

RESEARCH

Open Access



Environment shapes the fecal microbiome of invasive carp species

Jessica J. Eichmiller¹, Matthew J. Hamilton², Christopher Staley², Michael J. Sadowsky^{1,2*} and Peter W. Sorensen^{1*}

Abstract

Background: Although the common, silver, and bighead carps are native and sparsely distributed in Eurasia, these fish have become abundant and invasive in North America. An understanding of the biology of these species may provide insights into sustainable control methods. The animal-associated microbiome plays an important role in host health. Characterization of the carp microbiome and the factors that affect its composition is an important step toward understanding the biology and interrelationships between these species and their environments.

Results: We compared the fecal microbiomes of common, silver, and bighead carps from wild and laboratory environments using Illumina sequencing of bacterial 16S ribosomal RNA (rRNA). The fecal bacterial communities of fish were diverse, with Shannon indices ranging from 2.3 to 4.5. The phyla *Proteobacteria*, *Firmicutes*, and *Fusobacteria* dominated carp guts, comprising 76.7 % of total reads. Environment played a large role in shaping fecal microbial community composition, and microbiomes among captive fishes were more similar than among wild fishes. Although differences among wild fishes could be attributed to feeding preferences, diet did not strongly affect microbial community structure in laboratory-housed fishes. Comparison of wild- and lab-invasive carps revealed five shared OTUs that comprised approximately 40 % of the core fecal microbiome.

Conclusions: The environment is a dominant factor shaping the fecal bacterial communities of invasive carps. Captivity alters the microbiome community structure relative to wild fish, while species differences are pronounced within habitats. Despite the absence of a true stomach, invasive carp species exhibited a core microbiota that warrants future study.

Keywords: Fecal microbiome, Invasive carp, Microbiota, Community structure, 16SrRNA gene

Background

Aquatic invasive species are among the greatest threats to aquatic ecosystems. In particular, species of carp, such as the common, bighead, and silver carp, can consume large quantities of food and disrupt food chains, while potentially out-competing native species and reaching great densities in invaded ranges [1–4]. The common carp is the most widespread invasive fish in the world. It was introduced to the USA over a century ago, and it has gradually spread throughout lakes and rivers. Once established, common carp act as ecosystem engineers, uprooting aquatic vegetation and increasing phosphorus availability, resulting in eutrophication and ecosystem

degradation [5, 6]. In contrast, bighead and silver carps, members of a group known as the “Asian carps,” were recently introduced into the USA in the 1970s. Their range, however, is rapidly expanding and now stretches from the lower Mississippi River to its northern reaches and tributaries.

Although the potential ecosystem impacts of silver and bighead carp are not well known, as large filter feeders they could potentially affect native fish species directly by inducing changes in zooplankton communities [7, 8]. While the management of common carp has occasionally been successful through poisoning entire lakes or manipulating weaknesses in their life history characteristics (e.g., manipulation of predators in spawning habitats) [9], no control strategies have been successful in reducing the population of silver and bighead carps. Novel options for control must therefore come

* Correspondence: sadowsky@umn.edu; soren003@umn.edu

¹Department of Fisheries, Wildlife, and Conservation Biology, Minnesota Aquatic Invasive Species Research Center, University of Minnesota, Twin Cities, Saint Paul, MN 55108, USA

Full list of author information is available at the end of the article

through a more thorough understanding of the biology of these invasive fishes.

The microbiome, the collection of microorganisms associated with an animal, is essential for optimal growth and survival of the host species [10–12]. In particular, the digestive tract microbiota plays an integral role in the breakdown of food, provision of energy, vitamin production, and shaping innate immunity. In humans and other mammals, the gut microbiota has been shown to have broad effects on health and behavior [13]. In fish, the gut microbiota has largely been studied in the context of aquaculture in order to identify or examine the effect of probiotics to enhance growth or health [14–16]. A better understanding of the gut microbiome of fishes, however, might reveal potential for control of these invasive species, since dysbiosis of the gut microbiome has been found to contribute to disease manifestation in humans and other vertebrates [17]. Manipulation of gut microbiota to influence health has recently received greater attention in humans through procedures such as fecal microbiota transplantation [18, 19]. Thus, characterizing the gut microbiome of invasive fishes is an important step toward understanding the community that comprises this “hidden organ,” which might be eventually exploited for species control purposes.

