

Effect of *Scutellariae Radix* extract on experimental dextran-sulfate sodium-induced colitis in rats

Ho-Lam Chung, Grace Gar-Lee Yue, Ka-Fai To, Ya-Lun Su, Yu Huang, Wing-Hung Ko

Ho-Lam Chung, Department of Physiology, Faculty of Medicine, The Chinese University of Hong Kong, Shatin, Hong Kong, China
Grace Gar-Lee Yue, Institute of Chinese Medicine, The Chinese University of Hong Kong, Shatin, Hong Kong, China

Ka-Fai To, Department of Anatomical and Cellular Pathology, Faculty of Medicine, The Chinese University of Hong Kong, Shatin, Hong Kong, China

Ya-Lun Su, Institute of Materia Medica, Chinese Academy of Medical Sciences and Peking Union Medical College, Beijing 100050, China

Yu Huang, Wing-Hung Ko, Department of Physiology and Li Ka Shing Institute of Health Sciences, Faculty of Medicine, The Chinese University of Hong Kong, Shatin, Hong Kong, China
Supported by a direct grant for research from The Chinese University of Hong Kong, No. 2041075 awarded to Wing-Hung Ko

Correspondence to: Wing-Hung Ko, Department of Physiology and Li Ka Shing Institute of Health Sciences, Faculty of Medicine, The Chinese University of Hong Kong, Shatin, Hong Kong, China. whko@cuhk.edu.hk

Telephone: +852-26096781 Fax: +852-26035022

Received: May 3, 2007 Revised: July 4, 2007

to forskolin were suppressed after the induction of colitis. The stimulated ion transport activity of DSS-rats treated with SRE displayed significant improvement in the secretory responsiveness.

CONCLUSION: SRE was effective in treating acute DSS-induced ulcerative colitis, as gauged by reduced clinical disease, improved macroscopic and histological damage scores, and enhanced recovery of normal colonic secretory function.

© 2007 WJG. All rights reserved.

Key words: Ulcerative colitis; *Scutellariae Radix*; Inflammatory bowel disease; Colonic ion transport; Traditional Chinese medicine

Chung HL, Yue GGL, To KF, Su YL, Huang Y, Ko WH. Effect of *Scutellariae Radix* extract on experimental dextran-sulfate sodium-induced colitis in rats. *World J Gastroenterol* 2007; 13(42): 5605-5611

<http://www.wjgnet.com/1007-9327/13/5605.asp>

Abstract

AIM: To investigate the effect of *Scutellariae Radix* extract (SRE) on ulcerative colitis (UC) in rats induced by dextran-sulfate sodium (DSS).

METHODS: Colitis was induced in male Sprague-Dawley (SD) rats (170-180 g) by 4% dextran sulfate sodium (DSS, wt/v; MW 54000) in drinking water for 8 d. The treated rats received 4% DSS and SRE orally (100 mg/kg per day). Control rats received either tap water or SRE only. Macroscopic assessment which included body weight changes, fecal occult blood and stool consistency were determined daily. At the appointed time, the rats were sacrificed and the entire colons were removed. The colon length and the myeloperoxidase (MPO) activity were measured. The severity of colitis was graded by morphological and histological assessments. The ion transport activity of the colonic mucosa was assessed by electrophysiological technique.

RESULTS: Rats treated with oral administration of 4% DSS regularly developed clinical and macroscopic signs of colitis. Treatment with SRE relieved the symptoms, including the reduction in body weight, shortening and ulceration of the colon. Administration of SRE also significantly reduced the histological damage induced by DSS. Moreover, the *Isc* responses of the colonic mucosa

INTRODUCTION

Ulcerative colitis is a worldwide, chronic, idiopathic, inflammatory bowel disease (IBD) of the rectal and colonic mucosa. In the past, this disease was thought to occur infrequently in the Asia Pacific region. However, new evidence is showing that IBD is on the rise in the region, including in Hong Kong and mainland China^[1-3]. Although glucocorticoid and salicylazosulfapyridine have been mainly used for the treatment of this disease, their side effects remain a major clinical problem. Therefore, there is an increasing interest in using traditional Chinese medicines (TCM) as alternative therapy in addition to the conventional therapies that are used to treat UC^[4].

The dried root of *Scutellaria baicalensis* Georgi (*Scutellaria Radix*, common name Huangqin) is widely used in TCM. It is officially listed in the Chinese Pharmacopoeia and is one of the most widely used Chinese herbal medicines for the treatment of bacterial infection of the respiratory and gastrointestinal tract. In Japan and China, *Scutellariae Radix* has been employed for centuries as an important medicine to treat chronic inflammatory and ulcerative disease. The main components of *Scutellariae Radix* (and of all *Scutellaria* species) are baicalein, baicalin and wogonin.

Scutellariae Radix and its major flavonoids possess multiple biological and pharmacological effects, including anti-inflammation^[5], anti-viral^[6], anti-tumor^[7], anti-proliferative^[8] and anti-bacterial^[9], *etc.* Recent studies also suggest that *Scutellariae Radix*-containing TCM formula, such as Oren-gedoku-to (Huang Lian Jie Du Tang) may have therapeutic potential against murine colitis^[10-12].

In this study, an experimental model of UC was established in SD rats using DSS. The effect of the total extract of *Scutellariae Radix* on DSS-rats was evaluated using macroscopic, histological, biochemical and electrophysiological assessments.

