Title: Ecological and methodological drivers of species' distribution and phenology responses
to climate change
Running head: Species' responses to climate change
Christopher J. Brown ¹ *, Mary I. O'Connor ² , Elvira S. Poloczanska ^{1,3} , David S. Schoeman ⁴ ,
Lauren B. Buckley ⁵ , Michael T. Burrows ⁶ , Carlos M. Duarte ⁷ , Benjamin S. Halpern ^{8,9,10} , John
M. Pandolfi ¹¹ , Camille Parmesan ^{12, 13} , Anthony J. Richardson ^{3,14}
1. The Global Change Institute, The University of Queensland, St Lucia, Queensland,
Australia
2. Department of Zoology and Biodiversity Research Centre, University of British Columbia,
Vancouver, BC, Canada V6T1Z4
3. CSIRO Oceans and Atmosphere, EcoSciences Precinct, Dutton Park, Brisbane, QLD 4102,
Australia
4. School of Science and Engineering, University of Sunshine Coast, Maroochydore, DC,
Qld, Australia
5. Department of Biology, University of Washington, Seattle, WA 98115-1800.
6. Department of Ecology, Scottish Association for Marine Science, Marine Institute, Oban,
Argyll, UK, PA37 1QA, UK
7. King Abdullah University of Science and Technology (KAUST), Red Sea Research Center
(RSRC), Thuwal, 23955-6900, Saudi Arabia

23	8. National Center for Ecological Analysis and Synthesis, 735 State St. Suite 300, Santa
24	Barbara, CA, 93101 USA
25	9. Bren School of Environmental Science and Management, University of California, Santa
26	Barbara, CA 93106 USA
27	10. Imperial College London, Silwood Park Campus, Buckhurst Road, Ascot SL57PY, UK
28	11. ARC Centre of Excellence for Coral Reef Studies and School of Biological Sciences, The
29	University of Queensland, St Lucia, Queensland 4072, Australia
30	12. Marine Institute, Plymouth University, Drakes Circus, Plymouth, Devon PL4 8AA
31	13. Department of Geological Sciences, University of Texas at Austin, Austin, Texas, USA
32	14. Centre for Applications in Natural Resource Mathematics, School of Mathematics and
33	Physics, The University of Queensland, St Lucia, Queensland, 4072, Australia
34	
35	* Corresponding author.
36	Ph: +61 (7) 37 359 268
37	Email: christopher.brown@griffith.edu.au
38	Current address: Australian Rivers Institute, Griffith University, 170 Kessels Road, Nathan,
39	Queensland, Australia
40	
41	Keywords: Global warming, range-shift, range-edge, fishing, publication bias, marine
42	ecosystem, season, meta-analysis, time-series, tropics.

43 Type of paper: Primary Research Article

44 Abstract

45 Climate change is shifting species' distribution and phenology. Ecological traits, such as 46 mobility or reproductive mode, explain variation in observed rates of shift for some taxa. 47 However, estimates of relationships between traits and climate responses could be influenced 48 by how responses are measured. We compiled a global dataset of 651 published marine 49 species' responses to climate change, from 47 papers on distribution shifts and 32 papers on 50 phenology change. We assessed the relative importance of two classes of predictors of the 51 rate of change, ecological traits of the responding taxa and methodological approaches for 52 quantifying biological responses. Methodological differences explained 22% of the variation 53 in range shifts, more than the 7.8% of the variation explained by ecological traits. For 54 phenology change, methodological approaches accounted for 4% of the variation in 55 measurements, whereas 8% of the variation was explained by ecological traits. Our ability to 56 predict responses from traits was hindered by poor representation of species from the tropics, 57 where temperature isotherms are moving most rapidly. Thus, the mean rate of distribution 58 change may be underestimated by this and other global syntheses. Our analyses indicate that 59 methodological approaches should be explicitly considered when designing, analysing and 60 comparing results among studies. To improve climate impact studies, we recommend that: (1) 61 re-analyses of existing time-series state how the existing datasets may limit the inferences 62 about possible climate responses; (2) qualitative comparisons of species' responses across 63 different studies be limited to studies with similar methodological approaches; (3) meta-64 analyses of climate responses include methodological attributes as covariates and; (4) that 65 new time series be designed to include detection of early warnings of change or ecologically 66 relevant change. Greater consideration of methodological attributes will improve the accuracy 67 of analyses that seek to quantify the role of climate change in species' distribution and 68 phenology changes.

69 Introduction

70	A large number of marine (Poloczanska et al., 2013) and terrestrial (Parmesan & Yohe,
71	2003) species have shifted their distributions and phenology in recent decades, indicating that
72	climate change is driving a global biological response. For example, recent climate change
73	has driven the invasion of pest species (Ling et al., 2008), contributed to declines in
74	commercially important fish species (Beaugrand & Kirby, 2010) and appears to be increasing
75	mismatch in the seasonal timing between predators and their prey (Edwards & Richardson,
76	2004, Barbraud & Weimerskirch, 2006).
77	Despite an overall broad consistency in species' responses to climate change, there is
78	considerable variability in magnitudes and patterns of responses (Parmesan, 2007,
79	Poloczanska et al., 2013, Sunday et al., 2015). Variability poses a challenge to ecological
80	science and management of species impacted by climate change, because it hinders
81	predictions of future responses. Analyses across many species have examined how
82	combinations of taxonomic identity, ecological traits and local environmental variables may
83	explain variability in responses (Perry et al., 2005, Wolkovich et al., 2012, Pinsky et al.,
84	2013, Poloczanska et al., 2013, Sunday et al., 2015). All of these approaches base their
85	inferences on standardised measures of distribution and phenology, yet observed responses to
86	climate change may also depend on how distribution and phenology are measured
87	(Wolkovich <i>et al.</i> , 2012).
88	Measurements of distribution and phenology are influenced by a suite of decisions that are
89	made in two stages of all studies: their sampling design and data analysis (Brown et al., 2011,
90	Bates et al., 2014a). In the sampling design phase researchers decide how species' variables
91	are measured. For example, distribution can be measured as mean latitude of a populations'
92	geographic extent (e.g. Perry et al., 2005) or by measuring the most extreme latitudes where
93	a species is found (e.g. Robinson et al., 2015). Similarly, the phenology of breeding events

94	can be measured by censusing a population throughout a season to determine the peak
95	breeding date, or as the first and last individuals to breed (e.g. Fitter & Fitter, 2002, Barbraud
96	& Weimerskirch, 2006). Measures of distribution and phenology based on the most extreme
97	individuals rather than variables that represent the distribution of individuals within a
98	population may lead to very different estimates of climate change response rates. For
99	example, single individuals may by chance have extreme responses (Brown et al., 2011) and
100	measurements using single individuals are susceptible to detection biases (Cook et al., 2012,
101	Bates et al., 2015).
102	In the analysis phase, most marine climate change studies include only a single predictor –
103	temperature – to explain changes in distribution or phenology, and thus do not explicitly
104	consider other drivers of change (Brown et al., 2011). It is unknown whether studies that do
105	not account for other potential anthropogenic drivers, such as fishing, eutrophication and
106	habitat loss, could lead to higher estimates of impacts of climate change. For example, an
107	investigation of changes in the distribution of North Sea cod showed fishing pressure explains
108	part of the observed biological changes (Engelhard et al., 2014).
109	A greater understanding of how different methodological approaches affect detection of
110	observed responses to climate change will benefit climate change ecology in four main ways.
111	First, studies that analyse existing data sets to test for climate impacts, need to account for
112	historical choices made about field data collection that could limit the ability to detect
113	species' responses to climate change. For instance, uneven sampling effort along coasts
114	means museum records of species occurrences may misrepresent historical range boundaries
115	(Shoo et al., 2006, Przesławski et al., 2012). Second, many regional studies compare rates of
116	change with other studies in their discussions of how ecological traits influence a species'
117	response (e.g. Richardson, 2008). Comparisons of change may also need to consider
118	differences in methodological approaches across studies, such as how occurrence data are

