



## Effect of Ginger Nanoparticles on Hepato-renal Toxicity Induced by Carbon Tetrachloride in Rats

Sanaa Yasser Abd-Elrhman<sup>1</sup>, Hanaa Mostafa Abd El- Fattah<sup>1</sup>, Gehan M. Morsy<sup>1</sup>  
and Shima Elmasry<sup>1\*</sup>

<sup>1</sup>Department of Biochemistry and Nutrition, Faculty of Women for Arts, Science and Education, Ain Shams University, Cairo, Egypt.

### Authors' contributions

This work was carried out in collaboration among all authors. Author HMAEF designed the study and wrote the protocol and wrote the first draft of the manuscript. Author SYAE managed the analyses of the study, performed the statistical analysis and managed the literature searches. Authors GMM and SE wrote the protocol, wrote the first draft of the manuscript and performed the statistical analysis. All authors read and approved the final manuscript.

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### ABSTRACT

**Aims:** The current study was developed to investigate the influence of ginger (G), ginger nanoparticles (GNPs) and ginger nano-base (GNB) on hepato-renal toxicity induced by carbon tetrachloride (CCl<sub>4</sub>) in rats in comparison with silymarin (SM).

**Place and Duration of Study:** Department of Biochemistry and Nutrition, Faculty of Women for Arts, Science and Education, Ain Shams University.

**Methodology:** Fifty-four adult male Sprague-Dawley rats were divided into 6 groups. Group (1): Rats received distilled water orally and injected intraperitoneally (i.p.) with single dose of corn oil (1 ml/kg b.wt). Group (2): Rats were injected with single dose of CCl<sub>4</sub> diluted with corn oil (1:1) (1 ml/kg b.wt. i.p.) at the 4<sup>th</sup> week of experiment. Groups (3), (4) and (5): Rats were orally received 50 mg /kg b.wt./day of G, GNPs and GNB, respectively for 8 weeks and injected with CCl<sub>4</sub> as group 2. Group (6): Rats were orally received 100 mg /kg b.wt /day of SM for 8 weeks and injected with CCl<sub>4</sub> as group 2.

\*Corresponding author: E-mail: shima\_elmasry33@yahoo.com, Shima.elmasry@women.asu.edu.eg;

**Results:** Our results documented that injection with  $\text{CCl}_4$  caused significant increase ( $p < 0.05$ ) in liver function tests [serum alanine aminotransferase (ALT), aspartate aminotransferase (AST) and alkaline phosphatase (ALP) activities], kidney function tests [serum creatinine, urea, uric acid and cystatin C] and serum levels of malondialdehyde (MDA), Nitric oxide (NO), tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ) and interleukin 1 beta (IL-1 $\beta$ ). On the other hand, there was a significant decrease ( $p < 0.05$ ) in the serum total antioxidant capacity (TAC), Hepatic catalase (CAT) and superoxide dismutase (SOD) enzymes activity, with histopathological changes in liver and kidneys tissues. Oral administration of G, GNPs, GNB and SM caused an enhancement of liver and kidney function, decreasing serum oxidants and inflammatory markers levels while increasing the activities of antioxidant enzymes, also an improvement of organs histopathological changes was observed.

**Conclusion:** Our data proved that using ginger in the form of GNPs and GNB are more efficient in ameliorating hepato-renal toxicity induced by  $\text{CCl}_4$  than using native ginger as evidenced by biochemical analysis and histological examination of liver and kidneys tissues.

**Keywords:** Carbon tetrachloride; hepatotoxicity; nephrotoxicity; ginger nanoparticles; ginger nanobase; *Zingiber officinale*; silymarin.

## 1. INTRODUCTION

Liver is the organ engaged in digestion, secretion, biosynthesis, storage of vital nutrients and detoxification of toxicants. Hepatotoxicity is an adverse effects or injuries that occur to the liver due to exposure to hepatotoxins as carbon tetrachloride ( $\text{CCl}_4$ ), paracetamol, ethanol, galactosamine and anti-tuberculosis drugs [1]. Extravagant intake of these chemicals led to enormous production of free radicals, induce lipid peroxidation and resulting in structural and functional destroys to the membrane and finally causing dangerous toxicity to hepatic cells [2].

Kidneys are the main organs for the secretion of endogenous and exogenous substances. All toxic chemicals and metabolites that are removed by the urine pass through the kidney. Chemicals which induced renal toxicity may occur in the kidney either directly, via biotransformation process in the proximal tubular cells, or indirectly, through the formation of toxic metabolites in the liver which have nephrotoxic effects which leading to acute renal failure [3].

Carbon tetrachloride ( $\text{CCl}_4$ ) is largely used as an industrial chemical that evaporates simply in the encompassing environment. The liver has a wonderful tendency for  $\text{CCl}_4$  as it comprises cytochrome P450 enzyme system which converts  $\text{CCl}_4$  to trichloromethyl radical ( $\text{CCl}_3\cdot$ ) and peroxy trichloromethyl ( $\text{CCl}_3\text{O}_2\cdot$ ) radicals. The mass production of these toxic metabolites may covalently bind to the proteins and lipids of plasma membrane or interact with oxygen which invades unsaturated fatty acids initiating lipid peroxidation. Additionally, overproduction of free radicals may lead to attrition or deactivation

of antioxidant enzymes. These effects cause damage of unsaturated fatty acids and resulting in diverse pathological problems. Pharmacokinetically, metabolic activation of  $\text{CCl}_4$  happens not only in the liver but also in other organs like lung, heart, testis, brain and kidney [4,5].

Medicinal plants possess the potential of preventing acute and chronic diseases. The protective effect of plants is often maintained through an increased expression of antioxidants and scavenging of free radicals. Antioxidants present in medicinal plants protect against hepatic and renal damage by increasing superoxide dismutase (SOD) and catalase (CAT) activities and lowering lipid peroxidation [6].

Ginger (*Zingiber officinale* Rosco) a member of the family Zingiberaceae, is traditionally used as a food additive (spice) worldwide to enhance the flavor of food and a common medicinal herb, which contains various phytochemical compounds, as polyphenols including flavonoids and tannins with anti-inflammatory, antioxidant, anti-diabetic, hypo-lipidemic and anti-carcinogenic activities. Ginger has antioxidant activity owing to the presence of gingerols, shogaols, zingerone, gingerdiol and zingiberene [7]. Many studies had shown that ginger has hepatoprotective [7,8] and nephroprotective effects [9,10] by improving the liver and kidney biomarkers levels, decreasing inflammation, enhancing the antioxidant activities and prevent lipid peroxidation induced by toxicants.

There are challenges with the application of nanoscale compared with microscale materials. These include higher exposure per unit mass,

small size: Large surface area ratio, different ways of exposure due to smaller size (i.e. dermal penetration), different distribution to tissues depending on their different size or surface coating, chemistry or particle charge and the properties of a nanoscale material that may alter absorption, digestion, metabolism or excretion in the body. Nanotechnology holds excitement and promise in all areas of science [11].

Plants contain natural nanoparticles which obtained by simple preparation when compared with synthetic nanoparticles which involved financial and potential preparation difficulties [12]. Edible plants nanoparticles can spread from intestine to the liver, so they may have an influential function in treatment pathway and drug delivery to the target cells and enhancement of therapeutic outcomes. Furthermore, nanoparticles present in natural compounds supply a secure carrier for ameliorating drug bioavailability in the cells [11]. Milling herbs to nanoscale increase their exposed surface area, hence, enhancing their active compounds solubility and bioavailability [13].

Many studies demonstrated that ginger nanoparticles (GNPs) improve the antioxidant activity by increasing level of antioxidant enzymes and decrease oxidative stress [14]. Also, it has hepatoprotective effect against liver damage [12] and provide protection against hepatotoxicity and nephrotoxicity induced by acetaminophen in comparison with ginger extract [15], additionally, it prevents nephrotoxicity induced by emamectin benzoate [13].

