AIP The Journal of Chemical Physics

Encounter dynamics of a small target by a polymer diffusing in a confined domain

A. Amitai, C. Amoruso, A. Ziskind, and D. Holcman

Citation: J. Chem. Phys. **137**, 244906 (2012); doi: 10.1063/1.4772403 View online: http://dx.doi.org/10.1063/1.4772403 View Table of Contents: http://jcp.aip.org/resource/1/JCPSA6/v137/i24 Published by the American Institute of Physics.

Additional information on J. Chem. Phys.

Journal Homepage: http://jcp.aip.org/ Journal Information: http://jcp.aip.org/about/about_the_journal Top downloads: http://jcp.aip.org/features/most_downloaded Information for Authors: http://jcp.aip.org/authors

ADVERTISEMENT





Encounter dynamics of a small target by a polymer diffusing in a confined domain

A. Amitai,^{1,a)} C. Amoruso,^{1,a)} A. Ziskind,² and D. Holcman¹

¹Ecole Normale Supérieure, Institute of Biology (IBENS), Group of Computational Biology and Applied Mathematics, 46 rue d'Ulm, 75005 Paris, France ²Department of Neuroscience, Center for Theoretical Neuroscience, Columbia University,

1051 Riverside Drive Unit 87 Kolb Research Annex, New York, New York 10032, USA

(Received 12 September 2012; accepted 28 November 2012; published online 28 December 2012)

We study the first passage time for a polymer, that we call the narrow encounter time (NETP), to reach a small target located on the surface of a microdomain. The polymer is modeled as a freely joint chain (beads connected by springs with a resting non zero length) and we use Brownian simulations to study two cases: when (i) any of the monomer or (ii) only one can be absorbed at the target window. Interestingly, we find that in the first case, the NETP is an increasing function of the polymer length until a critical length, after which it decreases. Moreover, in the long polymer regime, we identified an exponential scaling law for the NETP as a function of the polymer length. In the second case, the position of the absorbed monomer along the polymer chain strongly influences the NETP. Our analysis can be applied to estimate the mean first time of a DNA fragment to a small target in the chromatin structure or for mRNA to find a small target. © *2012 American Institute of Physics*. [http://dx.doi.org/10.1063/1.4772403]

I. INTRODUCTION

Polymers such as DNA or mRNA often have to move inside constrained environment such as the nucleus or the cytoplasm before reaching a small, strategic target. Such targets include nuclear pores located on the nuclear envelope or ribosomes dispersed in the cytoplasm involved in protein synthesis:¹ mRNA must exit from the nucleus via passive diffusion in order to synthesize proteins (in the absence of any active transport of the RNA in the nucleus or the cytoplasm²). The task of finding a small nuclear pore can also arise in the context of gene delivery, where DNA fragments have to enter the nucleus. Finally, during the process of double strand DNA repair, DNA ends have to search for one another in the confined chromatin environment,³ and can also localize to the membrane periphery to interact with the nuclear pores.⁴

Previous studies of polymer in confined domains focused on static properties^{5,6} such as translocation,^{7–9} or reptation¹⁰ through a cylindrical tube and their dynamics.¹¹ During translocation, the polymer is threaded through a pore and diffuses until it exits the other side. The polymer has to overcome an effective barrier generated by each monomer on the surface membrane. In contrast, the encounter process we are studying here differs significantly and we shall study the search process of a small target by a freely diffusing polymer. This search process shares some similarities with the polymer looping problem¹²⁻¹⁴ where the critical time scale is defined by the duration for the two polymer ends to meet. We present here a numerical study for the motion of a freely joint chain¹³ polymer in a confined microdomain. Two different types of kinetics for diffusion-controlled processes in dense polymer systems have been discussed in Ref. 15, depending on the root mean square displacement of the active polymer site (compact and noncompact exploration). When the polymer is very small and its motion is dominated by its center of mass diffusion, then the narrow encounter time of the polymer (NETP) is precisely the mean first passage time for a Brownian particle to a small target, also known as the narrow escape time (NET).^{16–24} In a space of dimension *d*, it is given by

$$\langle \tau_{2d} \rangle = \frac{A}{\pi D} \ln \frac{1}{\varepsilon} + O(1),$$
 for $d = 2,$
(1)

$$\langle \tau_{3d} \rangle = \frac{|\Omega|}{4\varepsilon D \left[1 + \frac{L(\mathbf{0}) + N(\mathbf{0})}{2\pi} \varepsilon \log \varepsilon + O(1) \right]}, \text{ for } d = 3,$$
(2)

where A is the area (in two dimensions) and ε is the ratio of the absorbing region of the boundary to the total length of the boundary. In three dimensions, $|\Omega|$ is the volume, ε is the radius of the small absorbing target, and L(0) and N(0) are the two principal curvatures at the origin. D is the diffusion constant. The order one term in each expression depends on the initial position of the moving particle.²² These formulas have been extended to the case of multiple windows.^{22,25,26} We study here how the polymer length controls the mean time for a monomer to reach a small target (a disk of radius ε) located on the boundary of a microdomain. Once a monomer hits this disk, the target is found. In the context of chemical reactions, this time is the reciprocal of the forward reaction rate for diffusion limited processes. The present analysis of the NETP complements previous computations of the forward rate.²⁷⁻³² It may also be used to estimate the first time to activate a gene by a factor located on the DNA, which is

^{a)}A. Amitai and C. Amoruso contributed equally to this work.

different from the classical activation due to a transcription factor.^{33–36} However, at this stage a final analytical formula has to be derived.³⁷

The paper is organized as followed: we first present the freely joint chain polymer, where the monomers are represented by beads connected by elastic springs. We run Brownian simulations and estimate the NETP in two cases:

- 1. When *any* of the polymer monomer can be absorbed at the target region, which we designate $\langle \tau_{any} \rangle$.
- 2. When only one of the monomers can escape through the target region, which we call $\langle \tau_{mon} \rangle$.

