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Effects of $1\alpha,25$ dihydroxyvitamin D₃ and testosterone on miRNA and mRNA expression in LNCaP cells

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Abstract

Background: There is evidence from epidemiological and *in vitro* studies that the biological effects of testosterone (T) on cell cycle and survival are modulated by 1,25-dihydroxyvitamin D₃ (1,25(OH)₂D₃) in prostate cancer. To investigate the cross talk between androgen- and vitamin D-mediated intracellular signaling pathways, the individual and combined effects of T and 1,25(OH)₂D₃ on global gene expression in LNCaP prostate cancer cells were assessed.

Results: Stringent statistical analysis identifies a cohort of genes that lack one or both androgen response elements (AREs) or vitamin D response elements (VDREs) in their promoters, which are nevertheless differentially regulated by both steroids (either additively or synergistically). This suggests that mechanisms in addition to VDR- and AR-mediated transcription are responsible for the modulation of gene expression. Microarray analysis shows that fifteen miRNAs are also differentially regulated by 1,25(OH)₂D₃ and T. Among these miR-22, miR-29ab, miR-134, miR-1207-5p and miR-371-5p are up regulated, while miR-17 and miR-20a, members of the miR-17/92 cluster are down regulated. A number of genes implicated in cell cycle progression, lipid synthesis and accumulation and calcium homeostasis are among the mRNA targets of these miRNAs. Thus, in addition to their well characterized effects on transcription, mediated by either or both cognate nuclear receptors, 1,25(OH)₂D₃ and T regulate the steady state mRNA levels by modulating miRNA-mediated mRNA degradation, generating attenuation feedback loops that result in global changes in mRNA and protein levels. Changes in genes involved in calcium homeostasis may have specific clinical importance since the second messenger Ca²⁺ is known to modulate various cellular processes, including cell proliferation, cell death and cell motility, which affects prostate cancer tumor progression and responsiveness to therapy.

Conclusions: These data indicate that these two hormones combine to drive a differentiated phenotype, and reinforce the idea that the age dependent decline in both hormones results in the de-differentiation of prostate tumor cells, which results in increased proliferation, motility and invasion common to aggressive tumors. These studies also reinforce the potential importance of miRNAs in prostate cancer progression and therapeutic outcomes.

Background

Prostate cancer is the most commonly diagnosed non-cutaneous cancer in American males and is the second leading cause of cancer-related deaths in males in North America [1]. Androgens, including testosterone (T) and its active metabolite 5 α -dihydrotestosterone (5 α -DHT),

are important for the development and growth of early stage prostate tumors and exert their effects via androgen receptor (AR) [2-4]. Androgen ablation has been one of the mainstays for the treatment of early stage, organ-confined prostate cancer along with surgery and radiation therapy.

Several epidemiological studies have suggested that adequate levels of vitamin D are critical for the prevention of various solid tumors, including breast, ovarian and colon cancers [5,6]. The risk of developing and

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dying of these cancers appears to be inversely correlated with sun exposure, and/or vitamin D status, suggesting that vitamin D has chemopreventive properties [7]. Some studies have also suggested that vitamin D may play a role in prostate cancer prevention [8,9], but the data are less convincing than in other tumors and several recent meta-analyses have found weak or no associations between serum 25-hydroxyvitamin D₃ (25(OH)₂D₃) levels and tumor incidence and progression [10-13]. In addition, the effects of 1,25(OH)₂D₃ on tumor growth in the TRAMP, LPB-Tag transgenic and Nkx3.1;PTEN mutant mouse models of prostate cancer have produced conflicting results [14-16]. However, a variety of *in vitro* studies demonstrate that 1,25(OH)₂D₃ or its non-calcemic analogs (EB1089; CB 1093; Gemini analogs) induce apoptosis in a variety of prostate cancer cell lines including LNCaP, LNCaP C4-2, ALVA-3, LAPC-4, DU-145 and PC-3 [17-20]. These effects appear to occur through a combination of G₀/G₁ cell cycle arrest, apoptosis, differentiation and inhibition of angiogenesis [21-25]. In contrast, other studies have shown that 1,25(OH)₂D₃ induces cell cycle arrest but not apoptosis [26-28]. These disparate effects of 1,25(OH)₂D₃ on prostate tumor biology appear to be dictated predominantly by the androgen status of the mice [16] or the level of androgen in the culture medium [17,29], suggesting that in prostate cancer, there may be significant cross talk between androgen-mediated growth and vitamin D₃-mediated cell cycle arrest and differentiation which may influence tumor initiation and progression, and impact tumor growth and affect subsequent therapeutic intervention [17].

MicroRNAs (miRNAs) are a class of small non-coding, single-stranded RNAs that post-transcriptionally modulate the steady state levels of mRNA by targeting the 3' untranslated regions (3'UTR) of mRNAs. Recent studies have found that aberrant miRNA expression is closely associated with prostate cancer initiation and progression [30,31]. Several miRNAs that possess either oncogenic (miR-221/222, miR-21, miR-125b) [32-36] or tumor suppressor roles (miR-34 cluster, miR-146a, miR-200c) [37,38] have been identified in prostate cancer and some of these are associated with the castration resistant phenotype [34], or hormone-independent growth of prostate cancer [33]. Neither the effect of 1,25(OH)₂D₃ on miRNA levels in prostate cancer cell lines, nor the interaction with androgen signaling to modulate mRNA and miRNA transcription have been investigated. However, the importance of a regulatory loop involving miR-106b and p21 mRNA which is modulated by 1,25(OH)₂D₃ in non malignant prostate cells has recently been described [39]. The experiments described in this manuscript examine effects of testosterone and 1,25(OH)₂D₃, administered alone or in combination, on the mRNA and

miRNA expression in LNCaP cells and demonstrate that cross talk between VDR- and AR-mediated signaling significantly influences the biology of prostate cancer cells. Using concurrent microarray analyses in LNCaP cells of both miRNA and mRNA, we have found that androgen-mediated transcription of both mRNA and miRNA is enhanced by 1,25(OH)₂D₃, either additively or synergistically, highlighting the extensive cross talk between the two receptors. Many of the gene targets of T and 1,25(OH)₂D₃ have significant clinical relevance. The data demonstrate that while androgens may play a central role in the development of prostate cancer, declining T levels common in older patients may play a significant role in tumor progression, particularly in patients who are also vitamin D deficient.

