

1 Full Title

2 Effects of an urban sanitation intervention on childhood enteric infection and diarrhea in Maputo,
3 Mozambique: a controlled before-and-after trial

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Abstract

We conducted a controlled before-and-after trial to evaluate the impact of an onsite urban sanitation intervention on the prevalence of enteric infection, soil transmitted helminth re-infection, and diarrhea among children in Maputo, Mozambique. A non-governmental organization replaced existing poor-quality latrines with pour-flush toilets with septic tanks serving household clusters. We enrolled children aged 1-48 months at baseline and measured outcomes before and 12 and 24 months after the intervention, with concurrent measurement among children in a comparable control arm. Despite nearly exclusive use, we found no evidence that intervention affected the prevalence of any measured outcome after 12 or 24 months of exposure. Among children born into study sites after intervention, we observed a reduced prevalence of *Trichuris* and *Shigella* infection relative to the same age group at baseline (<2 years old). Protection from birth may be important to reduce exposure to and infection with enteric pathogens in this setting.

Introduction

Rapid urbanization has led to the expansion of informal settlements in many low- and middle-income countries (LMICs). Such settlements often have very limited sanitation infrastructure (UN-Habitat, 2016). Separation of human waste from human contact can prevent exposure to enteric pathogens that cause infection, diarrhea (Liu et al., 2016), and potentially long-term health effects such as environmental enteric dysfunction (EED) (Kosek et al., 2017), linear growth deficits (Rogawski et al., 2018), impaired cognitive development (MAL-ED Network Investigators, 2018), and reduced oral vaccine immunogenicity (Parker et al., 2018). Children living in densely populated slum areas where fecal contamination is pervasive and sanitation

infrastructure is limited may be at an increased risk of adverse health effects due to frequent exposure to enteric pathogens (Ezeh et al., 2017; Fink, Günther, & Hill, 2014).

Household-level sewerage has demonstrated health benefits (Barreto et al., 2010, 2007; Norman, Pedley, & Takkouche, 2010) and remains an important long-term goal for many urban settings despite limited evidence from controlled trials (Norman et al., 2010; Wolf et al., 2018). Such systems may not be feasible short-term solutions due to cost, space, and logistical constraints, challenges that have also impeded their evaluation via randomized trials (Norman et al., 2010). Further, in densely populated areas, there may not be space for household-level sanitation of any type. Shared sanitation is a subject of considerable debate but may represent the only near-term sanitation option in some settings (Evans et al., 2017; Heijnen et al., 2014; Tidwell et al., 2020). Yet, while shared, onsite systems may fill the growing need for safe sanitation in rapidly expanding urban areas in LMICs, to date, there has been little evidence of their health impacts in these settings. Recent large-scale, rigorous evaluations of onsite sanitation interventions and combined water, sanitation, and hygiene interventions have demonstrated mixed effects on health (Clasen et al., 2014; Humphrey et al., 2019; Luby et al., 2018; Null et al., 2018; Patil et al., 2014; Pickering, Djebbari, Lopez, Coulibaly, & Alzua, 2015) but all were conducted in rural areas with household-level interventions, and their findings may have limited generalizability to urban areas. A recent meta-analysis estimated that non-sewered interventions reduced the risk of self-reported diarrhea by 16% but did not estimate effects on objective health outcomes, such as enteric infection (Brown & Cumming, 2019), and could not stratify estimates by rural versus urban setting given the lack of evidence in urban areas (Wolf et al., 2018). To-date, no controlled

trials of urban onsite sanitation have been conducted despite over 740 million urban residents relying on such technologies (Berendes, Sumner, & Brown, 2017).

The Maputo Sanitation (MapSan) trial was the first controlled trial to evaluate an onsite, shared sanitation intervention in an urban setting and the first to use the prevalence of enteric infection, as detected by molecular methods, as the primary study outcome (Brown et al., 2015). The study was located in densely populated, low-income, informal neighborhoods of Maputo, Mozambique where the sanitary conditions are poor and disease burden high (Knee et al., 2018). As of 2017, only half of urban residents in Mozambique had access to at least basic sanitation infrastructure, 3% had access to sewerage, and 9% shared sanitation with multiple households, often in poor neighborhoods where space and resources are limited (UNICEF/WHO, 2019). We investigated whether an engineered, onsite, shared sanitation intervention could reduce enteric infection and diarrhea in young children living in these low-income, densely populated neighborhoods in Maputo, Mozambique.

Results

The MapSan trial was a controlled before-and-after trial designed to evaluate the impact of an onsite sanitation intervention on child health after 12 and 24 months of follow-up. The intervention consisted of pour-flush toilets to septic tanks with soakaway pits to discharge the liquid portion of the waste. A non-governmental organization (NGO) delivered the intervention to clusters of households known as compounds, replacing the existing poor-condition shared facilities. Control compounds did not receive the intervention and continued to use their poor-condition sanitation for the duration of the study. We assessed several measures of child health, including enteric infection measured via stool-based molecular methods, soil-transmitted

helminth (STH) re-infection measured via Kato-Katz, and diarrhea measured via caregiver report in both intervention and control children during three phases: baseline (pre-intervention), 12-month follow-up, and 24-month follow-up. Children were eligible for baseline enrollment if they were less than four years old (1-48 months old). At follow-up, children were eligible for enrollment if they were less than four years old or if they would have been less than four years old during baseline.

We enrolled 987 children in 495 compounds during the baseline phase (February 2015 – February 2016) and collected stool samples (whole stool or diaper samples containing liquid diarrhea) from 765 children (78%) (Figure 1). During the 12-month follow-up phase (March 2016 – April 2017), we enrolled or revisited 939 children in 438 compounds and collected 805 stool samples (86%). During the 24-month follow-up phase (April 2017 – August 2018), we enrolled or revisited 1001 children in 408 compounds and collected stool samples from 922 (90%). To improve the success rate of stool sample collection during the 12- and 24-month follow-up visits, we collected rectal swabs from children who did not provide a whole stool sample after multiple collection attempts. The proportion of each type of sample (whole stool, diaper sample, and rectal swab) was similar between arms at each phase (Supplemental Figure 1). Fewer than 5% of all samples were diapers and approximately 7% of 12-month samples and 25% of 24-month samples were rectal swabs (Supplemental Table 1). The NGO delivered interventions to 15 control compounds after baseline and children in those compounds were censored at the time of intervention receipt (Figure 1). Children living in control compounds that independently upgraded their latrines were included in the main analyses. However, as inclusion of these control children may have diluted the intervention effect, they were excluded from

sensitivity analyses designed to understand the impact of the intervention when compared with controls with poor-condition sanitation throughout the study. Children in intervention and control compounds were enrolled at similar rates during each phase (Supplemental Figure 2). Due to migration out of the compound, we collected longitudinal data from 62% of children (59% controls, 67% interventions) between baseline and 12-month and 51% of children (46% controls, 58% interventions) between baseline and 24-month.

At baseline enrollment, intervention compounds had more residents, households, and on-premise water taps than controls, though the number of shared latrines was similar (Table 1). Animals were observed in over half of all compounds. Intervention and control households had similar wealth scores, though intervention households had more members and were more crowded while control households more often had walls made of sturdy materials. All households used a municipal water tap as their primary drinking water source with 78% reporting use of a tap on the compound grounds. At baseline, latrines used by intervention households more often had pedestals or slabs, drop-hole covers, and sturdy walls compared with controls. Consistent with previous estimates in urban Maputo (Satterthwaite, Beard, Mitlin, & Du, 2019), open defecation was rare in our study population with only one control household reporting open defecation at baseline. Baseline characteristics of intervention and control children were similar: the average age at enrollment was 23 months (SD = 13), 51% were female, and 32% were still breastfeeding (Table 1). The age distributions of intervention and control children were similar at baseline and both follow-up phases (Supplemental Figure 3).

We used the Luminex Gastrointestinal Pathogen Panel (GPP), a qualitative multiplex molecular assay, to simultaneously test for 15 enteric pathogens in stool samples, including nine bacteria,

three protozoa, and three viruses. We detected ≥ 1 bacterial or protozoan enteric infection, our pre-defined primary outcome, in 78% (591/753) of children with stools available at baseline. We measured our pre-defined secondary outcome, ≥ 1 STH re-infection, using the Kato-Katz microscope method and detected ≥ 1 STH in 45% (308/698) of stools at baseline. The prevalences of pre-defined outcomes, individual pathogens, and pathogen types were similar between the intervention and control arms at baseline (Table 2). The prevalence of most bacterial, protozoan, and STH infections increased with age while the prevalence of enteric viruses decreased with age (Supplemental Table 2 and Supplemental Figure 4).

The characteristics of children with repeated observations (including baseline) were similar to characteristics of children measured at baseline only and to characteristics of children measured at 12-month and/or 24-month only with the exception of age-related characteristics (Supplemental Tables 3-6). Over half of the children enrolled after baseline were born into study sites (336/622 [54%], Figure 1).

Our main analyses included observations from all eligible children enrolled at baseline (mean sampling age 664 days, SD=393) and the 12-month (940 days, SD=498) and 24-month (1137 days, SD=603) follow-up visits (Table 2). We used a difference-in-difference (DID) analysis to estimate the intervention effect and adjust for baseline differences between intervention and control compounds. We present effect estimates from the DID analyses as prevalence ratios (ratio of ratios). To assess the validity of the parallel trend assumption, a key assumption of DID analyses, we ran “placebo tests” by replacing outcomes with variables unrelated to the intervention, such as child age, respondent role, and presence of animals. Placebo tests showed no effect of the intervention on these variables, suggesting the parallel trend assumption was

valid. We found no evidence that the intervention had an effect on the prevalence of any bacterial or protozoan infection (adjusted PR 1.04, 95% CI [0.94 – 1.15]), or any STH re-infection (1.11 [0.89 – 1.38]) 12 months after implementation (Table 2) despite household respondents reporting almost exclusive use of the intervention latrine (97%, 404/417). The prevalence of diarrhea remained fairly constant in both arms in all three phases with the exception of the 12-month measure in the control arm which was lower, resulting in a larger effect estimate with low precision (1.69 [0.89-3.21]).

The intervention had no meaningful effect at 12 months on the prevalence of infection with any of the three pathogen types measured by the GPP (bacterial, protozoan, viral), pathogen coinfection, or on any individual pathogen (Table 2). There was poor precision in the effect estimates for infrequently detected pathogens, evident from their wide confidence intervals. Therefore, some estimates suggestive of a large protective or detrimental effect (*Campylobacter*, *C. difficile*, *E. coli* O157, STEC, Norovirus GI/GII, Adenovirus 40/41) may have arisen by chance. While the National Deworming Campaign (NDC) provided albendazole to all compound members following baseline, during 12-month visitation only 58% of caregivers (56% control, 60% intervention) confirmed that their child was dewormed during these visits. A sensitivity analysis restricted to children confirmed to have been dewormed produced similar results to the main analysis (Supplemental Table 7). By the 12-month visit, 19 control compounds (19/240 [8.0%]) had independently upgraded their facilities to pour-flush toilets. Results from sensitivity analyses excluding children living in control compounds with independently upgraded facilities were consistent with the main results (Supplemental Table 8).

There was no evidence that the intervention had an effect on the prevalence of any bacterial or protozoan infection, any STH re-infection, or diarrhea after 24 months among all enrolled children (Table 2). We also found limited evidence of effect on the prevalence of any pathogen type or coinfection with ≥ 2 GPP pathogens 24 months after intervention. Results for several individual outcomes were suggestive of a protective (STEC, *E. coli* O157, *Cryptosporidium*, STH coinfection) or adverse (*Campylobacter*, *C. difficile*) effect, but evidence was weak as estimates were accompanied by wide confidence intervals and chance discoveries were possible given multiple comparisons. At the 24 month visits, caregivers confirmed baseline and/or 12-month deworming more frequently for intervention children (339/502 [68%]) than for control children (286/499 [57%]). Adjustment for deworming status or time since deworming had no impact on effect estimates (Supplemental Table 7). Excluding children from control compounds which independently upgraded their facilities by the 24-month visit (35/211 compounds, [17%]) did not impact the results (Supplemental Table 8).