119	used to estimate ranges. Third, new time-series are being initiated with the aim of measuring
120	future distribution and phenology change (e.g. Robinson et al., 2015). Greater understanding
121	of how different methodological approaches can affect measured responses to climate change
122	could assist the design of new time-series. For instance, inconsistent sampling effort through
123	time may bias measured rates of change (Bates et al., 2015). Finally, meta-analyses of
124	existing studies will produce more accurate estimates if they standardise for study differences,
125	or constrain comparisons to be among studies with similar methods (Parmesan, 2007,
126	Przeslawski et al., 2012). For instance, differences in rates of range shifts among European
127	butterflies, birds and plants could be a consequence of taxonomic identity, geographic biases
128	or differences in the metrics used (Parmesan and Yohe 2003).
129	Here we examine potential causes of variability in observed marine species distribution and
130	phenology responses to climate change using a meta-analysis. First, we ask whether
131	measurements of distribution and phenology change from the peer-reviewed literature are
132	representative of different taxa, oceans and methodological approaches. Then we conduct a
133	meta-analysis on rates of response, to ask how ecological traits, study design and
134	measurement approaches influence rates of distribution and phenology change. This enables
135	us to ask how study measurement approaches may bias measured rates of change and affect
136	inferences about the biological drivers of change. Finally, we investigate how different
137	measurement approaches are used for different taxa and discuss the implications of
138	measurement biases on the global understanding on climate change responses.
139	Methods

140 Database

141 We analysed the database of peer-reviewed studies of observed impacts of climate change on

- 142 marine organisms compiled by Poloczanska et al. (2013). We used a subset of 61 studies
- 143 where rates of range change in phenology or distribution were reported or could be calculated,

144	and updated the database with a further 18 studies published in 2011-2014, for a total of 79
145	studies (Supplemental online tables). In the original database and the update, we made every
146	effort to include every dataset and study that met our criteria; thus we believe this to be a
147	comprehensive dataset rather than a small subset of available data.
148	Three criteria were applied for inclusion of peer-reviewed studies in the database: (1) Authors
149	must have inferred or directly tested for trends in marine ecological and climate variables
150	from field observations; (2) observations spanned at least 19 years; and (3) studies included
151	data after 1990. Studies relying purely on modelling or experimental data were excluded.
152	Data spanning at least twenty years is a common cut-off used in syntheses of climate change
153	impacts (Rosenzweig et al., 2008). This length of time gives confidence that biological
154	changes might be driven by long-term climate change rather than yearly climate variability.
155	We chose to use nineteen years as the minimum time span, since several large studies had
156	durations of 19 years. Requiring studies to also have data after 1990 ensures that there are
157	observations in recent decades when the anthropogenic climate signal has been strongest.
158	Inclusion of all peer-reviewed studies resulted in some pseudo-replication of observations. In
159	some cases, multiple studies analysed the same raw dataset, and in other cases multiple
160	metrics of change were reported for a single species in a region. In such cases, only the most
161	recent study for a given data-set was included in the main database. Non-independent
162	observations were removed from the database, using a decision tree based on data and
163	analysis quality (Poloczanska et al., 2013). Following this process 47 distribution studies with
164	485 observations of change and 32 phenology studies with 156 observations of change
165	remained and were included in our analyses.
166	Analysis of rates of change

First, we summarize measurements of distribution and phenology change by methodologicalattributes, taxon, latitude, and for phenology, season. We then conducted analyses to examine

Page 8 of 37

169 how rates of change were influenced by ecological traits and methodological approaches. 170 Rates of change, in kilometres per decade or days per decade, were obtained from individual 171 studies in the database, either directly as reported in the text, calculated from figures, or by 172 contacting the study's authors. Distribution changes were recorded as positive where they 173 were consistent with a response to climate change (generally polewards, but see Burrows et 174 al., 2011) and negative if they were not consistent with climate change. Phenology changes 175 were recorded as negative for shifts to earlier dates and positive for shifts to later dates. 176 We related rates of change to a suite of predictors using a generalised linear modelling 177 approach. The response variables were the rate of change in either distribution or phenology. 178 Predictors were divided into two categories: methodological approaches and ecological traits. 179 For methodological attributes we considered the frequency of sampling (continuous, irregular 180 or comparison of two periods). Fewer sampling points for studies that compare two periods of 181 time (e.g. repeating a historical survey), may mean less accurate (either higher or lower) rates 182 of change, because intervening fluctuations are ignored. We considered the number of species 183 in a study; studies reporting on more species were expected to have lower rates of change 184 because they are less likely to be influenced as strongly by publication bias (Parmesan, 2007). 185 We also considered whether non-climatic drivers of change were accounted for in the study. 186 We expect slower rates of change in studies with non-climatic drivers because confounding 187 influences on the response could explain some of the observed variation. For distribution, we 188 considered whether rates were generated from abundance (or relative abundance) metrics or 189 from presence data on species occurrence at sites. Occurrence-based data were expected to 190 have higher rates of change because they are more susceptible to the outlying influences of a 191 single individual. Similarly for phenology, we considered whether the magnitude of change in 192 timing was related to whether the measure was an abundance metric, or the timing of the most 193 extreme individual (e.g. first or last arrival – the temporal analogue of single sightings on a

194 range edge). Extreme timings were expected to have higher rates of change (Moussus *et al.*,195 2010).

196 For ecological traits we considered whether life-history development type (direct 197 development with no larval phase, meroplanktonic, planktonic), exploitation status 198 (commercially targeted or not), the mean latitude of the observations for a species and the 199 depth range of the organism (coastal, demersal or pelagic) could influence the rates of change 200 measured (Poloczanska et al., 2013). For the depth range, species were classified based on the 201 life-history stage that was studied and coastal species where those constrained to terrestrial 202 (seabirds), intertidal (e.g. barnacles) or near-shore (e.g. anemones) habitats (Poloczanska et 203 al., 2013). Additionally, for distribution we considered whether the measurement was made 204 for a leading (poleward) or trailing (equatorward) edge, or for the population centre. For 205 phenology, we considered the season of the measurement. Where available, ecological traits 206 were extracted during the review of each study, based on what the individual studies reported. 207 Latitude, range edge and season could also be considered as methodological approaches, 208 however we classified them as ecological traits because ecological expectations can be given 209 for their effect on climate responses (Davis et al., 2010). 210 We also included climate predictors in analyses: the velocity of climate change (km/decade)

211 for distribution and seasonal shift (days/decade) for phenology. The climate predictors

212 measure the expected rate of response if species are tracking thermal niches in space and time

213 (Loarie et al., 2009, Burrows et al., 2011). The indices were calculated for each study

following Burrows *et al.* (2011). In brief, we used a global database of monthly sea-surface

- temperatures, at a resolution of 1° (Rayner *et al.*, 2003). First, we spatially matched every
- species observation to a grid square. Where studies had a larger extent than a single grid-
- square, we matched them to a grid square at the centroid of a study's location, or the nearest
- 218 ocean cell if the centroid fell on land. We then calculated the decadal rate of temperature

