillopora capitata*, *Pavona varians*, and *Pavona gigantea*. The reef at Utría lacks the typical geomorphological zones (i.e., backreef, reef flat, forereef); it is a relatively broad reef flat followed by a seaward slope, where coral development gradually decreases to a depth of approximately 10 m. Reef coring and probing revealed that the uncemented *Pocillopora* framework was at least 2.0–4.5 m in height, with uninterrupted growth throughout the last 2000–3000 years, and mean vertical accretion rates of $2.47 \text{ m kiloyear}^{-1} \pm 0.15$ (SE) (Vargas-Ángel 2001; see Chap. 6, Toth et al.). Community structure surveys conducted later in 1996–98 (Vargas-Ángel 2001) indicated that mean percent live coral cover was 41 %. A temporal comparison of permanent transects placed since 2002 indicates that coral cover on this reef has declined on average by 65 % from 57 % in 2002 to 37 % in 2013 (M. López-Victoria and F. A. Zapata, unpublished data). In all of these surveys, coral composition was relatively homogeneous throughout the shallow reef flat, with intermingled turf algal patches and pockets of rubble and bioclastic sand. Absence of ecological zonation has been regarded as an indicator of “instability”, whereby chronic stress and environmental disturbances may offset the competitive interactions and biological processes leading to species habitat partitioning (Karlson 1999). In addition to chronic siltation and turbidity stress, this reef is subject to subaerial exposure during extreme low tides, recurrent ENSO events, and periodic intense upwelling episodes (Vargas-Ángel 2001; Zapata and Vargas-Ángel 2003).

Located approximately 70 km north of Utría, the Tebada reef is a small structure, covering roughly 4.5 ha, and situated in the southern end of the Gulf of Cupica. When

Table 5.6 Known zooxanthellate scleractinian coral species from the Colombian Pacific Ocean

Species	Malpelo	Gorgona	Ensenada de Utría	Tebada	Bahía Cupica	Cabo Marzo	Comments
F. ACROPORIDAE							
<i>Acropora valida</i> (Dana, 1846)		4, 5, 8					
F. FUNGIIDAE							
<i>Cycloseris curvata</i> (Hoeksema, 1989)		2, 5, 8					
<i>Diaseris distorta</i> (Michelin, 1843)			12				Only dead fragments found outside reefs
F. AGARICIIDAE							
<i>Gardineroseris planulata</i> (Dana, 1846)	1, 5, 8	2, 5, 9	10	12	12		
<i>Leptoseris papyracea</i> (Dana, 1846)		5					
<i>Pavona chiriquiensis</i> Glynn et al., 2001	11, 13	13					
<i>Pavona clavus</i> (Dana, 1846)	1, 5, 8	2, 3, 5, 8	5, 6, 10		12	14	
<i>Pavona frondifera</i> (Lamarck, 1816)		14			17		
<i>Pavona gigantea</i> Verrill, 1869	5, 8	2, 3, 5, 8	5, 6, 10	12, 17	12	14	
<i>Pavona maldivensis</i> (Gardiner, 1905)	18, 14						
<i>Pavona varians</i> Verrill, 1864	1, 5, 8	2, 3, 5, 8	5, 6, 10	12, 17	12		
F. POCILLOPORIDAE							
<i>Pocillopora capitata</i> Verrill, 1864	1, 8	2, 3, 5, 8	5, 6, 10	12		14	
<i>Pocillopora damicornis</i> (Linnaeus, 1758)		2, 3, 5, 8	5, 6, 10	12, 17	12	14	
<i>Pocillopora elegans</i> Dana, 1846	5	2, 5	5, 6, 10		12		
<i>Pocillopora eydouxi</i> Milne Edwards & Haime, 1860	8	2, 3, 5, 8	10			14	
<i>Pocillopora verrucosa</i> (Ellis and Solander, 1786)	20	20					
<i>Pocillopora meandrina</i> Dana, 1846		20					
<i>Pocillopora effusus</i> Veron, 2002	20	20					
<i>Pocillopora ligulata</i> Dana, 1846		20					
<i>Pocillopora woodjonesi</i> Vaughan, 1918		20					
<i>Pocillopora</i> sp.	15						
F. PORITIDAE							
<i>Porites evermanni</i> Vaughan, 1907		16					Inferred from results in Boulay et al. (2014), which indicate that ~50 % of massive <i>Porites</i> in coastal Ecuador and Panamá are <i>P. evermanni</i>
<i>Porites lobata</i> Dana, 1846	1, 5, 8	2, 3, 5, 8	10	12,17	12		
<i>Porites panamensis</i> Verrill, 1866		2, 3, 5, 8	5, 6			14	
F. SIDERASTREIDAE							
<i>Psammocora contigua</i> (Esper, 1797)		9, 19					Based on <i>Psammocora obtusangula</i> (Lamarck, 1816) as listed by Zapata and Vargas-Ángel (2003). <i>P. obtusangula</i> was synonymized with <i>P. contigua</i> (Esper, 1794) by Stefani et al. (2008)

(continued)

Table 5.6 (continued)

Species	Malpelo	Gorgona	Ensenada de Utría	Tebada	Bahía Cupica	Cabo Marzo	Comments
<i>Psammocora profundacella</i> Gardiner, 1898		9, 19		12			Based on <i>Psammocora superficialis</i> , Gardiner, 1898, as listed by Zapata and Vargas-Ángel (2003). <i>P. superficialis</i> was synonymized with <i>P. profundacella</i> Gardiner, 1898 by Benzoni et al. (2010)
<i>Psammocora stellata</i> Verrill, 1864		2, 3, 5, 8	5, 6, 10	12, 17	12		
Total number species = 27	13	24	12	8	9	6	

Numbers indicate information sources as indicated in the references and sources section. List prepared by Fernando A. Zapata with contributions by Bernardo Vargas-Ángel for Ensenada de Utría, Tebada and Bahía Cupica. 9 February 2014

1. Birkeland et al. (1975)
 2. Glynn et al. (1982)
 3. Cantera (1983)
 4. von Prael and Mejía (1985)
 5. von Prael and Erhardt (1985)
 6. von Prael (1986a)
 7. von Prael (1986b)
 8. von Prael (1990)
 9. Guzmán and Cortés (1993)
 10. Vargas-Ángel (1996)
 11. Garzón-Ferreira and Pinzón (1999)
 12. Vargas-Ángel (2001)
 13. Zapata et al. (2007)
 14. Zapata et al. (2008)
 15. Rodríguez-Ramírez and Zapata (2012)
 16. Boulay et al. (2014)
 17. B. Vargas-Ángel, personal observation, unpublished data
 18. J. Garzón-Ferreira, photographic record
 19. H.M. Guzmán, personal communication
 20. F.A. Zapata, personal observation
- Other references
- Stefani et al. (2008)
- Benzoni et al. (2010)

surveys were conducted in 1996–1998 mean live coral cover was 39 %. At that time the living coral community was dominated by a mixture of *Psammocora stellata* and *Psammocora profundacella*, which together amounted to nearly 90 % of the live reef cover (Vargas-Ángel 2001), in contrast with the reefs farther south at Ensenada de Utría and Gorgona Island, which were dominated by pocilloporid species (Zapata and Vargas-Ángel 2003). Four more coral species were present at Tebada than further south, including a few small colonies of *Pocillopora damicornis*, *Pavona varians*, *Pavona gigantea*, and *Gardineroseris planulata*; the latter is relatively uncommon on coastal reefs and in coral communities. The evident structural pattern of the Tebada reef was an increase of live coral cover with increasing distance from shore, related to a decrease in scleractinian species richness (Vargas-Ángel 2001, 2003). Reef probing on the reef flat in 1996 revealed that the Tebada reef framework was at least 4 m thick, and coring to

a depth of approximately 1 m revealed an uncemented framework composed mainly of *Psammocora* rubble (Vargas-Ángel, unpublished data). Based on its geographic location, it is reasonable to assume that the Tebada reef is subject to similar environmental stressors as those at Utría, particularly ENSO and upwelling stress events. The reef flat at Tebada, however, is approximately 1 m deeper than that at Utría; therefore, it is not subject to subaerial exposure during extreme low tides. Recent inspection of this site in April 2014 revealed that both *Pocillopora* and *Psammocora* were important members of this coral formation and that framework construction was modest (C.G. Muñoz and M. Rodríguez-Moreno, personal communication). While Vargas-Ángel et al. (2001) reported minimal coral bleaching at this reef during June 1998, local fishermen recalled that noticeable coral bleaching occurred here at some time during the 1997–1998 El Niño event.

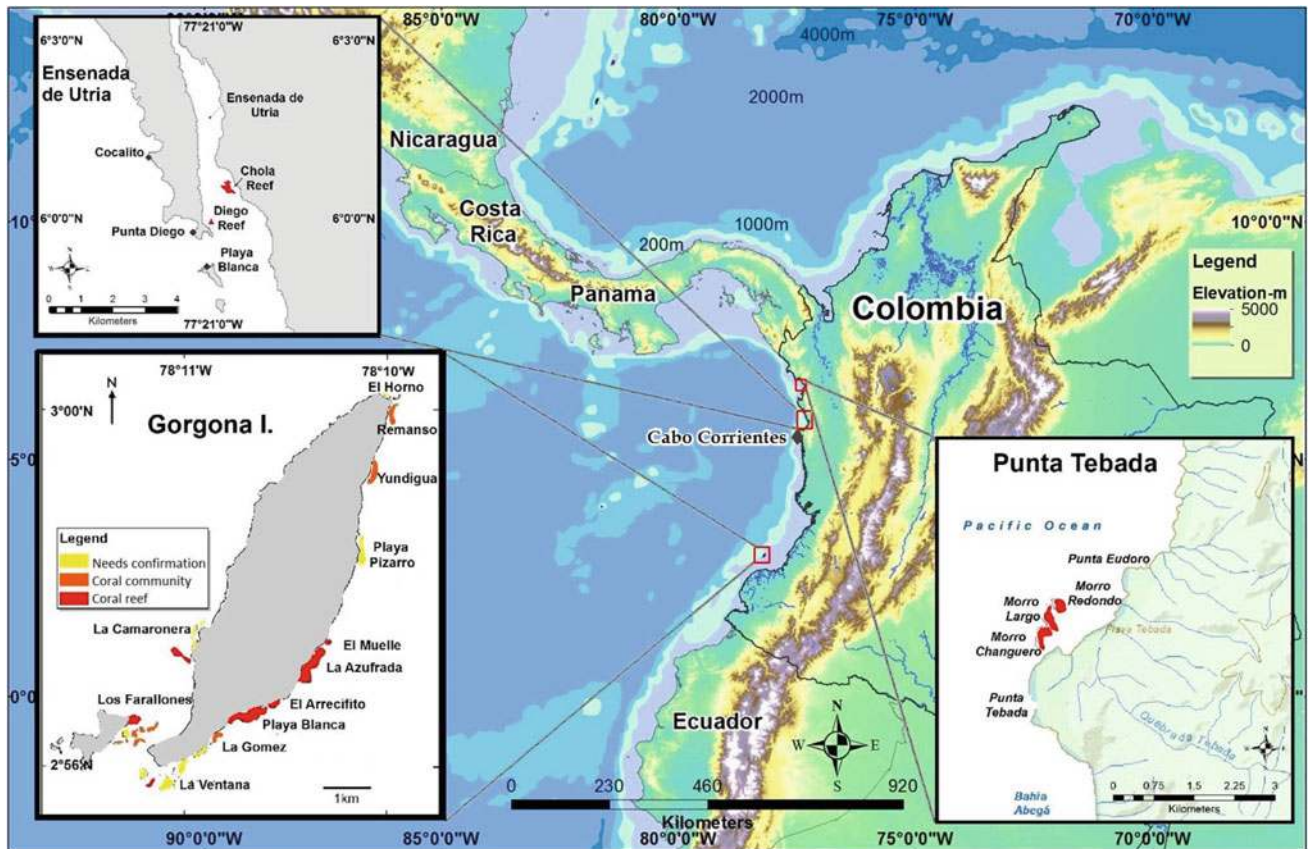


Fig. 5.18 Coral communities and coral reefs at Punta Tebada, Ensenada de Utria and Gorgona Island, Pacific Colombian (COL) coast. Red patches at Punta Tebada and Ensenada de Utria

denote high coral cover. Gorgona Is legend: red = structural coral reef (arrecife); orange = coral community (comunidad coralina); yellow = coral reef/community in need of confirmation (por confirmar)

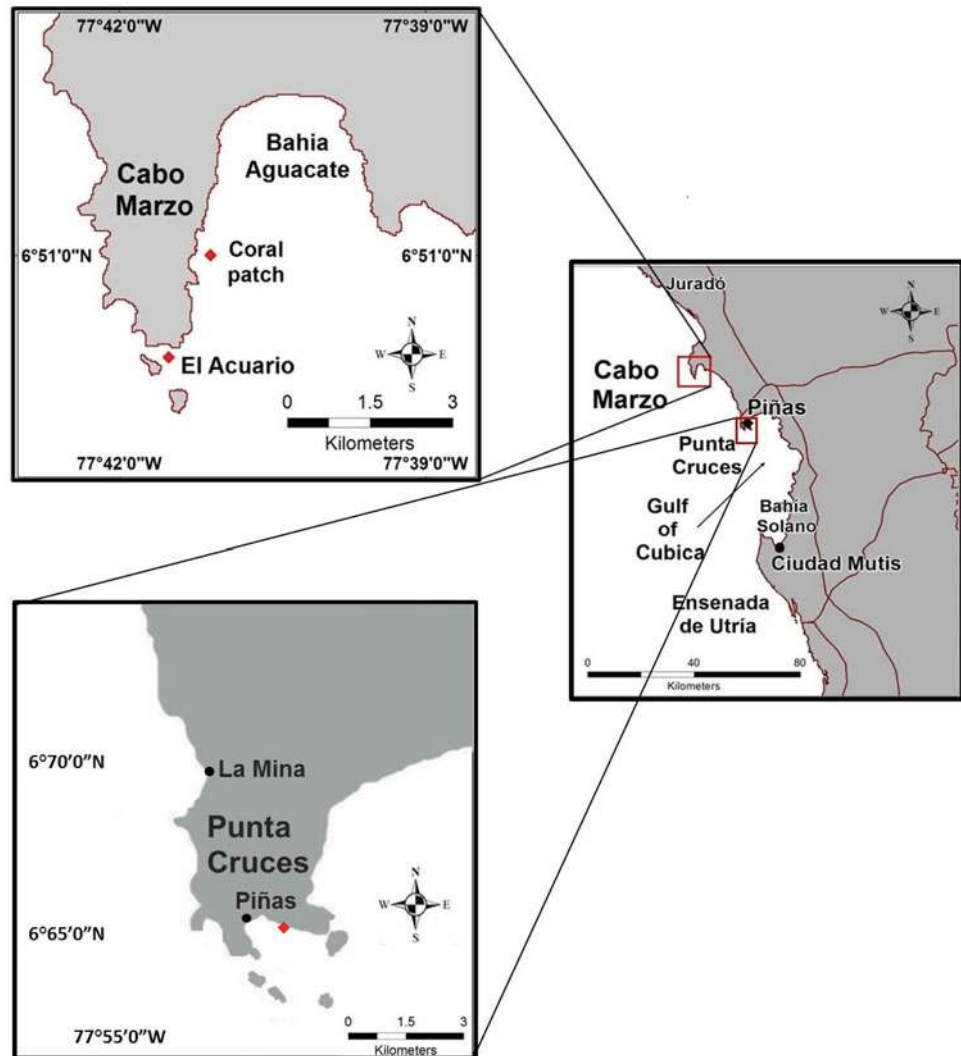
The coral reefs and coral communities of Gorgona Island are the largest and best developed in the Colombian Pacific region (Glynn et al. 1982; Zapata and Vargas-Ángel 2003). The largest structural coral reefs of Gorgona Island, La Azufrada and Playa Blanca reefs, are located on the eastern, leeward side of the island (Fig. 5.18). These reefs exhibit high coral cover, are more or less similar in size (~10 ha), structure and zonation, and are composed mainly of pocilloporid species with a significant presence of massive corals of the genera *Pavona* and *Gardineroseris* at the seaward reef base. A total of 24 species has been recorded for Gorgona Island (Table 5.6), but some of these records may have to be confirmed with more detailed work. The report of *Acropora valida* at Gorgona Island, a species known previously from the western Pacific but known today to be widespread across the Indo-Pacific region (Veron 2000), is intriguing: three colonies were found at 10 m depth on a rocky outcrop at the northernmost tip of Gorgona Island in 1983 (von Prahll and Mejía 1985). While the biogeographical implications of this report were potentially major given competing hypotheses regarding the origin of the eastern Pacific coral fauna (Dana 1975; Heck and McCoy 1978), despite repeated efforts to find new specimens, this species has never again been

observed at Gorgona Island or any other locality in the eastern Pacific.

