UC Irvine

UC Irvine Previously Published Works

Title

Endocannabinoid System and the Kidneys: From Renal Physiology to Injury and Disease.

Permalink

https://escholarship.org/uc/item/4vc072ps

Journal

Cannabis and cannabinoid research, 4(1)

ISSN

2578-5125

Authors

Chua, Janice T Argueta, Donovan A DiPatrizio, Nicholas V et al.

Publication Date

2019

DOI

10.1089/can.2018.0060

Peer reviewed

Cannabis and Cannabinoid Research Volume 4, Number 1, 2019 Mary Ann Liebert, Inc. DOI: 10.1089/can.2018.0060

REVIEW

Endocannabinoid System and the Kidneys: From Renal Physiology to Injury and Disease

Janice T. Chua, Donovan A. Argueta, Nicholas V. DiPatrizio, Csaba P. Kovesdy, A Nosratola D. Vaziri, Kamyar Kalantar-Zadeh, A and Hamid Moradi 1,5,*

Abstract

Introduction: As the prevalence of kidney disease continues to rise worldwide, there is accumulating evidence that kidney injury and dysfunction, whether acute or chronic, is associated with major adverse outcomes, including mortality. Meanwhile, effective therapeutic options in the treatment of acute kidney injury (AKI) and chronic kidney disease (CKD) have been sparse. Many of the effective treatments that are routinely utilized for different pathologies in patients without kidney disease have failed to demonstrate efficacy in those with renal dysfunction. Hence, there is an urgent need for discovery of novel pathways that can be targeted for innovative and effective clinical therapies in renal disease states. **Discussion:** There is now accumulating evidence that the endocannabinoid (EC) system plays a prominent role in normal renal homeostasis and function. In addition, numerous recent studies have described mechanisms through which alteration in the EC system can contribute to kidney damage and disease. These include a potential role for cannabinoid receptors in tubulo-glomerular damage and fibrosis, which are common features of AKI, interstitial nephritis, glomerulopathy, and other conditions leading to AKI and CKD.

Conclusion: These findings suggest that manipulating the EC system may be an effective therapeutic strategy for the treatment of kidney disease and injury. However, further mechanistic studies are needed to fully delineate the role of this system in various conditions affecting the kidneys. Furthermore, while most of the current literature is focused on the role of the EC system as a whole in renal pathophysiology, future studies will also need to clarify the contribution of each component of this system, including the EC mediators, in the pathogenesis of kidney disease and their potential role as part of a therapeutic strategy.

Keywords: acute kidney injury; chronic kidney disease; endocannabinoid; fibrosis; inflammation; nephropathy

Introduction

The kidneys play a central role in normal body homeostasis through a variety of functions, including removal of byproducts of metabolism, clearance of toxins, regulation of body volume status, electrolytes and systemic hemodynamics, and production of hormones such as erythropoietin and active vitamin D. Hence, it is not surprising that kidney damage is associated with significant morbidity and mortality. The latter is true

whether the decline in renal function is part of an acute process such as acute kidney injury (AKI) due to tubular necrosis, or a more chronic process such as chronic kidney disease (CKD) caused by hypertension (HTN) or diabetes. Furthermore, the mechanisms responsible for renal injury are complex and can be varied. While these mechanisms are regularly categorized based on the type of injury (acute or chronic) and anatomic part of the nephron affected (including the

¹University of California–Irvine, School of Medicine, Orange, California.

²Division of Biomedical Sciences, School of Medicine, University of California, Riverside, Riverside, California.

³Division of Nephrology, University of Tennessee Health Science Center, Memphis, Tennessee.

⁴Nephrology Section, Memphis Veterans Affairs Medical Center, Memphis, Tennessee.

⁵Nephrology Section, Tibor Rubin Veteran Affairs Health System, Long Beach, California.

^{*}Address correspondence to: Hamid Moradi, MD, Nephrology Section, Department of Medicine, Tibor Rubin Veteran Affairs Health System, 5901 E. 7th Street, Long Beach, CA 90822, E-mail: hamid.moradi@va.gov

[©] Janice T. Chua et al. 2019; Published by Mary Ann Liebert, Inc. This Open Access article is distributed under the terms of the Creative Commons License (http://creativecommons.org/licenses/by/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

glomerulus, tubules, mesangium, vasculature), there is significant overlap between these categories. For instance, there is evidence indicating that AKI can result in CKD. In addition, there is frequent overlap between the different anatomic sites of injury given that damage to one part of the nephron over a period of time can result in injury to other sites. For example, while diabetic kidney disease often manifests with glomerular injury and proteinuria, over an extended period of time it also results in tubulointerstitial damage and fibrosis leading to progressive CKD and end-stage kidney disease. Therefore, understanding the underlying pathways whose alterations can result in various forms of renal damage and injury can play an important role in devising effective therapies to prevent and treat kidney disease. In this regard, there is accumulating evidence that indicates that the endocannabinoid (EC) system plays a major role in normal renal physiology. In addition, there are data demonstrating that alterations of this pathway can lead to the pathogenesis of both acute and chronic kidney disease. Therefore, evaluation of the EC system can be a promising area of discovery, which may result in the generation of potentially novel therapies aimed at treating various forms of kidney disease.

The EC system comprises endogenous fatty acidderived ligands, their receptors, and the enzymes required for their biosynthesis and degradation.¹ The most wellcharacterized ECs are N-arachidonoyl ethanolamide, also known as anandamide (AEA), and 2-arachidonoylsn-glycerol (2-AG).² These lipid-derived molecules are generated on-demand by the metabolism of membrane phospholipids in response to various stimuli, including elevated intracellular calcium or metabotropic receptor activation.³ After production, they bind to the local cannabinoid receptors in an autocrine or paracrine manner, although measurable concentrations of these ligands can also be found in the blood, cerebrospinal fluid, and lymph. While the potential endocrine actions of these ECs remain an area of active research, it is well established that they act locally by binding with two widely studied cannabinoid receptors, cannabinoid subtype-1 (CB₁) and subtype-2 (CB₂).⁵ AEA and 2-AG can subsequently be taken up by cells through a high-affinity uptake mechanism^{6,7} and rapidly degraded through the action of the enzymes, fatty acid amide hydrolase (FAAH), and monoacylglycerol lipase (MGL), respectively.¹

While the role of the EC system has been initially a focus of extensive research in the central nervous system, over the course of the past two decades, a significant number of studies have confirmed its presence and importance in the peripheral organs, including the kidneys.

In this regard, substantial concentrations of ECs, the machinery required for their biosynthesis and degradation, as well as CB receptors have been detected in kidney tissue. ^{8,9} The effects produced by the actions of this system in the normal and pathological conditions of the kidney, however, have not been fully delineated given the many complexities involved in the production and breakdown of EC ligands. ⁸⁻¹¹ In addition, the differential distribution and actions of the CB₁ and CB₂ receptors in various structures and cell subtypes in the kidney can ultimately result in varied signaling outcomes whose overall impact will be difficult to predict. Accordingly, identifying the physiologic and pathophysiologic roles of the EC system in the field of nephrology remains an active area of exploration.