Each of these cases might be relevant under different conditions: in some cases, one of the two ends of the polymer has to reach the target, whereas in other situations, it might be sufficient for any part of the polymer to reach the target, such as in gene activation. We then present the NETP simulation results. To study the effect associated to the polymer length, we estimate the mean first passage time of a single absorbing monomer to the boundary of a total absorbing sphere, when the other monomers are reflected. We present various empirical results about the motion of a single monomer. Furthermore, we study the dependence of the NETP on the monomer's position along the chain and the distribution of the arrival times. Finally, we extend the NETP simulation analysis to include bending elasticity.

II. RESULTS

A. The narrow encounter time (NETP) for different polymer length

To study the NETP, we model the polymer motion with a freely joint chain (FJC)¹³ inside a convex domain Ω . Each polymer consists of *N* beads with coordinates $\mathbf{x} = (\mathbf{x}_1, ... \mathbf{x}_N)$, where consecutive beads are connected by springs of characteristic constant *k* and experience isotropic random collision forces. These forces are modeled as Brownian motions. The average bond length between neighboring monomers is given by

$$\langle |\boldsymbol{x}_k - \boldsymbol{x}_{k+1}| \rangle = l_0. \tag{3}$$

Because the DNA molecule is a quite unextendable at small scales below the persistence length of 50 nm, we decided to include this length in our simulations. Thus, contrary to the usual Rouse model³⁸ in which $l_0 = 0$, we study here the FJC where the equilibrium length between the nearest neighbors is nonzero with $l_0 = 50$ nm. For long polymer chains, it should not matter much which of the models (FJC or Rouse) to use, because they share many similarities.³⁸

The dynamics of the polymer is described by the overdamped Langevin-Smoluchowski equation:

$$\dot{\boldsymbol{x}} + \frac{1}{\gamma} \nabla U^N = \sqrt{2D} \dot{\boldsymbol{\eta}},\tag{4}$$

where η is a Gaussian random variable with zero mean and unit variance, γ is the friction coefficient, and U^N is the poJ. Chem. Phys. 137, 244906 (2012)

tential energy generated by the springs

$$U^{N}(\boldsymbol{x}) = \frac{k}{2} \sum_{k=1}^{N-1} (|\boldsymbol{x}_{k} - \boldsymbol{x}_{k+1}| - l_{0})^{2}.$$
 (5)

For all simulations, we use a circular or spherical microdomain Ω . To construct the initial configuration of the polymer, we put all monomers close to the origin folded on the top of each other and then ran simulations for a time long enough that depend on the size of the polymer. Indeed, for short polymers, we run simulations during a period of time $\frac{R^2}{D_{CM}} = NR^2/D$ (*R* is the radius of the sphere), which represent the mean time for the center of mass to explore the spherical domain. For longer polymers for which the radius of gyration is comparable to *R*, we run a preliminary simulation for a time equal to the longest relaxation time of the polymer $\frac{l_0^2N^2}{D}$.³⁸ To conclude, to make sure that the initial configuration of the polymer inside the sphere was chosen at equilibrium, we run an initial simulation up to time max{ $\frac{NR^2}{D}$, $\frac{l_0^2N^2}{D}$ }.

Most of the boundary is reflecting, except for the small absorbing window, where the polymer can be absorbed. Sample trajectories (Fig. 1) of the center of mass (initial position at the center of the domain) for different sets of parameters with beads and springs (blue), the boundary (green), and the absorbing target (red). The analytical estimation of the NETP remains a challenging problem, since the computation involves generalizing the narrow escape theory to the tubular neighborhood of the absorbing hole in dimension 2*N* or 3N.³⁷ However, this computation does not fall into the narrow escape methodology, ^{16–24} rather the computation requires to analyze the mean first passage time of a stochastic particle in a narrow band in high dimension, and thus many of the approximations used for a punctual particle are no longer valid. Obtaining such a formula for the polymer would be useful to better explore the complexity of the parameter space.

B. The NETP has a bell shape profile

To study the dependency of the NETP as a function of the polymer length *N*, we use the Euler's scheme (Eq. (A2)) and run Brownian simulations for a range of polymer lengths from N = 1 to N = 350, in dimensions two and three under the two scenarios described above. In Figs. 2–4, the NETP values are normalized to the NET for a single particle, which we obtained from Brownian simulations: $\tau_0 = 3s$ in dimension two and $\tau_0 = 15s$ in dimension 3, with parameters listed in Table I and the diffusion constant for a single monomer is $D = 0.04 \ \mu m^2/s.^{48}$ The NETP curve contains two phases that we shall now discuss (the NET τ_0 for one particle is referred in the figures as $\langle \tau_{2d} \rangle$ and $\langle \tau_{3d} \rangle$ in dimension 2 and 3, respectively).

1. The initial phase of the NETP increases linearly with the polymer length

In dimensions two and three, the NETP is initially an increasing function of the number of beads N, until a critical value N_c after which it is decreasing. With the parameter of



FIG. 1. Confined trajectories of FJC polymer. In a circular(2D)/spherical(3D) domain of radius 250 nm, the target size is $\varepsilon = 50$ nm. Trajectories of the center of mass (black) before absorption at the target (red). (a) Simulation with 1 bead, moving in 2 dimensions. (b)–(c) Simulations of a 4-bead FJC polymer, moving in 2 dimensions (b) and in 3 dimensions (c).

Table I, using Brownian simulations, we found that the critical value $N_c \in [10 - 15]$ (later on, we obtained the same critical length when only a single monomer can be absorbed). We were surprised to see that the polymer size associated with this critical number is much smaller than the critical length obtained from the radius of gyration $R_g \approx \sqrt{N_c l_0}/\sqrt{6} \approx 150$. This suggests that the the radius of gyration is not sufficient to characterize the effect of the confinement on the polymer dynamics.

In addition, for short polymer lengths, such that $N < N_c$, the NETP is largely determined by the motion of the center

TABLE I. Parameters of the simulations.

