

Ent-kaurane Diterpenoids from *Isodon rubescens* var. *lushanensis*

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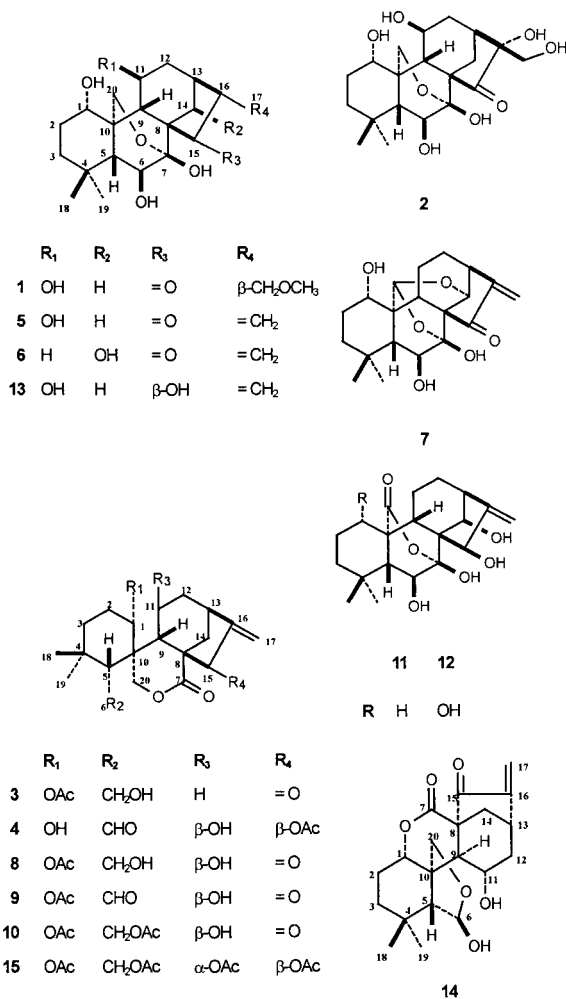
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Four new *ent*-kaurane diterpenoids lushanrubescensins F–I (1–4), together with 11 known ones, lasiodonin (5), oridonin (6), ponacidin (7), isodonoil (8), isodonal (9), rabdosin B (10), rabdoternins A and B (11 and 12), enmenol (13), epinodosin (14), and inflexusin (15), were isolated from *Isodon rubescens* var. *lushanensis*, and the structures were elucidated by spectroscopic analysis. The inhibitory effect against the K562, Bcap37, BGC823, BIU87, CA, CNE, and Hela cell lines of compounds 3 and 5–10 were evaluated.

Key words *Isodon rubescens* var. *lushanensis*; Labiatae; *ent*-kauranoid; lushanrubescensin F; lushanrubescensin I; cytotoxicity

Isodon rubescens (HELSM.) HARA (Labiatae), a perennial herb of the *Isodon* genus, which is a rich natural source of bioactive *ent*-kaurane diterpenoids,¹ is native to the Yellow River valley of China and used as an antitumor folk medicine in the treatment of esophageal and cardiac carcinoma in Henan Province, China. This medicinal plant was developed into a drug, which is in clinical use for its antibacterial and antiinflammatory activities.² Many phytochemical studies focused on this plant and its varieties, leading to the isolation of more than 20 *ent*-kaurane diterpenoids.³ From *I.*

rubescens var. *lushanensis* collected in Lushan Prefecture of Henan Province, five 20-nonoxygenated *ent*-kaurane diterpenoids, lushanrubescensins A–E, were reported previously,^{4–7} differing greatly from 7,20-epoxy or 6,7-seco *ent*-kauranoids isolated from *I. rubescens* and its varieties collected in other prefectures. In our reinvestigation of the constituents of *I. rubescens* var. *lushanensis*, 15 *ent*-kaurane diterpenoids including four new compounds (1–4) were isolated, none of which was a 20-nonoxygenated *ent*-kaurane diterpenoid. Compounds 3 and 5–10 were evaluated for their inhibitory effect against K562 and Bcap37 (human breast cancer cell lines), CA (human liver cancer cell line), CNE (human nasopharyngeal cancer cell line), BIU87 (human cystic cancer cell line), BGC823 (human stomach cancer cell line), and Hela cell lines. Herein, we report the isolation and structure elucidation as well as the cytotoxicity evaluation results.



