

Effect of plant growth regulators on the accumulation of indolizidine alkaloids in *Securinega suffruticosa* callus cultures

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Abstract In the presented work, plant growth regulators (PGRs) were tested for their influence on biomass growth and accumulation of therapeutically-relevant indolizidine alkaloids in *Securinega suffruticosa* (Phyllanthaceae) callus cultures. The study included 9 auxins [2,4-dichlorophenoxyacetic acid (2,4-D), 2,4,5-trichlorophenoxyacetic acid, indole-3-acetic acid (IAA), 4-chlorophenoxyacetic acid, indole-3-butyric acid, 4-amino-3,5,6-trichloropicolinic acid, β -naphthoxyacetic acid, 1-naphthaleneacetic acid, indole-3-propionic acid, one polar auxin transport inhibitor (2,3,5-triiodobenzoic acid) and 7 cytokinins [2-isopentenyladenine, 1-phenyl-3-(1,2,3-thiadiazol-5-yl)urea, *N*-(2-chloro-4-pyridinyl)-*N'*-phenylurea, diphenylurea, zeatin, kinetin (KIN), 6-benzylaminopurine]. The respective PGRs were applied at 0.5 and 5.0 mg l⁻¹, together with 5.0 or 0.5 mg l⁻¹ KIN or 2,4-D (for auxins and cytokinins, respectively). The calli subjected to different phytohormone combinations (39 modifications in total) were evaluated for growth and alkaloid content, and the results tested for statistical differences. The highest concentrations of securinine (1.73 mg g⁻¹ DW) and allosecurinine (3.11 mg g⁻¹ DW) were recorded in the callus grown in the presence of 0.5 mg l⁻¹ IAA and

5.0 mg l⁻¹ KIN. To the best of the authors' knowledge, this is the first such comprehensive report concerning the influence of PGRs on alkaloid accumulation in plant in vitro cultures.

Keywords Indolizidine alkaloids · Securinine · Auxins · Cytokinins · Phyllanthaceae

Abbreviations

AS	Allosecurinine
BAP	6-Benzylaminopurine
4-CPA	4-Chlorophenoxyacetic acid
CPPU	<i>N</i> -(2-chloro-4-pyridinyl)- <i>N'</i> -phenylurea (forchlorfenuron)
2,4-D	2,4-Dichlorophenoxyacetic acid
DPU	Diphenylurea
DW	Dry weight
G _i	Growth index
HPTLC	High performance thin layer chromatography
IAA	Indole-3-acetic acid
IBA	Indole-3-butyric acid
2iP	6-(γ,γ -Dimethylallylamino)purine (2-isopentenyladenine)
IPA	Indole-3-propionic acid
KIN	Kinetin
NAA	1-Naphthaleneacetic acid
BNOA	β -Naphthoxyacetic acid
PGR	Plant growth regulator
PCL	4-Amino-3,5,6-trichloropicolinic acid (picloram)
S	Securinine
SH	Schenk–Hildebrandt
2,4,5-T	2,4,5-Trichlorophenoxyacetic acid
TDZ	1-Phenyl-3-(1,2,3-thiadiazol-5-yl)urea (thidiazuron)
TIBA	2,3,5-Triiodobenzoic acid
ZEA	Zeatin

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Introduction

Securinine (S), the main pharmacologically active constituent of *Securinega suffruticosa* (Pall.) Rehd., Phyllanthaceae, is an indolizidine alkaloid and γ -aminobutyric acid_A receptor (GABA_AR) competitive antagonist. S is considered a promising agent for the prevention and treatment of neurodegenerative diseases, including Alzheimer's disease, amyotrophic lateral sclerosis and post-alcohol degeneration (Raj and Luczkiewicz 2008; Gupta et al. 2011). Apart from the above properties, S was shown to exhibit cytotoxic activity against colon (Rana et al. 2010) and breast cancer cell lines (Li et al. 2014), as well as inhibit toxoplasma growth (Holmes et al. 2011). The second main alkaloid of *S. suffruticosa*, allosecurinine (AS), acts as a cholinergic agent (Raj and Luczkiewicz 2008).

In previous years, in vitro cultures of *S. suffruticosa* were established in order to provide an efficient and reliable source of securinega alkaloids (Wysokińska 1979; Ide et al. 1986; Ide 1991; Yuan et al. 2005, 2007; Raj et al. 2009, 2015). Despite promising results in terms of alkaloid accumulation, these reports provided limited data on the influence of phytohormones on the accumulation of S and related compounds in the investigated biomasses. Given that the biosynthesis of alkaloids in plant cell cultures is strongly affected by both the type and concentration of PGRs used (Verpoorte et al. 1991), modifications of phytohormone composition can be considered a simple, yet effective tool for improving the productivity of an in vitro system. However, contrary to other alkaloid groups such as indole (Verpoorte et al. 1991; van der Heijden et al. 2004; Pietrosiuk et al. 2007) and diterpene derivatives (Sabater-Jara et al. 2010; Malik et al. 2011), reports concerning the influence of exogenous PGRs on indolizidine alkaloid accumulation are rather scarce and limited to the most commonly used phytohormones like 2,4-dichlorophenoxyacetic acid (2,4-D), 1-naphthaleneacetic acid (NAA), and kinetin (KIN) (Wysokińska 1979; Ide 1991; Yuan et al. 2007).