MATERIALS AND METHODS

Materials

Male SD rats, initially weighing 170-180 g, were housed five per cage and maintained in an animal holding room controlled at a constant temperature of $24^{\circ}\text{C} \pm 2^{\circ}\text{C}$ with a relative humidity of $70\% \pm 5\%$ and a 12 h light-dark cycle. Animals were fed on a standard pellet chow with free access to fresh tap water. The study was approved by the Animal Research Ethics Committee of our university. DSS was obtained from MP Biochemicals Inc. Hexadecyltrimethylammonium bromide, forskolin were obtained from Sigma. SRE was purified from the ground roots of *S. baicalensis* Georgi with hexane, acetone, and finally, with methanol as described previously^[13,14].

Experimental design

The induction of colitis was modified from a previously described work^[15]. The experiment lasted for 8 d. The rats were randomly divided into four groups. In the DSS Group, 4% DSS in drinking water *ad libitum* was given from d 0 to d 7. In DSS + SRE group, SRE (100 mg/kg per day) was orally administrated with exposure to 4% DSS drinking water. In the normal control group (Ctr), the rats had free access to a water bottle containing tap water. In Ctr + SRE group, SRE alone (100 mg/kg per day) was administered orally to the rats. In this model, the colonic damage was evaluated using macroscopic, histological, electrophysiological and biochemical assessments (see below). From d 3 to d 7, rats were sacrificed by CO_2 asphyxiation on each day. Postmortem, the entire colon was removed from the cecum to the anus and placed on an ice cold plate and cleaned of fat and mesentery. The length of each specimen was measured, which is an indirect marker of inflammation. The colon was divided into three parts (proximal, middle and distal) based on total length: 10% regions from three parts were fixed for histological examination; the distal portion was used for electrophysiological studies; and the adjacent distal 10% was snap-frozen in liquid nitrogen for later quantification of MPO activity.

Macroscopic assessment

Animals were checked daily for the three main clinical symptoms-body weight changes, stool consistency and fecal occult blood. Weight loss is usually observed in animals with colitis, thus body weight changes recorded

could be an indicator for the severity of colitis. The colonic damage was quantified by a clinical scoring system assessing stool consistency and rectal bleeding^[16]. For stool consistency, 0 points were given for well formed pellets, 2 points for pasty and semiformed stools that did not stick to the anus, and 4 points for liquid stools that did stick to the anus. Bleeding was scored 0 for no blood in hemocult, 2 points for positive hemocult, and 4 points for gross bleeding. These scores were added, forming a total clinical score that ranged from 0.0 (healthy) to 8.0 (maximal activity of colitis).

Histological assessment

For light microscopy, we used tissue samples from three parts (distal, middle and proximal) of colon of each animal fixed in 4% (40 g/L) buffered paraformaldehyde, dehydrated in increasing concentrations of ethanol, and embedded in paraffin. Thereafter, sections of tissue were cut at 5 μm on a rotary microtome, mounted on clean glass slides and dried overnight at 37°C . The sections were cleaned, hydrated, and stained with hematoxylin and eosin (HE) for histological evaluation of inflammatory infiltrate and tissue damage, according to standard protocols, and the slides were coded to prevent observer bias during evaluation. All tissue sections were examined in a blinded fashion by two investigators. Histological damage was scored using the criteria of Siegmund B *et al*^[17] which considers the inflammatory infiltrate (maximum score = 3) and tissue damage (maximum score = 3). Photographs of colon samples were digitized using a ZEISS Axioskop 2 plus camera. Analysis of the figures was carried out with Axio Vision 3.1 image analysis program.

Electrophysiological assessment

Mucosal ion transport was examined in colonic segments (approximately distal colon) mounted in Ussing chambers according to a well established protocol in our laboratory^[18]. In brief, tissues (surface area = 0.45 cm^2) were bathed in 20 mL of warm (37°C), oxygenated Krebs buffer. The spontaneous potential differences were maintained at 0 mV by a voltage clamp amplifier (Physiologic Instruments), and the short-circuit current (I_{SC} in μAcm^{-2}) was continuously measured as an index of electrogenic ion transport. A transepithelial potential difference of 1 mV was applied periodically, and the resultant change in current was used to calculate the transepithelial resistance (R_t) using Ohm's law. Stimulated ion transport was evoked by the addition of an adenylate cyclase-activating agent, forskolin (1 $\mu\text{mol/L}$), to the mucosal bathing solution. In all instances, the effect of the treatment was recorded as the maximum change in I_{SC} (ΔI_{SC}) to occur within 5 min.