Parameters	Description	Value
R	Radius of the circular/spherical domain	250 nm
а	Radius of the absorbing window	50 nm
l_0	Polymer persistence length	50 nm ⁴⁶
D	Diffusion constant	$4 \times 10^{-2} \mu \text{m}^2/\text{s}$ (Ref. 48)
γ	Friction coefficient	3.1×10^{-5} Ns/m
k	Spring constant	$1.75 \times 10^{-2} \mathrm{Nm^{-1}}$
		(Ref. 47)

of mass. When the polymer is far from the absorbing target, none of the beads will be able to reach it, until the center of mass has moved close to the target. The NETP thus reflects the mean first passage time of the center of mass to the target. In this limiting case, the center of mass undergoes Brownian motion with a diffusion constant inversely proportional to the number of beads: $D_{\rm CM} = \frac{D}{N}$. Thus, in the regime $N \ll N_c$, the NETP is approximately that of a single particle, but with a smaller diffusion constant. The expression for the NET ((1) and (2)) are inversely proportional to $D_{\rm CM}$, and thus we obtain the initial linear regime in N:

$$\begin{aligned} \langle \tau_{\text{any}} \rangle_{2d} &\approx N \langle \tau_{2d} \rangle, \\ \langle \tau_{\text{any}} \rangle_{3d} &\approx N \langle \tau_{3d} \rangle, \end{aligned}$$
 (6)

as confirmed in Fig. 2. However, for a polymer of length comparable to the size of the microdomain, the location of the center of mass does not determine anymore the NETP. In that regime, smaller subsections of the polymer can be close to the target even if the center of mass is far away. In addition, when any bead can be absorbed, increasing the polymer length results in a decrease in the NETP (Fig. 2). Interestingly, two regimes can be further distinguished for the decay phase.



FIG. 2. NETP for any monomer to the target, for different polymer lengths. The results are normalized to τ_0 (the NET for 1 bead), Eqs. (1) and (2). Depicted is the mean time for any of the beads to reach the target in dimension two (a) and three (b). Each data point represents the average over 2000 runs. The Brownian data are fitted by a double exponential function in dimension 2, $\frac{(\tau_{any}(N))_{2d}}{(\tau_{2d})} = a_2 \exp(-\alpha_2 N) + b_2 \exp(-\beta_2 N)$ with exponents $\alpha_2 = 0.0075$, $\beta_2 = 0.024$, and coefficients $a_2 = 0.23$, $b_2 = 2.17$ and in dimension 3 $\frac{(\tau_{any}(N))_{2d}}{(\tau_{2d})} = a_3 \exp(-\alpha_3 N) + b_3 \exp(-\beta_3 N)$ with exponents $\alpha_3 = 0.0082$, $\beta_3 = 0.030$ and coefficient $a_3 = 0.60$, $b_3 = 1.56$. (c) Probability distribution $P[\tau/\tau_0]$ of arrival times for any monomer to a small target (in three dimensions) for N = 100. The probability distribution of the arrival times to a small hole, located on the boundary of a sphere in three dimensions, is approximated by a sum of two exponentials. The datas are well approximated by a single exponential of the form $Pr\{\tau_{3d} = t\} = a \exp(-\alpha t)$ with a = 0.814, and $\alpha = 4.185$.

The decay phase of the NETP is approximated by an exponential

When any bead can be absorbed, the decay phase of the NEPT (Fig. 2) can be separated into two different regimes that can be described as followed. In the first one, the polymer moves freely until a monomer hits the absorbing boundary. The NETP is determined by the competitive effects of a decreased diffusion constant for the center of mass and an increased total polymer length. Increasing the polymer length leads to an effective smaller volume of the effective confining domain in which the polymer has to find the absorbing window. To investigate more specifically the dynamics of the polymer, we monitored in our simulations the time evolution of several polymers and observed at equilibrium various heterogeneous configurations with multiple pearl-like-structure (PLS) (Fig. 3). Thus, a polymer forms transient PLSs, sometimes a single PLS (Fig. 3(a)), or two transient small ones (Fig. 3(b)), or even three little ones (Fig. 3(c)). We conclude that a polymer of intermediate size is not simply uniformly distributed, rather its shape changes, making transient between various substructures, characterized as PLSs. These transients structures (Fig. 3(c)) suggest that the NETP as a function of N for a confined polymer cannot result from a simple scaling law, that can be analyzed from a steady state distribution.

In a second regime, when the length of the polymer becomes long enough, so that at least one monomer can always be found in the boundary layer (of size ε) of the absorbing hole,^{17,18} we expect the NETP to have a different decay as a function of *N* compared to the previous intermediate regime. In that case, the center of mass is strongly restricted due to the interaction of all the monomers with the microdomain



FIG. 3. Snapshots of the polymer configuration during the Brownian dynamics. A polymer (N = 100) is shown at (a) 7.5 ms, (b) 30 ms, and (c) 60 ms during the Brownian dynamics (the initial configuration of the polymer has uniformly distributed angles, with length l_0). This simulation reveals pearllike structures (PLSs): a large PLS in (a), two PLSs (b), and three small ones (c). The time step is $\Delta = 1.5 \times 10^{-4}$ s.



FIG. 4. Dynamics and distribution of a single monomer: (a) NETP for the three monomers: end, middle and N - 1 as a function of the polymer length N in three dimensions (Brownian simulations): the encounter time is normalized to the time τ_0 (for a single bead). Parameters are described in Table I. (b) Probability distribution $P[\tau/\tau_0]$ of arrival times for the end monomer to a small target (in three dimensions). The datas are well approximated by a single exponential of the form $P\{\tau_{3d} = t\} = a \exp(-\alpha t)$ with a = 1.014, $\alpha = 0.76$. Monomers radial distribution: (c) probability distribution function (pdf) of monomers position. The radial pdf is calculated from Brownian simulations for the end monomers N = 16 (points), N = 150 (full line), and the middle monomers N = 16 (points-line), N = 150 (dashed line). (d) The pdf of the center of mass for N = 16 (points), N = 48 (dashed line), N = 150 (full line). The normalized effective potential $U_N(r) = -k_B T \log (P(r))$ acting on a monomer was calculated from the radial pdf. (e) The potential for the end and middle monomers are significantly different, confirming that the boundary has different effect depending on the position of each monomer.