Point estimates of effect and associated confidence intervals were largely similar in unadjusted and adjusted models with few exceptions (e.g. ETEC at 24-month) (Table 2). Multivariable models for GPP outcomes and STH outcomes were adjusted for covariates selected *a priori* (child age, sex, caregiver education, and household wealth index). No other variables met our inclusion criteria for multivariable models, which included being imbalanced between intervention and control at baseline and meaningfully changing 12-month effect estimates ($>10\%$ change in prevalence ratios) (Supplemental Table 9). While the relationship between age and pathogen prevalence appeared to be non-linear for many pathogens (Supplemental Figure 4), the inclusion of a higher order age term (age squared) did not meaningfully change effect estimates

in the main or sub-group analyses (Supplemental Table 10). Three measures of seasonality were considered for inclusion in multivariable models to adjust for any difference in seasonal distributions of data collection: (1) a binary variable defining the ‘rainy’ (November – April) and ‘dry’ seasons (May – October) in Maputo, (2) a measure of cumulative rainfall (mm) in the 30 days prior to data collection, and (3) sine and cosine terms representing dates of sample collection. While there was some imbalance between arms in data collected during the wet and dry seasons at baseline (Supplemental Table 9), no measure of seasonality meaningfully changed effect estimates in the 12- and 24-month analyses and seasonality was excluded from final multivariable models (Supplemental Table 9 and 11). For diarrhea, two variables in addition to variables selected *a priori* met our inclusion criteria and were included in adjusted models: presence of a latrine drop-hole cover at baseline and reported use of a water tap located within the compound grounds at baseline (Supplemental Table 9). The magnitude of effect estimates were larger and confidence intervals wider for diarrhea in adjusted versus unadjusted models in the 12-month and 24-month analyses (Table 2). In addition to the main analyses which included all enrolled children, we also performed two sub-group analyses. The first included children who were born after the intervention was implemented (or after baseline in control compounds) and present at the 12- and/or 24-month follow-up visit. This analysis allowed us to evaluate the impact of the intervention on young children who were never exposed to the poor sanitation at baseline. The second sub-group analysis included only children with repeated measures at baseline and 12- and/or 24-month follow-up.

In sub-group analyses comparing children born into study compounds before the 24-month visit with children of similar ages at baseline (<2 years old), there was suggestive evidence that the

intervention reduced the prevalence of infection with any STH by half (n=522; adjusted prevalence ratio 0.51, [95% CI 0.27 - 0.95]), *Trichuris* by 76% (n=522; 0.24, [0.10 - 0.60]), and *Shigella* by 51% (n=630; 0.49, [0.28 - 0.85]) (Table 3). These effects were attenuated in sub-group analyses restricted to older children (>24 months) who were born before the intervention was implemented and present at the 24-month phase (Supplemental Table 12). We did not observe intervention effects among children born into the study by the 12-month visit, but the sample size was small, resulting in high uncertainty in effect estimates (Supplemental Table 13).

Longitudinal sub-group analyses explored the effect of the intervention on children with repeated measures at baseline and 12-month (for unadjusted analyses: n=870 data points [435 children with repeat measures] for GPP outcomes, n=572 [286] for Kato-Katz outcomes, and n=1112 [556] for diarrhea) and at baseline and 24-month (n=716 (358), n=402 (201), n=834 (417)). Effect estimates were consistent with results from the main analyses (Supplemental Tables 14-15) but less precise due to the reduced sample numbers.

Discussion

We found no evidence that this urban, onsite shared sanitation intervention was protective against our pre-specified child health outcomes of enteric infection, STH re-infection, or diarrhea. We also found no strong evidence that the intervention affected prevalence of any individual pathogen, pathogen type, or coinfection with ≥ 2 enteric pathogens or STH. In exploratory sub-group analyses, we found suggestive evidence that the intervention reduced the prevalence of any STH, *Trichuris*, and *Shigella* infections among children born into the study by the 24-month follow-up visit. Studying children born into intervention sites after implementation allowed us to examine the effect of the intervention from birth through the first two years of life. These results suggest that the intervention delayed pathogen exposure and the accumulation of enteric infections during early childhood, but it needs to be treated with caution as this was an exploratory subgroup analysis.

The trial was neither designed nor powered to detect differences in sub-groups of children such as those born after the intervention was implemented, potentially limiting our ability to detect small effects in such analyses. Further, all exploratory sub-group analyses included multiple comparisons, increasing the likelihood of chance discoveries. However, the magnitude of the effect estimates for the outcomes of any STH, *Trichuris*, and *Shigella* observed among children born into the study by the 24-month visit, and the directional consistency of effect estimates among most other outcomes in this sub-group analysis, strengthens the plausibility of these findings.

There are several reasons we observed suggestive evidence of an effect for some outcomes among this sub-group of young children but not among older children or in the main analyses.

Children's exposures vary by age, particularly as they become mobile and begin independent exploration of their environment. It is possible that the intervention reduced exposure via pathways that are important for very young children but may represent just minor pathways of exposure among older children (Kwong et al., 2020). Additionally, young children may experience fewer exposures outside of the compound. Reductions in exposure and subsequent infection early in life may delay or prevent the development of environmental enteric dysfunction (EED), a subclinical condition that affects the structure and function of the gut and may increase susceptibility to future infection (Keusch et al., 2014; Prendergast & Kelly, 2016). Results from the EED sub-study of the WASH Benefits cluster randomized controlled trial (cRCT) in Bangladesh suggest that the intervention delayed but did not prevent the onset of EED (Lin et al., 2019). If this intervention similarly delayed the development of EED among children born into intervention sites, they may have been less susceptible to infection than children of a similar age at baseline. Finally, some pathogens, like *Giardia* and certain STH, can cause persistent infections that can remain active for months or years if not treated (Else et al., 2020; Rogawski et al., 2017). The intervention would have no effect on such infections, highlighting the potentially important role of protection from birth.

Notably, both *Shigella* and *Trichuris* are primarily anthroponotic, and infection was strongly age-dependent in this study population (Knee et al., 2018). These factors may help explain the differing intervention effects observed both among pathogens and age groups. The intervention was unlikely to limit exposure to animal feces, reducing the likelihood that it would impact infection prevalence of zoonotic pathogens like *Campylobacter* or *Giardia*. The strong positive associations between age and prevalence for *Shigella* and *Trichuris* suggest that exposure

increases with age. This supports the hypothesis that the intervention may have reduced the overall frequency or intensity of exposure enough to impact *Shigella* and *Trichuris* infection among young children but not older children.

Rapid urbanization is expanding informal settlements and out-pacing the expansion of sanitation services in many cities, widening the gap in sanitation access between the urban rich and poor (UNICEF/WHO, 2019). To our knowledge, MapSan was the first trial to estimate the health impact of an urban, onsite shared sanitation intervention and the first to use enteric infection as the primary trial outcome. Most of the urban sanitation literature published to date has evaluated the expansion of sewerage, an important and ambitious goal that is out of reach for many cities in the near-term (Norman et al., 2010). Access to sewerage is associated with a 30-60% reduction of diarrheal disease depending on starting conditions, and an approximately 30% reduction in enteric parasite detection, though most studies are observational and few controlled trials exist (Barreto et al., 2010; Norman et al., 2010; Wolf et al., 2018).

Most studies of onsite sanitation interventions have occurred in rural areas. Despite good evidence that onsite sanitation is associated with reductions in diarrheal disease (M. C. Freeman et al., 2017; Wolf et al., 2018), several recent rural trials of basic sanitation and combined WASH interventions with good uptake and use reported mixed effects on child health outcomes including diarrhea, linear growth, and more recently, enteric infection (Ercumen et al., 2019; Grembi et al., 2020; Humphrey et al., 2019; Lin et al., 2018; Luby et al., 2018; Null et al., 2018; Pickering et al., 2019; Rogawski McQuade, Platts-Mills, et al., 2020).

The Sanitation, Hygiene, Infant Nutrition Efficacy (SHINE) trial in rural Zimbabwe found no impact of a combined WASH intervention on diarrhea, growth, or the prevalence of a suite of

enteric pathogens among children aged <12 months old but did report a small reduction in the number of parasitic pathogens detected.(Humphrey et al., 2019; Rogawski McQuade, Platts-Mills, et al., 2020)

While the WASH Benefits Bangladesh cRCT reported no effect of any WASH intervention on child growth, the sanitation, hygiene, and combined WASH study arms reduced the prevalence of diarrheal disease from 5.7% to 3.5% (Luby et al., 2018), accompanied by absolute reductions in *Giardia* prevalence of 6-9% among children aged 2-3 years in the same arms (Lin et al., 2018). The sanitation arm also reduced the prevalence of *T. trichiura* among children 2-3 years old (from 5.2% to 3.2%) but had no impact on *A. lumbricoides* or hookworm, the only other parasites detected frequently enough to estimate effects in that study (Ercumen et al., 2019). In a parallel analysis, only the water treatment and combined WASH interventions of the WASH Benefits Kenya cRCT reduced the prevalence *A. lumbricoides*, suggesting that the reduction in prevalence in the combined WASH arm may be attributable to the water treatment intervention (Pickering et al., 2019). The sanitation-only arm had no impact on any parasite measured, though *T. trichiura* was too infrequently detected to estimate effects (Pickering et al., 2019). An evaluation of a comprehensive suite of 34 enteric pathogens reported reduced prevalence and quantity of enteric viruses, but not bacteria or parasites, among children aged 14 months old in the combined WASH arms in the Bangladesh trial (Grembi et al., 2020). Together with our findings, these results suggest that sanitation and combined WASH interventions can reduce the prevalence of enteric infection in some settings but that effects may vary by pathogen, child age, intervention, and setting.

We previously published two baseline risk factor analyses to identify demographic, environmental, and WASH-related predictors of infection and environmental fecal contamination in our study setting prior to the intervention implementation (Holcomb et al., 2020; Knee et al., 2018). Age was an important predictor of infection, though the direction of its effect varied by pathogen type. Increasing age was associated with increased risk of bacterial and protozoan infections and decreased risk of viral infections (Knee et al., 2018). Other socio-demographic predictors of infection included breastfeeding, which was associated with a decreased risk of any infection (driven by its strong association with protozoan infection), and female sex which was associated with an increased risk of viral infection. Few sanitation-related or environmental variables were associated with infection at baseline and the magnitude of associations were often small. The presence of a latrine superstructure and drop-hole cover were associated with small reductions in risk of bacterial or protozoan infection, often only in unadjusted analyses, but other latrine features (e.g. presence of a cleanable slab) were not. The observation of feces or used diapers around the compound grounds was associated with increased risk of bacterial and protozoan infection but most other environmental and sanitary hazards were not (Knee et al., 2018).

Fecal contamination was common among all environmental reservoirs tested (water, soil, food preparation surfaces) at baseline. We detected one or more microbial markers of contamination in over 95% of environmental samples (Holcomb et al., 2020). *E. coli* was the most frequently detected and abundant marker of contamination among all sample types, and human-associated markers were most frequently detected in soil (59%) and stored drinking water (17%) samples. Measures of latrine quality that were associated with small reductions in infection risk (e.g. drop-

hole covers, latrine superstructures) were not associated with decreased odds of fecal contamination in this setting. Overall, we found few consistent relationships between markers of fecal contamination and environmental, WASH-related, and demographic characteristics at baseline (Holcomb et al., 2020).

While these results suggest WASH-related and environmental risk factors may be poor determinants of child health in this setting, the lack of heterogeneity in WASH conditions at baseline, given the selection criterion that compounds must share sanitation in “poor condition,” may have limited our ability to identify strong WASH-related predictors of infection or environmental fecal contamination. Results from a forthcoming companion study suggests the intervention had mixed effects on environmental fecal contamination. The intervention may have reduced the concentration of *E. coli* by an order of magnitude in soil collected from latrine entrances after 12 months, however, there was no effect on the prevalence or concentration of indicators of fecal contamination in any other environmental compartment sampled at that time (Holcomb et al., 2021). It is unlikely that the observed reductions in fecal contamination in soils alone would be sufficient to impact health outcomes in this setting. Other studies that have evaluated the impact of sanitation interventions on fecal contamination of the surrounding environment have found limited evidence of effect (Clasen et al., 2014; Ercumen, Mertens, et al., 2018; Ercumen, Pickering, et al., 2018; Fuhrmeister et al., 2020; Patil et al., 2014; Pickering et al., 2015; Gloria D. Sclar et al., 2016; Steinbaum et al., 2019).