Synthetic analogs and formulations of ginger have been developed, including combination with polymeric micelles [16] or nanoparticle-based encapsulation by poly-lactic-co-glycolic acids (PLGA) [14] or isolated ginger derived nanoparticles from ginger juice by using high power centrifugation [12], that exhibit greater chemical stability, systemic bioavailability and antioxidant, anti-inflammatory and antitumor activities than naturally occurring ginger. Therefore, we try to develop ginger nano-base particles by milling ginger with sodium bicarbonate [17], to increase ginger solubility and bioavailability.

*Silybum marianum* is a member of Asteraceae family, it is used to treat liver disease as a hepatoprotective herb. [18]. The hepatoprotective effect of silymarin is attributed to its dose-dependent antioxidant effect. This impact is

intermediated by catching the free radicals, reducing production of ROS and attenuating the lipid peroxidation. Other mechanisms include anti-apoptotic and anti-inflammatory actions via intervention with nuclear factor kappa-B (NF- $\kappa$ B), alternation of inducible nitric oxide (iNOS) and decreases in cyclooxygenase-2 (COX-2) expression [19]. Silymarin also has renoprotective mediator against nephrotoxic agents because of its antioxidant, anti-inflammatory and anti-apoptotic properties [20,21].

The present study was conducted to assess the influence of ginger (G), ginger nanoparticles (GNPs) and ginger nano-base (GNB) on liver and kidney toxicity induced by carbon tetrachloride (CCl<sub>4</sub>) in rats.

## 2. MATERIALS AND METHODS

### 2.1 Chemicals

Carbon tetrachloride (99%) was purchased from El-Gomhoria Company for Chemicals and Drugs; Cairo, Egypt. Silymarin (*Silybum marianum*) was purchased from Chemical Industries Development Company; Cairo, Egypt.

### 2.2 Plant Material

Fresh ginger rhizome roots (*Zingiber officinale Roscoe*) was purchased from Ministry of Agriculture; Giza, Egypt.

### 2.3 Experimental Animals

Adult male albino rats (Sprague -Dawley strain) weighing 150-180 g were obtained from animal Breeding House of National Research Center (NRC); Giza, Egypt. Animals were handled in this study in accordance with the Principles of laboratory animal care" (NIH publication No. 85-23, revised 1985). The experimental animals were housed individually in metallic cages with good ventilation and under conventional condition (22  $\pm$  3°C and natural light/dark cycle). All rats were received the standard commercial diet (obtained from National Research Center; Giza, Egypt) and water *ad libitum* for 7 days (acclimatization period) before starting the experiment.

### 2.4 Preparation of Ginger (G), Ginger Nanoparticles (GNPs) and Ginger Nano-Base (GNB)

Ginger rhizome roots were washed several times with water, cut into small pieces, then dried by

oven at 50°C for 2 hours, the dried ginger was ground to get fine ginger powder utilizing mixer and stored in sterilized glass container at room temperature for future use [22,14].

To prepare ginger nanoparticles (GNPs) about 50 mg of ginger powder was ground by mechanical ball milling [23] utilizing a planetary ball mill (Retsch PM 400, Germany) at (350 round/Sec) for 16 hours. The main episode in mechanical milling is the ball–powder–ball crashing, where the dehydrated powder was restricted between the crashed balls throughout milling with high speed forming fine powder in nano scales [22].

*Zingiber officinale* and turmeric (*curcuma longa*) are in the same botanical family (Zingiberaceae) and these plants have similar molecules. Also [6]-shogaol and [6]-gingerol from ginger have structural similarity with curcumin. So, they might have similar antioxidant and anti-inflammatory properties [24]. So that we applied the same method of preparation of curcumin nano-base in ginger to produce ginger nano-base, in an attempt for increasing the solubility and bioavailability of ginger as occur with curcumin nano-base.

For preparation of basic ginger nanoparticles (GNB), 50 mg fine ginger powder with low solubility in water was mixed with 50 mg sodium bicarbonate buffer, then ground by mechanical ball mill using a planetary ball mill (Retsch PM 400, Germany) at (350 round/Sec) for 15 hrs. The color of ginger changed from yellow to yellowish white as a result of the ginger sodium salt formation [17].

## 2.5 Evaluation of Ginger Nanoparticles (GNPs) Size by Using DLS

Dynamic light scattering (DLS) is an efficient method used to determine the nanoparticle size, size distribution and shape of particles in suspension through the brownian motion and doppler shift induced by a laser beam. The ginger nanoparticles or ginger nano-base was diluted with filtered de-ionized water to produce suspended solution and then placed in Elmasonic S 15 H apparatus at 20 - 25°C for 3 min to vibrate and distribute the nanoparticles in solution, then the solution was left for 10 min to allow stability of nanoparticles, after that the solution transferred to Malvern Zetasizer Nano ZS Apparatus to measure the nanoparticles size.

## 2.6 Determination of Total Phenols and Total flavonoids Content in Ginger

Total phenols were determined in ginger ethanolic extract by using the Folin-Ciocalteu method, while total flavonoids were determined by aluminum chloride colorimetric method according to Singleton et al. and Arvouet-Grand et al., respectively [25,26].

## 2.7 Determination of Main phenolic Compounds in Ginger

The concentration of [6]-gingerol, [8]-gingerol, [6]-shogol, [10]-gingerol, [8]-shogol and [10]-shogol in ginger was determined by using HPLC technique according to Cafino et al. [27].

## 2.8 Induction of Hepatotoxicity and Nephrotoxicity

Hepatotoxicity and nephrotoxicity were induced in rats by intraperitoneal (i.p) injection with single dose of carbon tetrachloride [1 ml / kg b.wt]] which diluted with corn oil (1:1) immediately before administration to rats at the end of the 4<sup>th</sup> week of the experiment [28].

## 2.9 Experimental Design

Fifty-four adult Sprague-Dawley rats were divided into six groups each of nine rats. One g of G, GNPs and GNB powder were separately suspended in 100 ml distilled water immediately before administration to rats. Olive oil has some protective effect on liver injury; hence, corn oil may be a better option as a vehicle for CCl<sub>4</sub>.

Rats were treated as follows: Group I: Normal control, rats were orally received distilled water by gastric tube daily for 8 weeks; Group II: CCl<sub>4</sub>, rats were orally received distilled water daily for 8 weeks, and injected intraperitoneally (i.p.) with single dose of CCl<sub>4</sub> (1 ml/kg/b.wt.) in corn oil (1:1) at the end of fourth week. Groups (III), (IV), (V): Rats were orally given G, GNPs and GNB (50 mg /kg b.wt. /day), respectively for 8 weeks, and injected with CCl<sub>4</sub> as mentioned in group II. Group VI: (SM), rats were orally received SM (100 mg /kg b.wt. /day) for 8 weeks and injected with CCl<sub>4</sub> as mentioned in group II.

## 2.10 Handling of Blood and Tissue Samples

At the end of empirical interval and after 12 hours fasting, rats were sacrificed under anesthesia,

and blood samples were collected from hepatic portal vein. Serum was obtained by centrifugation of blood at 3000 rpm for 15 minutes after that stored frozen at -20°C for biochemical tests. Liver and kidneys were separated and plotted free from adhering blood, washed with cold physiological saline (0.9% NaCl) and dried between filter papers. Part of liver and kidneys tissues were obtained and immediately preserved in 10% buffered formalin solution (pH 7.4) for microscopic examination, the other part was kept frozen at (-20°C) till used for biochemical analysis.

## **2.11 Biochemical Assessment**

### **2.11.1 Determination of liver function parameters**

Serum alanine aminotransferase (ALT) and aspartate aminotransferase (AST) activities were determined using colorimetric kit according to Murray [29], alkaline phosphatase (ALP) activity was determined by colorimetric method using Spectrum Diagnostic kit according to Belfield and Goldberg [30].

### **2.11.2 Determination of kidney function biomarkers**

The determination of serum creatinine was performed according to kinetic colorimetric method [31]. Serum urea and uric acid were determined using enzymatic colorimetric method according to Kaplan [32] and Fossati et al. [33], respectively. Serum cystatin C was determined based on sandwich ELISA assay technique according to the method described by Pergande and Jung [34].

