

End-product Inhibition of Photosynthesis in *Prunus cerasus* L. in Response to Whole-plant Source-Sink Manipulation

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Abstract. The source-sink ratio of 1-year-old, potted 'Montmorency' sour cherry (*Prunus cerasus*) trees was manipulated by partial defoliation (D) or continuous lighting (CL) to investigate the phenomenon of end-product inhibition of photosynthesis. Within 24 hours of D, net CO₂ assimilation rate (A) of the most recently expanded source leaves of D plants was significantly higher than nondefoliated (control) plants throughout the diurnal photoperiod. Between 2 and 7 days after D, A was 30% to 50% higher and stomatal conductance rate (g_s) was 50% to 100% higher than in controls. Estimated carboxylation efficiency (k) and ribulose-1,5-bisphosphate (RuBP) regeneration rate increased significantly within 2 days and remained consistently higher for up to 9 days after D. Leaf starch concentration and dark respiration rate decreased but sorbitol and sucrose concentration increased after D. The diurnal decline in A in the afternoon after D may have been due to feedback inhibition from accumulation of soluble carbohydrates (sucrose and sorbitol) in the cytosol. This diurnal decline indicated that trees were sink limited. By 9 days after D, photochemical efficiency was significantly higher than in control plants. In the long term, leaf senescence was delayed as indicated by higher A and g_s in combination with higher chlorophyll content up to 32 days after D. CL resulted in a significant reduction of A, g_s, k, variable chlorophyll fluorescence (F_v), photochemical efficiency, and estimated RuBP regeneration rate of the most recently expanded source leaves within 1 day. During the exposure to CL, A was reduced 2- to 3-fold and k was reduced up to 4-fold. The normal linear relationship between A and g_s was uncoupled under CL indicating that A was not primarily limited by g_s and since internal CO₂ concentration was not significantly affected, the physical limitation to A imposed by the stomata was negligible. The decrease in F_v and photochemical efficiency indicated that leaves were photoinhibited within 1 day. The decrease in instantaneous chlorophyll fluorescence after at least 1 day of CL indicated that there was a reversible regulatory mechanism whereby the damage to photosystem II reaction centers was repaired. Leaf chlorophyll content was not altered by 1, 2, or 3 days of exposure to CL, indicating that photooxidation of chlorophyll did not occur. The time to full photosynthetic recovery from CL increased as the duration of exposure increased. CL plants that were photoinhibited accumulated significant starch in the chloroplast in a companion study (Layne and Flore, 1993) and it is possible that an orthophosphate limitation in the chloroplast stroma was occurring. D plants that were continuously illuminated were not photosynthetically inhibited. After 7 days of CL, plants that were then partially defoliated yet remained in CL photosynthetically recovered within 5 days to pre-CL values. Under the conditions of this investigation, end-product inhibition of A occurred in young, potted sour cherry trees but the mechanism of action in D plants was different than in CL plants.

Plant growth and productivity may be limited by the production or use of photosynthetic carbon compounds (photoassimilates). Sucrose and starch are the primary end-products of leaf photosynthesis in most higher plants, where sucrose is the main translocated carbohydrate and starch is a temporary storage carbohydrate that accumulates in the chloroplast. Sorbitol, a sugar alcohol, is an additional important translocated form of photoassimilate in *Prunus* species (Loescher, 1987). The capacity of sink tissues to use

photoassimilate is referred to as sink demand or strength, which is the product of sink size and sink activity (Warren Wilson, 1972). Under sink-limiting conditions (Baysdorfer and Bassham, 1985), when source activity exceeds sink demand, photoassimilates may accumulate in the source leaf. Feedback inhibition of sucrose synthesis by sucrose accumulation may account for an inhibition in net CO₂ assimilation rate (A) (Herold, 1980). In addition to sucrose accumulation in the cytosol, starch accumulation in the chloroplast has also been correlated with an inhibition of A (Gucci et al., 1991a, 1991b; Schaffer et al., 1986).

Whole-plant manipulation to alter the source-sink ratio has been used for investigating the mechanisms involved in end-product inhibition of photosynthesis. Whole-plant partial defoliation (D) and continuous lighting (CL) are manipulation techniques that decrease and increase the source-sink ratio, respectively. Although actual sink demand remains unchanged after either treatment, the relative source activity is altered considerably. After D, there are fewer source leaves available to supply the necessary photoassimilate to the sinks so the relative demand per unit available leaf area increases. Often, as a result of D, photosynthetic enhancement of the remaining leaves occurs (Aoki, 1981; Baysdorfer and Bassham, 1985; Layne and Flore, 1992, 1993; Satoh et al., 1977; Tschaplinski and Blake, 1989a; Von Caemmerer

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and Farquhar, 1984; Wareing et al., 1968; Williams and Farrar, 1988). One would not expect end-product inhibition to occur in partially defoliated plants until a sink limited condition occurred. On the other hand, under CL conditions, the same number of source leaves remain but the duration over which photosynthesis can occur is greatly extended. Eventually, source supply will exceed sink demand and photoassimilates will accumulate leading to an end-product inhibition of A (Sawada et al., 1989). Much of the end-product inhibition research conducted has focussed on agronomic and herbaceous plants and, as yet, the definitive mechanisms involved remain to be fully elucidated. This study was conducted to investigate the underlying mechanisms involved in end-product inhibition of a woody plant, namely *P. cerasus*. For a recent review on photoassimilate distribution in *Prunus*, see Flore and Layne (1995).

In an earlier report (Layne and Flore, 1993), we documented some of the responses of young, potted sour cherry trees to D and

CL. In this paper, we report on five experiments that were conducted to determine how the biochemical and photochemical aspects of photosynthesis were affected in response to D and CL as it relates to end-product inhibition. Since diurnal fluctuations in gas exchange occur in many plant species, we conducted the first experiment to determine whether source-sink manipulation effects were stable throughout the day. We also wanted to determine if A was primarily limited by stomatal conductance rate (g_s) and whether the effect of CL could be reversed by D. In the second experiment, we investigated the early and long-term responses to D. We monitored diurnal gas exchange to determine if trees grown under constant light intensity in a growth chamber would demonstrate a diurnal rhythm of A and how D might affect that rhythm. Also, we wanted to determine what specific factors limit A in the short- and long-term and how D might bring about a photosynthetic rejuvenation in existing leaves. From Expt. 2 it was noted that dark respiration rate (R_d) decreased after D. The third experi-

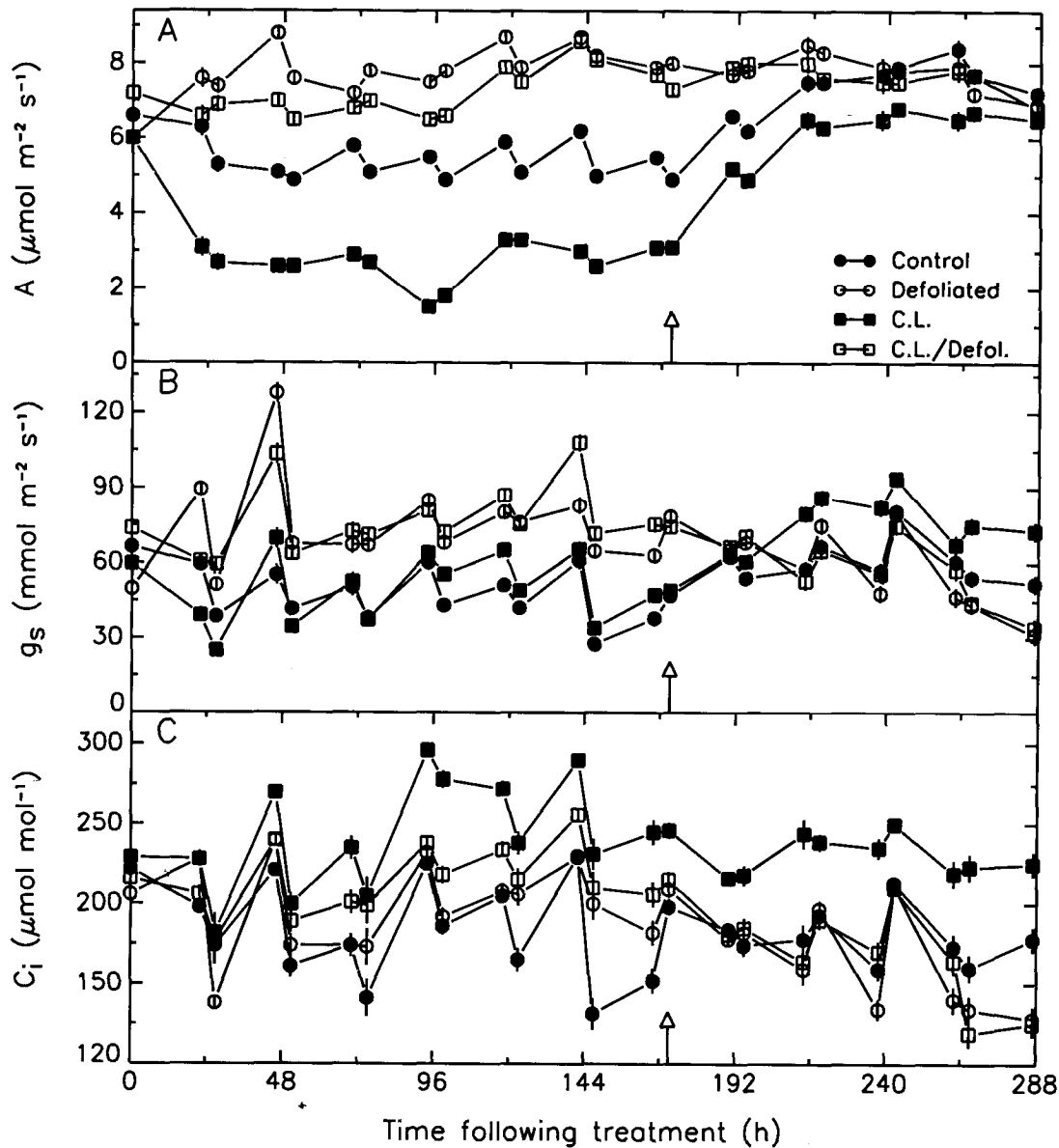


Fig. 1. Effect of partial defoliation (D) or continuous lighting (CL) on diurnal (A) net CO_2 assimilation rate (A), (B) stomatal conductance rate (g_s), and (C) internal CO_2 concentration (C_i) over time for 1-year-old, potted sour cherry trees. Treatments: control = 14-h photoperiod, nondefoliated (\bullet); D = 14-h photoperiod (\circ); CL = continuously illuminated, nondefoliated (\blacksquare); and CL/D = continuously illuminated, defoliated (\square). Gas exchange was measured at 1200 HR for 0 and 288 h after treatment and at 1000 and 1500 HR daily for Days 1–11. Control plants were defoliated on Day 7 after the 1500 HR measurement (note arrow). Each point represents the average (\pm SE) of 16 leaves.

ment was conducted to determine how D affected R_n and whether leaves became substrate-limited for respiration during the night. In addition, leaf carbohydrates were monitored to determine how carbohydrate partitioning changed after D. The fourth experiment was conducted to determine the effect of CL on diurnal gas exchange and several specific parameters in the short term. The effect of CL on photochemical events and the time to full photosynthetic recovery after CL were examined in experiment five to determine if photosynthetic machinery was damaged by CL. Findings from these experiments have been used to broaden our understanding of end-product inhibition of A in *Prunus cerasus*.

