

Review

Effects of rising temperatures and [CO₂] on the physiology of tropical forest trees

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Using a mixture of observations and climate model outputs and a simple parametrization of leaf-level photosynthesis incorporating known temperature sensitivities, we find no evidence for tropical forests currently existing ‘dangerously close’ to their optimum temperature range. Our model suggests that although reductions in photosynthetic rate at leaf temperatures (T_L) above 30°C may occur, these are almost entirely accountable for in terms of reductions in stomatal conductance in response to higher leaf-to-air vapour pressure deficits D . This is as opposed to direct effects of T_L on photosynthetic metabolism. We also find that increases in photosynthetic rates associated with increases in ambient [CO₂] over forthcoming decades should more than offset any decline in photosynthetic productivity due to higher D or T_L or increased autotrophic respiration rates as a consequence of higher tissue temperatures. We also find little direct evidence that tropical forests should not be able to respond to increases in [CO₂] and argue that the magnitude and pattern of increases in forest dynamics across Amazonia observed over the last few decades are consistent with a [CO₂]-induced stimulation of tree growth.

Keywords: review; photosynthesis; climate change; plant growth

1. INTRODUCTION

In an effort to guide thought as to how tropical forests may respond to climate change, there have been several reviews over the last few years, including Chambers & Silver (2004), Clark (2004) and Wright (2005), which have concluded that CO₂ is unlikely to have any positive effect on forest productivity. But with other projected changes in the global climate system, especially increasing temperatures, almost certainly to result in some form of tropical forest decline, it seems to us that nearly all these reviews are, at best, conceptually inconsistent. For example, Clark (2004) argues that increasing atmospheric [CO₂] should result in ‘little or no enhancement of biomass production rates’, which is equivalent to stating that the growth of tropical forests is currently not carbon limited. Yet, she also cites numerous examples of how higher temperatures might reduce tropical forest productivity through declined rates of net CO₂ assimilation or enhanced rates of respiration, which is, of course, equivalent to assuming the exact opposite. Similarly, when discussing CO₂, Wright (2005) suggests that ‘current photosynthesis levels meet, or even exceed, the carbon requirements for maintenance and growth’ but when discussing light availability concludes that ‘solar irradiance limits net primary production by closed-

canopy forests...because shade limits photosynthetic carbon uptake by most leaves’. Such inconsistencies suggest a need for a coherent and objective framework in which to assess the probable effects of rising temperatures and atmospheric [CO₂] on the physiology and growth of forest trees and this is the objective of this paper.

2. PHOTOSYNTHESIS, RESPIRATION AND PLANT GROWTH

We start with the simple question: ‘is the growth of tropical trees carbon limited?’, noting the relationship between photosynthesis and growth can be simply expressed as

$$N_P = G_P(1 - \phi), \quad (2.1)$$

where N_P is the rate of net primary production (new growth); G_P is the average rate of photosynthesis; and ϕ is the proportion of assimilated carbon lost through respiration of all organs (including the leaves at night) as well as through other processes such as volatile organic carbon emission (Harley *et al.* 2004) or exudation of organic acids and other carbohydrates from roots to the soil solution (Jones *et al.* 2003). From equation (2.1), we can reasonably infer that if N_P varies with a positive correlation to environmental factors known to stimulate G_P then positive evidence would be obtained that the growth of tropical forest trees is currently carbon limited.

It is extremely difficult to change ambient [CO₂] for forest trees in a long-term experimental setting but a

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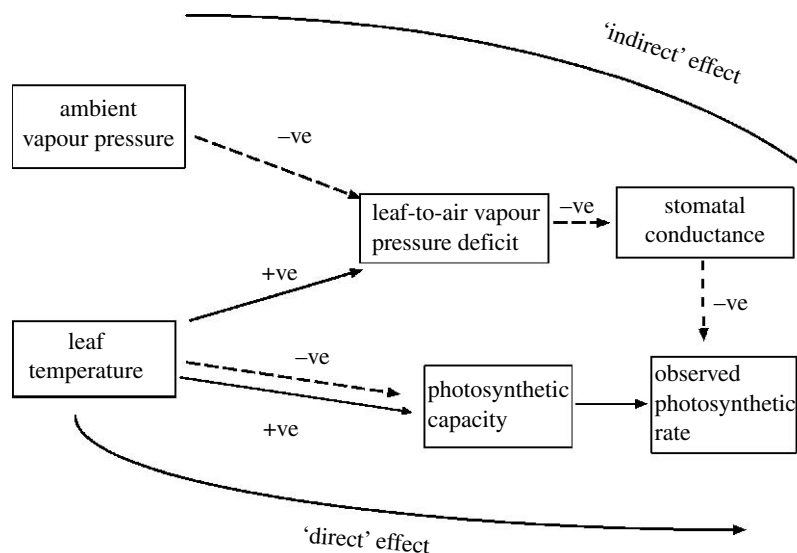


Figure 1. Schematic showing 'direct' and 'indirect' effects of temperature on leaf photosynthetic metabolism.

second strong environmental driver influencing tropical forest G_P is photon irradiance Q . Clarke & Clarke (1994) and Graham *et al.* (2003) showed that the N_P of neotropical trees responds positively to increases in Q on a seasonal or interannual basis. Keeling (2007) used a calibrated crown illumination index to show that tropical tree stem growth rates are positively correlated with tree canopy light exposure. Shading experiments show that the growth of young seedlings and saplings within tropical forests is almost always limited by light (Turner 2001). As far as we know, no mechanism other than a stimulation of photosynthesis by increased Q has been suggested to account for this. For understory plants in tropical forests, $[CO_2]$ also has a strong stimulatory effect on plant growth (Würth *et al.* 1998), as would be expected from the strong $[CO_2]$ dependence of quantum yield (Ehleringer & Björkman 1977) combined with the observation of shaded tropical forest seedlings often only just surviving very close to their growth compensation points (Turner 2001). Thus, the answer to our question seems to be 'yes' and a reasonable place to start any analysis of effects of temperature on forest productivity is thus through effects on photosynthesis and respiration—after which we consider direct long-term effects of $[CO_2]$.