219	change for each study's centroid using linear regressions of mean annual temperatures. The
220	time-period was chosen to match each study's duration. For distribution, we additionally
221	calculated the spatial gradient in temperature (degrees Celsius per kilometre) by taking the
222	mean temperature in each grid square across the each study's timespan, then calculating the
223	spatial gradient in temperature as the vector sum of the north-south and east-west components
224	of spatial change. For phenology, we additionally calculated the seasonal gradient in
225	temperature over the dates of each study, as the mean of the monthly temperature differences
226	over each season. Velocities were then calculated as the spatial or seasonal gradient in
227	temperature divided by the inter-annual trend (Burrows et al., 2011).
228	We used linear mixed-effects models to relate rates of change to the suite of methodological
229	and ecological predictors ('lme4' package in the R programming language (Bates et al.,
230	2014b)). Taxon was treated as a random effect because our main interest was to characterise
231	rates of change by ecological traits and measurement type, which are correlated with taxa. For
232	distribution and phenology we developed separate full models including all physical,
233	ecological and measurement predictors. Models were simplified using a step-wise selection
234	process, removing the variable that caused the greatest reduction in the Bayes Information
235	Criteria (BIC) at each step. The selection process stopped when no further predictors could be
236	removed without increasing the BIC. We chose to use the BIC over the Aikaike Information
237	Criteria because the BIC is less likely to include spurious predictors and it controls for sample
238	size (Burnham & Anderson, 2002). Thus, the final models could be considered conservative
239	in that they include only the strongest predictors of rates of response to climate change.
240	We plotted effect sizes for predictors included in the final models on term plots. Term plots
241	illustrate the modelled effects of a predictor relative to the mean of all predicted values. Term
242	plots are an appropriate way to display effect sizes when there is no control treatment,
243	because comparisons can be made across all predictions. A positive value for an effect on the

244	term plot indicates that a predictor increases the rate of an organism's response to climate
245	change. A negative value indicates the effect slows an organism's response to climate change.
246	Terms were presented with confidence intervals, which were estimated using bootstrapping
247	(using the 'boot' package in R, see Canty and Ripley (2014)).
248	We estimated the relative importance of methodological approaches when compared to
249	ecological traits by comparing the proportions of variance explained by each set of predictors.
250	We estimated variance explained by either measurement approaches or ecological traits as the
251	difference between the marginal R ² statistic (Nakagawa & Schielzeth, 2013) for the model
252	with all significant predictors and a model without the respective variables.
253	Following the analysis, we examined in more detail how inferences drawn from analysis of
254	the database may be influenced by the available studies. Specifically, we plot the frequency of
255	measurement for the variables that were significant predictors of distribution and phenology
256	change by taxa.
257	Results
258	Summary of distribution and phenology observations

259 Across all the distribution and phenology studies there were many biases in study attributes

and methodologies (Fig. 1). Of 47 distribution studies and 32 phenology studies, only 15 and

261 6 respectively had more than one species, although only 2 distribution studies and no

262 phenology studies had >10 species (Fig. 1a). Out of 485 distribution measurements,

263 occurrence-based measures of distribution were slightly more common than abundance-based

264 measures (Fig. 1b). For 156 phenology observations, abundance-based measures were more

- common than measurements of extreme individuals (Fig 1b). 38% of distribution responses
- 266 compared two points in time, whereas 85% of phenology responses were measured from
- 267 continuous time series (Fig. 1c). Most distribution and phenology data were collected in mid-

to-high latitudes, with phenology records, in particular, biased towards the northern
hemisphere and a remarkable paucity of observations for tropical species (Fig. 1d). There was
considerable bias in taxonomic representation; 41% of distribution records were for bony fish
and 19% for benthic algae (Fig. 1e), whereas 33% of phenology records were for seabirds and
51% for phyto and zoo-plankton, which were both poorly represented in distribution records
(3% and 1% respectively). Most benthic taxa had distribution observations, but few
phenology observations. Measurements of phenology tended to be made in spring and

summer, but rarely in autumn or winter (Fig. 1f).

276 Effects of ecological traits and methodological attributes on distribution rates of change

277 The final model for the rate of distribution change included one ecological trait and two

278 methodological approaches (Table 1, Fig 2). Estimates of change derived from irregular time-

series or those that were calculated by comparing two points in time tended to be faster than

280 continuous time-series (Figure 2). Occurrence-based measures of distribution change were

also faster than abundance-based measures. Demersal and pelagic species moved faster than

282 coastal species (intertidal species and seabirds). A model including these top-ranked

283 predictors suggests that phytoplankton have changed distributions faster than other taxa,

whereas benthic cnidarians and seabirds have changed the slowest (Fig. 2). The reduced

model explained 28% of the variance, with methodological approaches (sampling frequency

and type of measure) accounting for 22% of the variation in rates of change, and ecological

variables (depth zone) accounting for 7.8% (there was shared variance across methodological

and ecological variables, so the individual variables did not add up to the total variance

explained).

Ecological traits that were excluded from the final model included the range edge, which was not a parsimonious predictor of distribution change (Table 1). The velocity of climate change was also excluded from the final model, while the model estimated a positive effect of higher

velocities on distribution change, this effect was not strong. The number of species in each
study, a methodological variable, was also excluded from the final model, suggesting there is
no strong effect of publication bias in this analysis.

296 *Effects of ecological traits and methodological attributes on phenology rates of change*

297 The final reduced model for phenology change explained 14% of the variance in the data and

298 included four factors, timespan, season, inclusion of non-climatic factors and latitude (Table

299 1, Fig 3). It excluded sampling frequency, depth range, life-history development type,

300 exploitation status, seasonal climate shift, measurement type and the number of species in a

301 study. Studies that used shorter time-series were more likely to report earlier timings,

302 suggesting a slight publication bias, although the effect size was small. However, counter to

303 our expectations, studies that considered non-climatic factors estimated faster rates of change

than studies that did not. Phenological events at mid-latitudes were more likely to be slower

than at higher latitudes. The phenology of autumnal events typically shifted later, rather than

306 earlier. Overall, the effects of ecological traits and methodological attributes were small (8

and 4% of the variance respectively) compared to the random effect of taxa on rates of

308 change. Larval bony fish were most likely to be shifting events earlier, whereas, seabirds had

309 small changes in phenology or were likely to be shifting later.

310 *Differences in how responses are measured across taxa*

311 Next we examined how observations are distributed across taxa, ecological traits and

312 methodological approaches to explore the representativeness of climate research. We focus on

the ecological traits and methodological approaches that were significant predictors of rates of

314 change. Importantly, not all taxa had measurements with every methodological approach

315 (Figs 4 & 5). Lack of measurements indicates caution should be taken when extrapolating the

316 results of meta-analyses to poorly sampled taxa.

317 For distribution, occurrence-based measures (that tended to report larger distribution changes) 318 predominated over abundance-based measures. Most abundance-based measures came from 319 fish and larval fish studies, which typically use nets to sample species in fishery-related 320 surveys (Fig. 4). Occurrence-based measures were predominant in other taxa. Fishery-related 321 surveys also had many more continuous time-series, rather than comparisons of two points in 322 time. In particular, benthic molluses, benthic enidarians, macro-algae and other invertebrates 323 had no continuous time-series. 324 Although measurements of phenology change were faster in studies that considered non-325 climatic factors (Fig. 3a), there were very few studies (only 7) that considered non-climatic

327 climatic factors were mostly for seabirds, but there was also a small proportion for

328 phytoplankton and benthic crustaceans. All taxa were represented in data-sets with timespans

factors in their analyses (Fig. 5). Observations that come from studies that considered non-

329 of up to 50 years. Autumn and spring were also well represented, however many taxa did not

have phenological measurements in summer and winter. Most phenology records for most

331 species were at latitudes $>40^{\circ}$, only seabirds had measurements closer to the Equator.