The aim of the presented work was to investigate the influence of different combinations of PGRs (39 modifications in total) on biomass growth and the accumulation of S and AS in *S. suffruticosa* callus cultures. Consequently, the experiment led to the selection a phytohormone composition providing the highest concentrations of the investigated compounds. Correlations between callus growth and the accumulation of indolizidine alkaloids were established and possible relationships between the chemical structure of the PGRs used and the observed effects were also discussed. To the authors' knowledge, this is the first such comprehensive study on the influence of phytohormones on alkaloid accumulation in a single in vitro model.

Materials and methods

Plant material

The experiments were conducted with the use of previously established (Raj et al. 2009) *S. suffruticosa* callus cultures, grown on solidified (0.7 % w/v agar) Schenk–Hildebrandt (SH) medium (Schenk and Hildebrandt 1972) containing 5.0 mg l⁻¹ 2,4-D, 5.0 mg l⁻¹ KIN and 3 % w/v sucrose, and subcultured at 4-week intervals.

Experimental procedures and culture conditions

For the experiment, 1 g portions of *S. suffruticosa* callus, taken on 30 d of the growth cycle, were transferred to baby food jars (Sigma-Aldrich, St. Louis, US-MO) filled with 25 ml of the solidified (0.7 % w/v agar) SH media supplemented with 3 % (w/v) sucrose and the respective phytohormones (10 replicates per modification, 2 series of 5 repetitions). The cultures were maintained at 24 ± 1 °C under a 16/8 (light/dark) photoperiod (88 ± 8 μmol m⁻² s⁻¹, Philips TLD 35 fluorescent lamps). After 4-weeks, the calli were harvested and their growth indices (Gi) calculated as described by Yuan et al. (2007). After freeze-drying (Lyovac GT2, Fin-Aqua-Santasalo-Sohlberg Co, Tuusula, Finland), the samples were evaluated for S and SA content using the previously developed HPTLC-densitometric method (Raj et al. 2009).

PGRs used in the study were either sterile-filtered [forchlorfenuron (CPPU), diphenylurea (DPU), indole-3-acetic acid (IAA), 2-isopentenyladenine (2iP), thidiazuron (TDZ), 2,3,5-triiodobenzoic acid (TIBA); Schleicher & Schuell 0.2 μm CA filters, GE Healthcare, Little Chalfont, UK] into the pre-autoclaved media or co-autoclaved (1 bar, 120 °C, 20 min) with other media components (all reagents from Sigma-Aldrich, purified water produced by double distillation). The pH of the media was adjusted to 5.8 prior to autoclaving.

Experimental scheme

In order to test the effect of auxins and anti-auxins on the growth rate and accumulation of S and AS, the KIN-containing media were supplemented interchangeably with 2,4-D, IAA, NAA, TIBA, 4-chlorophenoxyacetic acid (4-CPA), indole-3-butyric acid (IBA), indole-3-propionic acid (IPA), β-naphthoxyacetic acid (BNOA), 2,4,5-trichlorophenoxyacetic acid (2,4,5-T) and picloram (PCL). The respective PGRs and KIN were applied at both 1:10 and 10:1 ratio—the media containing 5.0 or 0.5 mg l⁻¹ KIN were supplemented with 0.5 or 5.0 mg l⁻¹ auxin/auxin inhibitor, respectively. *S. suffruticosa* calli grown on

SH media supplemented solely with 0.5 or 5.0 mg l⁻¹ KIN (C1 and C2, respectively) and the callus maintained on PGR-free SH medium (C0) were the reference groups.

Analogously, the influence of cytokinins on callus growth and the accumulation of the indolizidine alkaloids was investigated by enriching the 2,4-D-supplemented media interchangeably with CPPU, DPU, 2iP, KIN, TDZ, 6-benzylaminopurine (BAP) and zeatin (ZEA). The cytokinins and 2,4-D were applied at both 1:10 and 10:1 ratio—the media containing 5.0 or 0.5 mg l⁻¹ 2,4-D were supplemented with 0.5 or 5.0 mg l⁻¹ auxins, respectively. *S. suffruticosa* calli grown on SH media supplemented solely with 0.5 or 5.0 mg l⁻¹ 2,4-D (C3 and C4, respectively) and the callus maintained on PGR-free SH medium (C0) were the reference groups.

In the next part of the study, the PGRs which provided the best results in terms of indolizidine alkaloid content (IAA and ZEA of auxin and cytokinin group, respectively) were applied jointly at 1:10 and 10:1 ratio (2 modifications: 0.5 mg l⁻¹ IAA + 5.0 mg l⁻¹ ZEA and 5.0 mg l⁻¹ IAA + 0.5 mg l⁻¹ ZEA). *S. suffruticosa* calli grown in the presence of 0.5 mg l⁻¹ IAA and 5.0 mg l⁻¹ KIN (modification providing the highest alkaloid concentration in both variable auxin and variable cytokinin groups) and on the PGR-free medium were the reference groups.

Statistical analysis

The differences in Gi values and S and AS concentrations observed between the calli grown on media with different PGR composition were tested for statistical significance ($p < 0.05$) using the analysis of variance, followed by Tukey's range test (Statistica 10.0 software, StatSoft, Tulsa, US-OK). Correlations between Gi and accumulation of S and A were also investigated for each of the experiments ($p < 0.05$).