surface (Fig. 4(d)). The NETP is determined by mean time for a monomer in the boundary layer of the absorbing window. It is still an unsolved problem to obtain an asymptotic estimate for that time. Using an optimal fit procedure, we obtain for the NETP an empirical scaling approximation with two exponentials

$$\frac{\langle \tau_{\text{any}}(N) \rangle_{2d}}{\langle \tau_{2d} \rangle} = a_2 \exp(-\alpha_2 N) + b_2 \exp(-\beta_2 N) \text{ for } d = 2$$
(7)

and

$$\frac{\langle \tau_{\text{any}}(N) \rangle_{3d}}{\langle \tau_{3d} \rangle} = a_3 \exp(-\alpha_3 N) + b_3 \exp(-\beta_3 N) \text{ for } d = 3.$$
(8)

 $\langle \tau_{3d} \rangle$ and $\langle \tau_{2d} \rangle$ are the NET for a single particle τ_0 , in dimension 3 and 2, respectively. In dimension two, the exponents are $\alpha_2 = 0.0075$, $\beta_2 = 0.024$ and coefficients $a_2 = 0.23$, $b_2 = 2.17$ and in dimension 3, $\alpha_3 = 0.0082$, $\beta_3 = 0.030$ and coefficient $a_3 = 0.60$, $b_3 = 1.56$ (Fig. 2). The empirical laws (7) and (8) remain to be derived analytically.

3. The arrival time is approximately Poissonian when any monomer can reach the target

Finally, we have shown in Fig. 2(c) that the histogram of arrival time of any monomer to the small target can be well approximated by a single exponential, suggesting that the arrival time be almost Poissonian. This result can appear quite surprising, but it is the effect of the small target, which select the long time behavior of the polymer.¹⁹

4. Phenomenological explanation of NETP bell shape

The bell shape nature of the NETP can be qualitatively explained using the NET Eqs. (1) and (2). Indeed, a small polymer can be considered as a quasi-particle of radius $R_g = l_0 \sqrt{N/6}$,¹⁰ evolving in an effective domain, which is the full domain minus its volume. Thus, the NETP is related to the mean first passage time of the quasi-particle with diffusion constant $D_{\rm CM} = \frac{D}{N}$ in the apparent domain of volume $V_a = \frac{4\pi}{3}(R - R_g)^3$, leading to a mean time proportional

$$\frac{V_a}{\varepsilon D_N} = N \frac{4\pi}{3\varepsilon D} (R - l_0 \sqrt{N/6})^3.$$
(9)

This phenomenological formula shows that the mean time has a maximum for $N_m = 25$ (parameter of Table I), which is an over estimation of the empirical value that we obtained from Brownian simulation N = 10. To summarize, when each bead can reach the absorbing target, the NETP exhibits two different behaviors: while for short polymer size, the dynamics can be abstractly described by a quasi-particle, leading to a maximum of the NETP, for longer polymers, the NETP decays with a single exponential, a behavior that was not expected. An analytical derivation of this result is still missing.

C. When a single monomer can be absorbed, the NETP depends on its location on the polymer chain

To study the impact on the NETP of the monomer location along the polymer chain that can be absorbed at the target, we ran a new set of Brownian simulations (Fig. 4). The polymer is confined in a sphere and only one monomer (we call it active monomer) can be absorbed, while all others are reflected at the target site. Interestingly, the simulation reveals that the location of the absorbing monomer on the chain can drastically affect the arrival time to the small target. Between the middle and the end monomer, we observe a factor 3 reduction in the arrival time. Interestingly, already taking the N-1th monomer compared to the last one is making a noticeable difference. In addition, the NETP increases to a different plateau that depends also on the monomer location: the plateau starts at a length N = 20 for the end monomer, while it is around N = 75 for the middle monomer. We note that the effect of the boundary starts on the polymer dynamics is seen already for N = 20, which is much smaller than the length of a polymer for which the gyration radius is comparable to the domain radius.

To analyze this behavior, we examined each phase separately. In the increasing phase (as a function of the polymer length, Fig. 4(a)), the arrival of the monomer to the target is mainly governed by the diffusion of the polymer center of mass, which can be approximated by a diffusing ball of diffusion constant $D_{\rm CM} = D/N$. However, for larger *N*, this approximation is not valid, rather the arrival time converges to a constant value that depends on the structure of the polymer where the stochastic dynamics is much richer than simple diffusion. The motion of the active monomer, which is affected by the entire polymer, depends on its length and the boundary of the confined domain.³⁹ Intuitively, the interaction of the active potential through their interaction with the boundary, which is different for the middle and the end monomer, leading to the major differences reported in Figs. 4(a) and 4(c).

For N large enough, the middle monomer is more confined than the end one, and by analogy with the diffusion of a stochastic particle in a potential well, the middle monomer has to surmount a higher potential barrier to reach the small target located on the boundary. To clarify this hand waving explanation, we approximate the active monomer motion as diffusion in a spherical symmetrical potential V inside our confined spherical domain for the polymer. The stochastic description is

$$\dot{X} = \frac{V(X)}{\gamma} + \sqrt{\frac{2kT}{\gamma}} \dot{\boldsymbol{w}}, \qquad (10)$$

where \boldsymbol{w} is the standard Brownian motion. Using the symmetry of the domain, the potential V has a single minimum at the center. Interestingly, in the high potential barrier approximation, the mean time to a small target does not depend on the specific shape of the potential, but rather on its minimum and maximum.⁴⁰ In that case, the mean time $\tau(N)$ to a target, which depends on the polymer size is given by

$$\tau(N) = \frac{(2\pi)^{3/2} \gamma \sqrt{kT}}{4a\omega_N^{3/2}} \exp\left[\frac{U_N}{kT}\right],\tag{11}$$

where $U_N(r)$ is the energy barrier generated by the polymer due to the presence of the boundary of the domain at the target site and ω_N is the frequency at the minimum $(U_N(r) \approx \frac{\omega_N}{2}r^2$ near 0).⁴⁰ The potential $U_N(r)$ can be recovered from formula (11) and the numerical simulations described in Fig. 4(c). We conclude that the position along the polymer chain of the monomer interacting with the target critically influences the search process.