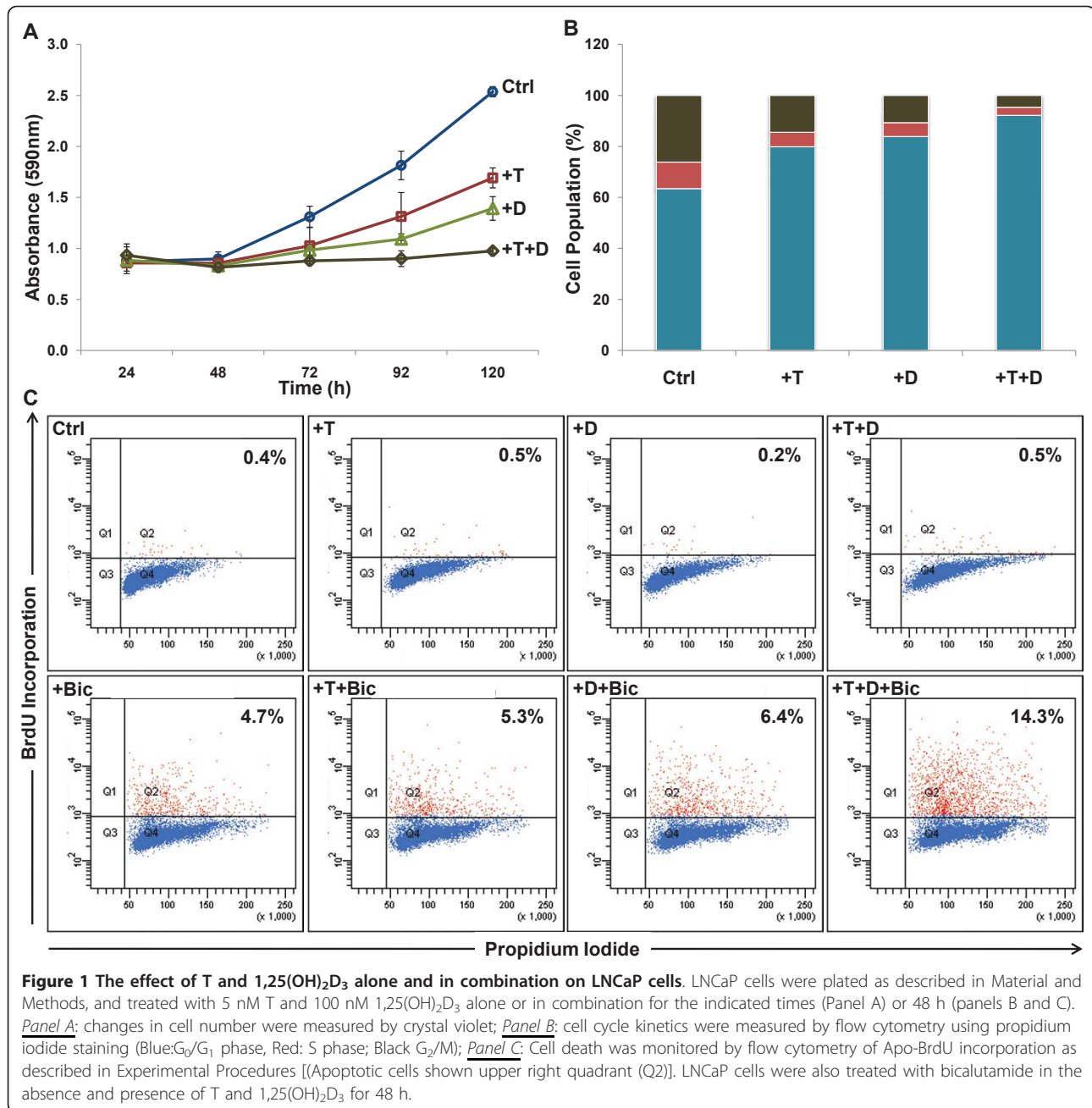
Results

Biological Response of LNCaP cells to T and 1,25(OH)₂D₃

1,25(OH)₂D₃ has variously been reported to induce G₀/G₁ cell cycle arrest or apoptosis in androgen-responsive LNCaP cells and other cell lines. In our hands, 100 nM 1,25(OH)₂D₃ and 5 nM T alone reduce growth of LNCaP cells as measured by crystal violet staining (Figure 1A). This correlates to the induction of G₀/G₁ cell cycle arrest in LNCaP cells (Figure 1B) with no evidence of apoptosis (Figure 1C). The combination of T and 1,25(OH)₂D₃ attenuates cell growth to a greater extent than either treatment alone (Figure 1A), which correlates to the nearly synchronous arrest of the cell populations in the G₀/G₁ phase of the cell cycle (Figure 1B). There was no evidence of cell death in LNCaP cells after treatment with 1,25(OH)₂D₃ alone or in combination with T, as monitored by changes in the sub G₀ population after staining with propidium iodide (not shown) or DNA fragmentation as measured by Apo-BrdU (Figure 1C). The lack of apoptosis in these cells is not due to defects in the apoptotic machinery, since bicalutamide induces apoptosis in LNCaP cells both in the absence and presence of T and 1,25(OH)₂D₃ (Figure 1C). The effects of T and 1,25(OH)₂D₃ on these parameters have been characterized at earlier and later time points with similar results (results not shown). These data demonstrate that AR- and VDR-mediated intracellular signaling pathways cooperate to modulate cell cycle kinetics in prostate cancer cells and attenuate their growth and proliferation without directly affecting apoptosis. They also demonstrate that the combination of T and 1,25(OH)₂D₃ does not block the sensitivity of the cells to bicalutamide.

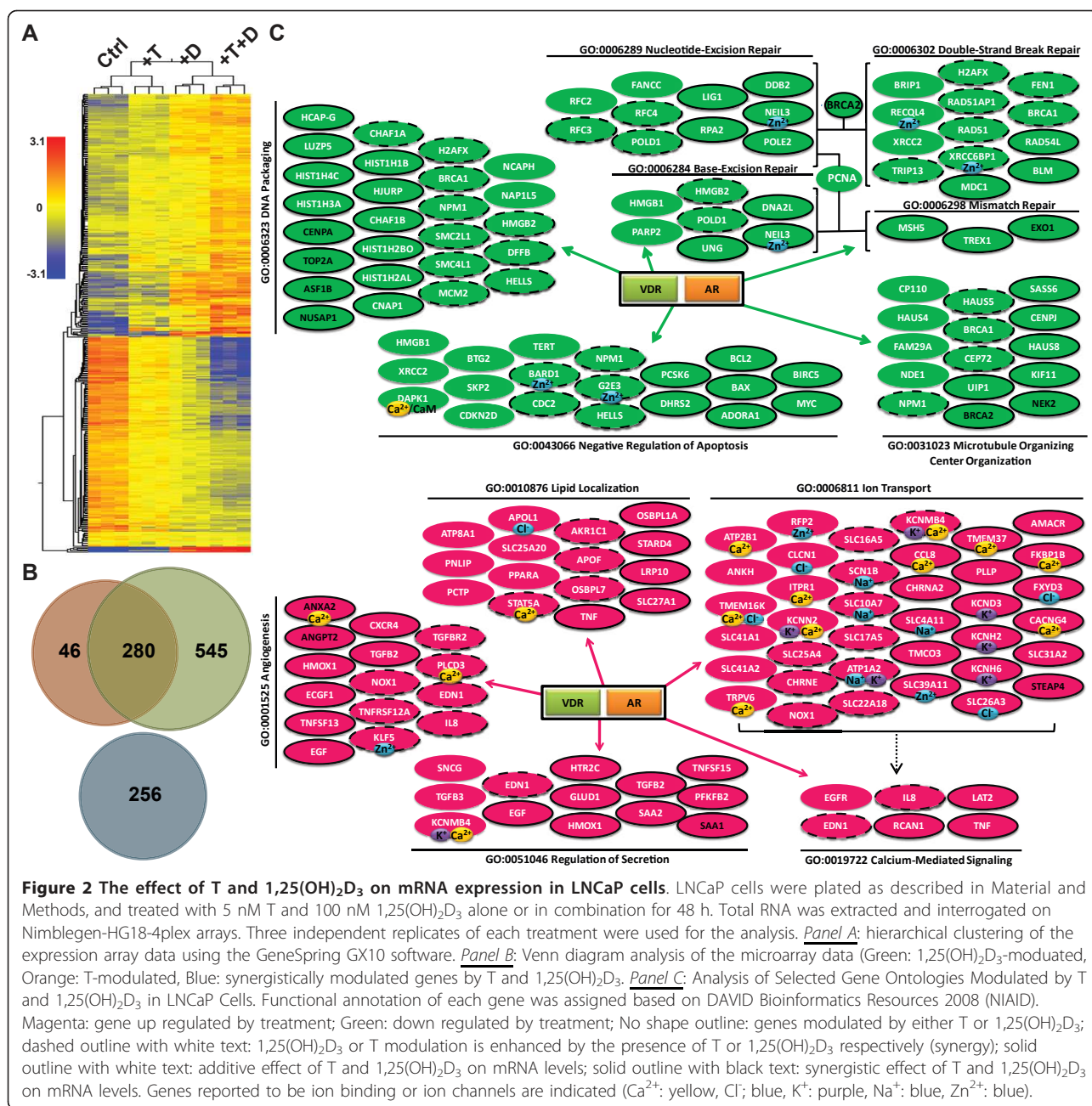
Effect of T and 1,25(OH)₂D₃ on gene expression in LNCaP cells

Total RNA samples obtained from LNCaP cells treated for 48 h with 5 nM T and 100 nM 1,25(OH)₂D₃ alone



or in combination were interrogated with Nimblegen-HG18-4plex whole genome microarrays. Gene expression profiles were clustered based on gene entities and treatment conditions. Treatment with T or 1,25(OH)₂D₃ alone and in combination shows distinct expression patterns that are tightly clustered by their treatment groups (Figure 2A). After filtering for the number of positive probes per gene, statistical analysis on the microarray array data with 1.5 fold cut-off generates a gene list that contains 1127 gene entities that are modulated by either T (326) or 1,25(OH)₂D₃ (825) or both additively (280)

in LNCaP cells. Omnibus testing demonstrates that the effect of T and D on the expression of these genes is highly significant ($p < 0.0001$). Many of the 825 genes regulated by 1,25(OH)₂D₃, identified in this array have been identified as vitamin D responsive genes in other studies [40-42]. Approximately 65% of T modulated genes (202 of 326) has been previously reported to be responsive to androgens (T, 5 α -dihydrotestosterone or R1881) in other *in vitro* systems (Androgen Responsive Gene Database: <http://argdb.fudan.edu.cn>). Thus, addition of exogenous T to the medium of LNCaP cells



identifies a significant new cohort of 124 mRNAs that are androgen responsive. Furthermore, T and 1,25(OH)₂D₃ also synergistically modulate 256 genes that are not significantly regulated by either hormone alone, and thus form a nonintersecting dataset (Figure 2B). These data suggest AR and VDR share many common gene targets and cooperate to regulate cellular processes in LNCaP cells.

Gene Set Enrichment Analysis using Pathways Studio and Gene Ontology analysis from DAVID Bioinformatics Resources (NIAID) were used to assess the

significance of the interactions between T and 1,25(OH)₂D₃ in LNCaP cells. As shown in Table 1, T alone significantly affects processes associated with cell division (particularly mitosis), microtubule based movement, chromosome segregation and progression through anaphase in LNCaP cells. 1,25(OH)₂D₃ alone also significantly affects the expression of genes associated with these cellular processes, in addition to those involved in calcium ion homeostasis and phosphoinositide-mediated signaling. Treatment with T and 1,25(OH)₂D₃ enhances the response of genes associated with these ontologies,

Table 1 Gene Set Enrichment Analysis of representative gene sets identified as significantly enriched after 1,25(OH)₂D₃ treatment in the presence of androgen in LNCaP cells.