Live coral and algal cover on these reefs appear to be highly dynamic, particularly on shallow areas due to recurrent subaerial exposures during extreme low tides (Zapata et al. 2010). While recurrent disturbances on the reef flat can at times reduce live coral cover to very low levels and increase algal turfs, high densities of herbivorous sea urchins and fishes readily remove filamentous turf algae leaving an exposed reef matrix composed of coral rubble cemented by calcareous algae that sexually-produced coral recruits eventually colonize. Coral recovery is further enhanced by asexual recruitment via colony fragmentation, which is very common and facilitated by the feeding activities of some fishes, particularly triggerfishes (Balistidae) and puffers (Tetraodontidae) (Palacios et al. 2014).

Coral formations of Gorgona Island have been studied since the 1970s and have been described in several accounts (von Prahll et al. 1979; Glynn et al. 1982; von Prahll and Erhardt 1985; von Prahll 1986a, b; Zapata 2001; Zapata et al. 2001; Zapata and Vargas-Ángel 2003). Nonetheless, significant coral formations continue to be found around the island. While Glynn et al. (1982) had previously described

Fig. 5.19 Cabo Marzo and Punta Cruces, northern-most known coral communities on coastal Colombia



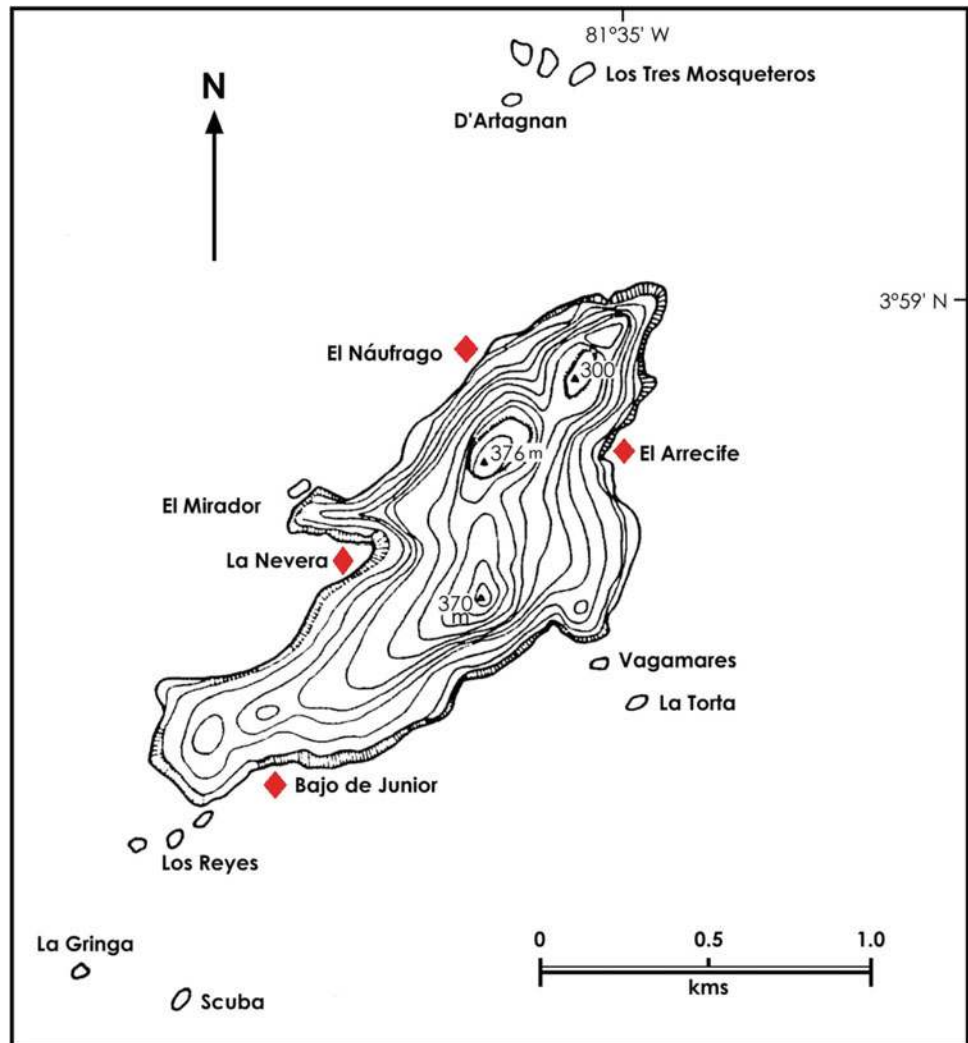
an incipient reef formation at La Camaronera, located on the western, wave-exposed coast, as “linear pocilloporid build-ups or spurs and hillocks of various sizes”, a different formation was found further offshore. This is a coral community that extends 8 ha and is made up of 11 species of corals, which cover 49 % of the substrate. Massive species of the genera *Pavona*, *Gardineroseris* and *Porites* are dominant, forming high topographic relief that provides habitat for a distinct fish community (Palacios and Zapata 2014). This coral formation contrasts with the incipient coral reef of La Ventana, also mentioned briefly by Glynn et al. (1982). This is a pocilloporid reef located on the south-eastern side of the island that remained undescribed until recently. While this is a small reef covering 1 ha, it exhibits very high live coral cover (73 %) because its low-lying reef framework is never subaerially exposed during extreme low tides. However, this dense pocilloporid stand provides low relief and constitutes a relatively monotonous habitat for a distinct but less equitable fish

community than that observed at La Camaronera (Palacios and Zapata 2014).

5.3.11 Malpelo Island (Colombia)

Malpelo Island, the easternmost and least remote of the five oceanic islands or archipelagos in the ETP, is a very small (1.6 km long) volcanic island located nearly 380 km off the Colombian continental coast, 630 km from Isla del Coco and 996 km from San Cristóbal, the nearest of the Galápagos Islands (Fig. 5.20). Malpelo’s marine environment is most similar to that of its closest oceanic neighbor, Isla del Coco, but in contrast to the latter, only a total of 13 zooxanthellate scleractinian species has been recorded at Malpelo (Table 5.6). A particularly interesting and possibly new undescribed species of *Pocillopora*, which exhibits an unusual morphology with a tightly submassive, hemispherical corallum (Fig. 5.21), was reported by Rodríguez-Ramírez

Fig. 5.20 Malpelo Island (MAL), Colombia. *Red diamonds* identify best-developed coral communities



and Zapata (2012). This rare coral, if a valid new species, would represent a possible endemic species for the island. Nonetheless, in terms of coral species composition, Malpelo is more similar to continental localities in El Salvador and Nicaragua than to the neighboring oceanic islands of Isla del Coco or the Galápagos (Fig. 5.2). However, this may be a result of poor species richness at Malpelo, El Salvador and Nicaragua rather than a reflection of similar environments with similar faunas. This interpretation is supported by results of similar analyses with the fish fauna in which oceanic islands in general, and Isla del Coco, Isla Malpelo and the Galápagos Islands in particular, were always most similar in species composition, and distinct from continental localities (Robertson and Cramer 2009).

While oceanographic conditions appear to be favorable for vigorous coral growth (but see below), steep rocky walls prevent the buildup of significant reef structures, and corals often exhibit unusual growth forms. For instance, some pocilloporids display horizontally-oriented colony growth with flattened branches, likely the result of strong wave

action (Fig. 5.22), while *Porites lobata* very commonly forms shingle-like structures on steeply inclined substrates (Fig. 5.23). Besides small, isolated coral patches and scattered coral colonies of *Pocillopora* spp. and *Porites lobata* along the rocky sublittoral, several sites around the island harbor coral communities, the largest of which is known as El Arrecife (The Reef). This community demonstrates a marked zonation pattern with pocilloporid colonies in shallow areas, *Porites lobata* and *Pavona clavus* at mid-depths and *Gardineroseris planulata* in the deepest areas. Strong oceanic swells and occasional rockfalls appear to upturn and break coral colonies affecting the development of coral formations around the island (Birkeland et al. 1975; Garzón-Ferreira and Pinzón 1999). Clear evidence of the effects of an unknown major disturbance was observed in July 2011 at El Arrecife, suggesting that the lack of reef building at this site may be due to recurrent sediment export during strong turbulence. Bastidas-Salamanca et al. (2014) reported a reduction in coral cover from ~45 % in 2011 to ~35 % in 2012 at El Arrecife and speculated that this may have been



Fig. 5.21 *Pocillopora* sp., possibly an undescribed species, El Arrecife, Malpelo Island, 11 m depth, 24 June 2007. Colony in foreground, approximately 0.5 m in diameter

caused by the mechanical action of a tsunami-like disturbance during a seismic event. However, the noticeable increase in variance around the mean coral cover values reported in 2012 does not lend support to this hypothesis because a major wave would probably have caused more uniform damage and suggests instead the action of a much more localized factor that affected some sites more than others.

Like other localities in the eastern Pacific in general and within Panama Bight in particular, Malpelo Island is exposed to high thermal variability. First, there is marked interannual thermal variability associated with ENSO events that occur at irregular intervals every few years. However, little is known about the effects of the 1982–83 El Niño warming event on Malpelo’s coral formations, although they may have been affected as much as other reefs in the region (Glynn et al. 1988). During the 1997–98 El Niño, coral bleaching and mortality were low at Malpelo Island, but such effects were also relatively low everywhere else in the Colombian Pacific compared to levels of bleaching observed in 1982–83 (Vargas-Ángel et al. 2001). Second, there is a markedly seasonal annual cycle with lower and more variable water temperature between January and March than

during the rest of the year, as in much of Panama Bight (Forsberg 1969; Stevenson et al. 1970; D’Croze and Robertson 1997; Zapata et al. 2011). During this period, the Intertropical Convergence Zone migrates from $\sim 8\text{--}10^\circ\text{N}$ to $\sim 2^\circ\text{N}$ and the north-east trade winds blow more strongly over the bight, producing a cyclonic pattern of water circulation and cold water upwelling in the middle of the bight, with a consequent shallowing of the thermocline (Rodríguez-Rubio et al. 2003; Devis-Morales et al. 2008). While cooling episodes may be common at Malpelo Island, in April 2009 an unusually cold event caused extensive bleaching of *Porites lobata* and several other coral species (Zapata et al. 2011). Thus, coral formations at Malpelo Island may also be limited by extreme temperature variations over several temporal scales.

5.3.12 Mainland Ecuador

Coral reef formations of relatively high diversity occur along the mainland coast of Ecuador from Esmeraldas to Santa Elena. At Esmeraldas, (El Faro point) near Cabo San Francisco, small patches of *Porites lobata* are present in the low



Fig. 5.22 *Pocillopora effusus*, horizontal and flattened growth form, 15 m depth, 24 June 2007, Malpelo Island (photo by F.A. Zapata)

intertidal zone. A *P. lobata* patch reef, about 1250 m² between 15 to 20 m depth, is present on the Surrones Bank, located approximately 6 km north of Pedernales (Fig. 5.24). Another small *P. lobata* patch reef occurs at 7 m depth at Ligüique in the Pacoche Wild Life Refuge. Coral diversity and reef development are greatest in the Machalilla National Park (Fig. 5.24). The Machalilla MPA lies within the coastal stretch from 2° 15'S to 1°00'N (Glynn 2003). A fringing reef located near the fishing village of Machalilla, at 1°28'S, is the southern-most known coral reef on the American Pacific coast. A mixed and diverse octocoral/coral community is present at Isote los Ahorcados, just south of the Machalilla reef. *Pavona clavus*, *Pavona gigantea*, *Pocillopora verrucosa*, and *Pocillopora damicornis* are the chief corals at this site. Small pocilloporid patches are also present at Pelado Islet, a new MPA located about 40 km south of the Machalilla MPA. And most recently (personal communication, 2015) Jesenia Zambrano has reported the presence of *Pocillopora* in the Puntilla Santa Elena MPA (see Fig. 21.7 in Chap. 21, Alvarado et al.).

Over the last decade, coral species richness has increased from 18 to 23 species (Fig. 5.6; Appendix). This increase is due to the recognition of four additional species of

Pocillopora and *Porites evermanni*. These new records probably represent long-term resident species and not recent arrivals. The relatively high species count of 23 for coastal Ecuador and no authenticated coral records for Peru underlines the limiting influence of the cool Peru Coastal Current, which reaches Talara just south of the Gulf of Guayaquil (Fig. 5.1; see Chap. 3, Fiedler and Levín). Pocilloporid corals have been reported (René Espinoza, personal communication), but unconfirmed, at Santa Clara Island, near 3° S, at the entrance to the Gulf of Guayaquil. The species composition of the coral fauna of mainland Ecuador is most similar to that of the southern Galápagos Islands (Fig. 5.2).