Biochemical assessment

The MPO activity was determined following a published protocol^[19]. Briefly, frozen tissue samples were weighed and suspended in potassium phosphate buffer (20 mmol/L, pH 7.4) at a ratio of 50 mg tissue to 1 mL of buffer. Tissue was homogenized by a polytron tissue homogenizer three times for 20 s, and homogenate was decanted into sterile

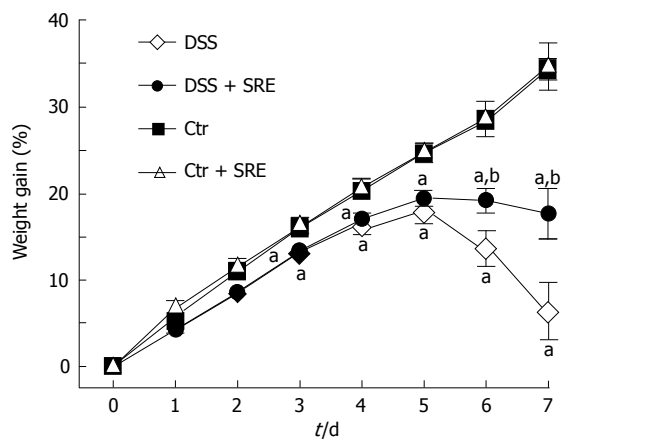


Figure 1 Figure showing the weight gain percentage (%) in different groups of rat from d 0 to d 7. Body weight was assessed daily and expressed as percentage increase of basal body weight. Values are expressed as the means \pm SE, ($n = 8$), ^a $P < 0.05$ vs Ctr group, ^b $P < 0.05$ vs DSS group, one-way ANOVA followed by Tukey multiple comparison test.

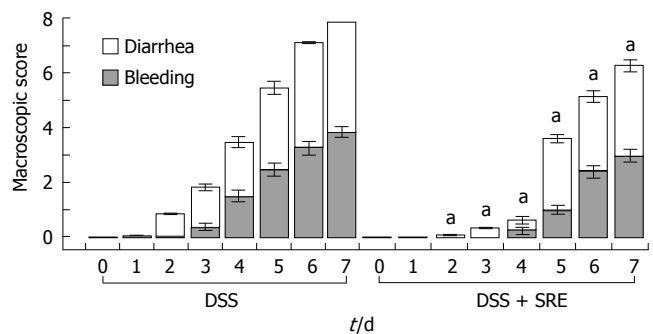


Figure 2 Ameliorative effect of SRE treatment on the time-course changes in the macroscopic score over the 8-d experimental period. The macroscopic score of Ctr and Ctr + SRE groups is 0 (data not shown). ^a $P < 0.05$ vs DSS group. Non-parametric data are expressed as the means \pm SE, Kruskal-Wallis One Way Analysis using Student-Newman-Keuls method ($n = 6-8$).

Eppendorf tubes and centrifuged at $10000 \times g$ for 10 min at 4°C . The pellet was then resuspended in 1 mL potassium phosphate buffer (50 mmol/L, pH 6.0) containing 5 mg/mL hexadecyltrimethylammonium bromide (TMB) at a tissue concentration of 50 mg/mL. Samples were sonicated three times for 10 s, freeze-thawed three times, and centrifuged at $10000 \times g$ for 5 min at 4°C . The reaction was started by mixing 20 μL of the supernatant at 25°C with 30 μL TMB. After 110 s, the reaction was terminated by addition of 50 μL of 0.18 mol/L H_2SO_4 . The change in absorbance was read at 450 nm. MPO activity (1 unit) was expressed as the amount of enzyme necessary for the degradation of 1 μmol H_2O_2 /min per 100 mg tissue at 25°C .

Statistical analysis

All data are presented as means \pm SE. One way ANOVA followed by post hoc statistics with Tukey test was used for statistical evaluation of the parametric data. Non-parametric data was analyzed by Kruskal-Wallis One Way Analysis using Student-Newman-Keuls method. $P < 0.05$ was considered as statistically significant. The values of n refer to the number of experiments undertaken using



Figure 3 Macroscopic view of the colon showing the changes in colon length in different groups of rat on d 7. Not much difference was observed between the colon of the Ctr (A) and Ctr + SRE group (B), but significant difference could be found in the DSS group (C) which displayed extensive hyperemia and edema. In the DSS + SRE group, the shortening of colon was less severe (D) when compared with the control (A).

different rats. Non-parametric data are expressed as the means \pm SE.

RESULTS

Macroscopic assessment

The weight gain % over the entire study period is shown in Figure 1. The weight gain % of rats in DSS group and DSS + SRE group were significantly lower than the Ctr group and Ctr + SRE group from d 3 until the end of experiment. The body weight of rats in DSS group then dramatically decreased from d 5 onwards. However, in the case of DSS + SRE group, the weight gain % became stabilized and the weight loss was found to be less severe than DSS group from d 6 to d 7. Figure 2 shows the macroscopic score recorded throughout the experimental period in DSS and DSS + SRE groups. Oral administration of SRE resulted in a significant reduction in the clinical activity of colitis compared with DSS-treated rats. Moreover, the occurrences of those clinical signs were found to be delayed in DSS + SRE group.

The colon length is a useful indication of colitis and is, therefore, measured as a marker of inflammation (Figure 3). After 8 d treatment with DSS in drinking water, there was a significant shortening of the colon length (Figure 3C) compared with the Ctr group (Figure 3A) and the Ctr + SRE group (Figure 3B). The oral administration of SRE significantly improved this inflammatory marker (Figure 3D). On d 7, the colon length of DSS group ($11.2 \text{ cm} \pm 0.4 \text{ cm}$, $n = 10$) was significantly shorter than the control group ($16.8 \text{ cm} \pm 0.6 \text{ cm}$, $n = 8$). The colon length of DSS-rats treated with SRE ($13.5 \text{ cm} \pm 0.4 \text{ cm}$, $n = 9$), however, was significantly longer than that of untreated rats.