332 Discussion and recommendations

326

333 The methodology used to standardize studies for meta-analyses can have considerable effects

334 on estimates for rates of response to climate change (Parmesan, 2007, Przesławski et al.,

2012). Typically, meta-analyses exclude some studies to achieve consistency, such as

excluding studies of single species to avoid publication bias (Parmesan & Yohe, 2003,

Parmesan, 2007, Poloczanska et al., 2013). Rather than excluding studies, we used linear

338 models to standardize for differences in approaches across studies. By including studies that

used different methods to measure change, we have quantified the size and direction of

- 340 methodological effects on estimates of distribution and phenology responses to climate
- 341 change. We found methodological differences explained 22% of the variation in range shifts,

342	more than the 7.8% of the variation explained by ecological traits. For phenology change,
343	methodological approaches accounted for 4% of the variation in measurements, whereas 8%
344	of the variation was explained by ecological traits. Our study bolsters other recent findings
345	that different approaches to observing a single pattern (e.g., a geographic distribution) can
346	lead to different estimates of change over time (Moussus et al., 2010, Wolkovich et al., 2012,
347	Bates et al., 2015). Below we discuss the implications of differences in study design and
348	biological traits on estimated rates of change.
349	Study design
350	We found studies comparing two points in time or using irregularly sampled time-series
351	measured greater rates of change than studies using continuous time-series. Continuous time-
352	series should quantify rates of change more accurately than infrequent sampling, because
353	infrequent samples confound short-term variability with long-term trends (Moussus et al.,
354	2010, Brown et al., 2011). Further, range shifts in response to climate change can be
355	confounded by inconsistent sampling effort when a species is unlikely to be detected at every
356	sampling event (Bates et al., 2015). Studies in our database that had infrequent sampling often
357	resurveyed sites that were sampled historically, so our result may also suggest some
358	publication and study-site selection bias towards places where ecological change has been
359	greatest.
360	Historical comparisons (i.e. resurveying) are an important way to create long-term studies,
361	where there has not been funding to support long-term sampling (e.g. Southward et al., 2005,
362	Przeslawski et al., 2012, Robinson et al., 2015). Studies of fish were more likely to have
363	continuous time-series, presumably because of their economic importance, whereas

364 observations for other taxa often came from comparisons of two points in time. We encourage

- authors to look for and publish resurveys of historical sampling, regardless of whether there
- 366 have been considerable changes in distribution, to help overcome potential publication biases.

367 Efforts to digitise and publish historical datasets (e.g. Engelhard et al., 2014), combined with 368 the growth of data journals with the mandate that data generated using public funds must be 369 made available, may lead to many such data sets surfacing in the future, providing a richer 370 and less-biased basis to assess responses to climate change. 371 Abundance-based estimates of distribution change were slower than occurrence-based 372 measures. Occurrence-based measures can be influenced by responses of single individuals or 373 by detectability of a species, so we expected their observed rate of change to be greater 374 (Brown et al., 2011, Bates et al., 2015). Occurrence-based measures may be more likely to 375 detect change, but also more susceptible to spurious affects. Occurrence and abundance 376 measures also reflect different aspects of distribution change (Bates *et al.*, 2014a). 377 Occurrence-based measures are sensitive to range expansion, whereas abundance-based 378 measures better reflect population establishment. As such, our analysis suggests that 379 population establishment occurs much more slowly than range expansion – taking the 380 difference in rate of change between abundance and occurrence-based measures, the analysis 381 suggests on average a lag of about 140 km/decade, which is of greater magnitude than rates of 382 change in individual species' range centres (Poloczanska et al., 2013). Further, this result 383 indicates that caution should be taken when extrapolating rates of change across different 384 locations. Ranges may expand rapidly as few individuals of a species occupy areas it was 385 previously absent from, but population establishment may follow more slowly (Bates et al., 386 2014a). The pattern of range expansion and population establishment is particularly important 387 when managing ecosystems dynamically as communities move into novel configurations with 388 climate change (Graham et al., 2014). 389 Studies with single species or short time-series are often excluded from meta-analyses 390 because of perceived publication bias toward publishing results consistent with climate

391 change (Parmesan & Yohe, 2003, Parmesan, 2007). Based on the analysis of length of study

as a predictor, we found no publication bias in distribution studies and only a small bias in
phenology studies. Publication bias may be less prevalent in marine than terrestrial studies
because overall there are more multi-species studies in marine ecosystems where sampling
methods tend to collect numerous organisms (e.g. fish and plankton) by nets (Richardson *et al.*, 2012).

397 We found that inclusion of non-climatic factors in the analysis increased the estimated rates of 398 phenology change, but had no effect on rates of distribution change. However, few studies 399 included non-climatic factors, so further investigation of how climate responses interact with 400 factors like fishing pressure and eutrophication is important. Phenology is sensitive to 401 multiple human impacts, and it may be that in the studies analysed here, those impacts are 402 also causing seasonal timings to occur earlier. Given the paucity of studies, further work is 403 required to assess the interaction between climate change and other variables (Parmesan et al., 404 2013).

405 *Ecological traits*

406 We found that pelagic and demersal species tended to move faster than coastal (inshore)

407 species. Coastal species such as kelps and rocky shore invertebrates may be less able to track

408 warming because their distributions are restricted to the coast, and hence subject to non-

409 climatic biogeographic barriers to simple latitudinal shifts, and their requirements for specific

410 largely rule out depth shifts (Broitman et al., 2008). For instance, limited availability of hard

substrates on Australia's East coast may limit pole-wards migration of rocky intertidal

412 organisms (Poloczanska et al., 2011).

413 Contrary to previous studies for terrestrial and marine ecosystems, we did not find that

414 leading edges moved faster than trailing edges (Parmesan et al., 1999, Sunday et al., 2012,

415 Poloczanska *et al.*, 2013). Range edges are more likely to be detected with occurrence-based

416 measures, because species tend to be rarer at their range edges (Sagarin *et al.*, 2006),

therefore, the measurement type and the position of measurements within a species' range may be confounded. In our analysis sufficient measurements of species abundance at range edges were available to distinguish the effects of range position and measurement type on the rate of distribution shift. We found that occurrence based measurements were generally faster than abundance based-measurements. Thus if measurement differences are not accounted for, studies may overestimate the rate of shift at range edges, because of the bias toward measuring edges using single individuals.

424 *Gaps in climate change ecology studies*

425 Our analyses revealed that many gaps remain in our understanding of distribution and 426 phenology responses to climate change. Gaps are a consequence of not only geographical 427 biases in sampling effort, but also of how different taxa are studied. In particular a strong 428 geographic bias exists towards temperate regions, where data-collection efforts have 429 historically been the greatest. Tropical regions, grossly underrepresented in current studies 430 (Lenoir & Svenning, 2015), are expected to display some of the highest rates of distribution 431 change, particularly in marine systems (Burrows et al., 2014) and the distributions of tropical 432 taxa may be particularly susceptible to warming (Sunday et al., 2012, Molinos et al., 2015). 433 Moreover, the subtropical and tropical ocean represents \sim 70% of the global ocean surface, 434 implying that the current paucity of studies of distribution and phenology shifts in the tropics 435 affects our capacity to extrapolate available data to global rates. Global rates of distribution 436 change estimated here are therefore likely lower than those that would be inferred if the 437 studies were randomly distributed across latitudes. 438 There were few long-term phenology studies in the tropics. While seasonality in temperature

439 is weaker in the tropics and our results suggest phenology change is slower toward lower

440 latitudes, warming can still drive temporal changes in species abundance, for instance blooms

441 of dangerous jellyfish (Jacups, 2010). In addition to warming, seasonality along tropical

442 coastlines can be driven by precipitation (van Schaik *et al.*, 1993, Chambers *et al.*, 2013). For
443 example, the timing of juvenile prawn migrations from rivers to the ocean is related to
444 cumulative rainfall in tropical river basins (Staples & Vance, 1986). Phenological response to
445 precipitation is more difficult to predict than warming-related responses because species may
446 shift earlier or later (Chambers *et al.*, 2013). Impacts of climate change on tropical seasonality
447 have historically been neglected and warrant further studies.

