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https://doi.org/10.1021/acs.jmedchem.8b00734

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Journal of Medicinal Chemistry

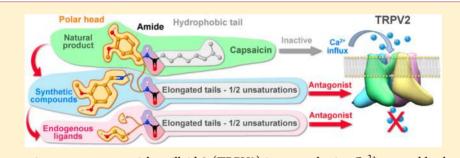
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¹ Elongation of the Hydrophobic Chain as a Molecular Switch: ² Discovery of Capsaicin Derivatives and Endogenous Lipids as Potent ³ Transient Receptor Potential Vanilloid Channel 2 Antagonists

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13 **(S)** Supporting Information



14 **ABSTRACT**: The transient receptor potential vanilloid 2 (TRPV2) is a nonselective Ca²⁺ permeable channel member of the

15 TRPV subfamily, still considered an orphan TRP channel due to the scarcity of available selective and potent pharmacological

16 tools and endogenous modulators. Here we describe the discovery of novel synthetic long-chain capsaicin derivatives as potent

17 TRPV2 antagonists in comparison to the totally inactive capsaicin, the role of their hydrophobic chain, and how the structure-

18 activity relationships of such derivatives led, through a ligand-based approach, to the identification of endogenous long-chain 19 fatty acid ethanolamides or primary amides acting as TRPV2 antagonists. Both synthetic and endogenous antagonists exhibited

20 differential inhibition against known TRPV2 agonists characterized by distinct kinetic profiles. These findings represent the first

example of both synthetic and naturally occurring TRPV2 modulators with efficacy in the submicromolar/low-micromolar

range, which will be useful for clarifying the physiopathological roles of this receptor, its regulation, and its targeting in

23 pathological conditions.

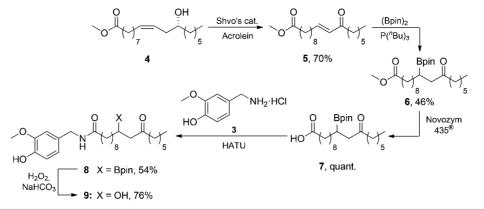
1. INTRODUCTION

²⁴ TRPV2 belongs to the polymodal transient receptor potential ²⁵ (TRP) superfamily of calcium-permeable nonselective cation ²⁶ channels, activated by a wide variety of physical and chemical ²⁷ stimuli. Due to its mechanosensor property, TRPV2 is ²⁸ considered a stretch-modulated channel and a regulator of ²⁹ calcium homeostasis in different tissues and organs, in ³⁰ particular the heart, where it is 10-fold more abundant than ³¹ in skeletal muscle.¹ Different lines of evidence suggest for ³² TRPV2 a key role in physiological cardiac function as well as in ³³ cardiomyopathies and dystrophic diseases.^{2–4} Besides the ³⁴ heart, TRPV2 is also found in the brain, vascular smooth ³⁵ muscle cells, the gastrointestinal tract, macrophages, and the ³⁶ urothelial tract, ⁵ and it is involved in a number of physiopathological processes, 6 including cancer, $^{7-9}$ particularly $_{37}$ of the urinary tract. $^{10-13}$ 38

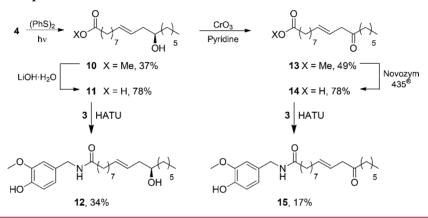
Despite its biological and pharmacological relevance, ³⁹ TRPV2 is still considered an orphan TRP channel due to ⁴⁰ the scarcity of selective drugs and known endogenous ligands. ⁴¹ The 2-aminoethoxydiphenyl borate (2APB) is one of the first ⁴² nonselective activators identified for rat TRPV2 (EC₅₀ = 129 ⁴³ μ M),¹⁴ although inactive at the human orthologue, suggesting ⁴⁴ a strong species specificity.^{15,16} *Cannabis sativa* derivatives such ⁴⁵ as Δ^9 -tetrahydrocannabinol (Δ^9 -THC), cannabidiol (CBD), ⁴⁶ and Δ^9 -tetrahydrocannabivarin (Δ^9 -THCV) are TRPV2 ⁴⁷ activators^{17,18} and so is *p*-(di-*n*-propylsulfamyl)benzoic acid ⁴⁸

Received: May 7, 2018 Published: September 3, 2018

Scheme 1. Synthesis of Compound 9



Scheme 2. Synthesis of Compounds 12 and 15



⁴⁹ (probenecid).¹⁹ However, all these agonists are known to ⁵⁰ modulate other TRP channels. Most TRPV channels are ⁵¹ proposed to be modulated also by phosphoinositide lipids.²⁰ ⁵² TRPV2-mediated Ca²⁺ influx has been reported following ⁵³ stimulation by endogenous lysophospholipids such as ⁵⁴ lysophosphatidylcoline (LPC) and lysophosphatidylinositol ⁵⁵ (LPI),²¹ LPC being a relatively potent activator (EC₅₀ = 3.4 ⁵⁶ μ M).²² To date, the nature of endogenous regulators of ⁵⁷ TRPV2 activity still remains elusive.²³

Also synthetic inhibitors of TRPV2 are either not specific or 59 endowed with low potency, as exemplified by ruthenium red 60 (IC₅₀ = 0.6 μ M),²⁴ a pore blocker that inhibits other 12 ion 61 channels;²⁵ La³⁺ and Gd³⁺;²⁶ citral;²⁷ the alkylated imidazole 62 SKF96365;¹⁶ tetraethylammonium and 4-aminopyridine, two 63 potassium channel blockers; 1-(2-(trifluoromethyl)phenyl)-64 imidazole, an inhibitor of capacitative Ca²⁺ entry;¹⁶ and 65 tranilast,²⁸ which has been used in several studies;^{29–34} even 66 though it has never been validated as TRPV2 antagonist.

⁶⁷ TRPV2 shares high sequence identity (>50%) with TRPV1, ⁶⁸ but its threshold of activation by temperature is higher (>52 ⁶⁹ $^{\circ}$ C)²⁴ and, unlike TRPV1, is not sensitive to capsaicin. The ⁷⁰ recently solved cryo-EM structures of both TRPV1 and ⁷¹ TRPV2,^{35,36} along with mutagenesis and computational ⁷² studies, showed that the TRPV1 binding site of capsaicin is ⁷³ not conserved in TRPV2. Furthermore, the replacement of ⁷⁴ critical residues leads to a mutant (TRPV2-Quad) against ⁷⁵ which capsaicin behaves as an antagonist rather than an agonist ⁷⁶ as in TRPV1.³⁷ These intriguing results prompted us to ⁷⁷ investigate a series of capsaicin bears a longer alkyl ⁷⁹ chain, featuring different length, unsaturation degree, and type of polar substituents. The structure–activity relationship 80 (SAR) of these synthetic compounds then suggested the 81 screening of structurally related endogenous lipids sharing at 82 least one functional group with the capsaicin derivativs, with 83 the aim of finding new endogenous modulators. 84

2. RESULTS

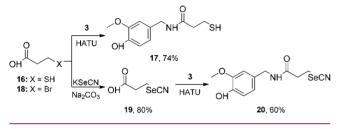
2.1. Synthesis. Commercial fatty acids such as ricinoleic 85 acid, oleic acid, and palmitic acid were used as starting material 86 to synthesize the 23 compounds tested. Scheme 1 shows the 87 s1 synthesis of the $\alpha_{,\beta}$ -unsaturated ketone 5 by the ruthenium- 88 catalyzed oxidation in anhydrous toluene of the homoallylic 89 alcohol of the methyl ricinoleate 4.38 Shvo's catalyst and 90 acrolein were used as catalyst and hydrogen scavenger, 91 respectively.³⁹ The addition of bis(pinacolato)diboron 92 $(Bpin)_2$ to the enone 5 in the presence of tri-*n*-butylphosphine 93 $(P("Bu)_3)^{40}$ yielded the β -boronketone **6** in 46% yield. 94 Enzymatically controlled hydrolysis⁴¹ of the methyl ester 6 95 with Novozym 435 lipase led to the carboxylic acid 7 96 quantitatively. This acid 7 was coupled, without any further 97 purification, with 4-hydroxy-3-methoxybenzylamine hydro- 98 chloride 3 by HATU⁴² and DIPEA in anhydrous DMF, 99 achieving the amide 8. The oxidative hydrolysis of the boron 100 substituent of the compound 8 led to the β -hydroxyketone 9 in 101 a 76% yield (Scheme 1).

The irradiation of alcohol 4 with diphenyl sulfide⁴³ in 103 isooctane in a photochemical reactor for 3 h led to the isomer 104 10 in 37% yield after several recrystallizations at -30 °C. This 105 compound was used to synthesize two new long-chain N- 106 vanillylamides (12, 15). The hydrolysis of the methyl ester of 107

108 10 led to the corresponding carboxylic acid 11. The 109 subsequent coupling of 11 with the 4-hydroxy-3-110 methoxybenzylamine hydrochloride 3 using the same con-111 ditions described above yielded compound 12 in a 34% yield. 112 Compound 10 was also oxidized with CrO_3 in pyridine⁴⁴ to 113 prepare the *trans* ketone 13 (49% yield), which was 114 enzymatically hydrolyzed to synthesize the corresponding 115 acid 14 in a 78% yield. Subsequently, 14 was coupled with the 116 vanillyl amine 3 to yield the (*E*)-*N*-(4-hydroxy-3-methoxyben-117 zyl)-12-oxooctadec-9-enamide 15 after purification by liquid 118 column chromatography (17% yield) (Scheme 2).

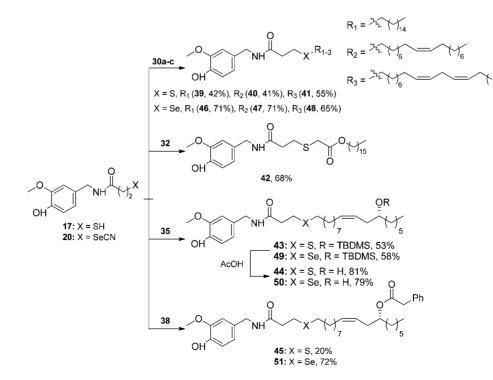
Scheme 3 shows the synthesis of the sulfur- and selenotion derivatives of 3. Mercaptopropionic acid 16 was coupled with

Scheme 3. Synthesis of Sulfur and Seleno Intermediates



121 4-hydroxy-3-methoxybenzylamine hydrochloride **3** using 122 HATU and DIPEA in anhydrous DMF, achieving the amide 123 **17** (74% yield). The synthesis of the seleno-derivatives started 124 with bromopropionic acid **18**, which was treated with KSeCN 125 in water: The neutralization with Na₂CO₃, yielded the 126 selenocyanatopropionic acid **19** in 80% without purification. 127 Finally, compound **19** was coupled with the 4-hydroxy-3-128 methoxybenzylamine hydrochloride **3** to obtain compound **20** 129 after purification by liquid column chromatography (60% 130 yield).

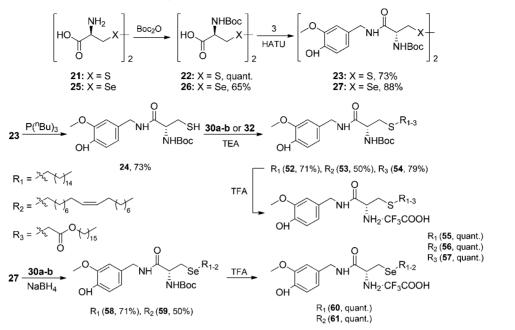
Scheme 4. Synthesis of No-Branched Sulfur- and Seleno-Derivatives



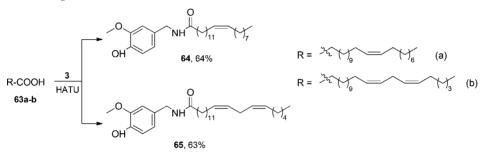
Amide 17 was S-alkylated with the previously synthesized 131 alkylating derivatives 30a-c, 32, and 35 (see Supporting 132 Information) in DMF and triethylamine, obtaining the long- 133 chain N-vanillylamides 39-43 and 45 in 41-68% yield. N- 134 Vanillylamide 44 was successfully achieved after removing the 135 TBDMS protecting group with acetic acid at room temper- 136 ature (81% yield). New long-chain N-vanillylamides were 137 obtained from compound 20, which was first treated with 138 NaBH₄ in ethanol at room temperature to remove the cyano 139 protection and regenerate the selenol group.⁴⁵ Subsequent Se- 140 alkylation was carried out in one-pot with the addition of 141 diverse set of alkylating reagents (30a-c, 35 ,and 38). N- 142 Vanillylamides 46-49 and 51 were synthesized in 71-87% 143 vields. Compound **50** was successfully prepared after removing 144 the TBDMS protecting group with acetic acid at room 145 temperature (79% yield) (Scheme 4). 146 s4

Scheme 5 shows the synthesis of amino-branched analogues. 147 s5 The first step consisted of the treatment of L-cystine 21 or L- 148 selenocystine 25 with Boc₂O in the presence of triethylamine 149 to afford the protected derivatives $\hat{\mathbf{22}}^1$ and $\mathbf{26}^2$ (quantitative 150 and 65% yield, respectively).46,47 These compounds were 151 coupled with 4-hydroxy-3-methoxybenzylamine hydrochloride 152 3 using EDCI. HOBt, and triethylamine (TEA) in anhydrous 153 DMF, achieving the amides 23 and 27 (74% and 88% yield). 154 The reduction of compound 23 with $P(^{n}Bu)_{3}$ in wet 155 dichloromethane afforded compound 24 in a 73% yield after 156 purification by liquid column chromatography. New long-chain 157 N-vanillylamides were afforded from compound 24, which was 158 S-alkylated with the previously synthesized alkylating deriva- 159 tives 30a-c and 32 in the presence of triethylamine, obtaining 160 the long-chain N-vanillylamides 52, 53, and 54 in moderate 161 yields (50-79% yield). The N-Boc deprotection was carried 162 out using trifluoroacetic acid⁴⁸ in dichloromethane yielding N- $_{163}$ vanillylamides 55, 56, and 57 as trifluoroacetic salts in 164 quantitative yields. Compound 27 was reduced with NaBH₄ 165

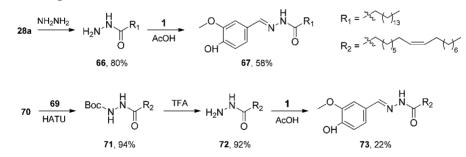




Scheme 6. Synthesis of Compounds 64 and 65



Scheme 7. Synthesis of Compounds 67 and 73



¹⁶⁶ in ethanol at room temperature to cleave the diselenium ¹⁶⁷ bond.⁴⁹ The Se-alkylation was carried out with the addition of ¹⁶⁸ the alkylating derivatives **30a,b** to afford the *N*-vanillylamides ¹⁶⁹ **58** and **59** in 74–88% yields. Finally, The *N*-Boc deprotection ¹⁷⁰ was carried out using the same conditions described above to ¹⁷¹ afford the *N*-vanillylamides **60** and **61** as trifluoroacetic salts. ¹⁷² Acids **63a,b**, which were previously obtained from the ¹⁷³ hydrolysis of their respective methyl esters **62a,b** (see ¹⁷⁴ Supporting Information), were coupled with the 4-hydroxy-¹⁷⁵ 3-methoxybenzylamine hydrochloride **3** using HATU and ¹⁷⁶ DIPEA in anhydrous DMF, achieving the amides **64** and **65** ¹⁷⁷ after purification by liquid column chromatography (64% and ¹⁷⁸ **63%** yield) (Scheme **6**). Methyl palmitate **28a** was treated with an excess of $_{179}$ hydrazine hydrate in ethanol to synthesize the palmitic acid $_{180}$ hydrazide **66** (80% yield). The addition of the aromatic $_{181}$ aldehyde vanillin **1** to compound **66** in the presence of acetic $_{182}$ acid in reflux conditions gave the Schiff's base compound **67** in $_{183}$ 58% yield. ⁵⁰A similar compound was synthesized starting from $_{184}$ oleaic acid **70**, which was coupled to *tert*-butyl hydrazine- $_{185}$ carboxylate **69** using HATU and DIPEA in DMF to yield the $_{186}$ oleylhydrazide **71** in a 94% yield. The *N*-Boc deprotection of $_{187}$ oleylhydrazide **71** with TFA in DCM for 2 h led to $_{188}$ oleylhydrazide **72** in 92% yield. Compound **72** refluxed with $_{189}$ vanillin **1** in the presence of acetic acid in methanol produced $_{190}$ the Schiff base **73** in 22% yield (Scheme 7).

f1 f1

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2.2. Biological Evaluation. 2.2.1 Capsaicin Derivatives Activate TRPV1 Channel. The capsaicin scaffold (Figure 194 1)⁵¹ can be ideally divided into three regions: head, neck, and

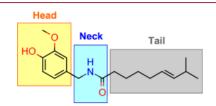


Figure 1. Chemical structure of capsaicin. The vanillyl head, the amide neck, and hydrophobic tail are shaded in yellow, cyan, and gray, respectively.

195 tail, formed by the vanillyl moiety, the amidic group, and the
196 lipophilic alkyl chain, respectively. Structural variations,
197 including incorporation of sulfur atom, into the head and the
198 neck regions have been described in the literature.⁵²⁻⁵⁵

Instead, the effect of a sulfur atom in the alkyl chain has been 199 200 less investigated. The recent availability of the 3D structure of 201 TRPV1⁵⁶ along with mutagenesis studies⁵⁷ allowed the 202 identification of the capsaicin binding site, where the alkyl 203 chain is hosted in a phenylalanine-rich hydrophobic region close to Thr550, a residue involved in H-bond interaction with 204 205 the ligand amide group. The presence of a sulfur atom near the 206 neck region should in principle lead to an increment of activity 207 due to favorable dipole-dipole and aromatic-sulfur inter-208 actions. Since sulfur can be substituted with selenium via 209 isosteric replacement, we also synthesized the corresponding 210 selenium analogs. Selenium is an essential trace element whose 211 role in medicine and biology is just starting to be elucidated. 212 Some selenium-containing compounds have provided protec-213 tion against many degenerative conditions, including cancer. 214 Thus, a series of novel capsaicin derivativs, i.e., 9, 12, 15, 39, 215 46, 55, 60, 42, 57, 44, 56, 40, 45, 65, 41, 48, 64, 47, 61, 51, 216 50, 67, 73, whose structures are reported in Tables 1 and 2, 217 featuring the same "head" and "neck" as capsaicin but differing 218 in length and nature of the hydrocarbon tail, were tested on 219 human TRPV1 heterologously expressed in human embryonic 220 kidney (HEK)-293 cells by fluorometric assay (see Tables S1 221 and S2 in Supporting Information). The predicted activities as 222 TRPV1 agonists were confirmed for many compounds within 223 the series, exhibiting EC₅₀ values from high-nanomolar to 224 subnanomolar range. A SAR analysis of the results also 225 disclosed the critical role of the region flanking the amide 226 group in modulating the activity. In fact, the insertion of a 227 positive charge next to the amide group was detrimental for 228 activity (compounds 55-57 and 60), and the introduction of 229 an imido group between the aromatic moiety and the amido 230 group led to totally inactive compounds (compounds 67 and 73). Conversely, the introduction of a single polar substituent 231 232 (hydroxyl, ester, or ketone) was well-tolerated, and the 233 introduction of a sulfur or selenium atom in the hydrophobic 234 tail even improved the activity. However, on the basis of the 235 antagonist activity exhibited by capsaicin on TRPV2-Quad,³⁷ 236 the new compounds were also tested on TRPV2 to determine 237 if the elongation and the functionalization of the alkyl chain 238 could elicit a functional response at this receptor.

239 **2.2.2. Capsaicin Derivatives Inhibit TRPV2 Channels** 240 **Activated by LPC.** The activity of the synthesized capsaicin 241 derivativs on TRPV2 was evaluated in vitro. The assays were 242 conducted using a fluorometric assay with rat TRPV2 heterologously expressed in HEK-293 cells. The tested 243 compounds did not significantly activate TRPV2-mediated 244 Ca^{2+} elevation in transfected HEK-293 cells. Instead, 245 preincubation (5 min) of TRPV2-HEK-293 cells with different 246 concentrations of the tested compounds, followed by 247 incubation with LPC (3 μ M), caused inhibition of intracellular 248 Ca^{2+} elevation due to TRPV2 response to LPC. The 249 corresponding IC₅₀ values are reported in Table 1. 250

The structure-activity relationships (SARs) of these 251 compounds suggested a critical influence on the capability to 252 exert TRPV2 antagonism of the alkyl chain and, in particular, 253 of its hydrophobicity, length, and degree of unsaturation. 254 Hydrophobicity is important since, as shown in Table 1, the 255 activity dramatically dropped after introduction in the chain of 256 polar substituents such as hydroxyl, keto, or ester groups 257 (these groups arising from esterification of the hydroxyl group) 258 or their combinations (42, 44, 50, 45, 51, 9, 12, 15). However, 259 the presence of an amino group next to the amide (55, 60, 56, 260 61), which had marginal effects for already active compounds, 261 by only slightly increasing their potency (60 vs 46), was 262 instead dramatic for those inactive compounds bearing a 263 hydroxyl or an ester moiety in the alkyl chain, whose activity 264 was completely rescued (see 42 vs 57). The complete recovery 265 of activity after introduction of an amino group next to the 266 amide in derivatives bearing a polar substituent in the alkyl 267 chain suggests that reinforcement of the polar interactions of 268 the "head" avoids the competition with the polar-substituted 269 alkyl chain for interaction with receptor polar residues in a 270 region where the polar head, but not the alkyl chain, should be 271 hosted to elicit a measurable effect. The chain is fairly more 272 tolerant to changes not substantially affecting the hydro- 273 phobicity of the alkyl group: replacement of sulfur with 274 selenium in the alkyl chain did not affect significantly ligand 275 activity (39 vs 46); its replacement with a carbon atom 276 determined an increase in potency (64 vs 40/47). While polar 277 functionalization of the alkyl chain caused a dramatic drop of 278 activity, amino or imino groups (67, 73) were well tolerated in 279 the region close to the amide moiety of capsaicin. In particular, 280 the imino derivatives were among the most active compounds 281 within the series (IC₅₀ = 0.28 and 0.12 μ M, respectively). Also 282 length and unsaturation degree of the alkyl chain significantly 283 affected the activities of the tested compounds. The C16:0 and 284 C18:0 saturated analogs were inactive, whereas the C20:0 285 derivative showed an IC₅₀ = 3.1 μ M. The insertion of a single 286 double bond in C18 chain (olvanil) dramatically increased the 287 antagonism, with $IC_{50} = 0.16 \ \mu M$. 288

Thus, the screening led to the identification of several very 289 potent TRPV2 antagonists, exhibiting IC_{50} values in the 290 subnanomolar to low-micromolar range. This result is quite 291 remarkable since, despite its close homology to TRPV1, 292 TRPV2 is insensitive to capsaicin, the residues being 293 responsible for capsacin binding and receptor activation in 294 TRPV1 not conserved in TRPV2.⁵⁸ 295

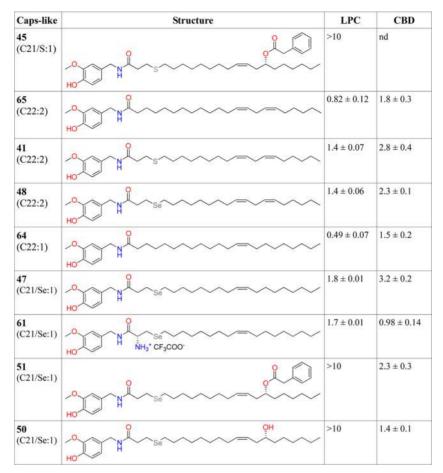
The most striking result from the SAR of capsaicin derivativs 296 against LPC is that the elongation of the alkyl chain of 297 capsaicin causes a switch of such scaffold from inactivity 298 toward potent antagonism at rat recombinant TRPV2. 299 Intriguingly, the dependence of TRPV2 modulation on the 300 length of the ligand alkyl chain has already been observed for 301 lysophospholipids, which require a carbon chain longer than 302 C12 to stimulate the receptor.²¹ 303

2.2.3. Capsaicin Derivatives Inhibit TRPV2 Channels 304 Activated by CBD. Due to different latency in the activation 305 f2

Caps-like	Structure	LPC	CBD
Palvanil (C16:0) ^a	HO	>10	>10
Stevanil (C18:0)	HOLD H	>10	>10
Olvanil (C18:1)	HO	0.16±0.02	1.7±0.1
Livanil (C18:2)	HO	2.6±0.2	2.1±0.1
9 (C18:0)	HO HO	>10	>10
12 (C18:1)		>10	7.5 ± 1.3
15 (C18:1)	HO	>10	4.4 ± 0.3
Eicosavan illamide (C20:0)	HOL	3.1 ± 0.2	>10
39 (C19/S)	HOL	3.8 ± 0.8	nd ⁶
46 (C19/Se)	HO	4.3 ± 0.9	nd
55 (C19/S)	HO NH3* CF3COO.	1.4 ± 0.2	nd
60 (C19/Se)	HO NH3 ⁺ CF3COO	1.2 ± 0.03	nd
42 (C21/S/O)	содуна в то в т	>10	nd
57 (C21/S/O)	HO LOF 2COO	1.4 ± 0.1	nd
44 (C21/S:1)	HO HO STATES	>10	nd
56 (C21/S:1)	HO NH ₃ * CF ₃ COO'	1.9 ± 0.1	nd
40 (C21/S:1)		2.5 ± 0.1	nd

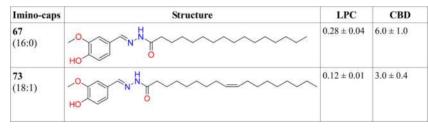
Table 1. Antagonist Potency of Capsaicin-like Compounds at TRPV2 against LPC (3 μ M) and CBD (2 μ M), Reported as IC₅₀ (μ M)

Table 1. continued



"In parentheses, number of C atoms in the alkyl chain followed by number of unsaturations. When heteroatom X occurs within alkyl chain, it is indicated as "/X". ^bnd: not determined.

Table 2. Antagonist Potency of Capsaicin-Imino Compounds at TRPV2 against LPC (3 μ M) and CBD (2 μ M), Reported as IC₅₀ (μ M)^{*a*}



^aIn parentheses, number of C atoms in the alkyl chain followed by number of unsaturations.