We now directly evaluate the monomers distribution inside our confined domain using a numerical approach. To further characterize the dynamics of a single monomer, we computed the equilibrium radial probability distribution function (pdf) P(r) of the monomers position. We found that the monomers, depending on their location along the chain, are differently distributed inside the confined domain (Figs. 4(c) and 4(d)). The middle monomer is more restricted to the center, away from the boundary compared to the end monomer, which explores in average a larger area. As the length of the chain increases, the middle monomer gets more restricted, while the end one does not seem to be much affected (Fig. 4(c)). The center of mass has a similar behavior as the middle monomer and its spatial distribution gets restricted for increasing polymer length. From the radial pdf, we decided to evaluate the effective potential $U_N(r)$ acting on a monomer by $U_N(r) = -k_B T \log (P(r))$ defined in Eq. (11). The potential acting on the center of mass is large enough so that it never arrived to the periphery during our simulations (Figs. 4(e) and 4(f)). Finally, similar to the case where all the monomers can interact with the target, we show in Fig. 4(b), that the arrival time distribution is well approximated by a single exponential, showing that the distribution is almost Poissonian. However, the rate depends on the location of absorbing monomer along the polymer chain.

1. Influence of the polymer length on the arrival time of an ensemble of absorbing monomers to the sphere boundary

To evaluate the effect of the polymer length on the monomer dynamics, we study the arrival time of the ensemble of *n* consecutive (the first one a polymer end) monomers to a sphere. As described in Sec. II C, the reflecting monomers can prevent the absorbing ones to reach the boundary of the sphere. To explore the arrival time as a function of *n*, we run Brownian simulations in a three dimensional sphere for various polymer sizes (*N* monomers) and absorbing lengths *n*, while the N - n remaining monomers are reflected on the surface. We have found (Fig. 5(a)) that the arrival time decays with *n*, independently of the polymer size (N = 40, 140, 220).

To further investigate the role of the polymer length on the arrival time, we studied the recurrence time of a single monomer, which is the time for that monomer to move from a sphere of radius $R_2 = \frac{R_1}{2}$ to the boundary (radius R_1 = 250 nm), as described in Figs. 5(b)-5(d). The recurrence time is increasing with the size of the polymer. However, it becomes independent on the polymer length for $N \ge 20$ (Fig. 5(e)) while the motion of a single monomer does not depend on the size of polymer. In the regime of large N, the longest relaxation time due to the internal modes of the polymer³⁸ is larger than the time it takes for a monomer to be absorbed at boundary (this situation happens when the radius of gyration of the polymer is larger than the radius of the sphere). To clarify that the time to absorption is independent of the total polymer length N, we simply recall that the motion of a single monomer in the large N regime is that of a correlated particle, that can be described as anomalous.³⁹ In that case, the mean square displacement is shown³⁹ to be independent of the polymer length. To conclude, Fig. 5(d) shows that the reflecting monomers prevent the absorbing ones to reach the surface of the sphere only below a certain number.

D. NETP with additional stiffness

In this final section, we explore the consequence of adding flexibility on the NETP, which can characterize complex DNA molecule containing various bound proteins. We account for the stiffness by including the bending energy

$$U_{bend}(\mathbf{x}) = \frac{\kappa_{ang}}{2} \sum_{i=1}^{N-1} (\mathbf{u}_{i+1} - \mathbf{u}_i)^2 = \kappa_{ang} \sum_{i=1}^{N-1} (1 - \mathbf{u}_i \cdot \mathbf{u}_{i+1}),$$
(12)



(a) Initial configuration (time t_1) (b) Intermediate configuration (time t_2) (c) Final configuration (time t_3)

FIG. 5. The recurrence time of a polymer between two concentric disks and the influence of the absorbing polymer size. A perfectly flexible polymer is introduced into the spherical domain and we preform a preliminary Brownian simulation until the equilibrium regime is achieved. After that, we estimate the first time t_1 that one of the end monomer hits the surface of the external sphere (a) then the first time t_2 that it enters into the sphere of radius R_2 (b) and finally the first time t_3 that it hits again the boundary of the external sphere (c). The recurrent time is average of $t_3 - t_1$ over many realizations. (d) recurrence time for the end monomers (polymer of length N = 60) in dimension three, computed as described in panel a-c. (e) Effect of the polymer reacting site size on the arrival time to a fully absorbing three dimensional sphere. The polymer is composed of *N* monomers, with *n* absorbing ones, the other N - n are reflected at the external sphere. We present the NETP to the boundary for three polymer sizes (40, 140, and 220). The results are normalized by the NET for a single Brownian particle $\langle \tau_{3d} \rangle$.

where $u_i = \frac{x_{i+1}-x_i}{|x_{i+1}-x_i|}$ is the unit vector connecting two consecutive monomers and x_i is the position of the *i*th monomer. This potential depends on the angle θ_i between two successive monomers with the relation $u_i \cdot u_{i+1} = \cos \theta_i$. The bending rigidity κ_{ang} is related to the persistence length L_p of the polymer by the expression⁴¹

$$L_p = \frac{\kappa_{ang} l_0}{k_B T},\tag{13}$$

where k_B is the Boltzmann constant, *T* is the temperature, l_0 has been defined above. The persistence length quantifies the stiffness of a polymer and can be characterized using the unit tangent vectors t(s) and t(0) at positions *s* and 0 along the polymer. Averaging over all starting positions, the expectation of ϕ , which is the angle between t(s) and t(0), falls off exponentially with the distance *s* along the polymer

$$\langle \hat{u}(s) \cdot \hat{u}(0) \rangle = e^{-s/L_P}.$$
(14)