As generally observed across the equatorial eastern Pacific, coastal Ecuadorian coral communities were severely impacted by El Niño-induced bleaching in 1982–83 and 1997–98 (Glynn 2003). Due to the remote locations of the coastal coral sites it has not been possible to monitor community responses or recovery after these disturbances. Thanks to the comprehensive cnidarian surveys and inventories of two MPAs compiled by Rivera and Martínez (2011), with the collaboration of Odalisca Breedy, it is possible to infer recovery since 1997–98, which appears to have been significant and widespread. Their photographic

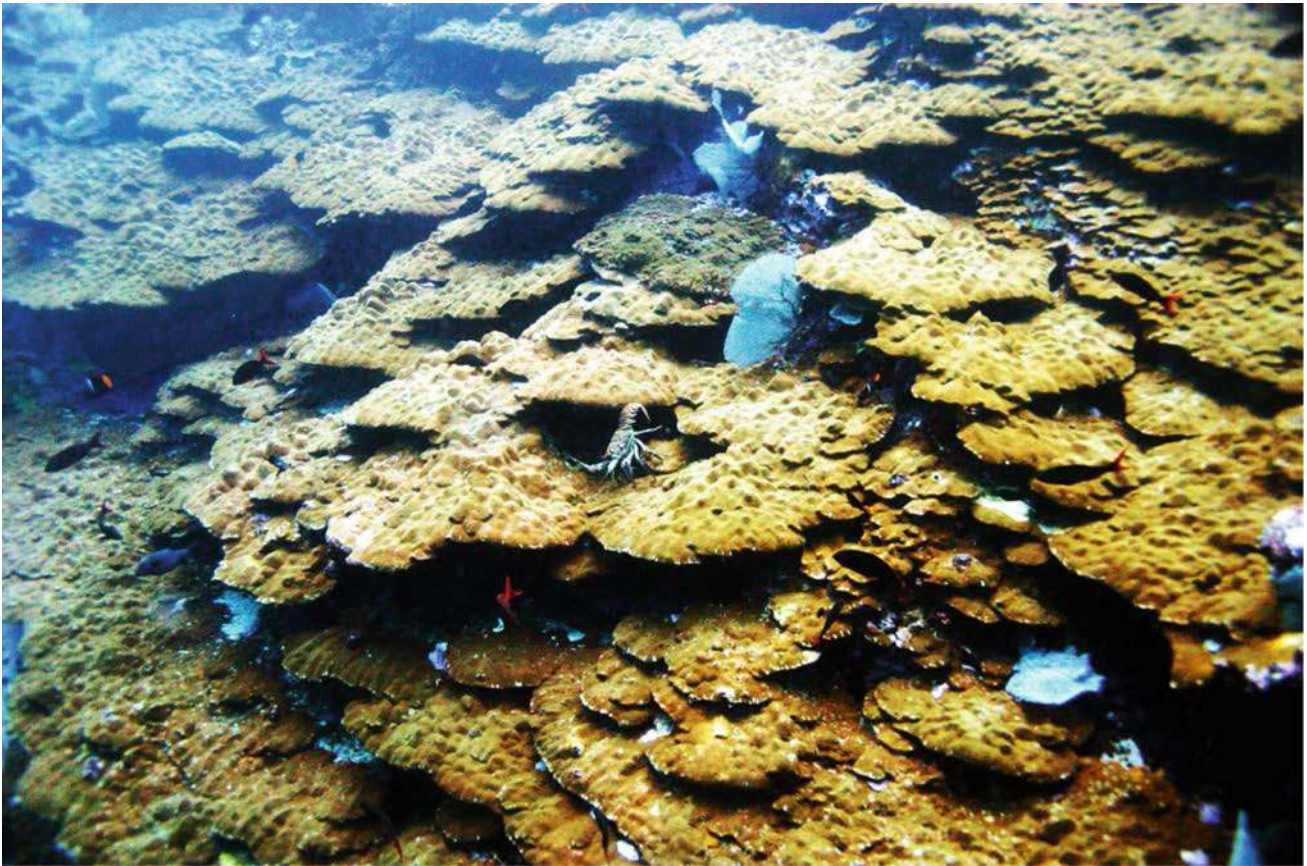


Fig. 5.23 *Porites lobata*, shingle-like growth form on inclined rocky substrate, 17 m depth, 25 June 2007, Malpelo Island (photo by F.A. Zapata)

guide illustrates numerous, robust, zooxanthellate species at several localities and depths.

Coral formations are best developed and most numerous at Isla de La Plata, about 30 km west of the mainland. Several patch and fringing coral reefs are present between 1 and 15 m depth along La Plata's east coast (Fig. 5.25a). Most of these formations are isolated and relatively small, ≤ 200 m in length; however, the largest fringing reef at Acuario is about 500 m long. *Pavona clavus*, some colonies 3–5 m in diameter, is the chief framework-building species at this locality. Farther south, at Palo Santo, *Pocillopora meandrina* is the principal reef-building species. A bathymetric assessment of zooxanthellate corals performed by Martínez et al. (2011) at five sites (Palo Santo, Acuario, Bahía Drake, Tres Piedra, El Faro) showed that *Pavona* and *Psammocora* exhibited a broad depth distribution with marked differences between species. *Pavona clavus* predominated from 1 to 6 m depth, whereas *Pavona gigantea* was present primarily from 6 to 15 m, with a few isolated colonies as shallow as 4 m. *Psammocora superficialis* colonies were found from 2 to 15 m, while *Psammocora stellata* occurred from 6 to 10 m only. *Pocillopora* species occupied mostly shallow depths, especially *Pocillopora ligulata* and *Pocillopora meandrina*,

which were not observed deeper than 5 m. *Pocillopora damicornis*, *Pocillopora verrucosa*, and *Pocillopora eydouxi*, however, reached depths of 10–11 m. The only colonies of *Porites lobata* observed at La Plata Island were present at the Tres Piedras site at 5 m depth.

The presence of six species of azooxanthellate scleractinians was also reported by Rivera and Martínez (2011). One of these (*Cladopsammia gracilis*), was especially abundant on the sides and underneath *Pavona clavus* colonies at the Acuario site. As in Panama, numerous gorgonian species were observed, but these generally were not common in coral reef habitats. Especially diverse and densely populated gorgonian communities were present at sites with strong currents, such as the shoals off Puntilla de Santa Elena reserve and south of Machalilla (Fig. 5.24).

Eastern Pacific coral reef development is generally confined to sheltered areas; the satellite image in Fig. 5.25b shows the prevailing swell direction from the SW and high wave assault on the western and southern island shores. Encrusting coralline algal communities are well developed along the exposed rocky coastal areas. The extent of coral reef development on mainland Ecuador now rivals and may exceed that in the Galápagos Islands. This was not the case

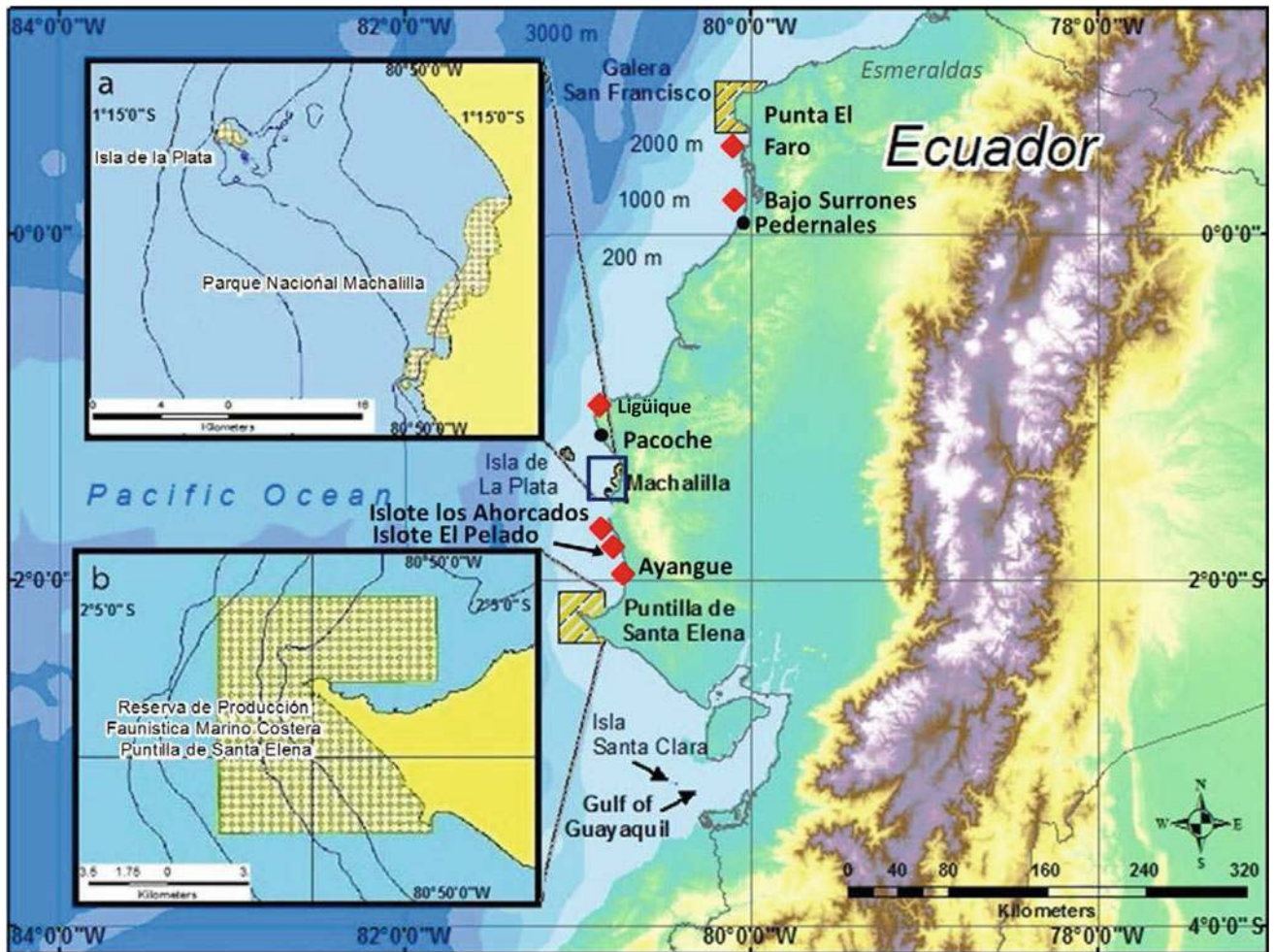


Fig. 5.24 Coastal Ecuador showing locations of three MPAs (Galera San Francisco, Machalilla, Puntilla de Santa Elena) and seven coral-rich sites (red diamonds) in Esmeraldas, Manabí, and Guayas provinces

before the ENSO activity of the past few decades, which has resulted in a marked reduction of coral communities and coral reefs in the southern Galápagos Islands. Since 2010, managers of the Machalilla National Park have installed nine permanent moorings in the MPA to prevent anchor damage from increasing tourism.

5.3.13 Galápagos Islands (Ecuador)

The Galápagos Islands are bathed in a confluence of westerly flowing surface currents that vary seasonally in magnitude and in latitude and are collectively referred to as the South Equatorial Current (SEC) in their passage towards the central Pacific. During years considered ENSO “neutral” the northerly island pinnacles of Wenman and Darwin (Fig. 5.26) are typically bathed in constitutively warmer waters from the southwesterly deviation of the NECC extending from the

Panama Bight. It strengthens its southerly extension as trade winds from the SE weaken during the equatorial warm season months (Dec–May). The southerly and central islands, however, are more exposed to the northerly extent of the South Pacific gyre, which transports surface water from the SE Peruvian oceanic and coastal systems, often referred to as the Humboldt Current. Separated by 1–2° of latitude from the aforementioned northwestern pinnacles, the central islands are as a result on average 1–2 °C cooler throughout the year (Banks 2002) harboring a greater relative proportion of coral genera such as *Pavona* spp. that persist under a wider range of fluctuating and cooler temperatures compared to *Porites* spp. that dominate the coral communities of warmer northerly islands (Banks et al. 2009).

The northeast to southwest SST gradient is exaggerated during the cold season months (Jun–Nov) and is reduced to less than 1 °C during the warm season (Dec–May) (Figs. 5.27 and 5.28). These regional differences notably disappear during

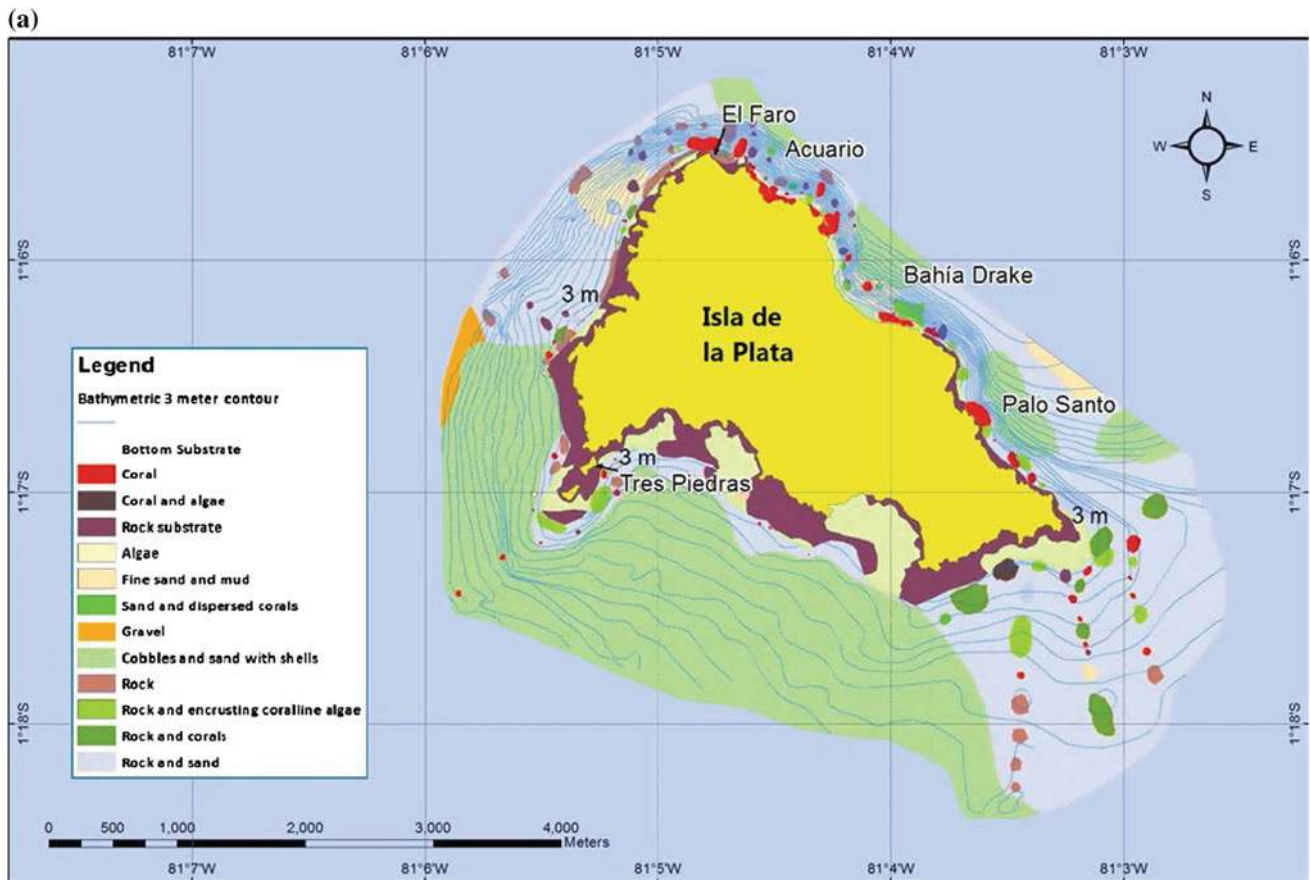


Fig. 5.25 a La Plata Island, Machalilla National Park, Ecuador. Substrate types are *color coded* (see legend), isobaths are shown at 3 m intervals. Modified after Rivera and Martínez (2011). b Satellite image of La Plata Island (2014 Digital Globe, Google Maps, April 2007)