Histological assessment

Histological damage was evaluated by the grading method described above in Materials and Methods. The occurrence of UC was confirmed on the basis of histological damage and inflammatory infiltrate as shown in Figure 4. Figure 5 summarized the damage scores from DSS rats and DSS rats treated with SRE. The microscopic score of samples

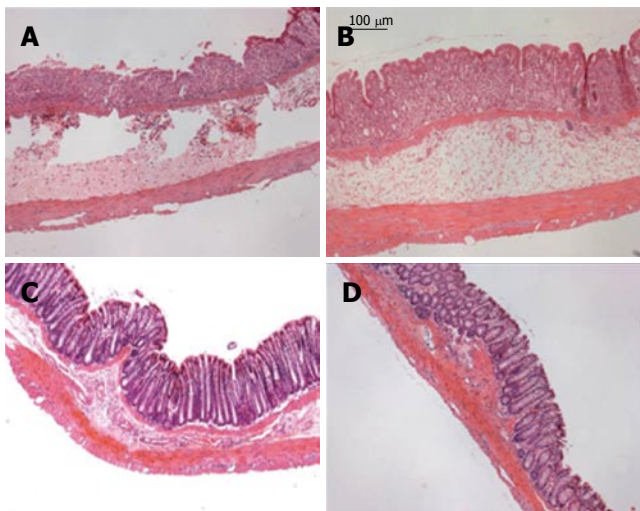


Figure 4 Histological sections from different groups of rats on d 7. **A:** DSS group showing extensive ulceration with a severe inflammatory cell infiltrate; **B:** DSS + SRE group showing recovery in the inflammatory cell infiltration with less severe ulceration; **C:** Non-colitic Ctr group showing the normal histology of the colon; **D:** Ctr + SRE group showing the normal morphology of the colon. (HE staining; original magnifications, x 100).

from different regions (distal, middle and proximal) of colon were not significantly different from each other (data not shown). The histological index began to increase on d 3 in both groups. However, the rats with SRE administration showed a significantly lower value than the untreated animals on d 6 and d 7.

Electrophysiological assessment

Electrogenic ion transport function was assessed by stimulating the colon with an adenylate cyclase activator, forskolin (1 $\mu\text{mol/L}$), using short-circuit current measurement technique (Figure 6). Under the experimental conditions, the increase in I_{SC} is mainly due to the cAMP-mediated Cl^- secretion *via* the cystic fibrosis transmembrane conductance regulator (CFTR)^[18]. The basal I_{SC} and R_t in the tissues were recorded and these parameters were not significantly different (data not shown) when tissues from different groups were compared. On d 7, the secretory response to the cAMP-elevating agent was significantly diminished in tissues from rats treated with DSS ($\Delta I_{SC} = 4.94 \pm 1.65 \mu\text{Acm}^{-2}$, $n = 6$) when compared with Ctr ($\Delta I_{SC} = 60.97 \pm 7.62 \mu\text{Acm}^{-2}$, $n = 8$) and Ctr + SRE ($\Delta I_{SC} = 61.48 \pm 6.78 \mu\text{Acm}^{-2}$, $n = 7$). On the other hand, the reduced responsiveness was ameliorated in the colitic rats with SRE administration ($\Delta I_{SC} = 20.67 \pm 3.30 \mu\text{Acm}^{-2}$, $n = 9$).

Biochemical assessment

The MPO activity in the groups without DSS treatment remained at a low value throughout the entire experiment and there was no effect of SRE on the control (d 7: Ctr $0.09 \pm 0.01 \text{ mU/mg}$, $n = 6$; Ctr + SRE $0.06 \pm 0.0003 \text{ mU/mg}$, $n = 5$). In comparison, rats with colitis were accompanied by a significant increase in MPO activity (d 7: $0.52 \pm 0.07 \text{ mU/mg}$, $n = 8$). On d 7, the increase in MPO activity ($0.34 \pm 0.05 \text{ mU/mg}$, $n = 8$) was significantly reduced in rats treated with SRE.

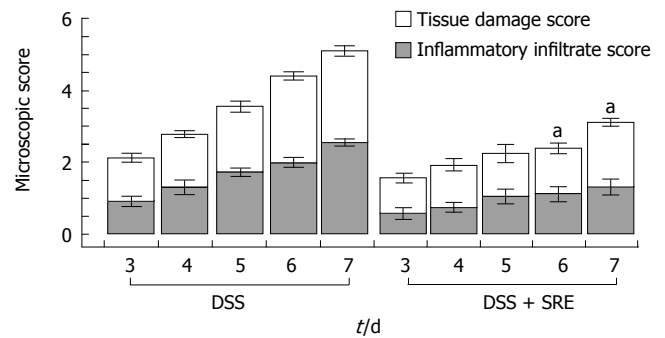


Figure 5 Ameliorative effect of SRE treatment on the time-course changes in the microscopic score from d 3 to d 7. The microscopic score of Ctr and Ctr + SRE groups is 0 (data not shown). ^a $P < 0.05$ vs DSS group. Non-parametric data are expressed as the means \pm SE, Kruskal-Wallis One Way Analysis using Student-Newman-Keuls method ($n = 4-5$).

