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EMPress enables tree-guided, interactive, and exploratory analyses of multi-omic datasets — Source link 🖸

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EMPress enables tree-guided, interactive, and exploratory analyses of multi-omic datasets

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- 32 Abstract
- 33 Standard workflows for analyzing microbiomes often include the creation and curation of
- 34 phylogenetic trees. Here we present EMPress, an interactive tool for visualizing trees in the
- 35 context of microbiome, metabolome, etc. community data scalable beyond modern large
- 36 datasets like the Earth Microbiome Project. EMPress provides novel functionality—including
- 37 ordination integration and animations—alongside many standard tree visualization features, and
- 38 thus simplifies exploratory analyses of many forms of 'omic data.
- 39

40

41 Main Text

42

43 The increased availability of sequencing technologies and automation of molecular methods 44 have enabled studies of unprecedented scale [1] prompting the creation of tools better suited to 45 store, analyze [2], and visualize [3] studies of this magnitude. Many of these tools, such as [4, 5, 46 6, 7], use phylogenies detailing the evolutionary relationships among features or dendrograms 47 that organize features in a hierarchical structure (e.g. clustering of mass spectra) [8]. The 48 challenge of enabling fully interactive analyses stems from the disconnect between feature-level 49 tools and dataset-level tools; few can interactively integrate multiple representations of the data 50 [9], and to our knowledge none scale to display large datasets. This is a key unresolved 51 challenge for the field: to allow researchers to contextualize community-level patterns 52 (groupings of samples) together with feature-level structure, i.e. which features lead to the 53 groupings explained in a given sample set. 54 55 Here, we introduce EMPress (https://github.com/biocore/empress), an open-source (BSD 3-56 clause), interactive and scalable phylogenetic tree viewer accessible as a QIIME 2 [2] plugin. 57 EMPress is built around the high-performance balanced parentheses tree data structure [10]. 58 and uses a hardware-accelerated WebGL-based rendering engine that allows EMPress to 59 visualize trees with hundreds of thousands of nodes using a laptop's web browser (Methods). 60 By integrating EMPress with the widely-used EMPeror software [3] within QIIME 2. EMPress

- 61 can simultaneously visualize a phylogenetic tree of features in a study coupled with an
- 62 ordination of the same study's samples. User actions in one visualization, such as selecting a
- set of samples in the ordination, update the other, providing context that would not be easily
 accessible with independent visualizations. This tight integration between displays streamlines
- 64 accessible with independent visualizations. This tight integration between displays streamlines 65 several use-cases elaborated below that previously required manual investigation or writing
- 66 custom scripts.
- 67

68 EMPress visualizations can be created solely from a tree, or users can provide additional

- 69 metadata files and a feature table to augment the tree. Using these common data files, users
- can interactively configure many visual attributes in the tree (see Methods and Figures for
- 71 examples).
- 72
- 73 Rather than providing a programmatic interface for the procedural generation of styled
- 74 phylogenetic trees [11, 12, FigTree (http://tree.bio.ed.ac.uk/software/figtree/)], EMPress
- 75 provides an interactive environment to support exploratory feature- and sample-level tree-based
- analyses. Many use-cases supported in EMPress accommodate community analysis tasks; this
- differs from Anvi'o [13] which is centered on the analysis of metagenomic assembled-genomes,
- pangenomes, etc.. PHYLOViZ [9], SigTree [14], and iTOL [15] are similar to EMPress in terms
- of their implementation (PHYLOViZ Online also uses WebGL), and/or use-cases (SigTree is
- 80 mostly used to visualize differential abundance patterns, and iTOL supports the visualization of
- 81 QIIME 2 tree artifacts). EMPress stands out in its scalability: iTOL claims trees with more than

82 10.000 tips to be "very large" (https://itol.embl.de/help.cgi), while EMPress readily supports trees 83 with over hundreds of thousands of tips, as shown in Fig. 1. Many visualization customization 84 options available in EMPeror, iTOL [15] and Anvi'o [13] are immediately accessible in EMPress' 85 interface. Continuous feature metadata can be visualized in tip-level barplots as a color gradient 86 and/or by adjusting the lengths of individual tips' barplots; categorical sample metadata 87 information can be visualized using a stacked barplot showing-for each tip-the proportion of 88 samples containing that tip stratified by category. These options are available on the user 89 interface and do not require programming or configuration files.

