

Effects of Ethanol on Membrane Lipids III. Quantitative Changes in Lipid and Fatty Acid Composition of Nonpolar and Polar Lipids of Mouse Total Liver, Mitochondria and Microsomes Following Ethanol Feeding¹

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ABSTRACT

The effects of ethanol on the total, nonpolar, and polar lipids of whole liver, mitochondria, and microsomes have been evaluated. Differences in the fatty acid composition of various lipid subclasses have been compared in control and ethanol treated mice. On the whole polyunsaturated fatty acids, especially arachidonic (20:4) and docosahexaenoic (22:6), were found to decrease. The significance of an enzymatic mechanism vs. a peroxidative mechanism to explain the results is discussed. Decreases also were observed in the ratios of arach-

idonate/linoleate following ethanol feeding. These changes are thought to be associated with decreases in the activity of the chain elongation-desaturation system.

INTRODUCTION

Both the size and the number of rat liver mitochondria have been shown to be affected by ethanol (1). In addition changes in fatty acid composition have been associated with membrane stability (2-4). Thus, it would seem that ethanol may have an effect on membrane structure and function, and this effect may be on changes in lipid composition. Liver triglyceride (TG) levels are reported to be influenced by the amount of ethanol given and the duration of exposure (5,6). On the other hand, Fallen et al. (7), found no increase in liver TG following the administration of ethanol. French et al. (8), studied the effects of ethanol on the important membrane constituents cholesterol

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TABLE I

Per Cent Distribution of Nonpolar Lipid and Polar Lipid in Total Liver, Mitochondria, and Microsomes, of Control^a and Ethanol Fed Mice^b

Lipid	Total liver		Mitochondria		Microsomes	
	Control	Alcohol	Control	Alcohol	Control	Alcohol
NPL						
TG	60.4	38.6	48.7	33.8	40.5	11.9
DG	11.7	10.6	22.6	16.8	14.1	13.0
CE	21.6	45.4	22.3	46.4	40.4	64.1
C	5.6	4.3	5.1	1.9	4.0	7.6
FFA	0.7	0.8	1.1	1.0	1.0	2.3
Remainder	—	0.3	0.2	0.1	—	1.1
PL						
CPG	49.2	46.4	63.8	65.5	43.1	35.7
EPG	26.6	21.8	28.2	19.6	42.7	18.2
DPG	8.8	16.1	0.8	2.6	4.2	31.4
SPG	5.1	2.3	0.8	3.5	3.3	4.9
CLPG	4.0	7.8	2.2	0.7	1.8	5.9
SM	3.6	1.1	1.1	4.0	2.3	2.8
Remainder	2.7	4.1	1.1	4.1	2.6	1.1

^aValues taken from Ref. 19.

^bNPL = nonpolar lipid, TG = triglyceride, DG = diglyceride, CE = cholesterol esters, C = cholesterol, FFA = free fatty acids, PL = polar lipid, CPG = choline phosphoglyceride, EPG = ethanolamine phosphoglyceride, DPG = diphosphatidyl glycerol (cardiolipin), SPG = serine phosphoglyceride, CLPG = choline lysophosphoglyceride, and SM = sphingomyelin.

TABLE II

Lipid Content of Total Liver Mitochondria and
Microsomes from Control and Ethanol Fed Mice
Lipid Wt (mg)^a

Tissue sample	TL	NPL	PL	Remainder ^b
Total liver				
Control	1549	805	720	24
Alcohol	1747	1200	540	7
Wt change	+198	+395	-180	-17
Per cent of total	+12.8	+25.5	-11.6	-1.1
Mitochondria				
Control	500	190	245	65
Alcohol	383	214	147	22
Wt change	-117	+24	-98	-43
Per cent total	-23.4	+4.8	-19.5	-8.6
Microsomes				
Control	416	174	211	31
Alcohol	316	133	172	11
Wt change	-100	-41	-39	-20
Per cent of total	-25.1	-9.8	-9.4	-4.8

^aWt are the total wt obtained from each group of 20 mice. TL = total lipid, NPL = non-polar lipid, and PL = polar lipid.