### **2.11.3 Assessment of inflammatory markers**

Serum tumor necrosis factor- alpha (TNF-  $\alpha$ ) and interleukin 1 beta (IL-1 $\beta$ ) levels were performed following sandwich ELISA technique using Rat ELISA kit according to Dowlati et al. [35] and Garabedian et al. [36], respectively.

### **2.11.4 Assessment of antioxidant and oxidative stress status**

Serum total antioxidant capacity (TAC) level was determined by enzymatic colorimetric method according to Koracevic et al. [37]. For assessment of hepatic CAT and SOD, the liver tissue was perfused with a phosphate buffer saline (PBS) solution, pH 7.4, containing 0.16 g/

ml heparin to remove any blood cell and clots. One gram of tissue was homogenized in 10 ml cold buffer (i-e, 100 mM potassium phosphate, pH 7.0, containing 2 mM EDTA). Then centrifuged at 4000 rpm for 15 minutes at 4 °C and the supernatant was removed. Hepatic CAT and SOD activities were determined colorimetrically using standard kit as described by Aebi [38] and Nishikimi et al. [39], respectively. Serum NO and MDA levels were determined by photometric method using standard kit according to Montgomery and Dymock [40] and Satoh [41], respectively.

## **2.12 Microscopic Examination of Liver and Kidneys Tissues**

Tissue samples from liver and kidneys were fixed in 10 % formalin solution for 24 hrs. After fixation, tissues were washed and dehydrated, then inserted in paraffin wax. The embedded tissues were sectioned to 5  $\mu$ m thick sections were subjected to microscopic examination. Hematoxylin and Eosin (H and E) stains were used with power magnification X 400 [42].

## **2.13 Statistical Analysis**

The data were statistically analyzed by one-way (ANOVA) and Post Hoc Tukey. A difference was considered significant when *p* was less than or equal to 0.05. The data was analyzed using (SPSS) statistics version 16.0 according to [43].

## **3. RESULTS AND DISCUSSION**

### **3.1 Characterization of Ginger Nanoparticles (GNPs) and Ginger Nano-base (GNB)**

The particle size of ginger nanoparticles and ginger nano-base were determined and evaluated according to size distribution by number. Fig. 1 shows that 99.5% of GNPs size was reached to 38.29 nm while Fig. 2 demonstrates that 99.9% of GNB size was reached to 22.05 nm.

Nanotechnology allow scientists to output nano-scale particles. The nanoparticles can effortlessly go among blood vessel walls, or even insert themselves into most of body cells when they are smaller than 20 nm or 50 nm which makes them an excellent choice for specialized drugs to be used in targeted delivery of plentiful doses of chemotherapeutic agents [44].

In the study of Mehanna et al., the ginger powder was ground using planetary ball mill device for 4 hours at 550 rpm to obtain ginger nanoparticles and the size of nanoparticles was evaluated by using DLS. They found that, 98.5% of GNPs reach to 75.59 nm [13].

### 3.2 Total Phenols and Flavonoids Content of Ginger

The data presented in (Table 1) indicate the total phenols expressed as mg of gallic acid equivalent (mg GAE /100 g of ginger) and total flavonoids expressed as mg of quercetin equivalent (mg QE /100 g of ginger). The results showed that each 100 g of the tested extract contains 2171.2 mg as gallic acid equivalent of total phenols and 343.75 mg as quercetin equivalent of total flavonoids.

Phenolic compounds are one of the principle groups of secondary metabolites in plants, they

have antioxidants and anti-inflammatory properties, regulate enzyme activity and gene expression [45]. Wijayanti et al. (2018) reported that the ethanol solvent is able to bind secondary metabolite compounds in the sample, they revealed that total phenolic content of 96% ethanolic extract of ginger was 485.1847 mg GAE/100 g dry weight [46].

Flavonoids constitute the largest percent of dietary polyphenols (more than 60%). Flavonoids apply their antioxidative activity by efficiently scavenging numerous free radical such as superoxide anion and peroxynitrite, by modulating oxidative stress-mediated enzyme activity and by catching the transition metals associated with processes of radical production [47]. Shirin and Prakash (2010) demonstrated that flavonoid content of 80% ginger ethanolic extract is 0.352 g quercetin/100 g ginger extract [48].

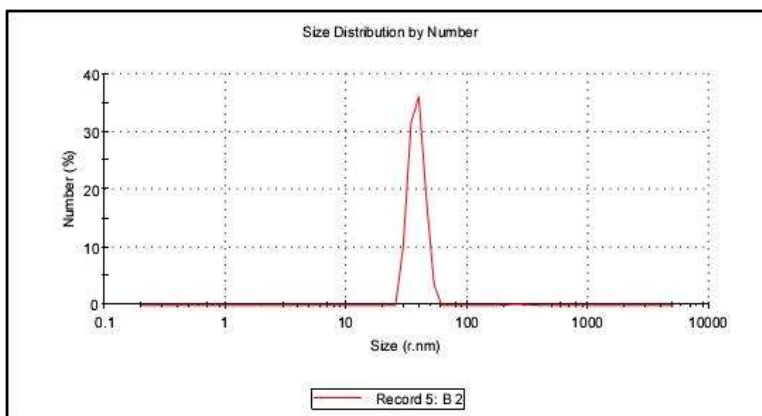


Fig. 1. Particle size of ginger nanoparticles (GNPs) was measured by dynamic light scattering (DLS) technique using a Malvern Zetesizer Nano ZS Apparatus

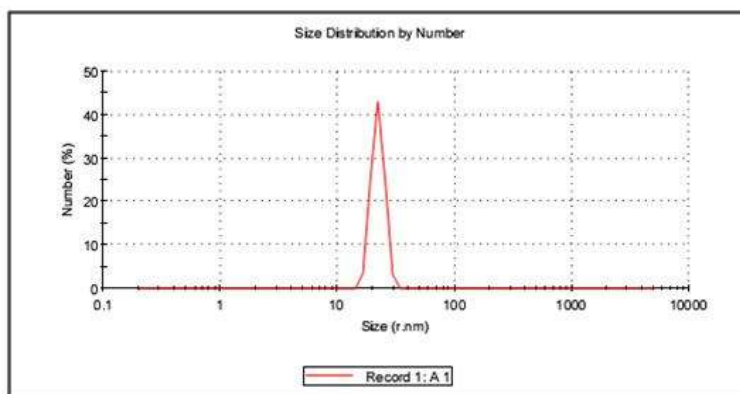


Fig. 2. Particle size of ginger nano-base (GNB) was measured by dynamic light scattering (DLS) technique using a Malvern Zetesizer Nano ZS Apparatus

**Table 1. Total flavonoid and phenolic contents of ginger ethanolic extract**

Active components	Value / 100 g of ginger
Total phenols	2171.2 mg as GAE
Total flavonoids	343.75 mg as QE

**3.3 The Concentration of Main Phenolic Compounds of Ginger**

Analysis of ginger extract using ultra high-performance liquid chromatography detected the presence of many active phenolic compounds. Fig. (3) indicate the percentage of main phenolic compounds in ginger extract as [6]-gingerol, [8]-gingerol, [6]-shogaol, [10]-gingerol, [8]-shogaol and [10]-shogaol. It is clear that [6]-gingerol represent the highest percentage (33.48%) of ginger phenolic components, while [10]-shogaol (2.03%) and [8]-shogaol (1.09%) were found in the lowest percentage.