³⁰⁶ profile between LPC and cannabidiol (CBD) (see Figure 2), ³⁰⁷ we also investigated the effect of a representative panel of ³⁰⁸ capsaicin derivatives against CBD to ascertain whether the ³⁰⁹ inhibitory activity/potency would vary against agonists ³¹⁰ exhibiting different kinetics of action. Also in this case, the ³¹¹ assays were conducted using a fluorometric assay with ³¹² recombinant rat TRPV2 heterologously expressed HEK-293 ³¹³ cells. The preincubation (5 min) of TRPV2-HEK-293 cells ³¹⁴ with different concentrations of the tested compounds, ³¹⁵ followed by incubation with CBD (2 μ M), caused an ³¹⁶ inhibition of the Ca²⁺ elevation due to the TRPV2 response ³¹⁷ to CBD. The corresponding IC₅₀ values of the tested ³¹⁸ compounds are reported in Table 1. While the trend identified ³¹⁹ in LPC antagonism for capsaicin derivatives bearing all carbon ³²⁰ atoms, selenium, or sulfur was substantially conserved, a

f2

different behavior was observed with those derivatives ³²¹ featuring polar substituents (i.e., **50/51**), since their activity ³²² against CBD was not negatively affected by these functional ³²³ groups, as instead observed against LPC. The imino-derivatives ³²⁴ **67** and **73** (see Table 2), i.e., the two most active compounds ³²⁵ against LPC (0.28 and 0.12 μ M, respectively), were less potent ³²⁶ against CBD (IC₅₀ = 6.0 and 3.0 μ M, respectively). The trend ³²⁷ of activity of C16:0, C18:0, and C18:1 derivatives was similar ³²⁸ to that observed for LPC, although C18:1 (olvanil) was less ³²⁹ potent as an antagonist (IC₅₀ = 1.7 μ M), whereas, different ³³⁰ from what observed with LPC, C20:0 was totally inactive. ³³¹ These results demonstrate a dependence of the antagonist ³³² activity on the type of agonist against which antagonism is ³³³ tested. ³³⁴ 500

450

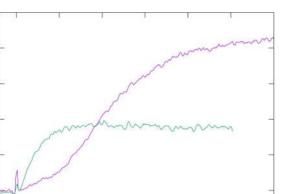
400

350

300

250

Intensity (a.u.)



LPC CBD 200 100 150 200 250 300 0 50 t(s)

Figure 2. TRPV2 is activated by LPC (3 μ M) and CBD (2 μ M). The graph shows the representative traces of $[Ca^{2+}]_i$ increase evoked by the two agonists in HEK293 cells overexpressing TRPV2.

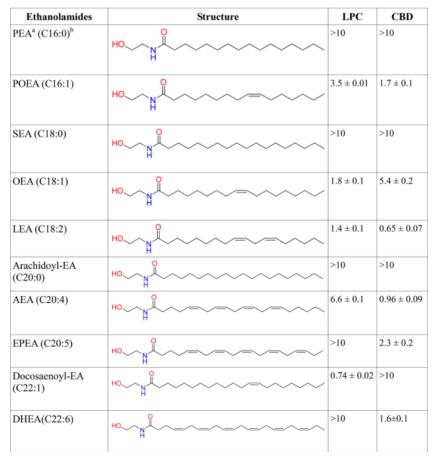
2.2.4. Evaluation of Endogenous Lipids as Potential 335 336 TRPV2 Antagonists. Since the activity of the tested 337 compounds appears to critically depend on the nature of 338 alkyl chain but is less affected by changes in the polar head, we

decided to ascertain the role of the head group of capsaicin, 339 i.e., the vanillyl moiety, by testing a series of naturally occurring 340 lipids bearing different polar heads and differing in length and 341 unsaturation of the alkyl chain in order to determine the 342 structural and functional requisites for TRPV2 modulation. 343

Article

2.2.5. Long-Chain Ethanolamides Exhibit Differential 344 Inhibition of TRPV2 upon Activation by LPC or CBD. To 345 evaluate the contribution of the aromatic moiety to the overall 346 activity, a panel of natural occurring ethanolamides differing in 347 length and unsaturation degree was tested for both agonism 348 and antagonism at TRPV2, using both LPC and CBD as 349 reference activators. Ethanolamides share with the tested 350 capsaicin derivativs the nature of both the alkyl chain and the 351 hydrophilic groups (amide and hydroxyl moieties) in the polar 352 head. The IC₅₀ values (against CBD 2 μ M and LPC 3 μ M) are 353 reported in Table 3. Ethanolamides featuring saturated alkyl 354 t3 chains, regardless of their lengths, were inactive against both 355 agonists, whereas the introduction of a single double bond was 356 sufficient to switch from inactivity to activity against both 357 agonists (see PEA vs POEA, or SEA vs OEA), similar to what 358 was already observed for capsaicin derivativs. However, while 359 the C20:0 capsaicin derivative was active against LPC, the 360 homolog ethanolamide was inactive. Moreover, while OEA was 361 less active than the counterpart olvanil, LEA was more potent 362 than livanil against both reference agonists. Increasing the 363

Table 3. Potency of Fatty Ethanolamides as Functional Antagonists at TRPV2 against LPC (3 μ M) and CBD (2 μ M), Reported as IC₅₀ (μ M)



"Abbreviations: EA, ethanolamide; PEA, palmitoyl ethanolamide; POEA, palmitoleoyl ethanolamide; OEA, oleoyl ethanolamide; LEA, lynoleoyl ethanolamide; arachidonoylethanolamide; EPEA, eicosapentaenoyl ethanolamide; DHEA, docosahexaenoyl ethanolamide. ^bIn parentheses, number of C atoms in the alkyl chain followed by number of unsaturations.

Table 4. Antagonist Potency of Fatty Amides at TRPV2 against LPC (3 µM) and CBD (2 µM), Reported as IC₅₀ (µM)

Amides	Structure	LPC	CBD
PA ^a (C16:0) ^b	H ₂ N	>10	>10
SA(C18:0)		>10	>10
OA (C18:1)	H ₂ N	2.1 ± 0.1	2.1 ± 0.2
LA (C18:2)		2.2 ± 0.1	1.2 ± 0.1
ErA (C22:1)	H ₂ N	0.67 ± 0.13	7.1 ± 0.7
Eicosanamide (C20:0)		>10	>10

^{*a*}Abbreviations: PA, palmitamide; SA, stearamide; OA, oleamide; LA, linoleamide; ErA, erucamide. ^{*b*}In parentheses, number of C atoms in the alkyl chain followed by number of unsaturations.

Table 5. Lack of Strong Antagonist Activity of Fatty Acids at TRPV2 against LPC (3 μ M) and CBD (2 μ M), Reported as IC₅₀ Values (μ M)

Acids	Structure	LPC	CBD
Palmitic acid (C16:0) ^a	Ю	>10	>10
Oleic acid (C18:1)	HOHO	>10	>10
Arachidic acid (C20:0)	но	>10	>10
Arachidonic acid (C20:4)	но	>10	>10
Erucic acid (C22:1)	но	>10	>10
Docosadienoic acid (C22:2)	но	>10	>10

^{*a*}In parentheses, number of C atoms in the alkyl chain followed by number of unsaturations.

Table 6. Slope Values from Linear Regression of Schild Analysis and t-Test Statistics

	LPC			CBD		
compd	slope ^a	N^{b}	P^{c}	slope ^a	N^{b}	P^{c}
61	-0.58 ± 0.087	4	<0.0024	-0.74 ± 0.048	4	< 0.002
olvanil	-0.77 ± 0.049	6	< 0.001	-0.55 ± 0.068	6	< 0.001
docosaenoyl-EA	-0.54 ± 0.046	6	< 0.001			
50				-0.63 ± 0.039	5	< 0.001

"Mean value \pm standard deviation. ^bNumber of experiments (each one performed at least in triplicate) used for Schild regression. ^cP values calculated from t test values for the "slope = 1 hypothesis".

³⁶⁴ number of double bonds increased the potency against CBD ³⁶⁵ but not LPC.

2.2.6. Long-Chain Primary Amides Exhibit Differafter ential Inhibition of TRPV2 Channels upon Activation by the LPC or CBD. To also evaluate the role of the hydroxyl group, we tested a series of amide derivatives. As for capsaicin- and the anolamine-derivatives, also for the amides the activity role thanolamine-derivatives, also for the amides the activity the activity bond. In particular, erucamide is active as TRPV2 antagonist with a potency comparable to that of its capsaicin derivative (0.67 vs 0.49 μ M) against LPC, but it is less potent than the $_{374}$ capsaicin counterpart against CBD (7.1 vs 1.5 μ M). As $_{375}$ observed with the ethanolamides, also the C20:0 amide $_{376}$ derivative was inactive against both activators (Table 4). $_{377}$ t4

2.2.7. Free Fatty Acids Are Poor Inhibitors of TRPV2 378 Channels. Finally, to investigate the role of the amide group, 379 we tested against both LPC and CBD a panel of long-chain 380 fatty acids, featuring alkyl chains comparable with those 381 occurring in the already-tested compounds. The results are 382 reported in Table 5. Fatty acids with alkyl chains from C16 up 383 t5 384 to C22 are by far less potent antagonists against both reference
385 agonists than the other classes of compounds bearing similar
386 alkyl chains, thus suggesting that the amide group is mandatory
387 for potent antagonism.

2.2.8. Schild Analysis on Selected TRPV2 Antagonists. 388 389 The effects of increasing concentrations of antagonist 61, 390 olvanil, and docosaenoyl-EA vs LPC and 61, olvanil, and 50 vs 391 CBD were tested against concentration-response curves of 392 LPC and CBD (where the effects of each concentration of 393 LPC and CBD were expressed as percent of their effect of 2 \times $394 \ 10^{-4}$ M in the absence of the antagonist) to calculate Schild's 395 plots. These compounds have been selected as representative 396 of antagonists active either against both activators (61, olvanil) 397 or selectively toward LPC (docosaenoyl-EA)/CBD (50) alone. 398 In all cases, the plots analyzed by linear regression gave slope 399 values significantly less than unity, as reported in Table 6, 400 indicative of a noncompetitive behavior. However, this result 401 may also be indicative of a nonequilibrium condition, and we 402 do not definitely rule out a competitive behavior.

3. DISCUSSION

403 Novel capsaicin derivativs, initially designed as TRPV1 404 agonists, behave as potent TRPV2 antagonists. The different 405 types of modifications introduced in these compounds 406 determine different agonist/antagonist profiles and, in 407 particular, opposite behaviors in terms of relative potency/ 408 efficacy within a derivative series on the two channels. In fact, 409 the insertion of a positive charge or an imido group close the 410 amido group, detrimental for TRPV1 agonism, is well-tolerated 411 for TRPV2 antagonism and even leads in some cases to an 412 increment or a rescue of activity. Conversely, the insertion of a 413 sulfur/selenium atom and/or the presence of a polar group, 414 which increases TRPV1 agonism, leaves unaffected, or even 415 decrease, TRPV2 antagonism.

Given the scarcity of known endogenous ligands for TRPV2, 416 417 the discovery of such long-chain capsaicin derivativs as potent 418 TRPV2 antagonists prompted us to investigate the following 419 classes of long-chain fatty acid derivatives with at least one 420 functional group in common with capsaicin derivatives as 421 potential TRPV2 modulators: (i) ethanolamides, (ii) primary 422 amides, and (iii) free fatty acids, to evaluate the role of the 423 amide group itself. Antagonists were found in both the 424 ethanolamide and primary amide, but not in fatty acid, series. Activities for both synthetic and endogenous ligands were 425 426 tested against either LPC or CBD as activators, since, on the 427 basis of their different kinetics of activation, CBD can be 428 defined as a direct TRPV2 agonist, whereas LPC induces 429 TRPV2 activation indirectly, via its G-protein-coupled 430 receptors and PI3,4 kinase mediated pathways.²¹ We found 431 that this different mode of activation is differentially counter-432 acted by the investigated compounds, which can be classified 433 as follows: (a) compounds endowed with similar antagonist 434 efficacy against both agonists, (b) compounds selectively active 435 against LPC, (c) compounds selectively active against CBD. 436 To determine the nature of antagonism, a Schild regression 437 was carried out for the representative members of each class, 438 i.e., olvanil, docoesanoyl-EA, and compound 50, and in all 439 three cases the antagonists behaved as noncompetitive ligands, 440 suggesting that these compounds may act as allosteric 441 antagonists. However, we cannot completely rule out a 442 competitive behavior since a Schild plot slope of <1 may 443 also suggest nonequilibrium conditions. Moreover, since the 444 hydrophobicity of the alkyl chain of the investigated

compounds is a critical requisite for LPC but not for CBD 445 inhibition, it is reasonable to speculate that a different binding 446 site is involved in LPC antagonism, with structural/functional 447 requisites different from those of CBD. This site might be 448 either on TRPV2 or on other targets activated by LPC in its 449 signaling cascade and would be the target of those compounds 450 selectively antagonizing activation by LPC. A common critical 451 requisite for activity of both ethanolamides and amides as 452 TRPV2 antagonists is the occurrence of at least one double 453 bond in the alkyl chain, since saturated lipids, regardless of the 454 length of their acyl chains, are totally inactive. This suggests 455 that a bent conformation of the alkyl chain is required for a 456 better accommodation into the active site, as previously 457 reported for other TRPV1 agonists.⁵⁹ Also C16:0 and C18:0 458 derivatives of capsaicin are inactive against both CBD and 459 LPC, whereas the C20:0 derivative is selectively active against 460 LPC. Instead, a different behavior is observed with imino- 461 capsaicin derivatives since they are active also when bearing 462 saturated alkyl chain. The aromatic moiety contributes to the 463 overall activity at TRPV2 of the compounds characterized in 464 the present work, since it occurs in the most active antagonists. 465

4. CONCLUSIONS

In summary, the search for structurally related synthetic or 466 endogenous lipids with structural similarity to capsaicin 467 derivative led to identification of olvanil and 73 as potent 468 TRPV2 antagonists against LPC (0.16 and 0.12 μ M, 469 respectively) and of LEA (linoleoylethanolamide) as potent 470 TRPV2 antagonist against CBD (0.65 μ M). This finding is 471 both surprising, since all other synthetic and endogenous 472 compounds tested here on TRPV2 behave as antagonists and 473 capsaicin is inactive at this channel, and of great physiological 474 importance, since novel potent endogenous antagonists were 475 been identified following this study.

In conclusion, starting from the testing of a series of 477 synthetic capsaicinoids as modulators of rat TRPV2, we 478 discovered not only new tools for the pharmacological 479 manipulation of the latter but also that previously described 480 endogenous lipids, i.e., long chain fatty acid ethanolamides and 481 primary amides, behave as negative modulators of this channel. 482 These data are of great potential importance given the 483 increasingly important role assigned to TRPV2 in temperature 484 sensing, pain, insulin secretion, immune response, muscle and 485 heart function, and cancer.⁵⁸

5. EXPERIMENTAL SECTION

5.1. Compounds. Stevanil, livanil, ethanolamides, amides, and 487 fatty acids when not described in the synthetic section have been 488 purchased from Cayman-Vinci Biochem. Palvanil and PEA are kind 489 gifts from Epitech Group SpA, Saccolongo, Padova, Italy, whereas 490 olvanil is a precious gift from Dr. Alberto Minassi, Dipartimento di 491 Scienze del Farmaco, Università del Piemonte Orientale, Novara, 492 Italy.

5.2. Synthetic Procedures. Reactions requiring anhydrous 494 conditions were performed in blazed or oven-dried glassware using 495 anhydrous solvents and under inert atmosphere (argon). The solvents 496 and reagents were purchase from Acros Organics, Sigma-Aldrich, 497 Fluka, Merk, Panreac, Strem Chemicals, or TCI Chemicals. 498 Petroleum ether, EtOAc, DCM, and MeOH were used without 499 further purification. In the case of anhydrous reactions, solvent and 500 reagents were properly dried. Acrolein was distilled at atmospheric 501 pressure and used immediately. The reactions were monitored until 502 completion by TLC on silica gel 60F-254 precoated plates (Merck). 503 Visualization of the compounds was performed by UV light (254 nm), 504

505 and staining was performed by immersion in a 5% solution of 506 concentrated H_2SO_4 in methanol or 5% w/v phosphomolibdic acid in 507 ethanol followed by heating. Flash column chromatography was 508 performed using silica gel (technical grade, 60 Å, 40–63 μ m) (Sigma-509 Aldrich) under air pressure. NMR spectra were recorded on a 510 MERCURYplus AS400 MHz Varian spectrometer. Chemical shifts 511 are reported in parts per million (ppm, δ units). Coupling constants 512 (J) are reported and expressed in hertz (Hz). Ssplitting patterns are 513 designated as br (broad), s (singlet), d (doublet), dd (double 514 doublet), t (triplet), q (quartet), dt (double triplet), td (triple 515 doublet), ddd (double double doublet), p (pentuplet), and m 516 (multiplet). All ¹³C NMR spectra were proton decoupled. High 517 resolution mass spectra (HR-MS) were recorded on at the Serveis 518 Cientificotècnics of Universitat de Lleida (SCT-UdL) and Servei de 519 Recursos Científics i Tècnics of Universitat Rovira i Virgili (URV) 520 with an Agilent G6510AA Q-TOF MS spectrometer in positive 521 electrospray ionization (ESI⁺) and Agilent LC1200 series coupled to 522 MS6210 TOF spectrometer in electrospray ionization (ESI⁺) 523 respectively. Mobile phase was composed of ACN/MeOH 50:50. 524 Flow rate: 0.6 mL/min. Infrared spectra were recorded on Jasco FT-525 IR 6300 using a diamond ATR crystal cell. Melting points were 526 measured using Gallenkamp capillary apparatus and are uncorrected. 527 Optical rotations were measured at 20 °C with a PerkinElmer 241 nc 528 polarimeter (λ = 589 Na, path length 1 dm). Some recorded values 529 were within the error limit of the polarimeter, and therefore it was not 530 possible to determine them. It has been indicated as $\left[\alpha\right]_{D}^{20} < 1^{\circ}$. 531 Analytical UPLC-MS was performed on a binary Acquity UPLC with 532 a Acquity PDA UPLC eLambda 800 nm triple quadrupole mass 533 spectrometer (Xevo TQ-S) using a Acquity UPLC BEH C18 50 × 2.1 534 mm, 1.7 μ m C18 column. UV detection = 210-500 nm, mass 535 spectrometry= ESI+ (scan 100-850 m/z). Flow rate was 0.3 mL/min 536 using a solvent gradient of B 100% over 6 min (total run time with 537 equilibration back to starting conditions = 2 min) where A = MeOH 538 and B = 85/15/0.2 MeOH/H2O/AcOH. Purities were measured by 539 UV absorption at 254 nm or TIC and are ≥95% unless otherwise 540 stated. Purity of final compounds was assessed by reversed-phase 541 UHPLC with UV diode array detection; all tested compounds were 542 >95% pure.

5.2.1. Procedure I. Amine Bond Formation. To a 0.35 M S44 solution of starting material in anhydrous DMF were added the amine S45 **3** (1.1 equiv), HATU (1.5 equiv), and DIPEA (3 equiv). The mixture S46 was stirred at room temperature for 20 h. To the mixture were added S47 EtOAc and brine, and the aqueous phase was extracted with EtOAc. S48 The combined organic phases were washed with 1 M HCl, saturated S49 solution of NaHCO₃ and brine. The organic phase was dried over S50 anhydrous Na₂SO₄, filtered and the solvent was removed under S51 reduced pressure. The crude residue was purified by silica gel column S52 chromatography.

553 5.2.2. Procedure II. Ester Hydrolysis. To a 0.2 M solution of 554 starting material in THF/H₂O (1:1) LiOH·H₂O (3 equiv) was added. 555 The mixture was stirred at room temperature until completion of the 556 reaction. The reaction mixture was acidified with 1 M HCl until pH 1 557 and extracted with EtOAc. The organic phase was dried over 558 anhydrous Na₂SO₄, filtered, and the solvent was removed under 559 reduced pressure to afford the corresponding compound.

5.2.3. Procedure III. Boc Protection. Et_3N (1.5 equiv) was s61 added to a 0.3 M aqueous solution of starting material, cooled in an s62 ice bath. Then Boc_2O (1.5 equiv) was added dropwise and stirred s63 overnight. After completion of the reaction, the solvent was s64 evaporated under reduced pressure. The residue was dissolved in s65 EtOAc, washed with 1 M HCl and brine, dried over anhydrous s66 Na₂SO₄, filtered, and evaporated under reduced pressure. The crude s67 residue was thoroughly washed with hexane several times.

568 **5.2.4.** Procedure IV. SS/SeSe Bond Cleavage. SS Bond 569 Cleavage. To a 0.15 M solution of starting material in wet THF 570 was added tri-*n*-butylphosphine $(P(^nBu)_3)$ (1.05 equiv). The reaction 571 mixture was stirred at room temperature for 2 h. After completion of 572 the reaction, the solvent was removed under reduced pressure to 573 afford the crude product, which was purified by silica gel column 574 chromatography. SeSe Bond Cleavage and Se-Alkylation. To a 0.13 M solution of 575 starting material in ethanol was added NaBH₄ (2.5 equiv) at 0 °C. 576 The reaction mixture was stirred for 20 min, followed by addition of 577 the respective iodinated compound. The reaction mixture was stirred 578 at room temperature for 16 h. Then, the reaction was quenched with 579 1 M HCl and extracted with EtOAc. The organic phase was dried 580 over anhydrous Na₂SO₄, filtered, and the solvent was removed under 581 reduced pressure. The crude residue was purified by silica gel column 582 chromatography. 583

5.2.5. Procedure V. Reduction of Methyl Ester. To a 0.2 M 584 solution of starting material in anhydrous THF, LiAlH₄ (2 equiv) was 585 added at 0 °C. The reaction mixture was stirred at room temperature 586 for 24 h. Then, the reaction was quenched with 1 M HCl, followed by 587 extraction with DCM. The combined organic phases were dried over 588 anhydrous Na_2SO_4 , filtered, and the solvent was removed under 589 reduced pressure. The solid residue was purified by silica gel column 590 chromatography. 591

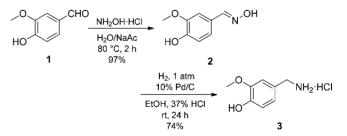
5.2.6. Procedure VI. lodination. To a 0.25 M solution of starting 592 material in toluene iodine (1.2 equiv), imidazole (3 equiv) and PPh₃ 593 (1.2 equiv) were added. The mixture was stirred at 90 °C for 2 h. The 594 solvent was evaporated under reduced pressure. The residue was 595 dissolved in EtOAc, washed with saturated aqueous solution of 596 KMnO₄, water, and brine, dried over anhydrous Na₂SO₄, filtered, and 597 evaporated under reduced pressure. The solid residue was purified by 598 silica gel column chromatography.

5.2.7. Procedure VII. S-Alkylation. To a 0.2 M solution of 600 starting material in DMF, TEA (1.5 equiv) and the corresponding 601 iodinated compound (1.12 equiv) were added. The reaction mixture 602 was stirred at 90 °C overnight. To the mixture were added EtOAc and 603 brine, and the aqueous phase was extracted with EtOAc. The 604 combined organic phases were washed with 1 M HCl, saturated 605 solution of NaHCO₃, and brine. The organic phase was removed under 607 reduced pressure. The crude residue was purified by silica gel column 608 chromatography.

5.2.8. Procedure VIII. TBDMS Deprotection. A 0.25 M solution 610 of the starting material in a mixture of $AcOH/THF/H_2O$ was stirred 611 at room temperature until deprotection was complete. The solvent 612 was evaporated under reduced pressure to obtain the reaction crude, 613 which was purified by silica gel column chromatography. 614

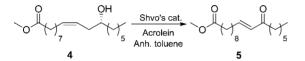
5.2.9. Procedure IX. Boc Deprotection. To a 0.3 M solution of 615 starting material in DCM, TFA (10 equiv) was added. The reaction 616 mixture was stirred for 1 h, followed by removal of the solvent under 617 nitrogen stream and drying in vacuo to afford the trifluoroacetate salt 618 of the compound.

5.2.10. Procedure X. Base Schiff Formation. To a 0.03 M 620 solution of starting material in MeOH, vanillin 1 (1 equiv) was added. 621 The mixture was refluxed for 2 h in the presence of small amount of 622 glacial AcOH. After cooling, the reaction mixture was filtered to 623 recover a solid, which was recrystallized from hot MeOH to afford the 624 corresponding compound. 625 g



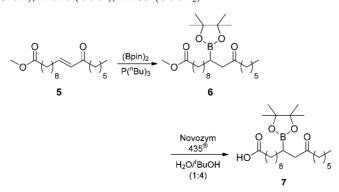
(E)-4-Hydroxy-3-methoxybenzaldehyde Oxime (2). Hydrox- 626 ylamine hydrochloride (2.37 g, 34.0 mmol) in H_2O (10 mL) and 627 sodium acetate trihydrate (4.48 g, 32.9 mmol) in H_2O (10 mL) were 628 successively added to a solution of vanillin 1 (5.00 g, 32.9 mmol) in 629 H_2O (30 mL). The reaction mixture was stirred at 80 °C for 2 h. The 630 reaction mixture was extracted with EtOAc, and the organic layer was 631 dried over anhydrous Na_2SO_4 and filtered. The solvent was 632 evaporated under reduced pressure to yield the oxime 2^1 (5.26 g, 633 634 97%) as an off-white solid. Mp = 118–119 °C. IR (ATR) ν = 3444, 635 3213, 3008, 2941, 1596, 1513, 1428, 1027, 969 cm⁻¹. ¹H NMR (400 636 MHz, (CD₃)₂SO) δ = 3.77 (s, 3H, CH₃O), 6.77 (d, 1H, *J* = 8.1 Hz, 637 H_{Ar}), 6.97 (dd, 1H, *J* = 8.1, 2.0 Hz, H_{Ar}), 7.16 (d, 1H, *J* = 2.0 Hz, 638 H_{Ar}), 7.99 (s, 1H, CH=N), 9.33 (s, 1H, OH), 10.84 (s, 1H, N-OH). 639 ¹³C NMR (101 MHz, (CD₃)₂SO) δ = 55.50 (CH₃O), 109.21 (C_{Ar}), 640 115.49 (C_{Ar}), 120.52 (C_{Ar}), 124.47 (CCHN), 147.85 (COH), 148.01 641 (CCH₃O), 148.10 (CH=N).

4-Hydroxy-3-methoxybenzylamine Hydrochloride (3). 37% 642 643 HCl (20 mL, 0.26 mol) and Pd/C (10 wt % loading) (20% w/w, 1.05 644 g) were added to a solution of 2 (5.2 g, 0.03 mol) in EtOH (150 mL). 645 The reaction mixture was hydrogenated at 1 atm at room temperature 646 for 24 h. The reaction mixture was filtered over Celite, and the solvent 647 volume was reduced under pressure. The residue was crystallized from 648 EtOAc and filtered to yield the amine hydrochloride salt 3^2 (4.2 g, 74%) as a white solid. Mp = 219–222 °C. IR (ATR) ν = 3112, 3024, 649 650 2805, 1763, 1377, 1033, 828, 670 cm⁻¹. ¹H NMR (400 MHz, $(CD_3)_2SO$ $\delta = 3.77$ (s, 3H, CH₃O), 3.83–3.90 (m, 2H, CH₂NH₂), 651 652 6.79 (d, 1H, J = 8.1 Hz, H_{Ar}), 6.85 (dd, 1H, J = 8.1, 2.0 Hz, H_{Ar}), 7.18 653 (d, 1H, J = 2.0 Hz, H_{Ar}), 8.40 (br, s, 3H, NH₂, HCl), 9.19 (s, 1H, 654 OH). ¹³C NMR (101 MHz, $(CD_3)_2SO$) δ = 42.19 (CH_2NH_2) , 55.70 655 (CH₃O), 113.45 (C_{Ar}), 115.27 (C_{Ar}), 121.74 (C_{Ar}), 124.64 (CCHN), 656 146.81 (COH), 147.51 (CCH₃O).