^bResults from less than 100% recovery from column separations.

(C) and cholesterol esters (CE). No differences were found between control and ethanol fed animals.

In animals fed ethanol for 21 days, Fallen et al. (9) found no change in total liver polar lipids or ethanolamine phosphoglyceride (EPG), while Lieber et al. (10) observed increases in both of these under similar conditions.

Recently, evidence was presented for a decrease in the polyunsaturated fatty acids of rat liver mitochondria lipids following acute ethanol intoxication (11). Earlier findings (12) suggested that in ethanol-treated rats a peroxidative decomposition of liver mitochondria

lipids may have occurred. A decrease in polyunsaturated fatty acids could be indicative of an *in vivo* peroxidative breakdown, since these acids would be most susceptible to this mechanism.

One cannot exclude the possibility of increased phospholipase activity following ethanol treatment. Recently evidence was presented showing an increase in plasmalogenase activity in mouse liver after ethanol feeding (13). Other data (14) suggesting that the polar lipid composition did not change after feeding ethanol would argue against the phospholipase hypothesis. However, in an earlier paper we

TABLE III

Effects of Ethanol on the Milligrams of Major Nonpolar Lipids and Polar Lipids in Mouse Liver^a

Tissue sample	NPL			PL		
	DG	TG	CE	EPG	CPG	DPG
Total liver						
Control	94	488	174	196	354	63
Alcohol	127	463	545	118	250	87
Wt change	+33	-25	+369	-78	-104	+24
Per cent of total	+2.1	-1.6	+23.8	-5.0	-6.7	+1.5
Mitochondria						
Control	43	93	42	69	156	
Alcohol	36	72	99	29	96	
Wt change	-7	-21	+57	-40	-80	
Per cent of total	-1.4	-4.2	+11.4	-8.0	-16.0	
Microsomes						
Control	25	71	70	87	89	9
Alcohol	17	16	85	31	62	54
Wt change	-8	-65	+15	-56	-17	+45
Per cent of total	-1.9	-15.6	+3.6	-13.5	-4.1	+10.8

^aValues given are for each group of 20 mice. Only lipids which represented 10% or more of either subfraction are considered. See footnote b, Table I for abbreviations.

TABLE IV
Changes in the Milligrams of Fatty Acids in Mouse Total Liver Lipids^a

Fatty acid	CE	DG	TG	EPG	CPG	DPG	Totals
Saturated							
14:0 C		7			18	3	28
A		10			10	7	27
Per cent change ^b		+3.2			-2.2	+6.3	0 ^c
16:0 C	49	18	97	59	66	8	297
A	68	32	85	23	42	17	267
Per cent change	+10.9	+14.9	-2.4	-18.4	-6.8	+14.3	-1.9
18:0 C		5	19	33	25	3	89
A		32	41	18	75	8	174
Per cent change		+28.7	+4.5	-7.6	+14.1	+7.9	+5.5
20:0 C		7					7
A		0					0
Per cent change		-7.4					-0.4
Unsaturated							
16:1 C	13	4	41		18	6	82
A	39	8	16		5	3	71
Per cent change	+14.9	+4.2	-5.1		-3.7	-4.8	-0.7
18:1 C	60	31	155	32	76	8	362
A	250	30	135	8	50	7	480
Per cent change	+109.2	-1.1	-4.1	-12.2	-7.3	-1.6	+4.7
18:2 C	20		128	10	7	29	194
A	85		117	19	10	36	267
Per cent change	+37.3		-2.2	+4.6	+0.8	+11.1	+4.7
20:4 C		10		37	75	5	127
A		3		20	39	2	64
Per cent change		-7.4		-8.7	-10.2	-4.8	-4.1
22:6 C		9		10	32		51
A		0		6	0		6
Per cent change		-9.6		-2.0	-9.0		-2.9

^aValues given are for each group of 20 mice and represent the average of four experiments. C = control, A = alcohol; see footnote b, Table I for other abbreviations.