Results

The effects of different PGR compositions on the growth rate (Gi value) and indolizidine alkaloid content of *S. suffruticosa* callus were presented in Fig. 1. The biomass maintained on the PGR-free medium (C0 modification—Fig. 2a) for 30 d was characterized by relatively low Gi value (218 %). AS concentration in the biomass (1.42 mg g⁻¹ DW) was over twofold higher than that of S (0.61 mg g⁻¹ DW).

2,4-D added at 0.5 mg l⁻¹ as the sole PGR (Fig. 1d, C3) significantly stimulated callus growth (Gi = 471 %, over twofold increase compared to C0 group) but this effect was not observed when 5.0 mg l⁻¹ concentration was applied (Fig. 1c, C4). 2,4-D decreased the accumulation of S and

AS in a dose-dependent manner, with stronger inhibitory effect at higher concentration.

The addition of 0.5 or 5.0 mg l⁻¹ KIN (C1 and C2, respectively, Fig. 1a, b) had no significant effect on callus growth, but significantly increased the accumulation of both alkaloids. No clear dose-related effects were observed, however, lower KIN concentration (C1) favoured AS production (3.12 mg g⁻¹, ca. twofold increase as compared to C0 (Fig. 1b).

Auxins/anti-auxins at 0.5 mg l⁻¹, applied jointly with 5.0 mg l⁻¹ KIN, exhibited variable effects on callus growth, ranging from inhibitory (TIBA, NAA) through neutral (IAA, CPA, IBA, IPA) to stimulatory (2,4,5-T, PCL, BNOA, 2,4-D). The combination of 0.5 BNOA and 5.0 KIN provided the highest growth rate (Gi = 502 %) of all PGR modifications (callus depicted in Fig. 2b). None of the auxins supplemented at 0.5 mg l⁻¹ had statistically significant stimulatory effect on S accumulation, however, IAA, BNOA and IPA significantly increased AS content. 2,4,5-T and IBA had significant negative effect on S production, whereas PCL and 2,4-D significantly decreased the accumulation of both alkaloids (Fig. 1a). The highest mean concentrations of S and AS (1.73 and 3.11 mg g⁻¹ DW) were recorded for the culture supplemented with 0.5 mg l⁻¹ IAA and 5.0 mg l⁻¹ KIN (Fig. 1a). Statistical analysis showed negative correlations between callus growth (Gi) and AS concentration (-0.27 , $p = 0.007$), as well as between Gi and the sum of alkaloids (-0.25 , $p = 0.013$).

The inclusion of 5.0 mg l⁻¹ auxin into the 0.5 mg l⁻¹ KIN-supplemented medium either did not affect (2,4,5-T, IAA, IBA, PCL, IPA, 2,4-D) or significantly stimulated callus growth (4-CPA, BNOA, NAA). The only exception was the anti-auxin TIBA which at 5.0 mg l⁻¹ significantly decreased the growth rate. As in the case of 0.5 mg l⁻¹ auxin/5.0 mg l⁻¹ KIN modifications, the highest Gi value (490 %) was recorded for BNOA (callus depicted in Fig. 2c). Of the PGRs tested, IAA, IBA, TIBA, BNOA and IPA had no effect on S accumulation whereas 2,4,5-T, 4-CPA, PCL, NAA and 2,4-D exerted significant inhibitory effect. Except for IAA and IPA, AS concentrations in the examined calli were significantly lowered in comparison to reference culture (C1). The combination of 5.0 mg l⁻¹ IAA and 0.5 mg l⁻¹ KIN provided the highest mean concentrations of S (1.42 mg g⁻¹ DW) and AS (2.42 mg g⁻¹ DW). The statistical analysis showed no important correlations between the growth rate (Gi) and indolizidine alkaloid accumulation.

As depicted in Fig. 1c, cytokinins at 0.5 mg l⁻¹, applied together with 5.0 mg l⁻¹ 2,4-D, had inhibitory (TDZ, CPPU), neutral (2iP, ZEA, KIN, BAP) or stimulatory (DPU, Gi = 427 %) effect on callus growth, as compared to C4 reference group (2,4-D only). All the cytokinins tested at 0.5 mg l⁻¹ significantly increased S