For our simulations, we use the value $\kappa_{ang} = 5$ (Ref. 42) that leads to $L_p = 250$ nm, which is the reported value of the persistence length of chromatin fibers.^{43,44} We use the rigid polymer dynamics obtained from the over-damped Langevin-Smoluchowski equation

$$\dot{\mathbf{x}} + \nabla U = \sqrt{2D} \dot{\mathbf{w}},\tag{15}$$

where the total potential $U = U(\mathbf{x})$ is the sum of two energy potentials

$$U(\mathbf{x}) = U^{N}(\mathbf{x}) + U_{bend}(\mathbf{x}), \qquad (16)$$

where $U^N(\mathbf{x})$ is the elastic potential defined in Eq. (5). Interestingly, the two potentials are of the same order of magnitude: using the maximum extensibility of DNA about 10% of its total length,⁴⁵ in that case the length $r - l_0 \sim 5$ nm $(r = |x_{k+1} - x_k|)$ and $U_{el}^N = \frac{1}{2}k(r - l_0)^2 \sim 2 \times 10^{-18}$ Nm, while the maximum energy between three consecutive monomers due to bending is $U_{bend} = k_{ang}(1 - \cos \theta) = k_B T * L_p/l_0 * \sim 5 \times 10^{-19}$ Nm. We simulate the arrival time of any monomer to the absorbing boundary and use the same fitting procedure as in Fig. 2. It leads in dimension 2 to (Fig. 6(a))

$$\frac{\langle \tau_{\text{any}} \rangle_{2d}}{\langle \tau_{2d} \rangle} = a_2 \exp(-\alpha_2 N) + b_2 \exp(-\beta_2 N)$$
(17)

with $a_2 = 0.37$, $b_2 = 2.9$, and $\alpha_2 = 0.02$, $\beta_2 = 0.18$ and in dimension 3 (Fig. 6(b)),

$$\frac{\langle \tau_{\text{any}} \rangle_{3d}}{\langle \tau_{3d} \rangle} = a_3 \exp(-\alpha_3 N) + b_3 \exp(-\beta_3 N)$$
(18)

with $a_3 = 0.28$, $b_3 = 1.64$, and $\alpha_3 = 0.01$, $\beta_3 = 0.12$. Compared to the nonflexible polymer, the maximum of the NETP is now shifted towards smaller values of *N*: the NETP is an increasing function of *N* for N < 6, and for large *N*, it is a decreasing function of *N*. We compare these results with the ones for flexible polymers: in a confined spherical cavity, flexible polymers have a tendency to fill the available space, with a probability of finding a monomer at the center of the cavity being higher than at the boundary, when the persistence length is comparable to the size of the sphere. On the contrary, for stiff polymers, the chain is forced to bend abruptly



FIG. 6. NETP for a semi-flexible polymer. The NETP is normalized to τ_0 (the NET for 1 bead, Eqs. (1) and (2)). The simulations are for any of the beads of a semiflexible polymer to reach the small target in dimension 2 (a) and 3 (b). Each data point is an average over 2000 realizations. An double exponential fit leads to $\frac{\langle i_2 \rangle}{\langle \tau_{2d} \rangle} = a_2 \exp(-\alpha_2 N) + b_2 \exp(-\beta_2 N)$ with $a_2 = 0.37$, $b_2 = 2.9$ and $\alpha_2 = 0.02$, $\beta_2 = 0.18$ and in dimension 3, $\frac{\langle i_3 \rangle}{\langle \tau_{2d} \rangle} = a_3 \exp(-\alpha_3 N) + b_3 \exp(-\beta_3 N)$. The same fit in three dimensions gives $a_3 = 0.28$, $b_3 = 1.64$ and $\alpha_3 = 0.01$, $\beta_3 = 0.12$. (c) Fraction of time a monomer spend near the boundary for a flexible and nonflexible polymer. (d) Snapshot of a nonflexible polymer that has the tendency to locate near the spherical boundary.

close to the surface and a large fraction of the polymer remains closed to the surface (Figs. 6(c) and 6(d)). Thus, for a target located on the surface, a monomer located on a nonflexible polymer should reach it in an almost two-dimensional process, decreasing the NETP.

We conclude that large enough nonflexible polymer find small targets faster than completely flexible ones, this is due to an increase probability to find monomers in the close vicinity of the boundary where the small target is located.

III. DISCUSSION AND CONCLUSION

We explored here by means of Brownian simulations, the mean time for part of a FJC polymer to find a small target located on the surface of a microdomain. When each of the monomers can be absorbed, our analysis reveals a bellshaped curve (Fig. 2). For a short polymer, it must approach close to the target for one monomer to be absorbed and the main contribution to the NETP is given by the diffusion of the center of mass of the polymer D_{CM} . Since D_{CM} is inversely proportional to N, the center of mass moves slower and the NETP increases with N. However, for longer polymers whose lengths are comparable to the size of the microdomain, some monomers can reach the target even if the center of mass is far away. In that regime, increasing the polymer length results in a decrease in the NETP. It is surprising that the empirical fit of our Brownian simulations requires two exponentials. It would be interesting to derive analytically this law and the expression for the associated parameters. Another consequence of the present analysis is the approximation of the arrival time of a monomer to the target by a single exponential, showing the Poissonian nature of this process. Indeed, hitting the target is a rare event. We further showed that the polymer dynamics can drastically affect the NETP of a single monomer to the target. In that case, a repulsive force near the boundary is generated by the other reflecting monomers (Fig. 4). Consequently, the NETP converges as the polymer increases to a constant value, which depends on the monomer position along the polymer chain.

Our analysis has several applications. In the context of chromatin dynamics in the nucleus, the long polymer regime shows that activation, defined as the arrival of one monomer to an active site by the chromatin segment, depends strongly on the chromatin length. Moreover, when the polymer represents a dsDNA break and the microdomain is the local confined domain generated around, our analysis shows that the mobile DNA segment (length of the polymer) significantly affects the search time: we found that increasing the length from small (free Brownian) to the maximum value leads to a factor larger that two difference (Fig. 2). To be quantitative, we can estimate the mean time for a dsDNA break to find a specific target such as the other strand, with the following parameters: the free DNA (persistent) length is 50 nm; it is located inside a chromatin sphere of radius 250 nm; the diffusion coefficient is $D = 4.10^{-2} \mu \text{m}^2/\text{s}$. We find using the result of Fig. 2 and formula (7)–(8) that the NETP (mean time for a monomer to find by diffusion a specific chromatin element of size 12.5 nm or the other dsDNA break) is around $\tau = [100 - 200]$ s.