448 There was systematic differences in types of observations across taxa, which suggests that we 449 have major gaps in our understanding of climate impacts. For instance, seabird studies that 450 measured changes in phenology with climate change were common, whereas there were only 451 two seabird studies of distribution that met the criteria for inclusion in our database. This is 452 likely to be because seabirds are most easily sampled at breeding colonies where there have 453 been long-standing monitoring programs (e.g. Barbraud & Weimerskirch, 2006), rather than 454 during their extensive foraging forays. In the future, the extensive and ongoing tracking 455 information collected using satellite tags should provide long-term information on distribution 456 shifts in feeding distributions, and potentially on shifts in breeding sites. In contrast to seabird 457 studies, studies of fish distribution were common, and studies of fish phenology rare. Fish 458 studies in the database often used fisheries data-sets for analyzing climate patterns. Fisheries-459 related surveys are large-scale, regular (usually annual) surveys of abundance indices. They 460 are typically annual so cannot be used to measure phenology, but they do often cover large 461 geographic areas, so are very suitable for distribution studies.

Few observations of marine phenology were available from autumn or winter, a potential temporal bias that also occurs in terrestrial studies of phenology (Gallinat *et al.*, 2015). The lack of data on phenology from autumn and winter could partially reflect the fact that many species perform many of their most important processes (e.g. feeding and reproduction) predominantly in spring and summer. There are also many more spring than autumn

467 observations for terrestrial ecosystems (Parmesan & Yohe, 2003). But unlike marine systems,

- terrestrial systems do have a few very long-term (e.g. grape harvests over 800 year, (Menzel,
- 469 2005)), and iconic (e.g. fall color indexes in New England, USA (Gallinat *et al.*, 2015))
- 470 autumn measurements. We found evidence that autumnal events were shifting later, rather
- than earlier, which is consistent with lengthening seasons. Measuring autumn phenology in
- 472 higher latitudes is therefore an important knowledge gap in both marine and terrestrial
- 473 systems. Because of this gap, we have little information on how growing seasons may be
- 474 extended by warming (for an example see Moore *et al.*, 2011).
- 475 *Recommendations for measuring change*
- 476 Based on the findings of our meta-analysis, we make several recommendations for measuring
- responses of organisms to climate change when analysing past studies of climate change
- 478 impacts or designing new studies.
- 479 (1) Re-analysis of existing time-series
- 480 A critical question is whether the time-series can be used to address the study's aim. For
- 481 instance, a database of species occurrences across space and time can be used to examine
- 482 colonisation of new areas, but is more limited in supporting inferences about the
- 483 establishment and persistence of new populations. Similarly for phenology, a time-series of
- the most extreme individuals breeding timing does not necessarily reflect significant change
- in a whole population, although changes in a few individuals may be an early warning for
- 486 population level change. Therefore, researchers should be careful to consider the potential
- 487 limitations and biases in data when conducting re-analyses
- 488 Covariates, particularly those not related to climate, are key to consider when analysing time-
- 489 series. A typical test is to ask whether warming is driving an observed change, with the null
- 490 hypothesis being warming is not a factor. Greater consideration of other alternatives is

491	important (Brown et al., 2011), including non-climate drivers of distribution and phenology.
492	For instance, changes in depth range of an organism could be driven by warming, but the
493	potentially confounding effect of fishing pressure changing with depth should also be
494	considered in such an analysis (Dulvy et al., 2008, Engelhard et al., 2014).
495	Broadening the scope of climate change studies to include other drivers will require greater
496	accessibility of data on human impacts – such as fishing and eutrophication at appropriate
497	time and space scales. Climate time-series data are widely available as free downloads,
498	whereas, data on other drivers are often less-easily available or do not exist at all. Efforts to
499	share currently closed databases, such as those on fishing and efforts to collect more data, for
500	instance using satellite images to map eutrophication, will enable better discrimination of
501	climate from other signals and thus more robust climate attribution.
502	(2) Qualitative comparisons with other studies
503	Qualitative comparisons among rates of change are common in regional or taxon-specific
503 504	Qualitative comparisons among rates of change are common in regional or taxon-specific studies of climate change (e.g. Richardson, 2008). The usual aim of qualitative comparisons is
504	studies of climate change (e.g. Richardson, 2008). The usual aim of qualitative comparisons is
504 505	studies of climate change (e.g. Richardson, 2008). The usual aim of qualitative comparisons is to give context for an observed rate of change being faster or slower than typical, and to
504 505 506	studies of climate change (e.g. Richardson, 2008). The usual aim of qualitative comparisons is to give context for an observed rate of change being faster or slower than typical, and to speculate about the ecological or environmental drivers of a difference. However, differences
504 505 506 507	studies of climate change (e.g. Richardson, 2008). The usual aim of qualitative comparisons is to give context for an observed rate of change being faster or slower than typical, and to speculate about the ecological or environmental drivers of a difference. However, differences will also be strongly influenced by measurement biases. Where possible, qualitative
504 505 506 507 508	studies of climate change (e.g. Richardson, 2008). The usual aim of qualitative comparisons is to give context for an observed rate of change being faster or slower than typical, and to speculate about the ecological or environmental drivers of a difference. However, differences will also be strongly influenced by measurement biases. Where possible, qualitative comparisons should be made with similar metrics used to measure observed change. For
504 505 506 507 508 509	studies of climate change (e.g. Richardson, 2008). The usual aim of qualitative comparisons is to give context for an observed rate of change being faster or slower than typical, and to speculate about the ecological or environmental drivers of a difference. However, differences will also be strongly influenced by measurement biases. Where possible, qualitative comparisons should be made with similar metrics used to measure observed change. For instance measurements of distribution based on occurrence at sites should not be compared
504 505 506 507 508 509 510	studies of climate change (e.g. Richardson, 2008). The usual aim of qualitative comparisons is to give context for an observed rate of change being faster or slower than typical, and to speculate about the ecological or environmental drivers of a difference. However, differences will also be strongly influenced by measurement biases. Where possible, qualitative comparisons should be made with similar metrics used to measure observed change. For instance measurements of distribution based on occurrence at sites should not be compared with measurements based on abundance averages, which are typically slower. As the number
504 505 506 507 508 509 510 511	studies of climate change (e.g. Richardson, 2008). The usual aim of qualitative comparisons is to give context for an observed rate of change being faster or slower than typical, and to speculate about the ecological or environmental drivers of a difference. However, differences will also be strongly influenced by measurement biases. Where possible, qualitative comparisons should be made with similar metrics used to measure observed change. For instance measurements of distribution based on occurrence at sites should not be compared with measurements based on abundance averages, which are typically slower. As the number of climate studies grows, it will become easier to compare studies that use similar methods.