Materials and Methods

Experiment 1: Diurnal gas exchange responses to source manipulation

Forty dormant 1-year-old sour cherry trees (*P. cerasus* 'Montmorency' on *P. mahaleb* rootstock) were planted in 11-liter plastic pots with 9.5 liters of sterilized greenhouse soil mix [5 sandy loam

:3 sphagnum peat: 2 torpedo sand (by volume), pH = 7.0]. All trees were cut to an active bud (0–10 cm above the bud union) and placed in an environmentally controlled greenhouse (day and night average temperatures were 28 and 23°C, respectively). During tree growth, lateral shoots were removed as they developed to ensure plant uniformity in terms of sink strength and plant size. Trees were watered to runoff every 3 days and Peter's soluble 20N–20P–20K fertilizer (500 ppm) was applied every 3 weeks. Pesticides [5-O-demethylavermectin (abamectin, Avid), cyano(4-fluoro-3-phenoxyphenyl)methyl-3-(2,2-dichloroethenyl)-2,2-dimethylcyclopropanecarboxylate (cyfluthrin, Decathlon), and d-(2-chlorophenyl)-d-(4-chlorophenyl)-5-pyrimidine-methanol (fenarimol, Rubigan) were applied as necessary according to label rates.

After 9 weeks of active growth in the greenhouse, leaf number and total leaf area were determined for each plant. Each plant had about 20–25 fully expanded leaves at this time and new leaves were still emerging. Gas exchange was measured on the four most-recently fully expanded, source leaves per plant using a portable photosynthesis system (model LCA2; Analytical Development

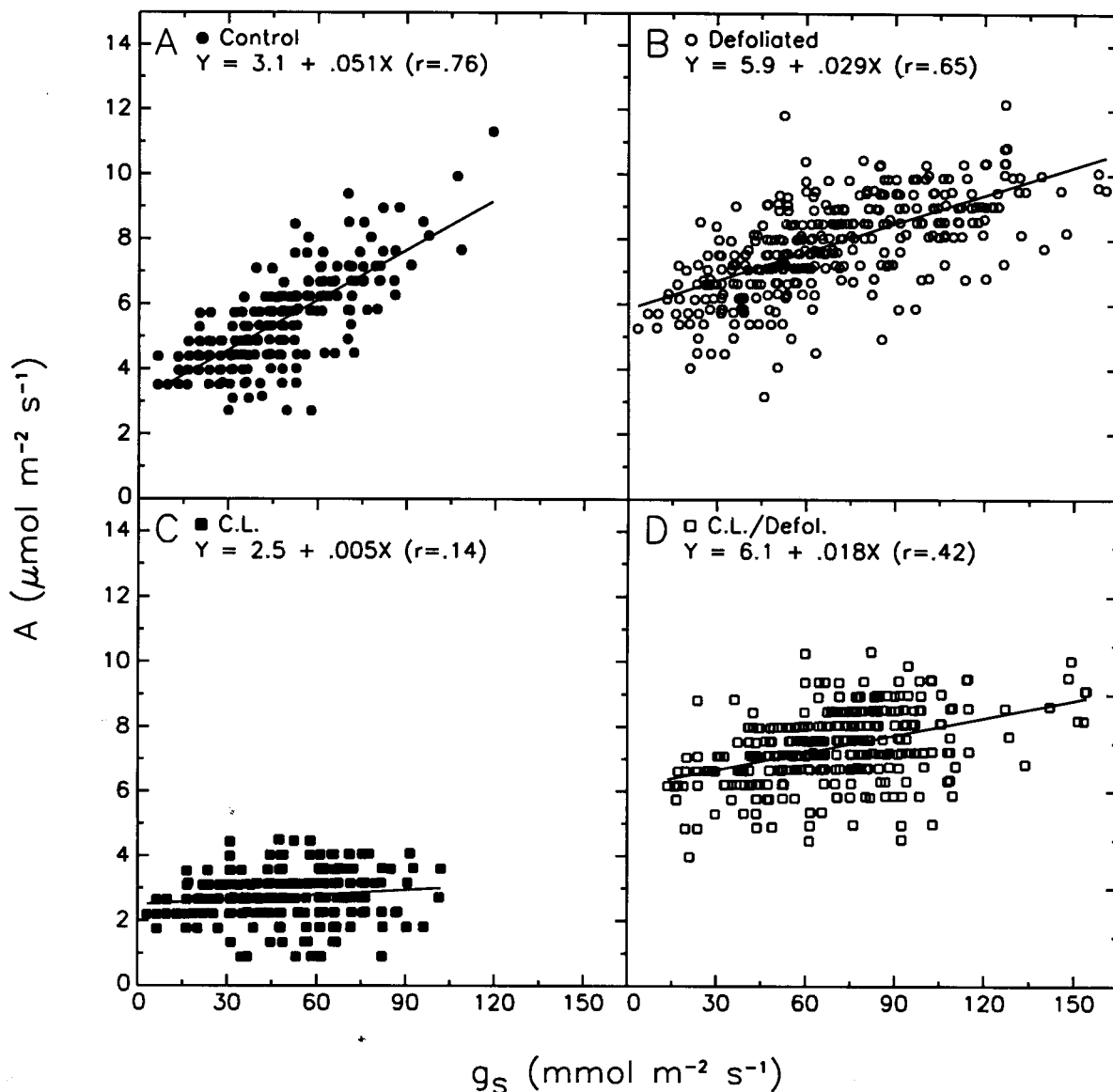


Fig. 2. Effect of partial defoliation (D) or continuous lighting (CL) on the relationship between net CO₂ assimilation rate (A) and stomatal conductance rate (g_s) overtime for 1-year-old, potted sour cherry trees. Treatments: control = 14-h photoperiod, nondefoliated (●); D = 14-h photoperiod (○); CL = continuously illuminated, nondefoliated (■); and CL/D = continuously illuminated, defoliated (D). Linear regressions appear about the raw data. Data for (A) and (C) are for Days 1–7 after treatment. Data for (B) and (D) are for Days 1–2 after treatment.

Co., Hoddesdon, U. K.) under the following conditions: flow rate = 400 ml·min⁻¹, leaf temperature = 30 ± 2C, vapor pressure deficit (VPD) = 3 kpa, ambient CO₂ = 350 ± 10 μl·liter⁻¹, and photosynthetically active radiation (PAR) > 1000 μmol·m⁻²·s⁻¹. The 16 most uniform plants were selected based on total leaf area and initial gas-exchange characteristics and then were moved from the greenhouse to a walk-in plant growth chamber (model PGV36; Conviron Systems of America, Pembina, N.D.). The growth chamber was programmed to the following settings: 0700 HR, air temperature = 28C, relative humidity (RH) = 50%, PAR = 800 μmol·m⁻²·s⁻¹; 2100 HR, air temperature = 23C, RH = 50%, PAR = 800 μmol·m⁻²·s⁻¹.

Plants were randomly assigned to one of each of the following treatments: 1) control = nondefoliated, 14-h photoperiod; 2) D = partially defoliated, 14-h photoperiod; 3) CL = 24 h photoperiod, nondefoliated; and 4) CL/D = 24 h photoperiod, D. For the D treatment, about 70%–75% of the total leaf area was removed by excising all expanded leaves below the fourth fully expanded leaf from the shoot apex. Plants that received a 14-h photoperiod were moved to the other side of the growth chamber behind a light screen (which reduced PAR to < 2 μmol·m⁻²·s⁻¹; the light compensation point was 14 μmol·m⁻²·s⁻¹) at 2100 HR and then they were returned to the illuminated side at 0700 HR each day. Immediately after the last measurement 7 days after treatment, the control and CL plants were partially defoliated as described above.

Statistical design. Plants were arranged in a completely random-

ized design. There were four replicate plants per treatment and the four most recently fully expanded leaves were measured per plant (16 measurements per treatment). Analysis of variance was conducted for treatment comparisons at each date. Linear regression analysis was used to determine the relationship between A and g_s. The PC version of SAS (Cary, N.C.) was used for statistical analysis.

Gas-exchange measurements. Leaf area was determined according to Kappes (1985), where LA = (length × width × 0.671). Gas exchange was measured in the growth chamber with the portable photosynthesis system daily on each of the four leaves per plant under the following conditions (flow rate = 400 ml·min⁻¹, leaf temperature = 32 ± 2C, VPD = 4 kpa, ambient CO₂ = 350 ± 10 μl·liter⁻¹, and PAR = 800 μmol·m⁻²·s⁻¹). On Days 0 and 12 after treatment, gas exchange was measured beginning at 1200 HR. On Days 1–11, gas exchange was measured beginning at 1000 HR and then again beginning at 1500 HR for each of the four leaves per plant. Gas-exchange parameters in all experiments were calculated using the computer program of Moon and Flore (1986).

Experiment 2: Short- and long-term effects of D

As described above, a similar group of dormant 1-year-old sour cherry trees were grown for 9 weeks in the greenhouse, and leaf number, total leaf area, and initial gas-exchange characteristics were determined for each plant. The 16 most uniform plants were selected based on total leaf area and initial gas-exchange characteristics. Selected plants were then moved from the greenhouse to

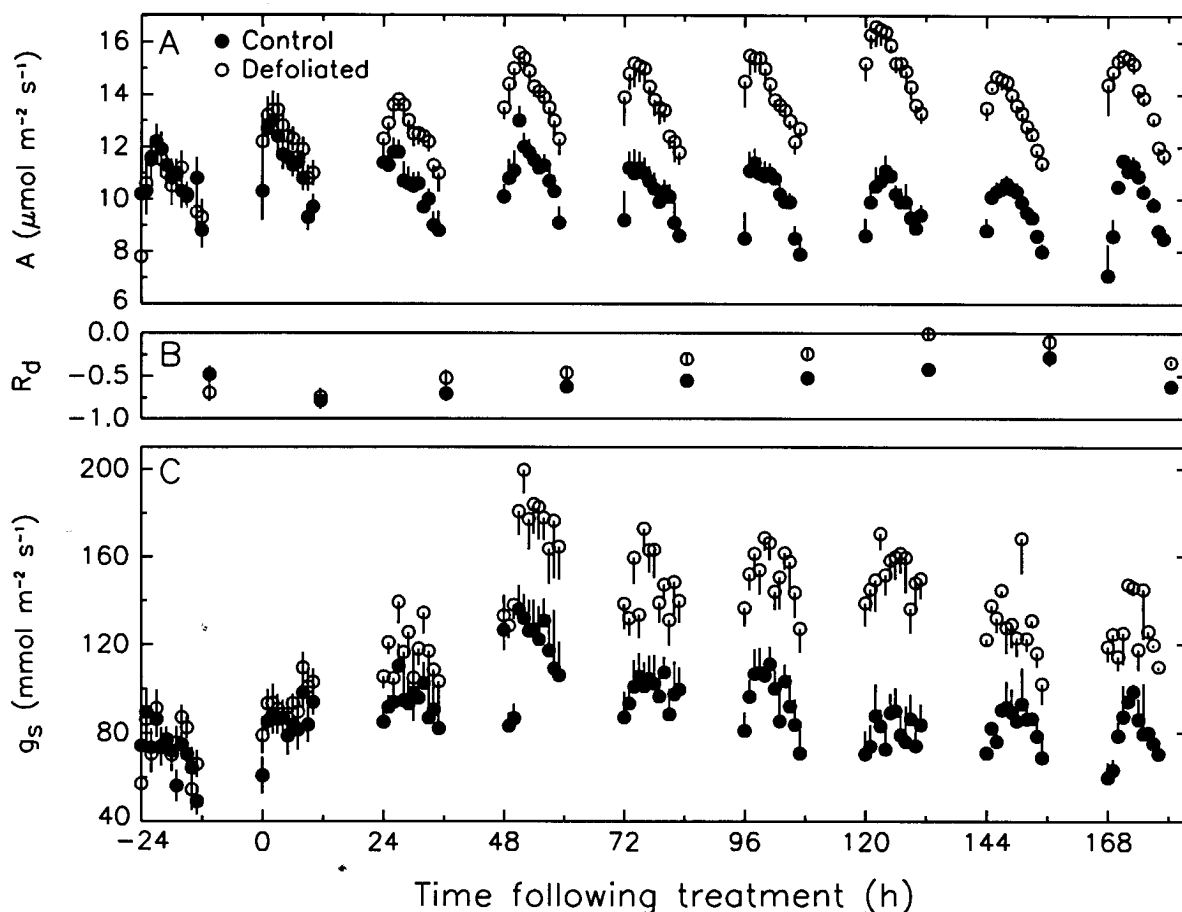


Fig. 3. Time course for the diurnal response of (A) net CO₂ assimilation rate (A), (B) dark respiration rate (R_d, expressed as negative CO₂ assimilation, same units as for A), and (C) stomatal conductance rate (g_s) to partial defoliation in 1-year-old, potted sour cherry trees. Treatments: control = non defoliated (●) and defoliated = partially defoliated plant (○). A and g_s were measured hourly from 0800 to 2000 HR and R_d was measured at 2130 HR daily. Each point represents the average (+, -, or ±SE) of six leaf measurements.