EC System and Normal Renal Physiology

It has been shown that CB₁ and CB₂ belong to a class of seven transmembrane domain G-protein-coupled receptors that are functionally dependent on the activation of heterotrimeric G_i/G₀ proteins.¹¹ Although the activation of both receptors results in the inhibition of adenylyl cyclase enzyme and increased activity of mitogen-activated protein kinase (MAPK), CB₁ activation has also been shown to stimulate nitric oxide synthase and directly control the activation of ion channels. The latter include the inwardly rectifying and A-type outward potassium channels, D-type outward potassium channels, and Ntype and P/Q-type calcium channels. 10,12,13 Despite the common G-protein subunit shared between CB1 and CB₂ receptors, their activation can produce opposing biological effects in normal and diseased states, in part due to the abundance and localization of these cannabinoid receptors and their EC ligands.

While the CB₁ receptor was initially thought to be localized to the central and peripheral nervous system, 12,14 it has been shown to be present in peripheral organs such as the kidneys. 15,16 For instance, the presence of functional CB₁ receptor has been demonstrated in proximal convoluted tubules, distal tubules, and intercalated cells of the collecting duct in the human kidney¹³ (Fig. 1). Furthermore, CB₁ receptor expression has also been found in other parts of the nephron in rodents, such as the afferent and efferent arterioles, 17 thick ascending limbs (TAL) of the loop of Henle, 18 and glomeruli, 19-23 as well as in various kidney cell subtypes such as glomerular podocytes, 24,25 tubular epithelial cells, 13,15,20,21,24,26-29 and cultured mesangial cells. 30,31 Similarly, the expression of CB2 receptors, although previously thought to be predominantly in immune cells,³² has also been

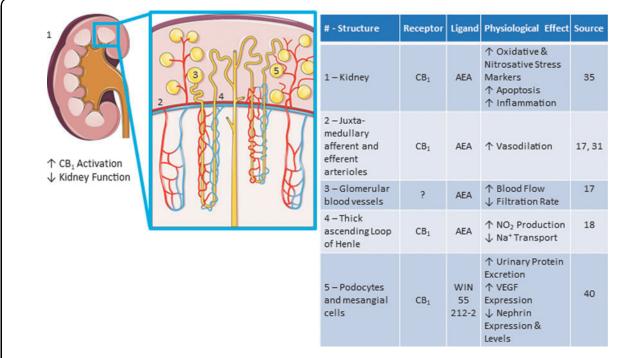


FIG. 1. Roles of the endocannabinoid system in healthy kidney. Images adapted from Servier Medical Art "Kidney" licensed under CC 3.0, https://smart.servier.com/smart_image/kidney

demonstrated in renal tissue.³³ For example, CB_2 receptor expression has been localized to podocytes,²⁵ proximal tubule cells,^{26,33,34} and mesangial cells³¹ in human and rat renal cortex samples.

In addition to differential expression of CB receptors in different tissues and cells, the complex regulation of the biosynthesis and degradation of the kidney's high basal levels of ECs through downstream enzymes contributes to the varied signaling effects of these ligands. $^{20,31,35-39}$ While the renal cortex displayed similar levels of AEA and 2-AG, AEA was demonstrated to be enriched in the kidney medulla compared with the cortex, while the levels of 2-AG in the medulla were similar to those of both ECs in the cortex.³⁹ Moreover, AEA is present in cultured renal endothelial and mesangial cells at low levels and can be synthesized from arachidonic acid and ethanolamine and catabolized by AEA amidase in these kidney cell subtypes.³¹ The expression of FAAH was shown to be augmented in the renal cortex (e.g., in the glomerulus, tubular system, and collecting ducts) in comparison to its low expression levels in the medulla.³⁹

Considering the diverse localization of the ECs and their receptors, as well as the complexities involved in their synthesis and catabolism, this system can play various roles in kidney function. Under normal conditions, the EC system is capable of regulating renal homeostasis as demonstrated by its control over renal hemodynamics, tubular sodium reabsorption, and urinary protein excretion. These effects are largely imparted through the activation of the CB_1 receptor. $^{17,18,31,39-41}$ In the following sections, we describe some effects of EC system activation on renal physiologic function (Fig. 1).

Renal hemodynamics

Under normal physiologic conditions, the EC system plays a critical role in the regulation of renal hemodynamics. For instance, it was shown that intravenous administration of AEA decreased glomerular filtration rate and increased renal blood flow in rodents, independent of changes in blood pressure. ¹⁷ In vitro studies showed that AEA can vasodilate juxtamedullary afferent or efferent arterioles^{17,31} through a CB₁-dependent process, normally inhibited by nitric oxide synthase, 31 to regulate glomerular filteration rate (GFR). The actions of the AEA signaling system are likely conducted through endothelial and mesangial cells, which are capable of producing and metabolizing AEA,31 as well as through the hyperpolarization of smooth muscle cells via the activation of potassium channels.⁴² It should be noted that there are also non-CB₁ receptor-dependent mechanisms

by which ECs can mediate a vasodilatory effect and thereby regulate renal hemodynamics.⁴³ Future studies need to further elucidate the role of the latter mechanisms in normal renal physiologic homeostasis.

Tubular sodium transport

AEA has been shown to have a regulatory effect on tubular sodium transport. In the medullary TAL of Henle's loop, AEA (through interaction with the CB₁ receptor) was shown to stimulate nitric oxide production, leading to an inhibition of sodium transport through the apical Na⁺/H⁺ transporter and Na⁺/K⁺/2Cl⁻ co-transporter. This was also associated with reduced oxygen consumption in the TAL portion of the nephron. This suggests that the activation of CB receptors via AEA can regulate renal blood flow as well as tubular handling of solutes, which can ultimately impact renal salt and water clearance.

Urinary protein excretion and modulation

To examine the role of glomerular CB₁ receptors in modulating urinary protein excretion, Hsu et al. used

CB₁ transgenic mice and rats treated with a selective CB₁ agonist. CB₁ receptor activation in the kidney, and specifically in the podocytes and mesangial cells of the glomerulus, increased urinary protein excretion. Increased activation and overexpression of CB₁ was also found to enhance vascular endothelial growth factor (VEGF) expression levels and subsequently reduce nephrin gene and protein levels, suggesting a potential pathway for podocyte dysregulation and proteinuria. Description

EC System and Renal Disease

The role of the EC system in renal pathology and dysfunction is an emerging area of research, which has been studied primarily in the context of CB receptors. Alterations of CB receptor expression and activity have been discovered in various renal diseases such as diabetic nephropathy, CKD, and different types of kidney injury (Fig. 2). Collectively, these studies on renal pathophysiology suggest that targeting the EC system may be of diagnostic and therapeutic value (Tables 1 and 2).