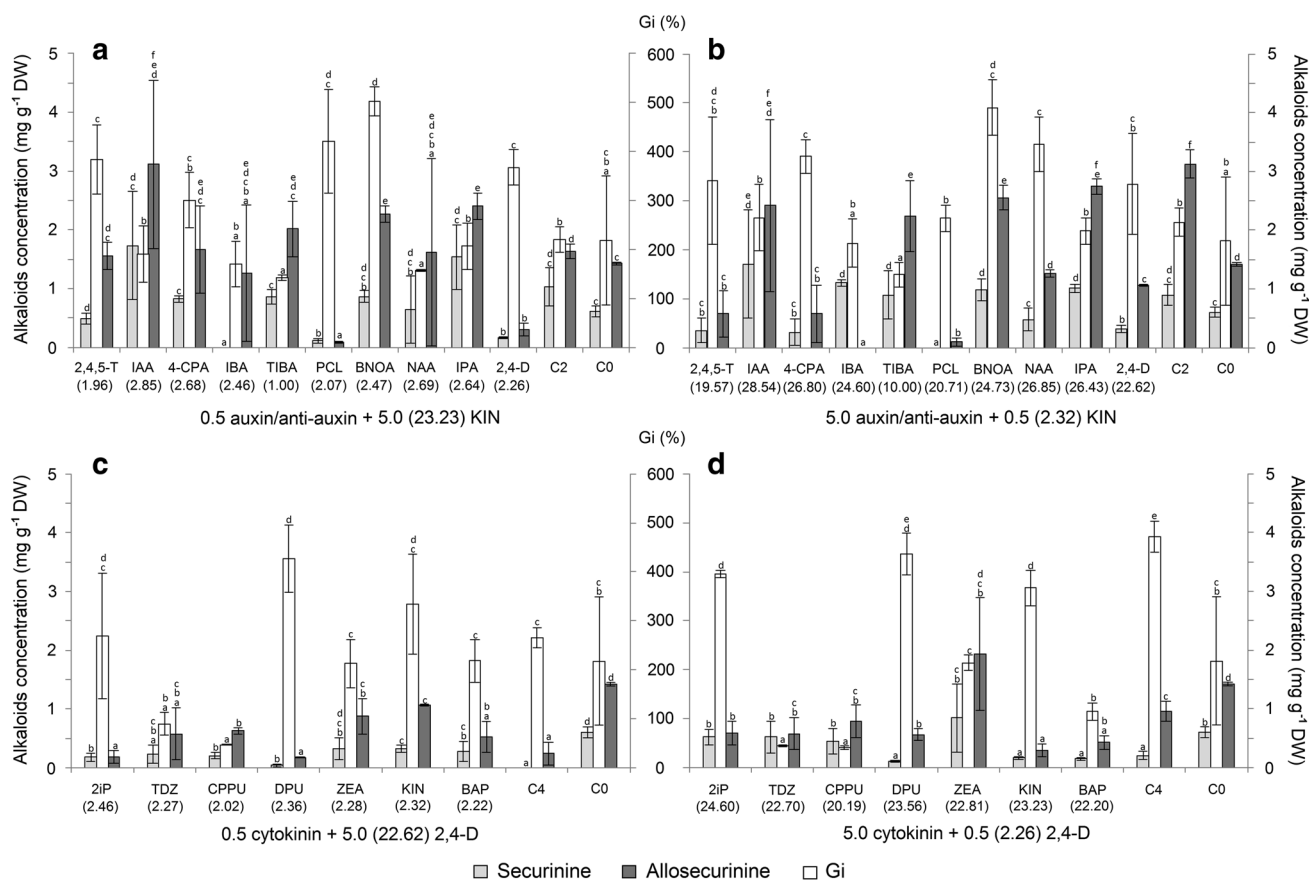


Fig. 1 The effect of different combinations of auxins/anti-auxins and cytokinins on the growth rate (Gi) and indolizidine alkaloid content of *S. suffruticosa* callus: **a** 0.5 mg l⁻¹ auxin/anti-auxin + 5.0 mg l⁻¹ KIN, **b** 5.0 mg l⁻¹ auxin/anti-auxin + 0.5 mg l⁻¹ KIN, **c** 0.5 mg l⁻¹ cytokinin + 5.0 mg l⁻¹ 2,4-D, **d** 5.0 mg l⁻¹ cytokinin + 0.5 mg l⁻¹

2,4-D (micromolar concentrations are given in *parentheses*). Control groups: C0 phytohormone-free medium, C1 0.5 mg l⁻¹ KIN, C2 5.0 mg l⁻¹ KIN, C3 0.5 mg l⁻¹ 2,4-D, C4 5.0 mg l⁻¹ 2,4-D. Different letters indicate significant differences between means ($p < 0.05$)

accumulation. The same effect was observed for AS, except for 2iP and DPU which had no significant effect on its concentration. The highest amounts of S and AS were recorded in the calli grown in the presence of 0.5 mg l⁻¹ ZEA (0.32 and 0.88 mg g⁻¹ DW, respectively) or 0.5 mg l⁻¹ KIN (0.32 and 1.06 mg g⁻¹ DW, respectively). At the same time, all phytohormone modifications tested provided lower indolizidine alkaloid concentrations compared to the PGR-free (C0) medium (Fig. 1c). No statistically significant correlations between callus growth and alkaloid concentrations were established.

When supplemented at 5.0 mg l⁻¹ and jointly with 0.5 mg l⁻¹ 2,4-D, the cytokinins had significant negative effect on callus growth. The inhibitory effect varied from ca. 20 % (2iP, KIN) to over 90 % decrease in Gi value recorded for TDZ and CPPU. DPU was the only PGR that did not significantly affect biomass growth (Gi = 436 %, comparable to C3 group). Of the cytokinins examined, DPU, KIN and BAP had no significant effect on S accumulation, whereas 2iP, TDZ and CPPU caused ca. twofold

increase in its concentration. AS accumulation was negatively influenced by 2iP, TDZ, DPU, KIN and BAP but remained statistically unchanged after TDZ and CPPU supplementation. The callus supplemented with 0.5 mg l⁻¹ ZEA and 5.0 mg l⁻¹ 2,4-D was characterized by the highest indolizidine alkaloid content (0.85 and 1.93 mg g⁻¹ DW of S and AS, respectively). The obtained results, however, were not significantly different than those of control (C0) group (Fig. 1d). Statistical analysis showed negative correlation (-0.31 , $p = 0.009$) between biomass growth (Gi) and S accumulation, however, no such relationship was established for AS and the sum of alkaloids.