90

91 Ordination plots computed from UniFrac distances are often used to visualize sample clustering

92 patterns in microbiome studies. However, interpreting the patterns in these plots—and

93 determining which features influence sample group separation—is not always straightforward.

94 While biplots show information about influential features alongside samples, the phylogenetic

95 relationships of these features are not immediately obvious. EMPress aids interpretation of

96 these plots by optionally providing a unified interface where the tree and ordination

97 visualizations are displayed side-by-side and "linked" through sample and feature identifiers

98 [16]. This combination allows for novel exploratory data analysis tasks. For example, selecting a

99 group of samples in the ordination highlights nodes in the tree present in those samples, and

100 vice versa (see Methods). This integration extends to biplots: clicking feature arrows in the

101 ordination highlights their placement in the tree. Lastly, EMPress allows visualizing longitudinal

102 studies by simultaneously showing the tree nodes unique to groups of samples at each

103 individual time point during an EMPeror animation (see Methods).

104

105 Using the first data release of the Earth Microbiome Project (EMP), we demonstrate EMPress' 106 scalability by rendering a 26,035 sample ordination and a 756,377 node tree (Figure 1A). To 107 visualize the relative proportions of taxonomic groups at the phylum level, we use EMPress' 108 feature metadata coloring to highlight the top 5 most prevalent phyla (see Methods). Next, we 109 add a barplot layer showing, for each tip in the tree, the proportions of samples containing each 110 tip summarized by level 2 of the EMP ontology (Animal, Plant, Non-Saline, and Saline), Paired 111 visualizations allow us to click on a tip in the tree and view the samples that contain that feature 112 in the ordination. This functionality is useful when analyzing datasets with outliers or mislabeled 113 metadata. Tip-aligned barplots summarize environmental metadata: for example, Figure 1B 114 shows the subset of samples (4,002) with recorded pH information and a barplot layer with the 115 mean pH where each feature was found. The barplot reveals a relatively dark section near many Firmicutes-classified features on the tree; in concert with histograms showing mean pH 116 117 for each phylum (Figure 1C), we can confirm that Firmicutes-classified features are more 118 commonly found in higher pH environments. 119

120 EMPress can be applied to various 'omic datasets. To illustrate this versatility we reanalyzed a

121 COVID-19 metatranscriptome sequencing dataset [17], a liquid chromatography mass-

122 spectrometry (LC-MS) untargeted metabolomic food-associated dataset [8], and a 16S rRNA

sequencing oral microbiome dataset [18]. Despite the vastly different natures of these datasets,

124 EMPress provides meaningful functionality for their analysis and visualization. Supplemental

125 Video 1 (supplementary-video-1.mp4) shows a longitudinal exploratory analysis using EMPress

and EMPeror representing a subset of SARS-CoV-2 genome data from GISAID. This paired
 visualization emphasizes the relationships in time and space among "community samples" and

- the convergence of locales in the United States with the outbreak in Italy (See Methods). The
- 129 interactive nature of EMPress allows rapid visualization of strains observed in a collection of
- 130 samples from different geographical locations.
- 131
- 132Figure 2A showcases Empress' ability to identify feature clusters that are differentially abundant
- in COVID-19 patients compared to community-acquired pneumonia patients and healthy
- 134 controls [17]. Clades showing KEGG enzyme code (EC) [19] annotations are collapsed at level
- 135 two except for lyases, highlighting feature 4.1.1.20 (carboxy-lyase diaminopimelate
- decarboxylase) that was more abundant in COVID-19 here and in an independent
- 137 metaproteomic analysis of COVID-19 respiratory microbiomes [20].
- 138

139 Recent developments in cheminformatics enabled the analysis and visualization of small 140 molecules in the context of a cladogram [8]. Using a tree that links molecules by their structural 141 relatedness, we analyzed untargeted LC-MS/MS data from 70 food samples (see Methods). 142 With EMPress' sample metadata barplots, we can inspect the relationship between chemical 143 annotations and food types. Figure 2B shows a tree where each tip is colored by its chemical 144 super class, and where barplots show the proportion of samples in the study containing each 145 compound by food type. This representation reveals a clade of lipids and lipid-like molecules 146 that are well represented in animal food types and seafoods. In contrast, salads and fruits are 147 broadly spread throughout the cladogram.