the walk-in plant growth chamber that was programmed as above, except that, at 2100 HR, PAR = 0 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$. D was accomplished as described above.

Gas-exchange measurements. Whole-leaf gas exchange was measured using the open gas-exchange system described by Sams and Flore (1982) and modified as follows: a) an infrared gas analyzer (model 225 MK3; Analytical Development Co.) was used to measure differential CO_2 concentrations at the inlet and outlet of the leaf chambers and b) air flow entering the chambers was regulated using the following Matheson Instruments (Horsham, Pa.): 8100 series flowmeters and 8200 series mass flow controllers connected to a model 8219 multichannel Dyna-Blender. Ambient CO_2 concentration was measured using a portable infrared gas analyzer and O_2 concentration was determined with an oxygen analyzer (model 0-260; Beckman Instruments, Irvine, Calif.). Trees to be measured were moved to an identical walk-in plant growth chamber (set to the same conditions as described above), which was equipped with the assimilation chambers. Unless otherwise indicated, gas-exchange measurements were conducted under the following room conditions: air temperature of 28C, 50% RH, PAR = 1000 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$. Four whole-leaf chambers were used simultaneously in which the following conditions were maintained, unless otherwise indicated: inlet CO_2 and O_2 , 350 \pm 15 $\mu\text{l}\cdot\text{liter}^{-1}$ and 21 kpa, respectively; leaf temp. 29 \pm 1C; VPD 1.0 \pm 0.2 kpa; inlet flow rate 3 liters $\cdot\text{min}^{-1}$.

Diurnal gas exchange. On three separate dates (three replicates over time) four plants were selected and two treatments were imposed on two plants each: 1) nondefoliated (control) and 2) D. The fourth fully expanded leaf of each plant was inserted into an individual leaf chamber (where it remained for the duration of the experiment). The day before D was designated as Day 0. On Day 1, the plants were partially defoliated at 0800 HR. Gas exchange was measured hourly during the photoperiod from 0800 to 2000 HR and then R_d was measured at 2130 HR after a 30 min exposure to darkness on Days 0-8. On Day 9, a response to photosynthetic

photon flux density (PPFD) was measured (as described below). Plants were then returned to the first walk-in growth chamber that was described above. The three replicates of this experiment were analyzed first by analysis of variance to determine if there was an effect of time on the responses measured. Since there was no significant difference in the responses of plants from the three replicates over time, the data from the replicates were pooled for subsequent statistical analyses.

Long-term effects. After the measurements of diurnal gas exchange and response to PPFD for the three replicates were completed, gas exchange was measured on the same four leaves of all twelve plants with the portable photosynthesis system. Since the experiment was replicated three times over time, the plants from each replicate had three different recovery durations. Leaf areas were measured for all leaves. For each plant, leaves 1-4 were sampled for chlorophyll (as described below) and then leaves were dried in an oven at 80C for 7 days so that specific leaf density (SLD) could be determined (dry weight/unit area).

Chlorophyll determination. Four discs (0.324 cm^2 each) were punched from the middle of the lamina (two on either side of the midrib) of each leaf using a paper holepunch. Fresh weight of discs was recorded and then chlorophyll was extracted in 8 ml n,n-dimethylformamide in darkness at 5C for 36 h. Absorbance of extracts was read at 664, 647, and 625 nm on a UV/Vis spectrophotometer (model U-3110; Hitachi, Tokyo). Calculations for chlorophyll *a*, chlorophyll *b*, and protochlorophyll (P) were made according to Moran (1982).

Response to CO_2 and low O_2 . From the same population of 16 plants, the 4 remaining plants were brought to the growth chamber in the laboratory. Gas exchange was measured on Days 0-6 and Day 9 after D, as described above. Carbon dioxide and low O_2 response were measured daily between 0930 and 1430 HR (as described below). All four plants were defoliated immediately after the last gas-exchange measurement on Day 0.

Gas-exchange measurements for changing environmental con-

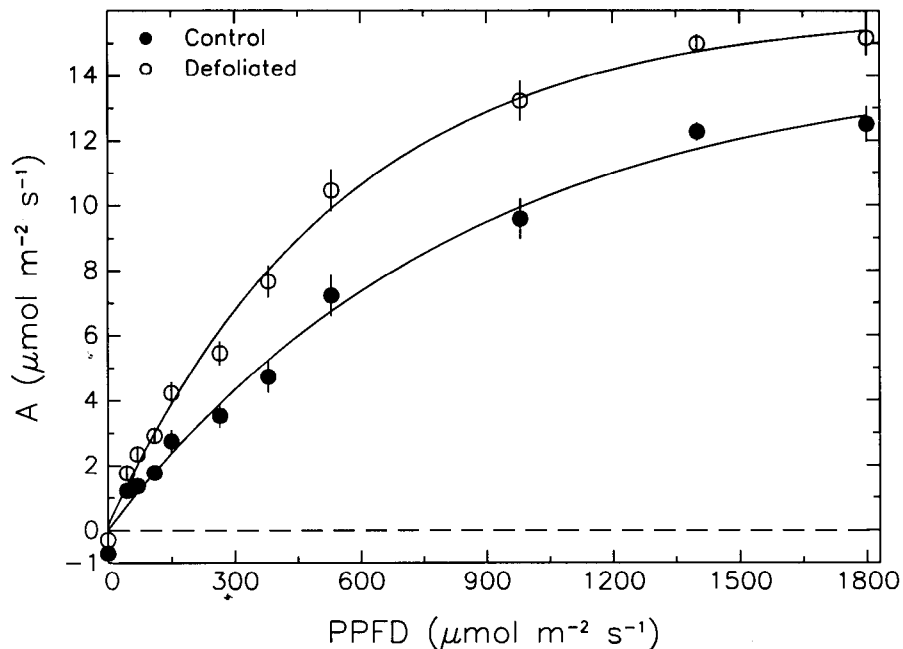


Fig. 4. Effect of partial defoliation on the response of net CO_2 assimilation rate (A) to photosynthetic photon flux density (PPFD) 9 days after treatment for 1-year-old, potted sour cherry trees. Treatments: control = nondefoliated (●) and defoliated = partially defoliated plant (○). Regression lines occur about the data. Data points that represent the average (\pm SE) of eight leaf measurements appear about the regressions.

ditions. Response to PPFD was determined by decreasing PPFD in a stepwise manner (15-min acclimation at each step allowed significant time for stomata to adjust) to the following levels 1800, 1400, 980, 530, 380, 265, 150, 110, 70, 45, and 0 $\mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$. Leaf temperature was maintained constant at all PPFD levels. Supplement light to obtain higher PPFDs in the growth chamber was provided by 400-W metal halide lamps. PPFD levels were produced using a combination of neutral density filters. PAR was measured with a recently calibrated PAR sensor (LI-190; LICOR, Lincoln, Neb.).

Photorespiration rate (R_p) was estimated by comparing A at ambient O_2 (21 kPa) with that at reduced O_2 (1.5 kPa). Response to CO_2 was determined by increasing the CO_2 concentration

stepwise to the following levels: 70, 100, 150, 200, 250, 300, 350, 400, 500, 700, and 900 $\mu\text{l}\cdot\text{liter}^{-1}$. Leaves were allowed to acclimate for 20 min at each CO_2 level, which allowed sufficient time for stomata to adjust.

Responses of A to PPFD and A to internal CO_2 concentration (C_i) were analyzed for each treatment by nonlinear regression. Curve fitting was performed using the Marquardt compromise method of successive approximations. The best fit curve, evaluated by analysis of residuals and r^2 , was the monomolecular asymptotic function (Hunt, 1980) of the type