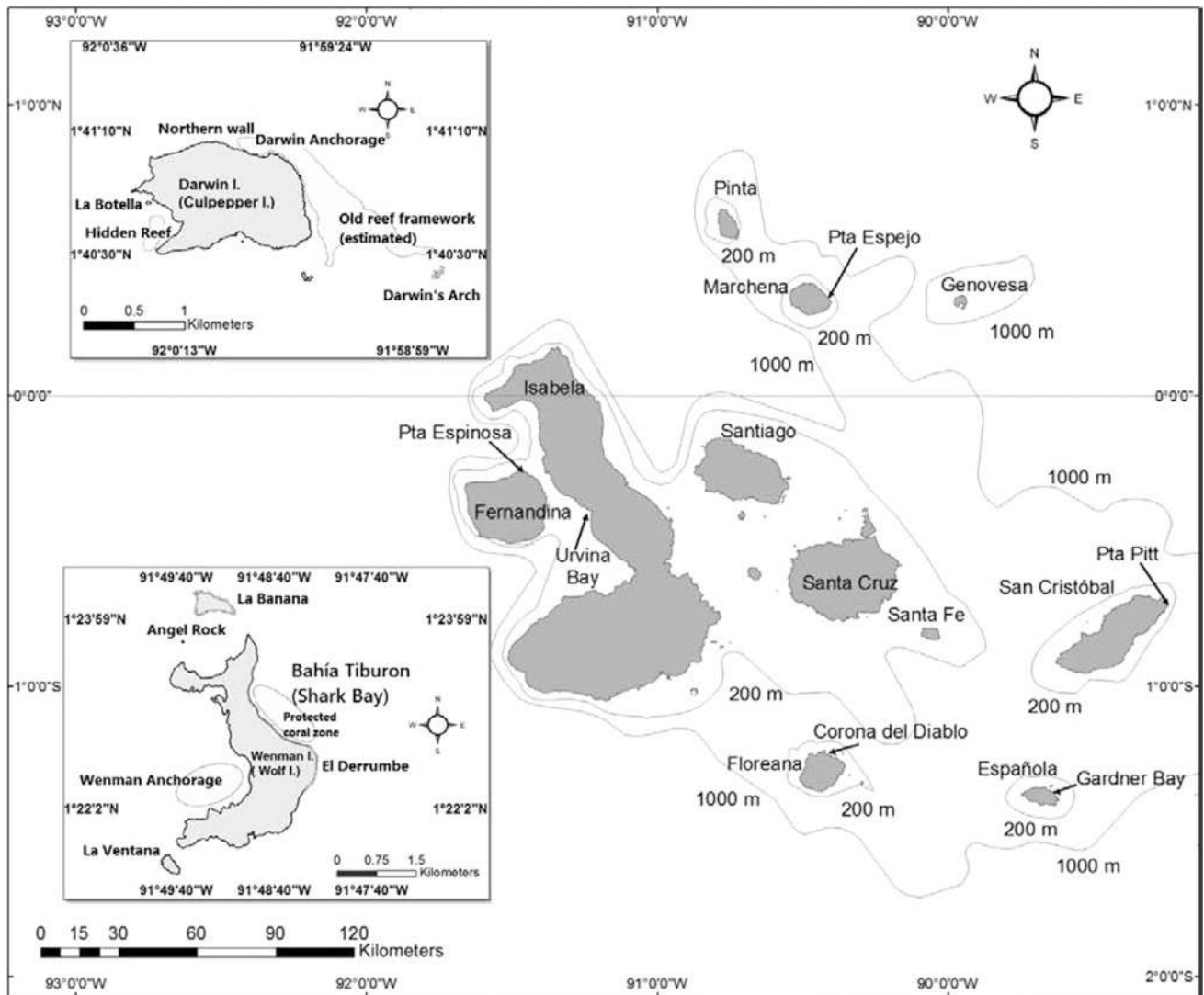


Fig. 5.26 Galápagos Islands (GAL), Ecuador. *Insets* of two northern-most islands Darwin (Culpepper) and Wenman (Wolf), located between $1^{\circ} 21'N$ and $1^{\circ} 41'N$

strong El Niño years when surface temperatures are elevated, sustained and homogenized across space and seasons, and conversely emphasized during La Niña episodes when trade winds and Humboldt transport are strengthened.

Coral species richness of the southern and northern Galápagos Islands is similar in spite of the large difference in habitable substrate areas (Dawson et al. 2009). This is likely a result of high connectivity between islands. Nineteen and 21 zooxanthellate scleractinian species are recognized as members of the coral faunas of the southern and northern islands respectively (Appendix). The coral fauna of the southern islands demonstrates its strongest affinity with the Ecuadorian mainland, and that of the northern islands with mainland Colombia (Fig. 5.2). Three species seemingly restricted to the southern islands, *Pavona chiriquiensis*,

Cycloseris curvata, and *Diaseris distorta*, are not important in reef building. Four species confined to the northern islands, *Leptoseris scabra*, *Pavona maldivensis*, *Pocillopora woodjonesi*, and *Porites evermanni*, are widespread Indo-Pacific species. All of these species except *P. woodjonesi*, which is rare, do contribute minimally to reef building in different areas of the eastern Pacific.

Development of coral communities and structural reefs is best expressed at Darwin (Fig. 5.29) and Wenman (Fig. 5.30), the two northern-most islands (Glynn et al. 2015). The largest known coral reef in the Galápagos Islands, located on the insular shelf between Darwin Arch and the eastern shore of Darwin Island, is nearly one km in length, and consists of vertically consolidated poritid stacks towering from 2 to nearly 4 m in height. While the chief frameworks

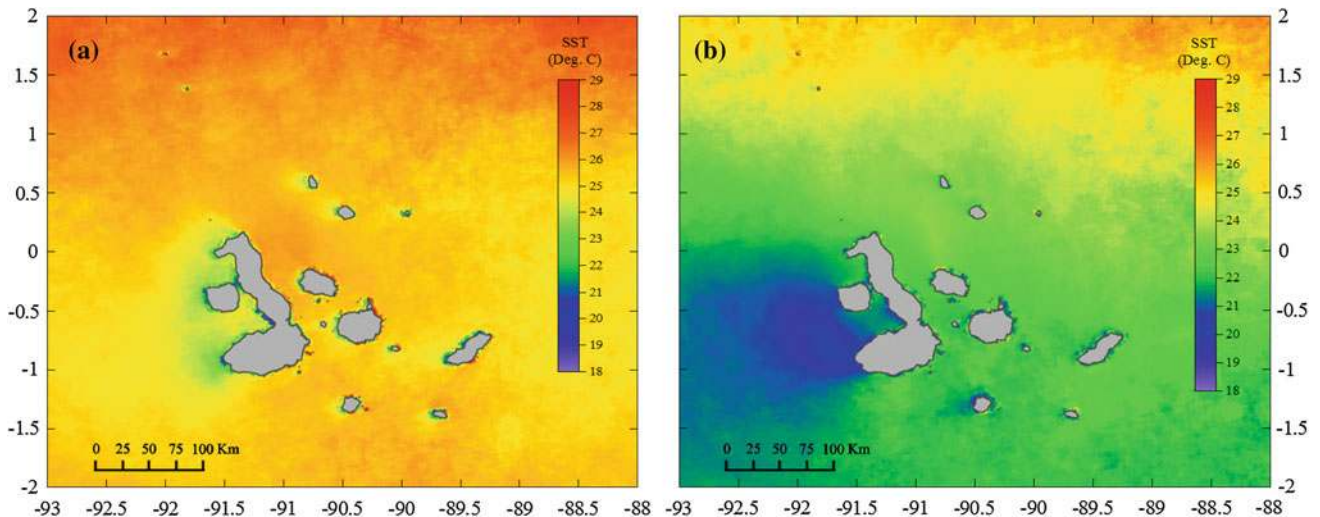


Fig. 5.27 Monthly averaged sea-surface temperatures (SSTs) across the archipelago during **a** the warm season (Dec–May) and **b** the cold season (June–Nov) derived from NASA MODIS-Aqua 1 km² remotely sensed data (2003–2008)

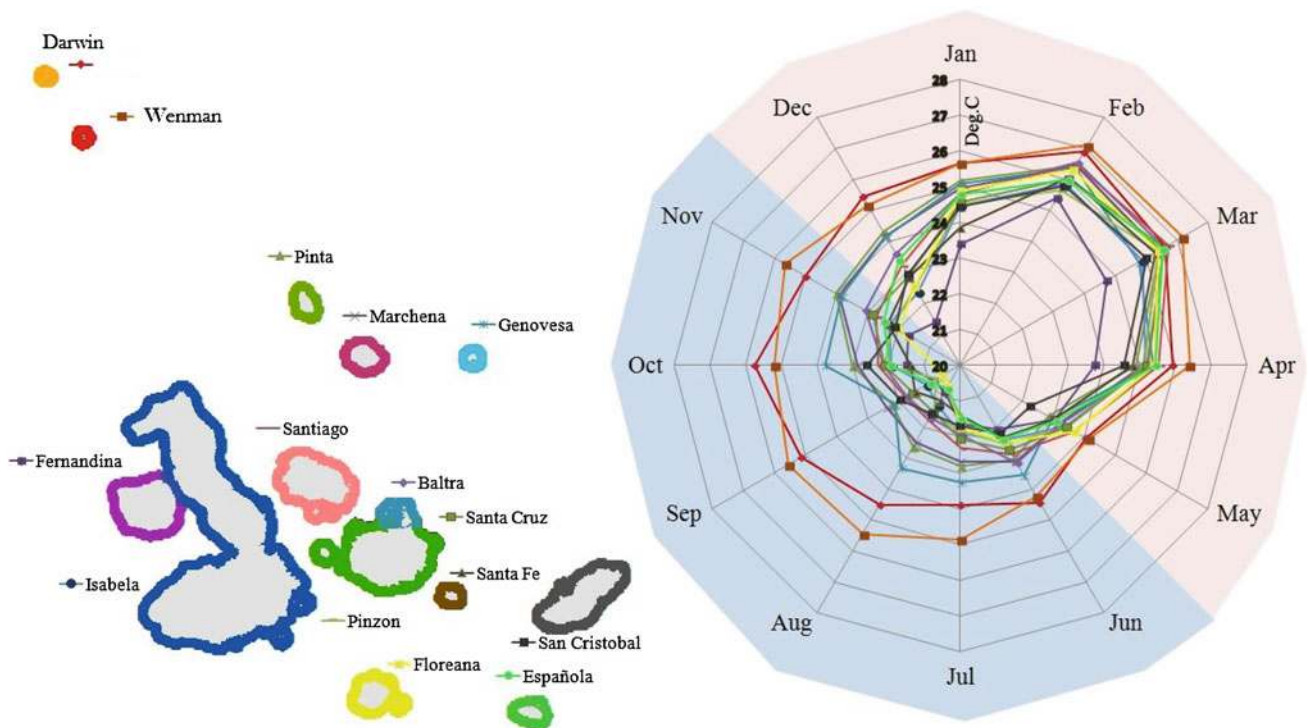


Fig. 5.28 Monthly patterns in mean SST for near-shore environments across each major island (calculated areas up to 5 km from island coastline using NASA MODIS-Aqua 1 km² remotely sensed data

2003–2008). Islands and respective monthly SST plots are color coded. Lowest and highest monthly SSTs occurred at Floreana in September (20–21 °C) and at Darwin in March (27–28 °C)

are at 12–16 m depth, live coral cover extends to 28 m depth along the NW/SE trending isobath. This reef has demonstrated notable recovery since the 1982–83 El Niño with live cover now exceeding 25 % over most of its length. The high cover of *Pocillopora* spp. before 1982–83 has virtually disappeared (Glynn, unpublished observation). We propose

naming this impressive structure Wellington Reef, in honor of Gerard M. Wellington who performed the first island-wide marine resource surveys in the Galapagos Islands.

Unusually high coral abundances and incipient reef development characterize the narrow insular shelf along a 1 km stretch at Bahía Tiburón, Wenman Island (Fig. 5.30).

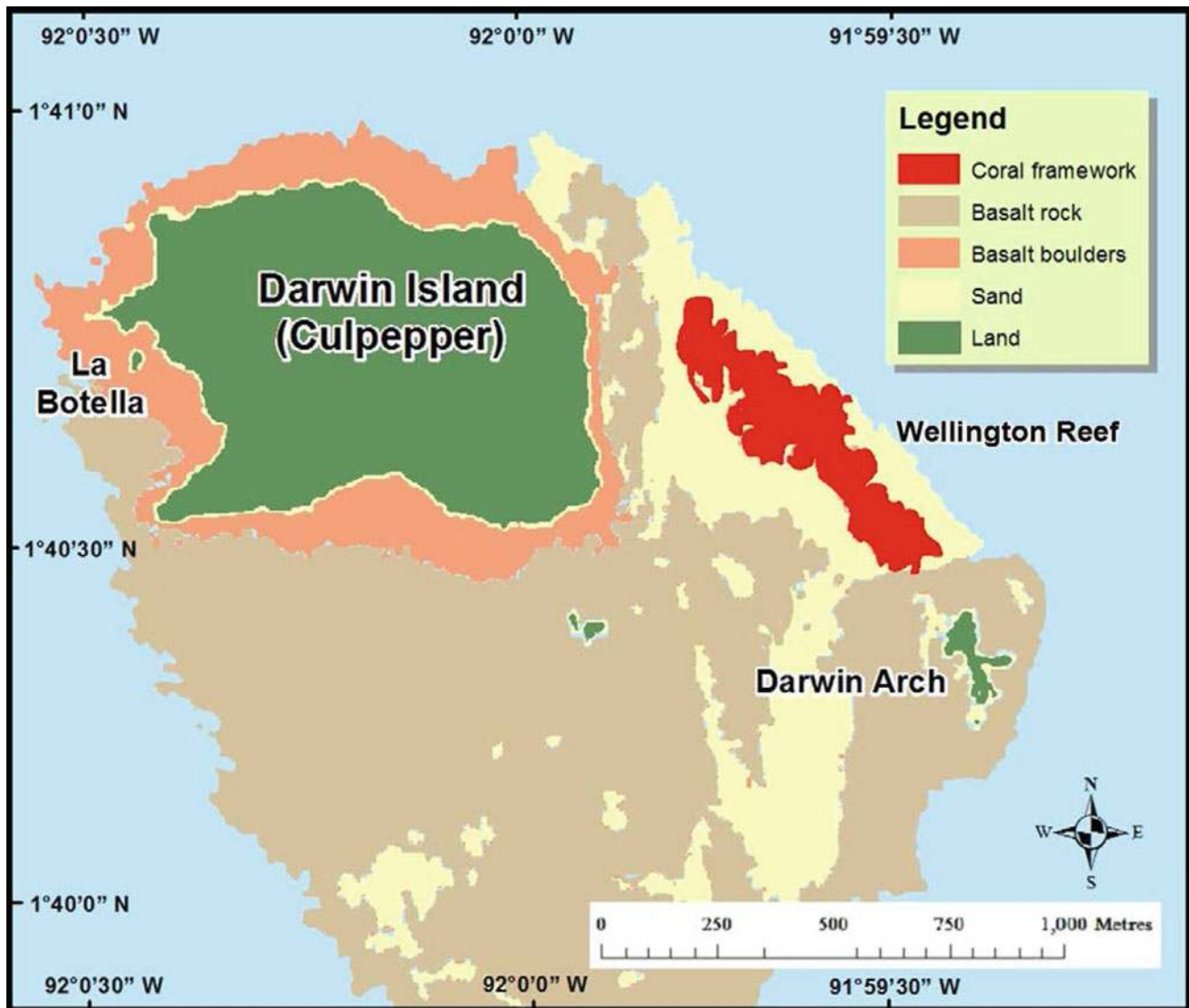


Fig. 5.29 Darwin Island, showing location of recently mapped coral framework of Wellington Reef

As at Darwin Island, massive *Porites* colonies predominate; however, large branching *Pocillopora* spp. colonies are numerous on rock boulders at shallow depths (<8 m), and *Pavona* spp. increase in abundance toward the deep (20–30 m) reef base.