515	covariates include measurement and ecological variables. Past studies have either ignored
516	these differences (Poloczanska et al., 2013) or tried to removed non-similar studies (Parmesan
517	& Yohe, 2003, Parmesan, 2007, Przesławski et al., 2012). While including additional
518	measurement variables in analysis did not significantly change our results when compared to
519	previous analyses (Poloczanska et al., 2013), it did shed important light on factors affecting
520	distribution and phenology change. Removing studies from analysis focuses on a subset of
521	potential biases (e.g. only including studies on multiple species or time series greater than a
522	certain length) is a blunt approach that does not consider multiple other potential sources of
523	bias (e.g. measurement type, latitude, non-climate factors) that are needed to provide more
524	robust estimates of climate change on species. Removing studies from analysis reduces the
525	power to detect real biological effects, and therefore should be avoided where possible.
526	(4) Design of new studies
526 527	(4) Design of new studies Numerous time-series are currently being started, with the aim of monitoring effects of
527	Numerous time-series are currently being started, with the aim of monitoring effects of
527 528	Numerous time-series are currently being started, with the aim of monitoring effects of climate change (e.g. Robinson <i>et al.</i> , 2015). Our findings provide some advice on designing
527 528 529	Numerous time-series are currently being started, with the aim of monitoring effects of climate change (e.g. Robinson <i>et al.</i> , 2015). Our findings provide some advice on designing sampling protocols. The aims of measuring the time-series should be explicitly defined and a
527 528 529 530	Numerous time-series are currently being started, with the aim of monitoring effects of climate change (e.g. Robinson <i>et al.</i> , 2015). Our findings provide some advice on designing sampling protocols. The aims of measuring the time-series should be explicitly defined and a protocol designed to address them. For example, a study that seeks to monitor invasion of pest
527 528 529 530 531	Numerous time-series are currently being started, with the aim of monitoring effects of climate change (e.g. Robinson <i>et al.</i> , 2015). Our findings provide some advice on designing sampling protocols. The aims of measuring the time-series should be explicitly defined and a protocol designed to address them. For example, a study that seeks to monitor invasion of pest species may focus on monitoring for occurrences, to obtain early warnings of ranges shifts. In
527 528 529 530 531 532	Numerous time-series are currently being started, with the aim of monitoring effects of climate change (e.g. Robinson <i>et al.</i> , 2015). Our findings provide some advice on designing sampling protocols. The aims of measuring the time-series should be explicitly defined and a protocol designed to address them. For example, a study that seeks to monitor invasion of pest species may focus on monitoring for occurrences, to obtain early warnings of ranges shifts. In contrast, a study that aims to detect ecologically significant might focus on monitoring
527 528 529 530 531 532 533	Numerous time-series are currently being started, with the aim of monitoring effects of climate change (e.g. Robinson <i>et al.</i> , 2015). Our findings provide some advice on designing sampling protocols. The aims of measuring the time-series should be explicitly defined and a protocol designed to address them. For example, a study that seeks to monitor invasion of pest species may focus on monitoring for occurrences, to obtain early warnings of ranges shifts. In contrast, a study that aims to detect ecologically significant might focus on monitoring abundance.

537 may be a more robust measure of change (Brown *et al.*, 2011). However, there may be trade-

- 538 offs in sampling effort that warrant consideration. Occurrences are cheaper to measure than
- abundances, particularly for rare species, so occurrence surveys may cover larger areas and a

540 greater range of species than abundance-based surveys could for the same cost. Further, 541 occurrence-based measures are easier to obtain from non-experts, such as through citizen 542 science programs or from observations from fishers (Robinson et al., 2015). Occurrence-543 based measures could therefore provide a more useful early warning of invasion of new 544 species, but do not necessarily indicate establishment of a new population. 545 A common approach to detecting climate change impacts is to resurvey sites that had 546 historical measurements of climate change. Such resurveys are important to fill data gaps, yet 547 our results suggest some selection bias for sites with greater change, at least for distribution 548 studies. It is important that resurvey sights that are selected randomly (or comprehensively) to 549 provide a less biased global view of climate change impacts, for instance by systematically 550 sampling across a species' entire range. 551 Large gaps remain in our knowledge of climate change responses in both terrestrial and 552 marine systems. Key amongst these is that there is three times as much information on 553 changes in distribution than phenology in the ocean, whereas on land there is 100 times more 554 information on phenology than on distribution change (comparing Poloczanska et al. 2013

with Rosenzweig *et al.*, 2008). Expanding terrestrial studies of species' distribution change

and marine studies of phenology change, particularly in autumn and winter, is important to

557 give a comprehensive view of life's responses to climate change.

558 Conclusion

- 559 We found that measurement biases can have a substantial effect on inferences about the
- 560 impacts of climate change on distribution and phenology. Greater consideration of
- 561 measurement bias in climate impacts studies will improve our understanding for how
- 562 measurement methods affect observations and ultimately contribute to a more representative
- view of the impacts of climate change on organisms.

564

Acknowledgements

565	This work was conducted as part of the Understanding Marine Biological Impacts of Climate
566	Change Working Group supported by the National Center for Ecological Analysis and
567	Synthesis, a centre funded by NSF (Grant #EF-0553768), the University of California, Santa
568	Barbara and the State of California. CJB acknowledges a University of Queensland
569	postdoctoral fellowship. JMP acknowledges support from the Australian Research Council
570	(CE0561435, DP130100250). We are grateful to all the workers on climate change impacts
571	which we have reviewed.
572	
573	References
574	Barbraud C, Weimerskirch H (2006) Antarctic birds breed later in response to climate change.
575	Proceedings of the National Academy of Sciences of the United States of America,
576	103 , 6248-6251.
577	Bates AE, Bird TJ, Stuart-Smith RD et al. (2015) Distinguishing geographical range shifts
578	from artefacts of detectability and sampling effort. Diversity and Distributions, 21, 13-
579	22.
580	Bates AE, Pecl GT, Frusher S et al. (2014a) Defining and observing stages of climate-
581	mediated range shifts in marine systems. Global Environmental Change, 26, 27-38.
582	Bates D, Maechler M, Bolker BM, Walker S (2014b) lme4: Linear mixed-effects models
583	using Eigen and S4. ArXiv e-print; submitted to Journal of Statistical Software,
584	http://arxiv.org/abs/1406.5823.
585	Beaugrand G, Kirby RR (2010) Climate, plankton and cod. Global Change Biology, 16,
586	1268-1280.

- 587 Broitman B, Blanchette C, Menge B et al. (2008) Spatial and temporal patterns of
- invertebrate recruitment along the west coast of the United States. EcologicalMonographs, 78, 403-421.
- Brown CJ, Schoeman DS, Sydeman WJ *et al.* (2011) Quantitative approaches in climate
 change ecology. Global Change Biology, **17**, 3697-3713.
- 592 Burnham KP, Anderson DR (2002) *Model selection and multimodel inference: a practical*
- 593 *information-theoretic approach*, New York, Springer.
- Burrows MT, Schoeman DS, Buckley LB *et al.* (2011) The pace of shifting climate in marine
 and terrestrial ecosystems. Science, 334, 652-655.
- 596 Burrows MT, Schoeman DS, Richardson AJ et al. (2014) Geographical limits to species-

range shifts are suggested by climate velocity. Nature, **507**, 492-495.

- 598 Canty A, Ripley B (2014) boot: Bootstrap R (S-Plus) Functions. (ed 1.3-13 RPV) pp Page.
- Chambers LE, Altwegg R, Barbraud C *et al.* (2013) Phenological changes in the Southern
 Hemisphere. Plos One, 8, e75514.
- 601 Cook BI, Wolkovich EM, Parmesan C (2012) Divergent responses to spring and winter
- warming drive community level flowering trends. Proceedings of the National
 Academy of Sciences, 109, 9000-9005.
- Davis CC, Willis CG, Primack RB, Miller-Rushing AJ (2010) The importance of phylogeny

to the study of phenological response to global climate change. Philosophical
Transactions of the Royal Society B-Biological Sciences, 365, 3201-3213.

- 607 Dulvy NK, Rogers SI, Jennings S, Stelzenmuller V, Dye SR, Skjoldal HR (2008) Climate
- change and deepening of the North Sea fish assemblage: a biotic indicator of warming
 seas. Journal of Applied Ecology, 45, 1029-1039.
- 610 Edwards M, Richardson AJ (2004) Impact of climate change on marine pelagic phenology
- and trophic mismatch. Nature, **430**, 881-884.