$$Y = B (1 * [1.0 - B(2) e^{-B(3)*X}])$$

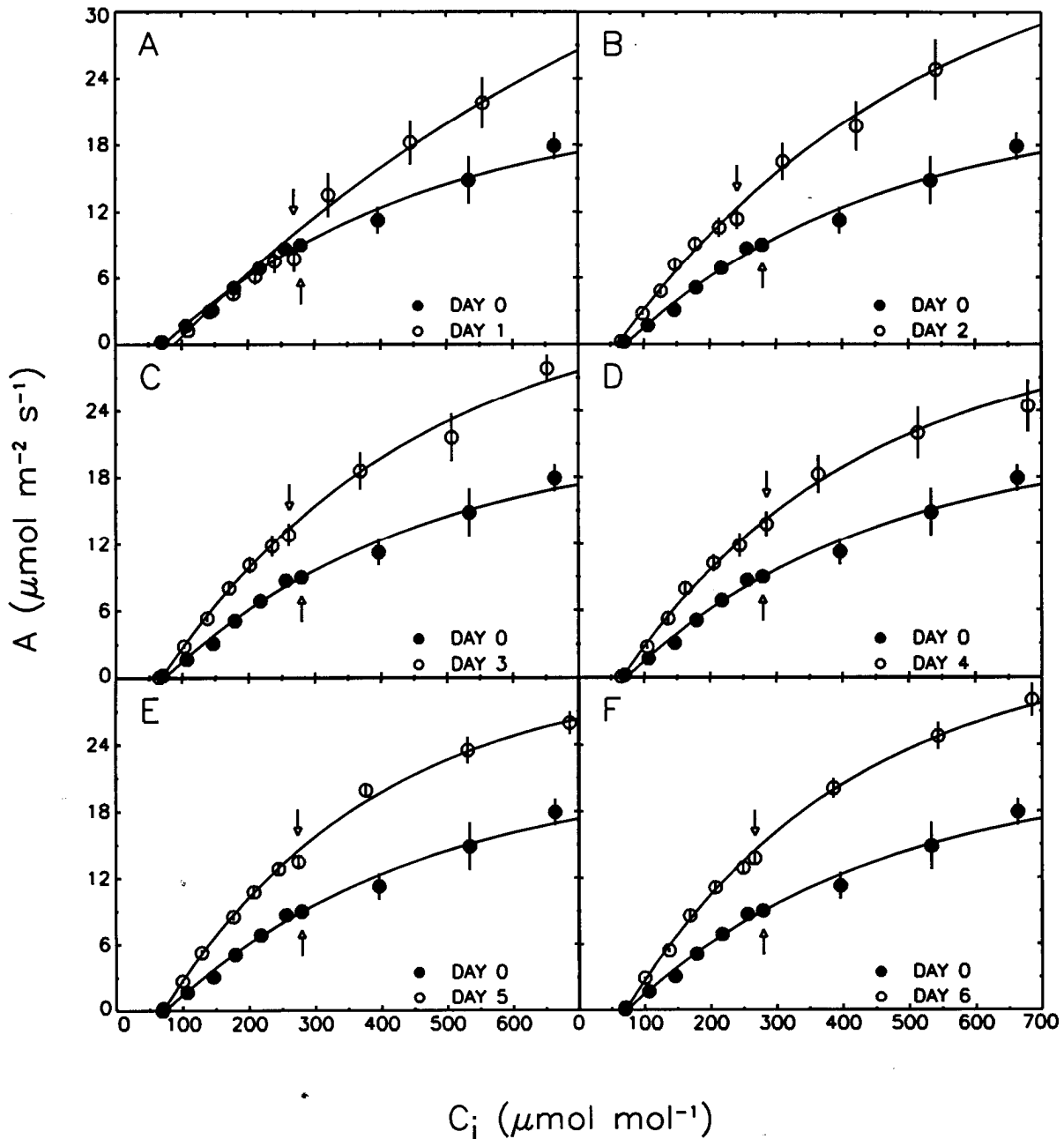


Fig. 5. Effect of partial defoliation (D) on the response over time of net CO_2 assimilation rate (A) to internal CO_2 concentration (C_i) of 1-year-old, potted sour cherry trees. Each section (A-F) compares the Day 0 pretreatment response (■) with the treatment response for the subsequent day (○). Treatments: Day 0 = before D; Days 1 (A), 2 (B), 3 (C), 4 (D), 5 (E), and 6 (F) correspond to 1, 2, 3, 4, 5, and 6 days after D, respectively. Regression lines occur about the data. Arrows point to the data collected at ambient CO_2 partial pressure ($350 \pm 25 \mu\text{l}\cdot\text{liter}^{-1}$). Data points that represent the average (\pm SE) of four leaf measurements appear about the regressions.

where B(1), B(2), and B(3) are the asymptotic value, minimum value and rate constant, respectively. This polynomial was selected because it provided direct estimates of specific physiological processes and it exhibited curvilinear features that represented the data. Individual leaf A vs. PPF and A vs. C_i(AC_i) response curves were developed using this polynomial.

For the A vs. PPF curve, dark respiration rate (R_d) was calculated at 0 μmol·m⁻²·s⁻¹ PPF. Light compensation point (cp) was extrapolated from the curve as the PPF level at which A was 0 μmol·m⁻²·s⁻¹. Photochemical efficiency (φ) was calculated as the slope of the A vs. PPF curve in the linear portion between 0 and 150 μmol·m⁻²·s⁻¹ PPF. From the AC_i response curve, CO₂ compensation point (Γ) was extrapolated as the C_i at which A was 0 μmol·m⁻²·s⁻¹. Estimated carboxylation efficiency (k) was calculated as the slope in the linear portion of the AC_i curve. Stomatal limitation to A (I_g) was calculated according to the differential method of Jones (1985). Maximum net CO₂ assimilation rate (A_{max}) was determined as the A at 900 μl·liter⁻¹ CO₂. Ribulose-1,5-bisphosphate (RuBP) regeneration rate limitations were expressed by a reduced A at saturating CO₂ concentrations (Farquhar and Sharkey, 1982). Linear regression analysis was used to determine photochemical and carboxylation efficiencies from the A vs. PPF and AC_i curves, respectively. Homogeneity of the independent regressions was evaluated by Student's *t* test (Steele and Torie, 1980).

Experiment 3: D effects on R_d and leaf carbohydrate content

As described above, a similar group of dormant 1-year-old sour cherry trees was grown for 9 weeks in the greenhouse, and leaf number, total leaf area, and initial gas-exchange characteristics were determined for each plant. The 15 most uniform plants were selected based on total leaf area and initial gas-exchange characteristics. Selected plants were moved from the greenhouse to the walk-in plant growth chamber in the laboratory. One day before D (designated as Day-1), whole-leaf gas exchange was measured on the four most recently expanded leaves of all 15 plants proceeding from 1000 until 1200 HR. R_d was measured at 2200 HR (after 1 h of darkness), 0200 HR (after 5 h of darkness), and 0600 HR (after 9 h of darkness; 1 h before the lights were turned on again). On Day 0, all 15 plants were partially defoliated between 2100 and 2200 HR. The fifth and sixth fully expanded leaves from the apex were immediately frozen in liquid N₂ and stored at -80C for carbohydrate analysis to provide a Day 0 leaf carbohydrate status for each plant. On Days 0-4, gas exchange was measured beginning at 1000 HR. One of the fifteen plants was destructively harvested at each

night measurement on Days 0-4. The 4 harvested leaves per plant were pooled as one sample, frozen in liquid N₂, and immediately stored at -80C.

Leaf carbohydrate analysis. Stored leaf samples (-80C) were lyophilized, weighed, and ground in a Wiley mill to pass through a 40-mesh screen. Subsamples (100 mg) were extracted three times each (20 rein) with 3.5 ml of 80% ethanol. The homogenates were centrifuged at 1500× g for 5 min after each extraction, supernatants were transferred into 50 ml polypropylene centrifuge tubes, and 5 ml of ultrapure water and 5 ml of reagent grade chloroform were added to partition the chlorophyll from the sample extract. The tubes were capped, shaken vigorously, and then centrifuged at 470x g for 3 min. The upper, clear aqueous phase containing the soluble sugars was transferred to a test tube and evaporated to dryness using a rotary evaporator equipped with a refrigerated condensation trap (model RT4104) at -103C (Speedvac SC200; Savant Instruments, Farmingdale, N.Y.). The pellet was also evaporated to dryness and it, along with the soluble sugar samples, was stored in a glass desiccator over an anhydrous desiccant.

Sorbitol, glucose, fructose, and sucrose were separated and quantified by high-performance liquid chromatography (HPLC) (series 4000i system equipped with a gradient pump, CarboPac PA1 column, autosampler, and pulsed amperometric detector connected to a model 4270 integrator; Dionex Corp., Sunnyvale, Calif.). The column was eluted with ultrapure water and 200 mM NaOH that had been sonicated for 30 rein, degassed with helium for 1 h, and then continuously degassed during the analysis. A combination of gradient and isocratic elution was used by programming the gradient pump as follows: 1) before sample injection, the column was gradient eluted with 65% water and 35% NaOH; 2) at time = 0 min, the sample was injected and the gradient was then adjusted to 75% water and 25% NaOH by time = 0.1 rein, and the elution remained at this ratio until time = 1.5 rein; 3) from time = 1.5 min to 9.0 min the gradient was adjusted to end up at 0% water and 100% NaOH; 4) from time = 9 min to 28 rein, the column was isocratically eluted with 100% NaOH. The flow rate of eluent was constant at 1 ml·min⁻¹. The column was maintained at room temperature.

The dry sample extract was prepared for HPLC analysis by adding 1 ml of ultrapure water, vortexing three times, each followed by at least 5 min to stand. The sample was then transferred to a 1.5-ml Eppendorf tube and spun at 10000× g for 10 min on a microfuge (Beckman Instruments,) to pellet any particulate matter that remained. The sample was then diluted 1:100 with ultrapure water, and a 0.5-ml aliquot was loaded into a sample injection vial.

Table 1. The effect of partial defoliation (D) on CO₂ compensation point (Γ), estimated carboxylation efficiency (k), net CO₂ assimilation rate (A₃₅₀) and internal CO₂ concentration (C_{i350}) at ambient CO₂ (350 μl·liter⁻¹), maximum A at 900 μl·liter⁻¹ CO₂ (A_{max}), stomatal limitation to A (I_g), and estimated photorespiration rate (R₁) of expanded leaves of 1-year-old, potted sour cherry trees with time.

Days after D	Γ (μmol CO ₂ /mol)	k (mol CO ₂ /m ² per sec)	A ₃₅₀ (μmol CO ₂ /m ² per sec)	C _{i350} (μmol CO ₂ /mol)	A _{max} (μmol CO ₂ /m ² per sec)	I _g (%)	R ₁ (%)
0	68.5	0.044	8.98	279	17.9	20.6	30.2
1	69.5	0.042 ^{NS}	7.71 ^{NS}	270 ^{NS}	21.8*	35.0***	27.3
2	60.5	0.083***	11.35**	242***	24.9***	36.2***	27.7
3	66.0	0.075***	12.83***	261*	27.8***	27.1**	25.3
4	66.8	0.080***	13.77***	285 ^{NS}	24.4***	19.4 ^{NS}	25.5
5	69.0	0.080***	13.46***	275 ^{NS}	26.0***	22.7 ^{NS}	33.1
6	68.8	0.084***	13.76***	267 ^{NS}	28.0***	25.2*	31.4
9	67.3	0.083***	13.48***	270 ^{NS}	26.0***	23.7 ^{NS}	25.8

^{NS} Means for each day are compared relative to Day 0 by orthogonal contrasts in each column. Means are averages of four leaf measurements.

^{NS}, *, **, ***Nonsignificant or significant at P = 0.05, 0.01, or 0.001, respectively. Absence of notation indicates no significant differences among days.

Each sample was analyzed three times. Soluble carbohydrates were identified and quantified on the basis of retention time and peak heights of known standards.

Starch in the pellet was determined using the method of Roper et al. (1988) modified as follows: samples were incubated at 55°C for 16 h with amyloglucosidase (Boehringer Mannheim 208 469) and then assayed calorimetrically using glucose oxidase (Sigma Chemical Co.). Absorbance at 440 nm was read with a UV/Vis spectrophotometer. Each sample was assayed three times.

Experiment 4: Short-term effects of continuous lighting

As described above, dormant 1-year-old sour cherry trees were grown for 9 weeks in the greenhouse, and leaf number, total leaf area, and initial gas-exchange characteristics were determined for each plant. The 16 most uniform plants were selected based on total leaf area and initial gas-exchange characteristics. Selected plants were moved from the greenhouse to the walk-in plant growth chamber programmed as above. The four most uniform plants were selected for this experiment based on the combined characteristics of leaf number, total leaf area, and initial gas-exchange characteristics. Preliminary findings from Expt. 1 indicated that A of the most recently expanded leaves of control sour cherry trees, identical to those described here, was constant for at least 2 weeks in the growth chamber when trees were at this stage of development. Based on this condition, we made comparisons of the effect of CL on the response observed for the same leaf prior to treatment rather than comparing with separate control plants. This permitted more replications for statistically evaluating the CL effects on the same leaves over time.

Diurnal gas exchange. The fourth fully expanded leaf of each plant was inserted into an individual leaf chamber (where it remained for the duration of the experiment). The day before CL was designated as Day 0. Plants remained in CL from 0700 HR on Day 0 until 2100 HR on Day 4. For Days 5–7, the growth chamber was reset to a 14-h photoperiod. Gas exchange was measured daily at 0900, 1230, 1500, and 2000 HR. CO₂ and O₂ responses were measured daily between 0930 and 1430 HR as described above.