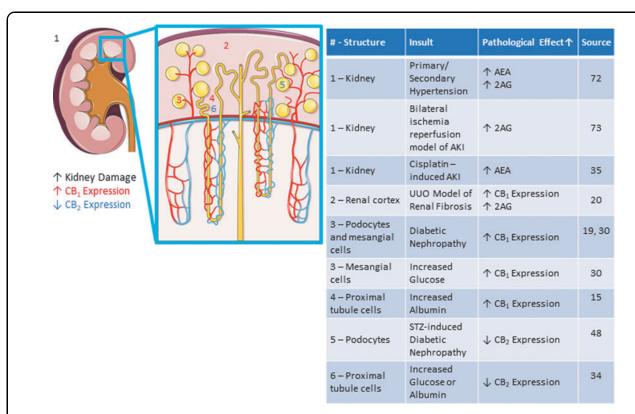


FIG. 2. Impact of various kidney disease models on receptors and ligands of the endocannabinoid system. Images adapted from Servier Medical Art "Kidney" licensed under CC 3.0, https://smart.servier.com/smart_image/kidney

Table 1. Impact of Targeting Cannabinoid Receptors in the Models of Kidney Disease

Mediator	Treatment	Location	Effect	Disease model	Source
CB ₁	AM251 (inverse agonist)		↓ Albuminuria	Mouse STZ-induced diabetic nephropathy	19
CB ₁	Rimonabant (inverse agonist)	Glomeruli and proximal tubules	↓ Apoptosis	Rat STZ-induced diabetic nephropathy	19,21,29
CB ₂	AM1421 (agonist)	Podocytes	↓ Albuminuria ↓ Monocyte infiltration □ Transposion of profileration	Mouse/rat obesity-related nephropathy	25,33
CB ₂	HU910 (agonist)	Podocytes	↓ Expression of profibrotic markers ¬Restored nephrin expression ↓ Albuminuria ↓ Mesangial matrix formation ↓ Fibronectin accumulation ↓ Sclerotic damage	ob/ob mice	49
CB ₁	JD5037 (inverse agonist)		↑ Renal structure and function ↓ Proteinuria	ZDF rats	24
CB ₁	AM251 rimonabant (inverse agonist)		↓ Body weight ↓ Systolic blood pressure ↓ Circulating leptin ↓ Albuminuria ↓ Circulating creatinine	DIO rats db/db mice	16,23
CB ₁	Rimonabant (inverse agonist)		↓ Glomerular fibrosis ↓ Tubular damage ↓ Renal hypertrophy	DIO rats	21,52–54
CB ₁	Knockout	Renal proximal tubule cells	↓ Lipid accumulation ↓ Liver kinase B1 activation ↓ AMP-activated protein kinase activity ↓ Fatty acid beta-oxidation	DIO mice	55
CB ₁	Rimonabant (inverse agonist)		↓ Monocyte chemoattractant protein-1 ↓ Macrophage infiltration	Mouse UUO	20
CB ₁	Win55,212-2 (agonist)	Glomeruli	↑ VEGF	Mouse/rat UUO	40
CB ₁ /CB ₂	ACPA/JWH133 (agonists)	Tubules	-Prevented damage	Mouse renal ischemia/reperfusion injury	56
CB ₁ /CB ₂	AM281/BCP (antagonist/agonist)		↓ Oxidative stress ↓ Tubular damage	Mouse cisplatin-induced renal damage	35,59–61
CB2	LEI-101 (agonist)		↓ Serum BUN ↓ Serum creatinine	Cisplatin-induced renal damage	59
CB2	Knockout		↑ Mortality ↑ Lung injury ↑ Bacteremia ↑ Neutrophil recruitment ↓ p38 MAPK at injury site	CLP model of sepsis in mouse	64
CB ₁ ?	Palmitic acid	Proximal tubule cells	↓ Apoptosis	Human hyperlipidemia/diabetic nephropathy	29
?	CBD	Tubules	↓ Serum creatinine ↓ Renal malondialdehyde ↓ Nitric oxide	Rat renal ischemia/reperfusion injury	57

Arrows represent an observed change p < 0.05. Question mark represents unconfirmed or unknown mediator.

BUN, blood urea nitrogen; CBD, cannabidiol; CLP, cecal ligation and puncture; STZ, streptozotocin; UUO, unilateral ureteral obstruction; VEGF, vascular endothelial growth factor; ZDF, Zucker diabetic fatty.

Diabetic nephropathy

It is well known that diabetes has major renal complications, including progressive kidney disease and pathology, a condition known as diabetic nephropathy. Diabetic nephropathy is characterized by glomerular hypertrophy and hyperfiltration, which can result in albuminuria, renal fibrosis, GFR decline, and end-stage renal disease. Several studies have examined the role of the EC system in diabetes-related podocyte, mesangial and tubular cell injury, as well as the func-

tion of CB receptor activation on the adverse outcomes of diabetic nephropathy (Fig. 2).

The evaluation of mouse models of diabetic kidney disease and renal tissue from humans with advanced diabetic nephropathy have shown elevated levels of CB₁ receptor expression in the kidney, and in particular in glomerular podocytes and mesangial cells. ^{19,30} In addition, *in vitro* studies have shown CB₁ receptor upregulation with exposure to increased glucose and albumin concentrations in mesangial cells³⁰ and

Ligand	Treatment	Location	Effect	Disease model	Source
AEA/2AG	URB597 (FAAH inhibitor)	Whole kidney	Inhibited ROS generation Augmented ROS-dependent phospholipid peroxidation	Rat primary/secondary HTN	72
AEA/2AG	URB597 (FAAH inhibitor)	Whole kidney	↑ Antioxidant defense ↑ CB ₁ /CB ₂ expression	Rat primary HTN	72
AEA/2AG	URB597 (FAAH inhibitor)	Whole kidney	 Pro-inflammatory responses ↑ CB₂ expression ↑ TRPV1 expression 	Rat secondary HTN	72
2AG	JZL184 (MGL inhibitor)	Serum Medulla	↓ Serum BUN ↓ Serum creatinine ↑ Tubular damage score	Mouse ischemia/reperfusion model of AKI	73
AEA/2AG	Administration	Human cultured podocytes	↑ Inflammation and injury ↓ Podocin ↓ Nephrin	Chronic high-glucose exposure	74
AEA	Administration	Mouse cultured podocytes	↑ Desmin gene expression -Blocked NLRP3 inflammasome activation -Ameliorated podocyte dysfunction	L-Hcys-induced podocyte injury	75

Table 2. Impact of Endocannabinoid Manipulation in the Models of Kidney Disease

2-AG, 2-arachidonoyl-sn-glycerol; AEA, anandamide; AKI, acute kidney injury; CB_1 , cannabinoid subtype-1; CB_2 , cannabinoid subtype-2; FAAH, fatty acid amide hydrolase; HTN, hypertension; Hcys, ι -homocysteine; MGL, monoacylglycerol lipase; ROS, reactive oxygen species.

