

SHORT REPORT

Open Access



Effects of senolytic drugs on human mesenchymal stromal cells

Clara Grezzella^{1†}, Eduardo Fernandez-Rebollo^{1†}, Julia Franzen¹, Mónica Sofia Ventura Ferreira², Fabian Beier² and Wolfgang Wagner^{1*} 

Abstract

Background: Senolytic drugs are thought to target senescent cells and might thereby rejuvenate tissues. In fact, such compounds were suggested to increase health and lifespan in various murine aging models. So far, effects of senolytic drugs have not been analysed during replicative senescence of human mesenchymal stromal cells (MSCs).

Methods: In this study, we tested four potentially senolytic drugs: ABT-263 (navitoclax), quercetin, nicotinamide riboside, and danazol. The effects of these compounds were analysed during long-term expansion of MSCs, until replicative senescence. Furthermore, we determined the effect on molecular markers for replicative senescence, such as senescence-associated beta-galactosidase staining (SA- β -gal), telomere attrition, and senescence-associated DNA methylation changes.

Results: Co-culture experiments of fluorescently labelled early and late passages revealed that particularly ABT-263 had a significant but moderate senolytic effect. This was in line with reduced SA- β -gal staining in senescent MSCs upon treatment with ABT-263. However, none of the drugs had significant effects on the maximum number of population doublings, telomere length, or epigenetic senescence predictions.

Conclusions: Of the four tested drugs, only ABT-263 revealed a senolytic effect in human MSCs—and even treatment with this compound did not rejuvenate MSCs with regard to telomere length or epigenetic senescence signature. It will be important to identify more potent senolytic drugs to meet the high hopes for regenerative medicine.

Keywords: Senolytic drugs, Senescence, Mesenchymal stromal cells, DNA methylation, Telomere attrition, ABT-263, Quercetin, Danazol, Nicotinamide riboside

Introduction

Senolytic drugs hold the perspective to specifically target senescent cells and thereby to rejuvenate tissues or organisms [1]. Several compounds have been suggested to possess senolytic effects, including navitoclax (ABT-263) [2], quercetin [1], danazol [3], and nicotinamide riboside [4]. ABT-263 inhibits BCL-2 protein family members, which are crucial regulators of the apoptosis pathway [2, 5, 6]. ABT-263 was shown to deplete senescent cells of human umbilical vein epithelial cells (HUVECs), IMR90 human lung fibroblasts, and murine embryonic fibroblasts, but not human primary pre-

adipocytes [5]. Danazol is a synthetic androgen with telomere elongating capacity, which has been used to target accelerated telomere attrition—a hallmark of aging and senescence [3]. Quercetin is a proteasome activator with anti-oxidant properties [7] that triggers apoptosis via the BCL-2 pathway [1]. Nicotinamide riboside increases levels of nicotinamide adenine dinucleotide (NAD⁺). Aged mice supplemented with nicotinamide riboside revealed increased lifespan and rejuvenated muscle stem cells [4].

Primary cells undergo a limited number of divisions before entering the state of replicative senescence. The process of senescence induces changes in morphology, metabolism, secretory phenotype, and differentiation potential of cells, thereby having a significant impact on experimental outcomes and affecting their therapeutic potential [8]. This applies particularly to mesenchymal stromal cells (MSCs), which raise high hopes in tissue

* Correspondence: wwagner@ukaachen.de

†Equal contributors

¹Helmholtz-Institute for Biomedical Engineering, Stem Cell Biology and Cellular Engineering, RWTH Aachen University Medical School, Pauwelsstraße 20, 52074 Aachen, Germany

Full list of author information is available at the end of the article

engineering and are concurrently tested in a multitude of clinical trials [9]. MSCs comprise a multipotent subset of cells, capable of differentiation towards osteogenic, chondrogenic, and adipogenic lineages. The selective removal of senescent MSCs from cultures might improve standardization and effectiveness of cell preparations for cell therapeutics in regenerative medicine. We have therefore directly compared the senolytic capacity of ABT-263, quercetin, danazol, and nicotinamide riboside in human MSCs during long-term culture.