Results and Discussion

Compound 1, obtained as colorless needles, showed the molecular ion peak at m/z 396.2165 in its high resolution electron impact (HR-EI)-MS, corresponding to the molecular formula C₂₁H₃₂O₇. This was corroborated by the ¹³C- and distortionless enhancement by polarization transfer (DEPT)-NMR spectra, which exhibited signals for all 20 carbons of the diterpene skeleton in addition to a methoxyl group.

On the basis of the characteristic signals of three methines (δ_C 61.5, 58.3, 29.8 due to C-5, 9, and 13), three quaternary carbons (δ_C 61.1, 42.5, 34.2 assignable to C-8, 10, and 4), two methyls (δ_C 32.9 and 22.3 attributable to C-18 and 19), an oxygenated methylene (δ_C 64.6, assigned as C-20), and a hemiketal quaternary carbon (δ_C 95.5, C-7), along with the structures of the compounds isolated previously from this plant, we assumed that compound 1 should be a 7,20-epoxy-*ent*-kaurane diterpenoid similar to lasiodonin (5),^{8,9} which was also isolated as one of the major constituents. Comparison of the NMR data between the two compounds confirmed the above deduction, and indicated that compound 1 was identical to 5 except for C-16 and C-17. The exomethylene group at C-16 of 5 was replaced by a methine (δ_C 58.4, C-16; δ_H 2.95, H-16) and an oxygenated methylene (δ_C 68.9, C-17; δ_H 3.75 and 3.62, H₂-17) in 1. Furthermore, the additional methoxyl was located on this oxygenated methylene carbon by the heteronuclear multiple bond connectivity (HMBC)

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correlation (Fig. 1) between the methoxyl protons (δ_{H} 3.19) with this carbon. Therefore, compound **1** was determined to be 16(*S*)-1 α ,6 β ,7 β ,11 β -tetrahydroxy-17-methoxy-7 α ,20-epoxy-*ent*-kaur-15-one by the nuclear Overhauser effect (NOE) (Fig. 1) between H-17 with H-12 β (δ_{H} 1.28) in the rotating frame Overhauser enhancement spectroscopy (ROESY) spectrum of **1**, and called lushanrubescensin F. All the NMR data of **1** were unambiguously assigned based on the two dimensional (2D)-NMR spectra including ^1H - ^1H correlation spectroscopy (COSY), ^1H -detected heteronuclear multiple quantum coherence (HMQC), HMBC, and ROESY.

Compound **2**, obtained as a white amorphous powder, exhibited a quasimolecular ion peak at m/z 399.2035 in its positive HR-FAB-MS in agreement with the molecular formula

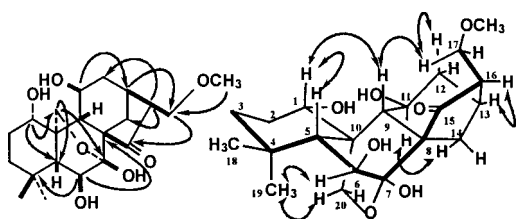


Fig. 1. Key HMBC (from H to C) and ROESY Correlations of **1**

$\text{C}_{20}\text{H}_{31}\text{O}_8$. The analyses of the ^1H - and ^{13}C -NMR data of **2** indicated that compound **2** was also a 7,20-epoxy-*ent*-kauranoid. On the basis of the comparison of the ^{13}C -NMR spectrum of **2** with that of **1** (Table 1), it was revealed that these two compounds were very similar except for two more hydroxyl groups and one less methoxyl group in **2** than in **1**. One of these hydroxyl groups was assigned to C-17 by the HMBC interactions of H₂-17 (δ_{H} 4.46) with C-15 (δ_{C} 223.3) and C-13 (δ_{C} 37.8). The other hydroxyl group was located at C-16 by the HMBC correlations arising from H₂-17 and H₂-14 (δ_{H} 3.51 and 2.50) with C-16 (δ_{C} 82.7). Furthermore, the relative configuration of C-16 was determined to be *R* by the NOE between H₂-17 with H-12 β (δ_{H} 2.42). Consequently, compound **2** was elucidated to be 16(*R*)-1 α ,6 β ,7 β ,11 β ,16,17-hexahydroxy-7 α ,20-epoxy-*ent*-kaur-15-one, and called lushanrubescensin G.