proximal tubule cells, respectively. ¹⁵ Furthermore, the CB_1 receptor has been found to be overexpressed in glomerular podocytes in experimental mice with diabetic nephropathy. ^{19,24} The potential consequences of the latter changes were shown in another study that found that hyperlipidemia, as induced by diabetic nephropathy, can be associated with palmitic acid–induced apoptosis in proximal tubular cells. These actions are mediated through upregulated CB_1 receptor expression. ²⁹

Given the evidence indicating a deleterious role for the CB₁ receptor in diabetic nephropathy, several studies have investigated the utility of CB₁ antagonist/inverse agonists as a potential therapeutic option for diabetic kidney disease. ^{15,20,24,46,47}

In a streptozotocin (STZ)-induced mouse model of diabetic nephropathy, albuminuria was reduced as a result of CB₁ receptor blockade through a selective CB₁ receptor antagonist. Similar findings were also reported in genetic mouse models of diabetic nephropathy. It was found that a marked reduction in proteinuria occurred through the preservation of glomerular podocytes and restoration of the expression of podocyte proteins nephrin, podocin, and zonula occludens-1. An addition, CB₁ antagonism was also found to be associated with decreased glomerular and proximal tubular apoptosis, ultimately leading to improvements in renal function.

In Zucker diabetic fatty (ZDF) rats, which develop type 2 diabetes due to obesity caused by a dysfunctional leptin receptor, chronic administration of a CB₁ receptor inverse agonist restored GFR, reduced proteinuria, and improved the markers of podocyte health through modulation of the renin-angiotensin system and inhibition of apoptosis.²⁴

While diabetic kidney disease is associated with increased expression of the CB₁ receptor in various parts of the nephron, there is also evidence that CB₂ receptor expression is significantly reduced. For example, STZinduced diabetic nephropathy in mice is associated with the downregulation of glomerular podocyte CB2 receptor expression. 48 Similarly, there is decreased expression of the CB₂ receptor in proximal tubule cells following exposure to elevated concentrations of albumin and glucose.³⁴ Furthermore, CB2 receptor activation has been shown to ameliorate albuminuria, restore podocyte protein expression, reduce monocyte infiltration, and decrease the expression of renal profibrotic markers^{25,33} in rats with obesity-related nephropathy. CB₂ agonism in obese diabetic nephropathy BTBR ob/ob mouse strain also reduced albuminuria, ameliorated dysfunctional nephrin expression in podocytes, and reduced mesangial matrix expansion, fibronectin accumulation, and sclerotic damage.⁴⁹

These studies demonstrate that antagonism of CB₁ receptors and activation of CB₂ receptors using selective pharmacological ligands is associated with the restoration of renal structure and function, specifically albuminuria and the expression of inflammatory markers, in genetic and experimental models of diabetic nephropathy.

Obesity-related kidney disease

Obesity is associated with and acts as a risk factor for the development of diabetic nephropathy,⁵⁰ with obese individuals possessing a higher risk of progressing

to end-stage renal disease.⁵¹ A study by Jenkin et al.³³ revealed a role of CB2 receptor activation in reducing the progression of obesity-related kidney dysfunction by decreasing proteinuria, creatinine clearance, and renal fibrotic markers. In contrast, ZDF rats treated with a CB₁ inverse agonist showed improved renal structure and function.²⁴ Surprisingly, these rats also displayed a marked increase in body weight compared with the weight stability of control mice, which was thought to be due to the development of extreme hyperglycemia in vehicle-treated controls.²⁴ In a separate study, CB₁ antagonism was shown to decrease albuminuria, reduce mesangial expansion, and ameliorate the expression of profibrotic and proinflammatory kidney proteins in lean and obese diabetic mouse models.²³ These findings suggest that the consequences of CB₁ modulation in diabetes can differ based on the experimental model, presence of obesity, and presence of hyperglycemia.

In other studies that utilized rats with diet-induced obesity, CB₁ receptor expression in the kidney is notably upregulated, and treatment with a CB₁ receptor antagonist reduced weight, systolic blood pressure, plasma leptin, albuminuria, and plasma creatinine levels. This is associated with the amelioration of glomerulopathy.¹⁶ Furthermore, studies using obese Zucker rats demonstrated that the CB₁ inverse agonist, rimonabant, ameliorated proteinuria in an animal model of obesityinduced nephropathy.21 Treatment with rimonabant partially restored creatinine clearance, reduced glomerulosclerosis and tubular-interstitial fibrosis, and lowered tubular damage and renal hypertrophy.21 It should also be noted that these findings may have been mediated by the effects of rimonabant and not related to the EC system. While obesity in fa/fa Zucker rats is caused by a mutation of the leptin receptor, rimonabant acts to increase leptin uptake by the kidney, which has been shown to reduce proximal tubule metabolic activity.⁵² Therefore, improvement in renal function in these rats may have occurred due to mechanisms related to leptin's role in proximal tubule cell metabolism, 52-54 as opposed to a direct action on the EC system.

Using a novel mouse strain lacking CB₁ receptors in renal proximal tubule cells, Udi et al.⁵⁵ found that CB₁ receptor deletion did not protect the mice from the deleterious metabolic effects associated with obesity, but significantly diminished obesity-induced lipid accumulation in the kidney. Furthermore, the stimulation of CB₁ receptors in renal proximal tubule cells was found to be associated with decreased activation of liver kinase

B1 and decreased activity of AMP-activated protein kinase, as well as reduced fatty acid beta-oxidation.⁵⁵ These findings indicate a potential relationship between renal proximal tubular epithelial cell CB₁ receptor and the pathologic effects of obesity-induced renal lipotoxicity and nephropathy.

In summary, the findings related to the CB₁ receptor highlight its partial potential in acting as a therapeutic target for obesity-induced renal disease. Further studies are needed to ascertain the efficacy of modulating CB₁ in the kidney to improve renal dysfunction independent of its effects on weight.

Renal interstitial disease and fibrosis

The CB₁ receptor has been shown to be upregulated in other renal disorders marked by interstitial inflammation and fibrosis, including acute interstitial nephritis.²⁰ Using unilateral ureteral obstruction (UUO) as an experimental model for renal fibrosis in mice, Lecru et al.²⁰ showed that CB₁ receptor expression was upregulated in UUO animals compared with controls. This is also associated with a marked increase in the renal content of 2-AG. Treatment of UUO mice with rimonabant reduced monocyte chemoattractant protein-1 synthesis and decreased macrophage infiltration.²⁰ It was also shown that CB₁ receptor activation led to enhanced VEGF levels, which subsequently reduced nephrin expression and protein levels.⁴⁰