3. TEMPERATURE AND PHOTOSYNTHESIS

As shown in figure 1, temperature can affect photosynthesis through modulation of the rates of activity of photosynthetic enzymes and the electron transport chain (Sage & Kubien 2007) and, in a more indirect manner, through leaf temperatures defining the magnitude of the leaf-to-air vapour pressure difference D , a key factor influencing stomatal conductances. These two processes are termed here 'direct' and 'indirect'.

(a) *Direct (mesophyll) effects*

Direct temperature effects on photosynthetic metabolism involve changes in the activity of ribulose-1,5-carboxylase/oxygenase (Rubisco—the main carboxylating enzyme of photosynthesis) as well as

processes associated with the regeneration of Rubisco's substrate, ribulose-1,5-bisphosphate (RuBP) through the Calvin cycle.

Temperature effects on Rubisco kinetics are complex with activation energies and Michaelis–Menten constants being affected (von Caemmerer 2000), but these temperature sensitivities are now reasonably well established (Bernacchi *et al.* 2001) and with an only modest sensitivity of RuBP carboxylation capacity to temperature (Sage & Kubien 2007; see figure 3). This temperature sensitivity varies little with genotype or growth conditions, although there may have been some genetic adaptation of Rubisco specificity to different levels of aridity (Galmes *et al.* 2005).

The maximum rate of RuBP regeneration, usually considered to be limited by the maximum rate of electron transport J_{max} , is generally more sensitive to temperature than RuBP carboxylation capacity with its temperature sensitivity also varying substantially with growth conditions and/or genotype (June *et al.* 2004). Typical response curves for J_{max} are shown in figure 2, with data from a modelling study of ecosystem flux data from forest near Manaus (Mercado *et al.* 2006), leaf-level measurements from the same tower (Tribuzy 2005) and from soya bean leaves in the laboratory (June *et al.* 2004). The latter also showed that inhibition of J_{max} at supraoptimal T_L is fully reversible. Although the mechanism by which this reversibility occurs is unknown, the decline in J_{max} at high T_L is associated with an increase in the cyclic flow of electrons around photosystem (PS) I possibly serving as an important mechanism for the protection of both PS II and lipid membranes under high-temperature conditions (Sharkey & Schrader 2006). The considerable variation in the temperature sensitivity of J_{max} with both species and growth conditions (June *et al.* 2004) contrasts with the relatively constant temperature sensitivity of RuBP carboxylation/oxygenation. One general 'rule of thumb' then is that enzyme-mediated processes tend to be invariant in their temperature responses, but that, due to potential changes in fluidity and lipid composition (Sung *et al.* 2003),

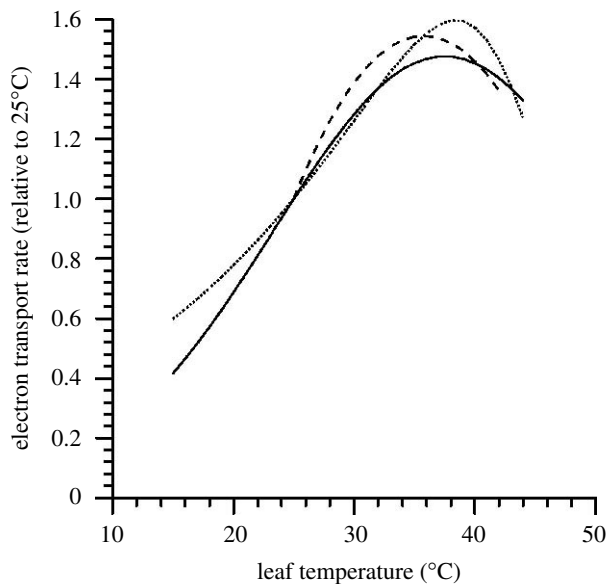


Figure 2. The temperature sensitivity of electron transport as deduced from the studies of June *et al.* (2004) with soya bean, and Tribuzy (2005) and Mercado *et al.* (2006) for Amazon forest trees. Dashed line, Tribuzy; dotted line, Mercado; solid line, June.

membrane-mediated processes may exhibit a considerable flexibility in temperature sensitivity according to growth conditions and genotype.

Although at lower [CO₂] a reduction in the activity of the membrane-bound Rubisco activase at high T_L may also limit photosynthesis (Sage & Kubien 2007), where reductions in enzyme activity occur they are usually irreversible and associated with enzyme denaturation at $T_L > 45^\circ\text{C}$. At such temperatures, irreversible destruction of the thylakoids may also occur, though the temperature at which this occurs depends upon the temperature at which leaves have developed (Berry & Björkman 1980).