In this setting, where fecal contamination was pervasive and burden of infection high, even considerable reductions in contamination and exposure may have been insufficient to realize measurable health gains as the intervention did not address all potential transmission pathways

(Briscoe, 1984; Julian, 2016; Robb et al., 2017). For example, the intervention did not address child feces disposal practices or handwashing behaviors and it is unlikely that the intervention infrastructure would have changed these (Majorin, Torondel, Chan, & Clasen, 2019). Previous studies of sanitation interventions have found no reduction in hand contamination (Ercumen, Pickering, et al., 2018), which has been associated with increased incident diarrheal disease in young children (Pickering et al., 2018). The intervention may not have reduced exposure via consumption of contaminated food – particularly foods contaminated prior to arrival in the compound – likely an important source of enteric pathogen transmission in some settings (Julian, 2016; Kwong et al., 2020). Children’s exposure to animal feces has been documented in rural, peri-urban, and urban settings and could be an important, unmitigated source of exposure to enteric pathogens in both intervention and control arms where animals were frequently observed (Delahoy et al., 2018; Kwong et al., 2020; Penakalapati et al., 2017). Observation of animals in compounds was examined as a potential confounder but did not change effect estimates.

The intervention was delivered at the compound level, not the community level, and was not designed to achieve any specified threshold of sanitation coverage in the study neighborhoods. Previous studies have suggested that achieving a certain level of community sanitation coverage may be necessary to reduce disease burdens (Barreto et al., 2007; Fuller & Eisenberg, 2016; Fuller, Villamor, Cevallos, Trostle, & Eisenberg, 2016; Harris, Alzua, Osbert, & Pickering, 2017; Jung, Lou, & Cheng, 2017; Spears, Ghosh, & Cumming, 2013; Wolf et al., 2018). For example, a study of a large-scale sewerage expansion in urban Brazil found that the intervention reduced diarrheal disease by 22%, with neighborhood coverage level being the single most important explanatory variable (Barreto et al., 2007). We did not measure neighborhood-level

sanitation coverage, but previous estimates show that while coverage is high and open defecation is limited (1%), only 9% of sanitation systems are safely managed (Satterthwaite et al., 2019). Further, in the Nhlamankulu district where many of our study sites are located, the majority of households (56%) rely on pit latrines serving individual households, most of which are in poor condition (Devamani, Norman, & Schmidt, 2014; Satterthwaite et al., 2019). Together with our results, this suggests that both the extent and quality of community coverage are likely important to reducing overall transmission. Sanitation coverage and quality may be especially important in urban areas given the proximity of compounds and the opportunity for person-to-person contact, neighborhood-level exposure, and for external sources of contamination (e.g. a neighbor's flooded pit latrine) to influence compound-level exposures (Barreto et al., 2007). We did not measure neighborhood-level exposures, which may be important for young children in slum settings (Ezeh et al., 2017; Medgyesi et al., 2019), and their impact on our health outcomes is unclear. In addition to neighborhood-level exposures, the transience of the study population meant that trips to and from provinces outside of Maputo, where exposures were varied and unmeasured, were common.

It is unlikely that our findings are due to poor intervention fidelity or use, a challenge encountered in some trials of rural sanitation interventions (Clasen et al., 2014; Patil et al., 2014). The use of the intervention required minimal behavior change as compound members switched from using their existing latrine in poor condition, which was removed following construction of the intervention latrine, to using the new hygienic latrine. The results of a forthcoming process evaluation demonstrate that 96% of intervention latrines were well-maintained two or more years after construction, suggesting continued use by compound members (Bick et al., 2021). Further,

only 3% of intervention compounds (8/270) had a secondary, non-intervention latrine in use after two or more years, indicating that members of most intervention compounds exclusively used the intervention latrines (Bick et al., 2021). It is possible that development in the study neighborhoods, including changes to sanitation facilities in control compounds, contributed to the limited effect of the intervention. However, results from sensitivity analyses that excluded control compounds with upgraded sanitation were consistent with results from the main analyses.

The two intervention designs we evaluated in this study – communal sanitation blocks and shared latrines – utilized the same basic sanitation technology but differed in the number of cabins and amenities available. While it is possible that this heterogeneity in design may have modified the effect of the intervention, this study was not powered to test this. Moreover, all intervention compounds were encouraged to independently upgrade their facilities by adding features like electricity and handwashing stations, or by connecting existing handwashing stations to the water supply, resulting in heterogeneity even within the two broad categories of intervention type.

While the NDC dewormed every study compound annually during the study period, it is possible that not all study participants received, or took, the medication and that the time between deworming and subsequent measurement of STH re-infection varied among children. Additionally, single-dose albendazole can have limited effectiveness against certain STH, notably *Trichuris* (Moser, Schindler, & Keiser, 2017). Inadequate or ineffective deworming could have limited our ability to detect an effect on STH outcomes. Sensitivity analyses adjusting for caregiver-confirmed deworming and for estimated time between deworming and re-infection measurement produced similar results to the main analysis.

There are several important limitations of this study. As the intervention was pre-planned and not implemented by the study team, we could not randomize its allocation, increasing the risk of confounding. We assessed potential confounding variables at baseline and used a DID analysis, which accounts for baseline outcome measures, to limit the effect of unmeasured, residual confounding. While we attempted to enroll intervention and control compounds with comparable numbers of residents, the NGO which identified and implemented the intervention selected most of the largest eligible compounds for intervention. This resulted in intervention compounds having a slightly higher mean number of residents than control compounds (Table 1). Crowding has been identified as a risk factor for pathogen transmission and poor health outcomes in other studies, (Halpenny, Koski, Valdés, & Scott, 2012; Rahman, Wojtyniak, Mujibur Rahaman, & Aziz, 1985; Rogawski McQuade, Shaheen, et al., 2020) though we found limited evidence of this in our study population at baseline (Knee et al., 2018). Further, we assessed the number of compound residents as a potential confounder but found that it did not meaningfully change the DID estimates for our pre-defined outcomes (Supplemental Table 9). We consider our analysis to be robust to small differences in study arms at baseline, however, we cannot exclude the possibility of residual confounding due to such differences, a limitation of non-randomized designs.

It was not possible to mask participants to their intervention status, and our measure of caregiver-reported diarrhea could be subject to respondent and recall biases. To reduce the risk of respondent bias, the MapSan field enumerator team and implementation team were different, and respondents were not informed explicitly that the MapSan team was evaluating the health effect of the intervention. To limit recall bias, we used a 7-day recall period (Arnold et al., 2013). Our

other pre-specified outcomes were objective measures of pathogen infection and not subject to the same biases (Brown & Cumming, 2019).

Due to the greater than expected losses to follow-up in both study arms, we were not able to follow all children enrolled at baseline through time as expected, but we still achieved our target enrollment numbers due to migration and births into study compounds. We conducted the originally planned longitudinal analysis as a sub-group analysis. It also served as a sensitivity analysis to estimate the impact of migration on our effect estimates. Results from this sub-group analysis were largely similar to results of the main analysis which treated measures as repeated cross-sections, though the reduction in sample size led to wider confidence intervals (Supplemental Tables 14-15). Measures of outcomes and covariates in children with and without repeated measures were mostly similar, further limiting the likelihood that changes in the study population biased our results.

While molecular detection of enteric pathogens in stool is evidence of pathogen exposure, it is not necessarily evidence of active infection, making its clinical significance less clear (Brown & Cumming, 2019). We assumed pathogen detection by the GPP indicated infection because the assay's limits of detection exceeded the median infectious dose of most pathogens. While the GPP detects many enteric pathogens recognized as important causes of childhood diarrhea in LMICs, (Liu et al., 2016) it does not detect all enteric pathogens of importance. Further, qualitative, cross-sectional analysis of stools does not provide information on the duration or intensity of infection or pathogen carriage. Quantitative results, like those produced by multiplex quantitative PCR panels, can be used to aid identification of etiologic agents of diarrhea, especially in cases of coinfection, and to differentiate between low-level enteric pathogen

detection of unknown clinical relevance and higher concentration shedding which is more clearly associated with disease (Liu et al., 2014, 2016; Platts-Mills, Liu, & Houpt, 2013). Some studies have demonstrated overall good performance of the GPP but observed elevated false positive detection rates for the *Salmonella* targets (Duong et al., 2016; Kellner et al., 2019). For this reason, we removed *Salmonella* results from our pre-specified outcome definition. Results from analyses including and excluding *Salmonella* were similar. In addition, some studies have observed reduced sensitivity or specificity for some GPP targets compared with qPCR-based methods, including norovirus, adenovirus, *Campylobacter*, *Yersinia enterocolitica*, ETEC, and *Salmonella*, though inconsistencies between studies exist and are likely due to differences in comparator assays or sample storage and processing (Chhabra et al., 2017; Deng et al., 2015; Duong et al., 2016; Huang et al., 2016; Zhan et al., 2020; Zhuo et al., 2017). Further, the lack of an adequate reference standard in most comparative studies complicates interpretation (K. Freeman et al., 2017).

Our ability to detect an effect on our primary outcome, the prevalence of ≥ 1 bacterial or protozoan infection, may have been limited by (1) the extended duration of shedding of some pathogens following active infection; (2) the overall high burden of disease in our study population, particularly among older children; and (3) residual confounding by age given the strong observed relationship between age and infection status (particularly for protozoan pathogens), all of which may have biased our results toward the null. Further, the intervention may have impacted the concentration of pathogens shed (Grembi et al., 2020; Lin et al., 2019), but our binary outcome was not sensitive to such differences. The qualitative nature of the GPP did not allow us to interrogate this question.

We analyzed a smaller number of stool samples for STH than for other enteric pathogens due to requirements of the Kato-Katz method used for STH detection. The Kato-Katz method can only be performed on whole, solid stool. Diarrheal samples and rectal swabs, the latter of which were introduced during the 12-month follow-up phase, were not eligible for STH analysis by Kato-Katz. Further, when limited stool material was collected, we prioritized the molecular analysis used for the primary outcome. While the smaller sample size available for the STH analyses may have reduced our ability to detect small effects, the proportions of whole stool, diarrheal diaper samples, and rectal swabs were similar between arms at each phase (Supplemental Table 1). This limited the potential impact that sample type could have on our results.

While the Kato-Katz method performs similarly to other microscope-based and molecular methods for detection of moderate to high intensity infections, it may be less sensitive than molecular methods in detecting low intensity infections (Benjamin-Chung et al., 2020; Cools et al., 2019). A recent study has also suggested reduced specificity of the Kato-Katz method for detection of low-intensity *A. lumbricoides* infections (Benjamin-Chung et al., 2020). In settings where low-intensity infections are common, or where STH may be targeted for elimination, methods with better diagnostic accuracy, like qPCR, may be considered.

We had limited ability to evaluate the impact of seasonality or weather-related trends on our effect estimates due to drought conditions during the 2015/2016 rainy season. We adjusted models for cumulative 30-day rainfall, a binary indicator of wet/dry season, and sine/cosine terms of sample collection date (Stolwijk, Straatman, & Zielhuis, 1999) but excluded all seasonality terms from final multivariable models because they did not meaningfully change effect estimates.

Our results demonstrate that access to hygienic, shared onsite sanitation systems was not sufficient to reduce enteric infection or diarrhea in children aged 6 years or younger (≤ 4 at baseline) 12-24 months after implementation. Results from our sub-group analysis of children born into intervention sites showed a substantial reduction in the prevalence of any STH, *Trichuris*, and *Shigella* infection, suggesting that children may require protection from birth to reduce or delay infection burdens. Our results do not suggest that shared sanitation is inadvisable in this setting, as we did not compare against household-level sanitation improvements, nor do they account for the many non-health related benefits associated with this intervention or upgraded sanitation generally (Caruso et al., 2018; G.D. Sclar et al., 2018; Shiras et al., 2018).