Methods

Cell culture

Mesenchymal stromal cells were isolated from the femoral bone marrow of three donors after orthopaedic surgery. All samples were taken after informed and written consent and the study was approved by the ethics committee of RWTH Aachen University Medical School (permit number: EK300/13). Culture medium consisted of Dulbecco's modified Eagle medium (DMEM, 1 g L⁻¹ glucose; PAA, Pasching, Austria) supplemented with 1% penicillin/streptomycin (PAA), 1% L-glutamine, 10% pooled human platelet lysate that was generated as described previously [10], and 0.1% heparin (5000 IU ml⁻¹; Ratiopharm, Ulm, Germany). Cells were cultured at 37 °C, in an atmosphere containing 5% CO₂, and passaged by trypsinization when they reached 90% confluency with a re-seeding density of 10,000 cells cm⁻². The immunophenotype and three-lineage differentiation potential of MSCs was validated as described previously [10, 11].

Senolytic compounds

Cells were treated either with ABT-263 (Selleck Chemicals, Houston, TX, USA), quercetin (Sigma-Aldrich, St. Louis, MO, USA), nicotinamide riboside (ChromaDex, Irvine, CA, USA), or danazol (Sigma-Aldrich). These compounds were dissolved in DMSO (VWR, Radnor, PA, USA) (ABT-263), ethanol (VWR) (quercetin and danazol), or phosphate-buffered saline (Sigma-Aldrich) (nicotinamide riboside), according to the manufacturer's instructions, and subsequently diluted in cell culture medium to working concentrations of 10 μM (ABT-263), 100 μM (quercetin and danazol), or 20 mM (nicotinamide riboside). If not stated otherwise, the compounds were applied only for 3 days.

Viability assays

Viability was measured with the ATPlite Kit (PerkinElmer, Waltham, MA, USA) according to the manufacturer's instructions. In addition, viability was measured with the AlamarBlue™ Cell Viability Reagent (ThermoFisher Scientific, Waltham, MA, USA) and the results were always consistent.

Co-culture of senescent and non-senescent cells

Mesenchymal stromal cells at early (passage 3) and late (before proliferation arrest) passages of the same donor were simultaneously taken into culture and stained with the PKH67 Green Fluorescent Cell Linker Kit and PKH26 Red Fluorescent Cell Linker Kit, respectively (both Sigma-Aldrich). These cells were subsequently mixed at a 1:1 ratio and treated with each of the four drugs for 1 day. Fluorescence was subsequently analysed by fluorescence microscopy (EVOS; Life Technologies GmbH, Darmstadt, Germany) or quantified on a FACS Canto II (BD, Heidelberg, Germany).

Senescence-associated beta-galactosidase staining

This assay was performed with the Senescence Detection Kit (Abcam, Cambridge, UK) according to the manufacturer's instructions.

Telomere length measurement

Genomic DNA was isolated with the NucleoSpin® Tissue kit (Macherey-Nagel, Düren, Germany). Telomere length was determined by monochrome multiplex quantitative PCR as described previously [12]. In brief, C_T values of telomere qPCR were compared to the corresponding C_T values of a single-copy gene qPCR of the same sample. The resulting T/S ratio correlates with the number of telomeric repeats.

Epigenetic senescence signature

Genomic DNA was isolated as already described and bisulfite converted with the EZ DNA Methylation™ Kit (Zymo Research, Irvine, CA, USA). DNA methylation was analysed by pyrosequencing at six senescence-associated CG dinucleotides (SA-CpG sites) related to the genes *GRM7*, *CASP14*, *CASR*, *SELP*, *PRAMEF2*, and *KRTAP13-3*, as described previously [13]. In brief, bisulfite-converted DNA was analysed on a PyroMark Q96 ID System (Qiagen, Hilden, Germany) and results were evaluated with the PyroMark CpG SW 1.0 software (Qiagen). The DNA methylation values were then implemented into linear regression models to predict passage numbers [14].

qRT-PCR analysis

RNA was isolated via a NucleoSpin RNA extraction kit (Macherey-Nagel) and analysed with a NanoDrop ND-1000 spectrophotometer (Thermo Scientific, Waltham, MA, USA). Semi-quantitative qRT-PCR was conducted with *Power SYBR* green master mix (Applied Biosystems, Foster City, CA, USA) and the following primers: *β-actin*, forward GGCACCACCTTCTACAAT and reverse AACATGATCTGGGTTCATCTTCTC; *p16*, forward GGTCGGGTAGAGGAGGT and reverse ATCATCATGACCTGGATCGG; *IL-6*, forward AGACAGCCACTCACCTCTT and reverse ACTCTTGTTACATGTC

TCCTTTCTC; *PAI-1*, forward CTGGTGAATGCCCTC TACTTC and reverse GGC GTGGTGA ACTCAGTATAG. Results were normalized to β -actin.