Acute kidney injury

There is accumulating evidence indicating the important role of CB₁ and CB₂ receptors and their modulation in the pathogenesis of various forms of AKI. With regard to ischemic AKI, selective CB₁ and CB₂ receptor agonists were found to have a dose-dependent effect in preventing tubular damage following renal ischemia/reperfusion injury in mouse kidney.⁵⁶ In a separate study, however, the administration of cannabidiol, a non-psychoactive constituent of cannabis with poorly defined pharmacological properties, led to a reduction in renal tubular injury in rats following bilateral renal ischemia/reperfusion.⁵⁷ Cannabidiol significantly attenuated the elevation of serum creatinine and renal malondialdehyde and nitric oxide levels associated with this condition.⁵⁷ In a more recent study, a triazolopyrimidine-derived CB2 receptor agonist was demonstrated to play a protective role in inflammatory renal injury following bilateral kidney ischemia/reperfusion.⁵⁸

A series of studies have demonstrated the deleterious role of CB₁ and the protective effects of CB₂ activation

on a nephrotoxic model of AKI in cisplatin-induced renal injury. $^{35,59-61}$ Inhibiting CB₁ receptor 35 or activating CB₂ receptor 59,60 limited oxidative stress and inflammation and reduced tubular damage in kidneys of animals with cisplatin-induced AKI. In addition, ß-Caryophyllene, a natural agonist of CB₂ receptor, dose-dependently protected against the deleterious effects of cisplatin-induced nephrotoxicity. 61

Furthermore, CB₁ and CB₂ receptors have been shown to play a role in renal apoptotic and inflammatory signaling pathways. 35,59,60 Indeed, activating CB₁ receptors is known to result in enhanced expression of oxidative/nitrosative stress markers, which activate p38, MAPK, and c-Jun N-terminal kinase pathways, as well as nuclear factor kappa-light-chain-enhancer of activated B cells-dependent transcription of downstream proinflammatory target genes. Ultimately, the activation of either route leads to apoptotic cell death and inflammation in the kidney.³⁵ Conversely, CB₂ receptor activation has been found to reduce proapoptotic signaling^{59,60} and mediate anti-inflammatory effects by attenuating immune cell infiltrates and inflammatory cytokine release.⁵⁹ Mukhopadhyay et al.⁵⁹ showed that a peripherally restricted CB2 receptor agonist (LEI-101), in a mouse model of cisplatin-induced nephrotoxicity, dependently attenuated renal dysfunction as measured by serum concentrations of blood urea nitrogen (BUN) and creatinine. The protective effects of CB₂ receptor activation in these studies were absent in CB₂ receptor knockout mice, suggesting that CB₂ receptors are a promising therapeutic target for reducing renal inflammation, oxidative/nitrosative stress, and apoptosis.

Another major contributor to AKI, which is associated with significant morbidity and mortality, is sepsisassociated kidney injury (SA-AKI).^{62,63} In a study using a cecal ligation and puncture (CLP) mouse model of sepsis, CB₂ receptor knockout mice demonstrated increased mortality, lung injury, bacteremia, neutrophil recruitment, and decreased p38 MAPK activity at the site of infection. 64 Treatment with a selective CB2 receptor agonist reduced the effects caused by CLP, such as inflammation, lung damage, and neutrophil recruitment, and ultimately improved survival.⁶⁴ These findings are in line with evidence demonstrating that following CB2 localization to leukocytes, their activation has been shown to mitigate leukocyte tumor necrosis factor-α-induced endothelial cell activation, adhesion and migration of leukocytes, as well as proinflammatory modulators. 65-68 Therefore, CB₂ receptor modulation may represent a novel therapeutic target in the treatment of SA-AKI.^{5,69,70}

The mechanism(s) by which the cannabinoid receptors modulate or recover tubular cell survival following acute damage are not well defined at this time. However, molecular differences in cannabinoid receptor mRNA and protein levels^{20,35,71} as well as differences in the physiological outcome of receptor activation are likely related to the type of AKI and to the abundance and localization of receptors.

EC ligands in renal health and disease. While many of the studies evaluating the role of the EC system in renal homeostasis and pathophysiology focused on CB receptors and their modulation, it is important to keep in mind that the overall effects of activation and inhibition of the EC system are dependent on various factors, only a portion of which is related to the activity of CB receptors. For example, the chief endogenous activators of the CB receptors, AEA and 2-AG, are present in substantial concentrations in the kidney^{8,9}; however, physiological responses elicited by these ligands under normal or pathological conditions have not been fully elucidated. Furthermore, detailed studies on how elevated or decreased levels of these ligands may impact renal function and pathology are scarce. For example, it is well known that AEA plays a role in the modulation of renal hemodynamics. ^{17,31} Infusion of this ligand was found to be associated with vasorelaxation of juxtamedullary afferent arterioles in vitro, 31 increased renal blood flow in rodents, 17 and alteration of tubular sodium transport. 18 While these effects may be partly mediated through the activation of CB1 and CB₂ receptors, it is important to highlight that these findings indicate the total effect of this ligand and it is difficult to identify exactly which receptors are activated in each segment of the nephron. Furthermore, there are CB receptor-independent effects that are not accounted for when the role of these ligands were to be assessed only in the context of CB receptors.

Recent studies have begun to address this important point by attempting to define the impact of these ligands in renal disease states. Biernacki et al.⁷² described alterations to the EC system in primary and secondary HTN, noting that these conditions resulted in renal oxidative stress through increased reactive oxygen species (ROS) and diminished levels of antioxidant enzymes. Despite the enhanced activity of FAAH and MGL in primary and secondary hypertensive rats, the levels of AEA and 2-AG in the kidney were significantly increased.⁷² Increasing endogenous levels of AEA by pharmacologically inhibiting its degradative enzyme,

FAAH, with a selective FAAH inhibitor, URB597, was found to have resulted in the inhibition of ROS generation in both types of hypertensive rats. These effects were mediated through improvement in antioxidant defense in the primary spontaneously hypertensive rat (SHR) kidney via the Nrf2 pathway, as well as through reduced proinflammatory responses in secondary hypertensive (DOCA-salt) rats.⁷² Furthermore, URB597 augmented ROS-dependent phospholipid peroxidation products and levels of ECs in both types of hypertensive kidneys, which resulted in enhanced CB receptor expression in SHR rats and enhanced expression of CB2 and TRPV1 receptors in DOCA-salt rats.⁷² Chronic treatment of Wistar normotensive control rats with URB597 similarly enhanced phospholipid oxidation in the kidney, comparable to its administration in DOCA-salt rats.⁷² Thus, while the EC system appears to play a protective role in HTN, the administration of a FAAH inhibitor did not significantly alter the proinflammatory or oxidative conditions caused by primary HTN, and only created imbalances between ECs, oxidants, and proinflammatory factors in secondary HTN, potentially leading to the development of kidney dysfunction.