Fish possess a gut microbiome that is distinct from other animals and the microbial communities of water and soil [20]. Previous studies have shown that the fish gut microbiome is dominated by members of the phyla *Proteobacteria*, *Firmicutes*, *Bacteroidetes*, *Actinobacteria*, and *Fusobacteria* [20], and both trophic level and salinity predominantly influence the fish gut microbial community [20–22]. While diet can also affect the gut microbiome, the significance and magnitude of the effect are variable [23–25]. The microbiota of prey items has been shown to influence the gut microbiome in three-spined stickleback; however, host genotype exhibited a larger effect [26]. Gut microbiome diversity was inversely related with dietary diversity in two species of freshwater fishes [27], whereas the effect of diet on Trinidadian guppies was negligible [28]. The gut microbiome can also reflect relative preference for cyanobacteria as a food source [29]. In silver carp, the gut microbiome has also been shown to be geographically and temporally variable [29].

Like other vertebrates, fish likely harbor a core microbiome. Roeselers et al. [30] identified a core microbiome of zebrafish through comparison of lab-raised and wild stocks. Further support of this concept was demonstrated in a reciprocal transplant of microbiota between zebrafish and mice [31]. After transplantation, the microbial community gradually shifted to resemble the typical structure of its new host. However, habitat changes, such as the transition from wild to captive environments can lead to dramatic changes in the gut microbiome of

fishes, including decreased gut microbiome diversity [25, 28, 32].

Although our understanding of the structure of the fish microbiome has increased in recent years, there are still important gaps in our current knowledge regarding the factors that shape the fish gut microbiome. The advent of metagenomics and high-throughput amplicon sequencing technologies has demonstrated that culture-based studies of the fish microbiome are inherently biased and do not reflect total community diversity [14, 16]. In the first study of carp using high-throughput sequencing, van Kessel et al. [33] found that nearly half of the sequences in captive carp belonged to the phylum *Fusobacteria*, which were conspicuously absent in culture-dependent studies. However, of the studies that have utilized metagenomic approaches to examine the gut microbiome of carp species, only two have included wild fish [22, 29]. In addition, only one of these studies was done in an invaded range. Ye et al. [29] compared the microbiome of silver carp to gizzard shad, a native fish species that is planktivorous, but does not share the same taxonomic order. This study found differences in microbiota between fish species that could be explained by gut morphology and feeding preferences. Additional research is needed to compare the gut microbiome among closely related species of carps in their invaded range and to characterize the core microbiome.

In this study, we characterized the fecal microbiomes of three species of invasive carps (silver, bighead, and common carps) and determined the relationships between microbial community structure and environment, diet, and fish species. The aims of this study were to determine (1) the effect of environment on the microbiome of invasive carps, (2) how microbial communities of fish species differ within environments and the extent to which diet plays a role, (3) to what extent a core microbiome exists among invasive carps, and (4) how differences in microbiome structure among environments and fish species affect inferred bacterial community function.

Our results shed new light on the understanding of the microbiome of invasive carps and highlight the dominant role of the environment in shaping the fish microbiome.