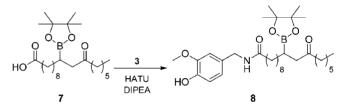
Methyl 12-Oxooctadec-(10E)-enoate (5). Shvo's catalyst (9 657 658 mg, 8 μ mol) and acrolein freshly distilled (390 μ L, 4.80 mmol) were 659 added to a solution of methyl ricinoleate 4 (500 mg, 1.60 mmol) in 660 anhydrous toluene (15 mL). The reaction mixture was purged with 661 N₂ and stirred under reflux for 45 min. The solvent was evaporated 662 under reduced pressure, and after the purification by silica gel column 663 chromatography (petroleum ether/ Et_2O 95:5) the enone 5^3 (348 mg, 664 70%) was obtained as a yellowish oil. $R_f = 0.50$ (petroleum ether/ 665 Et₂O 9:1). IR (ATR) ν = 2927, 2855, 1736, 1709, 1436, 1195, 1169, 666 1104, 979, 880, 752 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ = 0.86 (t, 667 3H, J = 6.9 Hz, CH_3), 1.23–1.33 (m, 14H, CH_2), 1.38–1.48 (m, 2H, 668 CH₂), 1.52-1.65 (m, 4H, CH₂), 2.18 (q, 2H, J = 6.4 Hz, CH₂), 2.29 669 (t, 2H, J = 6.9 Hz, CH_2), 2.51 (t, 2H, J = 6.9 Hz, $COCH_2$), 3.65 (s, 670 3H, CH₃O), 6.07 (dt, 1H, J = 15.9, 1.5 Hz, CH=CH), 6.80 (dt, 1H, J 671 = 15.9, 6.9 Hz, CH=CH). ¹³C NMR (101 MHz, CDCl₃) $\delta = 14.01$ 672 (CH₃), 22.48 (CH₂), 24.27 (CH₂), 24.86 (CH₂), 28.04 (CH₂), 28.96 673 (CH₂), 29.07 (4 × CH₂), 31.59 (CH₂), 32.38 (CH₂), 34.02 (CH₂), 674 40.08 (COCH₂), 51.41 (CH₃O), 130.28 (CH=CH), 147.20 (CH= 675 CH), 174.24 (COO-), 200.99 (COCH₂).

g

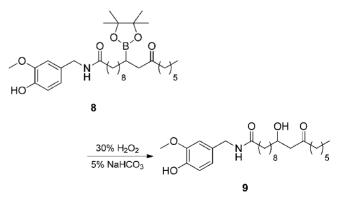


atmosphere. This solution was transferred to the tri-*n*-butylphosphine 683 solution. The reaction mixture was stirred at room temperature for 48 684 h. The crude was taken up in H₂O and extracted with petroleum 685 ether. The organic solution was dried over anhydrous Na₂SO₄, filtered 686 and the solvent was evaporated under reduced pressure to yield the β - 687 boron ketone **6** (190 mg, 46%) as a yellow oil after the purification by 688 silica gel column chromatography (petroleum ether/EtOAc 95:5). R_f 689 = 0.49 (petroleum ether/Et₂O 9:1). ¹H NMR (400 MHz, CDCl₃) δ = 690 0.84 (t, 3H, *J* = 6.9 Hz, CH₃), 1.18–1.28 (m, 30H, (CH₃)₄, CH₂), 691 1.34–1.39 (m, 1H, CHB), 1.49–1.60 (m, 4H, CH₂), 2.27 (t, 2H, *J* = 692 6.9 Hz, CH₂), 2.33 (td, 2H, *J* = 7.4, 3.7 Hz, COCH₂), 2.50 (d, 2H, *J* = 693 6.8 Hz, CHBCH₂CO), 3.64 (s, 3H, CH₃O).

12-Oxo-10-(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)- 695 **octadecanoic Acid (7).** Novozym 435 (83 mg, 50% w/w) was 696 added to a solution of the methyl ester **6** (190 mg, 0.43 mmol) in a 697 mixture of H₂O (308 μL) and *tert*-BuOH (922 μL). The reaction 698 mixture was stirred at 45 °C for 24 h. The mixture was filtered and the 699 solvent was evaporated under reduced pressure to yield the acid 7 700 (180 mg, quantitative) as a yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 701 = 0.87 (t, 3H, *J* = 6.9 Hz, CH₃), 1.20–1.34 (m, 30H, (CH₃)₄, CH₂), 702 1.38–1.44 (m, 1H, CHB), 1.51–1.58 (m, 2H, CH₂), 1.59–1.66 (m, 703 2H, CH₂) 2.30–2.40 (m, 4H, CH₂, COCH₂), 2.53 (d, 2H, *J* = 6.8 Hz, 704 CHBCH₂CO). 705 g

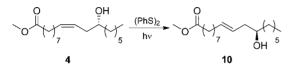


N-(4'-Hydroxy-3'-methoxybenzyl)-12-oxo-10-(4,4,5,5-tetra- 706 methyl-1,3,2-dioxaborolan-2-yl)octadecanamide (8). General 707 procedure I was applied to a solution of the acid 7 (175 mg, 0.41 708 mmol) dissolved in anhydrous DMF (6 mL), amine hydrochloride 709 salt 3 (69 mg, 0.45 mmol), DIPEA (200 μ L, 1.24 mmol), and HATU 710 (235 mg, 0.62 mmol). The amide 8 was obtained (125 mg, 54%) as a 711 brown oil after the purification by silica gel flash column 712 chromatography (petroleum ether/EtOAc 6:4). $R_f = 0.55$ (petroleum 713 ether/EtOAc 3:7). ¹H NMR (400 MHz, CDCl₃) δ = 0.87 (t, 3H, J = 714 6.7 Hz, CH₃), 1.21–1.31 (m, 30H, (CH₃)₄, CH₂), 1.35–1.41 (m, 1H, 715 CHB), 1.52-1.57 (m, 2H, CH₂), 1.61-1.67 (m, 2H, CH₂), 2.18 (t, 716 2H, J = 6.9 Hz, CH_2), 2.32–2.39 (m, 2H, $COCH_2$), 2.52 (d, 2H, J = 7176.7 Hz, CHBCH₂CO), 3.88 (s, 3H, CH₃O), 4.35 (d, 2H, J = 5.6 Hz, 718 CH₂NH), 5.64-5.71 (m, 1H, CH₂NH), 6.82 (ddd, 3H, J = 12.5, 9.9, 719 5.5 Hz, H_{Ar}). 720 g

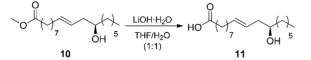


N-(4'-Hydroxy-3'-methoxybenzyl)-10-hydroxy-12-oxoocta- 721 decanamide (9). A volume of 5% w/v NaHCO₃ (2.5 mL, 1.49 722 mmol) was added to a solution of compound 8 (125 mg, 0.22 mmol) 723 and 2.5 mL of 30% H_2O_2 (0.02 mmol). The reaction mixture was 724 stirred at room temperature for 24 h. Saturated aqueous $Na_2S_2O_4$ 725 (0.25 mL) was added to decompose any remaining peroxide keeping 726 the temperature below 40 °C. The reaction mixture was diluted with 727 H_2O and extracted with EtOAc. The organic solution was dried over 728

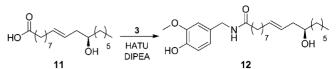
729 anhydrous Na2SO4 and filtered. The solvent was evaporated under 730 reduced pressure to yield the β -hydroxy ketone 9 (75 mg, 76%) as a 731 rosaceous solid after the recrystallization from Et₂O. Mp = 73-75 °C. 732 IR (ATR) ν = 3318, 2912, 2849, 1705, 1638, 1513, 1267, 1240, 1122, 733 718 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ = 0.88 (t, 3H, J = 6.9, Hz, 734 CH₃), 1.20-1.41 (m, 18H, CH₂), 1.40-1.50 (m, 2H, CH₂), 1.52-735 1.60 (m, 2H, CH₂), 1.60–1.68 (m, 2H, CH₂), 2.18 (t, 2H, J = 6.9 Hz, 736 CH₂), 2.41 (t, 2H, J = 6.9 Hz, COCH₂), 2.46-2.52 (m, 1H, 737 CHCH_{11a}CO), 2.59 (dd, 1H, J = 17.3, 1.8 Hz, CHCH_{11b}CO), 3.08 738 (br s, 1H, CHOH), 3.87 (s, 3H, CH₃O), 3.94-4.05 (m, 1H, CHOH), 739 4.35 (d, 2H, J = 5.7 Hz, CH_2NH), 5.69 (br s, 2H, OH, CH_2NH), 740 6.67–6.88 (m, 3H, H_{Ar}). ¹³C NMR (101 MHz, CDCl₃) δ = 14.16 741 (CH₃), 22.61 (CH₂), 23.73 (CH₂), 25.53 (CH₂), 25.87 (CH₂), 28.97 742 (CH₂), 29.34 (CH₂), 29.35 (CH₂), 29.48 (CH₂), 29.55 (CH₂), 31.70 743 (CH₂), 36.52 (CH₂), 36.96 (CH₂), 43.66 (CH₂NH), 43.84 744 (COCH₂), 49.06 (CHCH₂CO), 56.08 (CH₃O), 67.77 (CHOH), 745 110.85 (C_{Ar}), 114.53 (C_{Ar}), 120.93 (C_{Ar}), 130.56 (C_{Ar}), 145.25 (C_{Ar}), 746 146.84 (C_{Ar}), 172.99 (NHCO), 212.84 (COCH₂). HR-MS (ESI⁺), 747 m/z: [M + Na]⁺ calcd for C₂₆H₄₃NO₅Na 472.3033; found 472.3042.



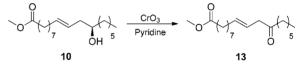
Methyl (12R)-Hydroxyoctadec-(9E)-enoate (10). Diphenyl 748 749 disulfide (56 mg, 0.26 mmol) was added to a solution of methyl 750 ricinoleate 4 (4 g, 12.8 mmol) in isooctane (120 mL). The reaction 751 mixture was placed in a photochemical reactor and irradiated for 3 h 752 with a Philips HP(L) 400 W medium-pressure mercury lamp. After 753 irradiation the solvent was removed under reduced pressure and the 754 crude reaction mixture was dissolved in hot petroleum ether (185 755 mL). The filtrate was cooled at -30 °C, and after 48 h a white solid 756 appeared. This solid was quickly filtered and recovered at -30 °C to 757 yield the compound 10^4 (1.49 g, 37%) as a yellowish oil at room 758 temperature. IR (ATR) ν = 3431, 2924, 2854, 1740, 1435, 1197, 759 1171, 969, 860, 724 cm⁻¹. $[\alpha]_D^{20}$ -0.2° (c 2.44, CHCl₃). ¹H NMR 760 (400 MHz, CDCl₃) $\delta = 0.87$ (t, 3H, I = 6.9 Hz, CH₃), 1.23–1.39 (m, 761 16H, CH₂), 1.39–1.48 (m, 3H, CH₂), 1.56–1.71 (m, 2H, CH₂), 762 1.97–2.09 (m, 3H, CH₂, H_{11a}), 2.18–2.26 (m, 1H, H_{11b}), 2.29 (t, 2H, 763 J = 6.9 Hz, CH_2), 3.53–3.61 (m, 1H, CHOH), 3.65 (s, 3H, CH_3O), 764 5.47-5.56 (m, 1H, CHCH), 5.47-5.56 (m, 1H, CHCH). ¹³C NMR 765 (101 MHz, CDCl₃) δ = 14.22 (CH₃), 22.75 (CH₂), 25.05 (CH₂), 766 25.79 (CH₂), 29.06 (CH₂), 29.20 (CH₂), 29.22 (CH₂), 29.49 (2 \times 767 CH₂), 31.97 (CH₂), 32.75 (CH₂), 34.22 (CH₂), 36.88 (CH₂), 40.85 768 (CHCH₂CHO), 51.57 (CH₃O), 71.06 (CHOH), 126.07 (CHCH), 769 134.69 (CHCH), 174.44 (COO-).



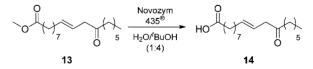
(12R)-Hydroxyoctadec-(9E)-enoic Acid (11). General proce-770 771 dure II was applied to a solution of compound 10 (200 mg, 0.64 772 mmol) dissolved in THF/H2O (3 mL, 1:1) and LiOH·H2O (46 mg, 1.92 mmol) to yield the fatty acid 11⁵ (150 mg, 78%) as a yellowish 773 solid after a recrystallization in hot petroleum ether. Mp = 49-51 °C. 774 $[\alpha]_{\rm D}^{20}$ +6.6° (*c* 1, EtOH). IR (ATR) ν = 3321, 3221, 3040, 2955, 2916, 775 776 2848, 1690, 1466, 1072, 959, 720, 682 cm⁻¹. ¹H NMR (400 MHz, 777 CDCl₃) $\delta = 0.88$ (t, 3H, J = 6.9 Hz, CH₃), 1.22–1.40 (m, 16H, CH₂), 778 1.40-1.50 (m, 4H, CH₂), 1.58-1.68 (m, 2H, CH₂), 1.97-2.11 (m, 779 3H, CH₂, H_{11a}), 2.18–2.28 (m, 1H, H_{11b}), 2.33 (t, 2H, J = 6.9 Hz, 780 CH₂), 3.54-3.63 (m, 1H, CHOH), 5.33-5.46 (m, 1H, CHCH), 781 5.45–5.58 (m, 1H, CHCH). ¹³C NMR (101 MHz, CDCl₃) δ = 14.24 782 (CH₃), 22.77 (CH₂), 24.79 (CH₂), 25.79 (CH₂), 29.02 (CH₂), 29.11 783 (CH₂), 29.15 (CH₂), 29.47 (CH₂), 29.50 (CH₂), 31.98 (CH₂), 32.73 784 (CH₂), 34.06 (CH₂), 36.86 (CH₂), 40.81 (CHCH₂CHO), 71.17 785 (CHOH), 126.05 (CHCH), 134.74 (CHCH), 179.27 (COOH). HR- MS (ESI⁺), m/z: $[M + Na]^+$ calcd for $C_{18}H_{34}O_3Na$ 321.240; found 786 g 321.2411. 787 g



N-(4'-Hydroxy-3'-methoxybenzyl)-(12R)-hydroxyoctadec- 788 (9E)-enamide (12). General procedure I was applied to a solution of 789 the acid 11 (70 mg, 0.23 mmol) dissolved in anhydrous DMF (3.3 790 mL), amine hydrochloride salt 3 (53 mg, 0.28 mmol), DIPEA (122 791 μ L, 0.70 mmol), and HATU (133 mg, 0.35 mmol). The compound 792 12 was afforded (35 mg, 34%) as an off-white solid after the 793 purification by silica gel flash column chromatography (petroleum 794 ether/EtOAc 6:4). $[\alpha]_{D}^{20} <+1^{\circ}$ (c 0.5, DCM). $R_{f} = 0.37$ (petroleum 795 ether/EtOAc 6:4). Mp = 73-75 °C. IR (ATR) ν = 3295, 2920, 2849, 796 1631, 1515, 1463, 1270, 1030, 959 cm⁻¹. ¹H NMR (400 MHz, 797 $CDCl_3$) $\delta = 0.88$ (t, 3H, J = 6.9 Hz, CH_3), 1.23–1.36 (m, 15H, CH_2 , 798 H_{13a}), 1.37-1.46 (m, 3H, CH₂, H_{13b}), 1.59-1.71 (m, 2H, CH₂), 799 1.96-2.09 (m, 3H, CH₂, H_{11a}), 2.14-2.27 (m, 3H, CH₂, H_{11b}), 3.53-800 3.61 (m, 1H, CHOH), 3.86 (s, 3H, CH₃O), 4.34 (d, J = 5.7 Hz, 2H, 801 CH₂NH), 5.35-5.44 (m, 1H, CHCH), 5.47-5.56 (m, 1H, CHCH), 802 5.72 (br s, 2H, CH₂NH, OH), 6.79 (ddd, 3H, J = 16.1, 9.9, 5.0 Hz, 803 H_{Ar}). ¹³C NMR (101 MHz, CDCl₃) δ = 14.23 (CH₃), 22.75 (CH₂), 804 25.79 (CH₂), 25.86 (CH₂), 29.06 (CH₂), 29.26 (CH₂), 29.35 (CH₂), 805 29.46 (CH₂), 29.49 (CH₂), 31.97 (CH₂), 32.73 (CH₂), 36.91 (CH₂), 806 36.96 (CH₂), 40.82 (CHCH₂CHO), 43.65 (CH₂NH), 56.07 807 (CH₃O), 71.07 (CHOH), 110.86 (C_{Ar}), 114.53 (C_{Ar}), 120.91 808 (C_{Ar}), 126.12 (CHCH), 130.54 (C_{Ar}), 134.68 (CHCH), 145.26 809 (C_{Ar}) , 146.84 (C_{Ar}) , 173.01 (NHCO). HR-MS (ESI^+) , m/z: [M + 810 H]⁺ calcd for C₂₆H₄₄NO₄ 434.3265; found 434.3293. 811 g

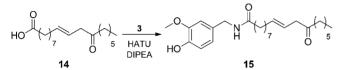


Methyl 12-Oxooctadec-(9E)-enoate (13). CrO₃ (960 mg, 9.6 812 mmol) and pyridine (1.5 mL, 19.2 mmol) were added to a solution of 813 compound 10 (500 mg, 1.6 mmol) in DCM (6 mL). The mixture was 814 vigorously stirred at room temperature for 2 h. The reaction mixture 815 was filtered over Celite and washed with 1 M HCl. The organic phase 816 was dried over anhydrous Na2SO4, filtered and the solvent was 817 evaporated under reduced pressure to yield the ketone 13⁶ (246 g, 818 49%) as a yellowish oil after the purification by silica gel column 819 chromatography (petroleum ether/Et₂O 98:2). $R_f = 0.48$ (petroleum 820 ether/Et₂O 9:1). IR (ATR) ν = 2925, 2854, 1738, 1715, 1435, 1362, 821 1195, 1170, 968, 725 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ = 0.87 (t, 822 $3H, J = 6.5 Hz, CH_3), 1.23-1.38 (m, 14H, CH_2), 1.51-1.64 (m, 4H, 823)$ CH_2), 1.96–2.08 (m, 2H, CH_2), 2.29 (t, J = 6.9 Hz, 2H, CH_2), 2.41 824 $(t, 2H, J = 6.9 \text{ Hz}, \text{COCH}_2), 3.07 (d, 2H, J = 5.2 \text{ Hz}, \text{CH}_2\text{CO}), 3.66 \text{ 825}$ (s, 3H, CH₃O), 5.45–5.56 (m, 2H, CH=CH). ¹³C NMR (101 MHz, 826 $CDCl_3$) $\delta = 14.16$ (CH₃), 22.63 (CH₂), 23.84 (CH₂), 25.06 (CH₂), 827 29.03 (CH₂), 29.06 (CH₂), 29.21 (2 × CH₂), 29.27 (CH₂), 31.73 828(CH₂), 32.67 (CH₂), 34.22(CH₂), 42.31 (COCH₂), 46.95 (CH₂CO), 829 51.57 (CH₃O), 122.13 (CHCH), 135.16 (CHCH), 174.42 (COO-), 830 209.95 (COCH₂). 831 g

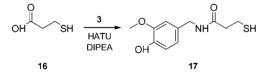


12-Oxooctadec-(9E)-enoic Acid (14). Novozym 435 (20 mg, 832 50% w/w) was added to a solution of the methyl ester **13** (20 mg, 833 0.06 mmol) in a mixture of H₂O (31 μ L) and *tert*-BuOH (138 μ L). 834 The reaction mixture was stirred at 45 °C for 24 h. The mixture was 835 filtered and the solvent was evaporated under reduced pressure to 836

837 yield the acid 14 (17 mg, 89%) as a white solid. Mp = 71–73 °C. IR 838 (ATR) ν = 3121, 2954, 2918, 2848, 1701, 1263, 1082, 962, 720, 689 839 cm^{-1.} ¹H NMR (400 MHz, CDCl₃) δ = 0.87 (t, 3H, *J* = 6.9 Hz, CH₃), 840 1.26–1.36 (m, 14H, CH₂), 1.50–1.58 (m, 2H, CH₂), 1.58–1.66 (m, 841 2H, CH₂), 1.98–2.08 (m, 2H, CH₂), 2.34 (t, 2H, *J* = 6.9 Hz, CH₂), 842 2.41 (t, 2H, *J* = 6.9 Hz, COCH₂), 3.08 (d, 2H, *J* = 5.2 Hz, CH₂CO), 843 5.44–5.57 (m, 2H, CHCH). ¹³C NMR (101 MHz, CDCl₃) δ = 14.17 844 (CH₃), 22.63 (CH₂), 23.85 (CH₂), 24.79 (CH₂), 29.03 (2 × CH₂), 845 29.12 (CH₂), 29.18 (CH₂), 29.26 (CH₂), 31.73 (CH₂), 32.66 (CH₂), 846 34.09 (CH₂), 42.32 (COCH₂), 46.95 (CH₂CO), 122.13 (CHCH), 847 135.17 (CHCH), 179.59 (COOH), 210.13 (COCH₂). HR-MS 848 (ESI⁺), *m*/*z*: [M + Na]⁺ calcd for C₁₈H₃₂O₃Na 319.2244; found 849 319.2267.

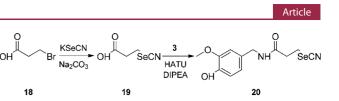


N-(4'-Hydroxy-3'-methoxybenzyl)-12-oxooctadec-(9E)-en-850 851 amide (15). General procedure I was applied to a solution of the acid 852 14 (210 mg, 0.71 mmol) dissolved in anhydrous DMF (10 mL), 853 amine hydrochloride salt 3 (148 mg, 0.78 mmol), DIPEA (400 μ L, 854 2.1 mmol), and HATU (404 mg, 1.06 mmol). The compound 15 was 855 obtained (52 mg, 17%) as an off-white solid after the purification by 856 silica gel flash column chromatography (petroleum ether/EtOAc 7:3). 857 Mp = 71–73 °C. R_f = 0.36 (petroleum ether/EtOAc 7:3). IR (ATR) 858 $\nu = 3393, 3312, 2917, 2850, 1703, 1636, 1554, 1509, 1242, 1125, 967,$ 859 705 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ = 0.87 (t, 3H, J = 6.9 Hz, 860 CH₃), 1.22-1.38 (m, 14H, CH₂), 1.50-1.58 (m, 2H, CH₂), 1.59-861 1.69 (m, 2H, CH₂), 1.97-2.04 (m, 2H, CH₂), 2.19 (t, 2H, J = 7.4 Hz, 862 CH₂), 2.40 (t, 2H, J = 7.4 Hz, COCH₂), 3.08 (d, 2H, J = 5.2 Hz, 863 CH₂CO), 3.87 (s, 3H, CH₃O), 4.35 (d, 2H, J = 5.7 Hz, CH₂NH), 864 5.47-5.52 (m, 2H, CHCH), 5.67 (s, 1H, CH₂NH), 5.73 (br s, 1H, 865 OH), 6.73–6.87 (6.79 (ddd, 3H, J = 12.5, 9.9, 5.0 Hz, H_{Ar}). ¹³C 866 NMR (101 MHz, CDCl₃) δ = 14.17 (CH₃), 22.63 (CH₂), 23.86 867 (CH₂), 25.86 (CH₂), 29.03 (CH₂), 29.05 (CH₂), 29.23 (CH₂), 29.26 868 (CH₂), 29.36 (CH₂), 31.73 (CH₂), 32.64 (CH₂), 36.96 (CH₂), 42.37 869 (COCH₂), 43.66 (CH₂NH), 46.89 (CH₂CO), 56.07 (CH₃O), 110.83 870 (C_{Ar}), 114.50 (C_{Ar}), 120.92 (C_{Ar}), 122.12 (CHCH), 130.56 (C_{Ar}), 871 135.11 (CHCH), 145.25 (C_{Ar}), 146.82 (C_{Ar}), 172.99 (NHCO), 872 210.08 (COCH₂). HR-MS (ESI⁺), m/z: [M + H]⁺ calcd for 873 C₂₆H₄₂NO₄ 432.3108; found 432.3137.



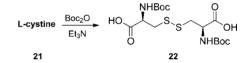
N-(4'-Hydroxy-3'-methoxybenzyl)-3-mercapto-874 875 propanamide (17). General procedure I was applied to a solution of 876 mercaptopropionic acid (1.2 mL, 12.68 mmol) dissolved in 877 anhydrous DMF (30 mL), amine hydrochloride salt 3 (2.65 g, 878 13.95 mmol), DIPEA (6.63 mL, 38.04 mmol), and HATU (7.23 g, 879 19.02 mmol). Compound 17 was obtained after silica gel column 880 chromatography (petroleum ether/EtOAc 5:5) as sticky oil (2.14 g, 74%). $R_f = 0.60$ (petroleum ether/EtOAc 4:6). IR (ATR) $\nu = 3425$, 881 882 2922, 2853, 1515, 836 cm⁻¹. ¹H NMR (400 MHz, (CH₃)₂CO) δ = 883 1.86 (t, 1H, J = 8.2 Hz, SH), 2.54 (t, 2H, J = 6.7 Hz, CH₂), 2.70–2.82 884 (m, 2H, CH_2SH), 3.80 (s, 3H, CH_3O), 4.31 (d, 2H, J = 5.9 Hz, 885 CH₂NH), 6.74 (d, 2H, J = 1.0 Hz, H_{Ar}, OH), 6.92 (s, 1H, H_{Ar}), 7.48 886 (s, 2H, H_{Ar} , CH₂NH). ¹³C NMR (101 MHz, (CH₃)₂CO) δ = 20.10 887 (CH₂SH), 39.71 (CH₂), 42.47 (CH₂NH), 55.33 (CH₃O), 111.25 888 (C_{Ar}), 114.66 (C_{Ar}), 120.16 (C_{Ar}), 130.83 (C_{Ar}), 145.61 (C_{Ar}), 147.36 (C_{Ar}), 170.16 (NHCO). HR-MS (ESI⁺), m/z: [M + H]⁺ calcd for 889 C₁₁H₁₅NO₃S, 242.0845; found 242.0861. 890

3-Selenocyanatopropanoic Acid (19). To a solution of 3bromopropionic acid **18** (1.5 g, 9.8 mmol) in water (3 mL) was added Na₂CO₃ until pH 7. A volume of 14 mL of 10% KSeCN (1.41

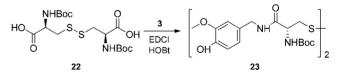


g, 9.8 mmol, 1 equiv) aqueous solution was added. The mixture 894 stirred at room temperature for 2 days. After removing partially the 895 solvent under reduced pressure, the crude was dissolved in Et₂O and 896 washed with 1 M HCl, water and brine. The organic solution was 897 dried over Na₂SO₄, filtered and the solvent was removed under 898 reduced pressure to yield the 3-selenocyanatopropanoic acid **19**⁷ as a 899 yellow oil (1.39 g, 80%) which was used in the next step without 900 further purification. IR (ATR) ν = 3024, 2649, 2152, 1703, 1401 901 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ = 3.07 (t, 2H, *J* = 6.4 Hz, 902 CH₂SeCN), 3.24 (dd, 2H, *J* = 6.4 Hz, CH₂CH₂SeCN), 9.52 (br s, 1H, 903 COOH). ¹³C NMR (101 MHz, CDCl₃) δ = 22.89 (CH₂SeCN), 34.90 904 (CH₂CH₂SeCN), 101.68 (SeCN), 176.86 (COOH).