As presented in Fig. 3, the combinations of IAA and ZEA (i.e. the PGRs of auxin and cytokinin groups which provided the highest indolizidine alkaloid accumulation—Fig. 1) had no significant effect on callus growth. The recorded concentrations of S (1.02–1.20 mg g⁻¹ DW) and AS (2.00–2.24 mg g⁻¹ DW) were significantly higher than those in the control group but also significantly lower in comparison to the combination of 0.5 mg l⁻¹ IAA and

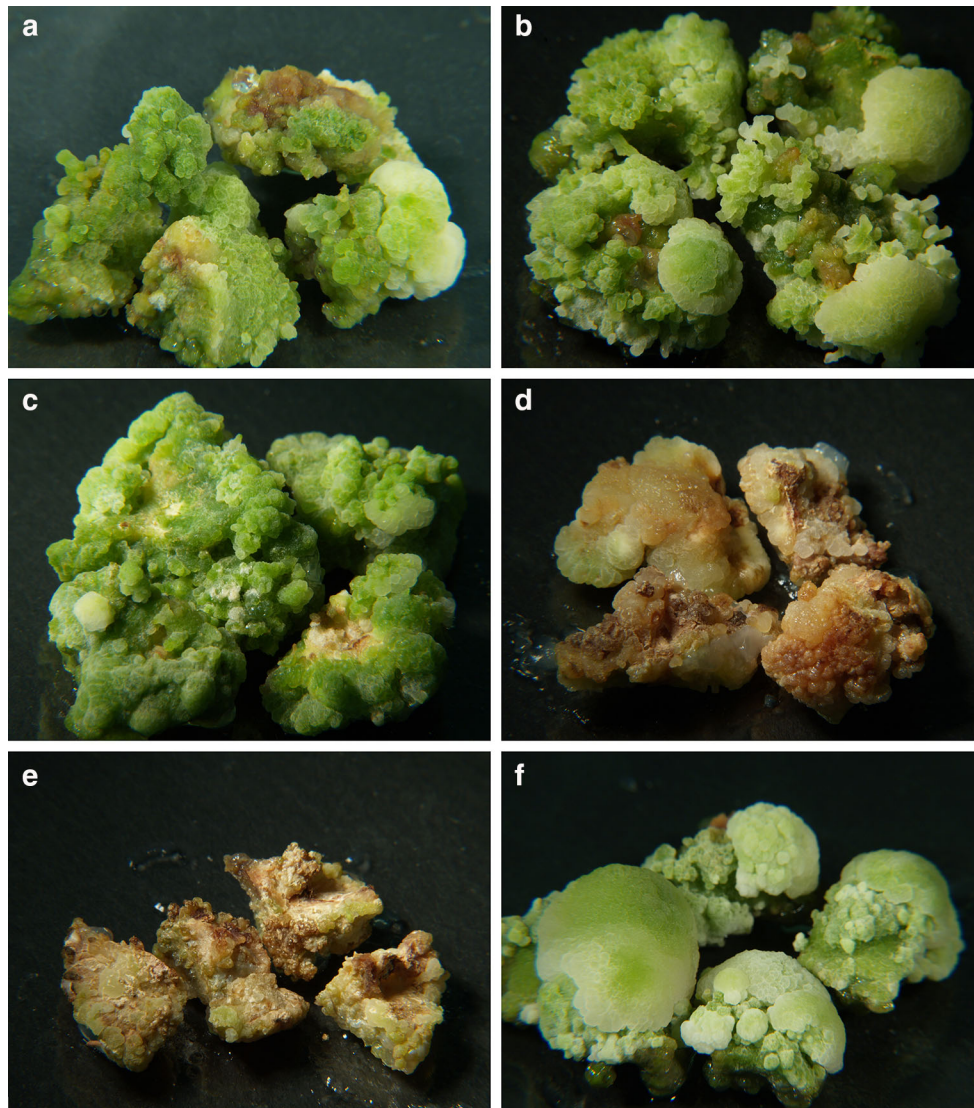


Fig. 2 *S. suffruticosa* calli grown using different PGR modifications: **a** PGR-free medium, **b** 0.5 mg l^{-1} BNOA + 5.0 mg l^{-1} KIN, **c** 5.0 mg l^{-1} BNOA + 0.5 mg l^{-1} KIN, **d** 0.5 mg l^{-1}

TDZ + 5.0 mg l^{-1} 2,4-D, **e** 5.0 mg l^{-1} TDZ + 0.5 mg l^{-1} 2,4-D, **f** 0.5 mg l^{-1} IAA + 5.0 mg l^{-1} KIN

5.0 mg l^{-1} KIN which consequently proved to be most effective PGR composition in the whole experiment (1.73 and 3.11 mg g^{-1} DW of S and AS, respectively).