Biological Process	Testosterone		1,25(OH) ₂ D ₃		T & D	
	Median Change	p-value	Median Change	p-value	Median Change	p-value
Mitosis	-1.54	3.51E-10	-1.98	3.63E-07	-3.63	1.27E-15
Microtubule-based movement	-1.62	2.54E-05	-2.11	1.17E-04	-4.19	5.78E-07
Chromosome segregation	-1.61	1.11E-03	-1.99	2.13E-03	-3.44	1.12E-04
Anaphase-promoting complex-dependent proteasomal ubiquitin-dependent protein catabolic process	-1.47	5.90E-03	n/a	n/a	-3.63	2.10E-03
DNA repair	n/a	n/a	-1.77	1.08E-02	-2.55	2.24E-03
DNA recombination	n/a	n/a	-2.02	4.30E-02	-3.42	1.41E-02
Phosphoinositide-mediated signaling	n/a	n/a	-1.73	4.97E-02	-2.40	1.60E-02
Elevation of cytosolic calcium ion concentration	n/a	n/a	2.02	1.76E-02	3.43	2.95E-02

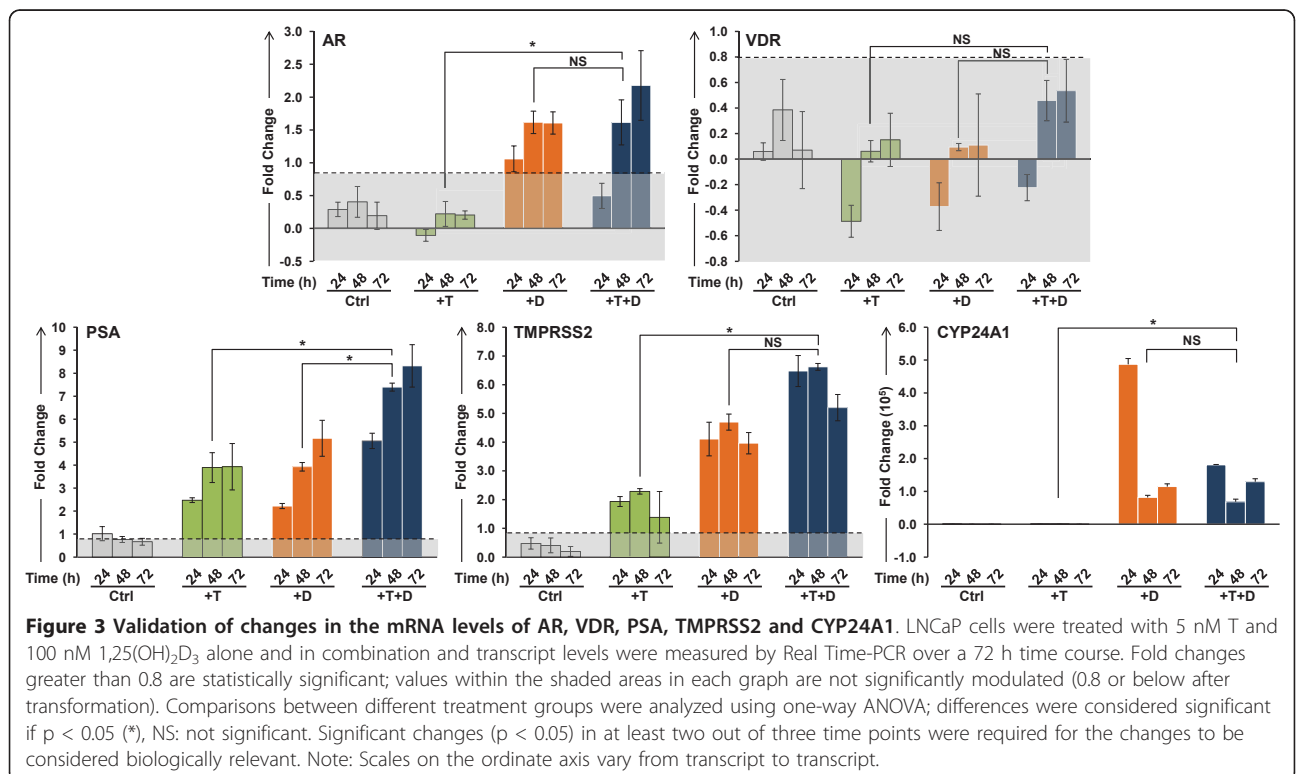
(False Discovery Rate <5%)

and, as revealed by comparing Figure 2C and additional files 1 and 2, the combination of the two hormones also additively or synergistically modulates a significantly greater number of genes than either hormone alone. It is evident that T and 1,25(OH)₂D₃ individually modulate the expression of many of the genes in these ontologies while the combination of the two hormones modulates a significant number of additional genes.

qPCR Validation of Microarray Analyses

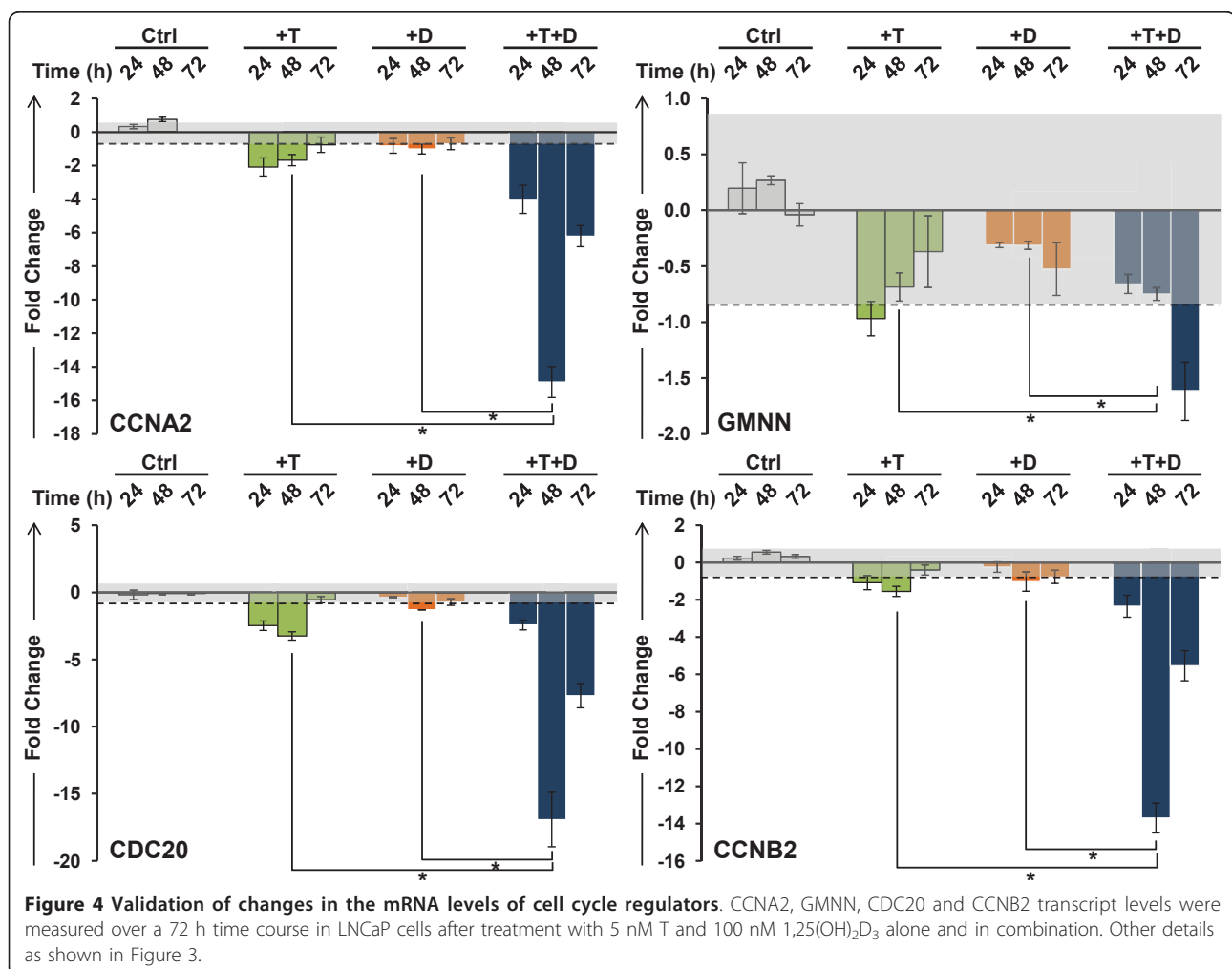
Validation of the microarray data of selected genes associated with the gene sets was performed in LNCaP cells after treatment with T and 1,25(OH)₂D₃ over a 72 h