Experiment 5: Time to full photosynthetic recovery after continuous lighting

A similar set of plants was grown for 9 weeks in the greenhouse, and leaf number, total leaf area, and initial gas-exchange characteristics

were determined for each plant. The most uniform 15 plants were selected based on leaf number, total leaf area, and initial gas-exchange characteristics and moved to the growth chamber described above.

Five plants each were randomly selected and subjected to either 1, 2, or 3 days of CL. Recovery was followed for at least 6 days after the CL treatment. Once a given group of five plants had received the appropriate CL treatment, they were moved to the other side of the growth chamber behind a light screen at 2100 HR and then returned to the illuminated side at 0700 HR each day. Once all plants had received their CL treatment, the chamber was reprogrammed to a 14-h photoperiod.

On each day for the duration of the experiment, gas exchange was measured beginning at 1000 HR using the portable photosynthesis system and chlorophyll fluorescence was measured beginning at 1500 HR (as described below). All measurements were made on the four most recently fully expanded leaves per plant. Each day after the chlorophyll fluorescence measurement, one leaf disc from each of the four leaves per plant was harvested for chlorophyll determination as described previously.

Chlorophyll fluorescence. A chlorophyll fluorescence measurement system (model CF-1000; P.K. Morgan Instruments, Andover, Mass.) was used. Leaves were dark acclimated for at least 15 min using dark acclimation cuvettes. Leaves were irradiated with 1000 μmol·m⁻²·s⁻¹ PPFD actinic light and chlorophyll fluorescence kinetics were recorded over a 60-sec sample time.

Results

Experiment 1

Gas exchange over time. One day after treatment, A was 2770 higher and 48% lower than control for D and CL plants, respectively (Fig. 1A). The g_s was 80% higher and 35% lower for D and CL plants, respectively (Fig. 1B). By 2 days after treatment, A was 47% higher and 57% lower than control for D and CL plants, respectively. The g_s was significantly higher in D plants (either 14- or 24-h photoperiod) than in control or CL plants (Fig. 1B). C_i was significantly greater for CL plants at most times of day from Day 2 on (Fig. 1C). Between 2 and 7 days after treatment, leaves from D and CL/D plants had significantly higher A and g_s than controls, while CL plants had significantly lower A. The photosynthetic reduction after CL was maintained as long as plants were under CL

Table 2. Long-term effects of partial defoliation (D) on net CO₂ assimilation rate (A), stomatal conductance rate (g_s), leaf chlorophyll *a*, chlorophyll *b*, and protochlorophyll (P) content, and specific leaf density (SLD) of expanded leaves of 1-year-old, potted sour cherry trees.^a

Treatment	A (μmol CO ₂ /m ² per sec)	g _s (mmol CO ₂ /m ² per sec)	Chlorophyll <i>a</i> (μg·cm ⁻²)	Chlorophyll <i>b</i> (μg·cm ⁻²)	P (μg·cm ⁻²)	SLD (mg·cm ⁻²)
<i>11 Days after D</i>						
Control	10.4	75.3	43.9	18.9	9.9	9.8
D	13.5	94.0	44.7	19.4	10.0	8.7
Significance	*	*	NS	NS	NS	NS
<i>23 Days after D</i>						
Control	10.1	68.3	49.5	21.6	10.2	8.8
D	14.1	137.0	50.0	21.1	10.3	9.0
Significance	**	***	NS	NS	NS	NS
<i>32 Days after D</i>						
Control	7.9	48.9	28.9	14.8	9.1	8.9
D	13.3	91.3	44.7	19.7	9.9	9.5
Significance	**	**	**	**	**	NS

^aMeans are averages of four replicates.

^{NS}, *, **, *** Nonsignificant or significant at *P* = 0.05, 0.01, or 0.001, respectively.

conditions. A , g_s and C_i of control plants fluctuated diurnally and were higher at 1000 than at 1500 HR most days. After 7 days, control and CL plants were D as described above. A and g_s steadily increased for these plants during the 5-day postdefoliation (recovery) period. C_i during this recovery phase for CL plants that were D remained fairly constant but significantly higher than the other treatments (Fig. 1C). By 12 days after the initiation of the experiment, A was not significantly different among the treatments. The photosynthetic inhibition from CL was completely ameliorated 5 days after D.

Relationship between A and g_s . Since plants from control and CL treatments were partially defoliated by Day 8, only the data for Days 1-7 were used for the regression analysis (Fig. 2 A and C). On the other hand, since plants from both the D and CL/D treatments were manipulated on Day 1 only, the data for Days 1-12 were used for the regression analysis (Fig. 2 B and D). From the regression analysis, the relationship between A and g_s over time was significant (at $P = 0.05$) for all treatments except CL. The strongest relationship between A and g_s was noted for leaves of control plants indicating that these leaves had the highest A per unit g_s (Fig. 2A). CL led to a significant uncoupling of the relationship between A and g_s (Fig. 2 C and D). An analysis of covariance was performed to determine if the regression coefficients were homogeneous for the four treatments. Based on the appropriate F test, the regressions were not homogeneous.

Experiment 2

Diurnal gas exchange. A pronounced diurnal response of A and g_s was observed for control and D plants in which A typically reached a maximum value at 1100 HR and then declined in the afternoon (Fig. 3 A and C). One day after treatment, A and g_s were significantly higher in the D plants at most times during the day. Between 2 and 7 days after D, A was 30% to 50% higher and g_s was 50% to 100% higher in leaves of D plants than in leaves of control plants. R_d was significantly lower (less negative) in leaves of D plants from 2 days after D (Fig. 3B).

Response to PPFD. A was significantly higher at each light intensity for D plants 9 days after treatment (Fig. 4). R_d was -0.7 VS. $-0.3 \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ for the control and D plants, respectively. The cp was 14 vs. $6 \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ for the control and D plants, respectively. The ϕ was 67% higher in leaves of D plants than in control plants (0.0226 vs. $0.0135 \text{ mol CO}_2 \text{ fixed/mol PPFD absorbed}$). Light saturation occurred near $1000 \mu\text{mol}\cdot\text{m}^{-2}\cdot\text{s}^{-1}$ PPFD.

Response to CO_2 and O_2 . A highly significant relationship was observed between A and C_i for each date of measurement ($r^2 = 0.96$, on average) (Fig. 5). The response of leaves 9 days after D was almost identical to that observed 6 days after D (Fig. 5F) and was not included in this figure. By 2 days after D, A was significantly higher at all CO_2 levels. (except $70 \mu\text{l}\cdot\text{liter}^{-1}$) and this remained constant through Day 9 (data not shown). Γ and R_i were not significantly affected by D (Table 1). Two days after D, k was 89% higher than it was before D and it remained significantly higher on each subsequent day of measurement (Table 1). By 2 and 4 days after D, net CO_2 assimilation at ambient CO_2 (A_{350}) was 26% and 53% higher, respectively, than it was before D (Table 1). By 2 and 3 days after D, internal CO_2 concentration at ambient CO_2 (C_{i350}) was reduced compared to the predefoliation values but the differences were not significant by Day 4 (Table 1). A at saturating CO_2 concentrations (Fig. 5) and A_{max} were significantly higher after D (Table 1). A_{max} was 56% higher by 6 days after D (Table 1). The I_g increased slightly between 1 and 3 days after D, but this was ameliorated thereafter (Table 1).

Long-term effects. A and g_s remained significantly higher in

leaves of D plants than in controls up to 32 days after D (Table 2). Chlorophyll a , chlorophyll b , and P were not significantly higher for leaves of D plants until 32 days after D. SLD was not significantly affected by D.

Experiment 3

Respiration and leaf carbohydrate content. Leaf A increased by 30% 1 day after D (Fig. 6A). By 3 or 4 days after treatment, A had almost doubled from the predefoliation value. R_d progressively decreased (became less negative) after D (Fig. 6B). R_d fluctuated during the dark period and was not consistently higher or lower at any given time during the night after D. Leaf starch content was significantly lower at the beginning of the night 48 h after D than

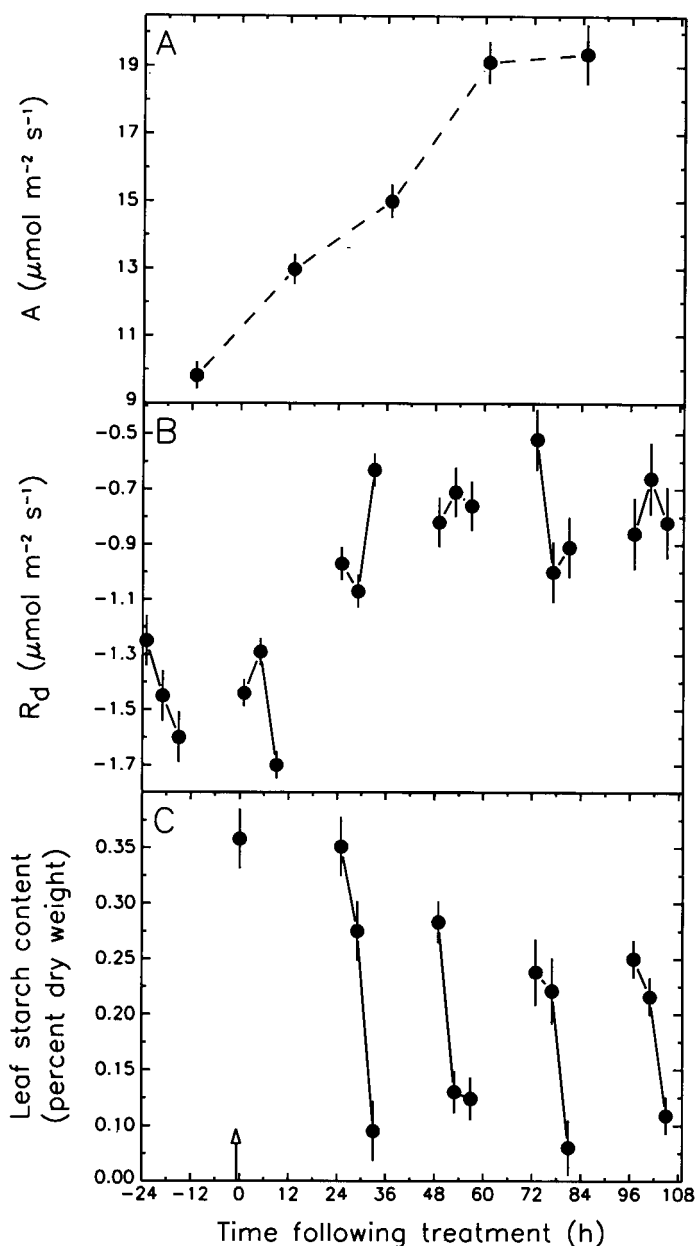


Fig. 6. Effects of partial defoliation (D) on (A) net CO_2 assimilation rate (A), (B) dark respiration rate (R_d , expressed as negative CO_2 assimilation), and (C) leaf starch content over time for 1-year-old, potted sour cherry trees. A was measured daily at 1000 HR. R_d was measured daily at 2200, 0200, and 0600 HR. The arrow indicates the time at which plants were partially defoliated. Each point represents the average (\pm SE) of at least four leaf measurements.

it was 24 h earlier (Fig. 6C). Starch content was reduced by almost 40% 4 days after treatment. Starch content was typically highest and lowest at the beginning and the end of the dark period, respectively. By 0600 HR, leaf starch was usually < 0.1% of the leaf dry weight. There was a significant correlation between leaf starch content and R_d ($r = -0.632$), where leaf starch content decreased as R_d decreased (data not shown). This reduction in R_d after D was consistently observed in four separate experiments (data not shown).