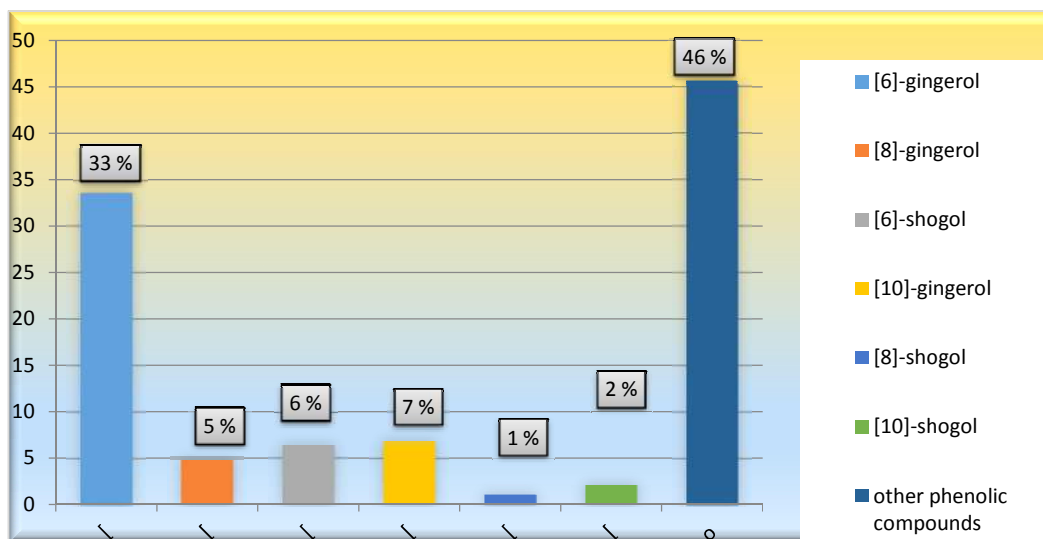
**3.4 Liver Function Biomarkers in Experimental Groups Intoxicated by CCl<sub>4</sub>**

A single dose of CCl<sub>4</sub> injection (1ml/kg b.wt.) resulted in hepatic damage as indicated by significant increase in serum ALT, AST and ALP enzymes activity (Table 2). Administration of G, GNPs, GNB and SM to CCl<sub>4</sub> administered rats significantly ( $P \leq 0.05$ ) lowered the activity of liver enzymes when compared with CCl<sub>4</sub> group. The most significant ameliorating effect on serum

ALT and ALP activities was seen in GNB treated group, while GNPs and SM treated groups showed the most significant enhancement in serum AST activity comparing to CCl<sub>4</sub> group.

CCl<sub>4</sub> intoxication is the experimental model of choice for induction of liver injury for many scientists all over the world. The damage of hepatocytes occurred by CCl<sub>4</sub> was suggested to be due to the fact that CCl<sub>4</sub> is metabolized by cytochrome P450 oxygenase enzyme system, producing trichloromethyl radical (CCl<sub>3</sub>·) and trichloromethyl peroxy radical (CCl<sub>3</sub>O<sub>2</sub>·) which invade lipids in the plasma membrane, resulting in initiation of fatty acids of phospholipid peroxidation and increase in MDA level with depletion in antioxidant enzymes, and finally leakage of plasma membrane and release of liver enzymes from hepatocyte [49].

The marked elevation of serum ALT, AST and ALP in CCl<sub>4</sub> treated group in our study indicates severe damage to hepatic tissue. These findings were in agreement with previous studies which indicated that administration of single dose of CCl<sub>4</sub> (1ml / kg B.W) significantly increased the serum level of ALT, AST and ALP enzymes in rats [49,50,51].



**Fig. 3. The percentage of main phenolic constituents of ginger that was indicated in the percentage of total phenols**

**Table 2. Serum alanine aminotransferase (ALT), aspartate aminotransferase (AST) and alkaline phosphatase (ALP) activities in the experimental groups (mean  $\pm$  SE).**

Parameters/ Groups	ALT (U/l)	AST (U/l)	ALP (U/l)
Control	11.049 $\pm$ 0.062 <sup>d</sup>	16.00 $\pm$ 0.543 <sup>e</sup>	106.81 $\pm$ 0.533 <sup>d</sup>
CCl <sub>4</sub>	18.197 $\pm$ 0.072 <sup>a</sup>	31.85 $\pm$ 0.169 <sup>a</sup>	180.19 $\pm$ 1.181 <sup>a</sup>
CCl <sub>4</sub> + G	13.978 $\pm$ 0.139 <sup>b</sup>	25.08 $\pm$ 0.126 <sup>b</sup>	120.75 $\pm$ 0.657 <sup>b</sup>
CCl <sub>4</sub> + GNPs	11.818 $\pm$ 0.096 <sup>c</sup>	23.12 $\pm$ 0.034 <sup>c</sup>	115.67 $\pm$ 0.744 <sup>c</sup>
CCl <sub>4</sub> + GNB	11.053 $\pm$ 0.139 <sup>d</sup>	24.48 $\pm$ 0.255 <sup>b</sup>	101.01 $\pm$ 0.919 <sup>e</sup>
CCl <sub>4</sub> + SM	11.582 $\pm$ 0.064 <sup>c</sup>	21.59 $\pm$ 0.578 <sup>d</sup>	106.47 $\pm$ 0.940 <sup>d</sup>

There is no significant difference between means have the same letters in the same column,  $n=9$  rats, ( $P \leq 0.05$ ). CCl<sub>4</sub>: Carbon tetrachloride, G: Ginger, GNPs: Ginger nano particles, GNB: Ginger nano- base, SM: Silymarin

Natural products contribute to the prevention, limitation and treatment of liver diseases. Ginger and silymarin are among the natural products with antioxidant effects so that they can be used to protect liver from hepatotoxicity [52]. In agreement with our finding, pretreatment with ginger extract (250 mg/ kg B.W) with CCl<sub>4</sub> (1 ml/kg B.W) caused significant decrease in serum ALT, AST and ALP, reversing the effect of CCl<sub>4</sub> due to the presence of gingerols and shogaols bioactive components that act as free radical scavenger and protect the liver cell membrane from oxidation and damage [53].

The nanoparticles of edible plant are highly reluctant to digestion by gastric, intestinal pancreatic juices and bile extract. Considerable quantities of plant nanoparticles are taken up by intestinal macrophages and stem cells. So that, the molecules involving lipids and RNA from plant nanoparticles may also possess an effect on the targeted cells positioned in the gut and liver [54]. The present study demonstrated that administration of ginger in the form of GNPs and GNB for 8 weeks exhibited more protection against CCl<sub>4</sub>- induced liver and kidney toxicity in rats in relative to native ginger.

In line with our result, Bakr et al. (2019) reported that administration of aqueous ginger extract (120 mg/kg B.W) and GNPs (120 mg/kg B.W) with acetaminophen caused significant decrease in serum ALT and AST activities and amelioration of microscopic examination of liver tissues as compared to acetaminophen group. The effect of GNPs on keeping live function tests was better than that of ginger extract [15].

Our results are also similar to the previous findings which showed that oral administration of ginger derived nanoparticles (GDNPs) with single dose of ethanol protect against alcohol-induced

liver injury through reduction of ALT and AST activities compared to ethanol group [55].

Ginger can protect liver due to its antioxidant properties which act as scavengers of free radicals, suppress lipid peroxidation and DNA destruction, that could ameliorate hepatocyte structure and function. It causes stabilization to the cell membrane, due to the presence of proteins and small peptide [7].

Silymarin was used in this study as a reference herbal drug for its well-identified hepatoprotective mechanism. The hepatoprotective actions of silymarin have been previously imputed to its antioxidant properties [56,57,58,59]. It was reported that silymarin inhibits liver injury by maintaining the integrity of the plasma membrane, inhibits the secretion of liver enzymes in blood, and suppresses apoptosis in the hepatocytes [60].

### 3.5 Kidney Function Biomarkers in Experimental Groups Intoxicated by CCl<sub>4</sub>

Our results revealed that rats treated with CCl<sub>4</sub> exhibited significant ( $P \leq 0.05$ ) increase in the serum urea, creatinine, uric acid and cystatin C levels compared to the control group (Table 3).

The treated groups showed a significant decrease ( $P \leq 0.05$ ) in the serum levels of creatinine, urea and cystatin C as compared to CCl<sub>4</sub>, with no significant difference between GNPs, GNB and SM groups. There was a significant ( $P \leq 0.05$ ) decrease in serum uric acid level in all treated groups except CCl<sub>4</sub> + G, which has no significant difference in comparison with CCl<sub>4</sub> group. The most effective treatment was appeared in GNPs comparing with GNB and SM.