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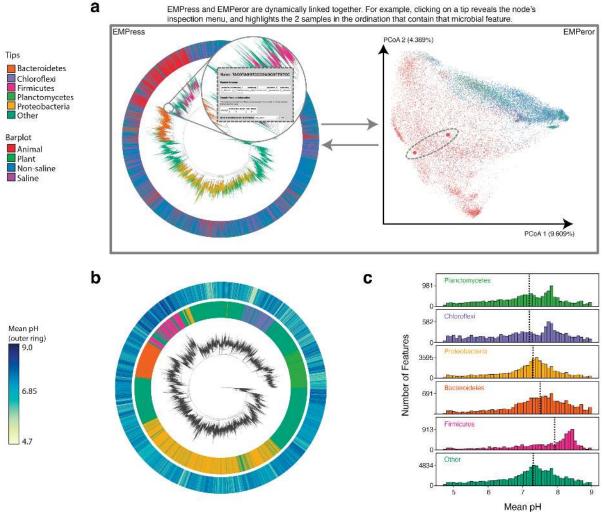
Lastly, in Figure 2C, we compare three differential abundance methods in an oral microbiome dataset [18] as separate barplot layers on a tree. This dataset includes samples (n=32) taken before and after subjects brushed their teeth (see Methods). As observed across the three differential abundance tools' outputs, all methods agree broadly on which features are particularly "differential" (for example, the cluster of Firmicutes-classified sequences in the bottom-right of the tree; see Methods), although there are discrepancies due to different methods' assumptions and biases.

- 156 Conclusions
- 157 By providing an intuitive interface supporting both categorically new and established
- 158 functionality, EMPress complements and extends the available range of tree visualization
- 159 software. EMPress can perform community analyses across distinct "omics" types, as
- 160 demonstrated here. Moving forward, facilitating the integration of multiple orthogonal views of a
- 161 dataset at a more generalized framework level (for example, using QIIME2's [2] visualization
- API) will be important as datasets continue to grow in complexity, size, and heterogeneity.

163 Acknowledgements

- 164 We thank members of the Knight Lab and IBM AIHL Bioinformatics team for feedback during
- 165 code reviews and presentations. We gratefully acknowledge the following Authors from the
- 166 originating laboratories responsible for obtaining the specimens and the submitting laboratories

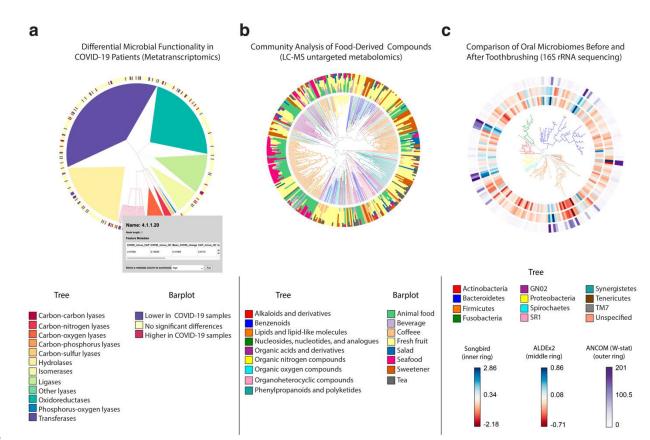
- 167 where genetic sequence data were generated and shared via the GISAID Initiative, on which a
- 168 portion of this research is based (Supplemental Table 1).
- 169
- 170 Funding
- 171 This work is partly supported by IBM Research AI through the AI Horizons Network and the UC
- 172 San Diego Center for Microbiome Innovation (to KC, MWF, JM, QZ, TZ, JS, ADS, SS, YVB,
- 173 RK); CCF foundation #675191 (RK and PCD), U19 AG063744 01 (RK, JMG, PCD), CDC
- 174 contract #75D30120C09795 (KGA and RK), U19 AI135995 (KGA).
- 175 Author Contributions
- 176 KC, QZ, YY, JM, TZ, JS, and RK conceived the original idea for the project. KC, MWF, GR, DM,
- 177 AG, SJ, ME, YY, ES, JM, TZ, QZ, YVB wrote source code and/or documentation for the project.
- 178 KC, MWF, AG, YVB wrote code to facilitate integration with EMPeror. LP, HCK, SS, ADS, YVB,
- 179 RK managed the project. KC, MWF, GR, NH, KLB, AT, JMG, LM, APC, NLM, CM, PCD, KGA,
- 180 LP, YVB analyzed and interpreted the datasets presented in this paper. KC, MWF, GR, DM,
- 181 NH, KLB, YVB contributed text to the methods section. All the authors contributed to the final
- 182 version of the manuscript.
- 183 Competing Interests
- 184 We declare none.