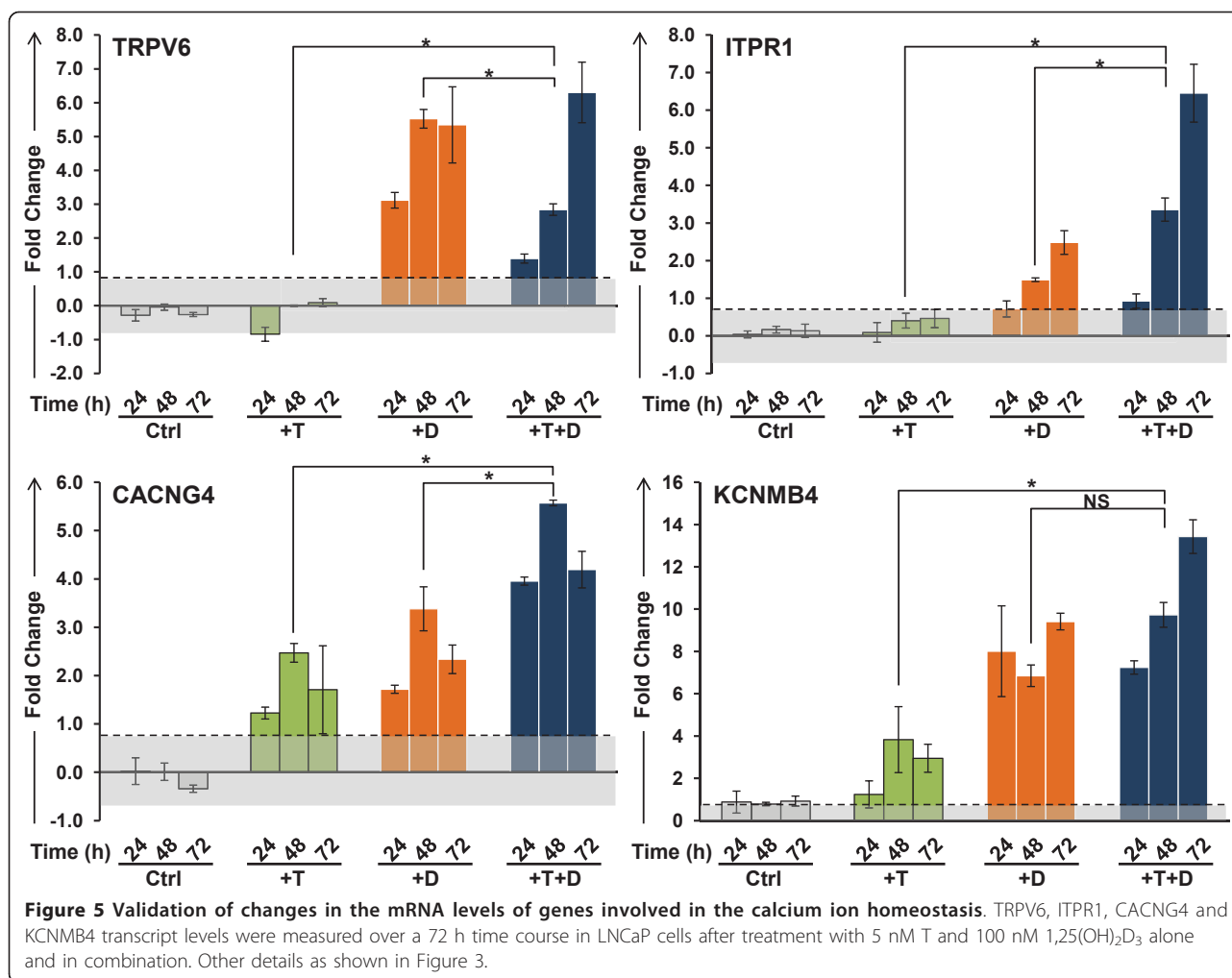
time course. The effects of these treatments on the expression of AR and VDR, as well as two well characterized androgen responsive genes, prostate specific antigen (PSA) and TMPRSS2, and CYP24A1, the classical VDR target gene are shown in Figure 3. Neither the AR nor VDR transcripts are significantly induced in LNCaP cells by T. However, 1,25(OH)₂D₃ alone or in combination with T increases the steady state level of AR mRNA at 48 and 72 h. This corresponds to a consistent increase in the level of the androgen receptor in the nucleus after treatment with 1,25(OH)₂D₃ in the absence or presence of T (additional file 3). Both hormones increase the transcript levels of PSA and



TMPRSS2, and the effect of the two hormones together is additive. The classic VDR target gene CYP24A1 is strongly induced by 1,25(OH)₂D₃, however T alone has little or no effect on its expression. These data demonstrate that the two intracellular signaling pathways are active in LNCaP cells and that the VDR-mediated signaling significantly affects the response of both androgen responsive PSA and TMPRSS2 genes. Representative qPCR validation data, grouped by GO classification, are shown in Figure 4 (cell cycle) and Figure 5 (ion homeostasis), and the changes in the expression of genes associated with other ontologies are shown in additional file 4 (cell survival and cell death), and additional file 5 (lipid metabolism, angiogenesis, DNA repair). It is clear from these data that a number of genes are modulated by the combination of the two steroids either additively (CYP2U1, HPGD, CXCR4, CACNG4, KCNMB4, GMNN) or synergistically (CCNA2, CDC20, CCNB2, Survivin/BIRC5, GADD45G, E2F1, ITPR1, BRCA1). Genes that are known to

modulate the cell cycle (CCNA2, CDC20, CCNB2) are down regulated by T and 1,25(OH)₂D₃ in a similar manner, reaching a nadir at 48 h after treatment (Figure 4). T and 1,25(OH)₂D₃ also display prolonged effects on the steady state levels of the transcripts when administered together and the transcript levels of the genes involved in prostaglandin and lipid metabolism (CYP2U1 and HPGD), and calcium homeostasis (TRPV6, ITPR1, and KCNMB4) show time-dependent increases throughout the 72 h time course (Figure 5). It is well established that ligand-activated AR and VDR bind to cognate response elements (ARE and VDRE respectively) to modulate gene transcription and thus the coordination between T and 1,25(OH)₂D₃ may be the consequence of receptor mediated transcription by both AR and VDR. While *in silico* searches for ARE and VDREs within 10 kb upstream or downstream of the structural genes shown to be modulated by T or 1,25(OH)₂D₃ or both, demonstrate that many of the genes have canonical response elements in their promoters





(additional file 6), nearly 50% of the genes identified in the expression microarray analysis appear to lack functional hormone response elements (either ARE or VDRE or both) in their promoters (additional file 6). For instance, genes that have previously been documented to be only 1,25(OH)₂D₃ inducible and/or contain VDRE at their promoters (KCNMB4, CXCR4) are also modulated by T (Figures 5 and additional file 4). Furthermore, genes such as CDC20 that lack both VDREs and AREs are down regulated by 1,25(OH)₂D₃ and T together while neither steroid alone has significant effects on gene expression (Figures 4 and additional file 5). These data demonstrate that 1,25(OH)₂D₃ has distinct effects on the regulation of transcript levels and that for a significant proportion the genes, the effects of 1,25(OH)₂D₃ require the presence of T for maximal effect. Furthermore, it appears that in addition to modulating transcription of the responsive genes, T and 1,25(OH)₂D₃ modulate the stability of the transcripts via modulation of miRNA expression.

Effect of T and 1,25(OH)₂D₃ on miRNA expression

To examine the effect of T and 1,25(OH)₂D₃ on miRNA expression, total RNA prepared from LNCaP cells 48 h after treatment using the same experimental paradigm outlined above was analyzed on the Agilent Human miRNA microarray v3, which contains 866 human miRNAs from the Sanger database v12.0. The miRNA expression profiles in LNCaP cells after treatment cluster to the different treatment groups are shown in Figure 6A. Statistical analysis with a stringent cut-off of 2.0 fold, identifies 15 miRNAs that are significantly modulated by T and 1,25(OH)₂D₃ (Table 2). The majority of miRNAs identified are up regulated by T and 1,25(OH)₂D₃ additively, including miR-29ab and miR-371/373 clusters. In contrast, two miRNAs, including miR-17 and miR-20a, members of the miR-17/92 cluster, are down regulated by T and 1,25(OH)₂D₃ synergistically. Bioinformatic analysis using the available miRNA target prediction databases (TargetScan Human v5.1) identifies approximately 8500 putative mRNA targets of these