Statistics

All experiments were performed with three independent biological replicas and the results are presented as mean \pm standard deviation (SD). Statistical significance was estimated by two-tailed paired Student's *t* test.

Results

In order to evaluate the specificity of these four different senolytic drugs we first tested the half-maximal effective concentration (EC50) for human MSCs (see Additional file 1: Figure S1). Based on that, we determined working concentrations for subsequent experiments: 10 μ M for ABT-263,

100 μ M for quercetin, 100 μ M for danazol, and 20 mM for nicotinamide riboside. Similar concentrations have been used previously for other cell types [1, 4, 5, 15]. However, it needs to be taken into account that the relatively high concentration of DMSO, which is required as solvent for ABT-263, can already exert toxic effects (see Additional file 1: Figure S2). To address whether the potentially senolytic drugs target specifically senescent MSCs we conceived a simple co-culture experiment: MSCs at early passages (passage 3) were labelled with PKH67 in green and corresponding cells at late passages (at proliferation arrest) were labelled with PKH26 in red. These cells were mixed 1:1, treated with the drugs for 1 day, and then analysed by fluorescence microscopy and flow cytometry (Fig. 1a, b; three biological replicas). ABT-263 treatment significantly reduced the proportion of senescent MSCs ($p = 0.018$; 8.1%

