Note



Effect of Jasmonates and Related Compounds on Seed Germination of *Orobanche minor* Smith and *Striga hermonthica* (Del.) Benth

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Jasmonates and related compounds were found to elicit the seed germination of the important root parasites, clover broomrape (*Orobanche minor* Smith) and witchweed [*Striga hermonthica* (Del.) Benth]. The stimulation of seed germination by the esters was more effective than by the corresponding free acids, and methyl jasmonate (MJA) was the most active stimulant among the compounds tested.

Key words: cucurbic acid; jasmonic acid; *Orobanche minor* Smith; seed germination; *Striga hermonthica* (Del.) Benth

Broomrapes (Orobanche spp.) and witchweeds (Striga spp.) are the two most important root parasitic weeds that cause considerable yield reduction to various crops in tropical and semi-tropical regions. 1) The seeds of these parasites have special germination requirements, including pretreatment (conditioning) in a warm, moist environment for several days and subsequent exposure to an exogenous germination stimulant.^{2,3)} Once germinated, these parasites will die within a week unless they attach to the roots of host plants. Therefore, inducing seed germination in the absence of host plants, termed "suicidal germination," is a promising strategy for depleting seed reserves in the soil. However, natural Striga and Orobanche germination stimulants such as strigol, 4,5) sorgolactone, 6) alectrol, 7) and dihydrosorgoleone8) are unstable in soil, and no useful and economic suicidal germination stimulants based on these molecules have been obtained. 3) Under these circumstances, germination stimulants structurally unrelated to the natural ones are important molecular probes to understand the germination mechanism of the parasites, and also to develop useful and economic germination stimulants. We recently found that the fungal metabolites, cotylenins and fusicoccins, which have diverse physiological effects on plants,9 could elicit the seed germination of Orobanche minor Smith and Striga hermonthica (Del.) Benth. 10) This suggests that other compounds with different chemistry may induce germina-

In the search for new germination stimulants, various compounds were first examined for their effects on O.

minor germination. In general, Orobanche spp. appeared to have stricter germination requirements than Striga spp., and the compounds which elicited Orobanche germination also induced Striga germination. So far, except for cotylenins and fusicoccins, only strigolrelated compounds have been reported to induce Orobanche germination.3) In contrast, in addition to cotylenins, fusicoccins, and strigol-related compounds. Striga germination is induced by other chemicals including ethylene, cytokinins, and auxins. Although various plant-growth regulators (gibberellins, cytokinins, auxins, abscisic acid, ethylene, and brassinosteroids) have been found not to induce Orobanche germination, 3,11) jasomonic acid (JA, Fig. 1) had not been examined for its effect on the seed germination of these parasites. Therefore, JA was included in the assay and found to induce the germination of O. minor. JA and related compounds were then examined for their effects on the seed germination of O. minor and S. hermonthica.

Seed germination assays were conducted as reported, 10,111) except that smaller glass Petri dishes (i.d. 4.5 cm) were used in this study. Each test compound was dissolved in acetone. An aliquot (0.1 ml) of the respective test solution was added to each Petri dish lined with a filter paper. The solvent was allowed to evaporate before conditioned seeds were placed on the filter paper and treated with distilled water (0.7 ml). Racemic mixtures of JA and related compounds were prepared as described elsewhere. 12,13) Seeds treated with 10^{-8} M (\pm)strigol, which had been synthesized as described in the literature, 14,15) or distilled water was used as the controls. Each treatment was replicated three times and conducted at least twice. The data presented in the tables are from single typical experiments, because the germination percentages may have varied from test to test.¹⁶⁾

In general, the response by seeds of both parasites to germination stimulants including (\pm)-strigol was rather low when compared to that reported earlier. This low germination response could have been due to the difference in size of the Petri dishes, or simply due to a season-dependent variation. Seasonal variations in response to germination stimulants have been reported for both *Striga* and *Orobanche* spp. Among the JA derivatives (1-3), methyl jasmonate (MJA, 1) at 10^{-4} M

Fig. 1. Structure of Jasmonic Acid (JA), Cucurbic Acid (CA), Their Related Compounds and Strigol.