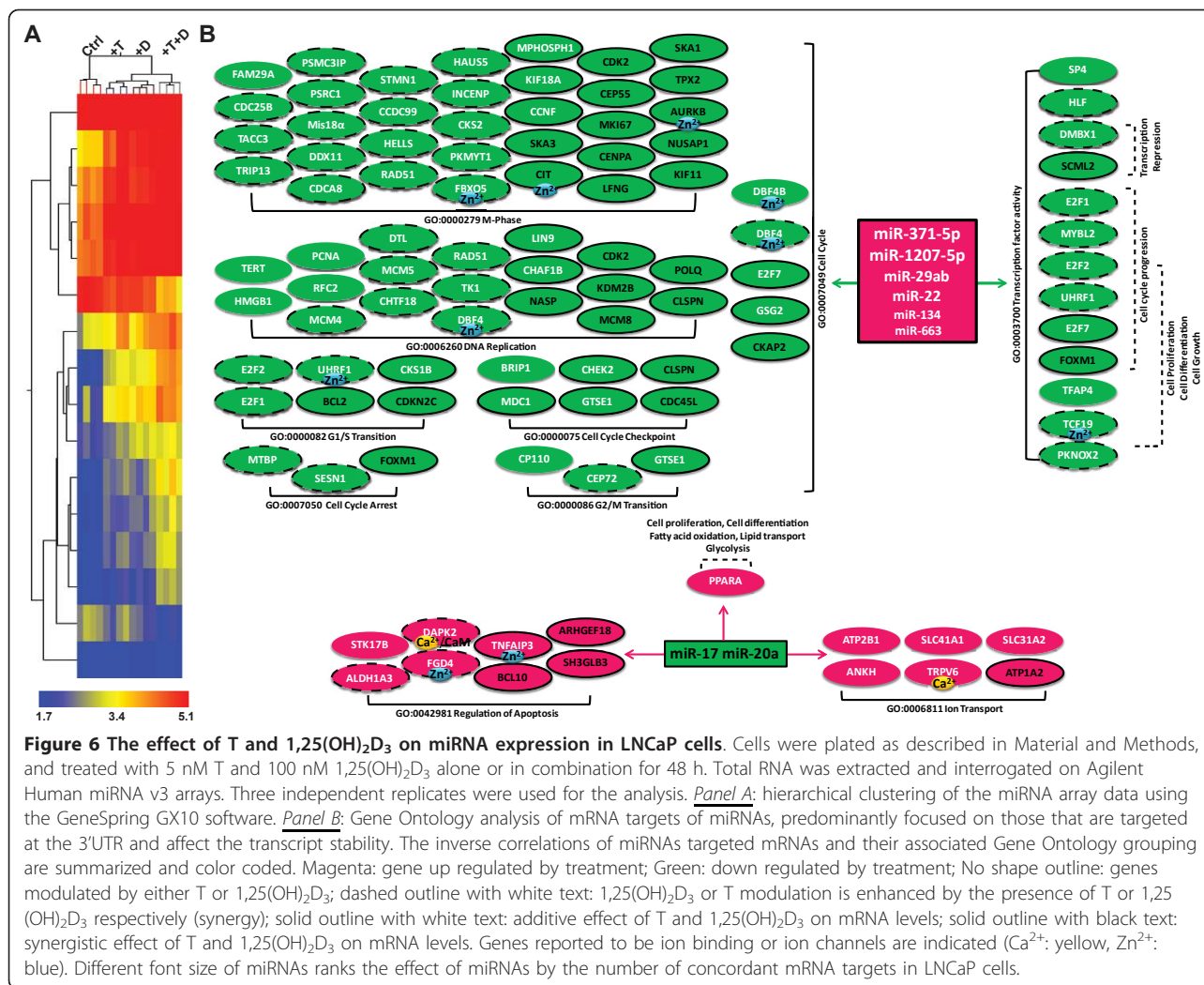


Table 2 miRNAs differentially modulated by 1,25(OH)₂D₃ and T alone or in combination in LNCaP cells, identified by Agilent Human miRNA microarray v3.

miRNA	Seed	Testosterone	1,25(OH) ₂ D ₃	T + 1,25(OH) ₂ D ₃
hsa-miR-17	aaagug	-1.237	-1.255	-2.337
hsa-miR-20a	aaagug	-1.220	-1.287	-2.318
hsa-miR-20b	aaagug	-1.174	-1.391	-2.273
hsa-miR-542-5p	cgggga	-1.232	2.304	1.197
hsa-miR-29b	agcacc	1.317	1.911	2.081
hsa-miR-1207-5p	ggcagg	1.756	1.637	2.242
hsa-miR-22	agcugc	1.691	1.745	2.312
hsa-miR-1915	cccagg	2.005	2.044	3.007
hsa-miR-29a	agcacc	2.122	2.577	3.108
hsa-miR-371-5p	cucaaa	1.918	2.163	3.197
has-miR-663	ggcggg	2.293	2.561	3.685
hsa-miR-134	gugacu	2.995	2.923	4.597
hsa-miR-135a*	auaggg	2.609	2.773	4.818
hsa-miR-1181	cgucgc	3.003	3.454	5.775
hsa-miR-629*	uucucc	3.143	3.263	6.160

The seed sequence corresponds to individual miRNAs is indicated.

miRNAs. However, most of these targets are not expressed in the prostate and are not identified as differentially regulated by T and 1,25(OH)₂D₃ by microarray. In total, 264 target transcripts are responsive to T and 1,25(OH)₂D₃ in LNCaP cells and show an inverse association with the targeting miRNA(s) (Figure 6B). This corresponds to approximately 23% of the genes modulated by T and 1,25(OH)₂D₃ in LNCaP cells. However this is at best a rough estimate since many of the targets of the modulated miRNAs identified by Target Scan have not yet been validated in LNCaP cells. A complete list of the miRNAs and their mRNA targets is provided in additional file 7.

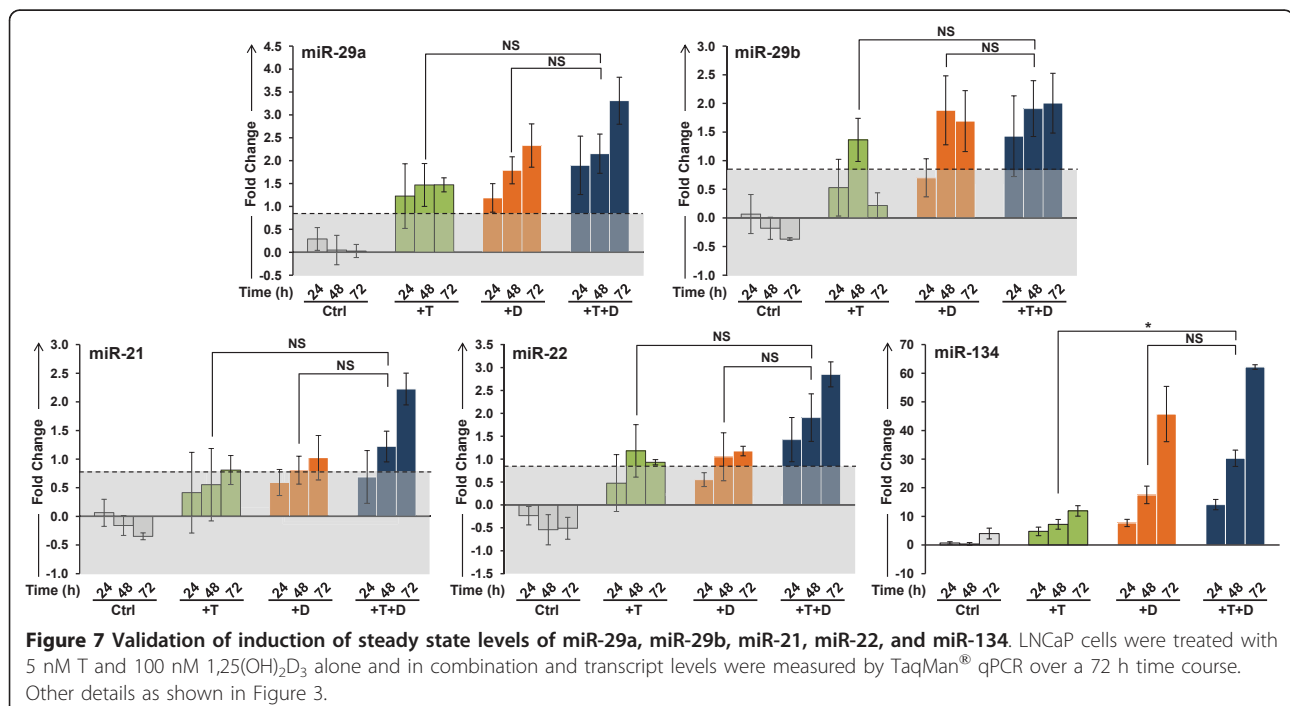
The effects of T and 1,25(OH)₂D₃ on the steady state levels of selected miRNAs were further assessed in LNCaP cells by TaqMan[®] qPCR to validate microarray data. T and 1,25(OH)₂D₃ alone showed time-dependent induction of miR-29a, miR-29b, miR-21, miR-22 and miR-134 expression in LNCaP cells while the combination of the two have a more rapid additive effects on these miRNAs (Figure 7). In contrast, neither steroid alone down regulates miR-17 and miR-20a of the miR-17/92 cluster, however the combination of T and 1,25(OH)₂D₃ significantly down regulates miR-17 and miR-20a, demonstrating the synergistic ability of T and 1,25(OH)₂D₃ to modulate miRNA levels (Figure 8). We have also assessed the changes of miR-18a, another member of the miR-17/92 cluster. Changes in miR-18a transcript levels showed a similar pattern as that of miR-17 and 20a, suggesting that T and 1,25(OH)₂D₃ together