Biogeographic patterns across the archipelago are greatly influenced by meso- and small-scale oceanography. Surface waters mix significantly with subsurface inputs from bathymetrically- and wind-driven upwelling processes across the archipelago. The eastward flowing Equatorial Undercurrent (EUC or Cromwell Current) shoals to 40–80 m depth over and around the Galápagos platform. The EUC brings 13–18°C water into the surface and is considered the most important oceanographic feature in maintaining extraordinary plankton productivity and unique

cold water habitats, particularly in the western exposed islands of Fernandina and Isabela (Fig. 5.26).

In contrast to the enabling Panamic influence in the north, the EUC favors solitary azooxanthellate corals and rather restricts the development of reef-building corals in the western regions, generating small scale patterns of cold water mixing around western shores of the central islands. These areas are relatively spatially consistent yet quite ephemeral where EUC filaments shoal around the bathymetry and mix into the upper euphotic zone (Shaeffer et al. 2008). Since widespread mortalities during the El Niño event of 1982–83, the patterning of suitable coral habitat in the southern islands is more heterogeneous compared to that of Wolf and Darwin. Although small patches of coral communities are common across the central isles (Pinta,

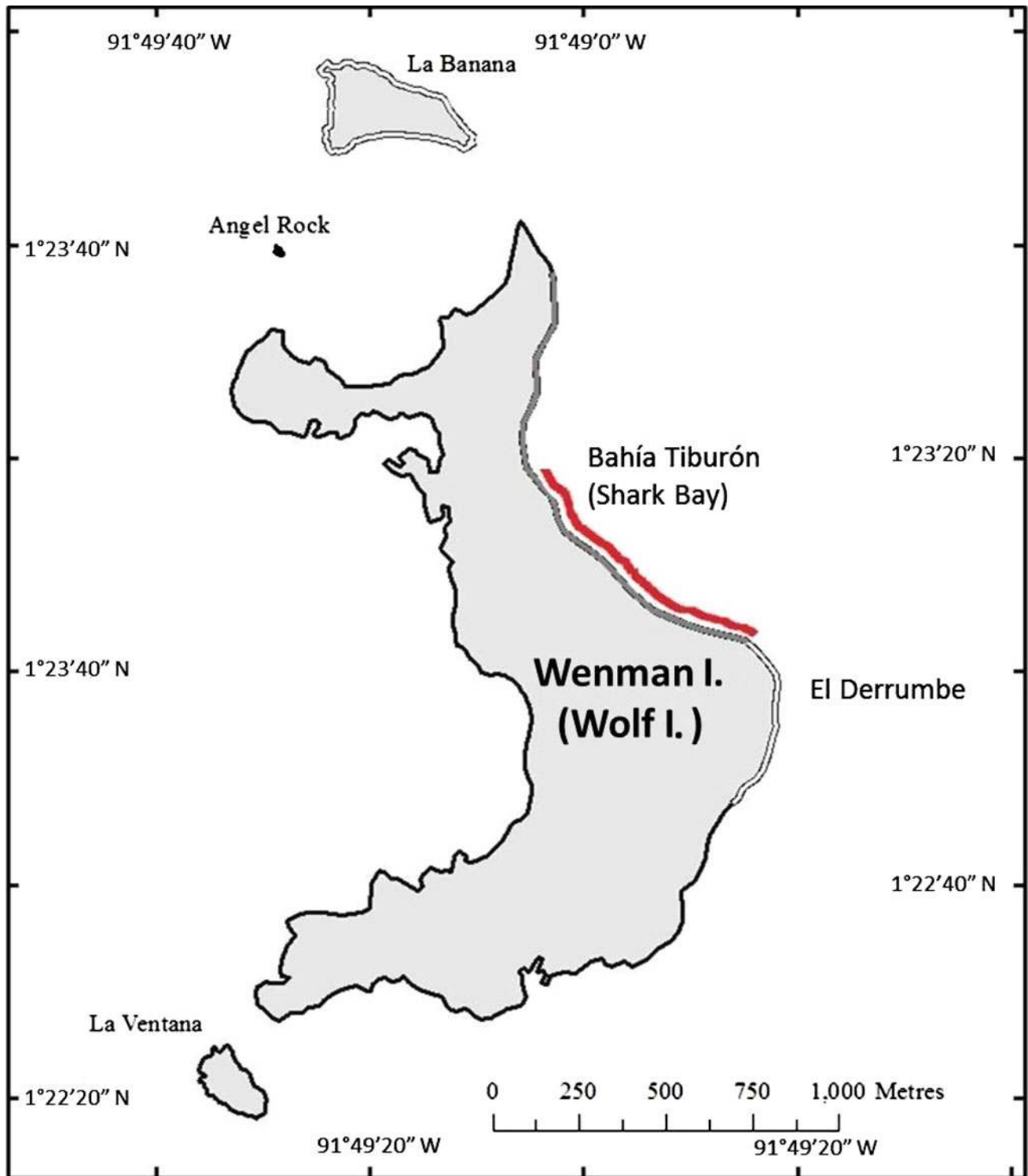


Fig. 5.30 Wenman Island, coral community development (*red line*) greatest along northeastern shelf at Shark Bay

Marchena, Genovesa), the largest extensions of zooxanthellate corals are found across the relatively sheltered northern leeward shores of Floreana Island, protected bays in eastern Santiago, Punta Pitt at the eastern end of San

Cristóbal and the southerly extent of Gardner Bay at Española Island (Fig. 5.26).

An interesting feature of the western islands of Isabela and Fernandina is that they show historical evidence of large

reef-forming corals in sheltered bays, despite the prevailing EUC upwelling and near absence of reef-building corals today. For example, there are large *Pavona clavus* colonies in Urvina Bay, Isabela, which was uplifted in the 1950s (Glynn and Wellington 1983; Colgan 1990, 1991; Dunbar et al. 1994). There are also reports of *Pocillopora* spp. persisting in shallow coastal pools such as those at Punta Espinosa, Fernandina, and Concha y Perla lagoon near Puerto Villamil, Isabela (Baums et al. 2014; Feingold and Glynn 2014). It seems possible that the strong stratification through solar heating at the surface in sheltered pools with low current could create a sub-habitat suitable for sporadic hermatypic coral growth. The downstream oceanic connectivity between coralline communities in the east and possible incursions of warmer water during strong El Niño years may also present opportunities for new recruitment over decadal scales.

Other unique coralline environments in the islands include extensive beds of free-living *Psammocora stellata* and *Porites lobata* composed of 3–14 cm diameter fragments across sand substrates at Punta Espejo, southeast Marchena Island (Feingold 2001). The fungiid free-living coral *Diaseris distorta* also persisted with a large population (>100,000 individuals) localized in patches across the sand sediment platform at 15–20 m depth east of Corona del Diablo (a small volcanic offshore crater at the northern tip of Floreana Island). Another fungiid, *Cycloseris curvata*, was present there in small numbers (30–40 individuals) (Feingold 1996).

Internal waves also influence local coral ecology. Kelvin waves that propagate to the east and pass in sets from the western Pacific shoal and deepen the thermocline over week-long periods in their passage across the islands. Tropical Instability Waves also form large scale front features with eddies that extend out from the upwelled equatorial cold tongue passing over the archipelago towards the west (Sweet et al. 2009). Both produce short-term changes in the water-column structure within larger seasonal signals. Kelvin waves being part of the El Niño trigger, and wider ENSO dynamic, are usually responsible for anomalous periods of alternating cool and heated surface waters that can last several weeks regardless of the season. Given strong surface heating of the upper mixed layer under the equatorial sun this can cause up to 10–12 °C thermal anomalies at 10 m depth over very short periods (4–6 days) as cold water extends into the shallow coastal zone.

Whereas warm-water stress upon corals in the region was largely evident in the widespread mortalities of the 1982–83 and 1997–98 El Niño events, cold water anomalies are pronounced and also impact coral health when combined with La Niña episodes (see Chap. 8, Glynn et al.). Significant mortalities of *Pocillopora* spp. were observed during the onset of the 2007–08 La Niña event. The upwelling

phase of a set of Kelvin waves coincided with the transition into the cold season, further exaggerated by La Niña conditions both exacerbating and prolonging cold-water shock to shallow water communities across the southerly islands. The gradual recovery observed since 1995 of *Pocillopora* spp. colonies monitored in Corona del Diablo (Floreana Island) was greatly impacted between 2007 and 2009 with numbers falling from 167 colonies to just 20 (Feingold and Glynn 2014) and similar patterns were observed across the south. Visual surveys along the northeast coast of Pinta in 2010, however, showed encouraging survivorship of large *Pocillopora* colonies, likely as a result of their positioning in transitory warmer water to the north.

These effects were also observed as far north as Wolf (Wenman) and Darwin islands, which harbor the greatest remaining massive coral frameworks (Glynn et al. 1983, 2003, 2008, 2015; Banks et al. 2009). Those species that persisted after the 1982–83 and the compounding 1997–98 El Niño event exhibited an interesting potential for redundancy in functional roles between alternating periods of warm water and cold-water stress. *Pavona clavus* colonies adjacent to *Porites lobata*, for example, remained healthy during heavy bleaching of *Porites* during cold water incursions in February 2008. This may have allowed for the maintenance of key ecological services (e.g., structure, refuge, and productivity) until the recovery of the more abundant *Porites* as warmer conditions returned. Although not yet observed directly, the reverse is likely to be true under strong heating stress for *P. clavus*. These observations also lend themselves to the possibility of selection for temperature-resistant zooxanthellae clades or “disaster” endosymbionts (Correa and Baker 2011; see Chap. 13, Baker et al.) persisting in those corals that survived temperature shock during recent strong ENSO events in northerly communities.

A reduced and clustered population following El Niño mortalities also presents other concerns. During surveys from 2006–2008 the average prevalence of coral tissue abnormalities across eight sites at Wenman and Darwin was over 23 % (Vera and Banks 2009), which is at least twice as high as that reported in Caribbean and Indo-Pacific regions. The species *Porites lobata* was most affected with 35 % showing symptoms of parasitism or illness. The condition identified as *Porites* trematodiasis was most evident. These findings support observations from other areas correlating the frequency of strong bleaching events and high host densities with disease outbreaks (Bruno et al. 2007; Cróquer and Weil 2009). Background climate-change scenarios of increased ocean acidification in a region already subjected to low pH-upwelled waters also presents unqualified risks for reduced coral calcification, increased infiltration of bioeroders, and diminished reef accretion (Manzello 2010; see Chap. 18, Manzello et al.).

Nonetheless a gradual seven-year recovery trend across the northerly coral frameworks over the 12–25 m platform between Darwin Arch and the east coast of Darwin Island (Glynn et al. 2009, 2015) is also corroborated by local observations of tissue regeneration and coral recruits close to existing colonies at other sites (Banks, Personal Communications). Wenman and Darwin, despite their small geographic size, make a disproportionately large contribution to the overall Panamic and Indo-Pacific subtidal biodiversity in the islands. Galápagos park authorities, in concert with local NGOs and the Ecuadorian Navy, have begun implementing low-impact tourism anchorages to complement zoned-protected areas physically signposted in 2006 (Merlen et al. 2009). Best-practice guidelines for tourism dive operations would help reduce physical damage by inexperienced divers to reefs, while fixed bouy anchorages should help minimize the significant damage caused by anchors and chains to slow growing colonies and encourage their future recovery.

The effect of the increased human footprint in the Galápagos Marine Reserve since the 1980s and its impact on habitat-forming corals is a topic of continuing speculation. Observations since the strong El Niño events suggest that climate disturbance when coupled with certain human disturbances (such as unsustainable extraction of resource species or chronic anchor damage) can cause broad shifts in ecosystem composition and biodiversity state that further inhibit natural recovery processes (Sonnenholzner et al. 2009; Edgar et al. 2010). Significant interactions between overfishing and ENSO may be particularly important in determining extinction risk given that since 1982–83 several azooxanthellate coral species have greatly reduced populations (e.g., *Tubastrea floreana*) or are even possibly already extinct (e.g., *Rhizopsammia wellingtoni*). Significant increases in the proportion of degraded corals and the bioerosion of weakened coral frameworks by the slate pencil sea urchin *Eucidaris galapagensis* have been linked to coral degradation and observed both *in situ* and upon tectonically uplifted colonies (Colgan 1990, 1991). The recovery of coral populations is further compounded by overgrazing scenarios inhibiting recruits through the notable proliferation of the sea urchins *Tripneustes depressus* and *Lytechinus semituberculatus*; these sea urchins may create barrens if there is a significant reduction in their natural reef predators (Sonnenholzner et al. 2009; Edgar et al. 2010). Any such biotic and indirect cascade effects will influence coral recovery intervals, population viability, and recruitment between strong El Niño events.

The 1997–98 El Niño event coincided with rapid extraction of lobster species *Panulirus gracilis* and *Panulirus penicillatus*, both candidate predators for coral burrowing molluscs such as *Lithophaga* spp. and the corallivore *Coralliophila violacea* at Wenman and Darwin Islands. Targeted and incidental fishing of intermediate-sized

predatory fish with regulatory roles such as the leather bass *Dermatolepis dermatolepis*, demersal labrids and other serranids (including the exported insular endemic bacalao *Mycteroperca olfax*) also accompany both lobster and sea cucumber fishing seasons. Multivariate reconstructions of benthic coverage (Edgar et al. 2010) from early (pre-1982–83 El Niño) photographic records and subsequent photo-quadrat and visual census monitoring indicate little change in benthic community composition between 1984 immediately after the strong El Niño event and some 20 years later in 2004. The footprint when removing natural biological controls of grazers through fisheries appears to be widespread in the archipelago and has resulted in high abundance of encrusting calcareous algae such as *Lithothamnium* sp. indicative of sea urchin barrens. Observations of increased *Eucidaris* sea urchin densities near to fishing ports (Sonnenholzner et al. 2009; Edgar et al. 2010) appears to support this hypothesis with analogies reported in other regions such as Hong Kong (Qui et al. 2014).