Methods

Sample collection

Wild fishes were collected from both river and lake habitats (Table 1). Bighead carp (*Hypophthalmichthys nobilis*), silver carp (*Hypophthalmichthys molitrix*), common carp (*Cyprinus carpio*), and freshwater drum (*Aplodinotus grunniens*) were caught from the Marseilles reach of the Illinois River, IL, USA (41° 21' 2'' N, 88° 26' 15'' W) in June and August 2013. Freshwater drums were collected for comparison to the carps because they are an abundant

Table 1 Description of fishes used in this study

Species	Common name	Habitat	Diet	Number
<i>Hypophthalmichthys nobilis</i>	Bighead carp	Laboratory	Algal feed mixture, see [72]	5
<i>Hypophthalmichthys molitrix</i>	Silver carp	Laboratory	Algal feed mixture, see [72]	5
<i>Cyprinus carpio</i>	Common carp	Laboratory	2.5-mm pellet feed (Oncor Fry, Skretting USA, Tooele, UT)	5
<i>Cyprinus carpio</i>	Common carp	Laboratory	Frozen brine shrimp (San Francisco Bay Brand, Newark, CA)	5
<i>Carassius auratus</i>	Goldfish	Laboratory	Flake food (Color Tropical Marine, Pentair Aquatic Ecosystems, Apopka, FL)	5
<i>Hypophthalmichthys nobilis</i>	Bighead carp	River	NA	19
<i>Hypophthalmichthys molitrix</i>	Silver carp	River	NA	20
<i>Cyprinus carpio</i>	Common carp	River	NA	16
<i>Cyprinus carpio</i>	Common carp	Lake	NA	13
<i>Aplodinotus grunniens</i>	Freshwater drum	River	NA	9

NA not applicable

co-occurring carnivorous fish, thus, enabling comparisons across trophic levels.

Asian carps were field-identified as either bighead or silver carp, and fin clips were taken for SNP analysis to determine species and exclude F₁ hybrids [34]. An additional 13 common carp were caught from Lotus (44° 52' 28'' N, 93° 31' 48'' W), Riley (44° 50' 10'' N, 93° 31' 17'' W), and Susan (44° 51' 5'' N, 93° 32' 27'' W) lakes in MN, USA, in August and September 2012. Four fish each were collected from Lotus and Riley lakes, while five were collected from Lake Susan.

Laboratory-housed bighead carp (*H. nobilis*), silver carp (*H. molitrix*), common carp (*C. carpio*), and goldfish (*Carassius auratus*) were also used in this study (Table 1). Common carp and goldfish were obtained from a commercial fish hatchery (Osage Catfisheries Inc., Osage Beach, MO; Hunting Creek Fisheries, PA, respectively). Bighead and silver carps were obtained from an experimental research facility (US Geological Survey, Columbia, MO, USA). Goldfish were included for comparison because they are closely related and hypothesized to have originated from a wild population of Prussian carp (*Carrassius gibelio*) and a model organism. All fish were juvenile development stage and had been housed in the laboratory for at least 6 months.

All laboratory fishes were kept at 18–20 °C, within flow-through tanks with constant aeration. Fish were fed ad libitum once daily, and food source was consistent for at least 3 weeks prior to sampling (Table 1). Fish were sampled from at a minimum of three different tanks. All fish were held in accordance with the University of Minnesota's Institutional Animal Care and Use Committee (IACUC) (Protocol: 1407-31659A).

Fecal specimens from wild fish were collected within 1 h of fish capture, whereas laboratory fishes were anesthetized in an aerated anesthetic bath (0.01 % MS-222; Syndel, CO, USA) prior to handling. Fecal specimens

were collected by manual stripping of live or recently deceased fish. Fish were stroked firmly from pelvic fins to anus, and fecal material was collected in a sterile microcentrifuge tube and stored frozen at –20 °C.

DNA extraction, PCR, and sequencing

DNA was extracted from frozen fecal samples using the QIAamp DNA Stool Mini Kit (Qiagen, Hilden, Germany) according to the manufacturer's protocol. The V6 hypervariable region of the 16S rRNA gene was amplified as previously described using a mixture of five forward primers and a barcoded reverse primer to amplify triplicate 50-μL reactions containing 25 ng of fecal DNA each [35]. PCR products were visualized on a 2 % agarose gel and purified using the QIAquick Gel Extraction Kit (Qiagen, Hilden, Germany). Replicate purified reactions were pooled, and DNA was quantified using the QuantiFluor-ST and the dsDNA System (Promega, Madison, WI, USA). Purified amplicons were pooled in equal concentrations.