(b) Indirect (stomatal) effects

As evaporative demand D increases, stomata tend to close to reduce the rate of water loss through transpiration. Associated with this stomatal closure is a reduction in CO₂ assimilation rate A due to a reduction in the rate of supply of CO₂ to the chloroplast (Farquhar & Sharkey 1982). For rainforest environments, the absolute humidity of the air tends to remain more or less constant over a day (e.g. Shuttleworth *et al.* 1985) and so it is diurnal fluctuations in T_L that drive the variations in D . This changing D as T_L varies over the day gives rise to an apparent temperature dependence of A that is actually associated with stomatal responses to variations in D (Koch *et al.* 1994). This is the indirect temperature response in figure 1.

4. TROPICAL FOREST PHOTOSYNTHETIC RESPONSE TO CLIMATE CHANGE

To quantify the importance of the above, we have developed a simple model of leaf-level photosynthesis, described in full in the electronic supplementary material. In brief, the model consists of standard equations of photosynthesis (Farquhar *et al.* 1980)

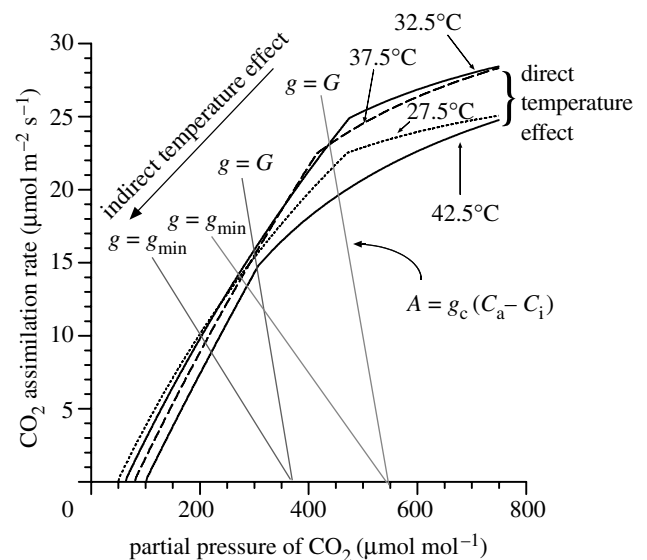


Figure 3. Modelled response of photosynthesis and stomatal conductance g to the intercellular/chloroplastic partial pressure of CO₂ for various leaf temperatures and with operating points at the modelled maximum stomatal conductance G of $0.6 \text{ mol m}^{-2} \text{ s}^{-1}$, along with that associated with the maximum leaf-to-air vapour pressure observed within the model for 2000 and 2040 with [CO₂] = 380 and $550 \text{ } \mu\text{mol mol}^{-1}$, respectively (denoted g_{min}). The lines intersecting the x-axis represent the ‘stomatal supply’ functions (Farquhar & Sharkey 1982) for $g = G$ and $g = g_{\text{min}}$ at [CO₂] = $380 \text{ } \mu\text{mol ml}^{-1}$ and [CO₂] = $550 \text{ } \mu\text{mol ml}^{-1}$.

interfaced with a hybrid of the stomatal models of Jarvis & Davies (1998) and Buckley *et al.* (2003), includes a surface energy balance, and is run for 2000 and 2040 using observational and model output for the region of Manaus in central Amazonia. For 2000, it is run for a current day [CO₂] of $380 \text{ } \mu\text{mol mol}^{-1}$. For 2040, it is run at [CO₂] of 380 and $550 \text{ } \mu\text{mol mol}^{-1}$. The latter is considered a likely [CO₂] to be occurring in 2040. The difference between model predictions for 2040 and 2000 with [CO₂] = $380 \text{ } \mu\text{mol mol}^{-1}$ gives an indication of the direct effect of predicted climate change on A . The difference between 380 and $550 \text{ } \mu\text{mol mol}^{-1}$ in 2040 illustrates the extent to which climate change effects on A will be modified by increases in [CO₂]. For all three [CO₂]/climate assumptions, the indirect temperature effect is quantified by comparing model predictions with g always set to its maximum value, $G = 0.6 \text{ mol m}^{-2} \text{ s}^{-1}$ (see electronic supplementary material), with predictions applying equation (E7), which allow stomata to respond to changes in D .

Using supply and demand functions (Farquhar & Sharkey 1982), figure 3 shows A as a function of intercellular/chloroplastic [CO₂], C , for $Q = 1500 \text{ } \mu\text{mol m}^{-2} \text{ s}^{-1}$. The different demand functions represent the direct temperature effects on A . The indirect (stomatal) supply functions are shown for both $g = G$ and for g at the maximum D occurring in the simulations. Over a wide range of T_L , the direct temperature effect is relatively small, with indirect effects, those being associated with reductions in C as g declines in response to increasing D , being much more significant.