It was demonstrated that acute subjection to  $\text{CCl}_4$  resulted in a significant drop in the renal function tests, which was noted through an increase in serum creatinine, urea and uric acid levels, this was in accordance with the present results [3,6]. A good explanation for  $\text{CCl}_4$ -induced nephrotoxicity was introduced by Akram et al. (2019), who suggested that  $\text{CCl}_4$ -reno toxic mechanism is almost like that of the liver, but cytochrome P-450 system is located in the renal cortex so  $\text{CCl}_4$  shows a high tendency to it. Renal toxicity caused a fast decay in the kidney functions that is principally ascribed to decline in glomerular filtration rate (GFR) and diminish the kidney capability to excrete the toxic metabolites formed in the body, resulting in abnormal detention of renal parameters [61].

Cystatin C is a non-glycosylated protein which produced in all nucleated cells, it is refined by glomeruli and then metabolized in proximal renal tubule cells following megalin-mediated endocytosis. Cys-C is the fundamental protector of the glomerular membrane from destruction by cysteine proteinases. The changed glomerular filtration rate also identified by the raised serum cystatin C because it is an preliminary marker of kidney problem than familiar markers (e.g. creatinine) and can reveal mild to moderate kidney problems [62]. In our study, the elevated level of Cys-C in  $\text{CCl}_4$  intoxicated rats indicated impairment in kidney function.

Our results were confirmed by previous study which documented that administration of aqueous ginger extract (500 mg/kg/day) along with  $\text{CCl}_4$  resulted in a significant decrease in the serum levels of creatinine, urea and uric acid compared to  $\text{CCl}_4$  group. The protective effect of ginger may be due to its antioxidant properties. Also, ginger contains vitamins A, B, C and E, magnesium and other trace elements, which behave as antioxidants against free radicals induced kidney injury [63]. Moreover, the existence of polyphenols and flavonoids in the ginger extract might be responsible for the antioxidant reno-protective activities and the lowering of elevated values of kidney function indices [64].

In agreement with our results, Bakr et al. (2019) reported that administration of aqueous ginger extract (120 mg/kg B.W) and GNPs (120 mg/kg B.W) with acetaminophen caused a significant decrease in serum urea and uric acid levels and enhancement of the kidney tissues as compared to acetaminophen group [15].

Silymarin has antioxidant and anti-inflammatory properties that may provide protective role against nephropathic processes. EL. Sayed et al. confirmed that administration of silymarin (200 mg/kg orally) with  $\text{CCl}_4$  caused significant decrease in serum level of creatinine and urea compared to  $\text{CCl}_4$  group [65].

### **3.6 Inflammatory Biomarkers in Experimental Groups Intoxicated by $\text{CCl}_4$**

It was observed from the present results that the mean value of serum TNF- $\alpha$  and IL-1 $\beta$  in  $\text{CCl}_4$  group were significantly ( $P \leq 0.05$ ) higher compared to control group. Administration of G, GNPs, GNB and SM attenuated the inflammatory effect of  $\text{CCl}_4$  which indicated by significant ( $P \leq 0.05$ ) reduction in TNF- $\alpha$  and IL-1 $\beta$  levels comparing to  $\text{CCl}_4$ -administrated group. Treatment with GNB has the best ameliorating effect on serum TNF- $\alpha$  level, while, GNPs and SM have the most enhancement effect on serum IL-1 $\beta$  level than other handlings comparing to  $\text{CCl}_4$  group (Table 4).

In the current study,  $\text{CCl}_4$ -treated rats exhibited severe inflammation and subsequently an overproduction of TNF- $\alpha$  and IL- $\beta$ . It was reported that intraperitoneal injection with (5 ml /kg B.W) of  $\text{CCl}_4$  resulted in significant raise in serum TNF- $\alpha$  and IL-1 $\beta$  levels which led to systemic inflammatory response syndrome. Absolute liberation of inflammatory agents can cause tissue injury.  $\text{CCl}_4$  stimulates macrophages and increases inflammatory cytokines that in turn intermediate the inflammatory responses and inflammation of liver. TNF- $\alpha$  trigger lymphocytes and neutrophils to motivate liberation of IL-1 $\beta$  [66].

Hepatic inflammation followed by liver injury developed by  $\text{CCl}_4$ -administration is attributed to activation of kupffer cells that lead to releasing reactive oxygen intermediates. ROS stimulate NF- $\kappa$ B and lead to massive expression of TNF- $\alpha$  and IL-1 $\beta$ . These cytokines expressed in the  $\text{CCl}_4$ -induced inflammatory process is related to the evolution of fibrosis. Protection from hepatic injury caused by  $\text{CCl}_4$  is associated with suppression of TNF- $\alpha$  level. So that, the inhibition of proinflammatory cytokines is used as indication for anti-inflammatory effect [67].

The active phenolic components of ginger have a strong anti-inflammatory property through suppression of TNF- $\alpha$  and IL-1 $\beta$  via inhibition of

**Table 3. Serum creatinine, urea, uric acid and cystatin C (Cys- C) levels in the experimental groups**

Parameters/Groups	Creatinine (mg/dl)	Urea (mg/dl)	Uric acid (mg/dl)	Cys – C (pg/ml)
Control	0.715 ± 0.012 <sup>c</sup>	39.675 ± 0.499 <sup>b</sup>	1.634 ± 0.025 <sup>d</sup>	0.962 ± 0.015 <sup>c</sup>
CCl <sub>4</sub>	1.078 ± 0.006 <sup>a</sup>	49.300 ± 0.653 <sup>a</sup>	2.989 ± 0.038 <sup>a</sup>	4.108 ± 0.126 <sup>a</sup>
CCl <sub>4</sub> + G	0.771 ± 0.011 <sup>b</sup>	38.901 ± 0.526 <sup>b</sup>	2.945 ± 0.053 <sup>a</sup>	1.231 ± 0.017 <sup>bc</sup>
CCl <sub>4</sub> + GNPs	0.726 ± 0.024 <sup>c</sup>	37.261 ± 0.271 <sup>c</sup>	2.212 ± 0.038 <sup>c</sup>	1.104 ± 0.018 <sup>b</sup>
CCl <sub>4</sub> + GNB	0.730 ± 0.006 <sup>c</sup>	37.571 ± 0.374 <sup>c</sup>	2.692 ± 0.079 <sup>b</sup>	1.159 ± 0.027 <sup>b</sup>
CCl <sub>4</sub> + SM	0.691 ± 0.015 <sup>c</sup>	37.259 ± 0.330 <sup>c</sup>	2.799 ± 0.019 <sup>b</sup>	1.194 ± 0.038 <sup>b</sup>

*There is no significant difference between means have the same letters in the same column, n= 9 rats, (P ≤ 0.05). CCl<sub>4</sub>: Carbon tetrachloride, G: Ginger, GNPs: Ginger nano particles, GNB: Ginger nano- base, SM: Silymarin*

COX-2 and NF- $\kappa$ B [68]. In the study performed by Mansour et al., (2019), treatment with ginger powder (108 or 216 mg/kg B.W/day) led to significant decrease in hepatic IL-1 $\beta$  and TNF- $\alpha$  as compared with di-ethylnitrosamine group (the same effect as CCl<sub>4</sub>) [68]. They suggested that the active components of ginger ([6]-gingerol and [6]-paradol) possess a strong anti-inflammatory activity [17].

GNPs were reported to reduce the chronic liver and kidney inflammation in rats by reducing the pro-inflammatory cytokines (TNF- $\alpha$  and IL-1 $\beta$ ) expression [69]. It was illustrated also that silymarin has anti-inflammatory properties which enable it to recur the inflammation caused by CCl<sub>4</sub> [51,70].