612	Engelhard GH, Righton DA, Pinnegar JK (2014) Climate change and fishing: a century of
613	shifting distribution in North Sea cod. Global Change Biology, 20, 2473-2483.
614	Fitter A, Fitter R (2002) Rapid changes in flowering time in British plants. Science, 296,
615	1689-1691.
616	Gallinat AS, Primack RB, Wagner DL (2015) Autumn, the neglected season in climate
617	change research. Trends in Ecology & Evolution, 30 , 169-176.
618	Graham NA, Cinner JE, Norström AV, Nyström M (2014) Coral reefs as novel ecosystems:
619	embracing new futures. Current Opinion in Environmental Sustainability, 7, 9-14.
620	Jacups SP (2010) Warmer Waters in the Northern Territory-Herald an Earlier Onset to the
621	Annual Chironex fleckeri Stinger Season. EcoHealth, 7, 14-17.
622	Lenoir J, Svenning JC (2015) Climate-related range shifts-a global multidimensional
623	synthesis and new research directions. Ecography, 38, 15-28.
624	Ling S, Johnson C, Ridgway K, Hobday A, Haddon M (2008) Climate driven range
625	extension of a sea urchin: inferring future trends by analysis of recent population
626	dynamics. Global Change Biology, 15, 719-731.
627	Loarie SR, Duffy PB, Hamilton H, Asner GP, Field CB, Ackerly DD (2009) The velocity of
628	climate change. Nature, 462, 1052-1055.
629	Menzel A (2005) A 500 year pheno-climatological view on the 2003 heatwave in Europe
630	assessed by grape harvest dates. Meteorologische Zeitschrift, 14, 75-77.
631	Molinos JG, Halpern BS, Schoeman DS et al. (2015) Climate velocity and the future global
632	redistribution of marine biodiversity. Nature Climate Change, in press.
633	Moore PJ, Thompson RC, Hawkins SJ (2011) Phenological changes in intertidal con-specific
634	gastropods in response to climate warming. Global Change Biology, 17, 709-719.
635	Moussus JP, Julliard R, Jiguet F (2010) Featuring 10 phenological estimators using simulated
636	data. Methods in Ecology and Evolution, 1, 140-150.

637	Nakagawa S, Schielzeth H (2013) A general and simple method for obtaining R2 from
638	generalized linear mixed effects models. Methods in Ecology and Evolution, 4, 133-
639	142.
640	Parmesan C (2007) Influences of species, latitudes and methodologies on estimates of
641	phenological response to global warming. Global Change Biology, 13, 1860-1872.
642	Parmesan C, Burrows MT, Duarte CM, Poloczanska ES, Richardson AJ, Schoeman DS,
643	Singer MC (2013) Beyond climate change attribution in conservation and ecological
644	research. Ecology Letters, 16, 58-71.
645	Parmesan C, Ryrholm N, Stefanescu C et al. (1999) Poleward shifts in geographical ranges of
646	butterfly species associated with regional warming. Nature, 399 , 579-583.
647	Parmesan C, Yohe G (2003) A globally coherent fingerprint of climate change impacts across
648	natural systems. Nature, 421, 37-42.
649	Perry AL, Low PJ, Ellis JR, Reynolds JD (2005) Climate change and distribution shifts in
650	marine fishes. Science, 308, 1912-1915.
651	Pinsky ML, Worm B, Fogarty MJ, Sarmiento JL, Levin SA (2013) Marine taxa track local
652	climate velocities. Science, 341, 1239-1242.
653	Poloczanska ES, Brown CJ, Sydeman WJ et al. (2013) Global imprint of climate change on
654	marine life. Nature Climate Change, 3 , 919-925.
655	Poloczanska ES, Smith S, Fauconnet L, Healy J, Tibbetts IR, Burrows MT, Richardson AJ
656	(2011) Little change in the distribution of rocky shore faunal communities on the
657	Australian east coast after 50 years of rapid warming. Journal of Experimental Marine
658	Biology and Ecology, 400, 145-154.
659	Przesławski R, Falkner I, Ashcroft MB, Hutchings P (2012) Using rigorous selection criteria
660	to investigate marine range shifts. Estuarine, Coastal and Shelf Science.

- 661 Rayner N, Parker DE, Horton E et al. (2003) Global analyses of sea surface temperature, sea
- ice, and night marine air temperature since the late nineteenth century. Journal of
- 663 Geophysical Research: Atmospheres (1984–2012), **108**.
- Richardson AJ (2008) In hot water: zooplankton and climate change. ICES Journal of Marine
 Science: Journal du Conseil, 65, 279-295.
- Richardson AJ, Brown CJ, Brander K *et al.* (2012) Climate change and marine life. Biology
 Letters, rsbl20120530.
- Robinson L, Gledhill D, Moltschaniwskyj N et al. (2015) Rapid assessment of an ocean
- warming hotspot reveals "high" confidence in potential species' range extensions.
 Global Environmental Change, **31**, 28-37.
- 671 Rosenzweig C, Karoly D, Vicarelli M *et al.* (2008) Attributing physical and biological

672 impacts to anthropogenic climate change. Nature, **453**, 353-U320.

- 673 Sagarin RD, Gaines SD, Gaylord B (2006) Moving beyond assumptions to understand
- abundance distributions across the ranges of species. Trends in Ecology & Evolution,
 21, 524-530.
- 676 Shoo LP, Williams SE, Hero J-M (2006) Detecting climate change induced range shifts:

677 Where and how should we be looking? Austral Ecology, **31**, 22-29.

678 Southward AJ, Langmead O, Hardman-Mountford NJ *et al.* (2005) Long-term oceanographic

and ecological research in the western English Channel. In: *Advances in Marine*

680 *Biology, Vol 47.* (eds Southward AJ, Tyler PA, Young CM, Fuiman LA) pp Page.

- 681 Staples D, Vance D (1986) Emigration of juvenile banana prawns Penaeus merguiensis from
- a mangrove estuary and recruitment to offshore areas in the wet-dry tropics of the Gulf
 of Carpentaria, Australia. Marine Ecology Progress Series, 27, 52.
- Sunday JM, Bates AE, Dulvy NK (2012) Thermal tolerance and the global redistribution of
 animals. Nature Climate Change, 2, 686-690.

686	Sunday JM, Pecl GT, Frusher S et al. (2015) Species traits and climate velocity explain
687	geographic range shifts in an ocean warming hotspot. Ecology Letters.
688	Van Schaik CP, Terborgh JW, Wright SJ (1993) The phenology of tropical forests: adaptive
689	significance and consequences for primary consumers. Annual Review of Ecology and
690	Systematics, 353-377.
691	Wolkovich EM, Cook B, Allen J et al. (2012) Warming experiments underpredict plant
692	phenological responses to climate change. Nature, 485, 494-497.
693	
694	

696 Tables

- 697 **Table 1** Analysis of rates of distribution and phenology change, with the Δ BIC calculated by
- adding (for non-significant variables) or removing (for significant variables) each variable
- from the reduced model. Variables with negative Δ BIC were not included in the reduced
- 700 models. N.A. Not applicable

Factor	Variable class	Distribution	Phenology data-
		data-set	set
		ΔΒΙC	ΔΒΙC
Abundance/occurrence	Methodological	23	-4.5
based measure	approach		
Data sampling	Methodological	64	-9.7
frequency	approach		
Non-climatic factors	Methodological	-6.1	2.8
considered	approach		
No. spp. in study	Methodological	-5.4	-4.0
	approach		
Timespan of study	Methodological	-5.4	0.2
	approach		
Depth category	Ecological trait	47	-9.5
Exploitation status	Ecological trait	-4.0	-4.9
Latitude	Ecological trait	-1.2	2.8
Planktonic larval	Ecological trait	-2.5	-5.6

dispersal type			
Range edge/centre	Ecological trait	-6.9	N.A.
Season	Ecological trait	N.A.	10.4
Velocity of climate	Ecological trait	-2.2	-4.6
change			