186 Figure 1. Earth Microbiome Project paired phylogenetic tree (including 756,377 nodes) and 187 unweighted UniFrac ordination (including 26,035 samples). (a) Graphical depiction of Empress' 188 unified interface with fragment insertion tree (left), and unweighted UniFrac sample ordination (right). Tips 189 are colored by their phylum-level taxonomic assignment; the barplot layer is a stacked barplot describing 190 the proportions of samples containing each tip summarized by level 2 of the EMP ontology. Inset shows 191 summarized sample information for a selected feature. The ordination highlights the two samples 192 containing the tip selected in the tree enlarged to show their location. (b) Subset of EMP samples with pH 193 information: the inner barplot ring shows the phylum-level taxonomic assignment, and the outer barplot 194 ring represents the mean pH of all the samples where each tip was observed (c) pH distributions 195 summarized by phylum-level assignment with median pH indicated by dotted lines. Interactive figures can 196 be accessed here.

197



198

199 Figure 2. EMPress is a versatile exploratory analysis tool adaptable to various -omics data types. 200 (a) RoDEO differential abundance scores of microbial functions from metatranscriptomic sequencing of 201 COVID-19 patients (n=8), community-acquired pneumonia patients (n=25), and healthy control subjects 202 (n=20). The tree represents the four-level hierarchy of the KEGG enzyme code. The barplot colors 203 significantly differentially abundant features (p<0.05) in COVID-19 patients. Clicking on a tip produces a 204 pop-up insert tabulating the name of the feature, its hierarchical ranks, and any feature annotations. 205 (b) Global FoodOmics Project LC-MS data. Stacked barplots indicate the proportions of samples (n=70) 206 (stratified by food) containing the tips in an LC-MS Qemistree of food-associated compounds, with tip 207 nodes colored by their chemical superclass. 208 (c) de novo tree constructed from 16S rRNA sequencing data from 32 oral microbiome samples. Samples 209 were taken before (n=16) and after (n=16) subjects (n=10) brushed their teeth; each barplot layer 210 represents a different differential abundance method's measure of change between before- and after-211 brushing samples. The innermost layer shows estimated log-fold changes produced by Songbird; the 212 middle layer shows effect sizes produced by ALDEx2; and the outermost layer shows the W-statistic 213 values produced by ANCOM (see Methods). The tree is colored by tip nodes' phylum-level taxonomic 214 classifications. Interactive figures can be accessed here.

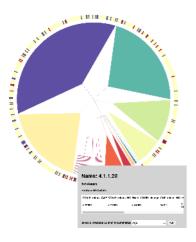
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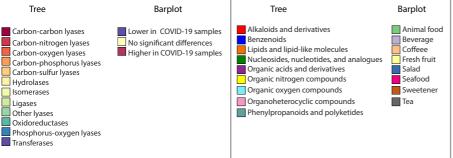
218 References

Thompson, L. R. *et al.* A communal catalogue reveals Earth's multiscale microbial diversity. *Nature* 551, 457–463 (2017).