Table 1. Model estimates for annual net CO₂ assimilation, maximum leaf temperature and maximum leaf-to-air vapour pressure difference for a leaf growing at the top of the canopy near Manaus for 2000 and 2040 in the absence of soil water deficits. (For 2040, simulations have been done both with the assumed [CO₂] for 2000 (380 μmol mol⁻¹) and for a more likely [CO₂] around that time of 550 μmol mol⁻¹. Two model assumptions for stomatal conductance g have been invoked: first, with $g \equiv 0.6 \text{ mol m}^{-2} \text{ s}^{-1}$ (minimal stomatal limitation); and secondly, and more realistically, with g responding to variations in leaf-to-air vapour pressure deficit and linking with leaf biochemistry according to equation (E5). The most likely values are shown in italics.)

model run	output parameter	2000 climate	2040 climate	
		[CO ₂] = 380 μmol mol ⁻¹	[CO ₂] = 380 μmol mol ⁻¹	[CO ₂] = 550 μmol mol ⁻¹
$g \equiv 0.6 \text{ mol m}^{-2} \text{ s}^{-1}$	annual net CO ₂ assimilation (mol C m ⁻² a ⁻¹)	287.6	294.8	379.3
	maximum leaf temperature (°C)	34.2	35.8	35.8
	maximum leaf-to-air vapour pressure difference (mmol mol ⁻¹)	20.8	26.8	26.8
interactive g from equation (E5)	annual net CO ₂ assimilation (mol C m ⁻² a ⁻¹)	207.4	188.7	271.1
	maximum leaf temperature (°C)	37.9	39.7	39.7
	maximum leaf-to-air vapour pressure difference (mmol mol ⁻¹)	33.0	40.7	40.8

The modelled annual rates of CO₂ fixation (G_P in equation (2.1)), and maximum simulated T_L and D are shown in table 1. The difference in G_P between $g \equiv G$ and g from equation (E5) for the 2000 climate (288 versus 207 mol C m⁻² a⁻¹) suggests an indirect temperature effect currently reducing G_P by approximately 30%. Maximum T_L and D are also much greater when g is allowed to vary (37.9 versus 34.2°C and 33.0 versus 20.8 mmol mol⁻¹, respectively) due to higher sensible heat fluxes associated with stomatal closure at high D .

Comparing G_P at [CO₂] = 380 μmol mol⁻¹ and $g \equiv G$ for 2000 and 2400 suggests a direct temperature effect of climate change of less than 2% (295 versus 288 mol C m⁻² a⁻¹) with the slight increase attributable to higher Q in 2040. However, when stomatal interactions with the environment are included, G_P is reduced from 207 to 187 mol C m⁻² a⁻¹. This 10% reduction is due to higher T_L and D under the 2040 climate. Nevertheless, once higher [CO₂] in 2040 is also taken into account, the indirect temperature effect reduction in G_P is more than negated by the increased availability of CO₂. Similar results have also been reported for a fully coupled simulation in a global circulation model (Bounoua *et al.* 1999) and changes in g in direct response to higher [CO₂] for our model are considered in the electronic supplementary material.

We conclude that temperature rises of the order of 1.5°C in Amazonia over the next 35 years or so are unlikely to have a significant direct effect on G_P . Lower g due to higher D may reduce G_P below the value that would otherwise occur, but this effect will be more than offset by higher [CO₂]. It is also important to recognize that current day indirect responses to temperature may bear little relationship to indirect effects of higher temperatures in the future. This is because increased temperatures associated with climate change will be accompanied by increases in sea temperatures, and therefore transiently increased evaporation from the oceans. Thus, on average, higher ambient humidities will occur and changes in D as global temperatures increase will be smaller than currently observed as temperatures vary on a daily or seasonal basis. Nevertheless, this effect may be offset by large-scale variations

in precipitation patterns. Indeed, we have used 2040 for the simulation of future climatic effects on G_P , because the Hadley Centre model used predicts significantly more rapid drying in the Amazon than most other general circulation models (GCMs), also with greater reductions in atmospheric humidity (Li *et al.* 2006). This leads to biome shifts after this time. Most other GCMs should therefore predict less severe increases in D than is the case here, and an even greater stimulation of G_P in 2040. Nevertheless, many of these precipitation estimates might also be substantially modified once detailed land-use change effects are taken into account (Moore *et al.* 2007).

5. PLANT RESPIRATION AND TEMPERATURE

Expressed as a proportion of G_P , plant respiration is ϕ in equation (2.1). It has long been known that for tropical forests ϕ tends to be higher than other ecosystems, typically ranging from 0.60 to 0.85 (Lloyd & Farquhar 1996). The reasons for this are unclear as available evidence suggests that, due to long-term acclimation, plants growing in warmer ecosystems should not necessarily have higher ϕ than their cooler counterparts (Atkin *et al.* 2005). One explanation, also consistent with a high proportion of tropical forest autotrophic respiration being below ground, is that tropical trees, growing on relatively infertile soils, need to invest a high proportion of their acquired carbon in the acquisition of phosphorus through mycorrhizal associations and via high rates of organic acid exudation (Lloyd *et al.* 2001). The suggestion of Chambers & Silver (2004) that much of this tropical tree respiration is simply 'wasteful' is without foundation.

Consistent with the idea that the growth of tropical trees may be carbon limited, enhanced respiration losses have been invoked as one explanation for tropical tree growth reductions associated with longer-term warming trends (Clark 2007; Feeley *et al.* 2007), even though this is also at odds with the high levels of carbohydrate reserves generally found in tropical trees (Würth *et al.* 2005) indicating that carbohydrate availability is not limiting for growth (Wright 2005).

But, in any case, the extreme apparent sensitivity of growth to temperature suggested by Clark (2007) and Feeley *et al.* (2007) requires a tropical tree respiration $Q_{10} > 5$ (see electronic supplementary material). This is in clear contradiction to observation (Meir *et al.* 2008) and other/additional explanations may exist. For example, the indirect detrimental effects of high temperatures on A demonstrated above may be linked to the growth reductions of tropical trees observed in warmer and drier years.