### 3.7 Oxidative Biomarkers in Experimental Groups Intoxicated by CCl<sub>4</sub>

Administration of CCl<sub>4</sub> (1 ml/kg b.wt.) was remarkably ( $P \leq 0.05$ ) elevated the oxidative biomarkers, MDA and NO levels, comparing to normal group. Co-administration of G, GNPs, GNB and SM with CCl<sub>4</sub> caused significant decline in serum MDA and NO relative to CCl<sub>4</sub> group. The supplementation with GNB caused the best improvement in the MDA level, while GNPs has the best influence on NO level when compared with toxic group (Table 5).

Our results revealed that CCl<sub>4</sub> caused increase in MDA level, the final product of tissue lipid peroxidation and stimulates the production of reactive nitrogen products as nitrite and NO, which afterward interacts with the superoxide anion to produce a peroxynitrite (ONOO<sub>2</sub>), a cytotoxic oxidant. Peroxynitrite caused DNA and mitochondrial membrane damage that evokes the process of the membrane fatty acid peroxidation or led to apoptosis. In corresponding to our study, it was reported that intraperitoneal injection with CCl<sub>4</sub> lead to

significant increase in serum NO and MDA levels. These findings assure that CCl<sub>4</sub> is a xenobiotic, caused oxidative stress and may injure hepatic cells [52].

It was illustrated that administration of ginger suspension (300 mg/kg/day) and CCl<sub>4</sub> (2 ml/kg B.W/ twice weekly) resulted in significant declines in serum MDA and NO levels compared to CCl<sub>4</sub> group. The beneficial effect of ginger is owing to the bioactive constituents like gingerols and shogaols that exhibited strong antioxidant activity. Gingerol inhibited lipid peroxidation induced by the ferric chloride-ascorbate system. Moreover, novel glucosides related to gingerdiol from ginger have antioxidant activity [71].

Additionally, Administration of silymarin and CCl<sub>4</sub> (2 ml/kg B.W) resulted in significant decrease in serum MDA and NO levels comparing to CCl<sub>4</sub> group. This effect was attributed to the antioxidant properties of silymarin and its ability to prevent lipid peroxidation and cell damage by oxidants [51].

### 3.8 Serum Total Antioxidant Capacity Level and Hepatic Antioxidant Enzymes Activity in Experimental Groups Intoxicated by CCl<sub>4</sub>

Table (6) clarifies that the serum TAC level and antioxidant defense enzymes (CAT and SOD) activity was significantly ( $P \leq 0.05$ ) declined in response to injection with CCl<sub>4</sub> compared to control group. Additionally, it was detected that serum TAC was significantly increased by treatment with SM, GNB, GNPs and G, respectively according to their effect, comparing to CCl<sub>4</sub> group. The activity of hepatic CAT and SOD was significantly raised in all treated groups comparing with CCl<sub>4</sub> group, the most significant increase in hepatic CAT levels was found in GNPs in relative to other treated groups.

**Table 4. Serum tumor necrosis factor- alpha (TNF- $\alpha$ ) and interleukin- 1 beta (IL-1 $\beta$ ) levels in the experimental groups (mean  $\pm$  SE)**

Parameters/Groups	TNF- $\alpha$ (pg/ml)	IL-1 $\beta$ (pg/ml)
Control	17.211 $\pm$ 0.164 <sup>e</sup>	33.657 $\pm$ 0.304 <sup>e</sup>
CCl <sub>4</sub>	92.969 $\pm$ 0.428 <sup>a</sup>	116.45 $\pm$ 0.159 <sup>a</sup>
CCl <sub>4</sub> + G	63.594 $\pm$ 0.639 <sup>b</sup>	74.451 $\pm$ 0.349 <sup>b</sup>
CCl <sub>4</sub> + GNPs	63.033 $\pm$ 0.441 <sup>b</sup>	69.442 $\pm$ 0.827 <sup>cd</sup>
CCl <sub>4</sub> + GNB	46.742 $\pm$ 0.323 <sup>d</sup>	70.869 $\pm$ 1.118 <sup>c</sup>
CCl <sub>4</sub> + SM	55.912 $\pm$ 0.468 <sup>c</sup>	68.916 $\pm$ 0.318 <sup>d</sup>

*There is no significant difference between means have the same letters in the same column, n= 9 rats, (P  $\leq$  0.05). CCl<sub>4</sub>: Carbon tetrachloride, G: Ginger, GNPs: Ginger nano particles, GNB: Ginger nano- base, SM: Silymarin*

**Table 5. Serum nitric oxide (NO) and malondialdehyde (MDA) levels in experimental groups (mean  $\pm$  SE)**

Parameters/Groups	NO ( $\mu$ mol/L)	MDA (nmol/ ml)
Control	0.392 $\pm$ 0.018 <sup>t</sup>	3.381 $\pm$ 0.074 <sup>d</sup>
CCl <sub>4</sub>	3.457 $\pm$ 0.074 <sup>a</sup>	8.111 $\pm$ 0.238 <sup>a</sup>
CCl <sub>4</sub> + G	1.896 $\pm$ 0.116 <sup>b</sup>	4.877 $\pm$ 0.186 <sup>b</sup>
CCl <sub>4</sub> + GNPs	0.770 $\pm$ 0.013 <sup>e</sup>	4.474 $\pm$ 0.085 <sup>bc</sup>
CCl <sub>4</sub> + GNB	0.986 $\pm$ 0.060 <sup>d</sup>	4.040 $\pm$ 0.145 <sup>c</sup>
CCl <sub>4</sub> + SM	1.298 $\pm$ 0.032 <sup>c</sup>	4.139 $\pm$ 0.156 <sup>c</sup>

There is no significant difference between means have the same letters in the same column, n= 9 rats, (P  $\leq$  0.05). CCl<sub>4</sub>: Carbon tetrachloride, G: Ginger, GNPs: Ginger nano particles, GNB: Ginger nano- base, SM: Silymarin

**Table 6. Serum total antioxidant capacity (TAC) level, hepatic Catalase (CAT) and hepatic superoxide dismutase (SOD) activities in the experimental groups (mean  $\pm$  SE)**

Parameters/Groups	TAC (mmol/l)	CAT (U/g)	SOD (U/g)
Control	1.496 $\pm$ 0.003 <sup>a</sup>	0.924 $\pm$ 0.003 <sup>a</sup>	249.75 $\pm$ 2.563 <sup>a</sup>
CCl <sub>4</sub>	0.686 $\pm$ 0.008 <sup>e</sup>	0.655 $\pm$ 0.002 <sup>e</sup>	132.51 $\pm$ 1.344 <sup>e</sup>
CCl <sub>4</sub> + G	1.224 $\pm$ 0.034 <sup>d</sup>	0.875 $\pm$ 0.001 <sup>d</sup>	168.24 $\pm$ 1.238 <sup>d</sup>
CCl <sub>4</sub> + GNPs	1.342 $\pm$ 0.013 <sup>c</sup>	0.904 $\pm$ 0.007 <sup>b</sup>	173.73 $\pm$ 1.669 <sup>d</sup>
CCl <sub>4</sub> + GNB	1.349 $\pm$ 0.0197 <sup>c</sup>	0.886 $\pm$ 0.003 <sup>c</sup>	188.11 $\pm$ 2.387 <sup>c</sup>
CCl <sub>4</sub> + SM	1.439 $\pm$ 0.011 <sup>b</sup>	0.898 $\pm$ 0.002 <sup>b</sup>	199.49 $\pm$ 2.027 <sup>b</sup>

There is no significant difference between means have the same letters in the same column, n= 9 rats, (P  $\leq$  0.05). CCl<sub>4</sub>: Carbon tetrachloride, G: Ginger, GNPs: Ginger nano particles, GNB: Ginger nano- base, SM: Silymarin

Hikal et al. showed a significant lowering in serum TAC and hepatic SOD and CAT activities after injection with CCl<sub>4</sub> (1 ml /kg B.W) in comparison with normal group, which go hand in hand with our results [50]. TAC points to all circumstances of the defense system against oxidation in body and has the powerful effect on diverse ROS production and elevated tendency to oxidative injury [66]. CCl<sub>4</sub> metabolism through cytochrome P450 enzyme system increase the oxidative stress in the body and in turn the defense antioxidant system try to scavenge these free radical, but excessive production and increased liberation of oxidant may lead to metabolic exhaustion and disruption of the antioxidant system, loss of homeostasis and finally oxidative destruction occur [3].