Compound **3**, obtained as colorless needles, showed an $[\text{M}+\text{H}]^+$ ion peak at m/z 391.2129 in its positive HR-FAB-MS, consistent with the molecular formula $\text{C}_{22}\text{H}_{31}\text{O}_6$ which was confirmed by its ^{13}C -NMR spectrum presenting signals for all 22 carbons of the molecular formula including two carbons of an acetoxy group. On the basis of the characteristic lactone carbonyl signal at δ_{C} 170.9 (s) due to C-7 and no-

Table 1. ^1H - and ^{13}C -NMR Data of Compounds **1**–**4** (500 and 125 MHz, in $\text{C}_5\text{D}_5\text{N}$, δ in ppm, J in Hz)

	1		2		3		4	
	δ_{H}	δ_{C}	δ_{H}	δ_{C}	δ_{H}	δ_{C}	δ_{H}	δ_{C}
1 β	4.16 ^{a)}	73.2 d	4.18 m	73.4 d	5.02 dd, 3.0, 10.6	77.0 d	5.69 dd, 2.8, 10.0	76.2 d
2 α	2.80 m	28.3 t	2.82 m	28.5 t	2.00–1.80 ^{a)}	24.3 t	2.05 m	24.4 t
2 β	1.84 m		1.86 m		2.00–1.80 ^{a)}		1.99 m	
3 α	1.43 br d, 12.4	39.8 t	1.42 m	39.9 t	1.33–1.40 ^{a)}	39.9 t	1.33 m	40.3 t
3 β	1.32 br t, 12.4		1.31 m		1.33–1.40 ^{a)}		1.45 m	
4		34.2 s		34.4 s		33.8 s		34.4 s
5 β	1.50 d, 4.8	61.5 d	1.53 d, 5.6	61.2 d	1.72 br s	53.5 d	4.41 s	61.5 d
6 α	4.16 ^{a)}	74.3 d	4.26 dd, 5.6, 10.0	74.7 d	3.82, 2H, br s	58.8 t	10.43 br s	205.1 d
6 β		95.5 s		95.8 s		170.9 s		175.4 s
8		61.1 s		61.2 s		58.6 s		53.3 s
9 β	1.86 ^{a)}	58.3 d	2.04 d, 11.2	58.3 d	3.20 dd, 4.0, 13.0	42.2 d	3.19 d, 10.0	42.6 d
10		42.5 s		42.7 s		44.3 s		43.8 s
11 α	4.79 m	63.3 d	4.88 m	63.5 d	1.80 ^{a)}	17.8 t	4.38 m	65.5 d
11 β					1.50 m			
12 α	1.75 br t, 12.8	30.4 t	2.81 m	32.3 t	1.30 ^{a)}	30.1 t	2.59 m	44.7 t
12 β	1.22 ^{a)}		2.42 m		1.33 ^{a)}		1.69 br t, 10.0	
13 α	2.50 br s	29.8 d	3.03 m	37.8 d	2.91 m	35.2 d	2.69 m	36.0 t
14 α	2.44 m	29.2 t	2.50 br d, 12.4	26.5 t	2.04 br d, 12.0	29.2 t	1.85 br d, 12.0	30.6 t
14 β	1.89 ^{a)}		3.51 dd, 2.4, 12.4		2.58 dd, 4.8, 12.0		2.28 dd, 5.0, 12.0	
15 α		224.3 s		223.3 s		202.5 s	4.92 br s	83.5 d
16	2.95 m	58.4 d		82.7 s		151.3 s		159.7 s
17	3.75 dd, 4.0, 10.0 3.62 dd, 8.0, 10.0	68.9 t	4.46 2H, s	63.7 t	5.96 s 5.34 s	118.3 t	5.45 s 5.20 s	109.6 t
18	1.17 3H, s	32.9 q	1.19 3H, s	33.1 q	0.99 3H, s	33.5 q	0.93 3H, s	33.3 q
19	1.07 3H, s	22.3 q	1.10 3H, s	22.5 q	0.78 3H, s	23.5 q	1.05 3H, s	24.1 q
20	4.64 d, 10.8 4.39 d, 10.8	64.6 t	4.75 d, 10.0 4.45 d, 10.0	64.7 t	5.12 d, 12.0 4.84 d, 12.0	68.9 t	5.51 d, 12.0 5.26 d, 12.0	67.1 t
1-OH	6.48 s		6.69 s					
6-OH	6.33 d, 11.0		6.59 d, 10.0					
7-OH	8.82 s							
11-OH	6.48 s							
1-OAc					2.16 s	170.2 s 21.4 q		
15-OAc							2.18 s	170.4 s 21.4 q

a) Overlapping signals.