In the longer term, there is no reason to believe that tropical trees should not be able to acclimate their respiration to increasing temperatures (Atkin *et al.* 2005) and, even if enhanced respiratory losses do occur in the future, they should be more than offset by the capability for increased G_P as [CO₂] increases simultaneously (table 1 and electronic supplementary material).

6. OTHER TEMPERATURE-RELATED FACTORS

Although we have focused on photosynthesis and respiration, other physiological processes may also be important. For example, reproductive processes such as flowering and fruit set may be especially sensitive to high temperatures (e.g. Sato *et al.* 2006). Another process that may become increasingly important as tropical forests warm may be the ability of plants to emit isoprene, a process thought to help maintain membrane stability under moderately high temperatures (Sharkey & Schrader 2006).

7. INCREASING [CO₂] AND PLANT GROWTH

Although an increased [CO₂] accompanying climate change should more than offset any detrimental effects of higher temperatures and increased D on tropical forest productivity, this does not necessarily mean that increasing [CO₂] should also be serving to stimulate growth rates above those which would otherwise occur. Indeed, although increasing [CO₂] provides a simple explanation for observed increases in recruitment and growth rates of Amazon forests over the last few decades (Lewis *et al.* 2004), this also being quantitatively consistent with theoretical predictions (Lloyd & Farquhar 1996), it has also been argued that a direct stimulation of plant productivity by CO₂ cannot account for these growth responses observed (Chambers & Silver 2004). The latter study did, however, make conservative assumptions regarding the extent to which CO₂ may stimulate tropical forest productivity (a 25% stimulation of N_P for an increase in [CO₂] from approx. 270 to 700 $\mu\text{mol mol}^{-1}$), so their result is not that surprising. In any case, it is not at all clear that experiments exposing plants to large-step changes in [CO₂], such as in typical CO₂ enrichment experiments, provide an adequate analogue for probable growth responses when [CO₂] is gradually increasing, such as is presently the case. For example, seedlings can only typically adjust their ratios of root to shoot by a factor of less than 0.2 in response to a doubling of [CO₂] (Curtis & Wang 1998), but for mature tropical forests an approximately threefold variation in the ratios of root to shoot exists, probably due to variations in nutrient

availability (Maycock & Congdon 2000; Powers *et al.* 2005). Thus, although it may be the case that nutrients, especially P, are becoming relatively more limiting for tropical forest growth as N_P and [CO₂] continue to increase (Chambers & Silver 2004), it would also be remarkable if tropical forest trees were not gradually increasing the proportion of biomass allocated below ground to facilitate relatively greater rates of nutrient acquisition. Moreover, numerous mechanisms exist that allow extra phosphorus to be taken up from the soil solution to support increased growth in response to higher [CO₂] (Lloyd *et al.* 2001). Plants, it seems, have ready access to what are often considered 'unavailable' phosphorus pools (Parfitt 1979; Johnson & Loeppert 2006), much of which should be available to support slowly increasing [CO₂]-mediated increases in growth (cf. Chambers & Silver 2004).

It has also been argued that because tropical tree carbohydrate concentrations, [CH₂O], are 'generally high', [CH₂O] must already be in excess with increasing [CO₂] unable to further stimulate plant growth (Körner 2003; Wright 2005). That plants growing under elevated [CO₂] often have higher [CH₂O] has also been taken as additional evidence for this idea (Körner 2003). Nevertheless, recent advances in our understanding of the signalling of growth responses in plants, in particular interactions between sugar and plant hormone signalling with nutrients and other growth limitations (Rolland *et al.* 2006), show that higher [CH₂O] is, in fact, usually associated with a stimulation of sink activity (i.e. faster rates of growth). Thus, increases in plant tissue [CH₂O] with higher [CO₂] do not mean that plant growth is not 'carbon' limited (Masle *et al.* 1990) and sugar signalling provides a simple explanation for why increased [CO₂] usually causing increases in both [CH₂O] and N_P . For a sugar sensing mechanism to work, it must be the case that, on average, any change in [CH₂O] gives rise to a less than proportional change in N_P . Conceptually this bears a strong resemblance to the general economic theory of Keynes (1936)—in particular the notion that in order for an economic system in which savings occur to be able to operate, it is necessary for the long-term ratio of expenditure to income to always be less than unity. In that respect, the need for plants to maintain considerable CH₂O reserves as insurance against drought, defoliation by pests or unexpected shading by competitors should not be discounted.

It is seedlings, saplings and trees growing under shaded conditions that tend to be the most carbon limited (Wright 2005) and plants adapted to and/or growing in shade are the most responsive to elevated [CO₂] (Curtis & Wang 1998; Kerstiens 2001). A strong sensitivity of tropical forest seedling growth to elevated [CO₂] has also been demonstrated (Würth *et al.* 1998), consistent with carbohydrate storage enhancing shade and stress tolerances for tropical forest seedlings (Myers & Kitajima 2007).

It thus seems likely to us that the currently observed accelerating dynamics of Amazon forests can reasonably be attributed to increases in [CO₂], mediated at the seedling stage, although other factors such as

changing light levels may, of course, also be involved (Wright 2005). Observations of increased growth being followed by increased mortality rates (Lewis *et al.* 2004) are also both conceptually and quantitatively consistent with ecosystem-level stimulations of G_p and N_p associated with slowly increasing [CO₂] (Lloyd & Farquhar 1996). Although this provides a plausible mechanism for the observed accelerating dynamics of tropical forests, there must be a limit to the maximum size that any forest can attain. Our inability to understand the basis of variations in aboveground carbon stocks for all but the driest Amazon forests (Saatchi *et al.* 2007) currently limits our understanding of how long any sequestration is likely to continue.