701

702 Figure legends

Fig 1 Frequency of different factors in studies of distribution and phenology: (a) number of

species in a study; (b) occurrence-based or abundance-based measures of distribution and

phenology; (c) sampling frequency; (d) latitude; (e) broad taxonomic groupings; and (f)

706 season, for

707 Fig 2 Term plot for analysis of rates of change in distribution using the final mixed effects 708 model, showing the final model (selected using BIC, Full model BIC = 5713, reduced model 709 BIC = 5686). For the fixed effects, points indicate mean predicted effects and bars are 95%710 confidence intervals. Taxa were treated as a random effect. Effects are standardised to the 711 overall mean, so positive effects indicate a tendency toward distribution change that is greater 712 and consistent with climate change, whereas negative effects indicate a tendency toward 713 smaller changes or changes that are not consistent with warming (though those two cannot be 714 distinguished in this figure). Note the varying scales for the y-axes; larger ranges indicate 715 larger effect sizes. 716 Fig 3 Term plot for analysis of rates of change in phenology using the final mixed effects

model, showing the final model (selected using BIC, Full model BIC = 1153, reduced model

- BIC = 1112). Points indicate mean predicted effects and bars the 95% confidence intervals.
- Taxa were treated as a random effect. Effects are standardised to the overall mean, so

720	negative effects (upwards on the y-axis) indicate a tendency toward phenology change that is
721	earlier and consistent with climate change, whereas positive effects indicate a tendency
722	toward smaller date changes or changes that are not consistent with warming. Note the
723	varying scales for the y-axes.
724	Fig 4 Proportion of distribution observations by taxa and each covariate used in the final
725	model for distribution rate of change. The maximum proportion of observations in any
726	category was 0.4.
727	Fig 5 Proportion of phenology observations by taxa and each covariate used in the final
728	model for distribution rate of change. The maximum proportion of observations in any
729	category was 0.4.

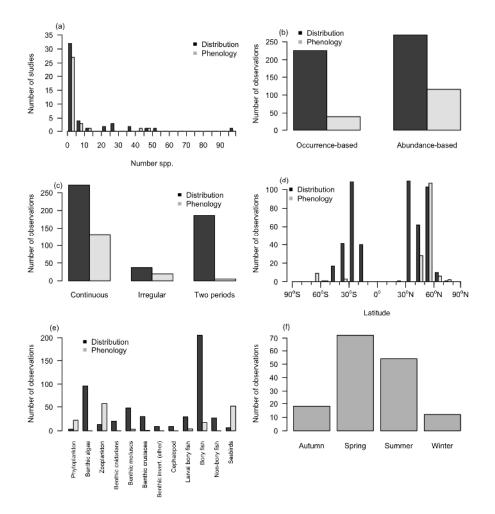


Fig 1 Frequency of different factors in studies of distribution and phenology: (a) number of species in a study; (b) occurrence-based or abundance-based measures of distribution and phenology; (c) sampling frequency; (d) latitude; (e) broad taxonomic groupings; and (f) season, for phenology. 704x939mm (72 x 72 DPI)

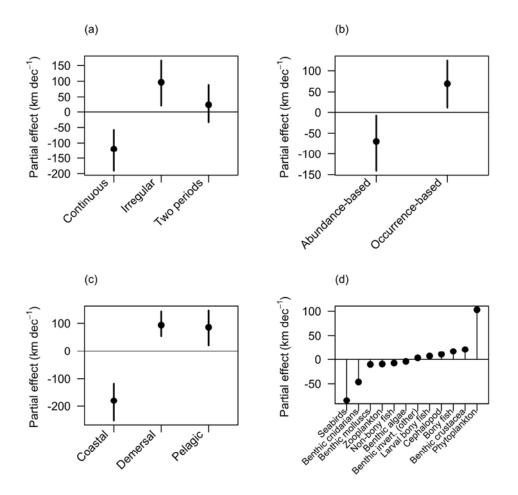


Fig 2 Term plot for analysis of rates of change in distribution using the final mixed effects model, showing the final model (selected using BIC, Full model BIC = 5713, reduced model BIC = 5686). For the fixed effects, points indicate mean predicted effects and bars are 95% confidence intervals. Taxa were treated as a random effect. Effects are standardised to the overall mean, so positive effects indicate a tendency toward distribution change that is greater and consistent with climate change, whereas negative effects indicate a tendency toward smaller changes or changes that are not consistent with warming (though those two cannot be distinguished in this figure). Note the varying scales for the y-axes; larger ranges indicate larger effect sizes.

80x80mm (300 x 300 DPI)

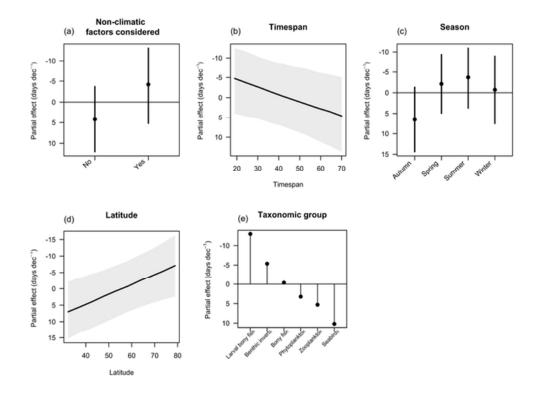


Fig 3 Term plot for analysis of rates of change in phenology using the final mixed effects model , showing the final model (selected using BIC, Full model BIC = 1153, reduced model BIC = 1112). Points indicate mean predicted effects and bars the 95% confidence intervals. Taxa were treated as a random effect. Effects are standardised to the overall mean, so negative effects (upwards on the y-axis) indicate a tendency toward phenology change that is earlier, whereas positive effects indicate a tendency toward smaller date changes or to a later date. Note the varying scales for the y-axes. 59x44mm (300 x 300 DPI)

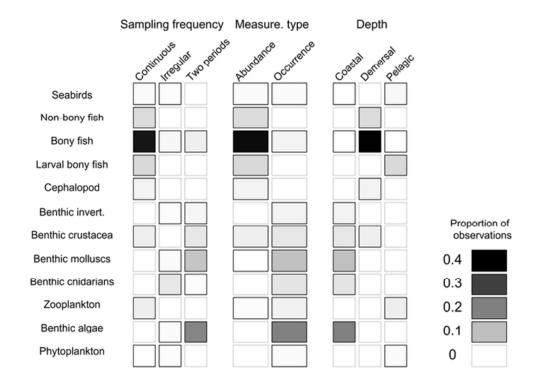


Fig 4 Proportion of distribution observations by taxa and each covariate used in the final model for distribution rate of change. The maximum proportion of observations in any category was 0.4. 59x44mm (300 x 300 DPI)

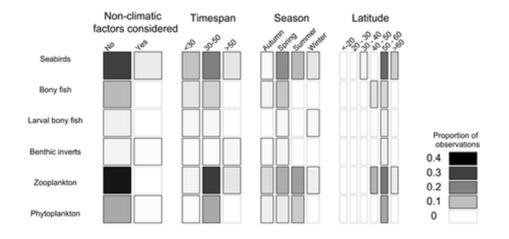


Fig 5 Proportion of phenology observations by taxa and each covariate used in the final model for distribution rate of change. The maximum proportion of observations in any category was 0.4. 39x19mm (300 x 300 DPI)