The need for effective sanitation solutions may be most urgent in densely populated, low-income, informal communities like our study setting where ubiquitous fecal contamination drives high infection burdens. Disease transmission in these settings may be driven by multiple interrelated pathways, complicated by frequent migration and the diversity of circulating pathogens, and therefore difficult to interrupt. While decades of research have demonstrated meaningful health gains following sanitation improvements, the results of this study, and other rigorous trials of sanitation interventions, suggest that the relationship between sanitation and health is complex, difficult to measure, and may not be generalizable across diverse settings and populations.

Methods

Study design and intervention

MapSan was a controlled before-and-after trial, and details of the study design and analysis plan have been published previously (Brown et al., 2015). We conducted the study in 17 densely

populated, low-income, informal neighborhoods in Maputo, Mozambique. The intervention was delivered to compounds, typically groups of three to five households (though larger and smaller compounds exist) often delineated by a wall or barrier, that shared sanitation and outdoor living space. Shared compound sanitation facilities are not considered public facilities. We collected data in an open cohort of children in intervention and control compounds at three time-points: baseline (pre-intervention), 12 months post-intervention, and 24 months post-intervention.

The NGO Water and Sanitation for the Urban Poor selected intervention compounds and designed and built 300 intervention facilities - pour-flush toilets discharging to septic tanks, the liquid effluent of which flows to the soil through soakaway pits (Supplemental Figures 5a and 5b). There were two intervention designs with the same basic sanitation technology: communal sanitation blocks (CSBs) and shared latrines (SLs) (Supplemental Figure 6a and 6b). The primary difference between CSBs and SLs was size. CSBs (n=50) included multiple stalls with toilets and served compounds of 21 or more people with one stall allocated per 20 residents. CSBs also included rainwater harvesting systems, a municipal shared water connection, elevated water tanks for storage of municipal water, a handwashing basin, a laundry facility, and a well-drained area for bathing. Shared piped water connections were part of the municipal water system and could be used for drinking in addition to other domestic purposes. Rainwater was intended for cleaning and flushing but not drinking. Shared latrines (n=250) were single-stall facilities serving fewer than 21 people. All septic tanks were sized to require emptying after approximately two years.

Intervention compounds were located in 11 neighborhoods of the Nhlamankulu and KaMaxakeni districts of Maputo (Supplemental Figure 7). The NGO selected intervention compounds using

the following criteria: (1) residents shared sanitation in poor condition as determined by an engineer; (2) the compound was located in the pre-defined implementation neighborhoods; (3) there were no fewer than 12 residents; (4) residents were willing to contribute financially to construction costs; (5) sufficient space was available for construction of the new facility; (6) the compound was accessible for transportation of construction materials and tank-emptying activities; (7) the compound had access to a legal piped water supply; and (8) the groundwater level was deep enough for construction of a septic tank. Intervention compounds were expected to pay approximately 10-15% of the construction costs (~\$64 for shared latrines and ~\$97 for CSBs) within one year of construction, with 25% of the total due upfront. Presence of a child was not a selection criterion and therefore not all intervention sites were included in the study. Opening of newly constructed intervention latrines occurred between February 2015 and February 2016. The study team used criteria 1, 3, 4, and 7 to select control sites that had at least one child younger than 48 months old in residence. We enrolled intervention and control compounds concurrently to limit any differential effects of seasonality or other secular trends on the outcomes (Supplemental Figure 2). Additionally, we attempted to enroll control compounds with similar numbers of residents as intervention compounds. Willingness to pay for facilities among controls was assessed using hypothetical versions of questions posed to interventions. Control compounds were located within the 11 intervention neighborhoods and six adjacent but similar neighborhoods due to the limited availability of eligible compounds remaining within intervention neighborhoods (Supplemental Figure 7). Intervention selection criteria (5), (6), and (8) were not used to select control sites as they were deemed to be related to intervention construction and maintenance and unlikely to influence our outcomes. It was not possible to blind participants or enumerators to intervention status.

598 *Participants*

599 We enrolled eligible children at three time points: baseline (0 months), 12 months post-
600 intervention, and 24 months post-intervention. Children aged 1- 48 months old were eligible for
601 baseline enrollment if we received written informed consent from a parent or guardian and if the
602 head of the compound provided verbal assent for the compound to be included in the study.
603 Children were eligible for enrollment at 12- and 24-month visits if they were aged 1-48 months
604 or if they were eligible for enrollment at baseline but absent during that study visit. Children who
605 moved into the compound fewer than six months before the 12-month or 24-month visit were not
606 eligible for enrollment during that phase given their limited exposure to their new compound.

607 *Ethics*

608 The study protocol was approved by the Comité Nacional de Bioética para a Saúde (CNBS),
609 Ministério da Saúde (333/CNBS/14), the Research Ethics Committee of the London School of
610 Hygiene & Tropical Medicine (reference # 8345), and the Institutional Review Board of the
611 Georgia Institute of Technology (protocol # H15160).

612 *Procedures*

613 Trained field enumerators completed consent procedures and surveys in the participant's
614 preferred language (Portuguese or Changana) and collected biological specimens from enrolled
615 children (Supplemental Methods). At baseline, we aimed to visit intervention compounds two
616 weeks prior to the opening of the new latrines. We scheduled follow-up visits to be 12 months
617 (± 2 weeks) and 24 months (± 2 weeks) from the date compound members began using their new
618 latrines, with visits to control compounds made concurrently (± 2 weeks).

We collected stool samples independently of reported symptomology. If we were unable to collect a stool sample after multiple attempts, a registered nurse collected a rectal swab after obtaining written consent for the procedure from a parent or guardian. Stool samples were kept cold and delivered to the Laboratory of Molecular Parasitology at the Instituto Nacional de Saúde (INS) within six hours of collection for analysis and storage at -80°C.

Samples were shipped frozen with temperatures monitors to the Georgia Institute of Technology (Atlanta, USA) where we used the xTAG Gastrointestinal Pathogen Panel (Luminex Corp, Austin, USA), a qualitative multiplex molecular assay, to detect 15 enteric pathogens in stool samples: *Campylobacter jejuni/coli/lari*; *Clostridium difficile*, toxin A/B; enterotoxigenic *Escherichia coli* (ETEC) LT/ST; Shiga-like toxin producing *E. coli* (STEC) stx1/stx2; *E. coli* O157; *Salmonella*; *Shigella boydii/sonnei/flexneri/dysenteriae*; *Vibrio cholerae*; *Yersinia enterocolitica*; *Giardia lamblia*; *Cryptosporidium parvum/hominis*; *Entamoeba histolytica*; adenovirus 40/41; norovirus GI/GII; and rotavirus. The GPP has been rigorously tested and extensively used for stool-based enteric pathogen detection (Chisenga et al., 2018; Claas, Burnham, Mazzulli, Templeton, & Topin, 2013; Deng et al., 2015; Duong et al., 2016; Huang et al., 2016; Kellner et al., 2019; Khare et al., 2014; Navidad, Griswold, Gradus, & Bhattacharyya, 2013; Patel, Navidad, & Bhattacharyya, 2014). We analyzed samples according to manufacturer instructions with the addition of elution steps for the pretreatment of rectal swabs and diaper material saturated with liquid stool (Supplemental Methods). Technicians at INS assessed stool samples for the presence of soil-transmitted helminths (STH) using the single-slide Kato-Katz microscope method (Vestergaard Frandsen, Lausanne, Switzerland).

Representatives of the National Deworming Campaign (NDC) at the Mozambican Ministério da Saúde (MISAU) offered single-dose albendazole (400 mg, 200 mg for children aged six to 12 months) to all eligible members of intervention and control compounds following sample collection activities of each phase. Eligibility was defined by the NDC and included compound members older than six months who were not pregnant.

Outcomes

For the 12-month analysis, we pre-specified the primary outcome as infection with one or more of the 12 bacterial or protozoan enteric pathogens detected by the GPP and secondary outcomes as re-infection with one or more STH as detected by Kato-Katz (following albendazole treatment at baseline), and seven-day period prevalence of caregiver-reported diarrhea. All three outcomes were considered secondary outcomes in the 24-month analysis. We defined diarrhea as the passage of three or more loose or liquid stools in a 24-hour period or any stool with blood (Arnold et al., 2013; Baqui et al., 1991). We excluded viral enteric pathogens from the primary outcome definition. The intervention may not have interrupted virus transmission due to their low infectious doses, high concentration shed in feces and extended period of shedding, environmental persistence, and capability for direct person-to-person transmission (Julian, 2016). Following reported specificity issues with the *Salmonella* target of the GPP, we removed it from our GPP-based outcome definitions (Duong et al., 2016; Kellner et al., 2019). In addition to the pre-specified outcomes, we evaluated the effect of the intervention on specific pathogen types (bacterial, protozoan, viral) and on individual pathogens. The results for other secondary outcomes listed in the trial registration (growth and environmental enteric dysfunction) will be published separately.

Statistical analysis

Our sample size calculation has been described previously (Brown et al., 2015). We included all enrolled children at each visit and analysed data as repeated cross-sectional observations. We examined the effect of the intervention at the 12-month and 24-month phases separately. We conducted two sets of exploratory sub-group analyses. The first assessed the effect of the intervention on children with repeat observations at baseline and 12-months and at baseline and 24-months visits. These longitudinal analyses also served as sensitivity analyses of the impact of participant migration on effect estimates. The second sub-group analysis compared children who were born into study sites after the intervention (or after baseline in controls) but before the 12-month or 24-month visit with children of a similar age group at baseline. For example, children born after baseline but before the 24-month visit were compared with children aged two years old or younger at baseline. These analyses allowed us to explore whether exposure to the intervention from birth would reduce enteric pathogen infection during the first 1-2 years of life.

We used a DID approach to assess the impact of the intervention on all outcomes at the 12- and 24-month visits. We used generalized estimating equations (GEE) to fit Poisson regression models with robust standard errors. Our GEE models accounted for clustering at the compound level because it was the highest level of nested data and the level of the intervention allocation (Bottomley, Kirby, Lindsay, & Alexander, 2016). We estimated the effect of the intervention as the interaction of variables representing treatment status (intervention versus control) and phase (pre- or post-intervention). Therefore, effect estimates from our DID analysis are presented as ratio measures (ratio of prevalence ratios) instead of absolute differences. Multivariable models were adjusted for covariates determined *a priori* as potentially predictive of our outcomes,

including child age and sex, caregiver's education, and household wealth. Given the important and potentially non-linear relationship between age and pathogen prevalence (Supplemental Figure 4), we also considered inclusion of a higher order age term (age squared) in our models (Supplemental Table 10). Additional covariates (Supplemental Table 9) were considered for inclusion in multivariable models if they were imbalanced between arms at baseline (>0.1 standardized difference in prevalence or mean) and resulted in a meaningful change in the DID effect estimate ($\pm 10\%$ change in 12-month DID prevalence ratio). We assessed the potential impact of seasonality on our results in three ways: (1) inclusion of binary indicator of wet (November – April) and dry (May – October) season in multivariable models, (2) inclusion of a variable representing cumulative rainfall (mm) 30 days prior to sample or survey collection in multivariable models, and (3) inclusion of sine and cosine functions of sample and survey dates in multivariable models (Supplemental Tables 9 and 11). We used the same statistical approach for sub-group analyses. All analyses were performed on complete case data, and a missing data table is presented in the supplemental information (Supplemental Table 16). We performed all statistical analyses with Stata version 16 (StataCorp, College Station, USA).

Registration

The trial was pre-registered at ClinicalTrials.gov (NCT02362932).

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Contributors

JB as principal investigator and OC as co-principal investigator designed the trial and drafted the study protocol with input from WPS, JK, DH, PK, JS, VZ, and RN. JK was the study manager, co-led laboratory work and data analysis with TS, and drafted the manuscript. TS curated the data, designed data collection tools and activities with JK, and produced figures. ZA led field data collection. CA, FB, DC, VC, EM, JMB, CR, WZ helped with sample organization and laboratory analysis. WPS designed the analytical approach and JK, DH, and AM helped refine it. JB and OC secured funding for the trial. All authors contributed critically to the final version of the manuscript.

Data sharing

De-identified participant data which underlie the results reported in this manuscript is publicly available on the MapSan trial Open Science Forum website (<https://osf.io/p5shk>). The published trial protocol can be accessed at: <https://bmjopen.bmj.com/content/5/6/e008215>.