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ELECTRONIC SUPPLEMENTARY MATERIAL

A: DESCRIPTION OF PHOTOSYNTHESIS MODEL

The stomatal model of Jarvis & Davies (1998) can be written (Buckley *et al.* 2003) as

$$g = \frac{G(A_m - A)}{1 + sD(A_m - A)} \quad , \quad (\text{E1})$$

where g is the stomatal conductance to water vapour diffusion, A is the observed net CO₂ assimilation rate, A_{max} is the value of A at saturating intercellular CO₂ concentration, c_p , and G is the maximum possible stomatal conductance which occurs when $D = 0$ and $A \rightarrow 0$, and s is a constant describing the response of g to changes in D , these assumed to be mediated via a direct sensing of the leaf evaporation rate (Mott & Parkhurst 1991). As discussed by Buckley *et al.* (2003) some sort of surrogate measure of guard cell [ATP], τ , may in fact be more appropriate than $(A_m - A)$ as a measure of how much faster CO₂ could be fixed if stomata did not limit its supply.

As shown by Farquhar & Wong (1984) and Buckley *et al.* (2003) τ may be modelled as taking on two different values: τ_c which applies when the ribulose biphosphate (RuBP) saturated rate of carboxylation, W_c , is greater than the rate which can be sustained by the current rate of electron transport, W with the alternative value, τ_j applying when $W_j < W_c$. As written for equations (A22) to (A24) in Buckley *et al.* (2003)

$$\tau_c = a_t - \kappa \frac{W_c}{W_j} \quad , \quad (\text{E2})$$

$$\tau_j = \frac{(a_t - \kappa) \left(\frac{V_r}{V_{\text{max}}} - 1 \right)}{\left[\frac{W_c}{W_j} \right] \frac{V_r}{V_{\text{max}}} - 1} \quad , \quad (\text{E3})$$

and

$$\tau \equiv \tau_o + \tau_c \quad \text{if } W_c > W_j \quad ,$$

$$\tau \equiv \tau_o + \tau_j \quad \text{else} \quad . \quad (\text{E4})$$

In equations (E2), (E3) and E4, a_t represents the total concentration of adenylates in the chloroplast (equal to $\tau + [\text{ADP}]$), κ is the concentration of photophosphorylation sites, V_r is the CO₂ and Rubisco saturated potential rate of carboxylation (i.e. the carboxylation rate that would occur if carboxylation were limited by the potential RuBP pool size only), V_{max} is the rate of carboxylation when limited by

Rubisco activity only (i.e. saturated with both CO₂ and RuBP) and τ_o represents a basal ATP level provided by other processes such as mitochondrial respiration.

We first rewrite equation (E1) in terms of τ for the RuBP saturated case as

$$g = \frac{G \tau_c / (\tau_o + a_t)}{1 + sD\tau_c / (\tau_o + a_t)} \quad , \quad (\text{E5})$$

where the scaling ensures that $g = G$ when the guard cell [ATP] supply is at its maximum possible value. Noting also that one can write (Buckley *et al.* 2003)

$$g = \frac{(0.23 + 1.37\omega)A}{c_a - p_i / p_t} \quad . \quad (\text{E6})$$

with ω being the ratio of the total and stomatal conductances to water vapour, c_a the ambient concentration of CO₂ and with p_i and p_t being the intercellular CO₂ partial pressure and the total ambient pressure respectively, combining equations (E2),(E4),(E5) and (E6), we obtain

$$\frac{(0.23 + 1.37\omega)A}{c_a - p_i / p_t} = \frac{G\tau_c}{\tau_o + a_t + sD\tau_c} \quad . \quad (\text{E7})$$

We then define A and τ_c in terms of their underlying biochemistry. As shown by Farquhar *et al.* (1980);

$$A = \left(1 - \frac{\Gamma^*}{p_i}\right) \cdot \min\{W_c, W_j\} - R_d \quad , \quad (\text{E8})$$

where Γ^* is the photorespiratory compensation point and with W_c and W_j expressed as

$$W_c = \frac{V_{\max} p_i}{p_i + K_c(1 + p_{O_2} / K_o)} \quad , \quad (\text{E9})$$

and

$$W_j = \frac{J p_i}{4(p_i + 2\Gamma^*)} \quad , \quad (\text{E10})$$

Combining equations (E2), (E4), (E5), (E7), (E8) and (E9), we obtain for the case where $W_c < W_j$

$$\begin{aligned} & \frac{(0.23 + 1.37\omega)V_{\max}(p_i - \Gamma^*)}{[p_i + K_c(1 + p_{O_2} / K_o)](c_a - p_i / p_t)} \\ &= \frac{G \left(\tau_o + a_t - \kappa \frac{4V_{\max}(p_i + 2\Gamma^*)}{J[p_i + K_c(1 + p_{O_2} / K_o)]} \right)}{\tau_o + a_t + sD \left(\tau_o + a_t - \kappa \frac{4V_{\max}(p_i + 2\Gamma^*)}{J[p_i + K_c(1 + p_{O_2} / K_o)]} \right)} \quad . \quad (\text{E11}) \end{aligned}$$