ticeable oxygenated methylene signals [δ_C 68.9 (t), C-20; δ_H 5.12 and 4.84 (d, $J=12.0$ Hz), H₂-20], compound **3** was inferred to be a 6,7-seco-7,20-olide *ent*-kauranoid, similar to isodonoiol (**8**),^{10,11} a known analogue also isolated in this study. Through comparison of the 1D-NMR data of two compounds (**3** and **8**), it was indicated that compound **3** was 11-dehydroxy-isodonoiol, which was confirmed by the HMBC and ROESY correlations of **3** (Fig. 2). Thus compound **3** was finally established to be 6-hydroxy-1 α -acetoxy-6,7-seco-7,20-olide-*ent*-kaur-16-en-15-one, and called lushanrubescensin H.

Compound **4**, an amorphous powder, was also indicated to be a 6,7-seco-7,20-olide-*ent*-kaurane diterpenoid by the analyses of its 1D-NMR spectra, with the molecular formula C₂₂H₃₀O₇ determined by the HR-EI-MS (Found 406.1976, Calcd 406.1992). By the comparison of the ¹³C-NMR data of **4** with those of isodonal (**9**),^{10,11} a known 6,7-seco-7,20-olide-*ent*-kauranoid obtained at the same time, it was revealed that **4** differed from **9** mainly at C-15. An acetoxy group was confirmed to be linked with C-15 of **4** by the HMBC correlations of H-15 (δ_H 4.92) with the acetoxy carbonyl carbon (δ_C 170.4), C-9 (δ_C 42.6), and C-17 (δ_C 109.6), and H₂-17 (δ_H 5.45 and 5.20) with C-15 (δ_C 83.5). This acetoxy group was further indicated to be β -orientated by the ROESY correlation between H-15 with H-14 β (δ_H 2.28). Thus compound **4** was assigned to be 1 α -hydroxy-15 β -acetoxy-6-oxo-6,7-seco-7,20-olide-*ent*-kaur-16-ene, and called lushanrubescensin I.

The structures of known compounds **5**–**15** were established to be lasiodonin (**5**),^{8,9} isodonoiol (**8**),^{10,11} isodonal (**9**),^{10,12} rabdoternins A and B (**11** and **12**),¹³ oridonin (**6**),¹⁴ ponigidin (**7**),¹⁵ radosin B (**10**),¹⁶ enmenol (**13**),¹⁷ epinododin (**14**),¹⁸ and inflexusin (**15**),¹⁹ by comparison of their spectral data with literature values. The reinvestigation results were very different from those reported in the literature,⁴ in which five 20-nonoxygenated *ent*-kauranoids had been reported from *I. rubescens* var. *lushanensis* collected in Lushan Prefecture of Henan Province, while the 15 *ent*-kauranoids obtained in our study from the same plant

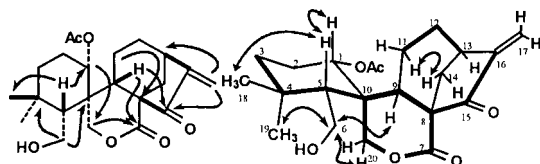


Fig. 2. Key HMBC (from H to C) and ROESY Correlations of **3**

collected in the same prefecture were 20-oxygenated. The different results showed that the structural types of secondary metabolites change with the different ecological environment or habitat of *I. rubescens* and its varieties.

Considering the application of this medical plant in drug production in China, we think it is necessary to perform further chemical and phytotaxological studies on this plant.

Compounds **3** and **5**–**10** were tested for their inhibitory effects against K562, Bcap37, BGC823, BIU87, CA, CNE, and Hela cell lines, with cisplatin as the positive reference substance. The results are presented in Table 2. Compounds **3**, **7** and **9** showed abilities to inhibit K562 cells comparable to that of cisplatin, while compounds **6** and **7** exhibited significant inhibitory effects against the CA cell line. But none of these compounds displayed an obvious inhibitory effect against Bcap37, BGC823, BIU87, CNE, and Hela cell lines.