With regard to other renal conditions, such as AKI, studies have shown varied responses to kidney injury in EC expression levels. Moradi et al. ⁷³ demonstrated that renal ischemia/reperfusion injury is associated with a significant increase in renal 2-AG content using a bilateral ischemia/reperfusion mouse model of AKI. It was found that the augmentation of kidney 2-AG concentrations following MGL inhibitor administration resulted in improved serum BUN, creatinine, and tubular damage score; however, the mRNA gene expression of renal inflammation and oxidative stress markers was not altered. Conversely, in a cisplatin-induced nephrotoxic model of AKI, cisplatin enhanced AEA but not 2-AG levels in renal tissue. ³⁵

To date, the mechanisms and conditions under which CB receptors are activated by ECs in the kidney—and subsequently the signaling cascades that result from this activation—have not been fully described. Studies have demonstrated conflicting results describing the role of AEA and CB₁ receptor activation in mediating glomerular podocyte injury. Jourdan et al.⁷⁴ showed that chronic exposure of human cultured podocytes to high glucose resulted in a significant upregulation in CB₁ receptor gene expression, which is also associated with an increase in cellular AEA and 2-AG. This is associated with signs of inflammation and podocyte injury, which manifest as

decreased podocin and nephrin and increased desmin gene expression.⁷⁴ In contrast, Li et al.⁷⁵ reported the protective functions of AEA following L-homocysteine (Hcys)-induced podocyte injury. AEA blocked Hcys-induced NLRP3 inflammasome activation in cultured podocytes and ameliorated podocyte dysfunction, ultimately precluding glomerular damage.⁷⁵ Therefore, while the former study demonstrated that an increase in CB₁ receptor gene expression accompanied by an upregulation in AEA and 2-AG is associated with podocyte injury, the latter study suggests that AEA exerts protective and anti-inflammatory effects in podocytes. Future studies are needed to investigate the role of EC ligands in CB receptor activation under varied conditions in renal health and disease.

Conclusion

The EC system has been found to regulate a variety of functions in renal health and disease states. Various components of the EC system, namely the CB₁ and CB₂ receptors and their major physiologic activators (AEA and 2-AG), have been localized to a variety of renal cell subtypes across different species. Consequently, the activation or inhibition of CB₁ and CB₂ can significantly impact renal function with beneficial or adverse effects. Altered CB receptor expression has been demonstrated in a number of renal diseases, including nephropathy, CKD, and AKI. These findings have led to the investigation of CB receptor manipulation using pharmacological agents, which have partly pointed to the CB receptors as potential therapeutic targets for renal dysfunction. An important result of these studies was the demonstration that CB₁ and CB₂ receptors act via separate pathways and modulate distinct downstream targets in the kidney, despite a largely homogenous distribution in the renal system.

More recently, the EC system has been studied for its association with a variety of renal disease states. Collectively, these studies suggest that the activity of ECs should be examined separately from their interactions with CB receptors, as conflicting results were seen in the biological responses elicited by ECs and the activation of their receptors.

In summary, significant focus has been placed on evaluating the role of CB receptors in renal function, homeostasis, and pathophysiology. While these endeavors have contributed significantly to our understanding of the role of the EC system in the kidney, important areas of opportunity remain for future research, especially the role of EC ligands as mediators of EC system

activity. At present, their role in renal physiology and pathophysiology remains to be fully elucidated. Furthermore, the clinical implications and relevance of EC system alteration will need to be further evaluated. Thus, while current data suggest that modulating EC system function and activity may provide a viable therapeutic intervention for renal dysfunction, future studies are essential to further elucidate the mechanisms through which ECs and CB receptors participate in renal physiology and disease, as well as the clinical context in which their stimulation or suppression could lead to beneficial or deleterious effects in the kidney.

Acknowledgments

The content of this article is the sole responsibility of the authors and in no way should be seen as official policy or interpretation by the US Department of Veterans Affairs or the US government. H.M. is supported by a career development award from the Office of Research and Development of the Department of Veterans Affairs, 1 IK CX 001043-01A2.

Author Disclosure Statement

No competing financial interests exist.

References

- Howlett AC. The cannabinoid receptors. Prostaglandins Other Lipid Mediat. 2002;68–69:619–631.
- DiPatrizio NV, Piomelli D. The thrifty lipids: endocannabinoids and the neural control of energy conservation. Trends Neurosci. 2012;35:403–411.
- 3. Pacher P, Batkai S, Kunos G. The endocannabinoid system as an emerging target of pharmacotherapy. Pharmacol Rev. 2006;58:389–462.
- Hillard CJ. Circulating endocannabinoids: from whence do they come and where are they going? Neuropsychopharmacology. 2018;43:155–172.
- Munro S, Thomas KL, Abu-Shaar M. Molecular characterization of a peripheral receptor for cannabinoids. Nature. 1993;365:61–65.
- Fowler CJ. The pharmacology of the cannabinoid system—a question of efficacy and selectivity. Mol Neurobiol. 2007;36:15–25.
- Beltramo M, Stella N, Calignano A, et al. Functional role of high-affinity anandamide transport, as revealed by selective inhibition. Science. 1997; 277:1094–1097.
- Kondo S, Kondo H, Nakane S, et al. 2-Arachidonoylglycerol, an endogenous cannabinoid receptor agonist: identification as one of the major species of monoacylglycerols in various rat tissues, and evidence for its generation through CA2+-dependent and -independent mechanisms. FEBS Lett. 1998;429:152–156.
- Ritter JK, Li G, Xia M, et al. Anandamide and its metabolites: what are their roles in the kidney? Front Biosci (Schol Ed). 2016;8:264–277.
- Hryciw DH, McAinch AJ. Cannabinoid receptors in the kidney. Curr Opin Nephrol Hypertens. 2016;25:459–464.
- Howlett AC, Blume LC, Dalton GD. CB(1) cannabinoid receptors and their associated proteins. Curr Med Chem. 2010;17:1382–1393.
- Pertwee RG. The pharmacology of cannabinoid receptors and their ligands: an overview. Int J Obes (Lond). 2006;30 Suppl 1:S13–S18.
- Larrinaga G, Varona A, Perez I, et al. Expression of cannabinoid receptors in human kidney. Histol Histopathol. 2010;25:1133–1138.
- Osei-Hyiaman D, DePetrillo M, Pacher P, et al. Endocannabinoid activation at hepatic CB1 receptors stimulates fatty acid synthesis and contributes to diet-induced obesity. J Clin Invest. 2005;115:1298–1305.
- 15. Jenkin KA, McAinch AJ, Zhang Y, et al. Elevated cannabinoid receptor 1 and G protein-coupled receptor 55 expression in proximal tubule cells