DISCUSSION

Ulcerative colitis is an inflammatory disease that causes ulcerations of the mucosa in the colon. The incidence of UC is around 1 in 1000 people with a higher prevalence among Caucasians^[20]. In the past, this disease has been considered to occur rarely in the Asia Pacific region, but recent evidence indicates that both UC and Crohn's disease (CD) are becoming increasingly prevalent in local populations^[3]. For example, from 1991 to 2000, there has been a three-fold increase in the number of cases of UC in China^[1]. In Hong Kong, from 1986-1989 to 1999-2001, there was also a three-fold increase in the incidence of CD in the Chinese population^[2]. Although the etiology of IBD remains essentially unknown^[21], results from many studies in human patients and animal models suggest that it may be related to an abnormal immune response in the gastrointestinal tract, possibly associated with genetic and environmental factors^[22,23]. Although progress has been made in the overall management of UC, the pharmacological treatments that are available are still unsatisfactory. Glucocorticoids, sulfasalazine and immunosuppressive drugs have been mainly used for the treatment and maintenance of UC, but the side effects or toxicity of these drugs remain a major clinical problem^[24]. As a result, there is an increasing interest in using TCM as alternative therapy in addition to the conventional therapies that are used to treat UC^[25]. In Japan and China, the most commonly used alternative remedies are herbal and these have been widely used in patients with mild to moderate disease, as well as an adjunct to therapy in patients with moderate to severe disease^[1]. Interestingly, a recent survey showed that one-third of both Chinese and Caucasian IBD patients had used complementary and alternative medicines and therapies^[4]. Several TCM formulae have been shown to possess an anti-colitic effect in rats^[26-28]. Recent studies suggest that *Scutellariae Radix* may have a pharmacological effect against murine colitis^[10-12] and therefore in this study we aim to further evaluate the therapeutic effect of SRE on DSS-induced rat colitis.

In the present study, 4% DSS in drinking water was administered for 8 d to induce acute colitis in rats. All DSS-treated rats showed numerous clinical symptoms such as body weight loss, diarrhea, bloody stools and shortening

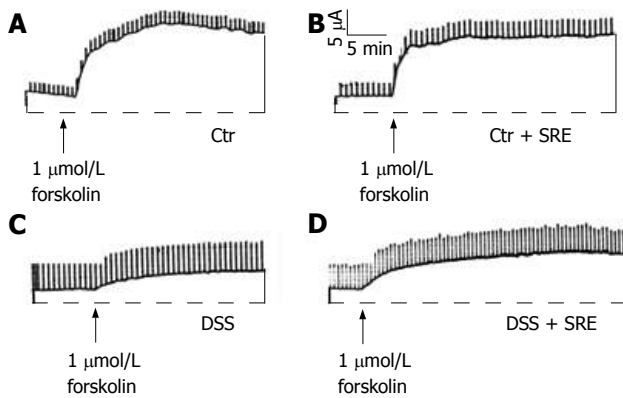


Figure 6 Representative tracings showing the change in I_{sc} evoked by forskolin in different groups of rat on d 7. All four groups showed an increase in ISC but with different magnitudes (A-D). The I_{sc} response was greatly reduced in the DSS group (C) when compared with the control (A). On the other hand, the reduction in secretory response appears to be ameliorated in the DSS + SRE group (D). The transient current pulses were the results of intermittently clamping the potential at 1 mV. The horizontal lines represent zero I_{sc} . The record is representative of at least six experiments.

of colon length. Body weight loss is a common symptom of UC because of the loss of body fluid and damage to the digestive system. In Ctr and Ctr + SRE groups, rats showed a steady increase in body weight throughout the experiment. In comparison, rats exposed to DSS had a lower rate of body weight increase followed by a dramatic decrease from d 5 onwards. Treatment with SRE to the colitic rats showed a less severe weight loss from d 6 to d 7. In DSS group, diarrhea and rectal bleeding occurred in d 1 and d 2, respectively. The SRE administration delayed the occurrence of both symptoms and with less severity. Colon shortening is always found in UC patients, which can act as an indirect marker of colonic inflammation. Although the colitic rats with SRE treatment also showed a decrease in colon length on d 7 when compared with the control, the shortening was much less severe than that of DSS rats. Taken together, the data suggested that SRE treatment could induce a decrease in the extent of colitis accompanied by reducing the severity and delaying the occurrence of the associated clinical symptoms.

Colitis induced by DSS was histologically characterized by severe disruption of tissue architecture, edema, a massive mixed immune cell infiltrate, ulceration and muscle thickening. To quantify the histological damage, we used a scoring system modified from a previous study^[17]. We found that the overall histological damage of colitis was significantly reduced by the SRE treatment on d 6 and d 7 of the experiment. Together with the reduced MPO activity, our results showed that SRE treatment ameliorated the DSS-induced colitis possibly *via* its anti-inflammatory and protective effect against the colonic tissue damage.

The colonic epithelium, in addition to its absorptive and secretory properties, presents an efficient barrier to commensal flora and pathogens^[29,30]. In fact, the secretion of water and electrolytes is one of the most important responses of the mucosa, purging the gut of offending agents and delivering anti-microbial mediators (e.g., antibodies) to the luminal surface^[31]. In addition, mucus forms a gel layer covering the mucosal surface, and it has been hypothesized that changes in mucin structure and/or

quantity may influence the protective functions of the mucosal surface, and affect the pathogenesis of IBD^[32]. However, published data on the electrolyte transport mechanisms in an inflamed colon are inconsistent^[33,34]. Some studies have documented the acute effects of immune and inflammatory agents in directly or indirectly stimulating intestinal anion secretion, while others do not support such a notion^[35,36]. In most cases, however, disease models of colonic mucosa exhibit reduced ion transport responses to secretory agonists, especially in the setting of chronic inflammation^[37-45].