^bRelative to the total amount (from Table III) of lipid subclass in question, i.e., cholesterol esters, diglyceride, etc. present in the control group. Only fatty acids representing 5% or more from each group are considered.

^cThe percentages given in this column are relative to the control total lipid values from Table II.

suggested increased enzymatic hydrolysis of polar lipids to explain the results of incorporation of palmitate-1-¹⁴C in mouse liver subcellular fractions (15).

In this article we examine the effects of chronic ethanol consumption on the levels of various nonpolar and polar lipids from mouse liver and also investigate changes in the composition of fatty acids associated with these lipids. Additional evidence in favor of the enzyme hydrolysis hypothesis will be presented by showing that the fatty acid composition of individual lipid classes is altered following ethanol ingestion for 21 days. These lipid changes and alterations in fatty acids were observed in the total liver, as well as the mitochondria and microsomes.

MATERIALS AND METHODS

Animals

Male Swiss-Webster mice weighing 24-26 g were used in this study. The control and

alcoholic groups were maintained for 21 days on water and ethanol, respectively, as described previously (13); each group consisted of 20 animals. Subcellular fractions were prepared by ultracentrifugation (16). Histological comparisons, between sections of liver from control and alcohol groups, using Safranin and Sudan Black stains (17) showed that fatty livers had not been produced in the alcoholic mice.

Lipid Extraction and Chromatography

The procedures for lipid extraction, separation into nonpolar and polar lipid fractions, and thin layer chromatography to obtain individual lipid subclasses, are described elsewhere (15,18). Quantitative lipid analysis was carried out, as previously described (19).

Gas Chromatography

Lipid subclasses representing 10% or more of the nonpolar lipid (NPL) and polar lipid (PL) were subjected to transesterification (20). Methyl esters were analyzed qualitatively by gas

TABLE V
Changes in the Milligrams of Fatty Acids
in the Mitochondrial Lipids of Mouse Liver^a

Fatty acid		CE	DG	TG	EPG	CPG	Totals
Saturated							
14:0	C	1	5		7	12	25
	A	8	3		2	5	18
	Per cent change ^b	+16.7	-4.6		-7.2	-4.5	-1.4 ^c
16:0	C	12	9	23	16	29	89
	A	10	10	10	7	19	58
	Per cent change	-4.8	+2.3	-14.0	-13.0	-6.4	-6.2
18:0	C		0.4	1	12	10	23
	A		9	13	3	27	52
	Per cent change		+20.0	+12.9	-13.0	+10.9	+5.8
20:0	C					8	8
	A					0	0
	Per cent change					-5.1	-1.6
Unsaturated							
16:1	C	1	1	6	0	11	19
	A	7	3	0	2	0.2	12
	Per cent change	+14.3	+4.6	-6.4	+2.9	-6.9	-1.4
18:1	C	16	15	40	9	51	131
	A	48	7	22	3	14	97
	Per cent change	+76.2	-18.6	-19.3	-8.7	-23.7	-6.8
18:2	C		2	19	2	8	31
	A		0	14	4	7	25
	Per cent change		-4.6	-5.4	+2.9	-0.6	-1.2
20:4	C		4	2	15	5	26
	A		0	1	6	13	20
	Per cent change		-9.3	-1.1	-13.0	+5.1	-1.2
20:6	C		4		4	19	27
	A		0		1	0	1
	Per cent change		-9.3		-4.3	-12.2	-5.2

^aValues given are for each group of 20 mice and represent the average of four experiments. C = control, A = alcohol; see footnote b, Table I for other abbreviations.