It was supposed that GNPs act as potent antioxidant compared to ginger extract. This may be attributed to that GNPs hold shogaols in bind shape compared to those carried by ginger extract that occur in free form. So that, GNPs carried low quantities of shogaols that required for acquiring the same effect on the hepatocytes compared with ginger extract. Shogaols organize many antioxidants and detoxication enzymes genetic expression [55,15].

Ginger has high antioxidant activity which was imputed to its high constituent of polyphenol

components such as ([6]-gingerol and it's derivatives) and diarylheptanoids, which possess a potent antioxidant activity [72]. The study of Hemieda et al. confirmed that, the antioxidant compounds of ginger were able to inhibit lipid peroxidation , scavenge the free radical such as NO, and suppress the pro-inflammatory mediator NO production by partially inhibiting enzymatic activity of inducible NO synthase [8].

### 3.9 Microscopic Examination of Liver Tissues in Experimental Groups Intoxicated by CCl<sub>4</sub>

The results of microscopic examination in our study provided supportive evidence for biochemical analysis. The histological examination of liver tissue from control rats detected normal hepatocyte structure (Fig. 4 A). Furthermore, the liver tissue from CCl<sub>4</sub> group revealed focal hepatocellular necrosis and apoptosis associated with mononuclear inflammatory cells infiltration (Fig. 4 B).

Liver section from CCl<sub>4</sub> + G group showed congestion of central vein and slight vacuolization of some hepatocytes (Fig. 5 A). Meanwhile, CCl<sub>4</sub> + GNPs and CCl<sub>4</sub> + GNB groups revealed slight cytoplasmic vacuolization of centrilobular hepatocytes (Figs. 5 B and 6 A), respectively. Congestion of central vein and

hepatic sinusoids was detected in CCl<sub>4</sub> + SM group (Fig. 6 B).

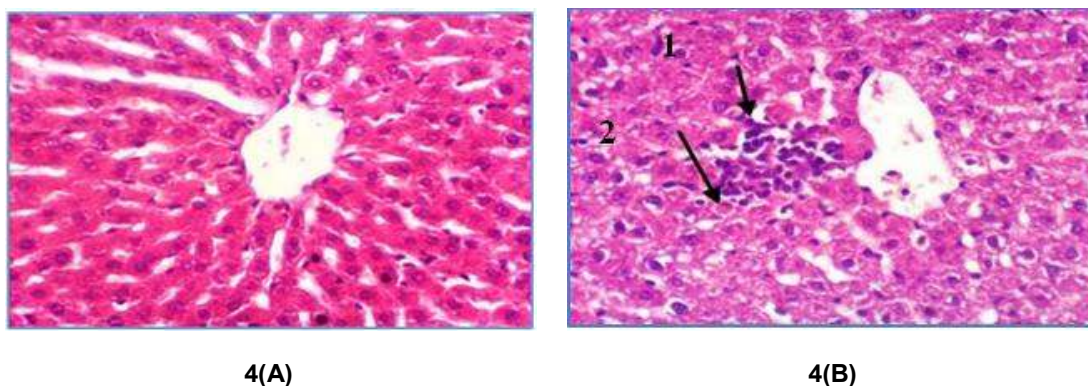
The microscopic examination provided supportive evidence for the hepatotoxic effect of CCl<sub>4</sub> that was evidenced by focal hepatocellular necrosis and apoptosis associated with mononuclear inflammatory cells infiltration, which agreed with previous studies [73,74].

The toxic effects of CCl<sub>4</sub> on liver that may ended by apoptosis or necrosis of hepatic cells is attributed to oxidative stress generated from CYT P450 metabolism by the highly reactive free radicals as CCl<sub>3</sub> and CCl<sub>3</sub>O<sub>2</sub>. These free radicals invade many molecules causing peroxidation of fatty acids of phospholipid, diminish of membrane integrity, leakage of liver enzymes, inflammation and finally liver injury. Prolonged liver injury leads to

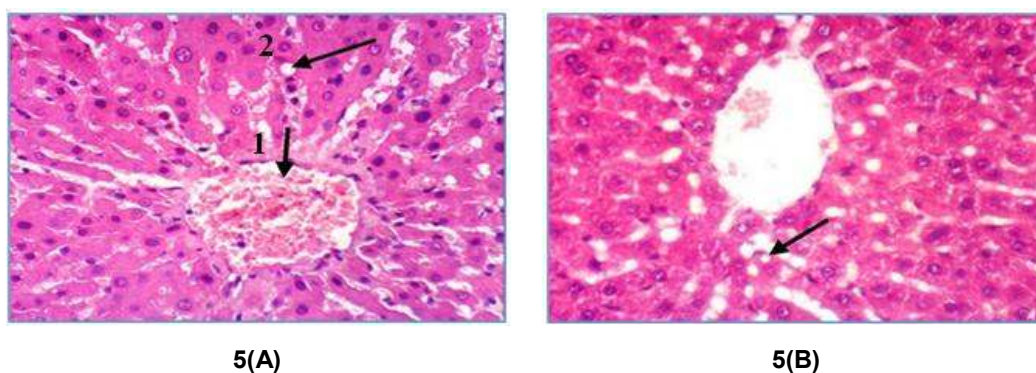
the development of liver fibrosis and cirrhosis [75].

We observed that ginger protected the architecture of hepatic tissue from damage by CCl<sub>4</sub> that induced liver necrosis, this effect may be owing to the presence of phenolic components as gingerols and shogaols which have antioxidant properties and are able to renewed endogenous antioxidant capacity with down regulating of the inflammatory response.

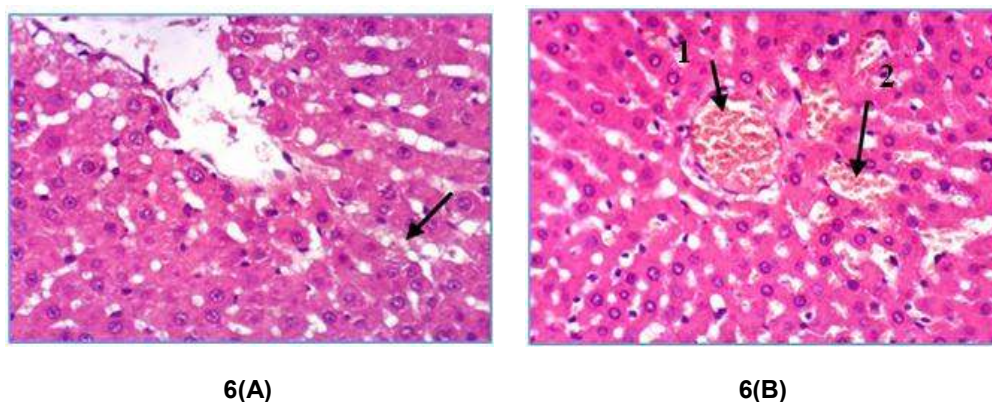
In the study of Bakr et al., the ginger nanoparticles- treated group followed by injection with CCl<sub>4</sub> showed more hepatoprotective and antioxidant impact than native ginger group, this may be due to that GNPs can afford digestion in stomach and intestine which enhance the delivery of nanoparticles and it's bioavailability within the hepatocytes. So, nanoparticles can reach to liver in considerable amounts [15].