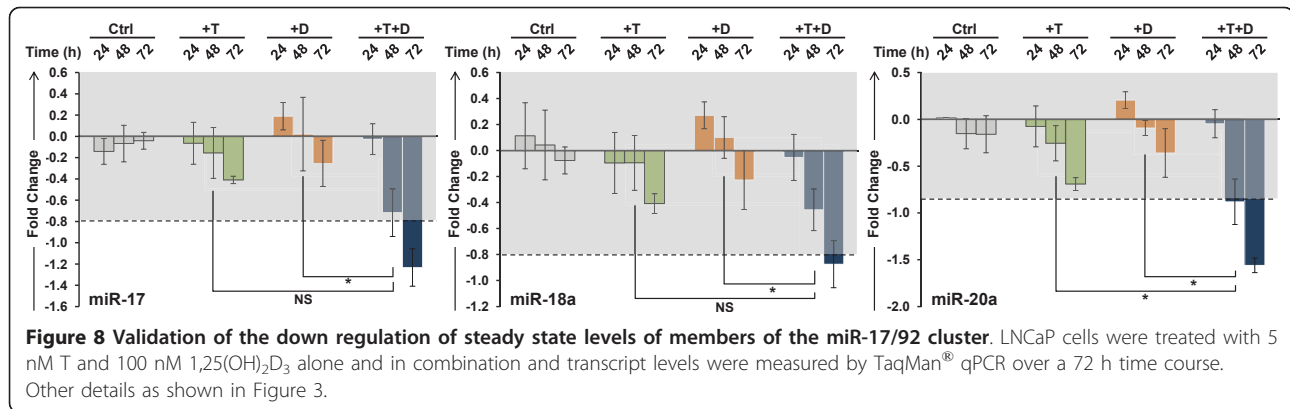
modulate all members of the miR-17/92 cluster rather than selectively down regulating individual members of the cluster.

In addition to the miRNAs identified from the microarray analysis, we have also assessed the effect of T and 1,25(OH)₂D₃ on miRNAs that have been reported to be deregulated in prostate cancer. The oncogenic miR-221, which has been reported to contribute to androgen-resistance phenotype is slightly down regulated by T and 1,25(OH)₂D₃ at 48 h, though the fold change is not statistically significant (additional file 8). Changes in the AR-inducible [35] and VDR-inducible [43] miR-125b are also not significant in LNCaP cells in this experimental paradigm (additional file 8). However, as already shown, the steady state levels of miR-21 are up regulated by T and 1,25(OH)₂D₃ in LNCaP cells with the highest fold-induction at 72 h in the presence of both hormones (Figure 7). This implies that the expression of miR-21 targets, such as PDCD4, may also be modulated by AR and VDR in LNCaP cells.

Discussion

The data presented here demonstrate that both T and 1,25(OH)₂D₃ modulate the mRNA and miRNA profiles in LNCaP cells, and that the combination of the two hormones modulates a significantly larger cohort of transcripts than either hormone alone. In most cases, the effects of the two hormones are additive or synergistic. Since the predicted changes in the cohort of proteins present in the cells would be expected to influence the





response to therapeutic agents, this raises questions regarding the interpretation of previous *in vitro* data describing the responsiveness of prostate cancer cell lines grown in the absence of these hormones.

The time frame we have chosen for the initial microarray experiments (48 h) and the times chosen for qPCR validation (24, 48 and 72 h) were selected based on previous experiments examining the induction of apoptosis in LNCaP cells by diverse agents including bicalutamide and Iejimalide [44]. These time points will not discriminate between primary effects of T and 1,25(OH)₂D₃ on transcription and secondary effects which may be due to either the modulation of miRNA, and subsequent degradation of target mRNAs or cascading regulation of transcription of other genes. However, this time frame is appropriate to examine the effects of induced miRNAs on transcript stability.

Bioinformatic analysis demonstrates that T and 1,25(OH)₂D₃ alter the expression of genes associated with several relevant gene ontologies that clearly have the potential to significantly influence the biology of the tumor cells, their interaction with the tumor microenvironment and their response to therapeutic intervention. Perhaps the most important of these changes relate to the regulation of calcium ion homeostasis and cell cycle progression. One of the primary physiological roles of 1,25(OH)₂D₃ is to regulate calcium and phosphorus metabolism in bone. However, 1,25(OH)₂D₃ also plays an important role in cell cycle regulation in many cancers, including breast, ovarian and colon cancer, through its interaction with the VDR [6,45-48]. In LNCaP cells, 1,25(OH)₂D₃ induces the expression of both voltage-gated (CACNG4) and non-voltage-gated (TRPV6) Ca²⁺ channels located on the plasma membrane. Though TRPV6 is primarily modulated by VDR, the full induction of ITPR1, annexin AII (ANXA2) and S100A10, major components of the TRPV6 auxiliary protein complex, requires the presence of both hormones, as does the induction of the highly Ca²⁺

sensitive PLC-δ3 (PLCD3). There is also a concurrent increase in ITPR1 in response to T and 1,25(OH)₂D₃ which has been shown to lead to increased signaling through the PI3K/AKT pathway [49]. This suggests that in response to T and 1,25(OH)₂D₃, the production of IP3 will increase, triggering the release of Ca²⁺ from endoplasmic reticulum (ER) stores via ITPR1, leading to the activation of TRPV6 through S100A10, resulting in an influx of Ca²⁺ and augmenting intracellular Ca²⁺ levels. In addition, diacylglycerol, produced by PLC mediated cleavage of PIP2, may activate PKC, to further increase the activity of TRPV6 via phosphorylation [50]. Together, these data suggest that the combination of T and 1,25(OH)₂D₃ elevates the intracellular Ca²⁺ level to establish a new homeostatic set point without inducing cell death. Since the induction of the execution phase of apoptosis in prostate cancer cells requires the elevation of intracellular free Ca²⁺ into the micromolar range [51], resetting the intracellular Ca²⁺ concentration closer to this threshold may render tumor cells more sensitive to apoptosis-inducing agents including doxorubicin, bicalutamide and radiation. It is well known that uncontrolled release of Ca²⁺ stores from the ER, for example in response to thapsigargin, initiates apoptosis via the activation of Ca²⁺-dependent caspases accompanied by release of cytochrome c through the mitochondrial permeability transition pore [52]. It is therefore likely that a fine-tuned Ca²⁺ balance within the cellular compartments in prostate cancer cells occurs after treatment with 1,25(OH)₂D₃ and thus prevents Ca²⁺ overload in the mitochondria and maintains other Ca²⁺-dependent signaling, which may include the modulation of miRNA expression [53]. Importantly, both T and 1,25(OH)₂D₃ appear to be required to establish the elevated homeostatic calcium levels. In this regard, the capacitative Ca²⁺ entry that has been shown to block the development of the apoptosis resistance phenotype such as that seen in Bcl-2 over-expressing LNCaP cells [54,55] may be equivalent to the elevated homeostatic Ca²⁺ level

induced by T and $1,25(\text{OH})_2\text{D}_3$, suggesting that adequate levels of the two hormones should also prevent the development of the apoptosis-resistant, or castration resistant, phenotype in prostate cancer.

Many of the changes in the steady state mRNA levels can be attributed to changes in transcriptional activity due to the presence of functional AREs and/or VDREs in the promoters of target genes. However, *in silico* searches suggests that the promoters of nearly 40% of the affected genes do not contain either response element. Both T and $1,25(\text{OH})_2\text{D}_3$ have been shown to induce rapid Ca^{2+} influx via store-operated Ca^{2+} (SOC) channels whose activities are mediated by membrane receptors (mVDR & mAR) and are related to the non-genomic action of AR and VDR [56-58]. In addition, the ligand-activated VDR can modulate transcription of some target genes through Sp-1 sites [59-61]. Therefore, it is likely that additional mechanisms are responsible for the extensive modulation seen in response to the two hormones.