Discussion

Modifications of phytohormone composition offer wide possibilities of improving cell culture productivity with respect to therapeutically relevant constituents (Murthy et al. 2014). However, difficulties can arise from often opposite effects of auxins and cytokinins towards secondary metabolism. Consequently, in order to provide high productivity, plant cell cultures need to be maintained in two-phase systems or supplemented with a balanced,

empirically optimized auxin/cytokinin composition (Verpoorte et al. 1991; Collin 2001; Luczkiewicz and Cisowski 2001; Ramawat and Mathur 2007).

In the presented work, 39 different PGR combinations were tested for their influence on the growth and indolizidine alkaloid content in *S. suffruticosa* callus cultures. Since the growth of *S. suffruticosa* callus was previously shown to be both auxin- and cytokinin dependent (Raj 2011), the experiment was conducted using auxin/cytokinin combinations. Given that the effects of phytohormones on primary and secondary metabolism are usually dose-dependent (Verpoorte et al. 1991; Ramawat and Mathur 2007), the media with opposite auxin/cytokinin ratios were tested—the experimental approach so far successfully applied in the studies concerning phytohormone-

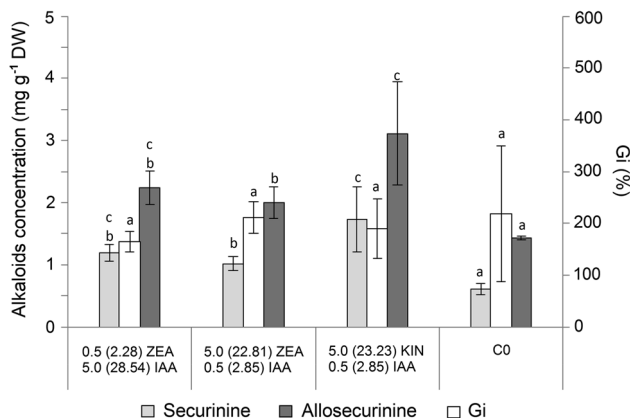


Fig. 3 The effect of the combinations of IAA with ZEA or KIN [concentrations given in mg l⁻¹ and μM (values in parentheses)] on the growth rate (Gi) and indolizidine alkaloid content of *S. suffruticosa* callus. C0: phytohormone-free medium. Different letters indicate significant differences between means ($p < 0.05$)

related accumulation of phenolics, including anthocyanins (Luczkiewicz and Cisowski 2001), isoflavones (Luczkiewicz et al. 2014) and phenolic acids (Szopa and Ekiert 2014), in plant cell cultures.

When applied alone, 2,4-D and KIN, respectively, had inhibitory and stimulatory effect on alkaloid accumulation which is in agreement with previous studies on cell cultures of alkaloid-bearing plants (Verpoorte et al. 1991; Ramawat and Mathur 2007). The experiments also demonstrated that simultaneous application of KIN and 2,4-D was not beneficial in terms of growth rates and alkaloid concentrations obtained. The influence of different combinations of the above PGRs on the growth and alkaloid content in *S. suffruticosa* callus was also investigated by other authors. Yuan et al. (2007) found that at the constant KIN concentration, the increase in 2,4-D content resulted in slower growth rate and lower alkaloid concentration. In another study, Ide (1991) tested several compositions of 2,4-D and KIN, the best of which yielded 0.06 mg g⁻¹ S (compared to 0.32 mg g⁻¹ S in the present work). Because of different PGR concentrations applied, the results of previous studies are not directly comparable with the current work. Nevertheless, the general conclusion is that a combination of 2,4-D and KIN can be used for both initiation and maintenance of *S. suffruticosa* cell cultures—while preserving their ability to accumulate alkaloids.

As indicated by previous studies, the replacement of 2,4-D with other auxins often resulted in significant increase of alkaloid accumulation in plant cell cultures (Verpoorte et al. 1991). In the present work, the combination of 0.5 mg l⁻¹ IAA and 5.0 mg l⁻¹ KIN provided the highest accumulation of S and AS among all PGR compositions tested. Positive effect on indolizidine alkaloid

accumulation was also noted for NAA, confirming the results by Wysokińska (1979), Ide (1991) and Yuan et al. (2007), as well as IPA whose influence on alkaloid metabolism has so far been scarcely investigated.

Contrary to the above PGRs, most auxins included in the study negatively influenced alkaloid accumulation in *S. suffruticosa*, with PCL exhibiting the strongest inhibitory effect. Also, no positive results were found for TIBA, an anti-auxin included in the experiments because of its stimulatory effect on secondary metabolism observed in previous work by the authors (Luczkiewicz et al. 2002, 2014).

When applied at 0.5 mg l⁻¹, cytokinins were shown to promote alkaloid accumulation in *S. suffruticosa*, thus reversing the negative effect of 2,4-D on secondary metabolism. Nevertheless, the recorded concentrations of S and AS were still lower than in C0 group (PGR-free medium). In general, media compositions involving purine-type cytokinins (especially ZEA and KIN) had more beneficial effects since they significantly increased alkaloid accumulation without negatively affecting callus growth. Phenylurea derivatives, on the other hand, either inhibited culture growth (TDZ and CPPU) or strongly increased biomass yield at the expense of alkaloid production (DPU).