For which it is possible to solve numerically for p_i and hence stomatal conductance, g using the approach outlined in Appendix 3 of Buckley *et al.* (2003). Note that in equation (E11) we have ignored the respiratory term of equation (E8) on the basis that, especially at high leaf temperatures, foliar

respiration is substantially inhibited in the light (Atkin *et al.* 2000). Likewise for the case where $W_j < W_c$ we write, also ignoring the R_d term

$$\frac{(0.23 + 1.37\omega)J(p_i - \Gamma^*)}{4(p_i + 2\Gamma^*)(c_a - p_i / p_t)} = \frac{G \left\{ \tau_o + a_t - \kappa \left(\frac{V_r}{V_{\max}} - 1 \right) / \left(\left[\frac{W_c}{W_j} \right] \frac{V_r}{V_{\max}} - 1 \right) \right\}}{\tau_o + a_t + sD \left\{ \tau_o + a_t - \kappa \left(\frac{V_r}{V_{\max}} - 1 \right) / \left(\left[\frac{W_c}{W_j} \right] \frac{V_r}{V_{\max}} - 1 \right) \right\}} \quad (E12)$$

which can also be solved numerically.

B. MODEL PHOTOSYNTHETIC PARAMETERS AND THEIR TEMPERATURE SENSITIVITIES

Based on the work of Domingues *et al.* (2004) we take a V_{\max} at 25 °C, $V_{m(25)}$, of 80 $\mu\text{mol m}^{-2} \text{s}^{-1}$ with the maximum rate of electron transport at 25 °C taken as $1.9V_{m(25)}$. The temperature sensitivity of V_{\max} is parameterised as in Bernacchi *et al.* (2003) but using the kinetic constants of von Caemmerer *et al.* (1994) calculated on the assumption that the leaf internal conductance to the diffusion to CO_2 is infinite, *visz* $K_c = 40.4 \text{ Pa}$ and $K_o = 24.8 \times 10^3 \text{ Pa}$. The temperature sensitivity of electron transport is as in June *et al.* (2004) with the dependence of the electron transport rate, J upon incoming irradiance (I) being described as the hyperbolic minimum of the J_{\max} and the product of I and F where F is the product of leaf absorbtivity to PAR and the effective quantum yield (Farquhar & Wong 1984).

As in Buckley *et al.* (2003) we take $\kappa = 2.5 |V_{m(25)}| \text{ mmol sites m}^{-2}$, $a_t = 12.6 |V_{m(25)}| \text{ mmol AxP m}^{-2}$ where the $|V_{m(25)}|$ indicates a numerical value only . i.e. = $V_{m(25)} / (\mu\text{mol m}^{-2} \text{s}^{-1})$, and with τ_o set to 1.6 mmol ATP m^{-2} . The ratio V_r / V_{\max} was taken as 2.27 (Farquhar & Wong 1984) and assumed to be independent of temperature. Based on observed stomatal responses to D as observed for Amazon forest from eddy covariance data (Mercado 2007) we took G as 0.6 $\text{mol m}^{-2} \text{s}^{-1}$ with $s = 0.122 \text{ mol}^{-1} \text{ mol}$.

One additional feature of our model is that it takes into account the observation that although the rate of electron transport through photosystem II may (reversibly) decline at leaf temperatures, T_L , above that optimal for electron transport, T_{opt} (June *et al.* 2004); this is also associated with an increase in the cyclic flow of electrons around photosystem I which probably serves as an important mechanism for the protection of both PS II and lipid membranes under high temperature conditions (Sharkey & Schrader 2006), as well as the maintenance of high ATP levels at supraoptimal T_L (Schrader *et al.* 2004). Thus in the simulations here, we simply set τ_j equal to the τ_j calculated to occur at T_{opt} for all $T_L > T_{\text{opt}}$. This is an important feature of the model which still requires experimental verification in terms of

stomatal responses to temperatures that are supraoptimal in terms of whole chain electron transport itself.

C. DRIVING VARIABLES AND THE LEAF ENERGY BUDGET

The model, which includes a simple energy balance as described in Lloyd *et al.* (1995), is run for a single sun exposed leaf at the top of the canopy and on an hourly basis using modelled values for air temperature, absolute humidity, wind-speed and incoming shortwave radiation in for both 2000 and 2040. Driving data for the hourly simulations in 2000 came from an updated version of New *et al.* (2000). For 2040, estimates were obtained as the difference between Hadley Centre GCM values for 2040 and 2000 added to the New *et al.* (2002) climatology values for 2000. In both cases hourly values were obtained using the climate generator which is part of the IMOGEN program (Huntingford *et al.* 2004). Boundary layer conductances and leaf energy budgets, also allowing for forced convection, were estimated as described in the Appendix of Ball *et al.* (1994) with an average leaf area for the Manaus tower site taken as 21 cm² (S. Patiño *et al.* unpublished data). The wind speed taken at the top of the canopy (where our theoretical leaf resides) was taken directly from the IMOGEN output.