221	2. Bolyen, E. et al. Reproducible, interactive, scalable and extensible microbiome data
222	science using QIIME 2. Nat. Biotechnol. 37 , 852–857 (2019).
223	3. Vázquez-Baeza, Y., Pirrung, M., Gonzalez, A. & Knight, R. EMPeror: a tool for
224	visualizing high-throughput microbial community data. <i>Gigascience</i> 2 , 16 (2013).
225	4. Lozupone, C. & Knight, R. UniFrac: a new phylogenetic method for comparing microbial
226	communities. Appl. Environ. Microbiol. 71, 8228–8235 (2005).
227	5. Washburne, A. D. et al. Phylogenetic factorization of compositional data yields lineage-
228	level associations in microbiome datasets. <i>PeerJ</i> 5, e2969 (2017).
229	6. Silverman, J. D., Washburne, A. D., Mukherjee, S. & David, L. A. A phylogenetic
230	transform enhances analysis of compositional microbiota data. Elife 6, (2017).
231	7. Morton, J. T. et al. Balance Trees Reveal Microbial Niche Differentiation. mSystems 2,
232	(2017).
233	8. Tripathi, A. et al. Chemically-informed Analyses of Metabolomics Mass Spectrometry
234	Data with Qemistree. <i>bioRxiv</i> 2020.05.04.077636 (2020)
235	doi:10.1101/2020.05.04.077636.
236	9. Nascimento, M. et al. PHYLOViZ 2.0: providing scalable data integration and
237	visualization for multiple phylogenetic inference methods. Bioinformatics 33, 128–129
238	(2017).
239	10. Cordova, J. & Navarro, G. Simple and efficient fully-functional succinct trees. Theoretical
240	Computer Science 656, 135–145 (2016).
241	11. Yu, G. Using ggtree to Visualize Data on Tree-Like Structures. Curr Protoc
242	Bioinformatics 69, e96 (2020).
243	12. Huerta-Cepas, J., Serra, F. & Bork, P. ETE 3: Reconstruction, Analysis, and
244	Visualization of Phylogenomic Data. Mol. Biol. Evol. 33, 1635–1638 (2016).
245	13. Eren, A. M. et al. Anvi'o: an advanced analysis and visualization platform for 'omics data.
246	PeerJ 3, e1319 (2015).
247	14. Stevens, J. R., Jones, T. R., Lefevre, M., Ganesan, B. & Weimer, B. C. SigTree: A
248	Microbial Community Analysis Tool to Identify and Visualize Significantly Responsive
249	Branches in a Phylogenetic Tree. Comput Struct Biotechnol J 15, 372–378 (2017).
250	15. Letunic, I. & Bork, P. Interactive Tree Of Life (iTOL) v4: recent updates and new
251	developments. <i>Nucleic Acids Res</i> 47 , W256–W259 (2019).
252	16. Becker, R. A., Cleveland, W. S. & Wilks, A. R. Dynamic Graphics for Data Analysis.
253	Statist. Sci. 2, 355–383 (1987).
254	17. Shen, Z. <i>et al.</i> Genomic Diversity of Severe Acute Respiratory Syndrome–Coronavirus 2
255	in Patients With Coronavirus Disease 2019. <i>Clin Infect Dis</i> 71 , 713–720 (2020).
256	18. Morton, J. T. <i>et al.</i> Establishing microbial composition measurement standards with
257	reference frames. <i>Nat Commun</i> 10 , 2719 (2019).
258	19. Kanehisa, M. Enzyme Annotation and Metabolic Reconstruction Using KEGG. <i>Methods</i>
259	Mol. Biol. 1611, 135–145 (2017).
260	20. Maras, J. S. <i>et al.</i> Multi-Omics integration analysis of respiratory specimen characterizes
261	baseline molecular determinants associated with COVID-19 diagnosis. <i>medRxiv</i>
262	2020.07.06.20147082 (2020) doi:10.1101/2020.07.06.20147082.
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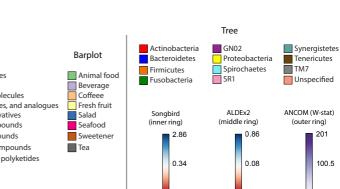
Differential Microbial Functionality in COVID-19 Patients (Metatranscriptomics)





b

Community Analysis of Food-Derived Compounds (LC-MS untargeted metabolomics)



-2 18

С

Comparison of Oral Microbiomes Before and After Toothbrushing (16S rRNA sequencing)



(outer ring)

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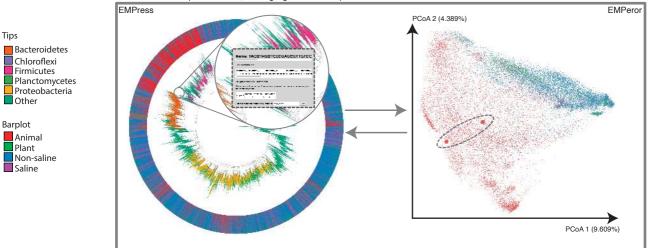
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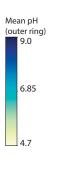
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EMPress and EMPeror are dynamically linked together. For example, clicking on a tip reveals the node's inspection menu, and highlights the 2 samples in the ordination that contain that microbial feature.

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