Competing interests

All authors have completed the ICMJE uniform disclosure form at www.icmje.org/coi_disclosure.pdf and declare: no support from any organization for the submitted work; no financial relationships with any organizations that might have an interest in the submitted work in the previous three years; no other relationships or activities that could appear to have influenced the submitted work.

Source data for Figures and Tables

Figure 1 source data 1 and source code 1

748 Table 1 source data 1 and source code 1

749 Table 2 source data 1 and source code 1

750 Table 3 source data 1 and source code 1

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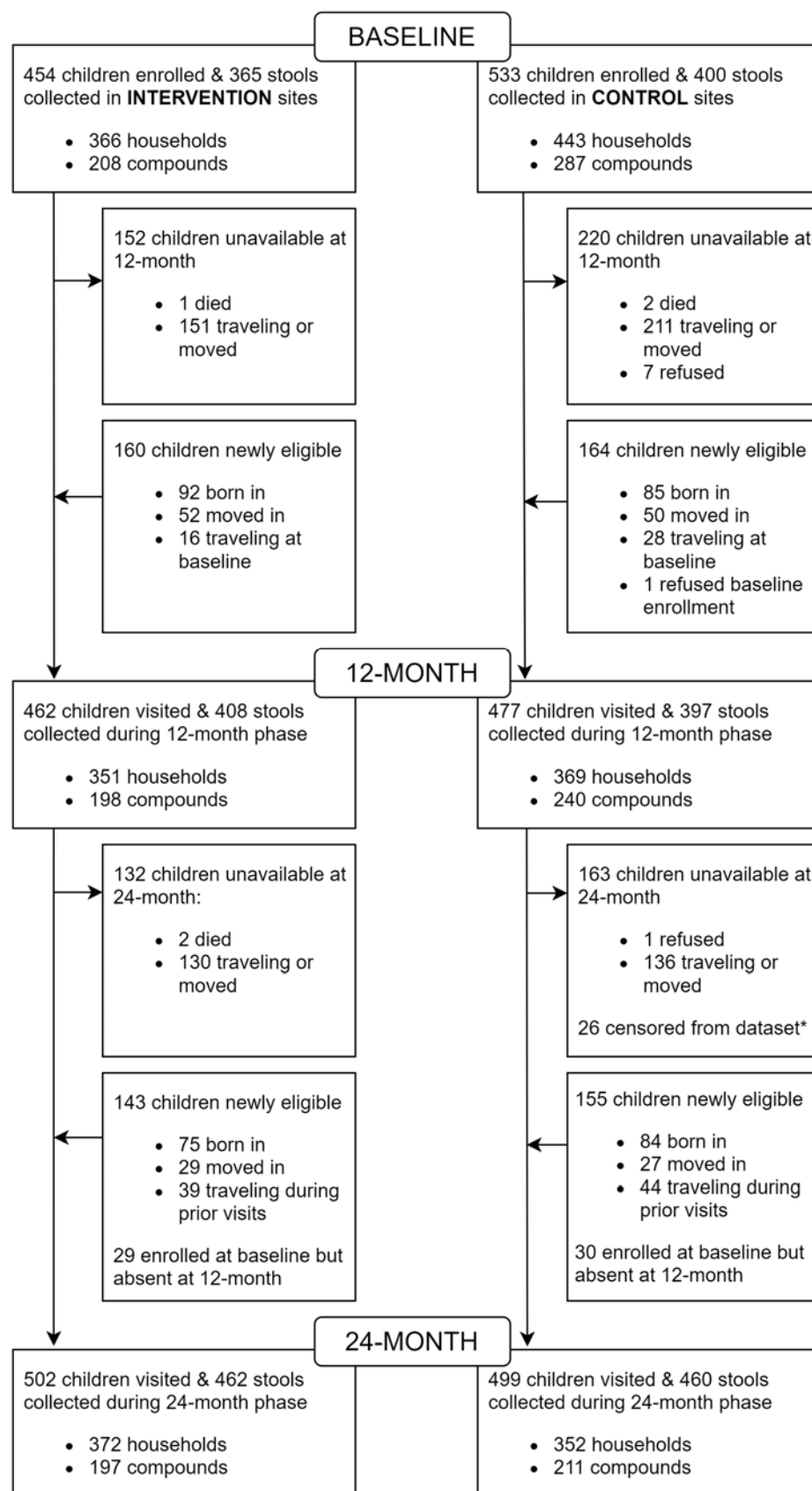
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1103 *Virology*, 96(May), 94–98. <https://doi.org/10.1016/j.jcv.2017.10.003>

1104



1106 Figure 1: Trial profile. †Eligible for enrollment at baseline and/or 12-month but traveling at time of visit. ‡Children
 1107 removed from 24-month analysis because their compound received an intervention after completion of the baseline
 1108 phase. Source files available in Figure 1 – source data 1 and Figure 1 – source code 1.

1109

Table 1: Baseline characteristics of enrolled children, households, and compounds

	Control		Intervention	
	N	n (%) or mean (SD)	N	n (%) or mean (SD)
Child level variables				
Age at survey, days [†]	520	700 (405)	441	694 (403)
Sex, female	520	266 (51%)	444	227 (51%)
Child is breastfed with or without complementary feeding	526	169 (32%)	448	143 (32%)
Child is exclusively breastfed	526	49 (9.3%)	448	37 (8.3%)
Child feces reported to be disposed of in a latrine	526	148 (28%)	448	141 (31%)
Child wears diapers	526	342 (65%)	447	294 (66%)
Caregiver completed primary school	528	287 (54%)	451	239 (53%)
Child's mother is alive	513	503 (98%)	435	426 (98%)
Respondent is child's mother	519	368 (71%)	443	284 (64%)
Household level variables				
Household population	441	5.4 (2.4)	365	6.1 (3.0)
Household wealth score, 0 (poorer) - 1 (wealthier) [*]	440	0.45 (0.10)	365	0.44 (0.10)
Household crowding, >3 persons/room	440	54 (12%)	365	60 (17%)
Household floor is covered [‡]	440	426 (97%)	365	333 (91%)
Household wall made of bricks, concrete, or similar [‡]	440	304 (69%)	365	215 (59%)
Household drinking water source inside compound	435	324 (74%)	360	294 (82%)
Latrine used by household has a ceramic or masonry pedestal [‡]	432	153 (35%)	359	142 (40%)
Latrine used by household has a drop-hole cover [‡]	434	232 (53%)	359	224 (62%)
Compound level variables				
Number of compound members	287	14 (6.2)	208	19 (12)
Number of households	287	3.8 (2.1)	208	4.4 (3.7)
Number of water taps in compound	283	0.98 (0.95)	207	1.4 (1.6)
Number of latrines in compound	287	1.0 (0.20)	207	1.1 (0.57)

Number of people sharing a latrine	285	14 (6.2)	197	17 (8.9)
Number of households sharing a latrine	285	3.7 (1.8)	197	4.0 (2.8)
Latrine walls made of brick, concrete or similar [‡]	282	72 (26%)	204	67 (33%)
Compound population density, persons/square meter [□]	281	0.071 (0.04)	205	0.087 (0.05)
Compound has electricity that normally functions	287	251 (87%)	208	189 (91%)
Compound is prone to flooding	287	184 (64%)	208	120 (58%)
Any animals observed in compound [‡]	287	170 (59%)	208	132 (63%)
Dog(s) observed [‡]	287	14 (4.9%)	208	14 (6.7%)
Chicken(s) or duck(s) observed [‡]	287	40 (14%)	208	30 (14%)
Cat(s) observed [‡]	287	149 (52%)	208	116 (56%)

Data are n (%) or mean (standard deviation) and collected by questionnaire unless otherwise noted. [†] Age range 32-1819 days, IQR 339-1021 days. Age distributions available in Supplemental Figure 3. ^{*} Assessed using Simple Poverty Scorecard for Mozambique (http://www.simplepovertyscorecard.com/MOZ_2008_ENG.pdf). [‡] Data collected by direct observation. [□] Calculated as # of people living in the compound divided by the area of the compound in square meters. Source files available in Table 1 – source data 1 and Table 1 – source code 1.

1114 **Table 2: Effect of the intervention on bacterial, protozoan, and STH infection and diarrhea at 12 and 24 months post-intervention.**

	Prevalence			12 month Prevalence ratio (95% CI), p-value *		24 month Prevalence ratio (95% CI), p-value □	
	Baseline	12-month	24-month	unadjusted	adjusted†	unadjusted	adjusted†
Any bacterial or protozoan infection‡							
Control	313/392 (80%)	334/395 (85%)	403/459 (88%)
Intervention	278/361 (77%)	347/408 (85%)	392/462 (85%)	1.04 (0.94 - 1.15), p=0.41	1.04 (0.94 - 1.15), p=0.41	1.00 (0.91 - 1.10), p=1.0	0.99 (0.91 - 1.09), p=0.89
Any STH infection‡							
Control	170/360 (47%)	143/283 (51%)	142/253 (56%)
Intervention	138/329 (42%)	150/305 (49%)	136/292 (47%)	1.12 (0.89 - 1.40), p=0.33	1.11 (0.89 - 1.38), p=0.35	0.94 (0.75 - 1.17), p=0.59	0.95 (0.77 - 1.17), p=0.62
Diarrhea‡							
Control	67/526 (13%)	40/430 (9.3%)	53/390 (14%)
Intervention	59/448 (13%)	59/436 (14%)	53/410 (13%)	1.41 (0.80 - 2.48), p=0.24	1.69 (0.89 - 3.21), p=0.11	0.92 (0.55 - 1.54), p=0.76	0.84 (0.47 - 1.51), p=0.56
Any Bacteria							
Control	271/392 (69%)	285/395 (72%)	345/459 (75%)
Intervention	227/361 (63%)	292/408 (72%)	324/462 (70%)	1.09 (0.95 - 1.25), p=0.25	1.09 (0.95 - 1.26), p=0.20	1.03 (0.90 - 1.18), p=0.69	1.00 (0.87 - 1.15), p=0.96
<i>Shigella</i>							
Control	179/392 (46%)	204/395 (52%)	269/459 (59%)
Intervention	152/361 (42%)	218/408 (53%)	245/462 (53%)	1.13 (0.91 - 1.39), p=0.28	1.12 (0.92 - 1.38), p=0.27	0.98 (0.80 - 1.20), p=0.86	0.95 (0.79 - 1.16), p=0.64
ETEC							
Control	116/392 (30%)	142/395 (36%)	127/459 (28%)
Intervention	110/361 (30%)	143/408 (35%)	126/462 (27%)	0.93 (0.68 - 1.28), p=0.66	0.96 (0.69 - 1.33), p=0.81	0.95 (0.67 - 1.35), p=0.77	0.83 (0.57 - 1.19), p=0.31
<i>Campylobacter</i>							
Control	39/392 (9.9%)	32/395 (8.1%)	48/459 (10%)
Intervention	21/361 (5.8%)	35/408 (8.6%)	34/462 (7.4%)	1.78 (0.89 - 3.56), p=0.10	1.68 (0.82 - 3.45), p=0.16	1.20 (0.60 - 2.39), p=0.60	1.28 (0.62 - 2.62), 0.50
<i>C. difficile</i>							
Control	22/392 (5.6%)	13/395 (3.3%)	13/459 (2.8%)