With new coastal-zoning reforms being considered, GMR coral communities present a curious dichotomy. They are particularly vulnerable before acidification and cascade effects after removal of regulatory species in their vicinity, yet are also a naturally resilient habitat subject to intense changes conditioned by a unique oceanography and millennia of ENSO fluctuations in the region. Moderation and mitigation of these effects through monitoring and feedback into management certainly deserve careful consideration given that novel climate stress is likely to be deterministic in the future survivorship of remaining Galápagos corals and the diverse communities they support.

5.3.14 Rapa Nui (Easter) and Salas y Gómez Islands (Chile)

Rapa Nui (Easter) and Salas y Gómez islands (both included under RPN, Appendix), the latter slightly more than 400 km east of Rapa Nui, share a common and relatively species poor shallow marine biota (Fig. 5.31). Two comprehensive reports by Castilla and Rozbaczylo (1987) and DiSalvo et al. (1988) offer excellent introductions to the marine habitats and biota of Rapa Nui. Studies focusing on the coral faunas and communities became available in the early to mid-2000s (Wellington et al. 2001; Hubbard and Garcia 2003; Glynn et al. 2003, 2007).

Only 15 species of zooxanthellate corals are known from the larger and more speciose Rapa Nui with nearly half of these in the genus *Pocillopora*. *Pocillopora verrucosa* exhibits high cover over mid-depths at several sites on the west coast and at Motu-Iti and Motu-Nui at the southern tip of Rapa Nui (Fig. 5.32). *Porites lobata* is the single poritid species present, and makes up the bulk of the coral cover

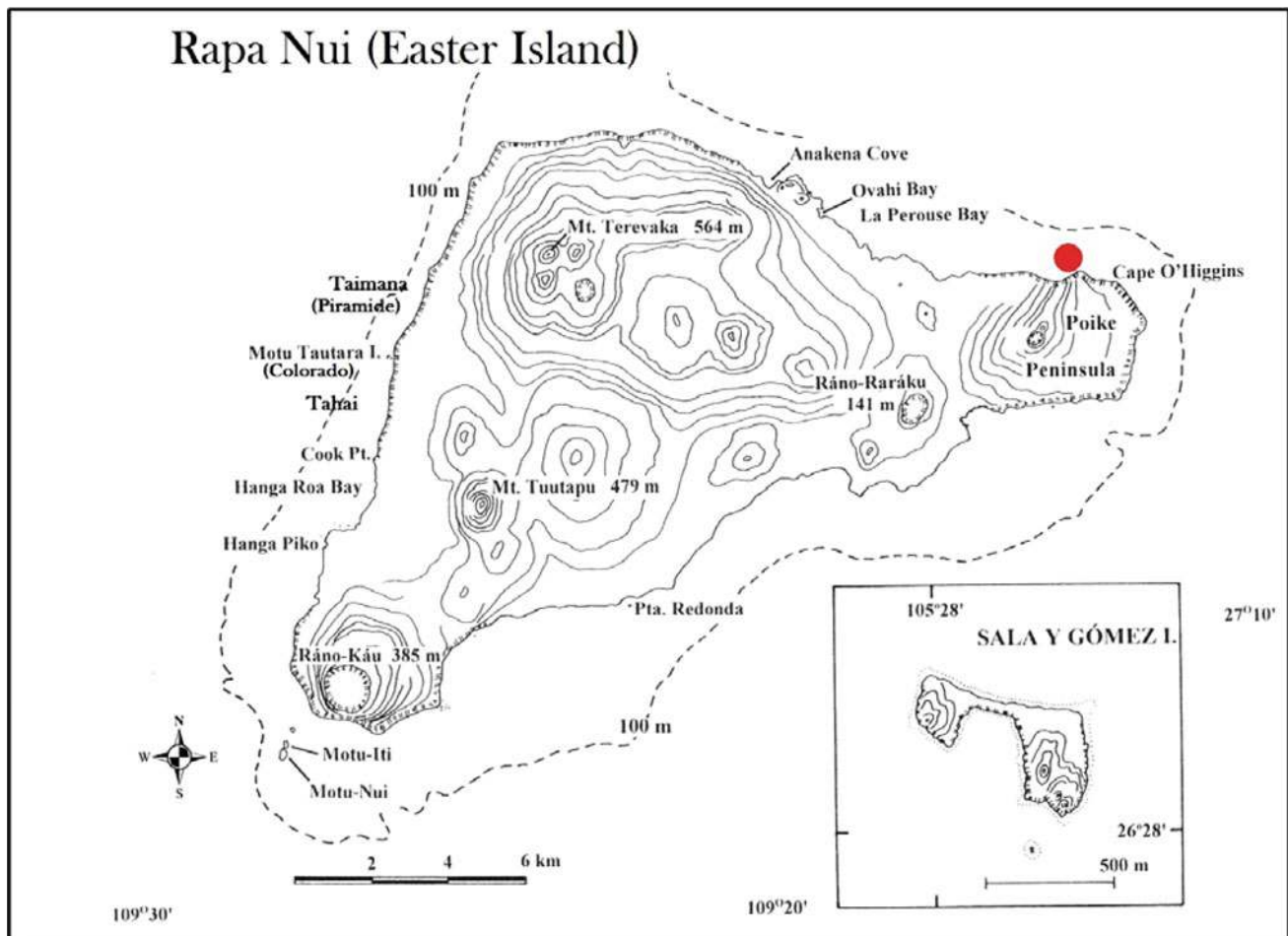


Fig. 5.31 Rapa Nui (Easter Island), Chile. Inset, Salas y Gómez Island, ~500 km east of Easter Island. Red dot west of Cape O'Higgins denotes location of coral reef framework (Hubbard and García 2003)

across all depths. Unlike its colony growth form elsewhere in the eastern Pacific, which is smooth to nodular and dome-shaped, at Rapa Nui the corallum takes on a variety of forms—columns, blades, spikes, cones, and marginal tabulae (Glynn et al. 2003). *Pavona* species, so prominent at other eastern Pacific sites, are absent from the Rapa Nui islands group. However, two species of faviid corals, *Leptastrea purpurea* and *Leptastrea transversa*, are present but they are minor components of RPN coral communities. These are the only members of the Faviidae known from the eastern Pacific region. While endemism is high in some groups (e.g., molluscs, 42 %; fishes, 77 %; Friedlander et al. 2013), all zooxanthellate scleractinian corals, except for the two faviid species, are widely distributed throughout the eastern Pacific and central/south Pacific regions (Appendix). Ongoing molecular genetic studies, however, suggest re-classification of *Pocillopora verrucosa* as a separate endemic species (D.

A. Paz-García, E. Wieters, F.J. García-de-León, E.F. Balart, unpublished data).

Traditionally, Rapa Nui has been considered as the eastern-most outpost of the Polynesian or southeast Pacific region (e.g., Veron 1995). However, an analysis of coral species similarities (presence/absence records) between the Pitcairn Island group and Rapa Nui indicates that this locality is more closely aligned with the eastern Pacific faunal group (Glynn et al. 2007). In support of this result, a dispersal barrier between Rapa Nui and the Pitcairn Islands was revealed through satellite-tracked surface drifter buoys. Surface currents were found to flow from east to west, forming an effective barrier to the eastward dispersal of planktonic larvae that occupy the upper surface of the ocean. Most recently, Veron et al. (2015) also concluded that Rapa Nui has its strongest affinity with the far eastern scleractinian zooxanthellate fauna. Reef-building coral species richness is



Fig. 5.32 *Pocillopora verrucosa* prominence in a mid-depth (12–15 m) coral community at Taimana, Rapa Nui west coast, 12 December 2012 (photo courtesy of Alba Medrano)

notably higher in the Pitcairn's Islands than at Rapa Nui. Veron (1995) reported 61 species in the Pitcairns; recent surveys have increased this number to 87 (Appendix).

Vibrant coral communities with high coral cover (40–80 %) characterize most sites along the now well-sampled western and northeast coasts of Rapa Nui, whereas corals appear to be scarcer on the less-studied, wave-battered southeast coast that is directly exposed to major oceanic swells (DiSalvo et al. 1988). Considering the high live coral cover at favorable sites, it is not surprising that incipient reef development has been observed. Massive *Porites* colonies in particular have relatively high skeletal extension rates, about 1 cm year^{-1} (Glynn et al. 2003), are longevous, and attain enormous dimensions (Figs. 5.33, 5.34 and 5.35). *Porites* frameworks were observed by Hubbard and Garcia (2003)

on the western and northern sides of the island. The largest of these is found northeast of Poike Peninsula, off Cape O'Higgins. Some coral buildups on elevated basalt substrates appear deceptively like true structural reefs (Fig. 5.34). These shelf locations are protected from severe wave assault from the southeast, originating in high latitude southern seas. Even though pocilloporid corals often occur as dense aggregations at sheltered sites, they do not form interlocking frameworks as at other ETP localities.

In general, two species, *Porites lobata* and *Pocillopora verrucosa*, account for the vast majority (95 %) of total coral cover across the island. While typically scarce, encrusting colonies of *Leptastrea* spp. can reach up to 10 % cover at some locations near Ovahe and Tahai. There they occur intermixed with *Porites* and *Pocillopora* in large patches



Fig. 5.33 Single colony of *Porites lobata*, ~10 m in diameter, off Anakena, 17 m depth. Photographed in August 1998 by Henry Tonnemacher, courtesy Dennis K Hubbard

(10s of square meters) on flat, mid-depth (10–12 m) platforms. Although still nominal in terms of their contribution to total coral cover, a notable increase in the prevalence of *Pocillopora eydouxi* has been observed over the past five years, particularly along the west coast (E. Wieters and H. Garcia, personal observation).

Generally, weak depth zonation patterns characterize changes in the relative abundances of the dominant coral community functional groups (Friedlander et al. 2013; Wieters et al. 2014). Total coral cover is lowest at shallow depths (<5 m), however, where branching *Pocillopora* spp. are nearly absent, massive *Porites* are predominant (average 38 % cover). Below this, *Pocillopora* increase in abundance to peak at mid depths (15–20 m), where their relative contribution to total coral cover is similar to that of *Porites* at most sites (average 38 and 36 %, respectively). Massive *Porites* again dominate in deeper habitats (>20 m), and *Pocillopora* become less abundant, but still reach 10–

20 % cover. Abundant live coral extends well below the 30 m depth limit with *Pocillopora* and *Porites* known to occur frequently to 60 m depth.

Across the northeast and west coasts, shifts in the relative abundance of species cause spatial differentiation in the functional structure of coral-dominated benthic communities and fish assemblages (Wieters et al. 2014). Though changes are best described by multivariate characterization of communities based on functional groups, some of the most apparent differences in terms of species are due to relatively higher abundances of *Pocillopora* and some macroalgal groups along the northwestern coast and tip of the island, as well as variable presence and abundances of the reef fish *Centropyge hotumatua*, *Acanthurus leucopareius* and *Chrysiptera rapanui*. Further studies have also identified contrasting communities at similar depths, with a preponderance of either corals or macroalgae (predominantly *Padina*). These alternative, coral/algal community configurations can exist



Fig. 5.34 *Porites lobata* community, off Anakena, 30 m depth. Photographed in August 1998 by Henry Tonnemacher, courtesy Dennis K. Hubbard

side-by-side (within hundreds to thousands of meters), and commonly present sharp boundaries between them, which remain unaltered for relatively long periods of time.

Published studies concerning ecological processes maintaining coral-dominated communities at Rapa Nui are virtually nonexistent. Ongoing experimental and comparative studies at a few locations suggest that, like elsewhere, grazing is a critical process involved in preventing macroalgae from overgrowing live corals. Evidence thus far also suggests grazing is especially effective when corals have been previously damaged and weakened by thermally-induced bleaching (Wieters, unpublished data). At Rapa Nui, where herbivorous fishes are generally small-bodied and in low abundance, sea urchins (*Diadema*) appear to be particularly important. While no previous studies have documented ecological effects or even the diet of benthic herbivores at Rapa Nui, comparisons with nearby Salas y Gomez reveal between-island differences in abundances of sea urchins and macroalgae (Friedlander et al. 2013), further suggesting the potential for a critical

top-down role by these benthic herbivores. Whether the effects and relative importance of sea urchins are currently magnified at Rapa Nui, due to the very apparent overfishing that seems responsible for reduced abundances of carnivorous fishes and other large predators, is not known. Obvious sea urchin-induced erosion of dead corals and limestone deposits is limited to a few specific locations of particularly high sea urchin densities (mean $19.0 \text{ ind. m}^{-2} \pm 2.6 \text{ SE}$), but there is no evidence that sea urchins cause mortality of established live colonies (Wieters, personal observation).

Severe, island-wide coral bleaching at Rapa Nui is known to have occurred twice in recent years: (1) following torrential rains and associated increased sedimentation in 1980 (Egaña and DiSalvo 1982), and (2) following prolonged sea warming associated with La Niña in 2000 (Wellington et al. 2001). This exceptionally strong La Niña event, centered at and above the equator in the northern hemisphere, resulted in two month-long elevated SSTs in the southern ocean, that extended over 73° of latitude from Rapa Nui to Fiji. During this event the bleaching threshold at Rapa Nui (26.1°C) was