Leaf sorbitol and sucrose content increased over time after D (Table 3). Sorbitol and sucrose content was always highest and lowest at the beginning and end of the dark period, respectively. On average, soluble carbohydrates comprised about 20% of the total leaf dry weight. The relative proportions for sorbitol, glucose, fructose, and sucrose were 45%, 5%, 4%, and 46%, respectively. D did not significantly alter the proportionate amounts of these four major soluble carbohydrates.

Experiment 4

Diurnal gas exchange. A decreased >2-fold after 1 day of CL (Fig. 7A). The g_s was significantly reduced by CL except at the end of Days 1 and 2 (Fig. 7B). The normal diurnal response of higher A and g_s in the morning and lower A and g_s in the afternoon disappeared under CL conditions (Fig. 7 A and B). Between 24 and 48 h of CL, there was a highly significant variation in g_s and C_i that was dampened somewhat during the following 48 h (Fig. 7 B and C). Returning plants to a 14-h photoperiod for 3 days was not long enough to overcome the inhibition of A resulting from 3 days of CL.

Response to CO_2 and O_2 . A was lower at each CO_2 level (except 70 and 900 $\mu\text{l}\cdot\text{liter}^{-1} CO_2$) each day in CL (Fig. 8 A–C) and during each of the 3 days after return to a 14-h photoperiod (Fig. 8 D–F). The relationship between A and C_i was not as strong (based on r^2 values for the regression functions) after CL. Γ was not altered by CL (Table 4). The k was reduced by almost 4-fold after 1 day of CL and remained significantly lower (by 2-fold) even 3 days after return to a 14-h photoperiod. A_{350} was reduced by >50% after 3 days of CL. C_{i350} was higher 1 day after CL but then not different from the pretreatment value on each subsequent day. At supraambient CO_2 levels (above 350 $\mu\text{l}\cdot\text{liter}^{-1}$), A was dramatically

reduced for CL trees (Fig. 8 A–C). By 2 and 3 days after CL, the stomata began to present an increased limitation to A, but this was relieved after return to a 14-h photoperiod (Table 4). R_d averaged about 35% for leaves and was not significantly affected by CL (data not shown).

Experiment 5

Time to full photosynthetic recovery. After 1 day of CL, A was significantly reduced in all trees (data not shown). Exposure to 1, 2, or 3 days of CL required 2, 4, or 5 days, respectively, before A of those same leaves recovered to the rate observed before treatment (data not shown).

Chlorophyll fluorescence. Estimated photochemical efficiency (ratio of variable fluorescence (F_v) to maximal fluorescence (F_m), F_v/F_m) was reduced by CL (Fig. 9A). CL for 1 day inhibited F_v/F_m significantly but full recovery occurred by the next day after return to a 14-h photoperiod. CL for 2 or 3 days resulted in an inhibition of F_v/F_m that took 2 to 3 days for full recovery. F_v was inhibited by CL (Fig. 9B). Instantaneous fluorescence (F_o) decreased after 1 day in CL. By 9 days after CL exposure, F_v and F_o were still significantly reduced relative to the pretreatment levels. The values for F_v/F_m for plants before CL (0.760, on average) are in close agreement with the values determined for other C_3 species (Bjorkman and Demmig, 1987).

change leaf chlorophyll content, or the chlorophylls to b ratio (data not shown). Chlorophyll content increased slightly over time after CL. Protochlorophyll content was not altered by CL and was 1.0 $\mu\text{g}\cdot\text{cm}^{-2}$, on average (data not shown).

Discussion

D reduced the source–sink ratio in sour cherry trees and, as a result, source activity became enhanced. When enhancement of A occurs after D, it indicates that source leaves were operating below their maximum photosynthetic potential (Tschaplinski and Blake, 1989a). In a previous study (Layne and Flore, 1992), we found that the photosynthetic enhancement of individual leaves to leaf area reduction was due to higher estimated photochemical and car-

Table 3. The effect of partial defoliation on soluble carbohydrate content of expanded leaves of 1-year-old, potted sour cherry trees over time.^z

Day	Time (HR)	Soluble carbohydrate content (% dry wt)				
		Sorbitol	Glucose	Fructose	Sucrose	Total
0	2200	6.7	1.1	1.1	8.4	17.3
1	2200	8.4	1.0	0.7	10.3	20.4
1	0200	6.8	1.0	0.9	8.8	17.5
1	0600	6.4	0.9	0.6	7.7	15.6
SE Day 1		0.2	0.06	0.05	0.8	0.9
2	2200	10.7	1.2	1.2	10.2	23.3
2	0200	10.7	0.8	0.7	9.0	21.2
2	0600	7.6	0.7	0.7	7.3	16.3
SE Day 2		0.4	0.01	0.05	0.5	0.3
3	2200	11.6	1.2	0.2	12.0	25.0
3	0200	8.4	1.4	0.2	8.6	18.6
3	0600	8.3	0.6	0.6	6.0	15.5
SE Day 3		0.4	0.01	0.01	0.1	0.2
4	2200	11.4	1.2	1.0	13.1	26.7
4	0200	10.0	1.2	0.6	10.6	22.4
4	0600	9.3	1.2	0.8	8.1	19.4
SE Day 4		0.3	0.04	0.10	0.4	0.4

^zData are means of three subsamples per sample. SEs apply to the means for each day, respectively.

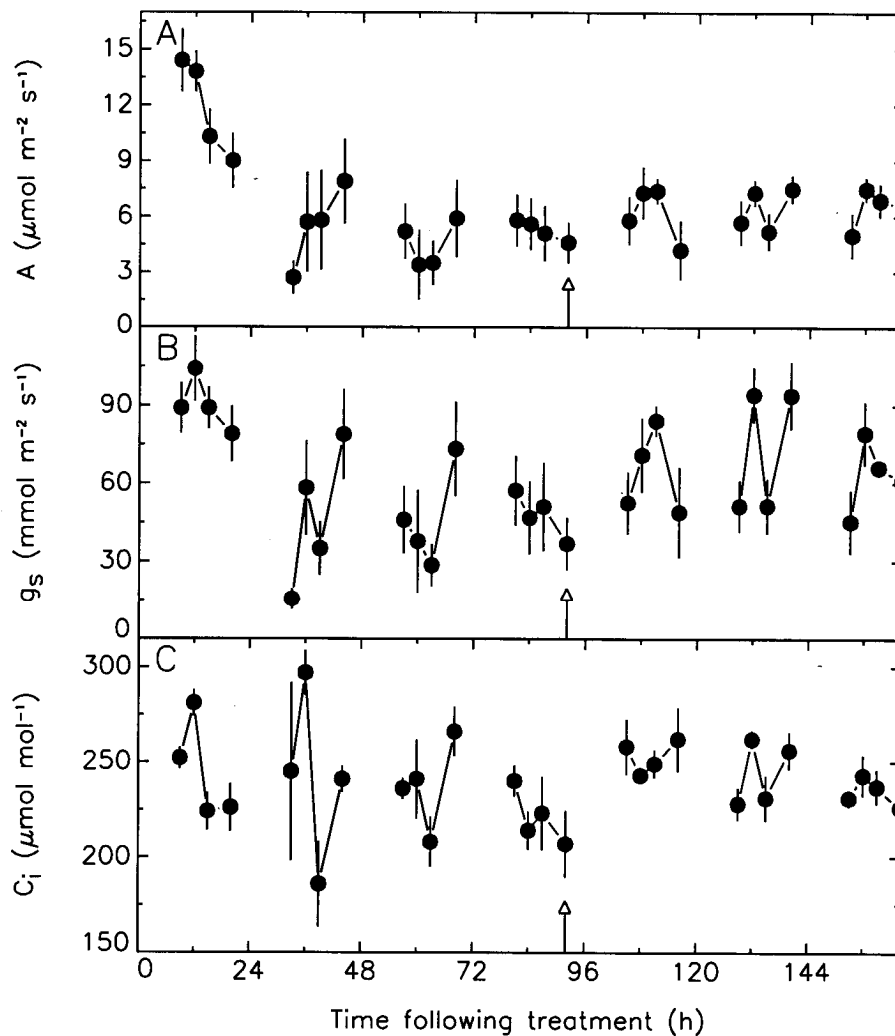


Fig. 7. Effects of continuous lighting (CL) on diurnal (A) net CO₂ assimilation rate (A), (B) stomatal conductance rate (g_s), and (C) internal CO₂ concentration (C_i) over time for 1-year-old, potted sour cherry trees. Gas exchange was measured at 0900, 1200, 0300, and 2000 hr for each day. Treatments: Day 0 (0–24 h) before CL, Day 1 (25–48 h), Day 2 (49–72 h), and Day 3 (73–96 h) represent 1, 2, and 3 days in CL, respectively; Day 4 (97–120 h), Day 5 (121–144 h), and Day 6 (145–168 h) represent recovery 1, 2, and 3 days after return to a 14-h photoperiod, respectively. Arrows point to the time at which plants were returned to a 14-h photoperiod. Each point represents the average (±SE) of four leaf measurements.

boxylation efficiencies and RuBP regeneration rate, and we suggest that a similar short-term mechanism is involved in source leaves of D plants noted here. Enhanced g_s also contributed to the photosynthetic enhancement that was observed. Tschaplinski and Blake (1989a) found a similar increase in A and g_s after decapitation of *Populus* trees. DeJong (1986) concluded that the higher photosynthetic rate of fruiting peach trees vs. those that had been defruited early in the season was due to enhanced g_s. Although we present evidence for a higher estimated carboxylation efficiency and stomatal conductance associated with D, Hudson et al. (1992) suggest that stomatal function is independent of total leaf rubisco activity. The enhancement of A was clearly not due to a reduced oxygenase activity of rubisco since R₁ was not affected by defoliation.

Tschaplinski and Blake (1989b) suggested that carbohydrate reserves in the stems of decapitated *Populus* trees played an important role in buffering short-term responses to altered sink demand. In our sour cherry trees, A was enhanced within 24 h of D. If the argument of Tschaplinski and Blake (1989b) applied to these young, potted sour cherry trees, then the reserve carbohydrate supply in their stems was small and it was rapidly depleted. Photosynthetic enhancement and senescence delay may continue

for weeks or months after D until a sink-limiting condition develops (Satoh et al., 1977). Reduced chlorophyll degradation, or enhanced synthesis may partly account for the short-term enhancement of g_s that was observed, but the long-term, sustained enhancement of A after D may have been due to the increased production of chloroplasts (Woolhouse, 1967) and plastid proteins (Wareing et al. 1968) and a reduced competition for root-derived cytokinin (Satoh et al., 1977; Wareing et al., 1968).