Finally, because a dsDNA break has been found to be directed to the nuclear boundary,⁴ we postulated that a specific mechanism should be involved. Indeed our analysis (Fig. 5) reveals that the reflective interaction of the polymer chain with the nuclear surface generates a local potential that prevents the break to approach the nuclear surface. Thus, by increasing the surface of the target, which can results from dephosphorylation, leads to a decrease (Fig. 5) of the NETP of the break to the nuclear surface. It would be interesting in a future study to add the effect of nucleosomes.³ The present work disregards the effect of an external field (such as an electric field), which could be induced in in vitro experiments. It would be interesting to add the effect of a field and explore the consequences on the NETP. In addition, the present study could also characterize the search of a target by a charged polymer. Indeed, because the Debye length is 0.8 nm (about the size of a monomer), the polymer, even if it is charged, cannot feel the target, showing that the present results are quite general.

ACKNOWLEDGMENTS

D.H.'s research is supported by an ERC starting grant.

APPENDIX: NUMERICAL METHODS AND PARAMETER CALIBRATIONS

1. Simulation procedure

For a chain of N beads (and N - 1 springs), we initialized the chain with its center of mass at the center of the domain, with the angle between adjacent springs random (uniformly distributed between 0 and 2π).

At each time step, the monomers (beads) move according to Eq. (A2). When a Brownian bead crosses numerically the boundary of the domain, it is reflected according to Snell's law (angle of incidence = angle of reflection). A monomer is "absorbed" at the target region if the line drawn from its position at the previous time step to its current position intersects with the target region of the boundary. We record the first time any of the beads reaches the absorbing region as τ_{any} . If the bead is not the active monomer, it is reflected back into the domain and the simulation continues. If the bead is an active monomer, the simulation ends and the final simulation time is recorded as τ_{mon} . All our results are displayed in reduced units. We have used the USFC Chimera software⁴⁹ for visual snapshots of the polymer dynamics.

2. Simulation parameters

a. Dimensions of the microdomain

We use for the microdomain, a disk and and a sphere in dimensions two and three, respectively, of radius R = 250 nm. For a size of the hole of 50 nm, we obtain that $\varepsilon = \frac{a}{R} = 0.05$.

b. Spring resting length I₀

The resting length of the spring connecting two neighboring bead to be 50 nm (approximately 150 base pairs).⁴⁶

c. Spring constant k

Using the direct measurements⁴⁷ of the elasticity of short stretches (30 base pairs) of double stranded DNA (dsDNA), the Young's Modulus of dsDNA is estimated to be 55 MPa.⁴⁷ We now estimate the spring constant *k*. Using that the ds-DNA was on average initially 8.5 nm, but when a force of 113.7 pN was applied, it stretches to an additional 6.5 nm, under the assumption that the dsDNA is still in the Hookean regime (i.e., stretching linearly), we obtain a spring constant of $k = F/x = 1.75 \times 10^{-2}$ N/m.

d. Integration algorithm and time step Δt

The simulation time step Δt is chosen such that each bead moves on average less than the distance Δx^* at each time step, where Δx^* is the smaller length scale of our system, which is the diameter of the small hole $2\varepsilon R$. In practice, we used Δx^* $= f \times (2\varepsilon R)$ where f is an extra precision parameter, fixed to 0.2. Hence, we used $\Delta t = (\Delta x^*)^2/2D = 1.5 \times 10^{-4}$ s. The numerical scheme is as follows:

$$\mathbf{x}(t + \Delta t) = \mathbf{x}(t) + \frac{1}{\gamma} \nabla U^N \Delta t + \eta \sqrt{2D\Delta t}, \quad (A1)$$

that is (for one of the non-end beads),

$$\begin{aligned} \mathbf{x}_{k}(t + \Delta t) \\ &= \mathbf{x}_{k}(t) - \frac{k}{\gamma} \left((\mathbf{x}_{k}(t) - \mathbf{x}_{k+1}(t)) - l_{0} \frac{\mathbf{x}_{k}(t) - \mathbf{x}_{k+1}(t)}{|\mathbf{x}_{k}(t) - \mathbf{x}_{k+1}(t)|} \right. \\ &+ (\mathbf{x}_{k}(t) - \mathbf{x}_{k-1}(t)) - l_{0} \frac{\mathbf{x}_{k}(t) - \mathbf{x}_{k-1}(t)}{|\mathbf{x}_{k}(t) - \mathbf{x}_{k-1}(t)|} \right) \Delta t \\ &+ \eta \sqrt{2D\Delta t}, \end{aligned}$$
(A2)

where η is a Gaussian random variable with zero mean and unit variance.

e. The friction coefficient γ and the diffusion constant D

The diffusion constant *D* of a DNA molecule has been estimated⁴⁸ to be $D = 4 \times 10^{-2} \,\mu\text{m}^2/\text{s}$. The friction coefficient is computed by using Einstein relation $D = \frac{k_B T}{\gamma}$, at room temperature and $\gamma = 10^{-7}$ Ns/m. The relaxation time constant of each spring is $\frac{\gamma}{k} \sim 10^{-5}$ s, much shorter than the time step we used. Our simulations occur in a regime where the springs relax to their resting length l_0 at every time step. Thus, the polymer keeps its original length $(N - 1)l_0$.

f. Simulation of the stiff polymer in the microdomain

We first introduced in the sphere a perfectly flexible polymer ($\kappa_{ang} = 0$), then we give it a stiffness by changing the value of κ_{ang} and a preliminary Brownian dynamics is performed to ensure equilibration of the polymer.