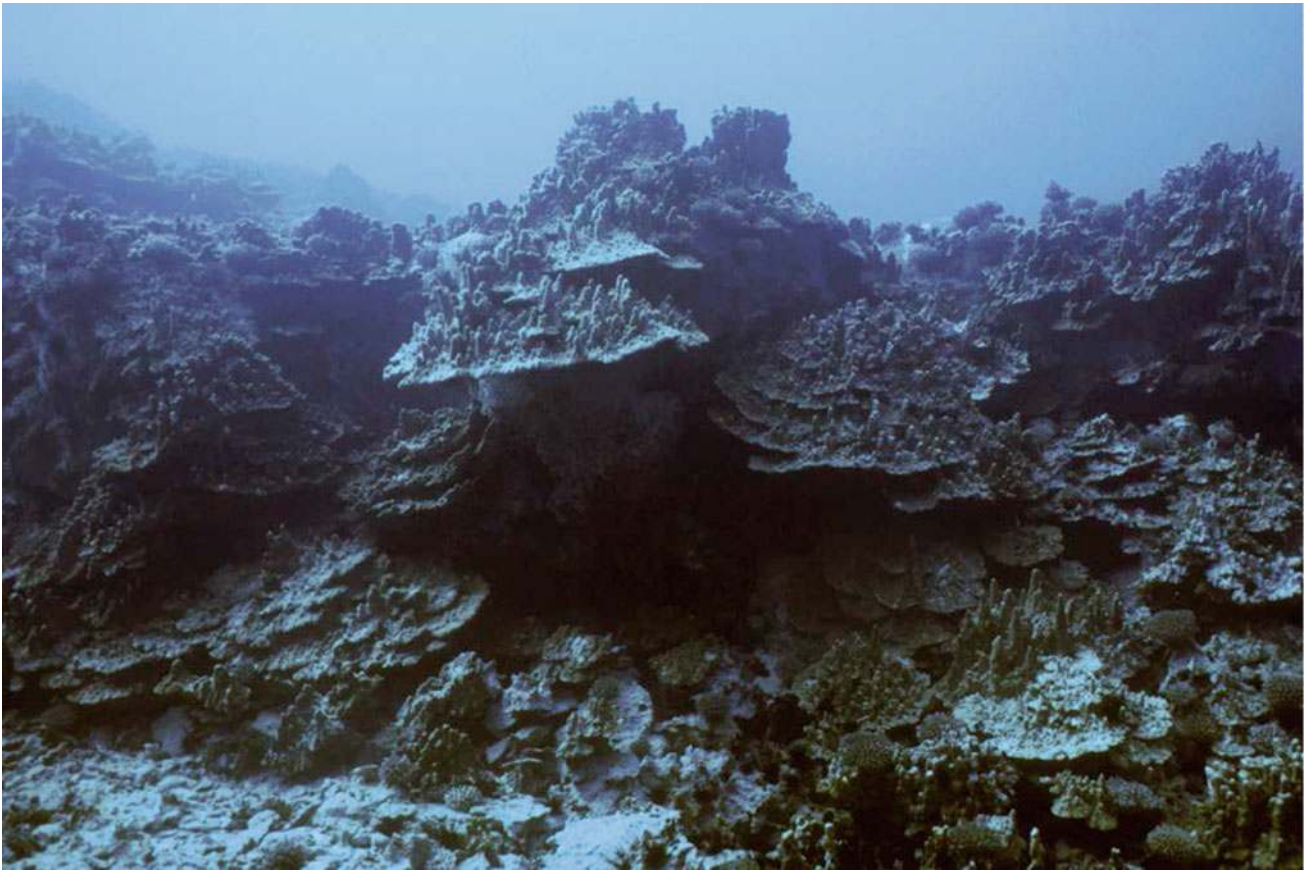


Fig. 5.35 Veneer of *Porites lobata* colonies overtopping a lava tube off Motu Tautara, ~25 m depth. Photographed in August 1998 by Henry Tonnemacher, courtesy of Dennis K. Hubbard

exceeded for 9–10 weeks in February and March. This thermally-driven disturbance led to high mortality of *Pocillopora* spp. to 20 m depth throughout the island. Within one year after the disturbance, greater than 80 % of *Pocillopora* colonies inhabiting shallow habitats (<9 m) were dead and a clear pattern of decreased mortality with increasing depth was documented (Fig. 5.36). While *Porites* experienced generally similar levels of bleaching compared with *Pocillopora* in 2000, the former appeared much more resistant with far fewer colonies having succumbed a year later. Initial recovery of *Pocillopora* was uneven spatially, with increased prevalence of coral tissue abnormalities and continued mortality observed up to 5 years after the massive bleaching at some sites, particularly in mid- to deep habitats (>13 m; compare mortality levels in 2001 and 2005, Fig. 5.36). The factors inhibiting or delaying recovery and causing the continued declines of *Pocillopora* at some sites are unknown, but preliminary evidence suggests that

depth-dependent resistance and other disturbances can be important. Despite the extensive bleaching, large scale mortality of one of the dominant species, and among-site variability in recovery, overall recovery of coral communities to date has been impressive, with total coral cover now similar to (deep habitats) or above (shallow habitats) the pre-bleaching levels (Wieters et al. 2014).

While no massive bleaching event similar to that documented in 2000 has been observed recently (as of 2014), regular surveys over the past 5 years show that bleaching of *Pocillopora* and *Porites* occurs every year. While the intensity/prevalence varies greatly among sites and years, the regular decline in zooxanthellae densities and/or degraded pigmentation suggests that these corals are frequently subjected to stressful conditions. This relatively mild bleaching rarely appears to cause coral mortality directly, but does appear to have consequences for other species in the community. Bleached colonies of both

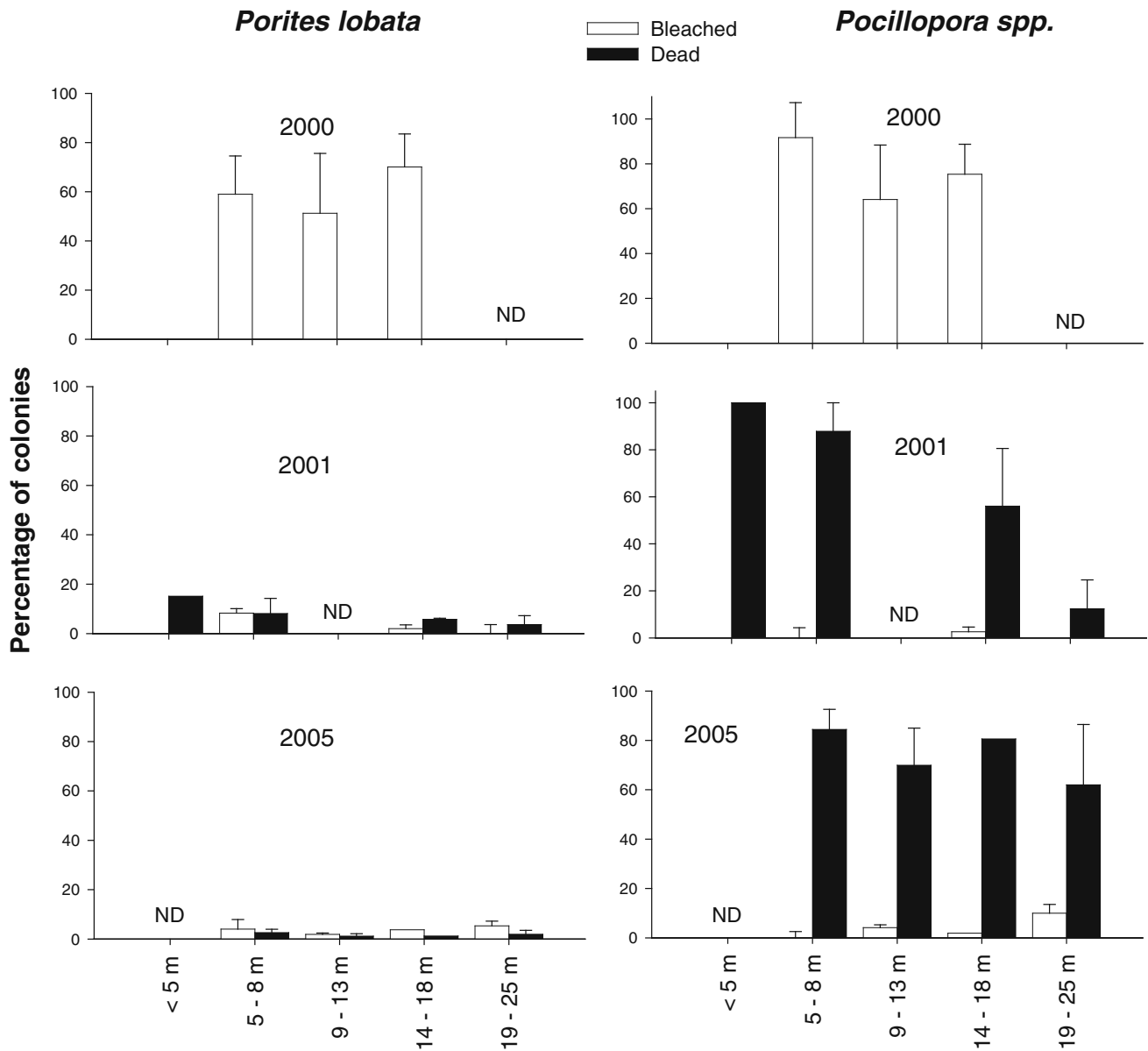


Fig. 5.36 Mean percentage (\pm SE) of *Porites* and *Pocillopora* colonies bleached or recently dead across varying subtidal depth zones. Error bars represent variation among sites 1–10s of km apart. ND no data available

Pocillopora and *Porites* are more likely to harbor associated fauna than their neighboring “healthy” (fully pigmented) counterparts. Moreover, the abundance and species richness of fauna are higher on bleached (but live) corals (Fig. 5.37). Increase in macroalgal abundance also occurs during years of more prevalent bleaching, though susceptibility of corals to overgrowth varied among sites and

appears to be associated with local coral energy reserves (lipid concentration, Fig. 5.38). These results suggest that disruptive stress that causes coral bleaching changes the habitat-modifying role of branching and massive corals as well as the intensity of competition with algae under natural conditions in the field. These hypotheses are currently being tested.

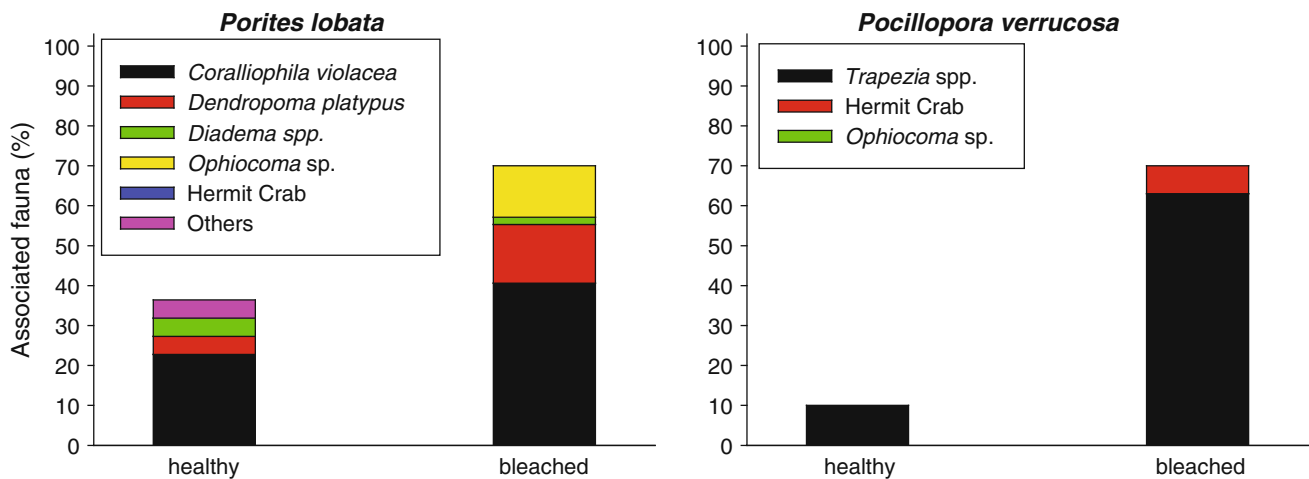


Fig. 5.37 Percentage of healthy and bleached *Porites* and *Pocillopora* colonies with diverse associated species of invertebrate functional groups. ‘Others’ include infrequently observed taxa such as sponges, polychaete worms, turrid snails, caridean shrimp, and non-trapeziid

brachyuran crabs. A minimum of 50 colonies of each species and condition was assessed at mid-depth (10–15 m) across four sites on the west coast of Rapa Nui

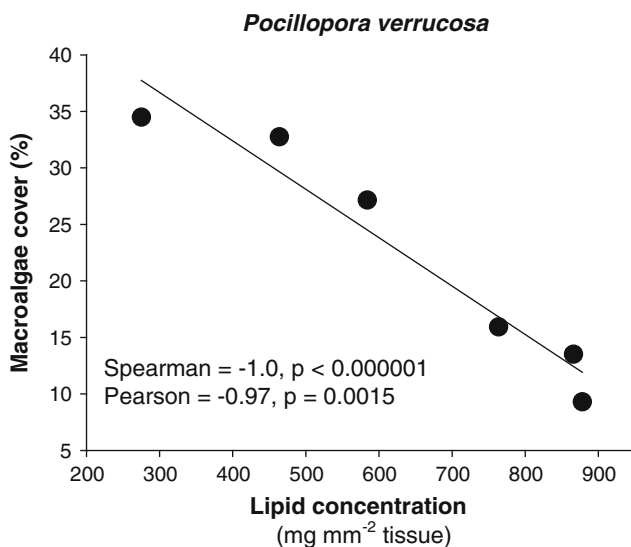


Fig. 5.38 Relationship between mean percentage cover of macroalgae and mean lipid concentration of *Pocillopora verrucosa* colonies at mid-depth (10–15 m) habitats during years of mild bleaching (>30 % cover bleached corals) across western and northeastern shores of Rapa Nui. Symbols are sites 1 to 10s of km apart

5.4 Conclusions

Coral reef-building potential exists across the entire eastern tropical Pacific region, from the Gulf of California to Rapa Nui, regardless of low species richness at the northern and southern latitudinal limits of coral distribution. With the exception of newly discovered ETP coral formations, e.g. El Salvador and Nicaragua, and small habitable sites, e.g.

Malpelo Island, coral species richness is relatively uniform at 20–25 species per locality across the eastern Pacific region. The presently recognized total species count of zooxanthellate corals in the ETP region is 47.

Concentrated reef development occurs chiefly on islands of the continental shelf and on distant oceanic islands, but also at some environmentally suitable coastal areas such as Huatulco (Mexico), along the shores bordering the southern Gulf of Papagayo and Golfo Dulce (Costa Rica), and at Machalilla (Ecuador). Massive colonies of *Porites*, *Pavona*, and *Gardineroseris* are usually the dominant reef-building corals in ETP oceanic island environments such as Clipperton Atoll, Isla del Coco, the northern Galápagos Islands, and Rapa Nui.

Coral species richness is relatively high in the northern latitudes of mainland Mexico and offshore in the Revillagigedo Islands, and farther south in the equatorial eastern Pacific, from Costa Rica to Ecuador. Reef building occurs in upwelling centers (Huatulco, Gulf of Papagayo, Gulf of Panama), and in several areas of the equatorial eastern Pacific, which experience aperiodic severe ENSO disturbances.

Thomas Dana’s hypothesis of the dispersal of central/west Pacific corals into the ETP via the NECC is indirectly supported by analyses of faunal affinities and molecular genetic evidence. Species-rich Palmyra Atoll in the northern Line Islands is in the path of the easterly flowing NECC and could likely serve as a source of coral propagules to be transported to the equatorial eastern Pacific. Surface current transport from the Line Islands via the NECC to Clipperton Atoll (a stepping stone) is also possible.

Potentially important corallivore and bioeroder species have caused greatest damage to corals after El Niño

warming/bleaching events. The crown-of-thorns seastar *Acanthaster planci* is moderately abundant in only three ETP areas—Gulf of California and Revillagigedo Islands (Mexico), and Gulf of Chiriquí (Panama), and has not been reported to overwhelm coral cover anywhere in the ETP. Population outbreaks of the black long-spined sea urchin, *Diadema mexicanum*, have been reported at several localities, usually following disturbances and high coral mortality, resulting in extensive bioerosion of reef frameworks and massive coral colonies. The slate pencil sea urchin *Eucidaris galapagensis* occurs at high population densities in the southern Galápagos Islands, where it has eroded reef frameworks and prevented the recovery of coral communities devastated by the severe 1982–83 El Niño event.

Competition for space among corals and macroalgae has been documented locally in El Salvador (*Acanthophora spicifera* vs. *Porites lobata*), Costa Rica (*Caulerpa sertularioides* vs. *Pocillopora* spp., *Psammocora stellata*) and Panama (*Caulerpa racemosa*, *C. sertularioides* vs. *Psammocora stellata*). Coral patches are sometimes overgrown and killed by algae, but such competitive dominance is usually ephemeral, with plant cover eventually subsiding and corals re-populating previously occupied substrates.

Despite diverse natural physical and biotic disturbances at both local (e.g., upwelling centers, bioeroder outbreaks) and regional (severe ENSO episodes) scales, ETP coral communities have generally demonstrated a high degree of resilience, with robust recovery at several localities in recent years.