The diurnal A response of individual leaves of plants under a constant light intensity in a growth chamber was similar to that observed in peach (Moing et al., 1992) or for whole trees measured outside in a whole-plant chamber under natural illumination conditions (Sams and Flore, 1983). The photosynthetic rates observed in this experiment were lower than that of field grown cherry trees (as reported in Sams and Flore, 1982). There are two possible explanations for this apparent discrepancy: 1) our trees were not grown outside but rather in the greenhouse and growth chamber and leaves were more typical of shade leaves in terms of morphology and A (Flore and Sams, 1986); and 2) we removed lateral shoots from the trees as they developed thereby reducing the number of competing sinks. The linear relationship between A and

g_s that we observed in sour cherry has been observed in several other tree crops including apple (Lakso, 1979), *Cacao* (Joly and Hahn, 1989) and *Populus* (Liu and Dickmann, 1992). Since this A vs. g_s relationship was quite strong throughout the diurnal photo-period, the decline in A was due, in part, to a similar decline in g_s . Tschaplinski and Blake (1989a) noted similar diurnal responses of A and relationships between A and g_s for rejuvenated stump leaves of decapitated *Populus* plants. Although Loveys et al. (1987) found that abscisic acid was relatively unimportant in the control of g_s in apricot (*P. armeniaca* L.), the possibility of a mild water stress induced stomatal closure in the afternoon cannot be completely tiled out since abscisic acid concentration and leaf water

potential were not measured. The trees in this experiment were watered regularly and exhibited no visual symptoms of water stress. Similar trees in another study (Layne and Flore, 1993) exhibited no change in leaf xylem water potential after D. It is currently not known whether plants of the genus *Prunus* contain a tight-binding inhibitor of rubisco (Seemann et al., 1985) that may lead to partial rubisco deactivation in the afternoon. But since gas-exchange measurements were made at or near a saturating light intensity, the deactivation of rubisco by carboxyarabinitol-1-phosphate appears unlikely (Berry et al., 1987).

D not only resulted in a significant enhancement of A , but carbon pools were significantly altered such that the relative

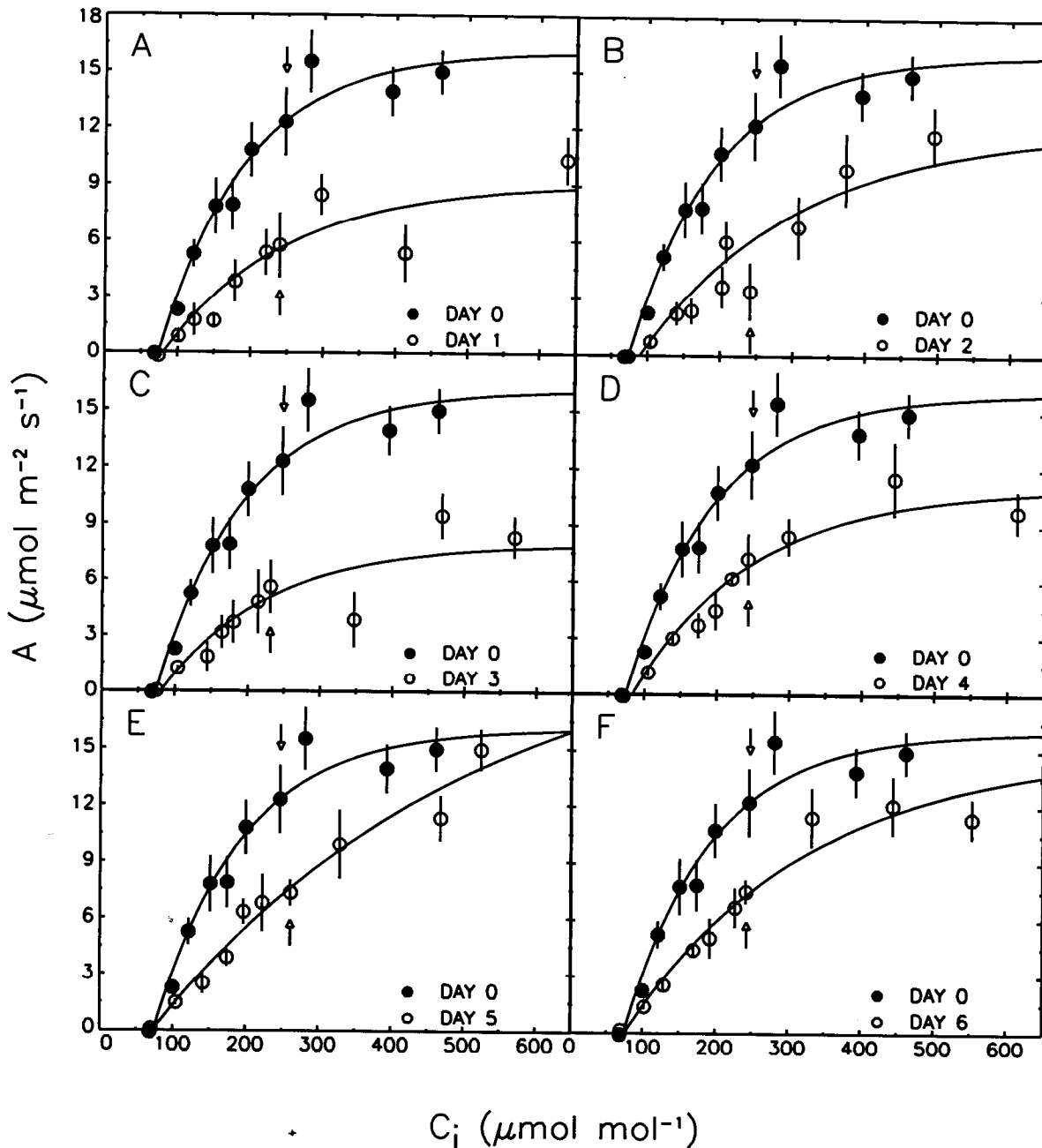


Fig. 8. Effect of continuous lighting (CL) on the response over time of net CO₂ assimilation rate (A) to internal CO₂ concentration (C_i) of 1-year-old, potted sour cherry trees. Each section (A-F) compares the Day 0 pretreatment response (●) with the treatment response for the subsequent day (○). Treatments: Day 0 = before CL; Days 1 (A), 2 (B), and 3 (C) represent 1, 2, and 3 days in CL, respectively; Days 4 (D), 5 (E), and 6 (F) represent recovery 1, 2, and 3 days after return to a 14-h photoperiod, respectively. Regression lines occur about the data. Arrows point to the data collected at ambient CO₂ partial pressure ($350 \pm 25 \mu\text{l-liter}^{-1}$). Data points that represent the average (\pm SE) of four leaf measurements appear about the regressions.

proportions of storage carbohydrate (starch) decreased while transport carbohydrates (sucrose and sorbitol) increased in the source leaf. Enhanced degradation of leaf starch may have provided additional substrates (e.g., glucose-6-phosphate, Glc-6-P) and, coupled with the enhancement of A noted, leaf sorbitol and sucrose biosynthesis may have exceeded the rate of export from the leaf leading to accumulation and a subsequent photosynthetic inhibition. This would indicate that, in the afternoon, trees were sink-limited. To our knowledge, there are no reports of Glc-6-P or sorbitol being directly involved in end-product inhibition of photosynthesis, but Moing et al. (1992) suggested that sorbitol biosynthesis is probably subject to a similar fine control mechanism as exists for sucrose biosynthesis. Although sorbitol accumulation has been associated with osmotic adjustment to short-term drought stress (Bielecki, 1982; Ranney et al., 1991) we did not observe any effect of D on leaf water potential (Layne and Flore, 1993). Direct inhibition of the rate of sucrose synthesis from sucrose accumulation in the cytosol has been suggested as a possible mechanism of feedback inhibition (Herold, 1980), yet Foyer (1988) has suggested that modulation of sucrose phosphate synthase activity occurs by an intermediate component (e.g., glucose, fructose) rather than by sucrose, *per se*. In contrast to the report of Stitt et al. (1990), where sucrose and starch accumulated in feedback-inhibited leaves, under our conditions, as sucrose content increased, starch content actually decreased overtime. In addition, thylakoid reactions, as estimated by chlorophyll fluorescence parameters, were not adversely affected by D (Layne and Flore, 1993).

The reduced R_d coupled with low starch concentrations indicated that leaves of D plants may have been substrate limited for mitochondrial respiration in the night. The variability of R_d during the dark period indicated that factors in addition to starch content were also involved in R_d noted. Avery et al. (1979) noted similar effects of D on potted apple trees, where A and R_d were related to substrate concentrations. Although we did not measure export rate, the fact that A increased while R_d decreased indicates that transport from the leaf probably increased after D (Avery et al., 1979). Avery et al. (1979) further suggested that A was modulated by removal of carbohydrates from the leaf and that there was a negative feedback mechanism via the phloem-transport system. Fondy and Geiger (1982) suggested that starch accumulation and degradation in source leaves of *Beta vulgaris* L. was controlled exogenously by the leaf sucrose concentration and endogenously by the photoperiod or photosynthetic duration. Mullen and Keller (1988) noted that the rate of assimilate export during the night for soybean leaves was closely associated with the rate of starch mobilization but not closely related with either leaf sucrose content or respira-

tory CO_2 loss. They further noted that starch stopped accumulating before the end of the light period and that there was a 75-min delay before starch was mobilized in the dark. It is possible that some of the depletion in leaf starch overtime was due to mobilization in the light as has been observed in cottonwood (Dickson and Larson, 1975).

CL altered the source-sink balance in sour cherry trees and led to an inhibition of leaf photosynthesis. Photosynthetic inhibition from CL has been observed in apple (Bohning, 1949) and sour cherry (Layne and Flore, 1993). Most of the decline in A after CL was from an increased mesophyll limitation as indicated by the significant decline in estimated carboxylation efficiency and RuBP regeneration rate. Although the stomatal limitation to A increased slightly as the duration of exposure to CL increased, C_i was not significantly reduced. Stomatal function has been found to be independent of total leaf rubisco activity (Hudson et al., 1992) and, hence, the physiological consequence of reduced g_s appeared to be negligible under CL conditions. Unlike our control trees in this experiment, or apple trees exposed to varying water stresses (Lakso, 1979), the linear relationship between A and g_s became uncoupled, indicating that A was not primarily limited by g_s under CL.

CL may have increased the mesophyll limitation to photosynthesis through one or a combination of the following mechanisms: 1) feedback inhibition (Foyer, 1988), 2) photoinhibition, or 3) photooxidation (Powles, 1984). Assuming that sink demand remained unchanged under CL, it is likely that the increase in photosynthesis duration resulted in a sink-limiting condition (Sawada et al. 1989). According to Furbank et al. (1987), a stromal orthophosphate (P_i) limitation may have occurred under sink-limiting conditions whereby chloroplast metabolism was restrained by limiting stromal and thylakoid reactions. In support of a P_i limiting condition, we found that leaf starch content significantly increased while sucrose and sorbitol pool concentrations actually decreased while plants were under CL (Layne and Flore, 1993). Since the concentration of soluble carbohydrates in the cytosol decreased under CL (Layne and Flore, 1993), it is unlikely that leaves were feedback inhibited by sucrose, *per se* (Foyer, 1988). On the other hand, starch accumulation in the chloroplast has been associated with an inhibition of A (Gucci et al., 1991a, 1991 b). It is possible that starch mobilization in the light (Dickson and Larson, 1975) was at such a low rate that starch accumulated to inhibitory concentrations causing one or more of the following to occur: 1) starch binding of Mg^{2+} required for rubisco activation (Plaut et al., 1987); 2) starch grain obstruction of the light path to the thylakoid membrane (Warren Wilson, 1966); 3) increasing the

Table 4. The effect of continuous lighting on CO_2 compensation point (Γ), carboxylation efficiency (k), net CO_2 assimilation rate (A_{350}), and internal CO_2 concentration (C_{i350}) at ambient CO_2 (350 μ l-liter⁻¹), and stomata] limitation to A (l_g) of expanded leaves of 1-year-old, potted sour cherry trees with time.