An important observation arising from the study is that PGR combinations of 0.5 mg l⁻¹ auxin and 5.0 mg l⁻¹ cytokinin employed in the study showed negative correlation between callus growth and alkaloid accumulation, indicating that the growth medium could not be optimized for both high biomass yield and secondary metabolite content. It also needs to be mentioned that several PGRs included in the study (e.g. PCL, TIBA, TDZ, CPPU, CPU) were not previously examined with respect to their influence on indolizidine alkaloid biosynthesis. Therefore, it is difficult to assess whether the observed effects follow a general rule or are rather species-specific.

In the final part of the study, *S. suffruticosa* calli were grown on the media supplemented with the combinations of auxin and cytokinin which provided the highest indolizidine alkaloid content (IAA and ZEA, respectively) in order to check for synergistic effects between both PGRs. The experiment proved unsuccessful, yielding lower concentrations of S and AS compared to IAA/KIN composition. Nevertheless, the maximum securinine content obtained in the present work (1.73 mg g⁻¹ DW) was higher in comparison to the reports by Wysokińska (1979) and Ide (1991) (1.17 and 0.4 mg g⁻¹ DW, respectively).

Conclusions

Cell growth and securinine concentration in *S. suffruticosa* callus cultures proved to be strongly affected by the type and amount of the PGRs applied. Of the phytohormone

compositions tested, the highest indolizidine alkaloid content was achieved using the combination of 0.5 mg l⁻¹ IAA and 5.0 mg l⁻¹ KIN, providing higher securinine concentration in comparison to previous reports. However, negative correlation between callus growth and indolizidine alkaloid accumulation indicates that developing a two-phase production system may be necessary to further improve alkaloid productivity of *S. suffruticosa* cell cultures.

Compliance with Ethical Standards

Conflict of interest The authors declare that they have no conflict of interest and the presented work is compliant with ethical standards of Plant Cell, Tissue and Organ Culture. All the authors read and approved the manuscript in its final form.