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Appendix: Coral Species Presence (1)/Absence (0) Data Matrix

Presence/absence of zooxanthellate scleractinian corals at 15 eastern Pacific and four central/south Pacific localities. Recent sources for each locality are listed in the caption to Fig. 5.7. No species re-assignments were attempted for the central/south Pacific taxa listed. *Pocillopora verrucosa* and *Pocillopora elegans* are treated as separate species. GOC, Gulf of California; TMM, tropical mainland Mexico; REV, Revillagigedo Islands; CLP, Clipperton Atoll; ELS, El Salvador; NIC, Nicaragua; CRM, Costa Rican mainland; COC, Cocos Islands; PAN, Panama; COL, Colombia; MAL, Malpelo Island; ECD, Ecuador; GLS, southern Galápagos Islands; GLN, northern Galápagos Islands; RPN, Rapa Nui (Easter Island); HAW, Hawaii Is., including the NW Is.; JOH, Johnston Atoll; PIT, Pitcairn Islands, including Pitcairn, Henderson, Oeno, and Ducie Islands; PAL, Palmyra Atoll.

Species	Locality																		
	GOC	TMM	REV	CLP	ELS	NIC	CRM	COC	PAN	COL	MAL	ECD	GLS	GLN	RPN	HAW	JOH	PIT	PAL
<i>Acropora abrotanoides</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
<i>Acropora aculeus</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Acropora acuminata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
<i>Acropora aspera</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Acropora austere</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
<i>Acropora cerealis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
<i>Acropora</i> cf. <i>solitaryensis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
<i>Acropora clathrata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Acropora cymbicyathus</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Acropora cytherea</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	1
<i>Acropora delicatula</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Acropora digitifera</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
<i>Acropora divaricata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
<i>Acropora elseyi</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1
<i>Acropora florida</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Acropora formosa</i> ^a	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Acropora gemmifera</i> ^b	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	1
<i>Acropora glauca</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Acropora globiceps</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
<i>Acropora granulosa</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Acropora humilis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	1
<i>Acropora hyacinthus</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
<i>Acropora latistella</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
<i>Acropora listeri</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
<i>Acropora longicyathus</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Acropora loripes</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	1
<i>Acropora lutkeni</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
<i>Acropora microclados</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
<i>Acropora microphthalma</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
<i>Acropora monticulosa</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Acropora multiacuta</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Acropora nana</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Acropora nasuta</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	1
<i>Acropora palmerae</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0

(continued)

Species	Locality																		
	GOC	TMM	REV	CLP	ELS	NIC	CRM	COC	PAN	COL	MAL	ECD	GLS	GLN	RPN	HAW	JOH	PIT	PAL
<i>Acropora paniculata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	1
<i>Acropora pocilloporina</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
<i>Acropora polymorpha</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Acropora polystoma</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Acropora reticulata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Acropora retusa</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
<i>Acropora robusta</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
<i>Acropora samoensis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
<i>Acropora secale</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
<i>Acropora selago</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	1
<i>Acropora</i> sp. (Line and Phoenix Is.)	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Acropora</i> sp. (slanted cones)	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Acropora</i> sp. B	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
<i>Acropora</i> sp. C	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
<i>Acropora spicifera</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Acropora squarrosa</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Acropora striata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
<i>Acropora subulata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
<i>Acropora syringodes</i> ^c	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Acropora tenuis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Acropora tortuosa</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
<i>Acropora valida</i>	0	0	0	0	0	0	0	0	0	1	0	0	0	0	1	1	1	1	1
<i>Acropora vaughni</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
<i>Acropora verweyi</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Acropora yongei</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
<i>Alveopora tizardi</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
<i>Alveopora verrilliana</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Astreopora</i> cf. <i>moretonensis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
<i>Astreopora expansa</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Astreopora gracilis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Astreopora listeri</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Astreopora myriophthalma</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
<i>Astreopora ocellata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0

(continued)

Species	Locality																		
	GOC	TMM	REV	CLP	ELS	NIC	CRM	COC	PAN	COL	MAL	ECD	GLS	GLN	RPN	HAW	JOH	PIT	PAL
<i>Astreopora suggesta</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Caulastrea cf. furcata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
<i>Coscinaraea columna</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0
<i>Coscinaraea wellsii</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
<i>Ctenactis echinata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Cycloseris curvata</i>	1	1	1	0	0	0	1	1	1	1	0	1	1	1 ⁱ	0	0	0	0	0
<i>Cycloseris cyclolites</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Cycloseris fragilis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Cycloseris sp.</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Cycloseris tenuis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1
<i>Cycloseris vaughani</i>	1	0	1	0	0	0	0	0	0	0	0	0	0	0	1	1	1	1	0
<i>Cyphastrea ocellina</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0
<i>Cyphastrea serailia</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
<i>Diaseris distorta</i>	1	1	1	0	0	0	1	1	1	1 ⁱ	0	1	1	0	0	1	0	0	0
<i>Echinophyllia aspera</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Echinophyllia echinata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Echinophyllia sp.</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Favia amicornum</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Favia chinensis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Favia fava</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Favia matthäi</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
<i>Favia pallida</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Favia rotumana</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
<i>Favia rotundata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Favia speciosa</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Favia stelligera</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
<i>Favites abdita</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Favites flexuosa</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Favites halicora</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Favites pentagona</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Favites russelli</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Fungia concinna</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Fungia fungites</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Fungia granulosa</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1
<i>Fungia horrida</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Fungia moluccensis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Fungia paumotensis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1

(continued)

Species	Locality																		
	GOC	TMM	REV	CLP	ELS	NIC	CRM	COC	PAN	COL	MAL	ECD	GLS	GLN	RPN	HAW	JOH	PIT	PAL
<i>Fungia repanda</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Fungia scutaria</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	1
<i>Gardineroseris planulata</i>	0	1	0	0	0	1	1	1	1	1	1	1	1	1	0	1	0	0	1
<i>Goniastrea australensis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
<i>Goniastrea edwardsi</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Goniastrea pectinata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Goniastrea retiformis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Halomitra pileus</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Herpolitha limax</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Hydnophora exesa</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Hydnophora microconos</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Hydnophora pilosa</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Hydnophora rigida</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Isopora brueggemanni</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1
<i>Isopora cuneata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Isopora palifera</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Leptastrea agassizi</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1
<i>Leptastrea bewickensis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1
<i>Leptastrea hawaiiensis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
<i>Leptastrea pruinosa</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	1
<i>Leptastrea purpurea</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	1	1
<i>Leptastrea</i> sp. A	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Leptastrea</i> sp. B	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Leptastrea</i> sp. C	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Leptastrea transversa</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	1
<i>Leptoria phrygia</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Leptoseris hawaiiensis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	0
<i>Leptoseris incrustans</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	0
<i>Leptoseris mycetoseroides</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1
<i>Leptoseris papyracea</i>	1	1	0	0	0	0	1	1	1	1	0	1	0	0	0	1	0	0	0
<i>Leptoseris scabra</i>	0	0	0	1	0	0	1	1	0	0	0	0	0	1	1	1	1	0	0
<i>Leptoseris solida</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1	0
<i>Leptoseris tubulifera</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
<i>Lobophyllia corymbosa</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1

(continued)

Species	Locality																		
	GOC	TMM	REV	CLP	ELS	NIC	CRM	COC	PAN	COL	MAL	ECD	GLS	GLN	RPN	HAW	JOH	PIT	PAL
<i>Lobophyllia hemprichii</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
<i>Merulina ampliata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Montastrea annuligera</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Montastrea curta</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
<i>Montipora aequituberculata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
<i>Montipora australiensis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
<i>Montipora bilaminata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
<i>Montipora caliculata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
<i>Montipora capitata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	1
<i>Montipora complanata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
<i>Montipora crassituberculata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
<i>Montipora danae</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Montipora dilatata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1
<i>Montipora efflorescens</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Montipora elschneri</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Montipora flabellata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	1
<i>Montipora foliosa</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Montipora foveolata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
<i>Montipora grisea</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
<i>Montipora hoffmeisteri</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1
<i>Montipora informis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Montipora lobulata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
<i>Montipora millepora</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Montipora monasteriata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Montipora nodosa</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
<i>Montipora patula</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	1
<i>Montipora peltiformis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Montipora socialis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Montipora</i> sp. 4 cf. <i>incrassata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	0
<i>Montipora tuberculosa</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1	1

(continued)

Species	Locality																		
	GOC	TMM	REV	CLP	ELS	NIC	CRM	COC	PAN	COL	MAL	ECD	GLS	GLN	RPN	HAW	JOH	PIT	PAL
<i>Montipora venosa</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
<i>Montipora verrilli</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	1
<i>Montipora verrucosa</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
<i>Pachyseris</i> sp.	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Pachyseris speciosa</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Parahalomitra robusta</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Pavona cactus</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Pavona chiriquiensis</i>	0	0	0	0	0	0	1	1	1	1	1	1	1	0	0	0	0	0	1
<i>Pavona clavus</i>	1	1	1	1	0	1	1	1	1	1	1	1	1	1	0	1	1	0	0
<i>Pavona divaricata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Pavona duerdeni</i>	1	1	1	1	0	0	0	0	1	0	0	0	0	0	0	1	1	0	1
<i>Pavona explanulata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Pavona frondifera</i>	0	0	0	0	0	0	1	1	1	1	0	0	0	0	0	0	0	0	1
<i>Pavona gigantea</i>	1	1	1	1	1	1	1	1	1	1	1	1	1	1	0	0	0	0	0
<i>Pavona maldivensis</i>	0	0	1	1	0	0	1	1	1	1	1	0	0	1	0	1	1	1	1
<i>Pavona minuta</i>	0	0	1	1	0	0	0	1	1	0	0	0	0	0	0	0	0	0	1
<i>Pavona</i> sp. cf. <i>pollicata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
<i>Pavona</i> sp. 1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
<i>Pavona varians</i>	0	1	1	1	0	1	1	1	1	1	1	1	1	1	0	1	1	1	1
<i>Platygyra daedalea</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
<i>Platygyra lamellina</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Platygyra pini</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Platygyra ryukyuensis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Platygyra sinensis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Plesiastrea versipora</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
<i>Pocillopora damicornis</i>	1	1	1	0	1	1	1	1	1	1	0	1	1	1	1	1	1	1	1
<i>Pocillopora danae</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0
<i>Pocillopora effusus</i>	0	1	0	1	1	0	0	0	0	1	1	1	1	1	0	0	0	0	0
<i>Pocillopora elegans</i>	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	0	0	1	0
<i>Pocillopora eydouxi</i>	1	1	1	1	0	0	1	1	1	1	1	1	1	1	1	1	1	1	1
<i>Pocillopora inflata</i>	0	1	0	0	0	0	1	1	1	0	0	1	1	1	0	0	0	0	0
<i>Pocillopora laysanensis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
<i>Pocillopora ligulata</i>	0	0	0	0	0	0	0	0	0	1	0	1	1 ^j	1 ^j	1	1	1	0	0

(continued)

Species	Locality																		
	GOC	TMM	REV	CLP	ELS	NIC	CRM	COC	PAN	COL	MAL	ECD	GLS	GLN	RPN	HAW	JOH	PIT	PAL
<i>Pocillopora meandrina</i>	1	1	1	1	1	0	1	1	1	1	0	1	1	1	1	1	1	1	1
<i>Pocillopora molokensis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	0
<i>Pocillopora</i> sp.	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Pocillopora</i> sp. 1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
<i>Pocillopora</i> sp. 2	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0
<i>Pocillopora</i> sp. 11 cf. <i>capitata</i>	1	1	1	0	1	0	1	1	1	1	1	1	1	1	0	1	1	0	1
<i>Pocillopora verrucosa</i>	1	1	1	1	1	1	1	0	1	1	1	1	1	1	1	1	0	1	1
<i>Pocillopora woodjonesi</i>	0	0	1	1	0	0	0	0	0	1	0	0	0	1	0	0	0	1	0
<i>Pocillopora zelli</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	1
<i>Porites</i> aff. <i>Annae</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	1	0
<i>Porites annae</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Porites amaudi</i>	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Porites australiensis</i>	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1
<i>Porites baueri</i>	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Porites brighami</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
<i>Porites compressa</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
<i>Porites duerdeni</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
<i>Porites evermanni</i>	0	1	0	1	0	0	1	1	1	1	0	1	0	1	0	1	1	0	1
<i>Porites hawaiiensis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
<i>Porites lichen</i>	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Porites lobata</i>	0	1	1	1	1	0	1	1	1	1	1	1	1	1	1	1	1	1	1
<i>Porites lutea</i> ^d	0	1	1	1	0	0	0	0	0	0	0	0	0	0	0	0	1	1	1
<i>Porites murrayensis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Porites panamensis</i>	1	1	1	0	1	1	1	0	1	1	0	1	0	0	0	0	0	0	0
<i>Porites pukoensis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
<i>Porites rus</i>	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	1	0	0	1
<i>Porites solida</i> ^e	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Porites</i> sp.	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Porites studeri</i> ^f	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
<i>Porites superfusa</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Porites sverdrupi</i>	1	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
<i>Porites vaughani</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Psammocora albopicta</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
<i>Psammocora brighami</i>	1	1	0	0	0	0	0	0	1	1	0	1	1	1	0	0	0	0	0
<i>Psammocora contigua</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Psammocora explanulata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
<i>Psammocora haimeana</i> ^g	1	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1

(continued)

Species	Locality																		
	GOC	TMM	REV	CLP	ELS	NIC	CRM	COC	PAN	COL	MAL	ECD	GLS	GLN	RPN	HAW	JOH	PIT	PAL
<i>Psammocora nierstraszi</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	0	1
<i>Psammocora obtusangula</i>	0	0	0	0	1	1	1	0	1	1	0	0	0	0	0	0	0	1	0
<i>Psammocora profundacella</i>	0	1	1	1	0	0	0	0	0	1	0	0	0	0	0	0	0	1	1
<i>Psammocora stellata</i>	1	1	1	1	1	1	1	1	1	1	0	1	1	1	1	1	1	0	1
<i>Psammocora superficialis</i> ^b	1	1	1	1	0	0	1	1	1	1	0	1	1	1	1	0	0	0	1
<i>Psammocora verrilli</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0	0	0
<i>Sandalolitha robusta</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Scolymia</i> cf. <i>vitiensis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
<i>Siderastrea glynni</i>	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
<i>Stylocoeniella guentheri</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	0
<i>Stylophora pistillata</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Symphyllia recta</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Turbinaria frondens</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Turbinaria reniformis</i>	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
<i>Turbinaria</i> sp.	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1
	19	26	25	21	11	10	25	22	27	28	13	23	20	22	15	62	44	87	163

^aWallace (1999) recognizes this species as *Acropora muricata*

^bWalsh et al. (2013)

^cA junior synonym of *A. longicyathus*

^dThis species is probably *Porites evermanni*

^eA frequently misnamed species

^fType locality in Hawaii

^gFrequently confused with *Psammocora nierstraszi*

^hFrequently confused with *Psammocora explanulata*

ⁱOnly dead skeletons present

^jRequires confirmation (see Hickman 2008)

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