Day	Dark ^y (h)	Light (h)	Γ (μ mol CO_2 /mol)	k (mol CO_2 /m ² per sec)	A_{350} (μ mol CO_2 /m ² per sec)	C_{i350} (μ mol CO_2 /mol)	l_g (%)
0	10	14	75.3	0.088	12.28	246.8	11.0
1	0	24	79.8	0.019***	5.72**	296.7**	18.2 ^{NS}
2	0	24	75.3	0.031***	3.45***	241.4 ^{NS}	58.1***
3	0	24	79.0	0.026***	5.60**	214.3 ^{NS}	30.5*
4	10	14	81.0	0.040***	7.26*	242.5 ^{NS}	27.3 ^{NS}
5	10	14	74.0	0.039***	7.33*	261.5 ^{NS}	29.9 ^{NS}
6	10	14	72.5	0.043***	7.54 ^{NS}	242.6 ^{NS}	29.3 ^{NS}

^yMeans for each day are compared relative to Day 0 by orthogonal contrasts in each column. Means are averages of four replicates.

^yDark and Light correspond to the number of hours in darkness and light during a 24 h cycle.

^{NS}, *, **, ***Nonsignificant or significant at $P = 0.05$, 0.01, or 0.001, respectively. Absence of notation indicates no significant differences among days.

diffusive path length or interfering with intercellular CO₂ transport (Nafziger and Keller, 1976); or 4) physical damage to chloroplasts (Schaffer et al., 1986; Wildman, 1967).

Thylakoid reactions including light trapping, electron transport, and ATP production via ATP synthase were probably limited under CL as indicated by the reduced F_v/F_m and estimated RuBP

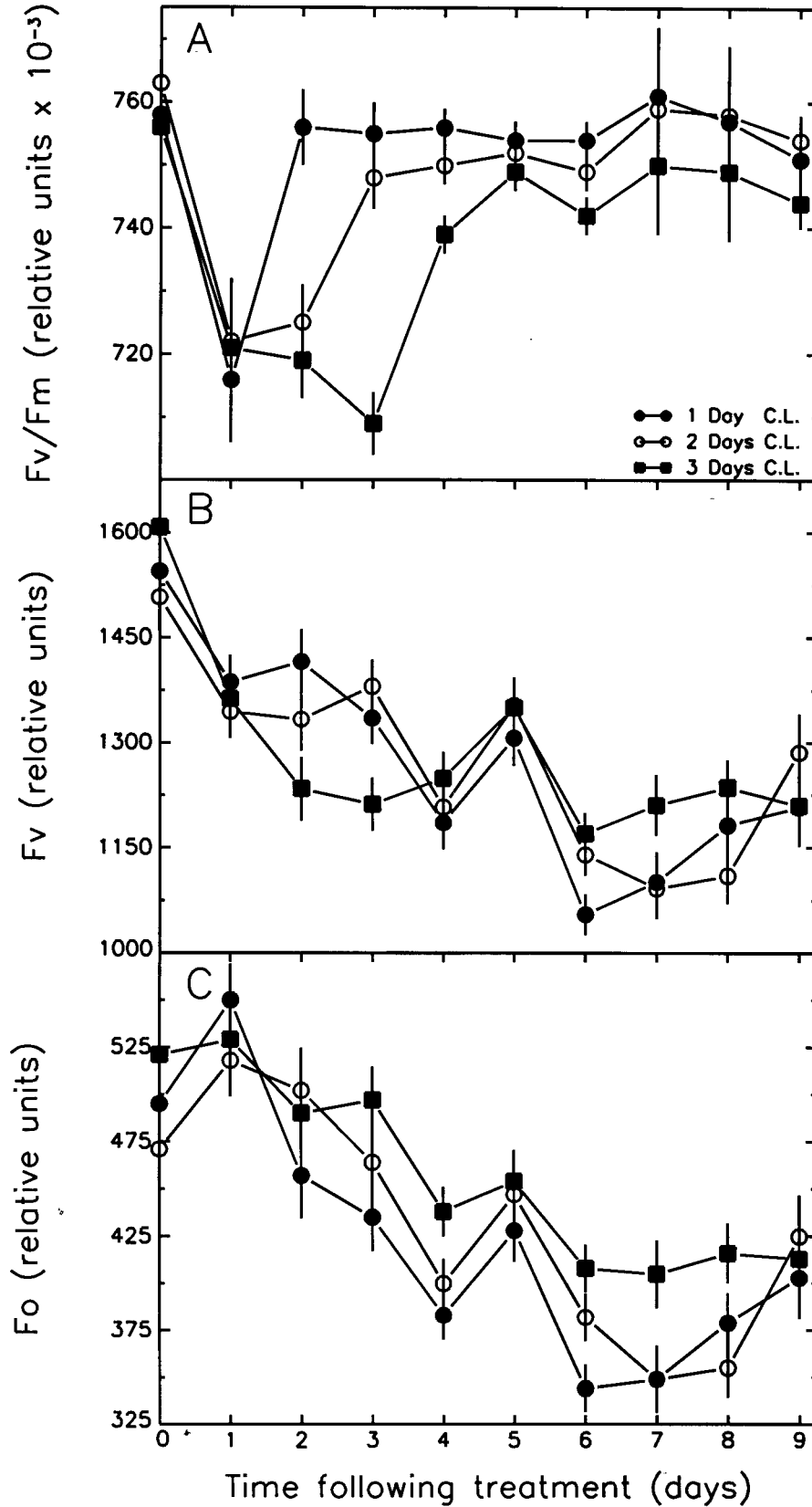


Fig. 9. Time course for the response of (A) estimated photochemical efficiency (F_v/F_m), (B) variable fluorescence (F_v), and (C) instantaneous fluorescence (F_o) to continuous lighting (CL) in 1-year-old, potted sour cherry trees. Treatments: 1 (●), 2 (○), and 3 (■) days in CL, respectively. Chlorophyll fluorescence was measured daily beginning at 1500 hr. Each point represents the average (\pm SE) of 20 leaf measurements.

regeneration rate. A P_i limitation in the chloroplast stroma should decrease the ATP : ADP ratio and depress the reduction of 3-phosphoglycerate and the regeneration of RuBP (Robinson and Walker, 1981). Since A and carboxylation efficiency were significantly inhibited under CL, substrate RuBP concentrations may have limited carboxylation as well. The P_i requirement of chloroplasts varies with O_2 pressure (Usuda and Edwards, 1982) and decreasing oxygenation (by decreasing O_2 pressure) lowers A by making P_i less available (Sharkey, 1985). We did not observe O_2 -insensitivity of A upon exposure to low O_2 (Sharkey, 1985) but we measured the low O_2 response at significantly higher leaf temperatures (28°C) than those reported by Sage and Sharkey (1987) where O_2 sensitivity was observed. To definitively verify whether P_i was limiting in the stroma or cytosol of feedback-limited leaves, nonaqueous fractionation techniques could be used (Sharkey and Vanderveer, 1989).

Photoinhibition, which precedes chlorophyll bleaching or photooxidation (Powles, 1984), also contributed to the decline in A under CL. Karukstis (1991) stated that photoinhibition of photosynthesis is manifested at the leaf, thylakoid, and photosystem II level by losses in CO_2 fixation, electron transport capacity, and F_v , respectively. Indeed, according to these conditions, sour cherry leaves exposed to CL were photoinhibited. Like Williams and Farrar (1988) using barley, we noted in cherry that as CL duration increased, the amount of reduction in F_v increased. Since the observed photoinhibition resulted in a decrease in F_0 overtime, this indicated that there was a reversible regulatory mechanism whereby the damage to photosystem II centers could be repaired (Krause, 1988). The restoration of F_v/F_m after return to a 14-h photoperiod in cherry leaves indicated that recovery from photoinhibition had indeed occurred (Krause, 1988). In studies on citrus, starch accumulation led to leaf chlorosis, senescence, and disruption of thylakoid membranes (Schaffer et al., 1986). Although there was a significant increase in leaf starch content during CL, up to 1.5% dry weight (Layne and Flore, 1993), we did not observe an effect of CL on leaf chlorophyll content. We assume that photooxidation did not occur in these experiments since no leaf chlorosis or necrosis was observed during 7 days of CL. If photooxidative damage did occur, it appears that the rate of damage did not exceed the rate of repair (Kyle, 1987). The full photosynthetic recovery after defoliation of CL plants may have been due to relief of end-product inhibition by depletion of accumulated starch (Sasek et al., 1985), reduced competition for root derived cytokinins (Satoh et al., 1977; Wareing et al., 1968) and/or production of new photosynthetic machinery (Satoh et al., 1977). Photosynthetic inhibition was not observed in partially defoliated plants that were continuously illuminated. It is possible that the export rate from source leaves to sinks may have been high enough to prevent carbohydrate accumulation under these conditions (Tschaplinski and Blake, 1989b).

The model plant system was used so that plant uniformity could be attained, environmental conditions could be precisely regulated, and consistent experimental manipulations could be achieved. We hoped thereby to eliminate the likely confounding effects that would occur in the field (weather, yield inconsistency, etc.). Use of a model plant system for detailed physiological studies that are difficult or impossible in the field is not without drawbacks. While maintaining source-sink uniformity in plant material, controlling environmental conditions, and ensuring that biotic or abiotic stress conditions did not confound the experiment, a compromise was made in being able to extrapolate these findings to how mature, fruiting trees would respond in the field (Gucci, 1988). By training trees to a single shoot for uniformity and utilization of walk-in,

plant growth chambers, we may have predisposed trees to a sink-limiting condition (Layne and Flore, 1993). In future studies, it would be desirable to allow a significant number of competing sinks (developing shoots) to develop to ensure that sink-limited conditions did not occur. Whether plants such as those just described would demonstrate a diurnal decline in A after D is not known. With the recent development of inflatable balloons for whole-tree gas-exchange measurements (Corelli-Grappadelli and Magnanini, 1993) and new gas-exchange equipment that can now be used in the field for the development of CO_2 , light, and VPD responses, more detailed physiological data can be collected in the field than before. Thus, whole-tree responses to source-sink alterations can be accurately monitored for mature, fruiting trees and the phenomenon and implications of end-product inhibition of A can be thoroughly investigated in the orchard.

In conclusion, under certain conditions, sink demand limits photosynthesis in young, potted sour cherry trees through the efficiency of photoassimilate production and use (Foyer, 1988). Manipulating whole plants to alter the source-sink ratio enabled us to investigate the possible mechanisms involved in the end-product regulation of photosynthesis in young, potted trees. Both whole-plant manipulation techniques that we used resulted in conditions where A was inhibited but the mechanism of action was probably different. The diurnal decline in A for D plants was associated with an increase in leaf sucrose and sorbitol content and a reduction in leaf starch indicating that, in the afternoon, trees were sink limited, and that sucrose and/or sorbitol were likely involved in the inhibition noted. We are currently unaware of any other reports linking leaf sorbitol content to an inhibition of source leaf A. Thylakoid reactions were not adversely affected by D. On the other hand, the decline in A after exposure to CL was in part due to photoinhibition and an accumulation of leaf starch (Layne and Flore, 1993). Whether chloroplasts became engorged with starch grains such that the thylakoid membrane became disrupted and electron transport through photosystem II was adversely affected as suggested by Wildman (1967) remains equivocal. Responses to CL also indicated that a P_i limitation existed but actual P_i concentrations were not determined. Finally, sour cherry A is affected by assimilate concentrations in the leaf. The precise points of regulation in the pathways to production of starch, sucrose, and sorbitol under sink-limiting conditions need to be determined.

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