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References

- Collin H (2001) Secondary product formation in plant tissue cultures. *Plant Growth Regul* 34:119–134. doi:[10.1023/A:1013374417961](https://doi.org/10.1023/A:1013374417961)
- Gupta K, Chakrabarti A, Rana S, Ramdeo R, Roth BL, Agarwal ML, Tse W, Agarwal MK, Wald DN (2011) Securinine, a myeloid differentiation agent with therapeutic potential for AML. *PLoS ONE* 6:e21203. doi:[10.1371/journal.pone.0021203](https://doi.org/10.1371/journal.pone.0021203)
- Holmes M, Crater AK, Dhudshia B, Thadani AN, Ananvoranich S (2011) *Toxoplasma gondii*: inhibitory activity and encystation effect of securinine and pyrrolidine derivatives on *Toxoplasma* growth. *Exp Parasitol* 127:370–375. doi:[10.1016/j.exppara.2010.09.002](https://doi.org/10.1016/j.exppara.2010.09.002)
- Ide A (1991) *Securinega suffruticosa*: In vitro culture and the formation of securinega alkaloids. In: Bajaj YPS (ed) *Biotechnology in agriculture and forestry*, vol 15: medicinal and aromatic plants III. Springer, Berlin, pp 420–431. doi:[10.1007/978-3-642-84071-5_25](https://doi.org/10.1007/978-3-642-84071-5_25)
- Ide A, Fujioka N, Nagano K, Tanaka N, Iwasaki K, Yamane Y, Koga D, Yagishita K, Nakao K, Kurisu Y, Fujioka N, Kohda H, Miyagawa H, Yamasaki K (1986) Growth and alkaloid production of callus culture induced from *Securinega suffruticosa*. *Phytochemistry* 26:145–148. doi:[10.1016/S0031-9422\(00\)81499-5](https://doi.org/10.1016/S0031-9422(00)81499-5)
- Li M, Han S, Zhang G, Wang Y, Ji Z (2014) Antiproliferative activity and apoptosis-inducing mechanism of L-securinine on human breast cancer MCF-7 cells. *Pharmazie* 69:217–223. doi:[10.1691/ph.2014.3802](https://doi.org/10.1691/ph.2014.3802)
- Luczkiewicz M, Cisowski W (2001) Optimisation of the second phase of a two phase growth system for anthocyanin accumulation in callus cultures of *Rudbeckia hirta*. *Plant Cell Tiss Organ Cult* 65:57–68. doi:[10.1023/A:1010652507981](https://doi.org/10.1023/A:1010652507981)
- Luczkiewicz M, Zárate R, Dembińska-Migas W, Migas P, Verpoorte R (2002) Production of pulchelin E in hairy roots, callus and suspension cultures of *Rudbeckia hirta* L. *Plant Sci* 163:91–100. doi:[10.1016/S0168-9452\(02\)00065-1](https://doi.org/10.1016/S0168-9452(02)00065-1)
- Luczkiewicz M, Kokotkiewicz A, Glod D (2014) Plant growth regulators affect biosynthesis and accumulation profile of isoflavone phytoestrogens in high-productive in vitro cultures of *Genista tinctoria*. *Plant Cell Tiss Organ Cult* 118:419–429. doi:[10.1007/s11240-014-0494-4](https://doi.org/10.1007/s11240-014-0494-4)
- Malik S, Cusidó RM, Mirjalili MH, Moyano E, Palazón J, Bonfill M (2011) Production of the anticancer drug taxol in *Taxus baccata* suspension cultures: a review. *Process Biochem* 46:23–34. doi:[10.1016/j.procbio.2010.09.004](https://doi.org/10.1016/j.procbio.2010.09.004)
- Murthy HN, Lee E-J, Paek K-Y (2014) Production of secondary metabolites from cell and organ cultures: strategies and approaches for biomass improvement and metabolite accumulation. *Plant Cell Tiss Organ Cult* 118:1–16. doi:[10.1007/s11240-014-0467-7](https://doi.org/10.1007/s11240-014-0467-7)
- Pietrosiuk A, Furmanowa M, Łata B (2007) *Catharanthus roseus*: micropropagation and in vitro techniques. *Phytochem Rev* 6:459–473. doi:[10.1007/s11101-006-9049-6](https://doi.org/10.1007/s11101-006-9049-6)
- Raj D (2011) *Badania biotechnologiczne nad akumulacją związków alkaloidowych w kulturach in vitro Securinega suffruticosa*, PhD Thesis, Wrocław Medical University
- Raj D, Luczkiewicz M (2008) *Securinega suffruticosa*. *Fitoterapia* 79:419–427. doi:[10.1016/j.fitote.2008.02.011](https://doi.org/10.1016/j.fitote.2008.02.011)
- Raj D, Kokotkiewicz A, Luczkiewicz M (2009) Densitometric HPTLC analysis of indolizidine alkaloids in the herb and in vitro cultures of *Securinega suffruticosa*. *J Planar Chromatogr Mod TLC* 22:371–376. doi:[10.1556/JPC.22.2009.5.11](https://doi.org/10.1556/JPC.22.2009.5.11)
- Raj D, Kokotkiewicz A, Luczkiewicz M (2015) Production of therapeutically relevant indolizidine alkaloids in *Securinega suffruticosa* in vitro shoots maintained in liquid culture systems. *Appl Biochem Biotechnol* 175:1576–1587. doi:[10.1007/s12010-014-1386-0](https://doi.org/10.1007/s12010-014-1386-0)
- Ramawat KG, Mathur M (2007) Factors affecting the production of secondary metabolites. In: Ramawat KG, Merillon JM (eds) *Biotechnology: secondary metabolites*. CRC Press, Boca Raton, pp 59–102. doi:[10.1201/b10756-4](https://doi.org/10.1201/b10756-4)
- Rana S, Gupta K, Gomez J, Matsuyama S, Chakrabarti A, Agarwal M, Agarwal A, Agarwal M, Wald D (2010) Securinine induces p73-dependent apoptosis preferentially in p53-deficient colon cancer cells. *FASEB J* 24:2126–2134. doi:[10.1096/fj.09-148999](https://doi.org/10.1096/fj.09-148999)
- Sabater-Jara AB, Tudela LR, López-Pérez AJ (2010) In vitro culture of *Taxus* sp.: strategies to increase cell growth and taxoid production. *Phytochem Rev* 9:343–356. doi:[10.1007/s11101-010-9167-z](https://doi.org/10.1007/s11101-010-9167-z)
- Schenk RU, Hildebrandt AC (1972) Medium and techniques for induction and growth of monocotyledonous and dicotyledonous plant cell cultures. *Can J Bot* 50:199–204. doi:[10.1139/b72-026](https://doi.org/10.1139/b72-026)
- Szopa A, Ekiert H (2014) Production of biologically active phenolic acids in *Aronia melanocarpa* (Michx.) Elliott in vitro cultures cultivated on different variants of the Murashige and Skoog medium. *Plant Growth Regul* 72:51–58. doi:[10.1007/s10725-013-9835-2](https://doi.org/10.1007/s10725-013-9835-2)
- van der Heijden R, Jacobs DI, Snoeijer W, Hallard D, Verpoorte R (2004) The *Catharanthus* alkaloids: pharmacognosy and biotechnology. *Curr Med Chem* 11:607–628. doi:[10.2174/0929867043455846](https://doi.org/10.2174/0929867043455846)
- Verpoorte R, van der Heijden R, van Gulik WM, ten Hoopen HJG (1991) Plant biotechnology for the production of alkaloids: present status and prospects. In: Brossi A (ed) *The alkaloids: chemistry and pharmacology*, vol 40. Academic Press, San Diego, pp 1–187. doi:[10.1016/S0099-9598\(08\)60268-0](https://doi.org/10.1016/S0099-9598(08)60268-0)
- Wysokińska H (1979) Wpływ niektórych regulatorów wzrostu i prekursorów biosyntezy alkaloidów na kulturę tkankową i komórkową *Securinega suffruticosa* (Pall.) Rehd., PhD Thesis, Medical University of Lodz
- Yuan W, Lu Z, Liu Y, Meng C, Cheng KD, Zhu P (2005) Three new podocarpane-type diterpenoids from callus of *Securinega suffruticosa*. *Chem Pharm Bull* 53:1610–1612. doi:[10.1248/cpb.53.1610](https://doi.org/10.1248/cpb.53.1610)
- Yuan W, Zhu P, Cheng K, Meng C, Wu F, Zhu H (2007) Callus of *Securinega suffruticosa*, a cell line accumulates dextro Securinega alkaloids. *Nat Prod Res* 21:234–242. doi:[10.1080/14786410701189781](https://doi.org/10.1080/14786410701189781)