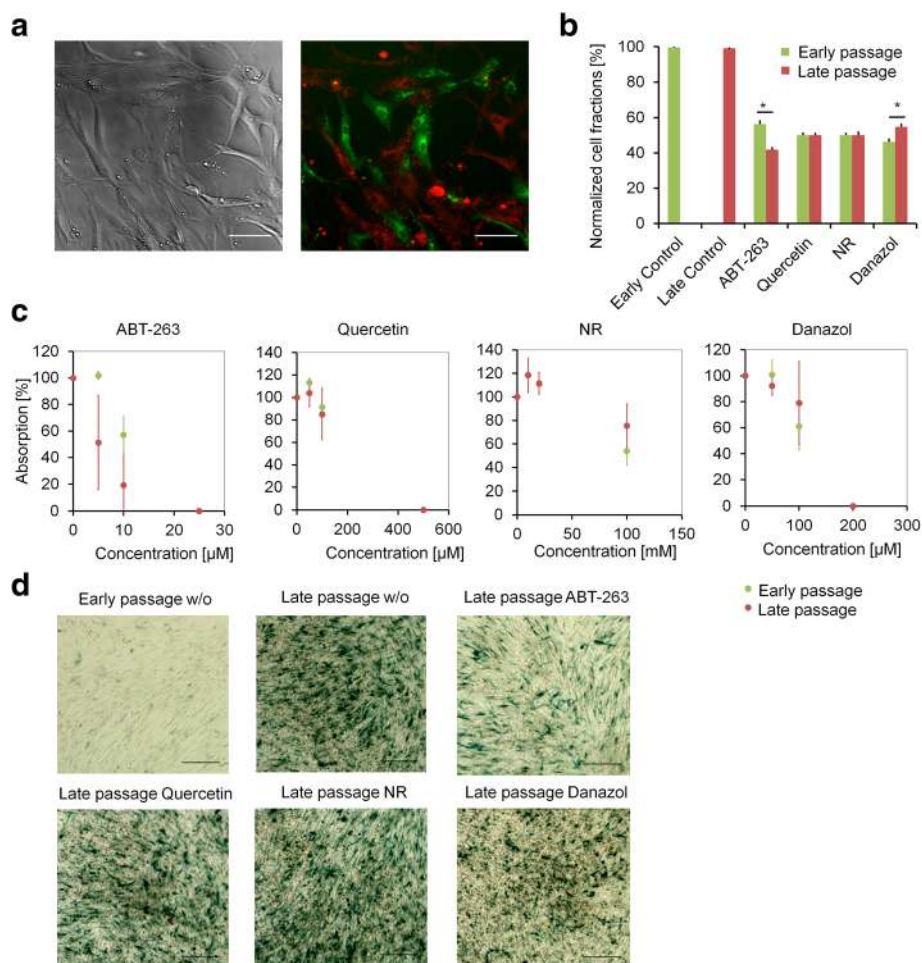


Fig. 1 ABT-263 has senolytic effects in human MSCs. **a** Exemplary phase-contrast and fluorescence microscopic images of non-senescent cells (PKH67, green, passage 3) and senescent cells (PKH26, red, passage 12). w/o = without additional drug; Scale bars = 100 μ m. **b** Normalized fractions of cells determined by flow cytometric quantification of red (senescent) and green (non-senescent) cells indicate that particularly ABT-263 reduced senescent cells (normalized to untreated controls, $n = 3$, mean \pm SD; * $p \leq 0.05$). **c** Dose-response curves analysed after 3 days of treatment with drugs and viability estimated by flow-cytometric assays (normalized to untreated controls, $n = 3$, mean \pm SD; individually assessed for early and late passage cells). **d** SA- β -gal staining performed either in non-senescent (passage 3) or senescent (passage 12) MSCs upon treatment with senolytic drugs for 3 days (exemplary images depicted, scale bars = 200 μ m). NR nicotinamide riboside

less senescent cells), although the specificity for senescent cells was relatively low. In contrast, quercetin and nicotinamide riboside did not affect the ratio of early and late passage cells and danazol even slightly increased the proportion of late passage cells ($p = 0.037$; 4.7% more senescent cells). Alternatively, we analysed dose–response curves in MSCs either at early or late passages ($n = 3$). These results support the notion that the toxic effect of ABT-263 was more pronounced on late passage cells (Fig. 1c). Furthermore, staining of senescence-associated beta-galactosidase (SA- β -gal) in MSCs at late passages was particularly reduced upon treatment with ABT-263 (Fig. 1d). Taken together, ABT-263 showed a mild senolytic effect in MSCs.

Removal of senescent cells might eliminate paracrine factors that trigger growth arrest [16] and thereby delay replicative senescence during culture expansion. To evaluate the effects on long-term growth curves of MSCs we supplemented each of the four drugs for either 3 days at passage 3 or during the entire culture period. Upon short-term treatment with quercetin and danazol, the maximal number of cumulative population doublings (cPDs) declined, probably due to the toxic effects, whereas nicotinamide riboside and ABT-263 did not impact on culture expansion (Fig. 2a; $n = 3$). For long-term

treatment we applied 50% reduced concentrations of the drugs and all of them accelerated the proliferation arrest (Fig. 2b; $n = 3$). Thus, none of these potentially senolytic drugs prolonged the culture expansion of MSCs.

Subsequently, we analysed the effect of the four senolytic drugs on molecular markers of senescence. Expression of the senescence-associated genes cyclin-dependent kinase inhibitor 2A (*CDKN2A* or *p16*), interleukin 6 (*IL-6*), and plasminogen activator inhibitor-1 (*PAI-1*) increased at later passages and was even slightly increased upon treatment with senolytic drugs (see Additional file 1: Figure S3). Telomere length was estimated by ratios in monochrome multiplex quantitative PCR, and showed a significantly decline in untreated MSCs during culture expansion ($p = 0.006$) [12]. Yet none of the senolytic drugs increased the average telomere length at later passages (Fig. 2c). Alternatively, we analysed the effect of the senolytic drugs on senescence-associated epigenetic modifications. DNA methylation changes at specific CpG sites are acquired in a highly reproducible manner during replicative senescence of MSCs. We have previously demonstrated that such epigenetic changes provide a robust surrogate marker to estimate the state of replicative senescence [13]. In fact, the results of our six CpG epigenetic senescence signature revealed good correlation of