The mechanism underlying the typical hyporesponsiveness of tissues from animal models of colitis has yet to be satisfactorily explained. For example, it has been reported that responsiveness to both Ca^{2+} - and cAMP-dependent secretagogues is reduced in mouse and rat colitis models when compared to normal tissue^[37-45]. Similar reduction in secretory responsiveness, as measured by changes in I_{sc} has also been observed in tissue resections from patients with IBD^[46,47]. It has been proposed that prolonged hyporesponsiveness to secretagogues is due to the upregulation of inducible nitric oxide synthase (iNOS), resulting in an ongoing synthesis of NO and chronic suppression of epithelial secretory function^[41,42]. However, another recent study suggests that it may be related to a disturbance of the enteric nervous system resulting in defective mucosal cAMP production and inhibition of ionic secretion, although the epithelial secretory machinery (e.g., CFTR) appears to be normal^[43]. Others suggested that prolonged hyporesponsiveness to secretagogues is due to the disruption of normal cholinergic control of ion secretion^[40,48]. Nonetheless, intestinal secretion is an important component of mucosal defense. Reduced secretory responses will compromise the ability of the mucosal defense mechanism to clear bacteria, bacterial products, or antigens away from the epithelium, which may then predispose the colon to inflammation^[49]. In this study, the I_{sc} responses of the colonic mucosa to forskolin were suppressed after the induction of colitis. Although the stimulated ion transport activity of DSS-rats treated with SRE was still reduced, they displayed improvement in the secretory responsiveness which may at least partly contribute to the therapeutic effect of SRE.

In summary, our findings indicated that SRE was effective in treating acute DSS-induced ulcerative colitis, as gauged by reduced clinical disease, improved macroscopic and histological damage scores, and enhanced recovery of normal colonic secretory function. The results support further evaluation of the therapeutic potential of SRE for the treatment of IBD.

ACKNOWLEDGMENTS

We thank Professor Woody WY Chan and Professor CH Cho for their technical advice in the performance of histological staining and MPO assay, respectively. We also thank Mr. Wallace CY Yip for his excellent technical help.

COMMENTS

Background

Scutellariae Radix, also known as Huangqin, is the dried root of *Scutellaria*

baicalensis Georgi (Lamiaceae). It is officially listed in the Chinese Pharmacopoeia and is one of the most widely used Chinese herbal medicines for the treatment of bacterial infection of the respiratory and gastrointestinal tract. In Japan and China, *Scutellariae Radix* has been employed for centuries as an important medicine. Although scientific evidence confirming the traditional use of *Scutellariae Radix* as an inflammatory modulator in experimental colitis is now accumulating, the physiological basis and the precise mechanism of action of *Scutellariae Radix* or its individual constituents remain largely unknown.

Research frontiers

Cytokine dysregulation is currently an important focus of both basic and clinical research in IBD, with recent immunologically based therapeutic interventions using highly specific agents demonstrating a promising clinical efficacy. The treatment of steroid-refractory Crohn's Disease with anti-TNF- α (infliximab) is an example of this kind of therapeutic approach. In addition, there is an increasing interest in using TCM as alternative therapy in addition to the conventional therapies that are used to treat IBD. However, there is still a paucity of scientific and clinical data so far to support the use of TCM in colitis patients. Therefore, there are unmet needs for further mechanistic, pharmacological and pharmacokinetic studies to delineate the biological basis underlying the therapeutic effects of TCM. More controlled clinical trials of the potential efficacy and safety of herbal TCM therapies are also required.

Innovations and breakthroughs

The authors showed for the first time that the therapeutic potential of *Scutellariae Radix* extract on experimental colitis may be related to the restoration of ion transport function of the colonic mucosa.

Applications

The results support further evaluation of the therapeutic potential of this herbal extract and its active component(s) for the treatment of IBD.

Terminology

Colonic ion transport: Intestinal fluid secretion is a passive process driven by osmotic forces generated by ion transport. In the colon, the main determinant of a lumenally-directed osmotic gradient is the mucosal transport of chloride ions into the lumen. Intestinal secretion is an important component of mucosal defense. Reduced secretory responses will compromise the ability of the mucosal defense mechanism to clear bacteria, bacterial products, or antigens away from the epithelium, which may then predispose the colon to inflammation.

Peer review

This is an interesting paper which shows the anti-inflammatory effect of *Scutellariae Radix* on experimental colitis. It also demonstrates that TCM has a scientific and biological basis for its effectiveness.