N-(4'-Hydroxy-3'-methoxybenzyl)-3-selenocyanato- 906 propanamide (20). General procedure I was applied to a solution of 907 compound 19 (1.3 g, 7.30 mmol), amine hydrochloride salt 3 (1.52 g, 908 8.03 mmol), DIPEA (3.82 mL, 21.9 mmol), and HATU (4.16 g, 909 10.95 mmol) in anhydrous DMF (20 mL). Compound 20 was 910 afforded after silica gel column chromatography (petroleum ether/ 911 EtOAc 5:5) as a white sticky solid (2.14 g, 60%). $R_f = 0.65$ (petroleum 912 ether/EtOAc 4:6). IR (ATR) $\nu = 3315$, 2924, 2853, 2148, 1638, 1235 913 cm⁻¹. ¹H NMR (400 MHz, (CH₃)₂CO) δ = 2.94 (t, 2H, J = 6.4 Hz, 914 $COCH_2$), 3.34 (t, 2H, J = 6.4 Hz, CH_2SeCN), 3.81 (s, 3H, CH_3), 915 4.30 (d, 2H, J = 5.8 Hz, CH_2NH), 6.75 (s, 2H, H_{Ar}), 6.91 (s, 1H, 916 H_{Ar}), 7.48 (s, 1H, OH), 7.72 (s, 1H, CH₂NH). ¹³C NMR (101 MHz, 917 $(CH_3)_2CO) \delta = 24.79 (CH_2SeCN), 34.84 (CH_2CH_2SeCN), 42.73 918$ (CH₂NH), 55.33 (CH₃O), 104.64 (SeCN), 111.35 (C_{Ar}), 114.72 919 (C_{Ar}) , 120.32 (C_{Ar}) , 130.19 (C_{Ar}) , 145.79 (C_{Ar}) , 147.38 (C_{Ar}) , 170.92 920 (NHCO). HR-MS (ESI⁺), m/z: [M + H]⁺ calcd for C₁₂H₁₅N₂O₃Se, 921 315.0248; found 315.0242. 922 g

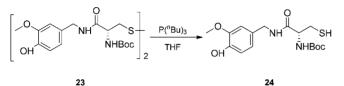


N,*N*-Di-Boc-L-cystine (22). General procedure III was applied to 923 L-cystine 21 (10 g, 41.67 mmol), Boc₂O (27.25 g, 124.85 mmol), and 924 Et₃N (17.5 mL, 125.38 mmol) in water (150 mL) to yield compound 925 22⁸ as a white solid, which was thoroughly washed with petroleum 926 ether several times (17.56 g, 96%). Mp: 145–146 °C. IR (ATR) ν = 927 3366, 2985, 2936, 1682, 1511, 1163, 1052, 868 cm^{-1.} ¹H NMR (400 928 MHz, (CD₃)₂SO) δ = 1.37 (s, 18H, Boc), 2.87 (dd, 2H, *J* = 13.5, 10.1 929 Hz, CHCH₂), 3.09 (dd, 2H, *J* = 13.5, 4.4 Hz, CHCH₂), 4.16 (td, 2H, 930 *J* = 10.1, 4.4 Hz, CHCH₂), 7.18 (d, 2H, *J* = 8.4 Hz, NH), 12.79 (s, 931 2H, COOH). ¹³C NMR (101 MHz, (CD₃)₂SO) δ = 28.60 932 (C(CH₃)₃), 52.96 (CHCH₂), 78.70 (C(CH₃)₃), 155.79 (NHCO₂), 933 172.82 (COOH). 934 g

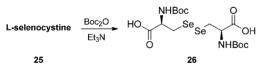


Di-[(2*R*)-*N*-Boc-amino-1-((4'-hydroxy-3'-methoxybenzyl)- 935 amino)-1-oxoprop-3-yl]disulfane (23). To a solution of com- 936 pound 22 (5 g, 11.35 mmol) in anhydrous DMF (50 mL) were added 937 HOBt (4.6 g, 34.05 mmol), Et₃N (4.74 mL, 34.05 mmol), and the 938 amine hydrochloride salt 3 (5.16 g, 27.24 mmol). The mixture was 939 stirred at 0 °C during 30 min. EDCI (6.52 g, 34 mmol) was added 940 and the mixture stirred at room temperature during 20 h. To the 941 mixture were added EtOAc and brine, and the aqueous phase was 942 extracted with EtOAc. The combined organic solutions were washed 943 with 1 M HCl, saturated NaHCO₃, and brine. The organic solution 944 was dried over anhydrous Na_2SO_4 , filtered and the solvent was 945

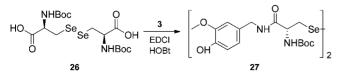
946 evaporated under reduced pressure. Compound **23** was afforded after 947 silica gel column chromatography (PE/EtOAc 1:9) as a white solid 948 (7.58 g, 94%). $R_f = 0.24$ (petroleum ether/EtOAc 1:9). Mp: 167–170 949 °C. $[\alpha]_D^{20}$ -67.42 (*c* 0.75, MeOH). IR (ATR) ν = 3330, 2975, 2935, 950 1658, 1511, 1272, 1033 cm⁻¹. ¹H NMR (400 MHz, (CD₃)₂SO) δ = 951 1.36 (s, 18H, C(CH₃)₃), 2.86 (dd, 2H, *J* = 13.0, 9.9 Hz, CHCH₂), 952 3.07 (dd, 2H, *J* = 13.0, 4.8 Hz, CHCH₂), 3.72 (s, 6H, CH₃O), 4.02– 953 4.32 (m, 6H, CHCH₂, CH₂NH), 6.55–6.72 (m, 4H, H_{Ar}, NHBoc), 954 6.79 (s, 2H, H_{Ar}), 7.06 (d, 2H, *J* = 8.4 Hz, H_A), 8.31 (t, 2H, *J* = 5.4 955 Hz, CH₂NH), 8.78 (br s, 2H, OH). ¹³C NMR (101 MHz, (CD₃)₂SO) 956 δ = 28.59 (C(CH₃)₃), 40.59 (CHCH₂), 42.40 (CH₂NH), 54.17 957 (CHCH₂), 55.92 (CH₃O), 78.73 (C(CH₃)₃), 111.82 (C_{Ar}), 115.53 958 (C_{Ar}), 119.88 (C_{Ar}), 130.37 (C_{Ar}), 145.76 (C_{Ar}), 147.85 (C_{Ar}), 155.70 959 (NHCO₂), 170.60 (NHCO). HR-MS (ESI⁺), *m*/z: [M + H]⁺ calcd 960 for C₃₂H₄₇N₄O₁₀S₂, 711.2734; found 711.2793.



N-(4'-Hydroxy-3'-methoxy)benzyl-(2R)-(Boc-amino)-3-961 962 mercaptopropanamide (24). General procedure IV (SS bond 963 cleavage) was applied to compound 23 (7 g, 9.86 mmol) dissolved in 964 THF (60 mL), P("Bu)₃ (2.55 mL, 10.35 mmol) in the presence of 965 water (1.3 mL). Compound 24 was afforded after silica gel column 966 chromatography (petroleum ether/EtOAc 5:5) as a white solid (5.11 g, 73%). $R_f = 0.42$ (petroleum ether/EtOAc 4:6). Mp: 108–110 °C. 967 968 $[\alpha]_{D}^{20}$ –15.65 (c 1.6, MeOH). IR (ATR) ν = 3456, 3327, 2989, 2934, 969 2847, 1678, 1513, 1240 cm⁻¹. ¹H NMR (400 MHz, CDCl₂) δ = 1.41 970 (s, 9H, $C(CH_3)_3$), 1.54 (t, 1H, J = 10.7 Hz, SH), 2.74 (ddd, 1H, J =971 13.8, 10.2, 6.1 Hz, CHCH₂), 3.09 (ddd, 1H, J = 13.6, 7.6, 4.6 Hz, 972 CHCH₂), 3.84 (s, 3H, CH₃O), 4.25-4.44 (m, 3H, CHCH₂) 973 CH₂NH), 5.48 (d, 1H, J = 7.8 Hz, CH₂NH), 5.81 (br s, 1H, OH), 974 6.67–6.89 (m, 4H, H_{Art} NHBoc). ¹³C NMR (101 MHz, CDCl₃) $\delta =$ 975 26.96 (CHCH₂), 28.23 (C(CH₃)₃), 43.47 (CH₂NH), 55.67 976 (CHCH₂), 55.93 (CH₃O), 80.69 (C(CH₃)₃), 110.47 (C_{Ar}), 114.44 977 (C_{Ar}), 120.58 (C_{Ar}), 129.66 (C_{Ar}), 145.12 (C_{Ar}), 146.74 (C_{Ar}), 155.46 978 (NHCO₂), 169.88 (NHCO). HR-MS (ESI⁺), m/z: [M + H]⁺ calcd 979 for C₁₆H₂₄N₂O₅SNa, 379.1298; found 379.1326.

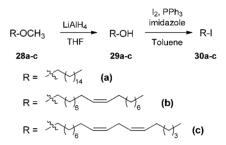


N,N-Di-Boc-L-selenocystine (26). General procedure III was applied to L-selenocystine 25 (1.5 g, 4.49 mmol), Boc₂O (3.24 g, 982 13.48 mmol), and Et₃N (1.88 mL, 13.48 mmol) in water (22 mL) to 983 yield compound 26⁹ as a yellow solid (1.55 g, 65%), which was used 984 in the next step without further purification. Mp: 145–147 °C. $[\alpha]_{20}^{D0}$ 985 –75.63 (*c* 1.5, DCM). IR (ATR) ν = 3364, 2979, 2557, 1698, 1662, 986 1506 cm⁻¹. ¹H NMR (400 MHz, (CD₃)₂SO) δ = 1.37 (*s*, 18H, 987 C(CH₃)₃), 3.10 (dd, 2H, *J* = 11.9, 10.2 Hz, CHCH₂), 7.17 (d, 2H, *J* 988 = 11.9, 4.7 Hz, CHCH₂), 4.06–4.21 (m, 2H, CHCH₂), 7.17 (d, 2H, *J* 989 = 8.3 Hz, NH), 12.79 (*s*, 2H, COOH). ¹³C NMR (101 MHz, 990 (CD₃)₂SO) δ = 28.61 (C(CH₃)₃), 31.38 (CHCH₂), 54.68 (CHCH₂), 991 78.71 (C(CH₃)₃), 155.71 (NHCO₂), 172.91 (COOH).



Di-[(2*R*)-*N*-Boc-amino-1-((4'-hydroxy-3'-methoxybenzyl)amino)-1-oxoprop-3-yl]diseleno (27). To a solution of compound
26 (1.5 g, 2.80 mmol) in anhydrous DMF (14 mL) were added HOBt
(1.14 g, 8.4 mmol), Et₃N (1.18 mL, 8.4 mmol), and the amine

hydrochloride salt 3 (1.27 g, 6.72 mmol). The mixture was stirred at 0 996 °C during 30 min. EDCI (1.61 g, 8.4 mmol) was added and the 997 mixture stirred at room temperature during 20 h. To the mixture were 998 added EtOAc and brine, and the aqueous phase was extracted. The 999 combined organic layers were washed with 1 M HCl, saturated 1000 NaHCO3, and brine. The organic phase was dried over anhydrous 1001 Na₂SO₄, filtered, and the solvent was evaporated under reduced 1002 pressure. Compound 27 was afforded after silica gel column 1003 chromatography (petroleum ether/EtOAc 1:9) as a white solid 1004 (1.98 g, 88%). $R_f = 0.26$ (petroleum ether/EtOAc 5:5). Mp: 93-95 1005 °C. $\left[\alpha\right]_{D}^{20}$ 42.94 (c 0.7, DCM). IR (ATR) ν = 3314, 2975, 2932, 1654, 1006 1513, 1157 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ = 1.26 (s, 18H, 1007 C(CH₃)₃), 3.12-3.30 (m, 4H, CHCH₂), 3.83 (s, 6H, CH₃O), 4.25 1008 (dd, 2H, J = 14.7, 5.4 Hz, CH₂NH), 4.48 (dd, 2H, J = 14.7, 6.5 Hz, 1009 CH_2NH), 4.75–4.94 (m, 2H, $CHCH_2$), 5.58 (d, 2H, J = 9.7 Hz, 1010 NHBoc), 5.63 (s, 2H, OH), 6.77 (ddd, 6H, $J = 12.5, 9.9, 5.0, H_{Ar}$), 1011 8.06 (t, 2H, J = 5.6 Hz, CH₂NH). ¹³C NMR (101 MHz, CDCl₃) $\delta = 1012$ 28.15 (C(CH₃)₃), 37.43 (CHCH₂), 43.28 (CH₂NH), 55.24 1013 (CHCH₂), 55.86 (CH₃O), 78.98 (C(CH₃)₃), 110.44 (C_{Ar}), 114.24 1014 (C_{Ar}) , 120.77 (C_{Ar}) , 130.03 (C_{Ar}) , 145.00 (C_{Ar}) , 146.58 (C_{Ar}) , 155.65 1015 (NHCO₂), 170.53 (NHCO). HR-MS (ESI⁺), m/z: [M + H]⁺ calcd 1016 for C₃₂H₄₆N₄O₁₀Se₂, 807.1623; found 807.1621. 1017 g



1-Hexadecanol (29a). General procedure V was applied to 1018 methyl palmitate **28a** (1 g, 3.69 mmol), LiAlH₄ (280 mg, 7.38 mmol) 1019 in anhydrous THF (20 mL). Compound **29a**¹⁰ was afforded after 1020 silica gel column chromatography (petroleum ether/Et₂O 9:1) as a 1021 white solid (875 mg, 98%). $R_f = 0.88$ (petroleum ether/Et₂O 9:1). 1022 Mp: 50–52 °C. IR (ATR) $\nu = 3320$, 3226, 2915, 2919, 2847, 1462 1023 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) $\delta = 0.87$ (t, 3H, J = 6.9 Hz, CH₃), 1024 1.15–1.41 (m, 24H, CH₂), 1.45–1.64 (m, 4H, CH₂, HOCH₂CH₂), 1025 3.62 (t, 2H, J = 6.9 Hz, HOCH₂CH₂). ¹³C NMR (101 MHz, CDCl₃) 1026 $\delta = 14.08$ (CH₃), 22.67 (CH₂), 25.74 (CH₂), 29.53 (CH₂), 29.43 1027 (CH₂), 29.60 (CH₂), 29.61 (CH₂), 29.65 (2 × CH₂), 29.67 (CH₂), 1028 29.68 (3 × CH₂), 31.91 (CH₂), 32.78 (HOCH₂CH₂), 62.99 1029 (HOCH₂CH₂).

(9Z)-Octadecen-1-ol (29b). General procedure V was applied to 1031 methyl oleate 28b (2.5 g, 8.43 mmol), LiAlH₄ (640 mg, 16.86 mmol) 1032 in anhydrous THF (50 mL). Compound 29b¹¹ was afforded after 1033 silica gel column chromatography (petroleum ether/Et₂O 9:1) as a 1034 brown oil (2.19 g, 97%). $R_f = 0.88$ (petroleum ether/Et₂O 9:1). IR 1035 (ATR) ν = 3320, 2921, 2852, 1463, 1055 cm⁻¹. ¹H NMR (400 MHz, 1036 $CDCl_3$) $\delta = 0.87$ (t, 3H, J = 6.9 Hz, CH_3), 1.16–1.41 (m, 22H, CH_2), 1037 1.47-1.62 (m, 2H, HOCH₂CH₂), 1.73 (s, 1H, OH), 2.00 (q, 4H, J = 10386.4 Hz, CH₂CH, CHCH₂), 3.61 (t, 2H, J = 6.9 Hz, HOCH₂CH₂), 1039 5.25-5.47 (m, 2H, CH=CH). ¹³C NMR (101 MHz, CDCl₃) δ = 1040 14.07 (CH₃), 22.65 (CH₂), 25.73 (CH₂), 27.16 (CH₂CH), 27.18 1041 $(CHCH_2)$, 29.22 (CH_2) , 29.30 $(2 \times CH_2)$, 29.40 (CH_2) , 29.49 1042 (CH₂), 29.50 (CH₂), 29.72 (CH₂), 29.74 (CH₂), 31.88 (CH₂), 32.75 1043 (HOCH₂CH₂), 62.93 (HOCH₂CH₂), 129.76 (CH=CH), 129.90 1044 (CH=CH).1045

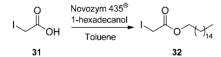
(9*Z*,12*Z*)-Octadecadien-1-ol (29c). General procedure V was 1046 applied to methyl linoleate 28b (1 g, 3.39 mmol), LiAlH₄ (257 mg, 1047 6.79 mmol) in anhydrous THF (30 mL). Compound 29c¹² was 1048 afforded after silica gel column chromatography (petroleum ether/ 1049 Et₂O 9:1) as a colorless oil (885 mg, 98%). $R_f = 0.88$ (petroleum 1050 ether/Et₂O 9:1). IR (ATR) $\nu = 3373$, 2926, 2855, 1719, 1463 cm⁻¹. 1051 ¹H NMR (400 MHz, CDCl₃) $\delta = 0.89$ (t, 3H, J = 6.9 Hz, CH_3), 1052 1.19–1.48 (m, 16H, CH_2), 1.51–1.61 (m, 2H, HOCH₂CH₂), 2.05 1053

1054 (q, 4H, *J* = 6.4 Hz, CH₂CH, CHCH₂), 2.77 (t, 2H, *J* = 6.9 Hz, 1055 CHCH₂CH), 3.59–3.67 (m, 2H, HOCH₂CH₂), 5.14–5.52 (m, 4H, 2 1056 × CH=CH). ¹³C NMR (101 MHz, CDCl₃) δ = 14.04 (CH₃), 22.55 1057 (CH₂), 25.61 (CHCH₂CH), 25.71 (CH₂), 27.18 (CH₂CH), 27.20 1058 (CHCH₂), 29.22 (CH₂), 29.33 (CH₂), 29.38 (CH₂), 29.48 (CH₂), 1059 29.63 (CH₂), 31.51 (CH₂), 32.78 (HOCH₂CH₂), 63.03 1060 (HOCH₂CH₂), 127.89 (CH=CH), 127.97 (CH=CH), 130.08 1061 (CH=CH), 130.08 (CH=CH).

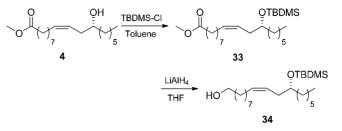
1-lodohexadecane (30a). General procedure VI was applied to 1063 compound **29a** (1 g, 4.12 mmol), iodine (1.25 g, 4.95 mmol), PPh₃ 1064 (1.3 g, 4.95 mmol), and imidazole (0.85 g, 12.36 mmol) in toluene 1065 (15 mL). Compound **30a**¹³ was afforded after silica gel column 1066 chromatography (petroleum ether) as a yellow oil (1.08 g, 75%). R_f = 1067 0.1 (petroleum ether). IR (ATR) ν = 2920, 2851, 1464, 1376, 1171, 1068 719 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ = 0.88 (t, 3H, *J* = 6.9 Hz, 1069 CH₃), 1.26 (s, 24H, CH₂), 1.34–1.41 (m, 2H, ICH₂CH₂CH₂), 1.75– 1070 1.87 (m, 2H, ICH₂CH₂), 3.18 (t, 2H, *J* = 6.9 Hz, ICH₂). ¹³C NMR 1071 (101 MHz, CDCl₃) δ = 7.21 (ICH₂), 14.11 (CH₃), 22.69 (CH₂), 1072 28.55 (CH₂), 29.36 (CH₂), 29.42 (CH₂), 29.55 (CH₂), 29.61 (CH₂), 1073 29.65 (2 × CH₂), 29.68 (2 × CH₂), 29.69 (CH₂), 30.51 (CH₂), 31.92 1074 (CH₂), 33.58 (ICH₂CH₂).

1-lodo-(9Z)-octadecene (30b). General procedure VI was 1075 1076 applied to compound 29b (2 g, 7.45 mmol), iodine (2.27 g, 8.94 1077 mmol), PPh3 (2.34 g, 8.94 mmol), and imidazole (1.52 g, 22.35 1078 mmol) in toluene (30 mL). Compound **30b**¹⁴ was afforded after silica 1079 gel column chromatography (petroleum ether/Et₂O 9:1) as a yellow 1080 oil (2.42 g, 86%). $R_f = 0.1$ (petroleum ether/Et₂O 9:1). IR (ATR) $\nu =$ 1081 2921, 2852, 1462, 1181 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) $\delta = 0.88$ 1082 (t, 3H, J = 6.9 Hz, CH_3), 1.16–1.48 (m, 22H, CH_2), 1.72–1.91 (m, 1083 2H, ICH_2CH_2), 2.01 (q, 4H, J = 6.4 Hz, CH_2CH , $CHCH_2$), 3.18 (t, 1084 2H, J = 6.9 Hz, ICH₂), 5.21–5.48 (m, 2H, CH=CH). ¹³C NMR 1085 (101 MHz, CDCl₃) δ = 7.24 (ICH₂), 14.10 (CH₃), 22.67 (CH₂), 1086 27.15 (CH₂CH), 27.21 (CHCH₂), 28.50 (CH₂), 29.16 (CH₂), 29.29 1087 (CH₂), 29.31 (CH₂), 29.51 (CH₂), 29.68 (CH₂), 29.75 (CH₂), 30.48 1088 (CH₂), 31.89 (CH₂), 33.55 (ICH₂CH₂), 129.73 (CH=CH), 129.98 1089 (CH=CH)

18-lodo-(6Z,9Z)-octadecadiene (30c). General procedure VI 1090 1091 was applied to compound 29c (850 mg, 3.18 mmol), iodine (968 mg, 1092 3.81 mmol), PPh₃ (1 g, 3.81 mmol), and imidazole (650 mg, 9.54 1093 mmol) in toluene (15 mL). Compound **30c**¹⁴ was afforded after silica gel column chromatography (petroleum ether) as a yellow oil (1.13 g, 1094 1095 95%). $R_{\rm f} = 0.1$ (petroleum ether/Et₂O 9:1). IR (ATR) $\nu = 3439$. 1096 2926, 2855, 1707, 1458, 1175 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ = 1097 0.89 (t, 3H, J = 6.9 Hz, CH_3), 1.18–1.50 (m, 16H, CH_2), 1.78–1.86 1098 (m, 2H, ICH₂CH₂), 2.05 (q, 4H, J = 6.4 Hz, CH₂CH, CHCH₂), 2.77 1099 (t, 2H, J = 6.9 Hz, CHCH₂CH), 3.18 (t, 2H, J = 6.9 Hz, ICH₂CH₂), 1100 5.25–5.50 (m, 2 × CH=CH). ¹³C NMR (101 MHz, CDCl₃) δ = 1101 7.20 (ICH₂), 14.07 (CH₃), 22.57 (CH₂), 25.63 (CHCH₂CH), 27.18 1102 (CH₂CH), 27.20 (CHCH₂), 28.50 (CH₂), 29.17 (CH₂), 29.30 (CH₂), 1103 29.34 (CH₂), 29.59 (CH₂), 30.48 (CH₂), 31.52 (CH₂), 33.55 1104 (ICH₂CH₂), 127.89 (CH=CH), 128.02 (CH=CH), 130.02 (CH= 1105 CH), 130.18 (CH=CH).

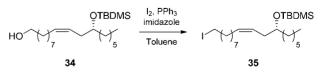


Hexadecyl 2-lodoacetate (32). To a solution of iodoacetic acid 1107 31 (500 mg, 2.69 mmol) in toluene (5 mL) were added 1-1108 hexadecanol (978 mg, 4.03 mmol, 1.5 equiv) and Novozym 435 (150 1109 mg). The reaction mixture was stirred at 50 °C for 2 days. The 1110 mixture was filtered off, EtOAc was added, and the organic phase was 1111 washed with saturated solution of NaHCO₃, water, and brine. The 1112 organic solution was then dried over Na₂SO₄, and the solvent was 1113 removed under reduced pressure. Compound 32¹⁵ was afforded after 1114 silica gel column chromatography (petroleum ether/Et₂O 9:1) as a 1115 yellow oil (562 mg, 51%). R_f = 0.36 (petroleum ether/Et₂O 9:1). IR 1116 (ATR) ν = 2920, 2851, 1733, 1259, 1089 cm⁻¹. ¹H NMR (400 MHz, 1117 CDCl₃) δ = 0.86 (t, 3H, *J* = 6.9 Hz, CH₃), 1.14–1.41 (m, 26H, CH₂),



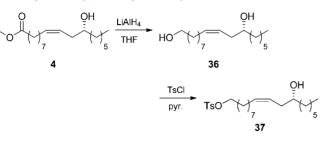
Methyl (12R)-[(tert-Butyldimethylsilyl)oxy]octadec-(9Z)- 1124 enoate (33). To a solution of methyl ricinoleate 4 (2 g, 6.4 1125 mmol) in DCM (40 mL) were added DMAP (31 mg, 0.25 mmol) 1126 and Et₃N (2.23 mL, 16 mmol). TBDMS-Cl was slowly added (1.5 g, 1127 9.92 mmol). The mixture was stirred at room temperature for 2 days. 1128 Then, the organic phase was washed with 1 M HCl, water, and brine, 1129 dried over anhydrous NaSO4 and the solvent was removed under 1130 reduced pressure. Compound 33¹⁶ was afforded after silica gel column 1131 chromatography (petroleum ether/Et₂O 9:1) as a colorless oil (2.37 1132 g, 87%). $R_f = 0.1$ (petroleum ether/Et₂O 9:1). $[\alpha]_D^{20}$ 9.98 (c 2.8, 1133 DCM). IR (ATR) ν = 2927, 2855, 1742, 1461, 1251 cm⁻¹. ¹H NMR 1134 (400 MHz, CDCl₃) δ = 0.04 (s, 6H, Si(CH₃)₂), 0.78–0.95 (m, 12H, 1135 SiC(CH₂)₂, CH₂), 1.16–1.46 (m, 18H, CH₂), 1.51–1.68 (m, 2H, 1136 $COCH_2CH_2$), 2.01 (q, 2H, J = 6.4 Hz, CH_2CH), 2.17 (t, 2H, J = 6.9 1137 Hz, CHCH₂), 2.29 (t, 2H, J = 6.9 Hz, COCH₂CH₂), 3.59–3.73 (m, 1138 4H, CH₃O, CH₂CHO), 5.29-5.51 (m, 2H, CH=CH). ¹³C NMR 1139 $(101 \text{ MHz}, \text{CDCl}_3) \delta = -4.59 (\text{SiCH}_3), -4.38 (\text{SiCH}_3), 14.06 (\text{CH}_3), 1140$ 18.11 (SiC(CH₃)₃), 22.61 (CH₂), 24.92 (COCH₂CH₂), 25.38 (CH₂), 1141 25.89 (SiC(CH₃)₃), 27.40 (CH₂CH), 29.10 (CH₂), 29.12 (CH₂), 1142 29.14 (CH₂), 29.45 (CH₂), 29.58 (CH₂), 31.87 (CH₂), 34.06 1143 (COCH₂CH₂), 35.23 (CHCH₂), 36.84 (CH₂), 51.38 (CH₃O), 72.37 1144 (CH₂CHO), 125.95 (CH=CH), 131.28 (CH=CH), 174.23 1145 (COOH). 1146