^bRelative to the total amount (from Table III) of lipid subclass in question, i.e., cholesterol esters, diglyceride, etc. present in the control group. Only fatty acids representing 5% or more from either group are considered.

^cThe percentages given in this column are relative to the control total lipid values from Table II.

liquid chromatography (GLC). Quantitative calculations were made using values for the various lipid classes present in normal mice, which we previously presented (19), and the values for ethanol fed mice reported in this paper (see Table I). GLC analysis was carried out using a Packard dual column Model 7800 instrument equipped with a hydrogen flame detector. A 6 ft glass column of either 14.5% ethylene glycol succinate-methyl silicone polymer (EGSS-X) coated on 100-120 mesh Gas Chrome P or 16% Apiezon M on 60-80 mesh Gas Chrome S was used. Samples were run isothermally at 190 C. The carrier gas was nitrogen at an inlet pressure of 32 psi and a flow rate of 140 ml/min. Peak areas were calculated as the product of peak ht and the width at half peak ht. Percentages are given in terms of peak areas. Corrections for the detector response to the various fatty acids have been incorporated in the percentages. Fatty acid methyl esters were identified by retention times relative to methyl stearate

(18:0) and by cochromatography with known standards.

RESULTS

The overall effects of ethanol on the lipids of the total liver and subcellular fractions are given in Table II. These data show that ethanol fed mice had 12.8% more lipid in the total liver than the controls and that this resulted from an increase in NPL exceeding the loss in PL. Mitochondria, on the other hand, showed a loss of 23.4% total lipid (TL) which can be attributed to losses in PL. Microsomes also showed an overall loss of lipid (25.1%), and this resulted from losses of both NPL and PL.

Table III shows that, in the total liver, the increase in NPL was due principally to CE which increased 300% (24% of the TL) and the loss of PL can be attributed to choline phosphoglyceride (CPG) and EPG, the losses in both cases representing more than 5% TL. The loss of mitochondrial and microsomal PL also

TABLE VI
Changes in Milligrams of Fatty Acids in the Microsomal Lipids of Mouse Liver^a

Fatty acid	CE	DG	TG	EPG	CPG	DPG	Totals
Saturated							
14:0 C		1		7		0.4	8
A		1		1		4	6
Per cent change ^b		0		-6.9		+40.0	-0.4 ^c
16:0 C	14	5	12	23	16	1	71
A	16	3	4	7	0	10	40
Per cent change	+2.8	+8.0	-11.3	-18.4	-18.0	+100	-7.4
18:0 C		1		6		9	40
A		3		0		20	31
Per cent change		+8.0		-8.4		+12.3	-2.2
20:0 C		4					4
A		0					0
Per cent change		-16.0					-1.0
Unsaturated							
16:1 C	6	1	5		0	1	13
A	6	1	1		16	4	28
Per cent change	0	0	-5.6		+18.0	+33.3	+3.6
18:1 C	28	8	23	13	11	1	84
A	33	5	8	4	4	4	58
Per cent change	+7.1	-12.0	-21.1	-10.3	-7.9	+33.3	-6.2
18:2 C	10		19	4	9	4	48
A	13		2	5	0	20	40
Per cent change	+4.3		-23.9	+1.1	-10.1	+17.8	-1.9
20:4 C		2		6	21	1	30
A		0		6	10	1	17
Per cent change		-8.0		0	-12.3	0	-3.1
22:6 C		2			12		14
A		0			0		0
Per cent change		-8.0			-13.5		-3.4

^aValues given are for each group of 20 mice and represent the average of four experiments. C = control, A = alcohol; see footnote b, Table I for other abbreviations.

^bRelative to the total amount (from Table III) of lipid subclass in question, i.e., cholesterol esters, diglycerides, etc. present in the control group. Only fatty acids representing 5% or more from either group are considered.