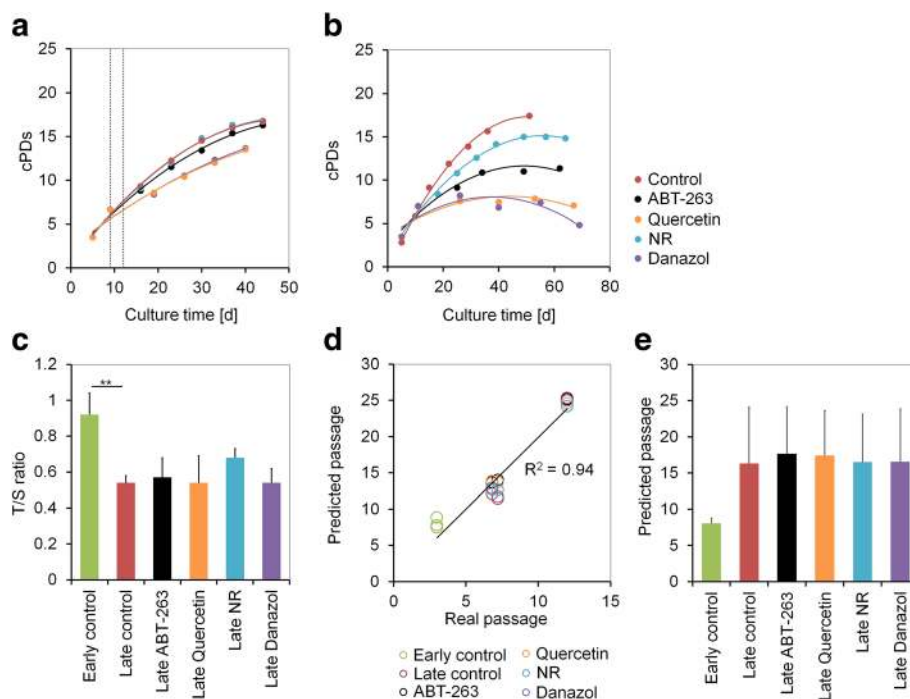


Fig. 2 Treatment with senolytic drugs did not support long-term expansion or affect molecular markers of senescence. Long-term growth curves of cumulative population doublings (cPDs) **a** after initial pulse treatment for 3 days (d; period indicated by dashed lines) and **b** with continuous treatment with drugs (results exemplarily depicted for one donor). **c** Relative telomere length measured as telomere to single copy gene (T/S) ratio (monochrome multiplex qPCR) of non-senescent and senescent MSCs with and without senolytic drug treatment ($n = 3$, mean \pm SD; $**p \leq 0.01$). **d** Comparison of real and predicted passage numbers in non-senescent and senescent MSCs with and without senolytic drug treatment. **e** Average number of predicted passages directly compared for the different treatments ($n = 3$, mean \pm SD). NR nicotinamide riboside

predicted and real passage numbers ($R^2 = 0.94$), although passage numbers were generally over-estimated. Notably, none of the tested drugs had significant impact on the senescence-associated DNA methylation changes (Fig. 2d, e).

Discussion

Recently, senolytic drugs raised high hopes for regenerative medicine, aging, and age-related diseases [1]. Quercetin has previously been demonstrated to be effective against senescent human endothelial cells and mouse MSCs [1]; ABT-263 was shown to specifically target senescent human HUVECs or fibroblasts [5]; nicotinamide riboside and danazol have been demonstrated to rejuvenate senescent stem cells [3, 4]. However, none of these compounds have been systematically analysed in human MSCs.

In this study, ABT-263 revealed moderate selectivity for senescent MSCs but also affected non-senescent cells, as described previously [17]. For quercetin, danazol, and nicotinamide riboside we did not observe any senolytic effect. Furthermore, none of the investigated drugs had a positive effect on culture expansion, telomere length, or epigenetic rejuvenation. The discrepancy from the aforementioned reports might be due to the different cell type used in this study. Thus, there is a need to identify more potent senolytic compounds for human MSCs. Recently some additional candidates were described [18, 19] and it is also conceivable that combinations of different drugs could better reduce the burden of senescent cells with less side effects. We used relatively high concentrations of the drugs as we anticipated that hereby the senolytic effect would be more pronounced. On the other hand, lower concentrations would allow reduction of toxic solvents and facilitate discerning the impact on survival and proliferation. Other cell cycle regulators, such as CDKN2B (p27) and the retinoblastoma proteins (RB1, RB2/P130, and P107), are well known to be functionally relevant for senescence of human MSCs [20]. Furthermore, senescence of MSCs is associated with several other changes—for example, in the secretome, autophagy, and metabolism—and these aspects should also be analysed. In addition, it might be conceivable that a senolytic compound modifies the senescence-associated secretory phenotype (SASP) without directly rejuvenating the cells. However, none of the investigated drugs had any positive effect on either the key cytokine IL-6 or PAI-1, which was shown to be part of crucial SASP regulation pathways [21]. The concept of senolytic drugs is attractive, but further research is required to specifically target the senescent cells within MSC preparations.