12R)-[(tert-Butyldimethylsilyl)oxy]octadec-(9Z)-en-1-ol 1147 (34). General procedure V was applied to compound 33 (2.20 g, 5.15 1148 mmol) with anhydrous LiAlH₄ (390 mg, 10.30 mmol) in dry THF 1149 (50 mL). Compound 34¹⁷ was afforded after silica gel column 1150 chromatography (petroleum ether/Et₂O 9:1) as a brown oil (1.91 g, 1151 93%). $R_f = 0.86$ (petroleum ether/Et₂O 9:1). $[\alpha]_D^{20}$ 13.21 (c 2.6, 1152 DCM). IR (ATR) ν = 3330, 2926, 2854, 1461, 1253, 1054 cm⁻¹. ¹H 1153 NMR (400 MHz, CDCl₃) δ = 0.04 (s, 6H, Si(CH₃)₂), 0.78–0.93 (m, 1154 12H, SiC(CH₃)₃, CH₃), 1.14-1.50 (m, 20H, CH₂), 1.51-1.62 (m, 1155 2H, HOCH₂CH₂), 2.04 (q, 2H, J = 6.4 Hz, CH₂CH), 2.18 (t, 2H, J = 11566.9 Hz, CHCH₂), 3.54–3.74 (m, 3H, HOCH₂CH₂, CH₂CHO), 1157 5.30–5.50 (m, 2H, CH=CH). ¹³C NMR (101 MHz, CDCl₃) δ = 1158 -4.58 (SiCH₃), -4.37 (SiCH₃), 14.07 (CH₃), 18.12 (SiC(CH₃)₃), 1159 22.61 (CH₂), 25.39 (CH₂), 25.72 (CH₂), 25.90 (SiC(CH₃)₃), 27.43 1160 (CH₂CH), 29.26 (CH₂), 29.38 (CH₂), 29.46 (CH₂), 29.49 (CH₂), 1161 29.64 (CH₂), 31.87 (CH₂), 32.77 (HOCH₂CH₂) 35.24 (CHCH₂), 1162 36.84 (CH₂), 63.00 (HOCH₂CH₂), 72.40 (CH₂CHO), 125.91 1163 (CH=CH), 131.36 (CH=CH). 1164 g



(12*R*)-[(*tert*-Butyldimethylsilyl)oxy]-1-iodooctadec-(9Z)-ene 1165 (35). General procedure VI was applied to compound 34 (1.8 g, 4.51 1166 mmol), iodine (1.37 g, 5.42 mmol), PPh₃ (1.42 g, 5.42 mmol), and 1167 imidazole (921 mg, 13.53 mmol) in toluene (20 mL). Compound 35 1168 was afforded after silica gel column chromatography (petroleum 1169 ether) as a colorless oil (1.86 g, 81%). $R_f = 0.1$ (petroleum ether/Et₂O 1170 9:1). [α]²⁰_D 7.12 (c 0.6, DCM). IR (ATR) ν = 2925, 2854, 1461, 1252, 1171

1172 1063 cm^{-1.} ¹H NMR (400 MHz, CDCl₃) δ = 0.05 (s, 6H, Si(CH₃)₂), 1173 0.80–0.97 (m, 12H, SiC(CH₃)₃, CH₃), 1.15–1.49 (m, 20H, CH₂), 1174 1.71–1.92 (m, 2H, ICH₂CH₂), 2.02 (q, 2H, *J* = 6.4 Hz, CH₂CH), 1175 2.18 (t, 2H, *J* = 6.9 Hz, CHCH₂), 3.18 (t, 2H, *J* = 7.1 Hz, ICH₂CH₂), 1176 3.57–3.75 (m, 1H, CH₂CHO), 5.29–5.52 (m, 2H, CH=CH). ¹³C 1177 NMR (101 MHz, CDCl₃) δ = -4.56 (SiCH₃), -4.35 (SiCH₃), 7.19 1178 (ICH₂), 14.09 (CH₃), 18.13 (SiC(CH₃)₃), 22.63 (CH₂), 25.40 (CH₂), 1180 29.31 (CH₂), 29.47 (CH₂), 29.61 (CH₂), 30.48 (CH₂), 31.89 (CH₂), 1181 33.55 (ICH₂CH₂), 35.25 (CHCH₂), 36.86 (CH₂), 72.38 (CH₂CHO), 1182 125.97 (CH=CH), 131.30 (CH=CH).

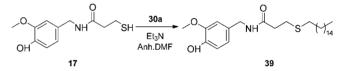


Octadec-(9Z)-ene-1-(12R)-diol (36). General procedure V was 1183 1184 applied to methyl ricinoleate 4 (2.50 g, 8 mmol) with LiAlH₄ (607 1185 mg, 16 mmol) in anhydrous THF (40 mL). Compound 36¹ 1186 afforded after silica gel column chromatography (petroleum ether/ 1187 Et₂O 9:1) as a colorless oil (1.95 g, 86%). $R_f = 0.82$ (petroleum ether/ 1188 Et₂O 9:1). IR (ATR) ν = 3329, 2923, 2853, 1458, 1053 cm⁻¹. ¹H 1189 NMR (400 MHz, CDCl₃) δ = 0.87 (t, 3H, J = 6.9 Hz, CH₃), 1.19-1190 1.39 (m, 18H, CH₂), 1.40-1.49 (m, 2H, CH₂), 1.51-1.58 (m, 2H, 1191 HOCH₂CH₂), 1.59 (br s, 2H, OH), 2.04 (q, 2H, J = 6.4 Hz, 1192 CH₂CH), 2.20 (t, 2H, J = 6.9 Hz, CHCH₂), 3.62 (m, 3H, 1193 HOCH₂CH₂, CH₂CHO), 5.29–5.47 (m, 1H, CH=CH), 5.47– 1194 5.66 (m, 1H, CH=CH). ¹³C NMR (101 MHz, CDCl₃) δ = 14.06 1195 (CH₃), 22.59 (CH₂), 25.68 (CH₂), 25.69 (CH₂), 27.36 (CH₂CH), 1196 29.17 (CH₂), 29.31 (CH₂), 29.33 (CH₂), 29.40 (CH₂), 29.59 (CH₂), 1197 31.81 (CH₂), 32.73 (HOCH₂CH₂), 35.32 (CHCH₂), 36.81 (CH₂), 1198 62.96 (HOCH₂CH₂), 71.49 (CH₂CHO), 125.14 (CH=CH), 133.39 1199 (CH=CH)

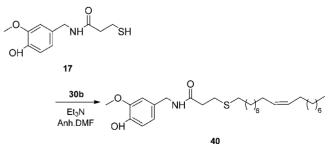
(12'R)-Hydroxyoctadec-(9'Z)-en-1-yl-4-methylbenzene-12.00 1201 sulfonate (37). To a solution of compound 36 (1.6 g, 5.62 mmol) in 1202 a mixture of DCM and pyridine (6 mL, 5:5) were added TsCl (1.07 g, 1203 5.62 mmol, 1 equiv) in portions and DMAP (27 mg, 0.22 mmol). The 1204 mixture was stirred at room temperature for 20 h. The mixture was 1205 washed with 1 M HCl and extracted with EtOAc. The organic phase 1206 was dried over Na2SO4, and the solvent was removed under reduced 1207 pressure. Compound 37¹⁹ was afforded after silica gel column 1208 chromatography (petroleum ether/Et₂O 7:3) as a yellow oil (1.11 g, 1209 45%). $R_f = 0.84$ (petroleum ether/Et₂O 7:3). $[\alpha]_D^{20}$ 4.40 (c 1.4, 1210 DCM). IR (ATR) $\nu = 2924, 2854, 1458, 1358 \text{ cm}^{-1}$. H NMR (400 1211 MHz, CDCl₃) $\delta = 0.88$ (t, 3H, J = 6.9 Hz, CH₃), 1.11–1.39 (m, 18H, 1212 CH₂), 1.39–1.54 (m, 2H, CH₂), 1.53–1.70 (m, 2H, OCH₂CH₂) 2.03 1213 (q, 2H, J = 6.4 Hz, CH₂CH), 2.20 (t, 2H, J = 6.9 Hz, CHCH₂), 2.44 1214 (s, 3H, CH₂C), 3.54-3.71 (m, 1H, CH₂CHO), 4.01 (t, 2H, J = 6.91215 Hz, OCH₂CH₂), 5.31-5.47 (m, 1H, CH=CH), 5.48-5.68 (m, 1H, 1216 CH=CH), 7.33 (d, 2H, J = 8.5 Hz, H_{Ar}), 7.78 (d, 2H, J = 7.9 Hz, 1217 H_{Ar}). ¹³C NMR (101 MHz, CDCl₃) δ = 14.06 (CH₃), 21.60 (CH₃C), 1218 22.59 (CH₂), 25.28 (CH₂), 25.69 (CH₂), 27.35 (CH₂CH), 28.78 1219 (OCH₂CH₂), 28.84 (CH₂), 29.10 (CH₂), 29.22 (CH₂), 29.32 (CH₂), 1220 29.56 (CH₂), 31.81 (CH₂), 35.34 (CHCH₂), 36.83 (C-CH₂), 70.64 1221 (OCH₂CH₂), 71.45 (CH₂CHO), 125.23 (CH=CH), 127.84 (2 × 1222 C_{Ar}), 129.76 (2 × C_{Ar}), 133.22 (C_{Ar}), 133.27 (CH=CH), 144.58 1223 (C_{Ar}).

g

1"-Hexyl-12"-(Tosyloxy)dodec-(3"Z)-en-(1"R)-yl-2-phenyla- 1224 cetate (38). To a solution of compound 37 (900 mg, 2.05 mmol) in 1225 anhydrous toluene (10 mL), phenylacetic acid (307 mg, 2.25 mmol, 1226 1.1 equiv), DCC (1.02 g, 5.13 mmol, 2.5 equiv), and DMAP (500 mg, 1227 4.1 mmol, 2 equiv) were added. The mixture was left stirred at room 1228 temperature overnight and then filtered off to remove DCU. The 1229 solvent was partially evaporated; the crude was dissolved in EtOAc 1230 and washed with 1 M HCl, water, and brine. The organic phase was 1231 dried over Na₂SO₄ and the solvent was removed under reduced 1232 pressure. Compound 38 was afforded after silica gel column 1233 chromatography (petroleum ether/EtOAc 8:2) as a colorless oil 1234 (935 mg, 82%). $R_f = 0.53$ (petroleum ether/EtOAc 8:2). $[\alpha]_D^{20}$ 16.91 1235 (c 5, DCM). IR (ATR) ν = 2925, 2855, 1730, 1361, 1187 cm⁻¹. ¹H 1236 NMR (400 MHz, CDCl₃) $\delta = 0.87$ (t, 3H, J = 6.9 Hz, CH₃), 1.11- 1237 1.39 (m, 18H, CH₂), 1.42-1.56 (m, 2H, CH₂), 1.58-1.67 (m, 2H, 1238 OCH_2CH_2), 1.97 (q, 2H, J = 6.4, CH_2CH_2), 2.13–2.38 (m, 2H, 1239 $CHCH_2$), 2.44 (s, 3H, CH_3C) 3.58 (s, 2H, $COCH_2$), 4.01 (t, 2H, J = 12406.9 Hz, OCH₂CH₂), 4.87 (p, 1H, J = 6.1 Hz, CH₂CHO), 5.19–5.37 1241 (m, 1H, CH=CH), 5.37-5.55 (m, 1H, CH=CH), 7.19-7.43 (m, 1242 7H, H_{Ar}), 7.79 (d, 2H, J = 8.0 Hz, H_{Ar}). ¹³C NMR (101 MHz, 1243 $CDCl_3$) $\delta = 14.04 (CH_3), 21.61 (CH_3C), 22.50 (CH_2), 25.17 (CH_2), 1244$ 25.31 (CH₂), 27.27 (CH₂CH), 28.80 (OCH₂CH₂), 28.88 (CH₂), 1245 29.04 (CH₂), 29.13 (CH₂), 29.27 (CH₂), 29.49 (CH₂), 31.66 (CH₂), 1246 31.89 (CHCH₂), 33.53 (CH₂), 41.74 (COCH₂), 70.64 (OCH₂CH₂), 1247 74.44 (CH₂CHO), 124.15 (CH=CH), 126.92 (C_{Ar}), 127.85 (2 \times 1248 C_{Ar}), 128.44 (2 × C_{Ar}), 129.20 (2 × C_{Ar}), 129.76 (2 × C_{Ar}), 132.57 1249 (CH=CH), 133.25 (C_{Ar}), 134.31 (C_{Ar}), 144.57 (C_{Ar}), 171.27 1250 (OCOCH₂). HR-MS (ESI⁺), m/z: [M+NH₄]⁺ calcd for $C_{33}H_{52}NO_5S$, 1251 574.3561; found 573.3563. 1252 g

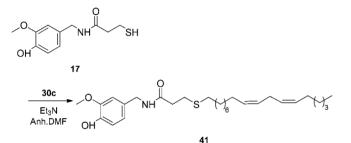


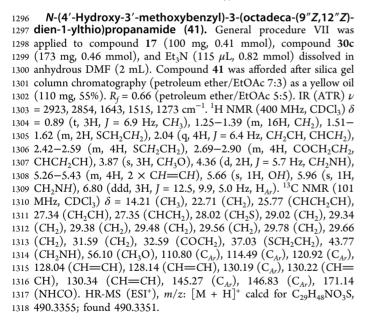
3-(Hexadecylthio)-N-(4'-hydroxy-3'-methoxybenzyl)- 1253 propanamide (39). General procedure VII was applied to 32 (150 1254 mg, 0.62 mmol), compound 30a (245 mg, 0.70 mmol), and Et₃N 1255 (175 µL, 1.24 mmol) dissolved in anhydrous DMF (4 mL). 1256 Compound 39 was afforded after silica gel column chromatography 1257 (petroleum ether/EtOAc 7:3) as a white solid (136 mg, 42%). Mp = 1258 72-73 °C. $R_f = 0.48$ (petroleum ether/EtOAc 5:5). IR (ATR) $\nu = 1259$ 2925, 2855, 1730, 1361, 1187 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ = 1260 0.88 (t, 3H, J = 6.9 Hz, CH_3), 1.23–1.32 (m, 24H, CH_2), 1.56–1.60 1261 (m, 4H, SCH₂CH₂), 2.40-2.58 (m, 4H, COCH₂S, SCH₂CH₂), 2.84 1262 $(t, 2H, J = 6.9 \text{ Hz}, CH_2S)$, 3.88 $(s, 3H, CH_3O)$, 4.37 (d, 2H, J = 5.7 1263)Hz, CH₂NH), 5.59 (s, 1H, CH₂NH), 5.90 (br s, 1H, OH), 6.81 (ddd, 1264 3H, J = 12.5, 9.9, 5.0 Hz, H_{Ar}). ¹³C NMR (101 MHz, CDCl₃) $\delta = 1265$ 14.28 (CH₃), 22.85 (CH₂), 28.04 (CH₂S), 29.05 (CH₂), 29.40 (CH₂), 1266 29.52 (CH₂), 29.69 (CH₂), 29.77 (CH₂), 29.81 (3 × CH₂), 29.85 (4 1267 × CH₂), 32.08 (CH₂), 32.63 (COCH₂), 37.07 (SCH₂CH₂), 43.80 1268 (CH_2NH) , 56.13 (CH_3O) , 110.80 (C_{Ar}) , 114.49 (C_{Ar}) , 120.97 (C_{Ar}) , 1269 130.24 (C_{Ar}), 145.28 (C_{Ar}), 146.84 (C_{Ar}), 171.12 (NHCO). HR-MS 1270 (ESI⁺), m/z: [M + H]⁺ calcd for $C_{27}H_{48}NO_3S$, 466.3355; found 1271 466.3378. 1272 g

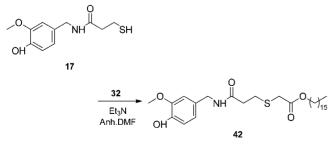


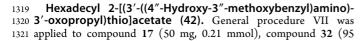
N-(4'-Hydroxy-3'-methoxybenzyl)-3-(octadec-(9"*Z*)-en-1- 1273 ylthio)propanamide (40). General procedure VII was applied to 1274 compound 17 (100 mg, 0.41 mmol), compound 30b (174 mg, 0.46 1275

1276 mmol), and Et₃N (115 μ L, 0.82 mmol) dissolved in anhydrous DMF 1277 (2 mL). Compound 40 was afforded after silica gel column 1278 chromatography (petroleum ether/EtOAc 5:5) as a white sticky 1279 solid (83 mg, 41%). $R_f = 0.73$ (petroleum ether/EtOAc 5:5). IR 1280 (ATR) ν = 3505, 3323, 2919, 2851, 1640, 1519 cm⁻¹. ¹H NMR (400 1281 MHz, CDCl₃) $\delta = 0.88$ (t, 3H, J = 6.9 Hz, CH₃), 1.23–1.37 (m, 22H, 1282 CH₂), 1.51–1.61 (m, 2H, SCH₂CH₂), 2.01 (q, 4H, J = 6.4 Hz, 1283 CH₂CH, CHCH₂), 2.44-2.55 (m, 4H, COCH₂, SCH₂CH₂), 2.83 (t, 1284 2H, J = 6.9 Hz, COCH₂CH₂), 3.88 (s, 3H, CH₃O), 4.37 (d, 2H, J =1285 5.7 Hz, CH₂NH), 5.28-5.40 (m, 2H, CH=CH), 5.64 (s, 1H, OH), 1286 5.94 (br s, 1H, CH₂NH), 6.81 (ddd, 3H, I = 12.5, 9.9, 5.0 Hz, H_{Ar}). $_{1287}$ ¹³C NMR (101 MHz, CDCl₃) δ = 14.26 (CH₃), 22.82 (CH₂), 27.33 1288 (CH₂CH), 27.36 (CHCH₂), 28.03 (CH₂S), 29.03 (CH₂), 29.35 1289 (CH₂), 29.39 (CH₂), 29.46 (2 × CH₂), 29.57 (CH₂), 29.66 (CH₂), 1290 29.76 (CH₂), 29.88 (CH₂), 29.91 (CH₂), 32.04 (CH₂), 32.61 1291 (COCH₂), 37.08 (SCH₂CH₂), 43.77 (CH₂NH), 56.11 (CH₃O), 1292 110.80 (C_{Ar}) , 114.49 (C_{Ar}) , 120.93 (C_{Ar}) , 129.93 (CH=CH), 130.11 1293 (CH=CH), 130.21 (C_{Ar}), 145.27 (C_{Ar}), 146.83 (C_{Ar}), 171.13 1294 (NHCO). HR-MS (ESI⁺), m/z: $[M + H]^+$ calcd for $C_{29}H_{50}NO_3S$, 1295 492.3511; found 492.3502.

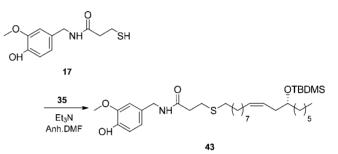




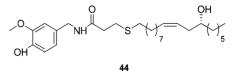




mg, 0.23 mmol), and Et₃N (60 μ L, 0.42 mmol) dissolved in 1322 anhydrous DMF (2 mL). Compound 42 was afforded after silica gel 1323 column chromatography (petroleum ether/EtOAc 6:4) as a white 1324 solid (75 mg, 68%). Mp: 59–60 °C. $R_f = 0.61$ (petroleum ether/ 1325 EtOAc 5:5). IR (ATR) ν = 3370, 3278, 2955, 2917, 2849, 1726, 1269 1326 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ = 0.88 (t, J = 6.9 Hz, 3H, CH₃), 1327 1.24-1.33 (m, 26H, CH₂), 1.57-1.65 (m, 2H, COOCH₂CH₂), 2.53 1328 $(t, 2H, J = 6.9 \text{ Hz}, \text{COCH}_2), 2.97 (t, 2H, J = 6.9 \text{ Hz}, \text{COCH}_2\text{CH}_2), 1329$ 3.24 (s, 2H, SCH₂), 3.88 (s, 3H, CH₃OH), 4.06 (t, 2H, J = 6.9 Hz, 1330 $COOCH_2CH_2$), 4.37 (d, 2H, J = 5.7 Hz, CH_2NH), 5.63 (br s, 1H, 1331 OH), 6.09 (br s, 1H, CH₂NH), 6.80 (ddd, 3H, J = 12.5, 9.9, 5.0 Hz, 1332 H_{4r}). ¹³C NMR (101 MHz, CDCl₃) $\delta = 14.26$ (CH₃), 22.83 (CH₂), 1333 25.96 (CH₂), 28.65 (CH₂), 29.26 (CH₂S), 29.36 (CH₂), 29.50 (CH₂), 1334 29.65 (CH₂), 29.72 (CH₂), 29.79 (CH₂), 29.79 (CH₂), 29.82 (CH₂), 1335 29.83 $(3 \times CH_2)$, 32.06 (CH_2) , 34.40 (SCH_2) , 36.55 $(COCH_2)$, 1336 43.76 (CH₂NH), 56.12 (CH₃O), 65.91 (COOCH₂), 110.77 (C_{Ar}), 1337 114.44 (C_{Ar}), 120.91 (C_{Ar}), 130.22 (C_{Ar}), 145.23 (C_{Ar}), 146.83 (C_{Ar}), 1338 170.75 (NHCO), 170.80 (COOCH₂). HR-MS (ESI⁺), m/z: [M + 1339 H]⁺ calcd for C₂₉H₅₀NO₅S, 524.3404; found 524.3437. 1340 g

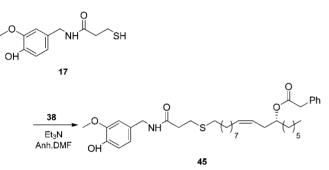


N-(4'-Hydroxy-3'-methoxybenzyl)-3-[(((12"R)-tert-butyl- 1341 dimethylsilyl)oxy)octadec-(9"Z)-en-1-ylthio]propanamide 1342 (43). General procedure VII was applied to compound 17 (100 mg, 1343 0.41 mmol), compound 35 (236 mg, 0.46 mmol), and Et₃N (120 μ L, 1344 0.82 mmol) dissolved in DMF (2 mL). Compound 43 was afforded 1345 after silica gel column chromatography (petroleum ether/EtOAc 5:5) 1346 as a yellow oil (135 mg, 53%). $R_f = 0.45$ (petroleum ether/EtOAc 1347 5:5). $\left[\alpha\right]_{D}^{20}$ -4.71 (c 0.45, DCM). IR (ATR) ν = 3370, 3278, 2955, 1348 2917, 2849, 1726, 1269 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ = 0.03 1349 (s, 6H, Si(CH₃)₂), 0.73-0.94 (m, 12H, SiC(CH₃)₃, CH₃), 1.14-1.42 1350 (m, 20H, CH_2), 1.47–1.67 (m, 2H, SCH_2CH_2), 2.00 (q, 2H, J = 6.4 1351 Hz, CH₂CH), 2.11-2.26 (m, 2H, CHCH₂), 2.41-2.57 (m, 4H, 1352 COCH₂, SCH₂CH₂), 2.83 (t, 2H, J = 6.9 Hz, COCH₂CH₂), 3.55- 1353 3.74 (m, 1H, CH₂CHO), 3.86 (s, 3H, CH₃O), 4.34 (d, 2H, J = 5.7 1354 Hz, CH₂NH), 5.27-5.51 (m, 2H, CH=CH), 5.76 (s, 1H, OH), 6.03 1355 (s, 1H, CH₂NH), 6.79 (ddd, 3H, J = 12.5, 9.9, 5 Hz, H_{Ar}). ¹³C NMR 1356 $(101 \text{ MHz}, \text{CDCl}_3) \delta = 4.57 (\text{SiCH}_3), -4.36 (\text{SiCH}_3), 14.09 (\text{CH}_3), 1357$ 18.13 (SiC(CH₃)₃), 22.62 (CH₂), 25.38 (CH₂), 25.91 (SiC(CH₃)₃), 1358 27.44 (CH₂CH), 27.87 (CH₂S), 28.87 (CH₂), 29.20 (CH₂), 29.28 1359 (CH₂), 29.44 (CH₂), 29.46 (CH₂), 29.60 (CH₂), 29.65 (CH₂), 31.87 1360 (CH₂), 32.43 (CH₂), 35.24 (CHCH₂), 36.84 (COCH₂, SCH₂CH₂), 1361 43.59 (CH₂NH), 55.93 (CH₃O), 72.38 (CH₂CHO), 110.66 (C_{Ar}), 1362 114.36 (C_{Ar}), 120.74 (C_{Ar}), 125.93 (CH=CH), 130.02 (C_{Ar}), 131.34 1363 (CH=CH), 145.12 (C_{Ar}), 146.71 (C_{Ar}), 171.04 (NHCO). HR-MS 1364 (ESI⁺), m/z: [M + H]⁺ calcd for C₃₅H₆₄NO₄SSi, 622.4307; found 1365 622.4307. 1366 g

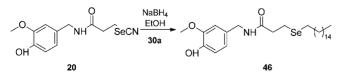


N-(4'-Hydroxy-3'-methoxybenzyl)-3-[((12"*R*)-hydroxy)- 1367 octadec-(9"*Z*)-en-1-ylthio]propanamide (44). General procedure 1368 VIII was applied to compound 43 (100 mg, 0.16 mmol) in AcOH/ 1369 THF/H₂O (1 mL, 6:2:2). Compound 44 was afforded after silica gel 1370 column chromatography (petroleum ether/EtOAc 6:4) as a colorless 1371 oil (66 mg, 81%). $R_f = 0.62$ (petroleum ether/EtOAc 5:5). $[\alpha]_{D}^{20}$ 1372

1373 –1.37 (c 0.4, DCM). IR (ATR) ν = 3290, 2923, 2852, 1645, 1514, 1374 1273 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ = 0.88 (t, 3H, J = 6.9 Hz, 1375 CH₂), 1.21-1.38 (m, 18H, CH₂), 1.41-1.49 (m, 4H, CH₂), 1.51-1376 1.60 (m, 2H, SCH₂CH₂), 2.04 (q, 2H, J = 6.4 Hz, CH₂CH), 2.22 (t, 1377 2H, J = 6.9 Hz, CHCH₂), 2.43–2.55 (m, 4H, COCH₂, SCH₂CH₂), 1378 2.83 (t, 2H, J = 6.9 Hz, COCH₂CH₂), 3.56–3.65 (m, 1H, CH₂CHO), 1379 3.88 (s, 3H, CH₃O), 4.37 (d, 2H, J = 5.7 Hz, CH₂NH), 5.34–5.46 1380 (m,1H, CH=CH), 5.50-5.60 (m, 1H, CH=CH), 6.00 (s, 1H, 1381 CH₂NH), 6.80 (ddd, 3H, J = 12.5, 9.9, 5.0 Hz, H_{Ar}). ¹³C NMR (101 1382 MHz, CDCl₃) δ = 14.23 (CH₃), 22.76 (CH₂), 25.86 (CH₂), 27.53 1383 (CH₂CH), 28.04 (CH₂S), 28.95 (CH₂), 29.28 (CH₂), 29.35 (CH₂), $1384 29.49 (2 \times CH_2), 29.71 (CH_2), 29.76 (CH_2), 31.98 (CH_2), 32.59$ 1385 (SCH₂CH₂), 35.49 (CHCH₂), 36.98 (COCH₂), 36.99 (SCH₂CH₂), 1386 43.81 (CH₂NH), 56.12 (CH₃O), 71.67 (CH₂CHO), 110.83 (C_{Ar}), 1387 114.52 (C_{Ar}), 120.94 (C_{Ar}), 125.31 (CH=CH), 130.13 (C_{Ar}), 133.59 1388 (CH=CH), 145.30 (C_{Ar}), 146.86 (C_{Ar}), 171.25 (NHCO). HR-MS 1389 (ESI⁺), m/z: $[M + H]^+$ calcd for $C_{29}H_{50}NO_4Si$, 508.3461; found 1390 508.3451.