Experimental

General Procedures Melting points were measured on an XRC-1 micro melting point apparatus and are uncorrected. Optical rotations were measured on a JASCO DIP-370 digital polarimeter. UV absorptions were obtained on a Shimadzu UV-2401PC UV-VIS recording spectrophotometer. IR spectra were determined on a Bio-Rad FTS-135 spectrophotometer with KBr pellets. MS were recorded on a VGAuto Spec-3000 spectrometer. 1D- and 2D-NMR spectra were run on Bruker AM-400 and DRX-50 instruments with tetramethylsilane (TMS) as an internal standard.

Plant Material The leaves of *I. rubescens* var. *lushanensis* were collected in Lushan Prefecture of Hennan Province in August 2000 and air-dried. The identity of the plant material was verified by Prof. Zhong-Wen Lin, and a voucher specimen (KIB-2000-10 Lin) is deposited in the Herbarium of the Department of Taxonomy, Kunming Institute of Botany, Chinese Academy of Science.

Extraction and Isolation The dried and powdered leaves (8.0 kg) were extracted with 70% Me₂CO and filtered. The filtrate was concentrated and partitioned successively between petroleum ether and water and then EtOAc and water. The EtOAc extract (370 g) was applied to column chromatography over a silica gel (100–200 mesh, 3.5 kg) column eluted with a system of CHCl₃–Me₂CO (10:0, 9:1, 8:2, 7:3, 6:4, 5:5). The CHCl₃ fraction was further chromatographed repeatedly over silica gel to afford **10** (80 mg) and **15** (10 mg). Then repeated chromatography of the CHCl₃–Me₂CO (9:1) fraction over silica gel yielded **14** (3.4 g), **3** (8.0 mg), **4** (16.0 mg), **5** (800 mg), **6** (3.0 g), and **7** (120 mg). Compounds **8** (20 mg), **9** (40 mg), **11** (5.0 mg), and **12** (2.0 mg) were isolated from the CHCl₃–Me₂CO (8:2) fraction in the same way. The CHCl₃–Me₂CO (7:3) fraction was similarly chromatographed on silica gel and refined with preparative TLC to yield compounds **1** (6.0 mg), **2** (2.0 mg), and **13** (20.0 mg).

Compound 1: Colorless needles, mp 202–203 °C. [α]_D²⁰ –77.3° ($c=0.11$, acetone). UV λ_{max} (MeOH) nm (log ϵ): 210 (2.91). IR (KBr) ν_{max} cm⁻¹: 3408, 2941, 2907, 1717, 1635, 1456, 1392, 1194, 1170, 1089, 1055. ¹H- and ¹³C-NMR (C₅D₅N) data: see Table 1. EI-MS m/z (%): 396 [M]⁺ (6), 378 (8), 364 (100), 346 (7), 328 (5), 300 (4). HR-EI-MS m/z : 396.2165 (Calcd for C₂₁H₃₂O₇, 396.2148).

Compound 2: White amorphous powder, [α]_D²⁰ –40.2° ($c=0.06$, acetone).

Table 2. Inhibitory Effects of Compounds **3** and **5**–**10** against Various Cell Lines

Compound	IC ₅₀ (μg/ml) for cell lines						
	K562	Bcap37	BGC823	BIU87	CA	CNE	Hela
3	3.56	13.42	8.91	ND	8.25	ND	>100
5	5.35	112.53	a)	a)	a)	a)	a)
6	4.37	8.32	ND	55.91	0.06	16.50	28.67
7	2.26	6.76	55.17	13.26	0.06	13.26	11.31
8	10.15	101.32	a)	a)	a)	a)	a)
9	2.29	28.64	79.87	ND	9.04	ND	>100
10	4.61	15.84	10.93	ND	>100	ND	>100
Cisplatin	3.84	1.54	2.54	4.34	0.88	6.54	3.60

a) Not tested. ND: not determined.

UV λ_{\max} (MeOH) nm (log ϵ): 212 (3.26). IR (KBr) ν_{\max} cm^{-1} : 3428, 2925, 1771, 1716, 1682, 1651, 1621, 1558, 1540, 1520, 1457, 1397, 1374. ^1H - and ^{13}C -NMR ($\text{C}_5\text{D}_5\text{N}$) data: see Table 1. Positive FAB-MS m/z : 399 $[\text{M}+1]^+$. Positive HR-FAB-MS m/z : 399.2035 (Calcd for $\text{C}_{20}\text{H}_{31}\text{O}_8$, 399.2019).