- ¹B. Alberts, A. Johnson, J. L. Raff, K. Roberts, and P. Walter, *Molecular Biology of the Cell*, 5th ed. (Garland Science, 2007).
- ²W. F. Marshall, A. Straight, J. F. Marko, J. Swedlow, A. Dernburg, A. Belmont, A. W. Murray, D. A. Agard, and J. W. Sedat, Curr. Biol. 7, 930 (1997).
- ³A. Groth, W. Rocha, A. Verreault, and G. Almouzni, Cell **128**, 721–733 (2007).
- ⁴S. Nagai, K. Dubrana, M. Tsai-Pflugfelder, M. B. Davidson, T. M. Roberts, G. W. Brown, E. Varela, F. Hediger, S. M. Gasser, and N. J. Krogan, Science **322**, 597–602 (2008).
- ⁵T. Sakaue and E. Raphael, Macromolecules **39**, 2621–2628 (2006).
- ⁶J. Dayantis and J. Sturm, Polymer **26**, 1631–1637 (1985).
- ⁷J. Chuang, Y. Kantor, and M. Kardar, Phys. Rev. E 65, 011802 (2002).
- ⁸C. T. A. Wong and M. Muthukumar, Biophys. J. **95**, 3619 (2008).
- ⁹P. Park and W. Sung, Phys. Rev. E **57**, 730 (1998).
- ¹⁰P.-G. de Gennes, *Scaling Concepts in Polymer Physics* (Cornell University, Ithaca, NY, 1979).
- ¹¹S. C. Weber, J. A. Theriot, and A. J. Spakowitz, Phys. Rev. E 82, 011913 (1998).
- ¹²R. W. Pastor, R. Zwanzig, and A. Szabo, J. Chem. Phys. **105**, 3878 (1996).
- ¹³N. M. Toan, G. Morrison, C. Hyeon, and D. Thirumalai, J. Phys. Chem. B 112, 6094 (2008).
- ¹⁴A. Amitai, I. Kupka, and D. Holcman, Phys. Rev. Lett. **109**, 108302 (2012).
 ¹⁵P. G. de Gennes, J. Chem. Phys. **76**, 3316 (1982).
- ¹⁶A. Singer, Z. Schuss, D. Holcman, and B. Eisenberg, J. Stat. Phys. **122**, 437–463 (2006).
- ¹⁷A. Singer, Z. Schuss, and D. Holcman, J. Stat. Phys. **122**, 465 (2006).
- ¹⁸A. Singer, Z. Schuss, and D. Holcman, J. Stat. Phys. **122**, 491 (2006).
- ¹⁹Z. Schuss, A. Singer, and D. Holcman, Proc. Natl. Acad. Sci. U.S.A. 104, 16098 (2007).
- ²⁰D. Holcman and Z. Schuss, J. Stat. Phys. 117, 191 (2004).
- ²¹M. J. Ward and J. B. Keller, SIAM J. Appl. Math. **53**, 770 (1993).
- ²²S. Pillay, M. Ward, A. Peirce, and T. Kolokolnikov, Multiscale Model. Simul. 8, 803–835 (2010).
- ²³I. V. Grigoriev, Y. A. Makhnovskii, A. M. Berezhkovskii, and V. Y. Zitserman, J. Chem. Phys. **116**, 9574 (2002).
- ²⁴J. Reingruber, E. Abad, and D. Holcman, J. Chem. Phys. **130**, 094909 (2009).
- ²⁵D. Holcman and Z. Schuss, Phys. Lett. A **372**, 3768–3772 (2008).
- ²⁶D. Holcman and Z. Schuss, J. Phys. A **41**, 155001 (2008).
- ²⁷G. Wilemski and M. Fixman, J. Chem. Phys. 58, 4009 (1973).
- ²⁸R. Zwanzig, Proc. Natl. Acad. Sci. U.S.A. **87**, 5856 (1990).
- ²⁹R. Zwanzig and A. Szabo, Biophys. J. 60, 671 (1991).
- ³⁰A. Szabo, K. Schulten, and Z. Schulten, J. Chem. Phys. **72**, 4350 (1980).
- ³¹K. Schulten, Z. Schulten, and A. Szabo, J. Chem. Phys. **74**, 4426 (1981).
- ³²A. Perico and M. Battezzati, J. Chem. Phys. **75**, 4430 (1981).
- ³³G. Malherbe and D. Holcman, Phys. Lett. A **374**(3), 466–471 (2010).
- ³⁴H. C. Berg and M. Purcell, Biophys. J. **20**, 193 (1977).
- ³⁵M. Slutsky and L. Mirny, Biophys. J. 87, 4021 (2004).
- ³⁶M. Coppey, O. Bénichou, R. Voituriez, and M. Moreau, Biophys. J. 87, 1640 (2004).
- ³⁷D. Holcman and I. Kupka, "Some questions related to modeling in cellular biology," J. Fixed Point Theory Appl. 7(1), 67–83 (2010).
- ³⁸M. Doi and S. F. Edwards, *The Theory of Polymer Dynamics* (Clarendon, Oxford, 1986).
- ³⁹A. Amitai, Y. Kantor, and M. Kardar, Phys. Rev. E 81, 011107 (2010).
- ⁴⁰A. Singer and Z. Schuss, "Activation through a narrow opening," SIAM J. Appl. Math. 68(1), 98–108 (2007).
- ⁴¹H. G. Bohr, Handbook of Molecular Biophysics: Methods and Applications (Wiley VCH, 2009).
- ⁴²M. Stevens, Biophys. J. 80, 130 (2001).
- ⁴³J. Langowski, Eur. Phys. J. E 19, 241 (2006).
- ⁴⁴G. van den Engh, R. Sachs, and B. J. Trask, Science **257**, 1410 (1992).
- ⁴⁵C. Bustamante, S. B. Smith, J. Liphardt, and D. Smith, Curr. Opin. Struct. Biol. **10**, 279–285 (2000).
- ⁴⁶P. J. Hagerman, Annu. Rev. Biophys. Biophys. Chem. **17**, 265–86 (1988).
- ⁴⁷T. Nguyen, S. Lee, K. Na, S. Yang, J. Kim, and E. Yoon, Nanotechnology 21, 075101 (2010).
- ⁴⁸M. R. Gartenberg, F. R. Neumann, T. Laroche, M. Blaszczyk, and S. M. Gasser, Cell **119**, 955–967 (2004).
- ⁴⁹See http://www.cgl.ucsf.edu/chimera for Chimera software.