Intervention	13/361 (3.6%)	17/408 (4.2%)	11/462 (2.4%)	1.95 (0.71 - 5.35), p=0.20	2.09 (0.77 - 5.64), p=0.15	1.32 (0.47 - 3.73), p=0.60	1.41 (0.46 - 4.30), p=0.54
<i>E. coli</i> O157							
Control	13/392 (3.3%)	19/395 (4.8%)	25/459 (5.5%)
Intervention	18/361 (5.0%)	14/408 (3.4%)	16/462 (3.5%)	0.48 (0.18 - 1.27), p=0.14	0.46 (0.18 - 1.21), p=0.12	0.43 (0.15 - 1.29), p=0.13	0.52 (0.17 - 1.59), p=0.25
STEC							
Control	3/392 (0.77%)	9/395 (2.3%)	17/459 (3.7%)
Intervention	10/361 (2.8%)	5/408 (1.2%)	15/462 (3.3%)	0.14 (0.03 - 0.67), p=0.014	0.15 (0.03 - 0.70), p=0.016	0.23 (0.05 - 1.03), p=0.055	0.24 (0.05 - 1.01), p=0.052
Any Protozoa							
Control	205/392 (52%)	236/395 (60%)	303/459 (66%)
Intervention	195/361 (54%)	259/408 (63%)	296/462 (64%)	1.04 (0.87 - 1.24), p=0.69	1.03 (0.86 - 1.22), p=0.76	0.93 (0.78 - 1.11), p=0.40	0.91 (0.76 - 1.09), p=0.29
<i>Giardia</i>							
Control	201/392 (51%)	230/395 (58%)	294/459 (64%)
Intervention	186/361 (52%)	251/408 (62%)	289/462 (63%)	1.06 (0.88 - 1.27), p=0.55	1.05 (0.88 - 1.25), p=0.58	0.96 (0.80 - 1.14), p=0.61	0.93 (0.78 - 1.11), p=0.44
<i>Cryptosporidium</i>							
Control	8/392 (2%)	8/395 (2%)	14/459 (3.0%)
Intervention	16/361 (4.4%)	15/408 (3.7%)	15/462 (3.3%)	0.89 (0.23 - 3.43), p=0.87	0.89 (0.24 - 3.31), p=0.86	0.46 (0.11 - 1.93), p=0.29	0.53 (0.13 - 2.14), p=0.37
Any virus							
Control	53/392 (14%)	52/395 (13%)	59/459 (13%)
Intervention	52/361 (14%)	45/408 (11%)	62/462 (13%)	0.77 (0.45 - 1.32), p=0.35	0.75 (0.44 - 1.27), p=0.29	0.96 (0.55 - 1.68), p=0.88	1.03 (0.57 - 1.86), p=0.92
Norovirus GI/GII							
Control	38/392 (9.7%)	44/395 (11%)	47/459 (10%)
Intervention	39/361 (11%)	37/408 (9.1%)	55/462 (12%)	0.71 (0.38 - 1.33), p=0.28	0.68 (0.36 - 1.27), p=0.23	1.00 (0.52 - 1.93), p=0.99	1.10 (0.55 - 2.18), p=0.79
Adenovirus 40/41							
Control	13/392 (3.3%)	9/395 (2.3%)	7/459 (1.5%)
Intervention	9/361 (2.5%)	9/408 (2.2%)	6/462 (1.3%)	1.34 (0.34 - 5.23), p=0.68	1.24 (0.32 - 4.83), p=0.76	1.18 (0.23 - 5.98), p=0.84	0.97 (0.18 - 5.19), p=0.97
Coinfection, ≥ 2 GPP							

pathogens							
Control	206/392 (53%)	237/395 (60%)	302/459 (66%)
Intervention	185/361 (51%)	257/408 (63%)	282/462 (61%)	1.08 (0.90 - 1.29), p=0.39	1.08 (0.91 - 1.29), p=0.37	0.95 (0.80 - 1.12), p=0.54	0.93 (0.79 - 1.10), p=0.41
<i>Trichuris</i>							
Control	139/360 (39%)	116/283 (41%)	124/253 (49%)
Intervention	117/329 (36%)	120/305 (39%)	117/292 (40%)	1.05 (0.82 - 1.35), p=0.68	1.01 (0.79 - 1.28), p=0.96	0.89 (0.69 - 1.16), p=0.40	0.86 (0.67 - 1.10), p=0.22
<i>Ascaris</i>							
Control	95/360 (26%)	82/283 (29%)	78/253 (31%)
Intervention	68/329 (21%)	87/305 (29%)	56/292 (19%)	1.26 (0.87 - 1.82), p=0.22	1.33 (0.92 - 1.93), p=0.13	0.80 (0.52 - 1.21), p=0.29	0.83 (0.54 - 1.27), p=0.39
Coinfection, ≥ 2 STH							
Control	64/360 (18%)	55/283 (19%)	60/253 (24%)
Intervention	47/329 (14%)	57/305 (19%)	37/292 (13%)	1.16 (0.76 - 1.77), p=0.50	1.17 (0.76 - 1.79), p=0.49	0.67 (0.40 - 1.13), p=0.13	0.63 (0.37 - 1.07), p=0.084

Prevalence results are presented as (n/N (%)). All effect estimates are presented as prevalence ratios (ratio of ratios) and estimated using generalized estimating equations to fit Poisson regression models with robust standard errors. *Analysis includes all children measured at baseline and 12-month visits. □ Analysis includes all children measured at baseline and 24-month visits. ‡Outcome was pre-specified in trial registration. All other outcomes are exploratory. †Pathogen outcomes adjusted for child age and sex, caregiver's education, and household wealth index. Reported diarrhea was also adjusted for baseline presence of a drop-hole cover and reported use of a tap on compound grounds as primary drinking water source. Sample sizes for adjusted analyses are slightly smaller than numbers presented in prevalence estimates due to missing covariate data. *Y. enterocolitica*, *V. cholerae*, *E. histolytica*, and rotavirus were detected in <2% of samples in each arm at each phase. Descriptive data for these pathogens are available in the Supplemental Table 2. Source files available in Table 2 – source data 1 and Table 2 – source code 1.

1123 **Table 3:** Effect of intervention on bacterial, protozoan, and STH infection and reported diarrhea in children born into study sites post-intervention (post-baseline)
1124 but by 24-month visit compared with children of a similar age at baseline (<2 years old).

	Prevalence (<2 years old)		Prevalence ratio (95% CI), p-value	
	Baseline	24-month, Born-in	unadjusted	adjusted [†]
Any bacterial or protozoan infection‡				
Control	158/228 (69%)	79/106 (75%)
Intervention	129/201 (64%)	71/107 (66%)	0.96 (0.77 - 1.21), p=0.74	0.99 (0.80 - 1.22), p=0.92
Any STH infection‡				
Control	67/205 (33%)	25/68 (37%)
Intervention	52/183 (28%)	13/75 (17%)	0.52 (0.26 - 1.05), p=0.069	0.51 (0.27 - 0.95), p=0.035
Diarrhea‡				
Control	46/283 (16%)	18/105 (17%)
Intervention	43/238 (18%)	22/100 (22%)	1.20 (0.57 - 2.5), p=0.64	1.37 (0.47 - 4.03), p=0.57
Any Bacteria				
Control	142/228 (62%)	70/106 (66%)
Intervention	102/201 (51%)	51/107 (48%)	0.89 (0.66 - 1.20), p=0.44	0.90 (0.67 - 1.19), p=0.45
<i>Shigella</i>				
Control	67/228 (29%)	36/106 (34%)
Intervention	49/201 (24%)	15/107 (14%)	0.48 (0.28 - 0.83), p=0.009	0.49 (0.28 - 0.85), p=0.011
ETEC				
Control	70/228 (31%)	30/106 (28%)
Intervention	58/201 (29%)	24/107 (22%)	0.84 (0.46 - 1.52), p=0.56	0.85 (0.48 - 1.51), p=0.58
<i>Campylobacter</i>				
Control	27/228 (12%)	14/106 (13%)
Intervention	14/201 (7%)	13/107 (12%)	1.75 (0.63 - 4.87), p=0.29	1.75 (0.61 - 4.98), p=0.30
<i>C. difficile</i>				
Control	20/228 (8.8%)	7/106 (6.6%)
Intervention	13/201 (6.5%)	7/107 (6.5%)	1.33 (0.36 - 4.86), p=0.67	1.49 (0.41 - 5.44), p=0.55

<i>E. coli</i> O157				
Control	7/228 (3.1%)	3/106 (2.8%)
Intervention	9/201 (4.5%)	2/107 (1.9%)	0.45 (0.06 - 3.66), p=0.46	0.53 (0.07 - 4.24), p=0.55
STEC				
Control	1/228 (0.44%)	2/106 (1.9%)
Intervention	9/201 (4.5%)	1/107 (0.93%)	0.05 (0.00 - 1.13), p=0.059	0.05 (0.00 - 1.26), p=0.070
Any Protozoa				
Control	82/228 (36%)	47/106 (44%)
Intervention	74/201 (37%)	43/107 (40%)	0.84 (0.55 - 1.28), p=0.42	0.90 (0.62 - 1.30), p=0.58
<i>Giardia</i>				
Control	79/228 (35%)	44/106 (42%)
Intervention	68/201 (34%)	41/107 (38%)	0.90 (0.58 - 1.39), p=0.63	0.93 (0.64 - 1.36), p=0.70
<i>Cryptosporidium</i>				
Control	7/228 (3.1%)	5/106 (4.7%)
Intervention	12/201 (6%)	5/107 (4.7%)	0.45 (0.08 - 2.57), p=0.37	0.64 (0.12 - 3.51), p=0.61
Any virus				
Control	34/228 (15%)	18/106 (17%)
Intervention	36/201 (18%)	18/107 (17%)	0.83 (0.37 - 1.83), p=0.64	0.83 (0.37 - 1.87), p=0.66
Norovirus GI/GII				
Control	26/228 (11%)	12/106 (11%)
Intervention	26/201 (13%)	17/107 (16%)	1.24 (0.48 - 3.17), p=0.66	1.29 (0.49 - 3.41), p=0.61
Adenovirus 40/41				
Control	7/228 (3.1%)	4/106 (3.8%)
Intervention	7/201 (3.5%)	0/107 (0.0%)	.. □	.. □
Coinfection, ≥2 GPP pathogens				
Control	92/228 (40%)	52/106 (49%)
Intervention	74/201 (37%)	39/107 (36%)	0.82 (0.56 - 1.21), p=0.33	0.86 (0.59 - 1.24), p=0.41
<i>Trichuris</i>				

Control	48/205 (23%)	18/68 (26%)
Intervention	41/183 (22%)	5/75 (6.7%)	0.25 (0.09 - 0.68), p=0.006	0.24 (0.10 - 0.60), p=0.002
<i>Ascaris</i>				
Control	45/205 (22%)	16/68 (24%)
Intervention	29/183 (16%)	9/75 (12%)	0.70 (0.30 - 1.64), p=0.42	0.68 (0.30 - 1.54), p=0.36
Coinfection, ≥ 2 STH				
Control	26/205 (13%)	9/68 (13%)
Intervention	18/183 (9.8%)	1/75 (1.3%)	0.13 (0.02 - 1.08), p=0.059	0.12 (0.01 - 1.02), p=0.052

Analysis includes children <2 years old at baseline and children born into the study after baseline and <2 years old at the time of the 24-month visit. Prevalence results are presented as (n/N (%)). All effect estimates are presented as prevalence ratios (ratio of ratios) and estimated using generalized estimating equations to fit Poisson regression models with robust standard errors ‡Outcome was pre-specified in trial registration. All other outcomes are exploratory. †Pathogen outcomes adjusted for child age and sex, caregiver's education, and household wealth index. Reported diarrhea was also adjusted for baseline presence of a drop-hole cover and reported use of a tap on compound grounds as primary drinking water source. Sample sizes for adjusted analyses are slightly smaller than numbers presented in prevalence estimates due to missing covariate data. □Models would not converge due to sparse data. *Y. enterocolitica*, *V. cholerae*, *E. histolytica*, and rotavirus were detected in <2% of samples in each arm at each phase and excluded. Descriptive data for these pathogens are available in the Supplemental Table 2. Source files available in Table 3 – source data 1 and Table 3 – source code 1.

1134 *Supplemental Files*

1135 Supplemental Figure 1: Proportion of each type of sample collected during the baseline, 12-
1136 month, and 24-month phases.

1137 Supplemental Table 1: Number and proportion of sample types collected in each arm at each
1138 phase.

1139 Supplemental Figure 2: Enrollment and stool sample collection profile.

1140 Supplemental Figure 3: Distribution of age (years) of enrolled children at each phase.

1141 Supplemental Table 2: Age stratified baseline prevalence of health outcomes.

1142 Supplemental Figure 4: Prevalence of pathogens by age at baseline, 12-month, and 24-month
1143 phases.

1144 Supplemental Table 3: Baseline enrollment characteristics of children with and without repeated
1145 measures at the 12-month phase

1146 Supplemental Table 4: Baseline enrollment characteristics of children with and without repeated
1147 measures at the 24-month phase

1148 Supplemental Table 5: Balance of characteristics measured at 12-month visits between children
1149 with repeat observations at baseline and 12-month and children with observations at the 12-
1150 month phase only.