As demonstrated here, T and $1,25(\text{OH})_2\text{D}_3$ cooperatively modulate a circumscribed group of miRNAs, which mediate mRNA degradation depending on the complementarity of the miRNA seed sequence to sequences within the 3'UTR of the transcript. While alterations in individual miRNAs are not as profound as those seen in the mRNA profiles, many of the miRNAs share identical seed sequences, which results in a cumulative effect on the target transcripts. In LNCaP cells, most of the miRNAs identified in this study are up regulated in response to T and $1,25(\text{OH})_2\text{D}_3$, including miR-21, miR-22, miR-29ab, miR-134 and miR-371-5p and miR-1207-5p. Both miR-21 and miR-29b have previously been shown to be induced by R1881 in LNCaP and LAPC-4 prostate cancer cells [62]. Induction of these miRNAs results in substantial down regulation of several large cohorts of genes classified by GO. Thus, in addition to down regulating transcription through their cognate hormone response elements, T and $1,25(\text{OH})_2\text{D}_3$ can profoundly affect the stability of the transcripts encoding proteins that function in cell cycle control, cytoskeleton organization, and DNA damage repair among others. In this context, MYCBP, a positive regulator of c-Myc activity and a validated miR-22 target should be repressed by T and $1,25(\text{OH})_2\text{D}_3$, thus inhibiting c-Myc mediated transcription of E-box containing genes [63]. In breast cancer cells, this correlates with suppressed cell proliferation and anchorage-independent growth suggesting that increased expression of miR-22 at 48 h in prostate tumor cells by T and $1,25(\text{OH})_2\text{D}_3$ may be partially responsible for cell cycle arrest and the prevention of tumor progression. In LNCaP cells miR-134 is very significantly up regulated by $1,25(\text{OH})_2\text{D}_3$ in the absence and presence of T. The role of miR-134 in

prostate cancer has not been previously described, however the steady state level of miR-134 is modulated by members of the p53/p73/p63 family as part of a miRNA-tumor suppressor network [64]. Thus, increased expression of miR-134 may further contribute to the activity of T and $1,25(\text{OH})_2\text{D}_3$ to suppress tumor growth.

While miR-21 was not identified in the microarray as a T and/or $1,25(\text{OH})_2\text{D}_3$ regulated miRNA, it was identified in the more informative qPCR analysis as a target of T and $1,25(\text{OH})_2\text{D}_3$, and the increases in the steady state level of miR-21 are significant after 48 h of treatment. Previous studies in breast and pancreatic cancer cell lines have suggested that over expression of miR-21 may lead to increased cell proliferation and decreased apoptosis through the targeted degradation of tumor suppressor protein PDCD4 [62,65]. In prostate cancer cells, these effects would be anticipated to counteract the anti-proliferative effects of $1,25(\text{OH})_2\text{D}_3$, suggesting that not all interactions between AR- and VDR-mediated signaling are beneficial. Interestingly, neither of the other miRNAs that have been implicated in prostate tumor progression (miR-221 or miR-125b) is modulated by T nor $1,25(\text{OH})_2\text{D}_3$ in this *in vitro* model system. In addition to up regulating miRNAs that encode cell cycle regulatory proteins and calcium ion homeostasis, T and $1,25(\text{OH})_2\text{D}_3$ down regulate the expression of the oncomiR cluster, miR-17/92. These data suggest that miRNAs may play important roles in T- and $1,25(\text{OH})_2\text{D}_3$ -induced cell cycle arrest in prostate cancer cells. The concurrent analysis of mRNA and miRNA expression has demonstrated that many of the combined effects of T and $1,25(\text{OH})_2\text{D}_3$ are modulated by a small cohort of 15 miRNAs that are additively or synergistically regulated by the two hormones. Since the majority of men diagnosed with prostate cancer are likely to be vitamin D insufficient [66], these data may have a profound impact on our understanding of the molecular mechanisms underlying the chemopreventive and chemotherapeutic effects of vitamin D_3 . Based on the data presented here, vitamin D deficiency is likely to render tumor cells more aggressive and less sensitive to chemotherapy. Given the trend toward "active surveillance" for men diagnosed with early stage prostate cancer, understanding the cross talk between the androgen- and vitamin D-mediated cellular effects may have a significant impact on the care of men in the period between diagnosis and the initiation of treatment. If these two signaling pathways interact in tumor tissue as demonstrated here, individual variations in dietary vitamin D and/or sun exposure, as well as differences in circulating T levels, may greatly influence the rate of prostate tumor growth and the sensitivity of prostate cancer to hormone and other chemotherapies. In this

context, the age related decline in serum testosterone may also contribute to the progression of prostate cancer. Thus, maintaining serum testosterone and combined with supplementation of vitamin D may substantially slow disease progression for men diagnosed with very early stage cancer, extending the time between diagnosis and treatment.

Methods

Cell Culture

LNCaP human prostate cancer cells, obtained from American Type Culture Collection (Rockville, MD), were grown in RPMI-1640 medium (Invitrogen, Carlsbad, CA) supplemented with 10% FBS (Sigma-Aldrich, St Louis, MO), 100 U/mL penicillin and 100 µg/mL streptomycin. Cells were maintained at 37°C in a humidified atmosphere of 95% air/5% CO₂. For all the experiments performed in this study, with the exception of crystal violet assays, LNCaP cells were plated at a density of 1×10^6 cells per 150 cm² dish for 48 h prior to treatment with 5 nM T (Sigma-Aldrich) and 100 nM 1,25(OH)₂D₃ (Sigma-Aldrich) alone or in combination. The steroids were dissolved in ethanol, and control cells were treated with the same volume of vehicle.

Crystal Violet Assay

LNCaP cells were seeded at 20,000 cells/well in 24 well plates for 24 h prior to treatment with 5 nM T and 100 nM 1,25(OH)₂D₃ alone or in combination. Cells were fixed with 2% glutaraldehyde in PBS for 20 min at room temperature followed by staining with 0.1% crystal violet (Sigma-Aldrich) for 30 min. The crystal violet stain was solubilized in 0.2% Triton X-100 in ddH₂O for 30 min and the absorbance was read with Victor³V 1420 Multi-label Counter (PerkinElmer Inc, Waltham MA) at 590 nm. Three independent biological replicates were analyzed in triplicate.

Flow Cytometry

LNCaP cells were treated with 5 nM T and 100 nM 1,25(OH)₂D₃ as described above. Cells treated with either vehicle (EtOH) or 100 µM bicalutamide serve as the negative and positive control, respectively. For cell cycle analysis, the cells were treated for 24, 48 and 72 h and harvested by trypsinization, followed by 90% ethanol permeabilization overnight at -20°C. Permeabilized cells were stained with 5 µg/mL propidium iodide (Sigma-Aldrich) in the presence of 0.015 U/mL RNase (Roche Applied Science, Indianapolis, IN) in PBS for 20 min at room temperature. Samples were analyzed within 3 h of labeling on BD™ LSR II Flow Cytometer (BD Biosciences, San Jose, CA). A minimum of 10,000 events were analyzed for each experimental condition. Three

independent biological replicates for each treatment group were analyzed.

Apoptosis was analyzed using Apo-BrdU staining. Cells were harvested by trypsinization and fixed with 4% formaldehyde in PBS for 30 min on ice, followed by 70% ethanol permeabilization overnight at -20°C. Samples were enzymatically labeled with bromodeoxyuridine triphosphate in TdT reaction buffer (Br-dUTP, 2.5 mM cobalt chloride, and terminal transferase 24,000 U) for 1 h at 37°C to label the 3'-OH ends of fragmented DNA (Roche). DNA strand breaks were counterlabeled with FITC-conjugated anti-BrdU monoclonal antibody according to the manufacturer's directions (Phoenix Flow Systems, San Diego, CA). Cells were counterstained with 5 µg/mL propidium iodide for 30 min at room temperature. Samples were analyzed on BD™ LSR II Flow Cytometer (BD Biosciences) within 3 h of labeling and a minimum of 10,000 events were analyzed for each experimental condition. Three independent biological replicates for each treatment group were analyzed.

mRNA Microarray Analysis

Total RNA isolated from LNCaP cells was processed using standard protocols for Nimblegen arrays. Briefly 10 µg of total RNA was reverse transcribed to cDNA using oligo-dT primers and Superscript II (Invitrogen), converted to double stranded cDNA and Klenow labeled with Cy3-labeled random 9-mers before hybridization to Nimblegen-Human-HG18 4 × 72 microarrays at 42°C for 16 h using a Nimblegen Hybridization system. The arrays were washed and scanned on a Genepix 4000B scanner following which the data was extracted using NimbleScan software. Further data analysis was performed using GeneSpring GX10. The raw dataset is available as a curated dataset at GEO (SuperSeries GSE23815, SubSeries GSE17461). Three independent biological replicates for each treatment group were analyzed.

miRNA Microarray Analysis

Total RNA was processed and hybridized to Agilent Human miRNA microarrays using standard protocols. 100 ng of total RNA was dephosphorylated with calf intestinal phosphatase and end-labeled with Cy3-pCp by T4 RNA ligase prior to an overnight hybridization at 55°C onto Agilent Human miRNA v3 (Sanger release 12.0) microarrays. The arrays were washed and scanned on a high resolution GC2565CA Agilent Scanner using the manufacturer's recommended settings. The raw data was extracted using Agilent Feature Extraction software v10.1.1 and imported into GeneSpring GX10 for further analysis. The raw data is available as a curated dataset at GEO (SuperSeries GSE23815, SubSeries GSE23814).