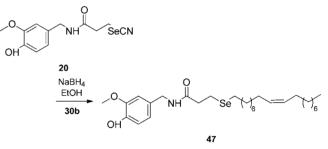


1391 1"-Hexyl-12"-[(3"'-((4""-hydroxy-3""-methoxybenzyl)-1392 amino)-3^m-oxopropyl)thio]dodec-(3^mZ)-en-(1^mR)-yl 2-phenyl-1393 acetate (45). General procedure VII was applied to compound 17 1394 (100 mg, 0.41 mmol), compound **38** (255 mg, 0.46 mmol), and Et₃N 1395 (115 µL, 0.82 mmol) dissolved in anhydrous DMF (2 mL). 1396 Compound 45 was afforded after silica gel column chromatography 1397 (petroleum ether/EtOAc 6:4) as a yellow oil (51 mg, 20%). $R_f = 0.78$ 1398 (petroleum ether/EtOAc 6:4). $[\alpha]_{D}^{20}$ 7.90 (c 0.4, DCM). IR (ATR) ν $1399 = 3290, 2924, 2853, 1729, 1646, 1514 \text{ cm}^{-1}$. ¹H NMR (400 MHz, 1400 CDCl₃) δ = 0.86 (t, 3H, J = 6.9 Hz, CH₃), 1.06–1.40 (m, 18H, CH₂), 1401 1.46–1.60 (m, 4H, CH_2 , SCH_2CH_2), 1.99 (q, 2H, J = 6.4 Hz, 1402 CH₂CH), 2.19–2.35 (m, 2H, CHCH₂), 2.44–2.56 (m, 4H, COCH₂, 1403 SCH₂CH₂), 2.83 (t, 2H, J = 6.9 Hz, COCH₂CH₂), 3.58 (s, 2H, 1404 OCOCH₂), 3.87 (s, 3H, CH₃O), 4.36 (d, 2H, J = 5.7 Hz, CH₂NH), 1405 4.86 (p, 1H, J = 6.2 Hz, CH₂CHO), 5.22–5.32 (m, 1H, CH=CH), 1406 5.39-5.48 (m, 1H, CH=CH), 6.04 (br s, 1H, CH₂NH), 6.80 (ddd, 1407 3H, J = 12.5, 9.9, 5.0 Hz, H_{Ar}), 7.21–7.34 (m, 5H, H_{Ar}). ¹³C NMR 1408 (101 MHz, CDCl₃) δ = 14.20 (CH₃), 22.66 (CH₂), 25.33 (CH₂), 1409 27.45 (CH₂CH), 27.01 (CH₂S), 28.99 (CH₂), 29.20 (CH₂), 29.32 1410 (CH₂), 29.37 (CH₂), 29.55 (CH₂), 29.69 (CH₂), 29.73 (CH₂), 31.82 1411 (CH₂), 32.04 (CHCH₂), 32.57 (COCH₂), 33.69 (CH₂), 36.91 1412 (SCH₂CH₂), 41.90 (OCOCH₂), 43.84 (CH₂NH), 56.11 (CH₃O), 1413 74.65 (CH₂CHO), 110.81 (C_{Ar}), 114.50 (C_{Ar}), 120.94 (C_{Ar}), 124.25 1414 (CH=CH), 127.09 (C_{Ar}), 128.60 (2 × C_{Ar}), 129.36 (2 × C_{Ar}), 1415 130.06 (C_{Ar}), 132.80 (CH=CH), 134.46 (C_{Ar}), 145.30 (C_{Ar}), 146.84 1416 (C_{Ar}), 171.37 (NHCO), 171.48 (OCOCH₂). HR-MS (ESI⁺), m/z: 1417 $[M + H]^+$ calcd for $C_{37}H_{56}NO_5S$, 626.3879; found 626.3870.

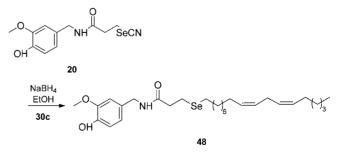


3-(Hexadecylseleno)-*N***-(4'-hydroxy-3'-methoxybenzyl)propanamide (46).** General procedure IV was applied to compound 1420 **20** (100 mg, 0.32 mmol), NaBH₄ (30 mg, 0.8 mmol), and compound 1421 **30a** (126 mg, 0.36 mmol) dissolved in EtOH (2 mL). Compound **46** 1422 was afforded after silica gel column chromatography (petroleum

ether/EtOAc 7:3) as a yellow sticky solid (166 mg, 71%). $R_f = 0.55$ 1423 (petroleum ether/EtOAc 7:3). IR (ATR) $\nu = 3504, 3317, 2917, 2848, 1424$ 1645, 1519 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ = 0.88 (t, 3H, J = 1425 6.9 Hz, CH₃), 1.22–1.36 (m, 26H, CH₂), 1.59–1.68 (m, 2H, 1426 SeCH₂CH₂), 2.53-2.62 (m, 4H, COCH₂, SeCH₂CH₂), 2.83 (t, 2H, J 1427 = 6.9 Hz, CH_2Se), 3.88 (s, 3H, CH_3O), 4.36 (d, 2H, J = 5.7 Hz, 1428 CH₂NH), 5.66 (s, 1H, CH₂NH), 5.88 (br s, 1H, OH), 6.80 (ddd, 3H, 1429 $J = 12.5, 9.9, 5.0 \text{ Hz}, \text{H}_{Ar}$.¹³C NMR (101 MHz, CDCl₃) $\delta = 14.26 \text{ 1430}$ (CH₃), 18.69 (CH₂Se), 22.83 (CH₂), 24.84 (SeCH₂CH₂), 29.31 1431 (CH₂), 29.49 (CH₂), 29.68 (CH₂), 29.75 (CH₂), 29.79 (2 × CH₂), 1432 29.83 $(4 \times CH_2)$, 30.08 (CH_2) , 30.74 (CH_2) , 32.06 (CH_2) , 38.03 1433 (COCH₂), 43.78 (CH₂NH), 56.12 (CH₃O), 110.83 (C_{Ar}), 114.49 1434 (C_{Ar}) , 120.96 (C_{Ar}) , 130.20 (C_{Ar}) , 145.28 (C_{Ar}) , 146.84 (C_{Ar}) , 171.41 1435 (NHCO). HR-MS (ESI⁺), m/z: [M + H]⁺ calcd for C₂₇H₄₈NO₃Se, 1436 514.2799; found 514.2795. 1437 g

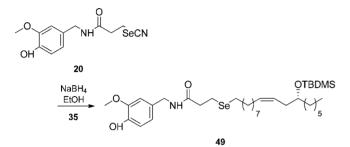


N-(4'-Hydroxy-3'-methoxybenzyl)-3-(octadec-(9"Z)-en-1- 1438 ylseleno)propanamide (47). General procedure IV was applied to 1439 compound 20 (200 mg, 0.64 mmol), NaBH₄ (59 mg, 1.6 mmol), and 1440 compound 30b (271 mg, 0.72 mmol) dissolved in EtOH (2 mL). 1441 Compound 47 was afforded after silica gel column chromatography 1442 (petroleum ether/EtOAc 7:3) as a yellow sticky solid (244 mg, 71%). 1443 $\bar{R}_{f} = 0.71$ (petroleum ether/EtOAc 7:3). IR (ATR) $\nu = 3509, 3321, 1444$ 2919, 2850, 1646, 1519 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ = 0.88 1445 (t, 3H, J = 6.9 Hz, CH_3), 1.24–1.37 (m, 22H, CH_2), 1.60–1.68 (m, 1446 2H, SeCH₂CH₂), 2.01 (q, 4H, J = 6.4 Hz, CH₂CH, CHCH₂), 2.54- 1447 2.61 (m, 4H, COCH₂, SeCH₂CH₂), 2.84 (t, 2H, J = 6.9 Hz, 1448 COCH₂CH₂), 3.88 (s, 3H, CH₃O), 4.37 (d, 2H, J = 5.7 Hz, 1449 CH₂NH), 5.29-5.40 (m, 2H, CH=CH), 5.61 (s, 1H, OH), 5.83 (br 1450 s, 1H, CH₂NH), 6.82 (ddd, 3H, J = 12.5, 9.9, 5.0 Hz, H_{4r}). ¹³C NMR 1451 (101 MHz, CDCl₃) δ = 14.27 (CH₃), 18.70 (CH₂Se), 22.83 (CH₂), 1452 24.84 (SeCH₂CH₂), 27.35 (CH₂CH), 27.37 (CHCH₂), 29.29 (CH₂), 1453 29.40 (CH₂), 29.47 (2 × CH₂), 29.58, (CH₂) 29.67 (CH₂), 29.89 1454 (CH₂), 29.92 (CH₂), 30.08 (CH₂), 30.74 (CH₂), 32.05 (CH₂), 38.06 1455 (COCH₂), 43.80 (CH₂NH), 56.14 (CH₃O), 110.83 (C_{Ar}), 114.48 1456 (C_{Ar}) , 120.99 (C_{Ar}) , 129.94 (CH=CH), 130.11 (CH=CH), 130.22 1457 (C_{Ar}) , 145.29 (C_{Ar}) , 146.84 (C_{Ar}) , 171.37 (NHCO). HR-MS (ESI⁺), 1458 m/z: $[M + H]^+$ calcd for $C_{29}H_{50}NO_3Se$, 540.2956; found 540.2957. 1459 g

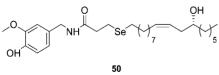


N-(4'-Hydroxy-3'-methoxybenzyl)-3-(octadeca-(9"*Z*,12"*Z*)- 1460 dien-1-ylseleno)propanamide (48). General procedure IV was 1461 applied to compound 20 (100 mg, 0.32 mmol), NaBH₄ (30 mg, 0.80 1462 mmol), and compound 30c (135 mg, 0.36 mmol) dissolved in EtOH 1463 (2 mL). Compound 48 was afforded after silica gel column 1464 chromatography (petroleum ether/EtOAc 7:3) as a yellowish oil 1465 (111 mg, 65%). R_f = 0.7 (petroleum ether/EtOAc 7:3). IR (ATR) ν = 1466 3288, 3008, 2923, 2852, 1644, 1514 cm⁻¹. ¹H NMR (400 MHz, 1467 CDCl₃) δ = 0.88 (t, 3H, *J* = 6.9 Hz, CH₃), 1.25–1.38 (m, 16H, CH₂), 1468

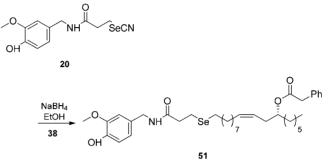
1469 1.59–1.68 (m, 2H, SeCH₂CH₂), 2.04 (q, 4H, J = 6.4 Hz, CH₂CH, 1470 CHCH₂), 2.54–2.61 (m, 4H, COCH₂, SeCH₂CH₂), 2.77 (t, 2H, J =1471 6.9 Hz, 2H, CHCH₂CH), 2.83 (t, 2H, J = 6.9 Hz, COCH₂CH₂), 3.88 1472 (s, 3H, CH₃O), 4.36 (d, 2H, J = 5.7 Hz, CH₂NH), 5.28–5.42 (m, 4H, 1473 2 × CH=CH), 5.66 (s, 1H, OH), 5.88 (br s, 1H, CH₂NH), 6.80 1474 (ddd, 3H, J = 12.5, 9.9, 5.0 Hz, H_A). ¹³C NMR (101 MHz, CDCl₃) δ 1475 = 14.21 (CH₃), 18.69 (CH₂), 22.70 (CH₂), 24.81 (SeCH₂CH₂), 25.77 1476 (CHCH₂CH), 27.33 (CH₂CH), 27.35 (CHCH₂), 29.26 (CH₂), 29.38 1477 (CH₂), 29.48 (CH₂), 29.56 (CH₂), 29.77 (CH₂), 30.06 (CH₂), 30.72 1478 (CH₂), 31.66 (CH₂), 38.02 (COCH₂), 43.78 (CH₂NH), 56.12 1479 (CH₃O), 110.82 (C_{Ar}), 114.48 (C_{Ar}), 120.95 (C_{Ar}), 128.04 (CH= 1480 CH), 128.14 (CH=CH), 130.19 (C_{Ar}), 130.22 (CH=CH), 130.34 1481 (CH=CH), 145.28 (C_{Ar}), 146.83 (C_{Ar}), 171.39 (NHCO). HR-MS 1482 (ESI⁺), m/z: [M + H]⁺ calcd for C₂₉H₄₈NO₃Se, 538.2799; found 1483 538.2761.



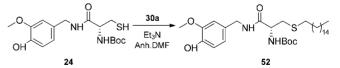
N-(4'-Hydroxy-3'-methoxybenzyl)-3-[(((12"R)-tert-1484 1485 butyldimethylsilyl)oxy)octadec-(9"Z)-en-1-ylseleno]-1486 propanamide (49). General procedure IV was applied to compound 1487 20 (100 mg, 0.32 mmol), NaBH₄ (30 mg, 0.80 mmol), and 1488 compound 35 (233 mg, 0.46 mmol) dissolved in EtOH (2 mL). 1489 Compound 49 was afforded after silica gel column chromatography 1490 (petroleum ether/EtOAc 7:3) as a yellow oil (124 mg, 58%). $R_f =$ 1491 0.54 (petroleum ether/EtOAc 7:3). $[\alpha]_{D}^{20}$ -2.21 (c 0.7, DCM). IR 1492 (ATR) ν = 3288, 2924, 2853, 1645, 1514. ¹H NMR (400 MHz, 1493 $\dot{\text{CDCl}}_3$ $\delta = 0.04$ (s, 6H, Si(CH₃)₂), 0.80–0.97 (m, 12H, SiC(CH₃)₃, 1494 CH₃), 1.15–1.32 (m, 20H, CH₂), 1.52–1.71 (m, 2H, SeCH₂CH₂), 1495 2.01 (q, 2H, J = 6.4 Hz, CH_2CH), 2.18 (t, 2H, J = 6.9 Hz, $CHCH_2$), 1496 2.58 (t, 4H, J = 6.9 Hz, $COCH_2$, $SeCH_2CH_2$), 2.84 (s, 2H, 1497 COCH₂CH₂), 3.58-3.70 (m, 1H, CH₂CHO), 3.89 (s, 3H, CH₃O), 1498 4.37 (d, 2H, J = 5.7 Hz, CH_2NH), 5.32–5.49 (m, 2H, CH=CH), 1499 5.58 (s, 1H, OH), 5.80 (s, 1H, CH₂NH), 6.81 (ddd, 3H, J = 12.5, 9.9, 1500 5.0 Hz, H_{Ar}). ¹³C NMR (101 MHz, CDCl₃) $\delta = -4.56$ (SiCH₃), 1501 -4.36 (SiCH₃), 14.09 (CH₃), 18.14 (SiC(CH₃)₃), 18.55 (CH₂Se), 1502 22.62 (CH₂), 24.71 (SeCH₂CH₂), 25.39 (CH₂), 25.91 (SiC(CH₃)₃), 1503 27.45 (CH₂CH), 29.13 (CH₂), 29.29 (CH₂), 29.44 (CH₂), 29.46 1504 (CH₂), 29.65 (CH₂), 29.68 (CH₂), 29.93 (CH₂), 31.88 (CH₂), 35.25 1505 (CHCH₂), 36.85 (CH₂), 37.90 (COCH₂), 43.64 (CH₂NH), 55.97 1506 (CH₃O), 72.39 (CH₂CHO), 110.65 (C_{Ar}), 114.31 (C_{Ar}), 120.83 1507 (C_{Ar}), 125.93 (CH=CH), 130.05 (C_{Ar}), 131.35 (CH=CH), 145.12 1508 (C_{Ar}), 146.66 (C_{Ar}), 171.19 (NHCO).



1509 **N-(4'-Hydroxy-3'-methoxybenzyl)-3-[((12**"*R*)-hydroxy)-1510 **octadec-(9**"*Z*)-**en-1-ylseleno]propanamide (50).** General proce-1511 dure VIII was applied to compound **49** (100 mg, 0.18 mmol) in 1512 AcOH/THF/H₂O (1 mL, 6:2:2). Compound **50** was afforded after 1513 silica gel column chromatography (petroleum ether/EtOAc 5:5) as a 1514 pale yellow oil (79 mg, 79%). $R_f = 0.77$ (petroleum ether/EtOAc 7:3). 1515 $[\alpha]_D^{20}$ -7.88 (*c* 0.3, DCM). IR (ATR) ν = 3288, 2923, 2852, 1646, 1516 1514 1273 cm^{-1.} ¹H NMR (400 MHz, CDCl₃) δ = 0.88 (t, 3H, *J* = 1517 6.9 Hz, CH₃), 1.21–1.39 (m, 18H, CH₂), 1.42–1.48 (m, 2H, 1518 COHCH₂), 1.58–1.67 (m, 2H, SeCH₂CH₂), 2.04 (q, 2H, *J* = 6.4 Hz, 1519 CH₂CH), 2.20 (t, 2H, *J* = 6.9 Hz, CHCH₂), 2.53–2.61 (m, 4H, COCH₂, SeCH₂CH₂), 2.83 (t, 2H, J = 6.9 Hz, COCH₂CH₂), 3.57– 1520 3.65 (m, 1H, CH₂CHO), 3.87 (s, 3H, CH₃O), 4.36 (d, 2H, J = 5.7 1521 Hz, CH₂NH), 5.34–5.45 (m, 1H, CH=CH), 5.49–5.60 (m, 1H, 1522 CH=CH), 5.73 (br s, 1H, OH), 5.93 (br s, 1H, CH₂NH), 6.80 (ddd, 1523 3H, J = 12.5, 9.9, 5.0 Hz, H_{Ar}). ¹³C NMR (101 MHz, CDCl₃) $\delta = 1524$ 14.23 (CH₃), 18.71 (CH₂Se), 22.76 (CH₂), 24.81 (SeCH₂CH₂), 1525 25.86 (CH₂), 27.53 (CH₂), 29.21 (CH₂), 29.35 (CH₂), 29.49 (2 × 1526 CH₂), 29.75 (CH₂), 30.00 (CH₂), 30.69 (SeCH₂CH₂), 31.98 (CH₂), 1527 35.50 (CHCH₂), 36.98 (CH₂), 38.04 (COCH₂), 43.79 (CH₂NH), 1528 56.13 (CH₃O), 71.65 (CH₂CHO), 110.85 (C_{Ar}), 114.51 (C_{Ar}), 1529 120.97 (C_{Ar}), 125.31 (CH=CH), 130.21 (C_{Ar}), 133.58 (CH=CH), 1530 145.29 (C_{Ar}), 146.85 (C_{Ar}), 171.39 (NHCO). HR-MS (ESI⁺), m/z: 1531 [M + H]⁺ calcd for C₂₉H₅₀NO₄Se, 556.2905; found 556.2901. 1532 g

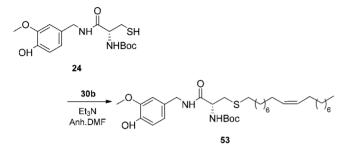


1"-Hexyl-12"-[(3"'-((4""-hydroxy-3""-methoxybenzyl)- 1533 amino)-3"-oxopropyl)seleno]dodec-(3"Z)-en-(1"R)-yl 2-phe- 1534 nylacetate (51). General procedure IV was applied to compound 1535 20 (100 mg, 0.32 mmol), NaBH₄ (30 mg, 0.80 mmol), and 1536 compound 38 (200 mg, 0.36 mmol) dissolved in EtOH (2 mL). 1537 Compound 51 was afforded after silica gel column chromatography 1538 (petroleum ether/EtOAc 5:5) as a yellow oil (155 mg, 72%). $R_f = 1539$ 0.58 (petroleum ether/EtOAc 5:5). [a]²⁰_D 14.78 (c 1.8, DCM). IR 1540 (ATR) $\nu = 3291, 2924, 2853, 1729, 1645, 1514 \text{ cm}^{-1}$. ¹H NMR (400 1541 MHz, CDCl₃) δ = 0.89 (t, 3H, J = 6.9 Hz, CH₃), 1.20–1.40 (m, 18H, 1542 CH₂), 1.50-1.58 (m, 2H, SeCH₂CH₂), 1.61-1.71 (m, 2H, 1543 $COHCH_2$), 2.01 (q, 2H, J = 6.4 Hz, CH_2CH), 2.23–2.37 (m, 2H, 1544 CHCH₂), 2.60 (t, 4H, J = 6.9 Hz, COCH₂, SeCH₂CH₂), 2.86 (t, 2H, 1545 J = 6.9 Hz, COCH₂CH₂), 3.61 (s, 2H, OCOCH₂), 3.89 (s, 3H, 1546 $CH_{3}O$), 4.38 (d, 2H, J = 5.7 Hz, $CH_{2}NH$), 4.90 (p, 1H, J = 6.3 Hz, 1547 CH₂CHO), 5.26-5.35 (m, 1H, CH=CH), 5.42-5.51 (m, 1H, 1548 CH=CH), 5.75 (s, 1H, OH), 5.98 (br s, 1H, CH₂NH), 6.83 (ddd, 1549) 3H, J = 12.5, 9.9, 5.0 Hz, H_{Ar}), 7.16–7.42 (m, 5H, H_{Ar}). ¹³C NMR 1550 (101 MHz, CDCl₃) δ = 14.18 (CH₃), 18.68 (CH₂), 22.63 (CH₂), 1551 24.76 (SeCH₂CH₂), 25.30 (CH₂), 27.43 (CH₂CH), 29.18 (CH₂), 1552 29.23 (CH₂), 29.35 (CH₂), 29.53 (CH₂), 29.66 (CH₂), 30.02 (CH₂), 1553 30.68 (CH₂), 31.79 (CH₂), 32.01 (CHCH₂), 33.66 (CH₂), 37.97 1554 (COCH₂), 41.87 (OCOCH₂), 43.74 (CH₂NH), 56.09 (CH₃O), 1555 74.62 (CH₂CHO), 110.82 (C_{Ar}), 114.48 (C_{Ar}), 120.91 (C_{Ar}), 124.22 1556 (CH=CH), 127.06 (C_{Ar}), 128.57 (2 × C_{Ar}), 129.33 (2 × C_{Ar}), 1557 130.17 (C_{Ar}), 132.78 (CH=CH), 134.42 (C_{Ar}), 145.26 (C_{Ar}), 146.83 1558 (C_{Ar}), 171.41 (NHCO), 171.46 (OCOCH₂). HR-MS (ESI⁺), m/z: 1559 $[M + H]^+$ calcd for $C_{37}H_{56}NO_5Se$, 674.3324; found 674.3315. 1560 g

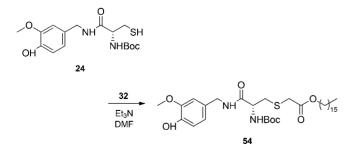


(2*R*)-Boc-amino-3-(hexadecylthio)-*N*-(4'-hydroxy-3'- 1561 methoxybenzyl)propanamide (52). General procedure VII was 1562 applied to compound 24 (200 mg, 0.56 mmol), compound 30a (220 1563 mg, 0.63 mmol), and Et₃N (0.16 mL, 1.12 mmol) in anhydrous DMF 1564 (5 mL). Compound 52 was afforded after silica gel column 1565 chromatography (petroleum ether/EtOAc 6:4) as a white solid 1566 (230 mg, 71%). *R_f* = 0.29 (petroleum ether/EtOAc 5:5). Mp: 76–77 1567 °C. [α]_D²⁰ –2.28 (*c* 0.6, DCM). IR (ATR) ν = 3449, 3336, 2918, 2850, 1568 1681, 1659, 1513 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ = 0.87 (t, 3H, 1569

1570 *J* = 6.9 Hz, CH₃), 1.15–1.35 (m, 26H, CH₂), 1.42 (s, 9H, C(CH₃)₃), 1571 1.47–1.60 (m, 2H, SCH₂CH₂), 2.52 (td, 2H, *J* = 6.9, 1.7 Hz, 1572 SCH₂CH₂), 2.84 (dd, 1H, *J* = 13.7, 6.9 Hz, CHCH₂S), 2.98 (dd, 1H *J* 1573 = 13.7, 5.5 Hz, CHCH₂S), 3.86 (s, 3H, CH₃O), 4.25 (d, 1H, *J* = 5.7 1574 Hz, CH₂NH), 4.29–4.45 (m, 2H, CHCH₂S), 5.39 (d, 1H, *J* = 5.7 Hz, 1575 CH₂NH), 5.70 (s, 1H, OH), 6.67 (t, *J* = 5.5 Hz, 1H, NHBoc), 6.78 1576 (ddd, 3H, *J* = 12.5, 9.9, 5.0 Hz, H₄). ¹³C NMR (101 MHz, CDCl₃) δ 1577 = 14.25 (CH₃), 22.82 (CH₂), 28.39 (C(CH₃)₃), 28.92 (CH₂), 29.36 1578 (CH₂), 29.49 (CH₂), 29.65 (CH₂), 29.74 (CH₂), 29.78 (2 × CH₂), 1580 34.61 (CHCH₂S), 43.68 (CH₂NH), 54.25 (CHCH₂S), 56.08 1581 (CH₃O), 80.59 (C(CH₃)₃), 110.63 (C_A), 114.50 (C_A), 120.76 1582 (C_A), 129.81 (C_A), 145.24 (C_A), 146.83 (C_A), 155.51 (NHCO₂), 1584 C₃₂H₅₇N₂O₅S, 581.3988; found 581.3978.