D. A NOTE ON STOMATAL RESPONSES TO CO₂

Although not explicitly included in our model, equation (2) gives rise to stomatal responses to ambient [CO₂] through the τ term in equation (E2). This is because [ATP] decline as [CO₂] increase. Nevertheless, the response in the model is complex, with stomata tending to be *relatively* less responsive to [CO₂] at high light and at high D (Buckley *et al.* 2003). The degree of stomatal closure in response to an increase in [CO₂] from 380 $\mu\text{mol mol}^{-1}$ to 550 $\mu\text{mol mol}^{-1}$ is thus quite small in our simulations. Clarke (2004) have, however, suggested that an increase in leaf temperatures associated with such stomatal closure may be critical in reducing tropical tree photosynthesis, perhaps even pushing some trees beyond their thermal limits, this being akin to the notion of “stomatal suicide” (Randall *et al.* 1996). We have thus tested the potential likelihood of this effect by reducing g_{max} by 25% (i.e. to 0.48 mol m⁻² s⁻¹) and rerunning the fully interactive 2040 scenario with [CO₂] = 550 $\mu\text{mol mol}^{-1}$. This gives rise to a substantial reduction in the simulated Gross Primary Productivity, G_p (reduced from 271.1 to 232.9 mol C m⁻² a⁻¹) due to substantially lower p_i , but only marginal increases in the maximum simulated leaf temperature and leaf-to-air vapour pressure deficit (from 39.7 to 40.4 °C and from 40.7 to 43.7 mmol mol⁻¹ respectively). Although photosynthetic rates are substantially reduced by imposed stomatal closure in response to higher [CO₂], the simulated G_p for 2040 at an ambient [CO₂] of 550 $\mu\text{mol mol}^{-1}$

is still substantially higher than for 2000 for which the ambient $[\text{CO}_2] = 380 \mu\text{mol mol}^{-1}$. According to these simulations then, there is no reason to assume that any stomatal closure at higher $[\text{CO}_2]$ should push tropical tree leaves dangerously close to their thermal limit or reduce their photosynthetic productivity below what is currently the case.

E. RESPIRATION, TEMPERATURE AND TROPICAL FOREST PRODUCTIVITY

Feeley *et al.* (2007) reported that, especially for their Pasoh site, a significant decline in community-level relative growth rates (RGR) of around 50% (for trees > 10 cm diameter at breast height) may have been attributable to increased respiration rates associated with an increase in minimum daily air temperatures of at most $0.7 \text{ }^\circ\text{C}$ over a 14 year period. One interesting question then is what the temperature sensitivity of respiration would have to be for this hypothesis to hold.

This can be simply calculated by first modifying equation (1) in the main text,

$$N_p = G_p(1 - \phi_o - \phi_m) \quad , \quad (\text{E13})$$

where ϕ_o represents the loss of carbon associated with the conversion of photosynthate to organic matter (typically around 0.2 and insensitive to temperature – see for example Lloyd & Farquhar 1996) and ϕ_m quantifies the loss of carbon through “maintenance” respiratory processes, also expressed as a fraction of G_p . Taking a recent estimate for ϕ_m of 0.45 (Malhi *et al.* 2008) and with ϕ_o as 0.2 (and so with φ in equation (1) of the main text equal to 0.65), this means that ϕ_m would have to increase from 0.45 to 0.625 (i.e. by $\sim 39\%$) in order for N_p to decline by about 50%, suggesting relative sensitivity of maintenance respiration to temperature, B , of approximately $0.39/0.70 \sim 0.56 \text{ }^\circ\text{C}^{-1}$. From such a calculation we can easily estimate Q_{10} as $\exp[10B]$ for which we obtain a Q_{10} for plant respiration of considerably more than 30.

Such a calculation is based on the assumption that the decline in stem productivity observed is proportionally the same throughout the entire plant. It may be, however, that new stem and structural root growth represent only the carbohydrate “leftovers” once carbohydrate for new leaf and fine root production have been utilised (Lloyd & Farquhar 1996). In which case, the required increase in ϕ_m would be considerably less. But even with only about 30% of new growth going into boles and structural roots, but with new growth associated with new leaves, branches and fine roots (which generally constitute about 70% of N_p (Malhi *et al.* 2008) conserved, then the decrease in overall N_p would be only ca 15% with a temperature sensitivity for ϕ_m of $\sim 0.17 \text{ }^\circ\text{C}^{-1}$. This leads to a Q_{10} of greater than 5 which still seems too high. Nevertheless, if that were to actually be the case, then it is worth pointing out that just looking at stem growth rates must also be considered as giving a greatly amplified

view of any changes in overall tree growth rates with time. And by corollary this also applying to any observed increases such as reported in Baker *et al.* (2004).

We further note that is by no means clear that long-term temperature effects will be of the same magnitude as interannual variations (Atkin *et al.* 2005), but even if so, for the projected increase in air temperatures between 2000 and 2040 being about 1.5 °C, then with a $Q_{10} = 2.3$ for Amazon forest respiration (Meir *et al.* 2001) and with no acclimation (and using the parameters above) ϕ_m would be expected to increase only by about 14% from 0.45 to around 0.51; i.e. φ in equation (1) of the main text would increase from 0.65 to 0.71. Thus, even taking a worse case scenario by allowing for a 25% reduction in stomatal conductances and no acclimation of respiration to increasing temperatures at all by 2040, then N_p would only decline by about 7% with the most probable value almost most certainly being much less than this and more likely a significant increase. For example, at the other extreme (assuming no stomatal closure and allowing for full acclimation of plant respiration) then N_p would be modelled to increase by about 33% between 2000 and 2040 and with stem growth rates actually doubling over that 40 year period if it were to be the case that all increased N_p is channelled towards boles and fine roots (as discussed above).

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