**Fig. 4. (A):** Liver section of rat from normal group showing the normal histological structure of hepatic lobule. **(B):** Liver section of rat from CCl<sub>4</sub> group showing focal hepatocellular necrosis (1) and apoptosis associated with mononuclear inflammatory cells infiltration (2). (H & E X 400)



**Fig. 5. (A):** Liver section of rat from CCl<sub>4</sub> + G group showing congestion of central vein (1) and slight vacuolization of some hepatocytes (2). **(B):** Liver section of rat from CCl<sub>4</sub> + GNPs group showing slight cytoplasmic vacuolization of centrilobular hepatocytes (H & E X 400)



**Fig. 6. (A): Liver section of rat from CCl<sub>4</sub> + GNB group showing slight cytoplasmic vacuolization of sporadic hepatocytes. ( B): Liver section of rat from CCl<sub>4</sub> + SM group showing congestion of central vein (1) and hepatic sinusoids (2) (H & E X 400)**

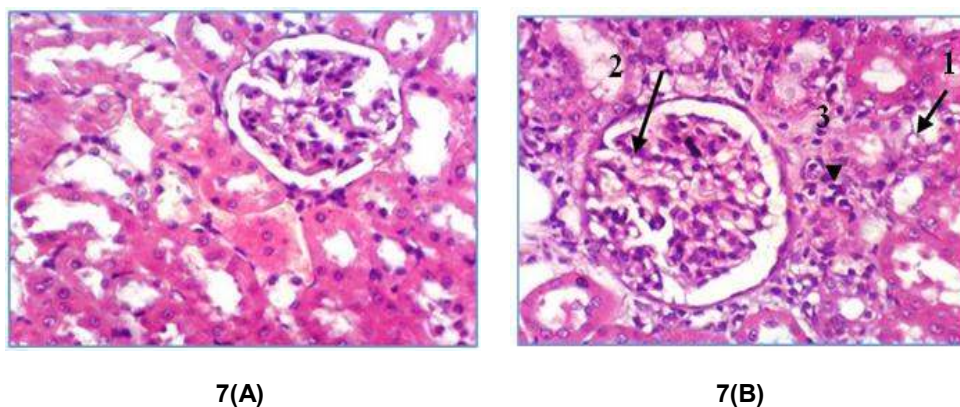
Pretreatment with silymarin inhibit liver necrosis and apoptosis caused by CCl<sub>4</sub>. Silymarin is a well-documented hepatoprotective agent, it comprises of flavonolignans complex and it is known for its antioxidant, anti inflammatory, antifibrotic impacts on the liver tissue. Also, it plays an important role as membrane-stabilizing agent, free radicals scavenging and suppression of lipid peroxidation and modulation of hepatocyte Ca<sup>+2</sup> [67,75].

### 3.10 Microscopic Examination of Kidneys Tissues in Experimental Groups Intoxicated by CCl<sub>4</sub>

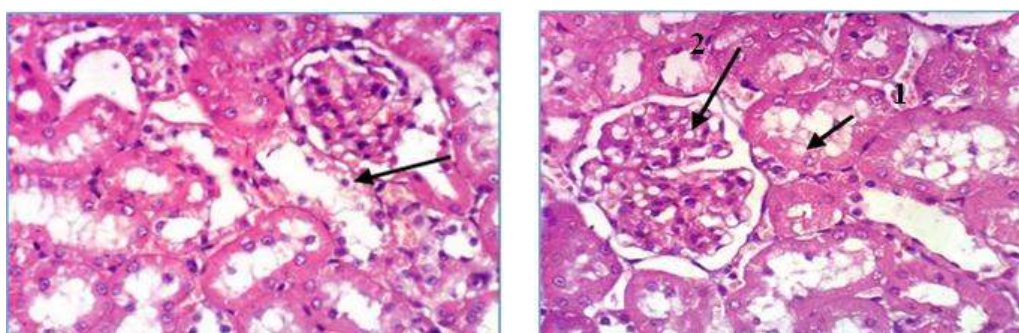
Kidney section of rats from control group showed normal renal tissues and histological structure of renal parenchyma, well defined glomeruli and Bowman's capsule, normal renal tubuli and

normal interstitial tissue with normal blood vessels (Fig. 7 A). Intraperitoneal injection with CCl<sub>4</sub> (1 ml/kg B.W) caused vacuolization of epithelial lining renal tubules and endothelial lining glomerular tuft as well as periglomerular inflammatory cells infiltration as shown in (Fig. 7 B). Slight vacuolization of epithelial lining renal tubules and endothelial lining glomerular tuft were found in all treated groups (Figs. 8 A, 8 B, 9A, 9B).

The changes occur in kidneys tissue by CCl<sub>4</sub> were induced by oxygen-derived free radicals and CCl<sub>4</sub> metabolites produced from CCl<sub>4</sub> metabolism by CYT P450 enzymes in renal cortex leading to lipid peroxidation in kidney cells. The changes of antioxidant status with CCl<sub>4</sub> may potentially cause nephropathies or renal disorders [61].



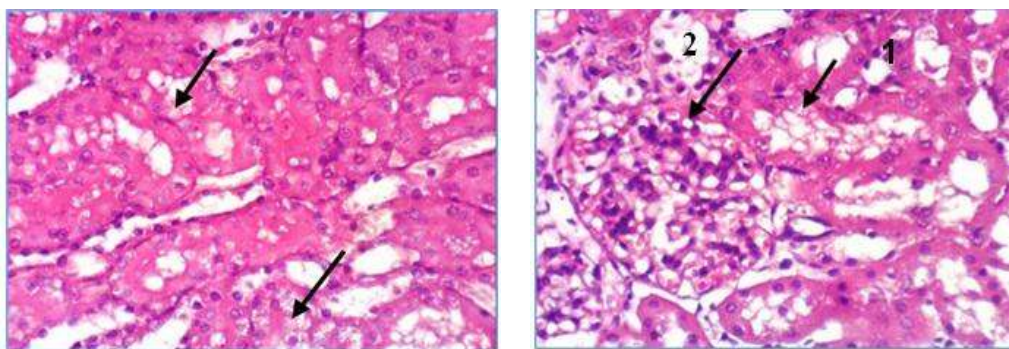
**Fig. 7. (A): Kidney section of rat from normal group showing the normal histological structure of renal parenchyma. (B): Kidney section of rat from CCl<sub>4</sub> group showing vacuolization of epithelial lining renal tubules (1) and endothelial lining glomerular tuft (2) as well as periglomerular inflammatory cells infiltration (3) (H & E X 400)**



8(A)

8(B)

**Fig. 8. (A):** Kidney section of rat from  $\text{CCl}_4$  + G group showing slight vacuolization of epithelial lining renal tubules.(B): Kidney section of rat from  $\text{CCl}_4$  + GNPs group showing slight vacuolization of epithelial lining renal tubules (1) and endothelial lining glomerular tuft (2) (H & E X 400)



9(A)

9(B)

**Fig. 9. (A):** Kidney section of rat from  $\text{CCl}_4$  + GNB group showing slight vacuolization of epithelial lining renal tubules (B): Kidney section of rat from  $\text{CCl}_4$  + SM group showing vacuolization of epithelial lining renal tubules (1) and endothelial lining glomerular tuft (2) (H & E X 400)

The preventive effect of ginger and silymarin contra  $\text{CCl}_4$  in organs including kidney, is attributed to the antioxidant, anti-inflammatory, scavenging the free radicals and membrane stabilizing effect [53].

Administration of GNPs and GNB resulted in enhancements in microscopically structures of kidneys tissues in rats as in liver tissue that impaired by  $\text{CCl}_4$  due to antioxidant, anti-apoptotic effect of them [15].

#### 4. CONCLUSION

From our study we concluded that, administration of  $\text{CCl}_4$  for 8 weeks led to hepatotoxicity and nephrotoxicity. Co-administration of G, GNPs, GNB and SM with  $\text{CCl}_4$  protect the liver and

kidney tissues from damage due to their high content of phenols and flavonoids, which enhance the antioxidant activities and protect against oxidative stress and lipid peroxidation. GNPs and GNB are more hepatoprotective and reno-protective against  $\text{CCl}_4$ - induced toxicity relative to the native ginger which may be due to their increased solubility and bioavailability.

#### ETHICAL APPROVAL

All authors hereby declare that "Principles of laboratory animal care" (NIH publication No. 85-23, revised 1985) were followed, as well as specific national laws where applicable. All experiments have been examined and approved by the appropriate ethics committee"

## COMPETING INTERESTS

Authors have declared that no competing interests exist.

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