- and whole kidney exposed to diabetic conditions. Clin Exp Pharmacol Physiol. 2015;42:256–262.
- Jenkin KA, O'Keefe L, Simcocks AC, et al. Chronic administration of AM251 improves albuminuria and renal tubular structure in obese rats. J Endocrinol. 2015;225:113–124.
- Koura Y, Ichihara A, Tada Y, et al. Anandamide decreases glomerular filtration rate through predominant vasodilation of efferent arterioles in rat kidneys. J Am Soc Nephrol. 2004;15:1488–1494.
- Silva GB, Atchison DK, Juncos LI, et al. Anandamide inhibits transportrelated oxygen consumption in the loop of Henle by activating CB1 receptors. Am J Physiol Renal Physiol. 2013;304:F376–F381.
- Barutta F, Corbelli A, Mastrocola R, et al. Cannabinoid receptor 1 blockade ameliorates albuminuria in experimental diabetic nephropathy. Diabetes. 2010:59:1046–1054.
- 20. Lecru L, Desterke C, Grassin-Delyle S, et al. Cannabinoid receptor 1 is a major mediator of renal fibrosis. Kidney Int. 2015;88:72–84.
- Janiak P, Poirier B, Bidouard JP, et al. Blockade of cannabinoid CB1 receptors improves renal function, metabolic profile, and increased survival of obese Zucker rats. Kidney Int. 2007;72:1345–1357.
- Lin CL, Hsu YC, Lee PH, et al. Cannabinoid receptor 1 disturbance of PPARgamma2 augments hyperglycemia induction of mesangial inflammation and fibrosis in renal glomeruli. J Mol Med (Berl). 2014;92:779–792.
- Nam DH, Lee MH, Kim JE, et al. Blockade of cannabinoid receptor 1 improves insulin resistance, lipid metabolism, and diabetic nephropathy in db/db mice. Endocrinology. 2012;153:1387–1396.
- Jourdan T, Szanda G, Rosenberg AZ, et al. Overactive cannabinoid 1 receptor in podocytes drives type 2 diabetic nephropathy. Proc Natl Acad Sci U S A. 2014;111:E5420–E5428.
- Barutta F, Piscitelli F, Pinach S, et al. Protective role of cannabinoid receptor type 2 in a mouse model of diabetic nephropathy. Diabetes. 2011; 60:2386–2396.
- Jenkin KA, McAinch AJ, Grinfeld E, et al. Role for cannabinoid receptors in human proximal tubular hypertrophy. Cell Physiol Biochem. 2010;26:879– 886.
- Tam J, Cinar R, Liu J, et al. Peripheral cannabinoid-1 receptor inverse agonism reduces obesity by reversing leptin resistance. Cell Metab. 2012; 16:167–179.
- 28. Sampaio LS, Taveira Da Silva R, Lima D, et al. The endocannabinoid system in renal cells: regulation of Na(+) transport by CB1 receptors through distinct cell signalling pathways. Br J Pharmacol. 2015;172:4615–4625.
- Lim JC, Lim SK, Han HJ, et al. Cannabinoid receptor 1 mediates palmitic acid-induced apoptosis via endoplasmic reticulum stress in human renal proximal tubular cells. J Cell Physiol. 2010;225:654–663.
- Lim JC, Lim SK, Park MJ, et al. Cannabinoid receptor 1 mediates high glucose-induced apoptosis via endoplasmic reticulum stress in primary cultured rat mesangial cells. Am J Physiol Renal Physiol. 2011;301:F179– F188.
- Deutsch DG, Goligorsky MS, Schmid PC, et al. Production and physiological actions of anandamide in the vasculature of the rat kidney. J Clin Invest. 1997;100:1538–1546.
- Lee SF, Newton C, Widen R, et al. Differential expression of cannabinoid CB(2) receptor mRNA in mouse immune cell subpopulations and following B cell stimulation. Eur J Pharmacol. 2001;423:235–241.
- Jenkin KA, O'Keefe L, Simcocks AC, et al. Renal effects of chronic pharmacological manipulation of CB2 receptors in rats with diet-induced obesity. Br J Pharmacol. 2016;173:1128–1142.
- Jenkin KA, McAinch AJ, Briffa JF, et al. Cannabinoid receptor 2 expression in human proximal tubule cells is regulated by albumin independent of ERK1/2 signaling. Cell Physiol Biochem. 2013;32:1309–1319.
- 35. Mukhopadhyay P, Pan H, Rajesh M, et al. CB1 cannabinoid receptors promote oxidative/nitrosative stress, inflammation and cell death in a murine nephropathy model. Br J Pharmacol. 2010;160:657–668.
- Koga D, Santa T, Fukushima T, et al. Liquid chromatographic-atmospheric pressure chemical ionization mass spectrometric determination of anandamide and its analogs in rat brain and peripheral tissues. J Chromatogr B Biomed Sci Appl. 1997;690:7–13.
- 37. Matias I, Petrosino S, Racioppi A, et al. Dysregulation of peripheral endocannabinoid levels in hyperglycemia and obesity: effect of high fat diets. Mol Cell Endocrinol. 2008;286(1–2 Suppl 1):566–578.
- Long JZ, LaCava M, Jin X, et al. An anatomical and temporal portrait of physiological substrates for fatty acid amide hydrolase. J Lipid Res. 2011; 52:337–344.