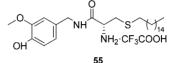


(2R)-Boc-amino-N-(4'-hydroxy-3'-methoxybenzyl)-3-(octa-1585 1586 dec-(9"Z)-en-1-ylthio)propanamide (53). General procedure VII 1587 was applied to compound 24 (100 mg, 0.42 mmol), compound 30b 1588 (179 mg, 0.47 mmol), and Et₃N (117 µL mL, 0.84 mmol) dissolved 1589 in DMF (2 mL). Compound 53 was afforded after silica gel column 1590 chromatography (petroleum ether/EtOAc 7:3) as a white solid (127 1591 mg, 50%). Mp: 43-44 °C. $R_f = 0.58$ (petroleum ether/EtOAc 7:3). 1592 $[\alpha]_D^{20}$ 0.26 (c 1.2, DCM). IR (ATR) ν = 3450, 3333, 2918, 2850, 1593 1514, 1240 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ = 0.88 (t, 3H, J = 1594 6.9 Hz, CH₃), 1.18-1.38 (m, 22H, CH₂), 1.42 (s, 9H, C(CH₃)₃), 1595 1.48–1.61 (m, 2H, SCH_2CH_2), 2.01 (q, 4H, J = 6.4 Hz, CH_2CH_2) 1596 CHCH₂), 2.45–2.58 (m, 2H, SCH₂CH₂), 2.84 (dd, 1H, J = 13.7, 6.91597 Hz, $CHCH_2S$), 3.00 (dd, 1H, J = 13.7, 5.5 Hz, $CHCH_2S$), 3.88 (s, 1598 3H, $CH_{3}O$), 4.24 (dd, 1H, J = 12.5, 6.1 Hz $CH_{2}NH$), 4.30–4.48 (m, 1599 2H, CHCH₂S), 5.22–5.44 (m, 3H, CH=CH, CH₂NH), 5.59 (s, 1H, 1600 OH), 6.61 (t, 1H, J = 5.5 Hz, NHBoc), 6.80 (ddd, 3H, J = 12.5, 9.9, 1601 5.0 Hz, H_{Ar}). ¹³C NMR (101 MHz, CDCl₃) δ = 14.10 (CH₃), 22.66 1602 (CH₂), 27.18 (CH₂CH), 27.20 (CHCH₂), 28.24 (C(CH₃)₃), 28.76 1603 (CH₂), 29.18 (CH₂), 29.23 (CH₂), 29.29 (CH₂), 29.30 (CH₂), 29.40 1604 (SCH₂CH₂), 29.50 (CH₂), 29.59 (CH₂), 29.68 (CH₂), 29.73 (CH₂), 1605 29.75 (CH₂), 31.88 (CH₂), 32.66 (SCH₂CH₂), 34.44 (CHCH₂S), 1606 43.55 (CH₂NH), 54.12 (CHCH₂S), 55.94 (CH₃O), 80.57 $1607 (C(CH_3)_3), 110.45 (C_{Ar}), 114.31 (C_{Ar}), 120.64 (C_{Ar}), 129.68$ 1608 (C_{Ar}), 129.76 (CH=CH), 129.95 (CH=CH), 145.10 (C_{Ar}), 1609 146.65 (C_{Ar}), 155.55 (NHCO₂), 170.37 (NHCO). HR-MS (ESI⁺), 1610 m/z: $[M + H]^+$ calcd for C₃₄H₅₉N₂O₅S, 607.4145; found 607.4138.

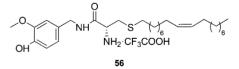


1611 Hexadecyl 2-[((2'*R*)-Boc-amino-3'-((4"-hydroxy-3"-1612 methoxybenzyl)amino)-3'-oxopropyl)thio]acetate (54). Gener-1613 al procedure VII was applied to compound 24 (200 mg, 0.56 mmol), 1614 compound 35 (258 mg, 0.63 mmol), and Et_3N (160 μ L, 1.12 mmol) 1615 dissolved in anhydrous DMF (2 mL). Compound 54 was afforded

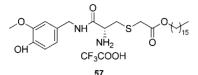
after silica gel column chromatography (petroleum ether/EtOAc 7:3) 1616 as a white solid (282 mg, 79%). Mp: 74–75 °C. $R_f = 0.75$ (petroleum 1617 ether/EtOAc 7:3). $[\alpha]_D^{20}$ -8.04 (c 1, MeOH). IR (ATR) ν = 3493, 1618 3326, 2917, 2849, 1655, 1518 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ = 1619 0.88 (t, J = 6.9 Hz, 3H, CH₃), 1.17–1.35 (m, 26H, CH₂), 1.42 (s, 9H, 1620 $C(CH_3)_3$, 1.55–1.65 (m, 2H, COOCH₂CH₂), 2.88 (dd, 1H, J = 1621 13.7, 6.9 Hz, CHCH₂S), 3.07 (dd, 1H, J = 13.7, 6.9 Hz, CHCH₂S), 1622 3.35 (s, 2H, SCH₂), 3.87 (s, 3H, CH₃OH), 4.07 (t, 2H, J = 6.9 Hz, 1623 COOCH2CH2), 4.25-4.49 (m, 3H, COCHCH2, CH2NH), 5.47- 1624 5.69 (m, 2H, CH₂NH, OH), 6.73–6.87 (m, 3H, H_{Ar}), 7.04 (t, 1H, J = 16255.0 Hz, NHBoc). ¹³C NMR (101 MHz, CDCl₃) δ = 14.09 (CH₃), 1626 22.66 (CH₂), 25.78 (CH₂), 28.26 (C(CH₃)₃), 28.44 (CH₂), 29.20 1627 (CH₂), 29.33 (CH₂), 29.48 (CH₂), 29.55 (CH₂), 29.62 (CH₂), 29.63 1628 (CH_2) , 29.65 (CH_2) , 29.67 $(3 \times CH_2)$, 31.90 (CH_2) , 34.70 1629 (SCH₂CH₂), 35.89 (CHCH₂S), 43.50 (CH₂NH), 53.59 (CHCH₂S), 1630 55.93 (CH₃O), 66.07 (COOCH₂), 80.35 (C(CH₃)₃), 110.42 (C_{Ar}), 1631 114.28 (C_{Ar}), 120.61 (C_{Ar}), 129.70 (C_{Ar}), 145.00 (C_{Ar}), 146.62 (C_{Ar}), 1632 155.46 (NHCO₂), 170.00 (NHCO), 171.34 (COOCH₂). HR-MS 1633 (ESI⁺), m/z: [M + H]⁺ calcd for C₃₄H₅₉N₂O₇S, 639.4043; found 1634 639.4040 1635 g



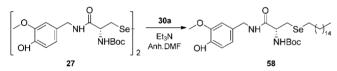
2-(Hexadecylthio)-1-[N-(4'-hydroxy-3'-methoxybenzyl)- 1636 carbamoyl]-(1R)-ethylammonium Trifluoroacetate (55). Gen- 1637 eral procedure IX was applied to compound 52 (200 mg, 0.34 mmol), 1638 TFA (0.26 mL, 3.4 mmol) in DCM (1 mL). Compound 55 was 1639 afforded after flushing nitrogen and drying in vacuo as a yellow oil 1640 (195 mg, quantitative). $[\alpha]_{D}^{20}$ -6.67 (c 0.6, DCM). IR (ATR) ν = 1641 3093, 2921, 2852, 1779, 1667, 1153 cm⁻¹. ¹H NMR (400 MHz, 1642 $CDCl_3$) $\delta = 0.88$ (t, 3H, J = 6.9 Hz, CH_3), 1.21–1.31 (m, 26H, CH_2), 1643 1.45-1.54 (m, 2H, SCH₂CH₂), 2.48 (t, 2H, J = 6.9 Hz, SCH₂CH₂), 1644 2.85-3.03 (m, 2H, CHCH2S), 3.83 (s, CH3O), 4.22-4.38 (m, 3H, 1645 CHCH₂S, CH₂NH), 6.52 (br s, 2H, NH₂), 6.68-6.85 (m, 4H, OH, 1646 H_{Ar}), 7.55 (t, 1H, J = 5.0 Hz, CH₂NH). ¹³C NMR (101 MHz, 1647 $CDCl_3$) $\delta = 14.26 (CH_3), 22.85 (CH_2), 28.82 (CH_2), 29.30 (CH_2), 1648$ 29.32 (CH₂), 29.52 (CH₂), 29.65 (CH₂), 29.74 (2 × CH₂), 29.84 1649 (CH_2) , 29.86 $(4 \times CH_2)$, 32.08 (CH_2) , 32.50 (SCH_2CH_2) , 33.06 1650 (CHCH₂S), 44.38 (CH₂NH), 52.72 (CHCH₂S), 56.01 (CH₃O), 1651 110.67 (C_{Ar}), 114.71 (C_{Ar}), 116.86 (CF_3COOH), 120.92 (C_{Ar}), 1652 128.31 (C_{Ar}), 145.52 (C_{Ar}), 146.95 (C_{Ar}), 161.37 (CF₃COOH), 1653 167.54 (NHCO). HR-MS (ESI⁺), m/z: $[M + H]^+$ calcd for 1654 C₂₇H₄₉N₂O₃S, 481.3458; found 481.3497. 1655 g



1-[N-(4'-Hydroxy-3'-methoxybenzyl)carbamoyl]-2-(octa- 1656 dec-(9"Z)-en-1-ylthio)-(1R)-ethylammonium Trifluoroacetate 1657 (56). General procedure IX was applied to compound 53 (100 mg, 1658 0.16 mmol), TFA (120 µL, 1.64 mmol) in DCM (1 mL). Compound 1659 56 was afforded after flushing nitrogen and drying in vacuo as a yellow 1660 oil (98 mg, quantitative). $[\alpha]_{\rm D}^{20}$ 0.62 (c 2.2, DCM). IR (ATR) $\nu = 1661$ 2922, 2853, 1662, 1199, 1133 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ = 1662 0.87 (t, 3H, J = 6.9 Hz, CH_3), 1.21–1.35 (m, 22H, CH_2), 1.43–1.51 1663 $(m, 2H, SCH_2CH_2), 2.00 (q, 4H, J = 6.4 Hz, CH_2CH, CHCH_2), 2.45 1664$ $(t, 2H, J = 6.9 \text{ Hz}, \text{SCH}_2\text{CH}_2), 2.94 (d, 2H, J = 6.0 \text{ Hz}, \text{CHCH}_2\text{S}), 1665$ 3.78 (s, 3H, CH₃O), 4.13-4.34 (m, 3H, CHCH₂S, CH₂NH), 5.26- 1666 5.43 (m, 2H, CH=CH), 6.70 (ddd, 3H, $J = 12.5, 9.9, 5.0 \text{ Hz}, H_{Ar}$), 1667 7.87 (t, 1H, J = 5.0 Hz, CH₂NH). ¹³C NMR (101 MHz, CDCl₃) δ = 1668 14.25 (CH₃), 22.83 (CH₂), 27.37 (CH₂CH, CHCH₂), 28.86 (CH₂), 1669 29.34 (CH₂), 29.41 (CH₂), 29.44 (CH₂), 29.46 (CH₂), 29.47 (CH₂), 1670 29.61 (CH₂), 29.68 (CH₂), 29.82 (CH₂), 29.85 (CH₂), 29.92 (CH₂), 1671 32.05 (CH₂), 32.66 (SCH₂CH₂), 32.96 (CHCH₂S), 44.0 (CH₂NH), 1672 1673 52.77 (CHCH₂S), 55.96 (CH₃O), 110.71 (C_{Ar}), 114.67 (C_{Ar}), 120.74 1674 (C_{Ar}), 128.82 (C_{Ar}), 129.90 (CH=CH), 130.11 (CH=CH), 145.27 1675 (C_{Ar}), 146.93 (C_{Ar}), 167.76 (NHCO). HR-MS (ESI⁺), m/z: [M + 1676 H]⁺ calcd for $C_{31}H_{51}N_2O_3S$, 507.3615; found 507.3616.

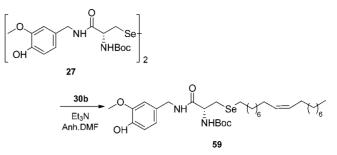


2'-Hexadecyloxy-1-[N-(4"-hydroxy-3"-methoxybenzyl)]-1677 1678 carbamoyl-2-[(oxoethyl)thio]ethan-(1R)-ammonium Trifluor-1679 oacetate (57). General procedure IX was applied to compound 54 1680 (200 mg, 0.31 mmol), TFA (240 µL, 3.1 mmol) in DCM (1 mL). 1681 Compound 57 was afforded after flushing nitrogen and drying in 1682 vacuo as a yellow oil (201 mg, quantitative). $\left[\alpha\right]_{\rm D}^{20}$ -7.53 (c 0.4, 1683 MeOH). IR (ATR) ν = 2917, 2850, 1662, 1176, 1131 cm⁻¹. ¹H NMR 1684 (400 MHz, CDCl₃) δ = 0.88 (t, J = 6.9 Hz, 3H, CH₃), 1.18–1.34 (m, 1685 26H, CH₂), 1.53-1.64 (m, 2H, COOCH₂CH₂), 2.98-3.14 (m, 2H, 1686 CHCH₂S), 3.37 (s, 2H, SCH₂), 3.82 (s, 3H, CH₃OH), 3.99-4.11 (m, 1687 2H, COOCH₂CH₂), 4.22-4.43 (m, 3H, COCHCH₂, H₂NH), 6.67-1688 6.83 (m, 3H, H_{Ar}), 7.94 (t, 1H, J = 5.0 Hz, CH₂NH). ¹³C NMR (101 1689 MHz, CDCl₃) δ = 14.26 (CH₃), 22.84 (CH₂), 25.87 (CH₂), 28.43 $1690 (CH_2), 29.35 (CH_2), 29.51 (2 \times CH_2), 29.64 (CH_2), 29.73 (CH_2),$ 1691 29.81 (CH₂), 29.83 (CH₂), 29.85 (3 × CH₂), 32.08 (CH₂), 34.65 1692 (CH₂), 34.95 (CH₂), 44.24 (CH₂NH), 53.08 (CHCH₂S), 55.99 1693 (CH₃O), 67.26 (COOCH₂), 110.62 (C_{Ar}), 114.64 (C_{Ar}), 120.80 1694 (C_{Ar}), 128.61 (C_{Ar}), 145.35 (C_{Ar}), 146.91 (C_{Ar}), 167.33 (NHCO), 1695 172.72 (COOCH₂). HR-MS (ESI⁺), m/z: [M + H]⁺ calcd for 1696 C₂₉H₅₁N₂O₅S, 539.3513; found 539.3557.

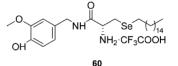


(2R)-Boc-amino-3-(hexadecylseleno)-N-(4'-hydroxy-3'-1697 1698 methoxybenzyl)propanamide (58). General procedure III was 1699 applied to compound 27 (200 mg, 0.25 mmol), NaBH₄ (24 mg, 0.62 1700 mmol), and compound 30a (197 mg, 0.56 mmol) dissolved in EtOH 1701 (2 mL). Compound 58 was afforded after silica gel column 1702 chromatography (petroleum ether/EtOAc 7:3) as a white solid 1703 (231 mg, 74%). $R_f = 0.37$ (petroleum ether/EtOAc 6:4). Mp: 75–76 1704 °C. $[\alpha]_{D}^{20}$ –5.24 (*c* 1.3, DCM). IR (ATR) ν = 3281, 3008, 2924, 2854, 1705 1666, 1516 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) δ = 0.88 (t, 1H, J = 1706 6.9 Hz, CH_3), 1.17–1.38 (m, 26H, CH_2), 1.42 (s, 9H, J = 4.9 Hz, 1707 C(CH₃)₃), 1.58-1.69 (m, 2H, SeCH₂CH₂), 2.46-2.67 (m, 2H, 1708 SeCH₂CH₂), 2.83 (dd, 1H, J = 12.8, 6.9 Hz, CHCH₂Se), 3.05 (dd, 1709 1H, J = 12.8, 5.2 Hz, CHCH₂Se), 3.88 (s, 3H, CH₃O), 4.22-4.36 (m, 1710 1H, CHCH₂Se), 4.37 (d, 2H, J = 5.7 Hz, CH₂NH), 5.33 (s, 1H, 1711 CH₂NH), 5.58 (s, 1H, OH), 6.55 (t, 1H, J = 5.5 Hz, NHBoc), 6.80 1712 (ddd, 3H, J = 12.5, 9.9, 5.0 Hz, H_{Ar}). ¹³C NMR (101 MHz, CDCl₃) δ $1713 = 14.10 (CH_3), 22.67 (CH_2), 25.37 (SeCH_2CH_2), 25.88 (CHCH_2Se),$ 1714 28.24 (C(CH₃)₃), 29.13 (CH₂), 29.34 (CH₂), 29.51 (CH₂), 29.59 1715 (CH₂), 29.63 (3 × CH₂), 29.66 (CH₂), 29.67 (2 × CH₂), 29.81 1716 (CH₂), 30.51 (CH₂), 31.90 (CH₂), 43.54 (CH₂NH), 54.63 1717 (CHCH₂Se), 55.95 (CH₃O), 80.37 (C(CH₃)₃), 110.49 (C_{Ar}), 1718 114.32 (C_{Ar}), 120.65 (C_{Ar}), 129.68 (C_{Ar}), 145.10 (C_{Ar}), 146.67 1719 (C_{Ar}), 155.30 (NHCO₂), 170.46 (NHCO). HR-MS (ESI⁺), m/z: [M $1720 + H^{+}$ calcd for $C_{32}H_{57}N_2O_5Se$, 629.3433; found 629.3431.

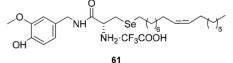
1721 (2*R*)-Boc-amino-*N*-(4'-hydroxy-3'-methoxybenzyl)-3-(octa-1722 dec-(9"Z)-en-1-ylseleno)propanamide (59). General procedure 1723 III was applied to compound 27 (200 mg, 0.25 mmol), NaBH₄ (24 1724 mg, 0.62 mmol) and compound 30b (212 mg, 0.56 mmol) dissolved 1725 in EtOH (2 mL). Compound 59 was afforded after silica gel column 1726 chromatography (petroleum ether/EtOAc 6:4) as a yellow oil (287 1727 mg, 88%). $R_f = 0.66$ (petroleum ether/EtOAc 7:3). $[\alpha]_D^{20} - 4.90$ (c 1728 1.4, DCM). IR (ATR) $\nu = 3444$, 3337, 2919, 2850, 1676, 1511 cm⁻¹. 1729 ¹H NMR (400 MHz, CDCl₃) $\delta = 0.88$ (t, 3H, J = 6.9 Hz, CH_3),



1.16-1.39 (m, 22H, CH₂), 1.42 (s, 9H, C(CH₃)₃), 1.57-1.68 (m, 1730 2H, SeCH₂CH₂), 2.01 (q, 4H, J = 6.4 Hz, CH₂CH, CHCH₂), 2.44- 1731 2.70 (m, 2H, SeCH₂CH₂), 2.83 (dd, 1H, J = 12.8, 6.9 Hz, 1732 $CHCH_2Se$), 3.05 (dd, 1H, J = 12.8, 5.2 Hz, $CHCH_2Se$), 3.88 (s, 1733 3H. $CH_{3}O$), 4.26–4.35 (m, CHCH₂Se), 4.37 (d, 2H, J = 5.7 Hz, 1734 CH₂NH), 5.23–5.43 (m, 3H, CH=CH, CH₂NH), 5.60 (s, 1H, OH), 1735 6.56 (t, 1H, J = 5.5 Hz, NHBoc), 6.79 (ddd, 3H, J = 12.5, 9.9, 5.0 Hz, 1736 H_{Ar}). ¹³C NMR (101 MHz, CDCl₃) δ = 14.10 (CH₃), 22.66 (CH₂), 1737 25.36 (SeCH₂CH₂), 25.90 (CHCH₂Se), 27.18 (CH₂CH), 27.20 1738 $(CHCH_2)$, 28.24 $(C(CH_3)_3)$, 29.11 (CH_2) , 29.23 (CH_2) , 29.30 (2×1739) CH₂), 29.41 (CH₂), 29.50 (CH₂), 29.72 (CH₂), 29.75 (CH₂), 29.80 1740 (CH₂), 30.50 (CH₂), 31.88 (CH₂), 43.55 (CH₂NH), 54.42 1741 $(CHCH_2Se)$, 55.95 (CH_3O) , 80.57 $(C(CH_3)_3)$, 110.48 (C_{Ar}) , 1742 114.31 (C_{Ar}), 120.66 (C_{Ar}), 129.68 (C_{Ar}), 129.76 (CH=CH), 1743 129.95 (CH=CH), 145.10 (C_{Ar}), 146.65 (C_{Ar}), 155.54 (NHCO₂), 1744 170.43 (NHCO). HR-MS (ESI⁺), m/z: [M + H]⁺ calcd for 1745 C34H59N2O5Se, 655.3589; found 655.3583. 1746 g



2-(Hexadecylseleno)-1-[N-(4'-hydroxy-3'-methoxybenzyl)- 1747 carbamoyl]-(1R)-ethylammonium Trifluoroacetate (60). Gen- 1748 eral procedure IX was applied to compound 58 (200 mg, 0.32 mmol), 1749 TFA (240 µL, 3.2 mmol) in DCM (1 mL). Compound 60 was 1750 afforded after flushing nitrogen and drying in vacuo as a yellow oil 1751 (201 mg, quantitative). $[\alpha]_D^{20}$ 0.65 (c 1.4, MeOH). IR (ATR) ν = 1752 3425, 3316, 2916, 2849, 1658, 1187 cm⁻¹. ¹H NMR (400 MHz, 1753 $CDCl_3$) $\delta = 0.88$ (t, 3H, J = 6.9 Hz, CH_3), 1.20–1.34 (m, 26H, CH_2), 1754 1.53-1.61 (m, 2H, SeCH₂CH₂), 2.55 (t, 2H, J = 6.9 Hz, SeCH₂CH₂), 1755 2.85-3.01 (m, 2H, CHCH2Se), 3.82 (s, 3H, CH3O), 4.21-4.37 (m, 1756 3H, CHCH₂Se, CH₂NH), 6.73 (ddd, 3H, J = 12.5, 9.9, 5.0 Hz, H_{Ar}), 1757 7.53 (t, 1H, J = 5.0 Hz, CH₂NH), 7.98 (br s, 1H, OH), 9.42 (br s, 2H, 1758 NH₂). ¹³C NMR (101 MHz, CDCl₃) δ = 14.25 (CH₃), 22.84 (CH₂), 1759 23.51 (CHCH₂Se), 25.89 (CH₂), 27.72 (CH₂), 29.22 (CH₂), 29.51 1760 (CH₂), 29.64 (CH₂), 29.73 (CH₂), 29.80 (CH₂), 29.81 (CH₂), 29.83 1761 (CH_2) , 29.85 (3 × CH_2), 30.19 (CH_2) , 32.08 (CH_2) , 44.50 1762 (CH₂NH), 53.54 (CHCH₂Se), 55.94 (CH₃O), 110.72 (C_{Ar}), 1763 114.77 (C_{Ar}), 116.78 (CF₃COOH), 120.96 (C_{Ar}), 128.09 (C_{Ar}), 1764 145.43 (C_{Ar}), 146.96 (C_{Ar}), 160.81–162.0 (CF₃COOH), 167.72 1765 (NHCO). HR-MS (ESI⁺), m/z: [M + H]⁺ calcd for C₂₇H₄₉N₂O₃Se, 1766 529.2903; found 529.2905. 1767 g

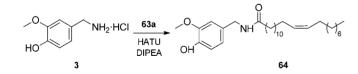


1-[*N*-(4'-Hydroxy-3'-methoxybenzyl)carbamoyl]-2-(octa-1768 dec-(9"*Z*)-en-1-ylseleno)-(1*R*)-ethylammonium Trifluoroace-1769 tate (61). General procedure IX was applied to compound 59 (200 1770 mg, 0.30 mmol), TFA (230 μ L, 3 mmol) in DCM (1 mL). 1771 Compound 61 was afforded after flushing nitrogen and drying in 1772 vacuo as a yellow oil (199 mg, quantitative). [α]_D²⁰ -2.58 (*c* 0.3, 1773 DCM). IR (ATR) ν = 2922, 2853, 1666, 1199 cm⁻¹. ¹H NMR (400 1774 1775 MHz, CDCl₃) $\delta = 0.87$ (t, 3H, J = 6.9 Hz, CH₃), 1.22–1.34 (m, 22H, 1776 CH₂), 1.51–1.61 (m, 2H, SeCH₂CH₂), 2.00 (q, 4H, J = 6.4 Hz, 1777 CH₂CH, CHCH₂), 2.54 (t, 2H, J = 6.9 Hz, SeCH₂CH₂), 2.93 (d, 2H, 1778 J = 6.4 Hz, CHCH₂Se), 3.81 (s, 3H, CH₃O), 4.17–4.34 (m, 3H, 1779 CHCH₂Se, CH₂NH), 5.28–5.42 (m, 2H, CH=CH), 6.72 (ddd, 3H, 1780 J = 12.5, 9.9, 5.0 Hz, H_A), 7.64 (t, 1H, J = 5.5 Hz, CH₂NH). ¹³C 1781 NMR (101 MHz, CDCl₃) $\delta = 14.26$ (CH₃), 22.83 (CH₂), 23.56 1782 (CHCH₂Se), 25.89 (SeCH₂CH₂), 27.37 (CH₂CH, CHCH₂), 29.24 1783 (CH₂), 29.42 (CH₂), 29.47 (CH₂), 29.47 (CH₂), 29.59 (CH₂), 29.68 1784 (CH₂), 29.87 (CH₂), 29.91 (CH₂), 29.92 (CH₂), 30.26 (CH₂), 32.06 1785 (CH₂), 44.17 (CH₂NH), 53.40 (CHCH₂Se), 56.00 (CH₃O), 110.73 1786 (C_{Ar}), 114.68 (C_{Ar}), 120.87 (C_{Ar}), 128.61 (C_{Ar}), 129.90 (CH=CH), 1787 130.12 (CH=CH), 145.38 (C_{Ar}), 146.93 (C_{Ar}), 167.76 (NHCO). 1788 HR-MS (ESI⁺), m/z: [M + H]⁺ calcd for C₂₉H₅₁N₂O₃Se, 555.3059; 1789 found 555.3067.

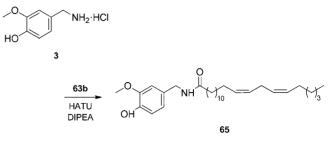
R-COOCH ₃	LiOH·H ₂ O	R-COOH	$R = \frac{\gamma_2}{\gamma_3} + \frac{\gamma_3}{\gamma_3} + \frac{\gamma_4}{\gamma_5} $ (a)
62a-b	-		R = 72 (b)

(13Z)-Docosenoic Acid (63a). General procedure II was applied 1790 1791 to a solution of methyl (13Z)-docosenoate 62a (500 μ L, 1.23 mmol) 1792 dissolved in THF/H₂O (6 mL, 1:1) and LiOH·H₂O (155 mg, 3.70 1793 mmol) to yield compound 63a as a white solid (360 mg, 86%). Mp: 1794 30-32 °C. IR (ATR) ν = 2916, 2849, 1691, 1471 cm⁻¹. ¹H NMR 1795 (400 MHz, CDCl₂) $\delta = 0.88$ (t, 3H, I = 6.9 Hz, CH₂), 1.17–1.39 (m, 1796 28H, CH_2), 1.58–1.70 (m, 2H, OHCOCH₂CH₂), 2.02 (q, 4H, J = 1797 6.4 Hz, CH₂CH, CHCH₂), 2.34 (t, 2H, J = 6.9 Hz, OHCOCH₂CH₂), 1798 5.24–5.42 (m, 2H, CH=CH). ¹³C NMR (101 MHz, CDCl₃) δ = 14.09 (CH₂), 22.67 (CH₂), 24.67 (OHCOCH₂CH₂), 27.20 (CH₂CH₂) 1799 1800 CHCH₂), 29.05 (CH₂), 29.23 (CH₂), 29.30 (CH₂), 29.31 (2 × CH₂), 1801 29.42 (CH₂), 29.51 (CH₂), 29.53 (CH₂), 29.57 (CH₂), 29.59 (CH₂), 1802 29.76 $(2 \times CH_2)$, 31.90 (CH_2) , 34.01 $(OHCOCH_2CH_2)$, 129.86 1803 (CH=CH), 129.89 (CH=CH), 179.89 (OHCOCH₂CH₂).