Additional file

Additional file 1: Figure S1. Dose–response curves for EC50 determination. **Figure S2.** Dose–response curves of solvent controls. **Figure S3.** Expression analysis of senescence-associated genes. (PDF 1049 kb)

Abbreviations

cPD: Cumulative population doubling; DMEM: Dulbecco's modified Eagle medium; EC50: Half-maximal effective concentration; HUVEC: Human umbilical vein epithelial cell; MSC: Human mesenchymal stromal cell; SA- β -gal: Senescence-associated beta-galactosidase staining; SD: Standard deviation

Acknowledgements

The authors thank all of the patients in this study for their collaborative participation.

Funding

This work was supported by the Else Kröner-Fresenius-Stiftung (2014_A193), by the German Research Foundation (WA 1706/8-1), and by the German Ministry of Education and Research (01KU1402B).

Availability of data and materials

All data generated or analysed during this study are included in this published article.

Authors' contributions

CG, JF, EF-R, and WW designed the study. CG, JF, and EF-R performed experiments and analysed and formatted the data. MSVF and FB performed telomere length measurements. CG, JF, EF-R, and WW wrote the first draft of the manuscript and all authors read, edited, and approved the final manuscript.

Ethics approval and consent to participate

Mesenchymal stromal cells were isolated from the femoral bone marrow of three donors after orthopaedic surgery. All samples were taken after informed and written consent and the study was approved by the ethics committee of RWTH Aachen University Medical School (permit number: EK300/13).

Competing interests

RWTH Aachen Medical School has applied for a patent for the Epigenetic-Senescence-Signature. WW is cofounder of Cygenia GmbH that can provide services for this assay (www.cygenia.com). Apart from these, the authors declare no competing financial interests.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Author details

¹Helmholtz-Institute for Biomedical Engineering, Stem Cell Biology and Cellular Engineering, RWTH Aachen University Medical School, Pauwelsstraße 20, 52074 Aachen, Germany. ²Department for Hematology, Oncology, Hemostaseology and Stem Cell Transplantation, Faculty of Medicine, RWTH Aachen University, Aachen, Germany.

Received: 13 February 2018 Revised: 26 March 2018

Accepted: 28 March 2018 Published online: 18 April 2018

References

- Zhu Y, Tchkonina T, Pirtskhalava T, Gower AC, Ding H, Giorgadze N, et al. The Achilles' heel of senescent cells: from transcriptome to senolytic drugs. *Aging Cell*. 2015;14:644–58.
- Chang J, Wang Y, Shao L, Laberge R-MM, Demaria M, Campisi J, et al. Clearance of senescent cells by ABT263 rejuvenates aged hematopoietic stem cells in mice. *Nat Med*. 2016;22:78–83.
- Townsend DM, Dumitriu B, Liu D, Biancotto A, Weinstein B, Chen C, et al. Danazol treatment for telomere diseases. *N Engl J Med*. 2016;374:1922–31.
- Zhang H, Ryu D, Wu Y, Gariani K, Wang X, Luan P, et al. NAD⁺ repletion improves mitochondrial and stem cell function and enhances life span in mice. *Science*. 2016;352:1436–43.