- 39. Ritter JK, Li C, Xia M, et al. Production and actions of the anandamide metabolite prostamide E2 in the renal medulla. J Pharmacol Exp Ther. 2012:342:770-779.
- 40. Hsu YC, Lei CC, Shih YH, et al. Induction of proteinuria by cannabinoid receptors 1 signaling activation in CB1 transgenic mice. Am J Med Sci. 2015;349:162-168.
- 41. Li J, Wang DH. Differential mechanisms mediating depressor and diuretic effects of anandamide. J Hypertens. 2006;24:2271-2276.
- 42. Randall MD, Alexander SP, Bennett T, et al. An endogenous cannabinoid as an endothelium-derived vasorelaxant. Biochem Biophys Res Commun. 1996;229:114-120.
- 43. Zakrzeska A, Schlicker E, Baranowska M, et al. A cannabinoid receptor, sensitive to O-1918, is involved in the delayed hypotension induced by anandamide in anaesthetized rats. Br J Pharmacol. 2010;160:574-584.
- 44. Decleves AE, Sharma K. New pharmacological treatments for improving renal outcomes in diabetes. Nat Rev Nephrol. 2010;6:371-380.
- 45. Johnson DW, Jones GR, Mathew TH, et al. Chronic kidney disease and measurement of albuminuria or proteinuria: a position statement. Med J Aust. 2012:197:224-225.
- 46. Bermudez-Siva FJ, Serrano A, Diaz-Molina FJ, et al. Activation of cannabinoid CB1 receptors induces glucose intolerance in rats. Eur J Pharmacol. 2006;531:282-284.
- 47. Lafontan M, Piazza PV, Girard J. Effects of CB1 antagonist on the control of metabolic functions in obese type 2 diabetic patients. Diabetes Metab. 2007:33:85-95.
- 48. Barutta F, Grimaldi S, Franco I, et al. Deficiency of cannabinoid receptor of type 2 worsens renal functional and structural abnormalities in streptozotocin-induced diabetic mice. Kidney Int. 2014;86:979-990.
- 49. Zoja C, Locatelli M, Corna D, et al. Therapy with a selective cannabinoid receptor type 2 agonist limits albuminuria and renal injury in mice with type 2 diabetic nephropathy. Nephron. 2016;132:59-69.
- 50. Abrass CK. Overview: obesity: what does it have to do with kidney disease? J Am Soc Nephrol. 2004;15:2768-2772.
- 51. de Jong PE, Verhave JC, Pinto-Sietsma SJ, et al. Obesity and target organ damage: the kidney. Int J Obes Relat Metab Disord. 2002;26 Suppl 4:S21-
- 52. Briffa JF, Grinfeld E, McAinch AJ, et al. Short term exposure to elevated levels of leptin reduces proximal tubule cell metabolic activity. Mol Cell Endocrinol. 2014;382:38-45.
- 53. Briffa JF, McAinch AJ, Poronnik P, et al. Adipokines as a link between obesity and chronic kidney disease. Am J Physiol Renal Physiol. 2013;305: F1629-F1636.
- 54. Kaseda R, lino N, Hosojima M, et al. Megalin-mediated endocytosis of cystatin C in proximal tubule cells. Biochem Biophys Res Commun. 2007; 357:1130-1134.
- 55. Udi S, Hinden L, Earley B, et al. Proximal tubular cannabinoid-1 receptor regulates obesity-induced CKD. J Am Soc Nephrol. 2017;28:3518-3532.
- 56. Feizi A, Jafari MR, Hamedivafa F, et al. The preventive effect of cannabinoids on reperfusion-induced ischemia of mouse kidney. Exp Toxicol Pathol. 2008;60:405-410.
- 57. Fouad AA, Al-Mulhim AS, Jresat I. Cannabidiol treatment ameliorates ischemia/reperfusion renal injury in rats. Life Sci. 2012;91:284-292.
- 58. Nettekoven M, Adam JM, Bendels S, et al. Novel triazolopyrimidinederived cannabinoid receptor 2 agonists as potential treatment for inflammatory kidney diseases. Chem Med Chem. 2016;11:179-189.
- 59. Mukhopadhyay P, Baggelaar M, Erdelyi K, et al. The novel, orally available and peripherally restricted selective cannabinoid CB2 receptor agonist LEI-101 prevents cisplatin-induced nephrotoxicity. Br J Pharmacol. 2016; 173:446-458
- 60. Mukhopadhyay P, Rajesh M, Pan H, et al. Cannabinoid-2 receptor limits inflammation, oxidative/nitrosative stress, and cell death in nephropathy. Free Radic Biol Med. 2010;48:457-467.
- 61. Horvath B, Mukhopadhyay P, Kechrid M, et al. beta-Caryophyllene ameliorates cisplatin-induced nephrotoxicity in a cannabinoid 2 receptordependent manner, Free Radic Biol Med. 2012;52:1325-1333.
- 62. Angus DC, van der Poll T. Severe sepsis and septic shock. N Engl J Med. 2013;369:840-851.
- Swaminathan S, Rosner MH, Okusa MD. Emerging therapeutic targets of sepsis-associated acute kidney injury. Semin Nephrol. 2015;35:38-54.
- 64. Tschop J, Kasten KR, Nogueiras R, et al. The cannabinoid receptor 2 is critical for the host response to sepsis. J Immunol. 2009;183:499–505.

- 65. Sardinha J, Kelly ME, Zhou J, et al. Experimental cannabinoid 2 receptormediated immune modulation in sepsis. Mediators Inflamm. 2014;2014: 978678.
- 66. Rajesh M, Mukhopadhyay P, Batkai S, et al. CB2-receptor stimulation attenuates TNF-alpha-induced human endothelial cell activation, transendothelial migration of monocytes, and monocyte-endothelial adhesion. Am J Physiol Heart Circ Physiol. 2007;293:H2210-H2218.
- 67. Klein TW. Cannabinoid-based drugs as anti-inflammatory therapeutics. Nat Rev Immunol. 2005;5:400-411.
- Ramirez SH, Hasko J, Skuba A, et al. Activation of cannabinoid receptor 2 attenuates leukocyte-endothelial cell interactions and blood-brain barrier dysfunction under inflammatory conditions. J Neurosci. 2012;32:4004-4016.
- 69. Pacher P, Mechoulam R. Is lipid signaling through cannabinoid 2 receptors part of a protective system? Prog Lipid Res. 2011;50:193-211.
- 70. Klein TW, Newton C, Friedman H. Cannabinoid receptors and the cytokine network. Adv Exp Med Biol. 1998;437:215-222.
- 71. Soares RZ, Vuolo F, Dall'Igna DM, et al. Evaluation of the role of the cannabidiol system in an animal model of ischemia/reperfusion kidney injury. Rev Bras Ter Intensiva. 2015;27:383-389.
- 72. Biernacki M, Ambrozewicz E, Gegotek A, et al. Redox system and phospholipid metabolism in the kidney of hypertensive rats after FAAH inhibitor URB597 administration. Redox Biol. 2018;15:41-50.
- 73. Moradi H, Oveisi F, Khanifar E, et al. Increased renal 2arachidonoylglycerol level is associated with improved renal function in a mouse model of acute kidney injury. Cannabis Cannabinoid Res. 2016;1: 218-228
- 74. Jourdan T, Park JK, Varga ZV, et al. Cannabinoid-1 receptor deletion in podocytes mitigates both glomerular and tubular dysfunction in a mouse model of diabetic nephropathy. Diabetes Obes Metab. 2018;20:698-708.
- 75. Li G, Xia M, Abais JM, et al. Protective Action of Anandamide and its COX-2 metabolite against I-homocysteine-induced NLRP3 inflammasome activation and injury in podocytes. J Pharmacol Exp Ther. 2016;358:61-70.
- 76. Park F, Potukuchi PK, Moradi H, et al. Cannabinoids and the kidney: effects in health and disease. Am J Physiol Renal Physiol. 2017;313:F1124-F1132.

Cite this article as: Chua JT, Argueta DA, DiPatrizio NV, Kovesdy CP, Vaziri ND, Kalantar-Zadeh K, Moradi H (2019) Endocannabinoid system and the kidneys: from renal physiology to injury and disease, Cannabis and Cannabinoid Research 4:1, 10-20, DOI: 10.1089/ can.2018.0060.

Abbreviations Used

2-AG = 2-arachidonovl-sn-glycerol

AEA = anandamide

AKI = acute kidney injury

BUN = blood urea nitrogen

CB₁ = cannabinoid subtype-1

 $CB_2 = cannabinoid subtype-2$ CKD = chronic kidney disease

CLP = cecal ligation and puncture

EC = endocannabinoid

FAAH = fatty acid amide hydrolase

GFR = glomerular filteration rate

Hcys = L-homocysteine

HTN = hypertension

MGL = monoacylglycerol lipase

ROS = reactive oxygen species

SA-AKI = sepsis-associated kidney injury

SHR = spontaneously hypertensive rat

STZ = streptozotocin

 $\mathsf{TAL} \!=\! \mathsf{thick} \; \mathsf{ascending} \; \mathsf{limbs}$

UUO = unilateral ureteral obstruction

VEGF = vascular endothelial growth factor

ZDF = Zucker diabetic fatty