1151 Supplemental Table 6: Balance of characteristics measured at 24-month visits between children
1152 with repeat observations at baseline and 24-month and children with observations at the 24-
1153 month phase only.

1154 Supplemental Table 7: Sensitivity analysis assessing the impact of reported deworming on STH
1155 effect estimates 12 and 24 months after the intervention.

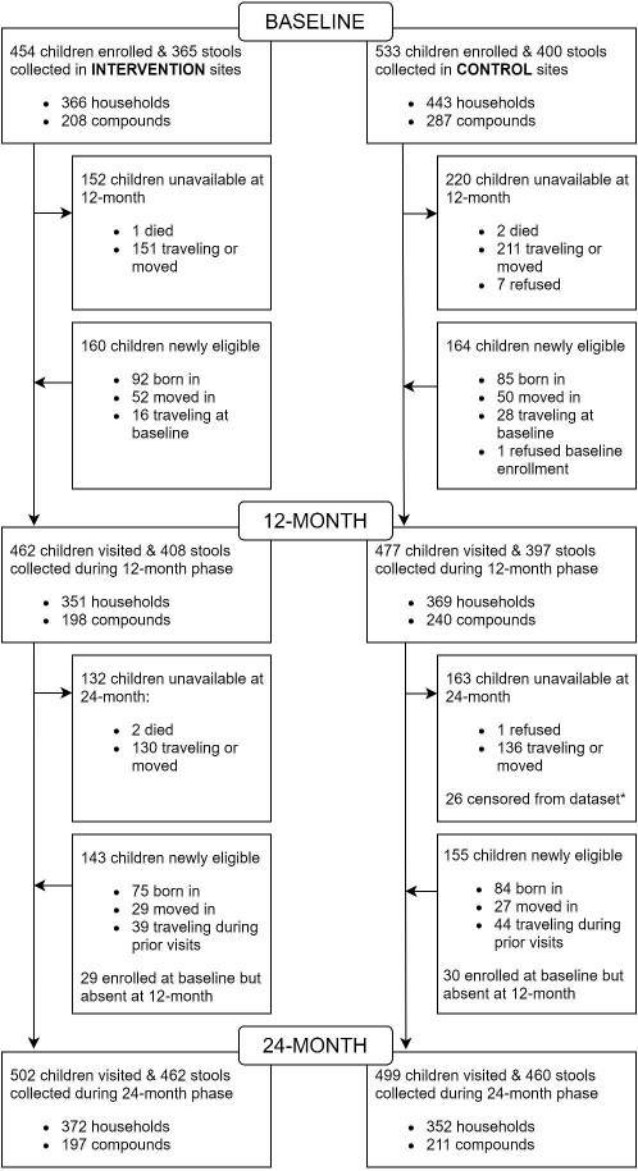
1156 Supplemental Table 8: Sensitivity analysis assessing impact of independent upgrading of control
1157 sanitation facilities on effect estimates.

1158 Supplemental Table 9: Confounding assessment for primary outcome and both secondary
1159 outcomes (any STH, diarrhea) at 12-month.

1160 Supplemental Table 10: Effect estimates (prevalence ratios) for main analyses and all sub-group
1161 analyses adjusted for *a priori* covariates and age-squared

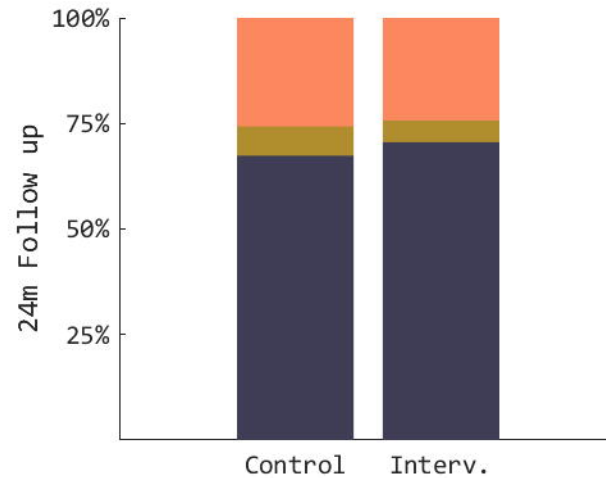
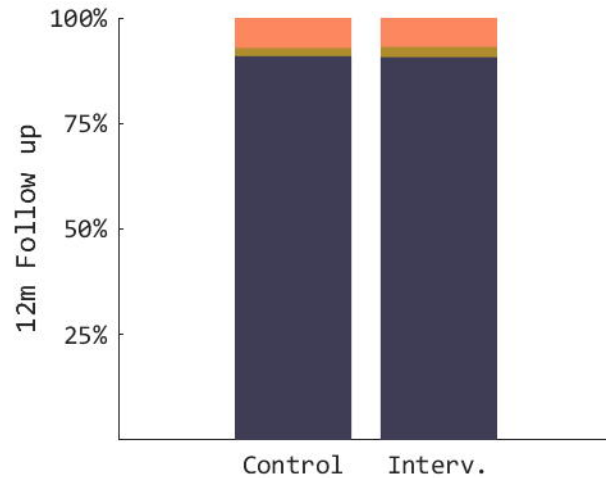
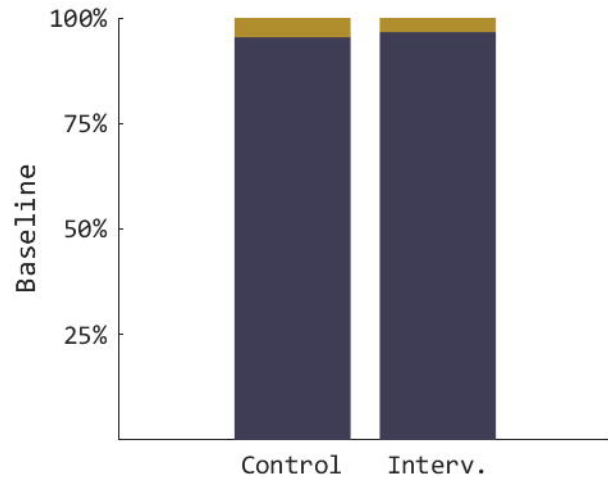
1162 Supplemental Table 11: Comparison of effect estimates (prevalence ratios) at 12- and 24 month
1163 adjusted for *a priori* covariates only and for *a priori* covariates and seasonality.

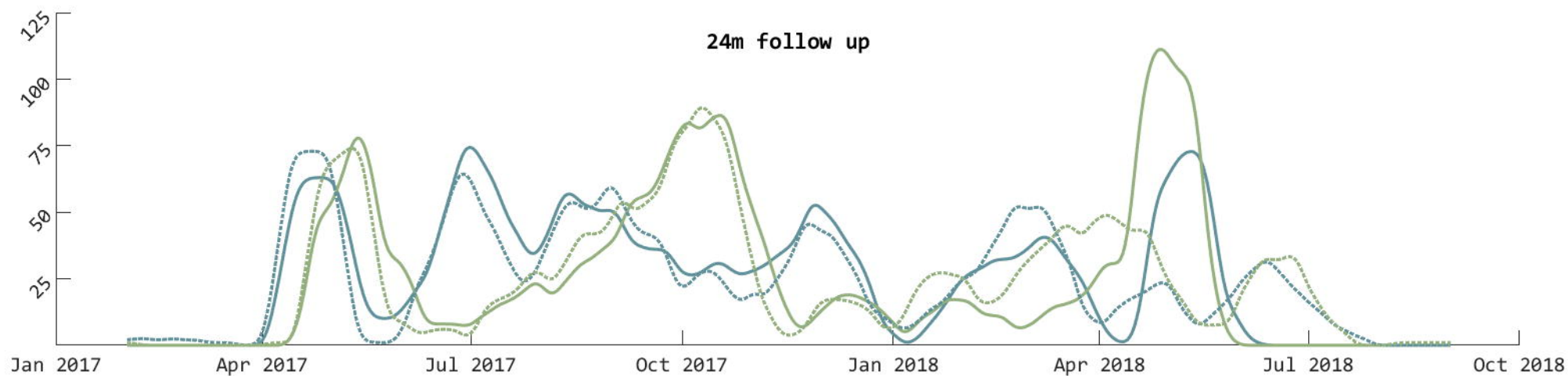
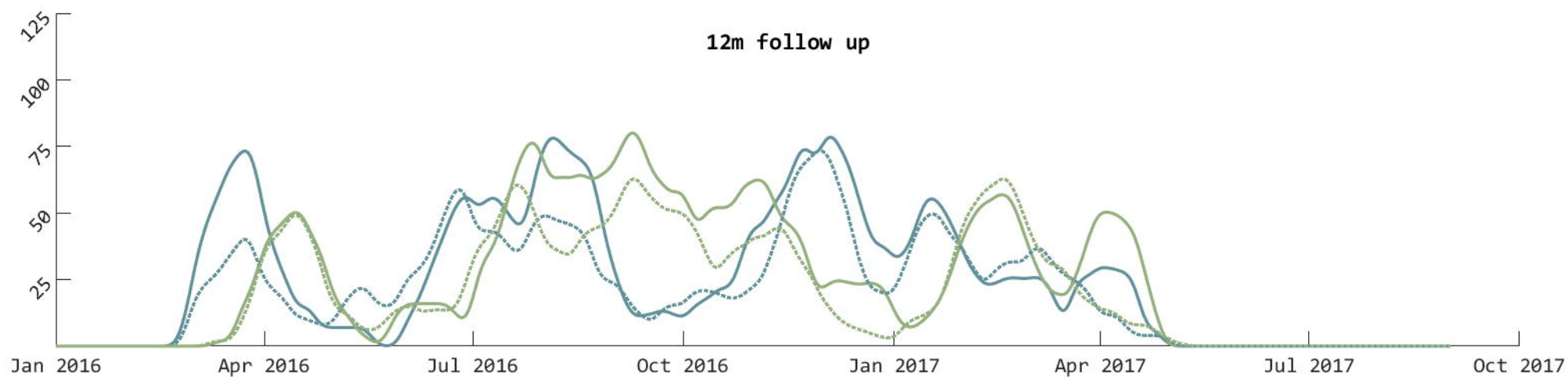
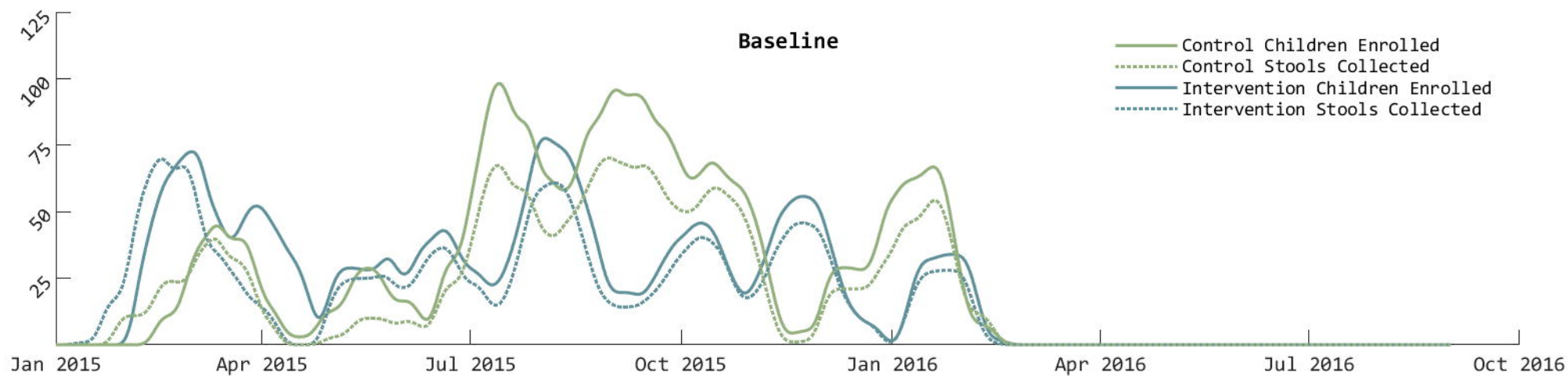
- 1164 Supplemental Table 12: Effect of the intervention on enteric infection and diarrhea in children >2
- 1165 years old after 24 months
- 1166 Supplemental Table 13: Effect of intervention on enteric infection and reported diarrhea in
- 1167 children born into study sites post implementation (post-baseline) and before 12-month visit
- 1168 Supplemental Table 14: Effect of the intervention on children with repeated observations at
- 1169 baseline and 12-month visit.
- 1170 Supplemental Table 15: Effect of the intervention on children with repeated observations at
- 1171 baseline and 24-month visit.
- 1172 Supplemental Figure 5a: Schematic of communal sanitation block design
- 1173 Supplemental Figure 5b: Construction of a soakaway pit for discharge of liquid effluent from
- 1174 intervention latrines.
- 1175 Supplemental Figure 6a: Photo of communal sanitation block as constructed
- 1176 Figure 6b: Photo of shared latrine as constructed
- 1177 Supplemental Figure 7: Map illustrating locations of intervention (n=208) and control sites
- 1178 (n=287) (compounds).
- 1179 Supplemental Methods: Consent procedures, survey administration, and specimen collection and
- 1180 analysis
- 1181 Supplemental Table 16: Outcome and covariate descriptions, coding, and % missing.
- 1182 Supplemental Table 17: Consort 2010 Checklist Extension for Cluster Trials.