5. Zhu Y, Tchkonja T, Fuhrmann-Stroissnigg H, Dai HM, Ling YY, Stout MB, et al. Identification of a novel senolytic agent, navitoclax, targeting the Bcl-2 family of anti-apoptotic factors. *Aging Cell*. 2016;15:428–35.
6. Willis S, Day CL, Hinds MG, DCS H. The Bcl-2-regulated apoptotic pathway. *J Cell Sci*. 2003;116:4053–6.
7. Chondrogianni N, Kapeta S, Chinou I, Vassilatou K, Papassideri I, Gonos ES. Anti-ageing and rejuvenating effects of quercetin. *Exp Gerontol*. 2010;45:763–71.
8. Wagner W, Horn P, Castoldi M, Diehlmann A, Bork S, Saffrich R, et al. Replicative senescence of mesenchymal stem cells: a continuous and organized process. *PLoS One*. 2008;3:e2213.
9. Wagner W, Ho AD, Zenke M. Different facets of aging in human mesenchymal stem cells. *Tissue Eng Part B Rev*. 2010;16:445–53.
10. Horn P, Bokermann G, Cholewa D, Bork S, Walenda T, Koch C, et al. Impact of individual platelet lysates on isolation and growth of human mesenchymal stromal cells. *Cytotherapy*. 2010;12:888–98.
11. Fernandez-Rebollo E, Mentrup B, Ebert R, Franzen J, Abagnale G, Sieben T, et al. Human platelet lysate versus fetal calf serum: these supplements do not select for different mesenchymal stromal cells. *Sci Rep*. 2017;7:5132.
12. Ventura Ferreira MS, Crysandt M, Ziegler P, Hummel S, Wilop S, Kirschnner M, et al. Evidence for a pre-existing telomere deficit in non-clonal hematopoietic stem cells in patients with acute myeloid leukemia. *Ann Hematol*. 2017;96:1457–61.
13. Koch CM, Jousseen S, Schellenberg A, Lin Q, Zenke M, Wagner W. Monitoring of cellular senescence by DNA-methylation at specific CpG sites. *Aging Cell*. 2012;11:366–9.
14. Franzen J, Zirkel A, Blake J, Rath B, Benes V, Papantonis A, et al. Senescence-associated DNA methylation is stochastically acquired in subpopulations of mesenchymal stem cells. *Aging Cell*. 2017;16:183–91.
15. Podhorecka M, Macheta A, Chocholska S, Bojarska-Junak A, Szymczyk A, Goracy A, et al. Danazol induces apoptosis and cytotoxicity of leukemic cells alone and in combination with purine nucleoside analogs in chronic lymphocytic leukemia. *Ann Hematol*. 2016;95:425–35.
16. Coppé J-P, Desprez P-Y, Krtolica A, Campisi J. The senescence-associated secretory phenotype: the dark side of tumor suppression. *Annu Rev Pathol*. 2010;5:99–118.
17. Baar MP, Brandt RMC, Putavet DA, Klein JDD, Derks KWJ, Bourgeois BRM, et al. Targeted apoptosis of senescent cells restores tissue homeostasis in response to chemotoxicity and aging. *Cell*. 2017;169:132–147.e16.
18. Zhu Y, Doornebal EJ, Pirtskhalava T, Giorgadze N, Wentworth M, Fuhrmann-Stroissnigg H, et al. New agents that target senescent cells: the flavone, fisetin, and the BCL-XL inhibitors, A1331852 and A1155463. *Aging (Albany NY)*. 2017;9:955–63.
19. Fuhrmann-Stroissnigg H, Ling YY, Zhao J, McGowan SJ, Zhu Y, Brooks RW, et al. Identification of HSP90 inhibitors as a novel class of senolytics. *Nat Commun*. 2017;8:422.
20. Alessio N, Capasso S, Ferone A, Di Bernardo G, Cipollaro M, Casale F, et al. Misidentified human gene functions with mouse models: the case of the retinoblastoma gene family in senescence. *Neoplasia*. 2017;19:781–90.
21. Özcan S, Alessio N, Acar M, Mert E, Omerli F, Peluso G, et al. Unbiased analysis of senescence associated secretory phenotype (SASP) to identify common components following different genotoxic stresses. *Aging (Albany NY)*. 2016;8:1316–29.

Submit your next manuscript to BioMed Central and we will help you at every step:

- We accept pre-submission inquiries
- Our selector tool helps you to find the most relevant journal
- We provide round the clock customer support
- Convenient online submission
- Thorough peer review
- Inclusion in PubMed and all major indexing services
- Maximum visibility for your research

Submit your manuscript at
www.biomedcentral.com/submit