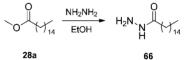
(13Z,16Z)-Docosadienoic Acid (63b). General procedure II was 1804 1805 applied to a solution of methyl (13Z,16Z)-docosadienoate 62b (25 1806 μ L, 0.07 mmol) in THF/H₂O (1 mL, 1:1) and LiOH·H₂O (9 mg, 1807 0.21 mmol) to yield compound 63b²⁰ as a sticky solid (23 mg, 1808 quantitative). IR (ATR) ν = 2922, 2853, 1708, 1458 cm⁻¹. ¹H NMR 1809 (400 MHz, CDCl₂) $\delta = 0.89$ (t, 3H, I = 6.9 Hz, CH₂), 1.17–1.45 (m, 1810 22H, CH_2), 1.53–1.72 (m, 2H, $COCH_2CH_2$), 2.05 (q, 4H, J = 6.41811 Hz, CH₂CH, CHCH₂), 2.34 (t, 2H, J = 6.9 Hz, COCH₂CH₂), 2.77 (t, 1812 2H, J = 6.9 Hz, CHCH₂CH), 5.24–5.44 (m, 4H, 2 × CH=CH). ¹³C 1813 NMR (101 MHz, CDCl₃) δ = 14.07 (CH₃), 22.58 (CH₂), 24.68 1814 (OHCOCH₂CH₂), 25.63 (CHCH₂CH), 27.20 (CH₂CH), 27.24 1815 (CHCH₂), 29.07 (CH₂), 29.24 (CH₂), 29.32 (CH₂), 29.36 (CH₂), 1816 29.43 (CH₂), 29.54 (CH₂), 29.58 (CH₂), 29.60 (CH₂), 29.68 (CH₂), 1817 31.53 (CH₂), 34.05 (OHCOCH₂CH₂), 127.94 (2 × CH=CH), 1818 130.17 (2 × CH=CH), 179.96 (OHCOCH₂CH₂).



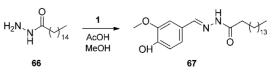
N-(4'-Hydroxy-3'-methoxybenzyl)docosa-(13Z)-enamide 1819 **N-(4'-Hydroxy-3'-methoxybenzyl)docosa-(13Z)-enamide** 1820 **(64).** General procedure I was applied to a solution of compound **63a** 1821 (200 mg, 0.59 mmol) in anhydrous DMF (5 mL), amine 1822 hydrochloride salt **3** (123 mg, 0.65 mmol), DIPEA (309 μL, 1.77 1823 mmol), and HATU (337 mg, 0.88 mmol). Compound **64** was 1824 afforded after silica gel column chromatography (petroleum ether/ 1825 EtOAc 6:4) as a sticky solid (179 mg, 64%). R_f = 0.42 (petroleum 1826 ether/EtOAc 5:5). IR (ATR) ν = 3489, 3315, 3304, 2918, 2849, 1827 1648, 1465 cm^{-1. 1}H NMR (400 MHz, CDCl₃) δ = 0.88 (t, 3H, *J* = 1828 6.9 Hz, CH₃), 1.23–1.36 (m, 28H, CH₂), 1.59–1.69 (m, 2H, 1829 COCH₂CH₂), 2.01 (q, 4H, *J* = 6.4 Hz, CH₂CH, CHCH₂), 2.19 (t, 1830 2H, *J* = 6.9 Hz, COCH₂CH₂), 3.87 (s, 3H, CH₃O), 4.34 (d, 2H, *J* = 1831 5.7 Hz, CH₂NH), 5.29–5.39 (m, 2H, CH=CH), 5.69 (s, 2H, OH, 1832 CH₂NH), 6.79 (ddd, 3H, *J* = 12.5, 9.9, 5.0 Hz, H_A). ¹³C NMR (101 MHz, CDCl₃) δ = 14.25 (CH₃), 22.82 (CH₂), 25.94 (COCH₂CH₂), 1833 27.35 (CH₂CH, CHCH₂), 29.46 (3 × CH₂), 29.50 (CH₂), 29.66 (2 × 1834 CH₂), 29.69 (CH₂), 29.75 (2 × CH₂), 29.83 (CH₂), 29.91 (CH₂), 1835 29.92 (CH₂), 32.04 (CH₂), 37.00 (COCH₂CH₂), 43.66 (CH₂NH), 1836 56.05 (CH₃O), 110.82 (C_{Ar}), 114.50 (C_{Ar}), 120.92 (C_{Ar}), 130.00 1837 (CH=CH), 130.04 (CH=CH), 130.51 (C_{Ar}), 145.26 (C_{Ar}), 146.83 1838 (C_{Ar}), 173.04 (COCH₂CH₂). HR-MS (ESI⁺), *m*/*z*: [M + Na]⁺ calcd 1839 for C₃₀H₅₁NO₃Na, 496.3767; found 496.3756. 1840 g



N-(4'-Hydroxy-3'-methoxybenzyl)docosa-(13Z,16Z)-diena- 1841 mide (65). General procedure I was applied to a solution of 1842 compound 63b (23 mg, 0.07 mmol) dissolved in DMF (1 mL), amine 1843 hydrochloride salt 3 (15 mg, 0.08 mmol), DIPEA (38 µL, 0.21 1844 mmol), and HATU (39 mg, 0.10 mmol). Compound 65 was afforded 1845 after silica gel column chromatography (petroleum ether/EtOAc 6:4) 1846 as a sticky oil (21 mg, 63%). $R_f = 0.40$ (petroleum ether/EtOAc 5:5). 1847 IR (ATR) $\nu = 3489$, 3316, 3302, 2919, 2849, 1639, 1518 cm⁻¹. ¹H 1848 NMR (400 MHz, CDCl₃) δ = 0.89 (t, 3H, J = 6.9 Hz, CH₃), 1.24– 1849 1.38 (m, 22H, CH₂), 1.59-1.70 (m, 2H, COCH₂CH₂), 2.05 (q, 4H, J 1850 = 6.4 Hz, CH_2CH , $CHCH_2$), 2.19 (t, 2H, J = 6.9 Hz, $COCH_2CH_2$), 1851 2.77 (t, 2H, J = 6.9 Hz, CHCH₂CH), 3.87 (s, 3H, CH₃O), 4.35 (d, 1852 2H, J = 5.7 Hz, CH_2NH), 5.28–5.43 (m, 4H, 2 × CH=CH), 5.59–1853 5.72 (m, 2H, OH, CH₂NH), 6.79 (ddd, 3H, J = 12.5, 9.9, 5.0 Hz, 1854 H_{Ar}). ¹³C NMR (101 MHz, CDCl₃) $\delta = 14.22$ (CH₃), 22.72 (CH₂), 1855 25.78 (CHCH₂CH), 25.94 (COCH₂CH₂), 27.35 (CH₂CH), 27.39 1856 (CHCH₂), 29.48 (2 × CH₂), 29.50 (2 × CH₂), 29.65 (CH₂), 29.70 1857 (CH_2) , 29.75 (2 × CH_2), 29.83 (CH_2) , 31.68 (CH_2) , 37.03 1858 (COCH₂CH₂), 43.68 (CH₂NH), 56.08 (CH₃O), 110.82 (C_{Ar}), 1859 114.49 (C_{Ar}), 120.95 (C_{Ar}), 128.09 (2 × CH=CH), 130.31 1860 (CH=CH), 130.34 (CH=CH), 130.53 (C_{Ar}), 145.26 (C_{Ar}), 1861 146.82 (C_{Ar}), 173.05 (COCH₂CH₂). HR-MS (ESI⁺), m/z: [M + 1862 Na]⁺ calcd for C₃₀H₄₉NO₃Na, 494.3610; found 494.3606. 1863 g



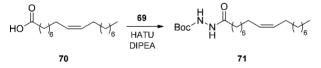
Hexadecanohydrazide (66). To a suspension of methyl 1864 palmitate 28a (1 g, 3.69 mmol) in ethanol (20 mL), hydrazyne 1865 hydrate (64%, 370 µL, 7.38 mmol, 2 equiv) was added. Then, the 1866 mixture was heated at 150 °C for 3 h. The mixture was cooled, and 1867 the solid precipitated was recovered by filtration to yield compound 1868 66^{21} as a white solid (800 mg, 80%). Mp: 110–111 °C. IR (ATR) ν = 1869 3315, 3288, 3199, 2956, 2917, 2848, 1627, 1535 cm⁻¹. ¹H NMR (400 1870 MHz, CDCl₃) $\delta = 0.88$ (t, 3H, J = 6.9 Hz, CH₃), 1.06–1.42 (m, 24H, 1871 CH₂), 1.55-1.74 (m 2H, NHCOCH₂CH₂), 2.08-2.23 (m, 2H, 1872 NHCOCH₂CH₂), 3.89 (br s, 2H, NH₂NH), 6.66 (s, 1H, NH₂NH). 1873 ¹³C NMR (101 MHz, CDCl₃) δ = 14.10 (CH₃), 22.67 (CH₂), 25.48 ₁₈₇₄ (NHCOCH₂CH₂), 29.25 (CH₂), 29.27 (CH₂), 29.34 (CH₂), 29.44 1875 (CH₂), 29.57 (CH₂), 29.62 (CH₂), 29.63 (CH₂), 29.64 (CH₂), 29.66 1876 (CH₂), 29.67 (CH₂), 31.90 (CH₂), 34.59 (NHCOCH₂CH₂), 173.97 1877 $(NHCOCH_2).$ 1878 g



N'-(4'-Hydroxy-3'-methoxybenzylidene)hexadecano-1879 1880 hydrazide (67). General procedure X was applied to compound 66 1881 (280 mg, 1.03 mmol), vanillin 1 (157 mg, 1.03 mmol), AcOH (60 μL, 1882 1.03 mmol) in MeOH (30 mL). Compound 67 was afforded as a 1883 white solid (242 mg, 58%) after recrystallization from hot MeOH. 1884 The ¹H NMR analysis confirmed the presence of the *cis* isomer of the 1885 imine as the minor product. Mp: 109–110 °C. IR (ATR) ν = 3202, 1886 3054, 2917, 2849, 1659, 1510 cm⁻¹. Trans isomer: ¹H NMR (400 1887 MHz, CDCl₃) $\delta = 0.88$ (t, 3H, J = 6.9 Hz, CH₃), 1.23–1.42 (m, 24H, 1888 CH₂), 1.69–1.78 (m, 2H, NHCOCH₂CH₂), 2.74 (t, 2H, J = 6.9 Hz, 1889 NHCOCH2CH2), 3.95 (s, 3H, CH3O), 5.86 (s, 1H, OH), 6.93 (d, 1890 1H, J = 8.2 Hz, H_{Ar}), 7.09 (dd, 1H, J = 8.2, 1.8 Hz, H_{Ar}), 7.25 (d, 1H, 1891 J = 1.8 Hz, H_{Ar}), 7.65 (s, 1H, HC=NNH), 9.02 (s, 1H, NHCO). Cis 1892 isomer: ¹H NMR (400 MHz, CDCl₃) δ = 2.28 (t, 2H, J = 6.9 Hz, 1893 NHCOCH₂CH₂), 3.94 (s, 1H, CH₃OH), 5.91 (br s, 1H, OH), 6.89 1894 (d, 1H, J = 8.2 Hz, H_{Ar}), 6.98 (dd, 1H, J = 8.2, 1.8 Hz, H_{Ar}), 7.49 (br 1895 s, 1H, H_{Ar}), 8.00 (s, 1H, HC=NNH), 8.46 (s, 1H, NHCO). The rest 1896 of signals are common to trans isomer. Trans isomer: ¹³C NMR (101 1897 MHz, CDCl₃) δ = 14.27 (CH₃), 22.85 (NHCOCH₂CH₂), 24.97 1898 (CH₂), 29.51 (CH₂), 29.59 (CH₂), 29.64 (CH₂), 29.72 (CH₂), 29.81 1899 $(2 \times CH_2)$, 29.85 $(4 \times CH_2)$, 32.08 (CH_2) , 32.96 $(NHCOCH_2CH_2)$, 1900 56.09 (CH₃O), 107.97 (C_{Ar}), 114.61 (C_{Ar}), 122.37 (C_{Ar}), 126.49 1901 (C_{Ar}), 143.20 (HC=NNH), 147.07 (C_{Ar}), 147.90 (C_{Ar}), 176.00 1902 (NHCO). Cis isomer: ¹³C NMR (101 MHz, CDCl₃) δ = 56.38 1903 (CH₃O), 107.86 (C_{Ar}), 114.13 (C_{Ar}), 123.80 (C_{Ar}), 126.20 (C_{Ar}). 1904 The rest of signals are common to *trans* isomer. HR-MS (ESI⁺), m/z: 1905 $[M + Na]^+$ calcd for $C_{48}H_{80}N_4O_6Na$, 831.5976; found 831.5968.

$$H_2N-NH_2 \xrightarrow{Boc_2O} H_2N_N \xrightarrow{N} Boc$$
68 69

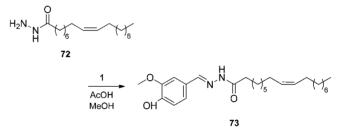
1906 **tert-Butyl Hydrazinecarboxylate (69).** Hydrazyne hydrate 68 1907 (64%, 1.52 mL, 31.2 mmol) was mixed with isopropanol (3 mL) at 0 1908 °C. Then, a solution of Boc₂O (6.8 g, 31.2 mmol, 1 equiv) in 1909 isopropanol (6 mL) was added dropwise. The reaction mixture turned 1910 cloudy upon addition and was stirred at room temperature for 2 h. 1911 The solvent was removed under reduced pressure and the residue was 1912 dissolved in DCM, washed with 1 M HCl and brine. The organic 1913 phase was dried over Na₂SO₄, and the solvent was removed under 1914 reduced pressure. The residue was recrystallized from hexane to yield 1915 compound **69**²² as a white solid (1.94 g, 47%). Mp: 38–40 °C. IR 1916 (ATR) ν = 3374, 3324, 2981, 1692, 1627, 1502 cm⁻¹. ¹H NMR (400 1917 MHz, CDCl₃) δ = 1.44 (s, 9H, C(CH₃)₃), 3.57 (s, 2H, NH₂) 6.00 (s, 1918 1H, NHCO). ¹³C NMR (101 MHz, CDCl₃) δ = 28.28 (C(CH₃)₃), 1919 80.42 (C(CH₃)₃), 158.22 (COO).



N'-(tert-Butyloxycarbonyl)octadec-(9Z)-enohydrazide (70). 1920 1921 General procedure I was applied to a solution of oleic acid 70 (1 g 1922 3.54 mmol) dissolved in DMF (30 mL), compound 69 (524 mg, 3.96 1923 mmol), DIPEA (1.85 mL, 10.62 mmol), and HATU (2.02 g, 5.31 1924 mmol). Compound 71^{23} was afforded after silica gel column 1925 chromatography (petroleum ether/EtOAc 7:3) as a yellow oil (1.32 1926 g, 94%). $R_f = 0.47$ (petroleum ether/EtOAc 6:4). IR (ATR) $\nu = 3280$, 1927 2924, 2854, 1729, 1673, 1242 cm⁻¹. ¹H NMR (400 MHz, CDCl₃) $\delta =$ 1928 0.86 (t, 3H, J = 6.9 Hz, CH_3), 1.16–1.40 (m, 20H, CH_2) 1.44 (s, 9H, 1929 C(CH₃)₃), 1.57-1.74 (m, 2H, NHCOCH₂CH₂), 1.90-2.07 (m, 4H, 1930 CH₂CH, CHCH₂), 2.11–2.28 (m, 2H, NHCOCH₂CH₂), 5.22–5.43 1931 (m, 2H, CH=CH), 6.85 (s, 1H, NHNH), 8.06 (s, 1H, NHNH). ¹³C 1932 NMR (101 MHz, CDCl₃) δ = 14.07 (CH₃), 22.64 (CH₂), 25.25 1933 (NHCOCH₂CH₂), 27.14 (CH₂CH), 27.18 (CHCH₂), 28.11 (C-1934 (CH₃)₃), 29.08 (CH₂), 29.17 (CH₂), 29.19 (CH₂), 29.27 (CH₂), 1935 29.29 (CH₂), 29.48 (CH₂), 29.67 (CH₂), 29.72 (CH₂), 31.86 (CH₂), 1936 33.97 (NHCOCH₂CH₂), 81.66 ($C(CH_3)_3$), 129.68 (CH=CH), 1937 129.93 (CH=CH), 155.85 (COC(CH₃)₃), 172.80 (NHCOCH₂).

$$H_2N$$
, N , H_{6}

Oleylhydrazine (72). To a solution of compound 71 (1 g, 2.52 1938 mmol) in DCM (3 mL), TFA (1.93 mL, 25.2 mmol, 10 equiv) was 1939 added. The mixture was stirred for 2 h at room temperature. Then, 1940 the solvent was partially evaporated. Water was added, and the pH 1941 was adjusted to 7 with saturated solution of NaHCO₃. The aqueous 1942 phase was extracted with DCM, and the organic solution was dried 1943 over Na2SO4 and filtered. The solvent was removed under reduced 1944 pressure to yield the compound 72 as a yellow solid (687 mg, 92%). 1945 Mp: 109–110 °C. IR (ATR) ν = 3316, 3214, 2919, 2849, 1628, 1596 1946 cm^{-1} . ¹H NMR (400 MHz, CDCl₃) $\delta = 0.87$ (t, 3H, J = 6.9 Hz, CH₃), 1947 1.12-1.42 (m, 20H, CH₂) 1.53-1.74 (m, 2H, NHCOCH₂CH₂), 1948 1.88-2.05 (m, 4H, CH₂CH, CHCH₂), 2.08-2.24 (m, 2H, 1949 NHCOCH₂CH₂), 3.97 (s, 2H, H₂N), 5.20–5.43 (m, 2H, CH= 1950 CH), 6.84 (s, 1H, NH). ¹³C NMR (101 MHz, CDCl₃) δ = 14.08 1951 (CH₃), 22.65 (CH₂), 25.46 (NHCOCH₂CH₂), 27.13 (CH₂CH), 1952 27.19 (CHCH₂), 29.07 (CH₂), 29.18 (CH₂), 29.22 (CH₂), 29.29 (2 × 1953 CH₂), 29.49 (CH₂), 29.66 (CH₂), 29.73 (CH₂), 31.87 (CH₂), 34.55 1954 (NHCOCH₂CH₂), 129.67 (CH=CH), 129.99 (CH=CH), 173.98 1955 (NHCOCH₂). 1956 g



N'-(4'-Hydroxy-3'-methoxybenzylidene)octadec-(9Z)-eno- 1957 hydrazide (73). General procedure X was applied to compound 72 1958 (300 mg, 1.01 mmol), vanillin 1 (153 mg, 1.01 mmol), AcOH (60 μL, 1959 1.01 mmol) in MeOH (30 mL). Compound 73 was afforded after 1960 silica gel column chromatography (petroleum ether/EtOAc 6:4) as a 1961 colorless oil (1.32 g, 94%). The ¹H NMR analysis confirmed the 1962 presence of the cis isomer of the imine as a minor product. IR (ATR) 1963 ν = 3452, 3194, 2921, 2852, 1650, 1211 cm⁻¹. Trans isomer: ¹H 1964 NMR (400 MHz, CDCl₃) $\delta = 0.87$ (t, 3H, J = 6.9 Hz, CH₃), 1.22- 1965 1.43 (m, 20H, CH₂), 1.69-1.78 (m, 2H, NHCOCH₂CH₂), 1.94- 1966 2.07 (m, 4H, CH₂CH, CHCH₂), 2.74 (t, 2H, J = 6.9 Hz, 1967 NHCOCH₂CH₂), 3.95 (s, 3H, CH₃O), 5.31-5.36 (m, 2H, CH= 1968 CH), 5.93 (br s, 1H, OH), 6.93 (d, 1H, J = 8.2 Hz, H_{Ar}), 7.10 (dd, 1969 1H, J = 8.2, 1.8 Hz, H_{Ar}), 7.25 (d, 1H, J = 1.8 Hz, H_{Ar}), 7.69 (s, 1H, 1970 HC=NNH), 9.43 (s, 1H, NHCO). Cis isomer: ¹H NMR (400 MHz, 1971 $CDCl_3$) $\delta = 2.28$ (t, 2H, J = 6.9 Hz, $NHCOCH_2CH_2$), 3.93 (s, 1H, 1972) CH₃OH), 5.36-5.39 (m, 2H, CH=CH), 5.97 (br s, 1H, OH), 6.89 1973 (d, 1H, J = 8.2 Hz, H_{Ar}), 6.97 (dd, 1H, J = 8.2, 1.8 Hz, H_{Ar}), 7.49 (d, 1974 1H, J = 1.8 Hz, H_{Ar}), 8.00 (s, 1H, HC=NNH), 8.62 (s, 1H, NHCO). 1975 The rest of signals are common to trans isomer. Trans isomer: ¹³C 1976 NMR (101 MHz, CDCl₃) δ = 14.26 (CH₃), 22.82 (NHCOCH₂CH₂), 1977 25.00 (CH₂), 27.34 (CH₂CH), 27.36 (CHCH₂), 29.35 (CH₂), 29.46 1978 (CH₂), 29.46 (CH₂), 29.49 (CH₂), 29.61 (CH₂), 29.66 (CH₂), 29.84 1979 (CH₂), 29.91 (CH₂), 32.04 (CH₂), 32.94 (NHCOCH₂CH₂), 56.08 1980 $(CH_{3}O)$, 108.06 (C_{Ar}) , 114.63 (C_{Ar}) , 122.32 (C_{Ar}) , 126.54 (C_{Ar}) , 1981 129.88 (CH=CH), 130.13 (CH=CH), 143.54 (HC=NNH), 1982 147.06 (C_{Ar}), 147.89 (C_{Ar}), 176.30 (NHCO). Cis isomer: ¹³C NMR 1983 $(101 \text{ MHz}, \text{CDCl}_3) \delta = 56.35 (CH_3O), 107.87 (C_{Ar}), 114.11 (C_{Ar}), 1984$ 123.79 (C_{Ar}), 126.16 (C_{Ar}), 147.24 (C_{Ar}), 147.73 (C_{Ar}). The rest of 1985 signals are common to trans isomer. HR-MS (ESI⁺), m/z: [M + Na]⁺ 1986 calcd for C52H84N4O6Na, 883.6289; found 883.6286. 1987

5.3. TRP Channels Assays. Assays of TRP-mediated elevation of 1988 $[Ca^{2+}]_i$ were performed as previously described.⁶⁰ HEK-293 (human 1989 embryonic kidney) cells wild-type or stably overexpressing recombi-1990 nant human TRPV1 or rat TRPV2 were grown on 100 mm diameter 1991

1992 Petri dishes as monolayers in Eagle's minimum essential medium 1993 (EMEM) supplemented with 1% nonessential amino acids, 10% fetal 1994 bovine serum (FBS), 50 U/mL penicillin plus 50 µg/mL 1995 streptomycin, and 2 mM glutamine, maintained under 5% CO2 at 1996 37 °C and only for the overexpressing cells selected by G-418 1997 (Geneticin, 600 mg mL⁻¹; Thermo-Fisher Scientific). On the day of 1998 the experiment, the cells were loaded for 1 h at 25 °C with the Ca²⁺ 1999 indicator Fluo-4-AM (Thermo-Fisher Scientific) 4 μ M in DMSO 2000 containing 0.02% Pluronic F-127 (Thermo-Fisher Scientific) in 2001 EMEM without FBS. After loading, cells were washed twice in 2002 Tyrode's buffer (145 mM NaCl, 2.5 mM KCl, 1.5 mM CaCl₂, 1.2 mM 2003 MgCl₂, 10 mM D-glucose, and 10 mM HEPES, pH 7.4), resuspended 2004 in the same buffer, and transferred, about 100 000 cells for each 2005 determination, to the quartz cuvette of the spectrofluorimeter (λ_{ex} = 2006 488 nm; λ_{em} = 516 nm) PerkinElmer LS50B equipped with PTP-1 2007 fluorescence Peltier system (PerkinElmer Life and Analytical Sciences, 2008 Waltham, MA, USA) under continuous stirring at 25 °C. Experiments 2009 were carried by measuring cell fluorescence before and after the 2010 addition of test compounds at various concentrations. The values of 2011 the effect on $[Ca^{2+}]_i$ in wild-type (i.e., not transfected with any TRP 2012 construct) HEK-293 cells were taken as baselines. Potency (EC50 2013 values) was determined as the concentration of test compounds 2014 exerting a half-maximal agonist effect (i.e., half-maximal increases in 2015 $[Ca^{2+}]_i$). The efficacy of the agonists was determined by comparing 2016 their effect to the maximal effect on $[Ca^{2+}]_i$ observed with 4 μM 2017 ionomycin. Antagonist/desensitizing behavior was evaluated against 2018 the agonist capsaicin 0.1 μ M (Sigma-Aldrich) for TRPV1 and the 2019 agonists lysophosphatidylcholine (LPC) (Sigma-Aldrich) 3 μ M and 2020 cannabidiol (CBD) 2 μ M (a kind gift by GW Pharmaceuticals) for 2021 TRPV2 by adding the test compounds in the quartz cuvette 5 min 2022 before stimulation of cells with the agonist. The effect on $[Ca^{2+}]_{i}$ 2023 exerted by agonist alone was taken as 100%. Data are expressed as the 2024 concentration exerting a half-maximal inhibition of agonist-induced 2025 $[Ca^{2+}]_i$ elevation (IC₅₀). Concentration-response curves were fitted 2026 by a sigmoidal regression with variable slope. Curve fitting and 2027 parameter estimation were performed with GraphPad Prism (Graph-2028 Pad Software Inc., San Diego, CA). Determinations were performed 2029 at least in triplicate. Statistical analysis of the data was performed by 2030 analysis of variance at each point using ANOVA followed by 2031 Bonferroni's test.

2032 ASSOCIATED CONTENT

2033 **Supporting Information**

2034 The Supporting Information is available free of charge on the 2035 ACS Publications website at DOI: 10.1021/acs.jmed-2036 chem.8b00734.

Tables S1 and S2 of TRPV1 activity and ¹H and ¹³C

- 2038 NMR spectra (PDF)
- 2039 Molecular formula strings and some data (CSV)

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2051 Notes

2052 The authors declare no competing financial interest.

A.S.M. is an employee of Epitech Group SpA. V.D.M. provides 2053 consultancy services and performs sponsored research for GW 2054 Research Ltd. 2055

ACKNOWLEDGMENTS

We gratefully acknowledge financial support from Universitat 2057 de Lleida, Ministerio de Educación, Cultura y Deporte and 2058 Banco Santander (Programa UdL-Impuls). The authors are 2059 grateful to the Serveis Cientifictècnics (SCT) of the Universitat 2060 de Lleida for providing us with spectroscopic and chromato- 2061 graphic facilities. We acknowledge Dr. Alberto Minassi, 2062 Dipartimento di Scienze del Farmaco, Università del Piemonte 2063 Orientale, Novara, Italy, for the kind gift of olvanil. 2064

ABBREVIATIONS USED

2065

2.056

TRPV2, transient receptor potential vanilloid 2; TRPV1, 2066 transient receptor potential vanilloid 1; EA, ethanolamide; 2067 LPC, lysophosphatidylcoline; CBD, cannabidiol; PEA, palmi- 2068 toyl ethanolamide; POEA, palmitoleoyl ethanolamide; OEA, 2069 oleoyl ethanolamide; LEA, lynoleoyl ethanolamide; AEA, 2070 arachidonoylethanolamide; EPEA, eicosapentaenoyl ethanola- 2071 mide; DHEA, docosahexaenoyl ethanolamide; PA, palmita- 2072 mide; SA, stearamide; OA, oleamide; LA, linoleamide; ErA, 2073 erucamide 2074

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