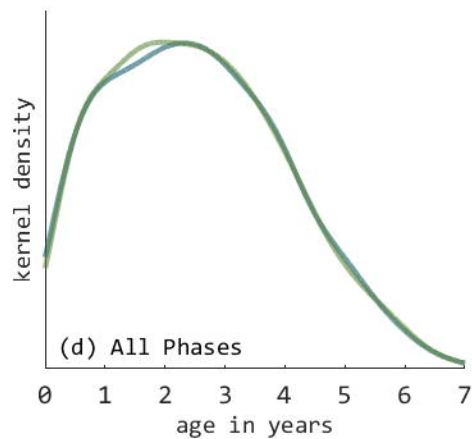
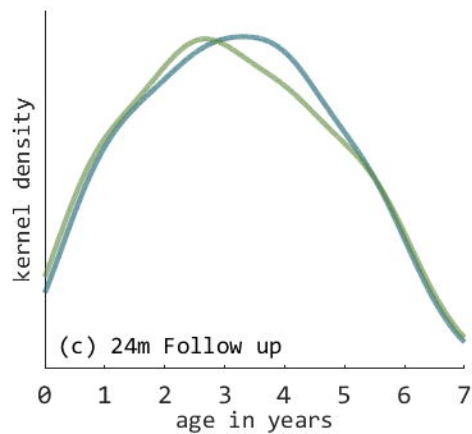
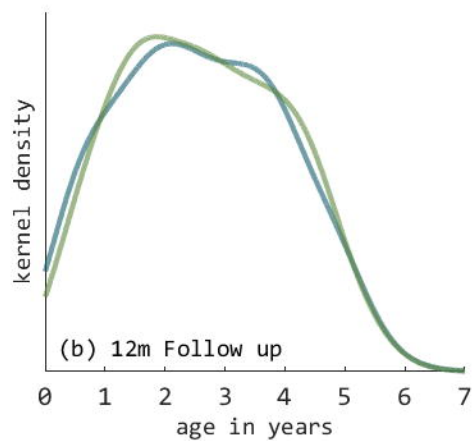
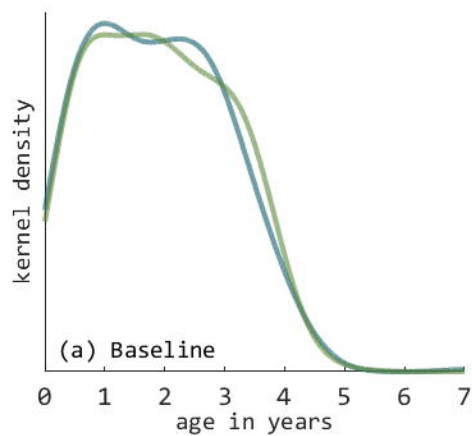


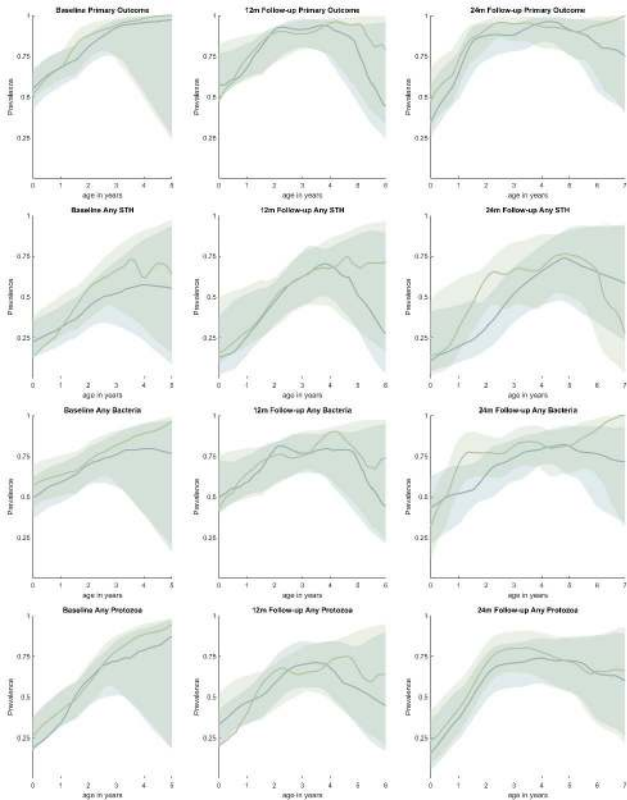
Types of Collected Samples

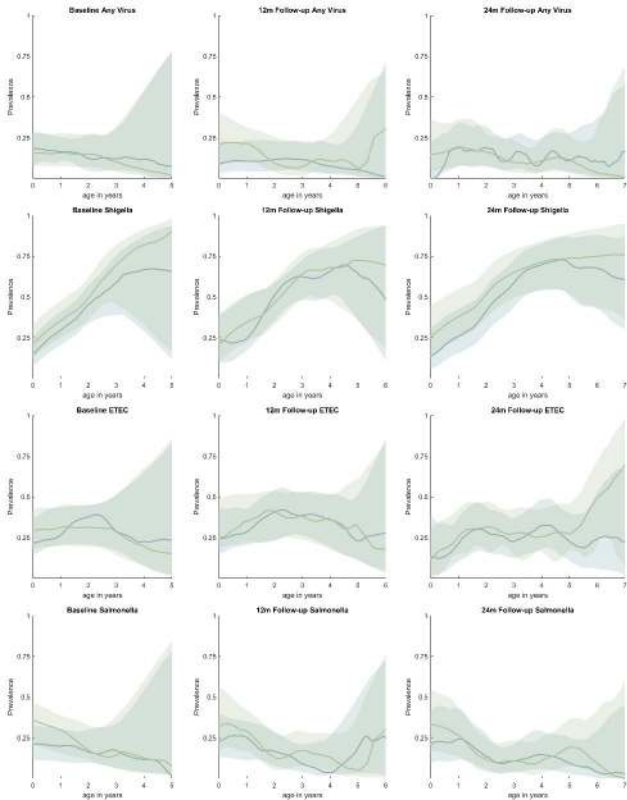
■ Stool ■ Diaper ■ Swab

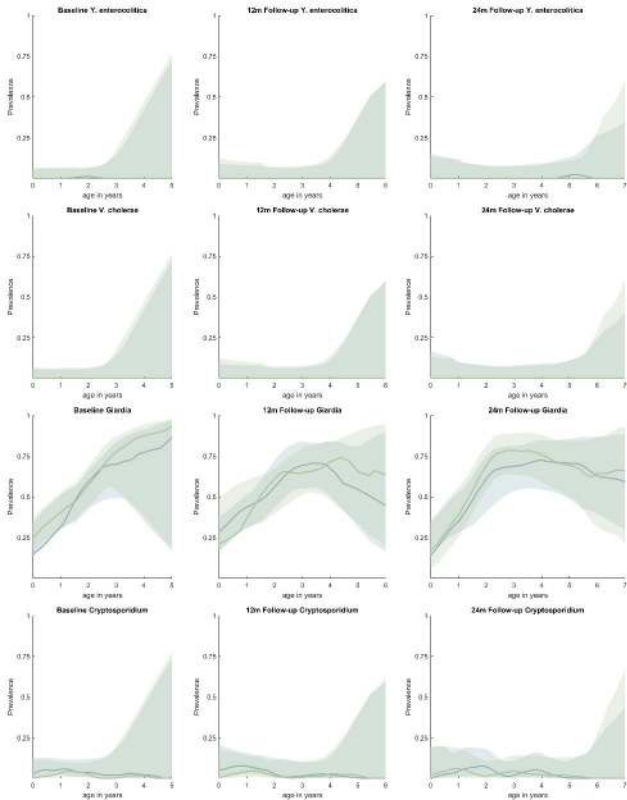


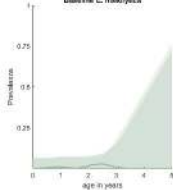
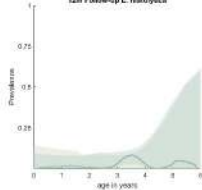
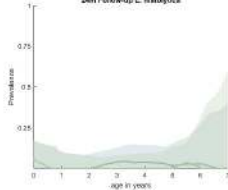
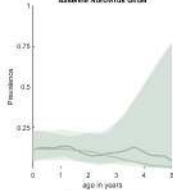
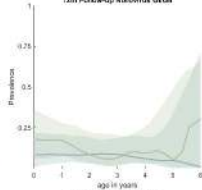
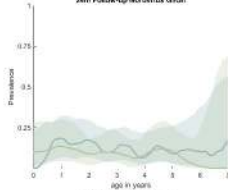
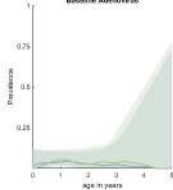
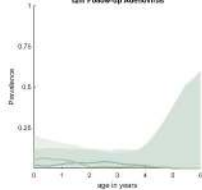
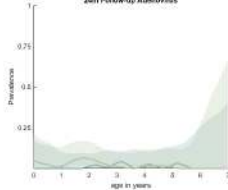
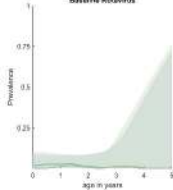
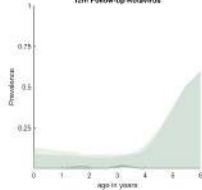
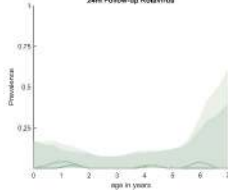


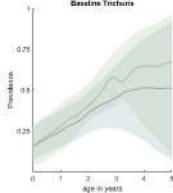
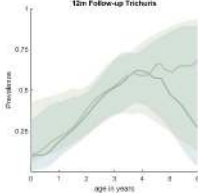
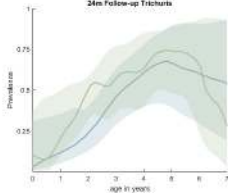
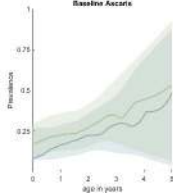
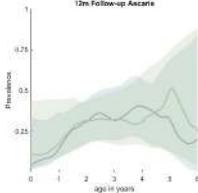
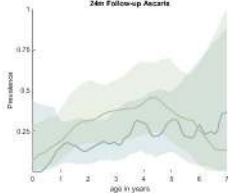








Baseline *E. histolytica***12m Follow-up *E. histolytica*****24m Follow-up *E. histolytica*****Baseline Norovirus GtGI****12m Follow-up Norovirus GtGI****24m Follow-up Norovirus GtGI****Baseline Adenovirus****12m Follow-up Adenovirus****24m Follow-up Adenovirus****Baseline Rotavirus****12m Follow-up Rotavirus****24m Follow-up Rotavirus**

Baseline Trichuris**12m Follow-up Trichuris****24m Follow-up Trichuris****Baseline Ascaris****12m Follow-up Ascaris****24m Follow-up Ascaris**

Elevated water storage tank (500L)

Door (entry)

Laundry basin

Pour-flush toilet
(pedestal design)

Septic tank