^cThe percentages given in this column are relative to the control total lipid values from Table II.

was due to CPG and EPG. Diposphatidyl glycerol (cardiolipin) (DPG) increased in all cases, although in mitochondria this lipid constituted less than the selected 10% cutoff value (Table I). The surprisingly high percentage of DPG in the total liver and microsomes is not understood. The ester-phosphorous ratio (19) of this isolated lipid indicated a relatively pure fraction; and, therefore, the per cent values are thought to be real. Perhaps mouse liver is different with respect to DPG than other tissues analyzed to date. TG decreased in the total liver and subcellular fractions, with the greatest change occurring in the microsomes (15% TL).

Tables IV-VI give the amounts of fatty acids present in the major lipid subclass. Several observations stand out and deserve noting. The amount of arachidonic (20:4) and docosahexanoic acids (22:6) decreased in all lipids containing them in the total liver, mitochondria, and microsomes of the ethanol treated mice, except in the CPG of mitochondria where 20:4 increased. No particular trend is evident in the

changes with regard to linoleic acid (18:2). Overall we see that this fatty acid increased by ca. 5% TL in the total liver and decreased by 1-2% in the mitochondria and microsomes respectively. Since the changes in 18:2 do not appear to be localized, the effect of ethanol to decrease the ratios of 20:4/18:2 (Table VII) is probably a general one rather than a specific effect on any particular lipid. Table IV shows that the large increase in CE in the total liver and mitochondria is primarily oleic acid (18:1).

DISCUSSION

The peroxidative hypothesis to explain decreases in polyunsaturated fatty acids following ethanol ingestion would appear to have little support in view of the work of others (11,21). Thus it would appear that the effect is more on the enzyme level, as we suggested earlier (15). Evidence has been presented suggesting that the characteristic fatty acid composition of phospholipids is determined by redistribution of the fatty acids after the nitrogenous base has been

attached (22). The activity of the fatty acyl-CoA:lysophosphatide transferases catalyzing this reaction recently has been shown to be enhanced with increased ethanol consumption (23). Thus, a mechanism of this type seems more likely than peroxidation. This is supported further when one examines the data in Tables V and VI. Here we see that, in both the mitochondria and microsomes of alcohol treated mice, a change has occurred in the fatty acid distribution of the major lipids. Our recent studies on the effects of ethanol on aldehydogenic lipids (13) would also support this finding.

Examination of Tables IV-VI also reveals that significant increases in the stearic acid (18:0) content of various lipids has occurred in the total liver and mitochondria of ethanol fed mice. This is in contrast to others (11) who observed decreases in this fatty acid in rats fed ethanol. The reason for this discrepancy is not understood; however, it could be related to experimental conditions, most notably diet and the time duration of the experiments. Corn oil-fed rats and coconut oil-fed rats gave opposite results with respect to fatty acids found in the CPG and EPG following ethanol treatment (23).

A decrease in the arachidonate-linoleate ratio of liver lipids following alcohol ingestion also has been observed by others (24). Similar changes in heart lipids also were reported (25). Such decreases have been suggested to account for membrane stability (3) and mitochondria fragility (2). It has been suggested (26) that the decrease observed in liver mitochondria was due to decreases in the activity of the chain elongation-desaturation system which converts linoleate to arachidonate. Since liver CoA content is decreased markedly by ethanol (27), a decrease in the activity of the enzyme system seems a logical explanation.

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TABLE VII

Effects of Ethanol on the Arachidonate-Linoleate Ratios in Lipids of Mouse Total Liver, Mitochondria, and Microsomes

Lipid fraction	Total liver	Mitochondria	Microsomes
Total lipids			
Control	0.65	0.90	0.62
Alcohol	0.24	0.69	0.42
Change	-0.41	-0.21	-0.20
Nonpolar lipids			
Control	0.67	0.28	0.07
Alcohol	0.01	0.07	0.00
Change	-0.66	-0.21	-0.07
Polar lipids			
Control	2.43	2.20	1.47
Alcohol	0.94	1.30	0.68
Change	-1.49	-0.90	-0.79

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