Compound 3: Colorless needles, mp 201—202 °C. $[\alpha]_{\text{D}}^{20} + 31.8^\circ$ ($c=0.32$, acetone). UV λ_{\max} (MeOH) nm (log ϵ): 229 (3.81). IR (KBr) ν_{\max} cm^{-1} : 3450, 1745, 1714, 1672, 1450, 1333. ^1H - and ^{13}C -NMR ($\text{C}_5\text{D}_5\text{N}$) data: see Table 1. Positive FAB-MS m/z : 391 $[\text{M}+1]^+$. Positive HR-FAB-MS m/z : 391.2129 (Calcd for $\text{C}_{22}\text{H}_{31}\text{O}_6$, 391.2121).

Compound 4: Amorphous powder, $[\alpha]_{\text{D}}^{20} + 45.2^\circ$ ($c=1.99$, MeOH). UV λ_{\max} (MeOH) nm (log ϵ): 212 (3.22). IR (KBr) ν_{\max} cm^{-1} : 3444, 2950, 2873, 1735, 1710, 1464, 1410, 1371, 1296, 1239, 1125. ^1H - and ^{13}C -NMR ($\text{C}_5\text{D}_5\text{N}$) data: see Table 1. EI-MS m/z (%): 404 $[\text{M}-2]^+$ (2), 386 (20), 358 (4), 344 (6), 326 (6), 298 (10), 148 (40), 107 (100). HR-EI-MS m/z : 406.1976 (Calcd for $\text{C}_{22}\text{H}_{30}\text{O}_7$, 406.1992).

Cytotoxicity against Human Tumor K562 Cells A cytotoxicity assay against the K562 cell line was performed using the improved MTT [3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-tetrazolium bromide] colorimetric method.²⁰ K562 cells at the log phase of their growth cycle (4×10^4 cells/ml) were added to each well (90 μl /well), then treated in five replicates at various concentrations of the samples (100, 10, 1, 0.1, 0.01 $\mu\text{g}/\text{ml}$), and incubated for 48 h at 37 °C in a humidified atmosphere of 5% CO_2 . Thereafter each well was treated with MTT solution 10 μl (5 mg/ml) and further incubated for 4 h. Then, a three-system solution of 10% SDS–5% isobutanol–0.012 ml/l HCl was added to each well (100 μl /well). The OD of each well was recorded at room temperature 12 h later on an ELISA reader (Bioteck EL-340, U.S.A.) at two wavelengths (570, 630 nm). In these experiments, the negative reference agents were isochoric normal saline, 1% DMSO, or 0.1% DMSO, and cisplatin was used as the positive reference substance at three concentrations (10, 1, and 0.1 $\mu\text{g}/\text{ml}$).

Cytotoxicity against Bcap37, BGC823, BIU87, CA, CNE, and HeLa Cell Lines Exponentially growing Bcap37, BGC823, BIU87, CA, CNE, or HeLa cells were dissociated with 0.25% trypsin in D-Hanks buffer. They were formed into a single-well suspension of density (4×10^4 cells/ml) by RPMI-1640 containing 10% heat-inactivated bovine serum. Ninety microliters of the cell suspension were added to each well of the 96-well microplate. Cultures were preincubated for 24 h in a 5% CO_2 atmosphere and 100% humidity incubator. After that, 10 μl of the test solution was added to each well of the 96-well microplate (each concentration was tested in quadruplicate wells). Then cells were incubated at 37 °C in a 5% CO_2 atmosphere for an additional 72 h. At the end of exposure, the cells were fixed by the addition of 50 μl of cold 50% trichloroacetic acid (TCA) at 4 °C for 1 h. After fixation, cultures were washed five times with tap water. When the plates were air-dried, TCA-fixed cells in each well were stained with a 0.4% 50 μl sulforhodamin B (SRB) solution in 1% acetic acid for 30 min. At the

end of the staining period, SRB was removed and cultures were rinsed four times with 1% acetic acid to remove unbound dye. After being rinsed, the cultures were air-dried until no standing moisture was visible. Then 10 mm unbuffered Tris solution (150 μl) was added to each well. Plates were agitated for 5 min on a gyratory shaker. Then the OD was read on a microplate reader at a wavelength of 570 nm.

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