qPCR Validation of Microarray Data

Changes in mRNA and miRNA identified by microarray were validated in independent biological replicates. LNCaP cells were plated and treated as previously described for 24, 48 and 72 h. Cells were harvested by trypsinization and total RNA (both mRNA and miRNA) was extracted using miRNeasy mini kit (Qiagen, Valencia, CA). Reverse transcription PCR reactions were performed with 1.5 µg total RNA using Taqman[®] Reverse Transcription Reagents (Applied Biosystems, Carlsbad, CA) to synthesize cDNA for mRNA expression analysis. The reaction mixture was incubated for 10 min at 25°C, 1 h at 37°C and 5 min at 95°C and kept at -20°C until further analysis. qPCR probes for each gene were designed using Primer-Blast (National Center for Biotechnology Information) with default settings and synthesized by Integrated DNA Technologies (Coralville, IA). The list of primers for each gene is available in additional file 9. SYBR Green reactions with SYBR[®] Green PCR Master Mix (Applied Biosystems) were analyzed using the ABI 7900HT Fast Real-Time PCR System (Applied Biosystems): 50°C for 2 min, 95°C for 10 min, 95°C for 15 sec and 60°C for 1 min, repeated for 40 cycles. Relative expression levels of each gene in real time were analyzed using the $2^{-\Delta\Delta C_T}$ method [67] and presented as ratio relative to the expression of the housekeeping gene GAPDH. For miRNA expression analysis, 10 ng total RNA were used to make cDNA with TaqMan[®] MicroRNA Reverse Transcription Reagents (Applied Biosystems). The reaction mixture was incubated for 30 min at 16°C, 30 min at 42°C and 5 min at 85°C and kept at -20°C until further analysis. TaqMan[®] MicroRNA Assays were used to evaluate miRNA expression according to manufacturer's protocol. TaqMan reactions were analyzed using the ABI 7900HT Fast Real-Time PCR System: 95°C for 10 min, 95°C for 15 sec and 60°C for 60 sec repeated for 40 cycles. Relative expression levels of each miRNA in real time were analyzed using the $2^{-\Delta\Delta C_T}$ method with U6 snRNA as the reference control. Each sample was replicated twice from three independent sets of RNA preparations. Results are tabulated as mean ± SD and presented as fold change after transformation to show divergence from no effect (zero fold change).

Statistical Analysis of Microarray Data

For mRNA microarray analysis in GeneSpring GX10, the gene list was filtered to exclude entities which showed low signal values across all samples (i.e. bottom 20th percentile). Statistically significant genes from each expression profile were selected using one-way ANOVA ($p \leq 0.05$). The multiple testing correction Benjamini and Hochberg false discovery rate (FDR) (p -value < 0.05) was integrated within each test. A fold change cut-off at

1.5 fold was implemented to generate the final list of differentially expressed genes. Genes passing the statistical tests were further assigned into their gene ontology (GO) grouping using DAVID Bioinformatics Resources v6.7 (NIAID) with the default setting, essentially as previously described [68]. Significantly regulated and enriched GO groups were selected for qPCR analysis.

For miRNA microarray analysis, the raw data was imported into GeneSpring GX10, log₂ transformed, normalized to the 75th percentile following which the entity list was filtered to exclude probes that showed low signal values across all treatment groups (i.e. bottom 20th percentile). This list was further filtered to only include entities that were marked "present;" or "marginal" in all 3 replicates for at least one of the 4 treatment groups. Entities with fold changes greater than 2.0 were considered significant when $p < 0.05$ using one-way ANOVA with Benjamini-Hochberg FDR post-test. Targets of miRNAs that passed the statistical test were identified using TargetScan Human v5.1 from Whitehead Institute for Biomedical Research. Gene targets whose mRNA expression levels were inversely proportional to the corresponding miRNA expression levels were considered concordant and were further analyzed for their enrichment in Gene Ontology grouping.

Additional material

Additional file 1: Analysis of Selected Gene Ontologies Modulated by T in LNCaP Cells. Functional annotation of each gene was assigned based on DAVID Bioinformatics Resources 2008 (NIAID). Magenta: gene up regulated by treatment; Green: down regulated by treatment; No shape outline: genes modulated by either T or 1,25(OH)₂D₃; dashed outline with white text: 1,25(OH)₂D₃ or T modulation is enhanced by the presence of T or 1,25(OH)₂D₃ respectively (synergy); solid outline with white text: additive effect of T and 1,25(OH)₂D₃ on mRNA levels; solid outline with black text: synergistic effect of T and 1,25(OH)₂D₃ on mRNA levels. Genes reported to be ion binding or ion channels are indicated (Ca²⁺: yellow, Zn²⁺: blue)

Additional file 2: Analysis of Selected Gene Ontologies Modulated by 1,25(OH)₂D₃ in LNCaP Cells. Functional annotation of each gene was assigned based on DAVID Bioinformatics Resources 2008 (NIAID). Magenta: gene up regulated by treatment; Green: down regulated by treatment; No shape outline: genes modulated by either T or 1,25(OH)₂D₃; dashed outline with white text: 1,25(OH)₂D₃ or T modulation is enhanced by the presence of T or 1,25(OH)₂D₃ respectively (synergy); solid outline with white text: additive effect of T and 1,25(OH)₂D₃ on mRNA levels; solid outline with black text: synergistic effect of T and 1,25(OH)₂D₃ on mRNA levels. Genes reported to be ion binding or ion channels are indicated (Ca²⁺: yellow, Zn²⁺: blue)

Additional file 3: Immunoblotting analysis of 5 nM T and 100 nM 1,25(OH)₂D₃ on nuclear AR and VDR expression. LNCaP cells were treated with 5 nM T and 100 nM 1,25(OH)₂D₃ alone and in combination for 48 h. Nuclear proteins were extracted and ran on 10% SDS-PAGE and transferred to PVDF membrane. Antibodies against AR (Millipore) and VDR (Santa Cruz) were used to detect the protein levels of AR and VDR in the nucleus. Lamin A/C was used as the loading control.

Additional file 4: Validation on changes in the mRNA levels of genes involved in cell death. GADD45G, STK17B, E2F1 and Survivin/BIRC5 transcript levels were measured over a 72 h time course in LNCaP

cells after treatment with 5 nM T and 100 nM 1,25(OH)₂D₃ alone and in combination. Other details as shown in Figure 3.

Additional file 5: Validation on changes in the mRNA levels of selected genes involved in lipid metabolism, angiogenesis, calcium induced signaling and DNA repair. CYP2U1, HPGD, CXCR4, ANXA2, EGFR and BRCA1 transcripts were measured over a 72 h time course in LNCaP cells after treatment with 5 nM T and 100 nM 1,25(OH)₂D₃. Fold changes greater than 0.8 are statistically significant; values within the shaded areas in each graph are not significantly modulated (0.8 or below after transformation). Comparisons between different treatment groups were analyzed using one-way ANOVA; differences were considered significant if $p < 0.05$ (*), NS: not significant. Significant changes ($p < 0.05$) in at least two out of three time points were required for the changes to be considered biologically relevant. Note: Scales on the ordinate axis vary from transcript to transcript.

Additional file 6: Differentially regulated mRNAs by T and 1,25(OH)₂D₃ in LNCaP cells after 48 h of treatment. Down regulated mRNAs are shaded in green and up regulated mRNAs are shaded in red. The presence (1) or absence (0) of hormone response elements are indicated based on the *in silico* searches for androgen response elements (AREs) and/or vitamin D response elements (VDREs) in the promoters of T and 1,25(OH)₂D₃ responsive genes identified from microarray analysis.

Additional file 7: miRNA targeted that are differentially regulated by T and 1,25(OH)₂D₃ in LNCaP cells after 48 h of treatment. Down regulated mRNAs/miRNAs are shaded in green and up regulated mRNAs/miRNAs are shaded in red.

Additional file 8: Effects of T and 1,25(OH)₂D₃ on the expression of miR-125b and miR-221 in LNCaP cells. LNCaP cells were treated with 5 nM T and 100 nM 1,25(OH)₂D₃ alone and in combination and transcript levels were measured by TaqMan[®] qPCR over a 72 h time course. Other details as shown in Figure S4.

Additional file 9: Sequence of Primers used for qPCR.

Acknowledgements

The research presented in this manuscript was supported by an operating grant from the USPHS (RO1 CA101114-04). The authors would like to thank David Frank and Marcy Kuentzel for excellent technical help with the microarray experiments; and Dr. Andrew Reilly for his help and insights into the analysis of the microarray data.

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Authors' contributions

WWW was responsible for execution of the experiments described in this manuscript and the analysis of the data. She also wrote the first draft of the manuscript. NC assisted with the analysis of the microarray data and SVC performed the microarray experiments and the initial bioinformatic analyses. JW was responsible for experimental design and editing the manuscript. MT was responsible for experimental design and editing the manuscript. All authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

Received: 8 March 2011 Accepted: 18 May 2011 Published: 18 May 2011

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doi:10.1186/1476-4598-10-58

Cite this article as: Wang *et al*: Effects of 1 α ,25 dihydroxyvitamin D3 and testosterone on miRNA and mRNA expression in LNCaP cells. *Molecular Cancer* 2011 **10**:58.