REFERENCES

- Jiang XL, Cui HF. An analysis of 10218 ulcerative colitis cases in China. *World J Gastroenterol* 2002; **8**: 158-161
- Leong RW, Lau JY, Sung JJ. The epidemiology and phenotype of Crohn's disease in the Chinese population. *Inflamm Bowel Dis* 2004; **10**: 646-651
- Ouyang Q, Tandon R, Goh KL, Ooi CJ, Ogata H, Fiocchi C. The emergence of inflammatory bowel disease in the Asian Pacific region. *Curr Opin Gastroenterol* 2005; **21**: 408-413
- Leong RW, Lawrance IC, Ching JY, Cheung CM, Fung SS, Ho JN, Philpott J, Wallace AR, Sung JJ. Knowledge, quality of life, and use of complementary and alternative medicine and therapies in inflammatory bowel disease: a comparison of Chinese and Caucasian patients. *Dig Dis Sci* 2004; **49**: 1672-1676
- Lin CC, Shieh DE. The anti-inflammatory activity of *Scutellaria rivularis* extracts and its active components, baicalin, baicalein and wogonin. *Am J Chin Med* 1996; **24**: 31-36
- Wu JA, Attele AS, Zhang L, Yuan CS. Anti-HIV activity of medicinal herbs: usage and potential development. *Am J Chin Med* 2001; **29**: 69-81
- Ikemoto S, Sugimura K, Yoshida N, Yasumoto R, Wada S, Yamamoto K, Kishimoto T. Antitumor effects of *Scutellariae radix* and its components baicalein, baicalin, and wogonin on bladder cancer cell lines. *Urology* 2000; **55**: 951-955
- Huang HC, Wang HR, Hsieh LM. Antiproliferative effect of baicalein, a flavonoid from a Chinese herb, on vascular smooth muscle cell. *Eur J Pharmacol* 1994; **251**: 91-93
- Kubo M, Kimura Y, Odani T, Tani T, Namba K. Studies on *Scutellariae radix*. Part II: The antibacterial substance. *Planta Med* 1981; **43**: 194-201
- Hong T, Kobayashi T, Song Q, Cyong J. Immunomodulatory action of Oren-gedoku-to on murine colitis model induced by TNBS. *J Traditional Medicines* 1999; **15**: 388-389
- Hong T, Jin G, Kobayashi T, Song Q, Cyong J. Effect of Oren-gedoku-to (Huang-Lian-Jie-Du-Tang) on the murine colitis induced by dextran sulfate sodium. *J Traditional Medicines* 2000; **17**: 66-72
- Hong T, Jin G, Cyong J. Effect of components of Oren-gedoku-to (Huang-Lian-Jie-Du-Tang) on the murine colitis induced by dextran sulfate sodium. *J Traditional Medicines* 2000; **17**: 173-179
- Chen ZY, Su YL, Bi YR, Tsang SY, Huang Y. Effect of baicalein and acetone extract of *Scutellaria baicalensis* on canola oil oxidation. *JAOCs* 2000; **77**: 73-78
- Su YL, Leung LK, Bi YR, Huang Y, Chen ZY. Antioxidant activity of flavonoids isolated from *Scutellariae rehderiana*. *JAOCs* 2001; **77**: 807-812
- Stucchi AF, Shofer S, Leeman S, Materne O, Beer E, McClung J, Shebani K, Moore F, O'Brien M, Becker JM. NK-1 antagonist reduces colonic inflammation and oxidative stress in dextran sulfate-induced colitis in rats. *Am J Physiol Gastrointest Liver Physiol* 2000; **279**: G1298-G1306
- Loher F, Bauer C, Landauer N, Schmall K, Siegmund B, Lehr HA, Dauer M, Schoenharting M, Endres S, Eigler A. The interleukin-1 beta-converting enzyme inhibitor pralnacasan reduces dextran sulfate sodium-induced murine colitis and T helper 1 T-cell activation. *J Pharmacol Exp Ther* 2004; **308**: 583-590
- Siegmund B, Lehr HA, Fantuzzi G. Leptin: a pivotal mediator of intestinal inflammation in mice. *Gastroenterology* 2002; **122**: 2011-2025
- Ko WH, Law VW, Yip WC, Yue GG, Lau CW, Chen ZY, Huang Y. Stimulation of chloride secretion by baicalein in isolated rat distal colon. *Am J Physiol Gastrointest Liver Physiol* 2002; **282**: G508-G518
- Mullane KM, Kraemer R, Smith B. Myeloperoxidase activity as a quantitative assessment of neutrophil infiltration into ischemic myocardium. *J Pharmacol Methods* 1985; **14**: 157-167
- Sandler RS, Eisen GM. Epidemiology of Inflammatory Bowel Disease. In: Kirsner JB, editor. *Inflammatory Bowel Disease*. Philadelphia: W.B. Saunders Company, 2000; 89-112
- Korzenik JR. Past and current theories of etiology of IBD: toothpaste, worms, and refrigerators. *J Clin Gastroenterol* 2005; **39**: S59-S65
- Farrell RJ, Banerjee S, Peppercorn MA. Recent advances in inflammatory bowel disease. *Crit Rev Clin Lab Sci* 2001; **38**: 33-108
- Farrell RJ, Peppercorn MA. Ulcerative colitis. *Lancet* 2002; **359**: 331-340
- Sands BE. Therapy of inflammatory bowel disease. *Gastroenterology* 2000; **118**: S68-S82
- Langmead L, Dawson C, Hawkins C, Banna N, Loo S, Rampton DS. Antioxidant effects of herbal therapies used by patients with inflammatory bowel disease: an in vitro study. *Aliment Pharmacol Ther* 2002; **16**: 197-205
- Cao YB, Zhang JD, Diao YY, Yan L, Wang DJ, Jia XM, Gao PH, Cheng MH, Xu Z, Wang Y, Jiang YY. Effects of Changtai granules, a traditional compound Chinese medicine, on chronic trinitrobenzene sulfonic acid-induced colitis in rats. *World J Gastroenterol* 2005; **11**: 3539-3543
- Guo SM, Tong HB, Bai LS, Yang W. Effect of traditional Chinese medicinal enemas on ulcerative colitis of rats. *World J Gastroenterol* 2004; **10**: 1914-1917
- Liu L, Wang ZP, Xu CT, Pan BR, Mei QB, Long Y, Liu JY, Zhou SY. Effects of Rheum tanguticum polysaccharide on TNBS-induced colitis and CD4+T cells in rats. *World J Gastroenterol* 2003; **9**: 2284-2288
- Mowat AM. Anatomical basis of tolerance and immunity to