5, $R_1=(Z)$ -3-pentenyl, $R_2=CH_2CO_2Me$; 8, $R_1=(Z)$ -3-pentenyl, $R_2=CH_2CO_2H$; 9, $R_1=(Z)$ -3-pentenyl, $R_2=CH_2CH_2OH$; 10, $R_1=$ pentyl, $R_2=CH_2CO_2Me$.

induced the highest germination of O. minor (Table 1). The activity of 7-epi-MJA (2) was less than 50% that of MJA. Furthermore, JA (3) was only 1/10 as active as MJA, suggesting that the ester function was preferable for high activity and/or, presumably, for penetration into the seeds. The structural and stereochemical requirements for germination-stimulative activity can be seen in more detail with the cucurbic acid (CA) derivatives (4– 12), of which the most active stimulant was 6-epi-9,10-dihydro-MCA (10). The ester, 6-epi-MCA (5), was about 2-fold as active as the free acid, 6-epi-CA (8). This difference in activity is relatively small when compared to that observed between MJA and JA. On the other hand, reduction of the carboxyl group to the alcohol, as in 9, resulted in a significant loss of activity. This implies that the carboxyl group played an important role in stimulating germination. When comparing the activities of com-

Table 1. Percentage Germination for Seeds of *O. minor* after Exposure to Solutions of Jasmonate and Its Related Compounds

Germination±SE (%)*			
No.	Compounds**	10 ⁻⁴ M	$10^{-5} \mathrm{M}$
1	MJA	67 ± 2.3	3±0.6
2	7-epi-MJA	29 ± 2.7	2 ± 0.5
3	JA	6 ± 0.8	1 ± 0.2
4	MCA	7 ± 1.0	0
5	6-epi-MCA	42 ± 2.3	1 ± 0.3
6	7-epi-MCA	22 ± 1.8	0
7	6,7-di- <i>epi</i> -MCA	4 ± 0.8	0
8	6-epi-CA	20 ± 1.4	0
9	6- <i>epi</i> -CA-ol***	2 ± 0.5	0
10	6-epi-9,10-dihydro-MCA	58 ± 2.0	3 ± 0.3
11	6-epi-CA lactone	0	0
12	6,7-di-epi-CA lactone	39 ± 2.3	0
	(\pm) -strigol****	(68 ± 2.0)	
	control	0	0

^{*} Mean \pm SE (n=3).

Table 2. Percentage Germination for Seeds of *S. hermonthica* after Exposure to Solutions of Jasmonate and Its Related Compounds

Germination ± SE (%)*			
No.	Compound**	$10^{-3} \mathrm{M}$	$10^{-4} \mathrm{M}$
1	MJA	50±1.2	12±2.3
3	JA	26 ± 1.9	6 ± 2.1
10	6-epi-9,10-dihydro-MCA	22 ± 1.8	6±1.3
	(\pm) -strigol***	(44 ± 3.6)	
	control	0	0

^{*} Mean \pm SE (n=3).

pounds 4-7 which have different relative configurations at the 3, 6 and 7 positions on the cyclopentane ring, the 6,7-cis configuration seems to have been preferable for high activity. For example, (3,6,7-all cis) 6-epi-MCA (5) was the most active. MCA (4) and 6,7-di-epi-MCA (7), the two 6,7-trans derivatives, were 1/6 and 1/10 as active as 6-epi-MCA (5), respectively. In contrast, the relative stereochemical relationship between the pentenyl side chain and the acetate moiety appears to have been of lesser importance; (3,7-trans) 7-epi-MCA (6) was 1/2 as active as (3,7-cis) 6-epi-MCA (5). In addition, with the cucurbic acid derivatives, the unsaturated pentenyl group at the 7 position seems to have reduced activity. The compound carrying an saturated alkyl side chain (6epi-9,10-dihydro-MCA, 10) was more active than 6-epi-MCA (5). The germination stimulative activity of the CA lactones (11 and 12) suggests that these two compounds may have interacted directly with the receptor(s) of germination stimulants, since 6-epi-CA lactone (11), which would afford the moderately active 6-epi-CA (8) after hydrolysis, was totally inactive, whereas 6,7-di-epi-CA lactone (12) was much more active than 6,7-di-epi-MCA (7). It is thus apparent that MJA (1) and 6-epi-9,10-dihydro-MCA (10) were the most active among the JA- and CA-related compounds, respectively. Furthermore, the high activity of 6-epi-9,10-dihydro-MCA (10) suggests that proper modification of the pentenyl side chain may further increase the activity, not only in CA derivatives but also in JA derivatives. At 10⁻⁵ M, these compounds induced only negligible germination.

MJA (1), JA (3) and 6-epi-9,10-dihydro-MCA (10) were then examined for their effects on *S. hermonthica* germination (Table 2). At 10⁻⁴ M, all the compounds elicited negligible germination (6-12%). At 10⁻³ M, MJA (1), JA (3) and 6-epi-9,10-dihydro-MCA (10) induced 22-50% germination. The relative order of activity is somewhat different from that observed with *O. minor* seeds, JA (3) being as active as 6-epi-9,10-dihydro-MCA (10).

JA and MJA display various physiological effects in plants, and they inhibit or promote seed germination depending on the plant species. ^{17,18)} This study has shown that JA and related compounds could induce seed germination of the root parasitic weeds, *O. minor* and *S. hermonthica*, which have special germination requirements. Although the activity of JA and related compounds was rather weak, proper modification of

^{**} Racemic mixture.

^{***} Alcohol derivative of 6-epi-CA.

^{**** (} \pm)-Strigol was tested at 10^{-8} M.

^{**} Racemic mixture.

^{*** (} \pm)-Strigol was tested at 10^{-8} M.

their structures, in particular, of the ester and alkenyl moieties, ¹³⁾ may lead to more active stimulants. Furthermore, the activity of individual enantiomers as germination stimulants should be examined. The stereochemistry of JA and related compounds appears to have had a pronounced effect on their activity.

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References

- 1) Stewart, G. R. and Press, M. C., The physiology and biochemistry of parasitic angiosperms. *Annu. Rev. Plant Physiol. Plant Mol. Biol.*, 41, 127-151 (1990).
- Worsham, A. D., Germination of witchweed seeds. In "Parasitic Weeds in Agriculture," Vol. 1, ed. Musselman, L. J., CRC Press, Boca Raton, FL, pp. 45-61 (1987).
- Joel, D. M., Steffens, J. C., and Matthews, D. E., Germination of weedy root parasites. In "Seed Development and Germination," ed. Kigel, J. and Galili, G., Marcel Dekker, New York, pp. 567-597 (1995).
- Cook, C. E., Whichard, L. P., Turner, B., Wall, M. E., and Egley, G. H., Germination of witchweed (*Striga lutea*): isolation and properties of a potent stimulant. *Science*, 154, 1189-1190 (1966).
- Cook, C. E., Whichard, L. P., Wall, M. E., Egley, G. H., Coggon, P., Luhan, P. A., and McPhail, A. T., Germination stimulants. II. The structure of strigol—a potent seed germination stimulant for witchweed (*Striga lutea Lour.*). *J. Am. Chem.* Soc., 94, 6198-6199 (1972).
- 6) Hauck, C., Müller, S., and Schildknecht, H., A germination stimulant for parasitic flowering plants from *Sorghum bicolor*, a genuine host plant. *J. Plant Physiol.*, **139**, 474-478 (1992).
- 7) Müller, S., Hauck, C., and Schildknecht, H., Germination

- stimulants produced by *Vigna unguiculata* Walp cv Saunders Upright. *J. Plant Growth Regul.*, **11**, 77–84 (1992).
- 8) Lynn, D. G., Steffens, J. C., Kamut, V. S., Graden, D. W., Shabanowitz, J., and Riopel, J. L., Isolation and characterization of the first host recognition substance for parasitic angiosperms. J. Am. Chem. Soc., 103, 1868-1870 (1981).
- 9) Marrè, E., Fusicoccin: a tool in plant physiology. *Annu. Rev. Plant Physiol.*, **30**, 273-288 (1979).
- 10) Yoneyama, K., Takeuchi, Y., Ogasawara, M., Konnai, M., Sugimoto, Y., and Sassa, T., Cotylenins and fusicoccins stimulate seed germination of *Striga hermonthica* (Del.) Benth and *Orobanche minor* Smith. *J. Agric. Food Chem.*, 46, 1583-1586 (1998).
- 11) Takeuchi, Y., Omigawa, Y., Ogasawara, M., Yoneyama, K., Konnai, M., and Worsham, A. D., Effects of brassinosteroids on conditioning and germination of clover broomrape (*Orobanche minor*) seeds. *Plant Growth Regul.*, 16, 153-160 (1995).
- Seto, H. and Yoshioka, H., An efficient and stereocontrolled syntheses of (±)-methyl epijasmonate and (±)-cucurbic acid. Chem. Lett., 1797–1800 (1990).
- 13) Seto, H., Kamuro, Y., Qian, Z.-H., and Shimizu, T., Structure-activity relationships of (±)-cucurbic acid analogs on the root growth of rice seedlings and height of young corn plants. *Nippon Noyaku Gakkaishi*, 17, 61-67 (1992).
- 14) Brooks, D. W., Bevinakatti, H. S., Kennedy, E., and Hathaway, J., Practical total synthesis of (±)-strigol. J. Org. Chem., 50, 628-638 (1985).
- Dailey, O. D., Jr., A new synthetic route to (±)-strigol. J. Org. Chem., 52, 1984-1989 (1987).
- 16) Mangnus, E. M., Stommen, P. L. A., and Zwanenburg, B., A standardized bioassay for evaluation of potential germination stimulants for seeds of parasitic weeds. J. Plant Growth Regul., 11, 91-98 (1992).
- 17) Sembdner, G. and Parthier, B., The biochemistry and the physiological and molecular actions of jasmonates. *Annu. Rev. Plant Physiol. Plant Mol. Biol.*, **44**, 569-589 (1993).
- 18) Creelman, R. A. and Mullet, J. E., Biosynthesis and action of jasmonates in plants. Annu. Rev. Plant Physiol. Plant Mol. Biol., 48, 355-381 (1997).