- intestinal antigens. *Nat Rev Immunol* 2003; **3**: 331-341
- 30 **Macdonald TT**, Monteleone G. Immunity, inflammation, and allergy in the gut. *Science* 2005; **307**: 1920-1925
- 31 **Chang E**. Intestinal Epithelial Function and Response to Mucosal Injury in Inflammatory Bowel Diseases. In: Kirsner JB, editor. *Inflammatory Bowel Disease*. Philadelphia: W.B. Saunders Company, 2000; 3-19
- 32 **Einerhand AW**, Renes IB, Makkink MK, van der Sluis M, Büller HA, Dekker J. Role of mucins in inflammatory bowel disease: important lessons from experimental models. *Eur J Gastroenterol Hepatol* 2002; **14**: 757-765
- 33 **Hawker PC**, McKay JS, Turnberg LA. Electrolyte transport across colonic mucosa from patients with inflammatory bowel disease. *Gastroenterology* 1980; **79**: 508-511
- 34 **Sandle GI**, Higgs N, Crowe P, Marsh MN, Venkatesan S, Peters TJ. Cellular basis for defective electrolyte transport in inflamed human colon. *Gastroenterology* 1990; **99**: 97-105
- 35 **Schmitz H**, Barmeyer C, Gitter AH, Wullstein F, Bentzel CJ, Fromm M, Riecken EO, Schulzke JD. Epithelial barrier and transport function of the colon in ulcerative colitis. *Ann N Y Acad Sci* 2000; **915**: 312-326
- 36 **Greig E**, Sandle GI. Diarrhea in ulcerative colitis. The role of altered colonic sodium transport. *Ann N Y Acad Sci* 2000; **915**: 327-332
- 37 **Burnett AL**, Donowitz M, Marshall FF. Inhibition of transport processes of intestinal segments following augmentation enterocystoplasty in rats. *J Urol* 1996; **156**: 1872-1875
- 38 **Schulzke JD**, Fromm M, Menge H, Riecken EO. Impaired intestinal sodium and chloride transport in the blind loop syndrome of the rat. *Gastroenterology* 1987; **92**: 693-698
- 39 **Bell CJ**, Gall DG, Wallace JL. Disruption of colonic electrolyte transport in experimental colitis. *Am J Physiol* 1995; **268**: G622-G630
- 40 **Sayer B**, Lu J, Green C, Söderholm JD, Akhtar M, McKay DM. Dextran sodium sulphate-induced colitis perturbs muscarinic cholinergic control of colonic epithelial ion transport. *Br J Pharmacol* 2002; **135**: 1794-1800
- 41 **MacNaughton WK**, Lowe SS, Cushing K. Role of nitric oxide in inflammation-induced suppression of secretion in a mouse model of acute colitis. *Am J Physiol* 1998; **275**: G1353-G1360
- 42 **Asfaha S**, Bell CJ, Wallace JL, MacNaughton WK. Prolonged colonic epithelial hyporesponsiveness after colitis: role of inducible nitric oxide synthase. *Am J Physiol* 1999; **276**: G703-G710
- 43 **Sánchez de Medina F**, Pérez R, Martínez-Augustin O, González R, Lorente MD, Gálvez J, Zarzuelo A. Disturbances of colonic ion secretion in inflammation: role of the enteric nervous system and cAMP. *Pflugers Arch* 2002; **444**: 378-388
- 44 **Kachur JF**, Keshavarzian A, Sundaresan R, Doria M, Walsh R, de las Alas MM, Gaginella TS. Colitis reduces short-circuit current response to inflammatory mediators in rat colonic mucosa. *Inflammation* 1995; **19**: 245-259
- 45 **Diaz-Granados N**, Howe K, Lu J, McKay DM. Dextran sulfate sodium-induced colonic histopathology, but not altered epithelial ion transport, is reduced by inhibition of phosphodiesterase activity. *Am J Pathol* 2000; **156**: 2169-2177
- 46 **Hubel KA**, Renquist KS. Ion transport in normal and inflamed human jejunum in vitro. Changes with electric field stimulation and theophylline. *Dig Dis Sci* 1990; **35**: 815-820
- 47 **Crowe SE**, Luthra GK, Perdue MH. Mast cell mediated ion transport in intestine from patients with and without inflammatory bowel disease. *Gut* 1997; **41**: 785-792
- 48 **Green CL**, Ho W, Sharkey KA, McKay DM. Dextran sodium sulfate-induced colitis reveals nicotinic modulation of ion transport via iNOS-derived NO. *Am J Physiol Gastrointest Liver Physiol* 2004; **287**: G706-G714
- 49 **Wood JD**. Neuro-immunophysiology of colon function. *Pharmacology* 1993; **47** Suppl 1: 7-13

S- Editor Liu Y L- Editor Roberts SE E- Editor Wang HF