DNA was sequenced using an Illumina HiSeq 2000 platform at the University of Minnesota Genomics Center (Saint Paul, MN, USA) with up to 20 pooled samples per lane for a total of six runs. Paired-end sequences (100-bp read length) were joined as previously described [19]. Sequencing results were submitted to NCBI under project number SRP071816.

Sequence processing

Sequence reads were processed using the mothur software package version 1.36.1, as previously described [35, 36]. Sequence reads that had ambiguous bases, more than one mismatch to primer sequences, homopolymers > 8 nt, and quality scores < 35 in a 50-nt window were removed. UCHIME [37] was used to identify possible chimeric sequences in mothur using the default parameters and the SILVA database release 102 of

bacterial reference sequences [38]. Sequences identified as possible chimeras were subsequently removed from the dataset. Sequences were aligned using the SILVA database [38]. The threshold for aligning the reverse complement sequence was set to 0.75, and all other settings were set to default parameters. OTUs were clustered by furthest neighbor at a 97 % similarity cutoff. OTUs were classified using a naïve Bayesian classifier and the Ribosomal Database Project (RDP) taxonomic database release 9 and mother training set 9, with a probability cutoff of 60 % [39]. Chloroplast sequences and sequences that were unclassified at the kingdom level were removed, which comprised 0.001 % of total reads. Sequences that had a frequency ≤ 10 were removed from the dataset [40], removing a total of 1.7 % of reads. The number of reads from each sample was normalized to 150,000 from a maximum of 1,841,796 by randomly subsampling.

Sequence analysis

The Shannon index of diversity (alpha diversity) was calculated for each experimental group (Table 1) using mothur. One-way ANOVA and Tukey post hoc tests were used to examine differences in OTUs observed (S_{obs}) and Shannon index of diversity using JMP, Version 10 (SAS Institute Inc., Cary, NC). The distances among environment-species groups were calculated based on Bray-Curtis dissimilarity [41]. The resulting distances were used to perform a hierarchical clustering of experimental groups using Unweighted Pair Group Method (UPGMA). Results were graphed using Tree-View v 1.6.6. Bray-Curtis dissimilarity distances were calculated among individual samples and used to perform ordination and statistical tests. Ordination was done using non-metric multidimensional scaling (NMDS) in two dimensions [42]. NMDS was done separately for lab and wild fishes. Ten iterations were performed, and the iteration resulting in the lowest stress was plotted. Analysis of molecular variance (AMOVA) was used to test for significance of observed groupings by testing whether the genetic diversity within each group was different from the pooled genetic diversity.

Analysis of similarity (ANOSIM) and Kruskal-Wallis test were done in mothur. ANOSIM was also used to compare the community composition (beta diversity) among sampling groups. Kruskal-Wallis test was used to examine significant differences in abundance of individual OTUs among groups. Results are discussed at the taxonomic level of order, due to short sequence reads [43]. All analyses were done at $\alpha = 0.05$.

Putative microbiota functions were predicted using PICRUSt [44]. OTUs were mapped to the Greengenes database version 13.5, and 85 % of genes were classified to a Tier 1 KO function. The weighted nearest

sequenced taxon index (NSTI) scores averaged 0.056 ± 0.018 , indicating a relatively good match to reference genomes (ideal NSTI ≤ 0.03). Functional predictions were assigned up to KO tier 2. Tier 1 KO were compared among fishes using one-way ANOVA and Tukey post hoc tests for wild and lab fishes separately. Student's t test was used to compare KO between wild and lab fishes and between lake and river environments for common carp. Due to numerous significant differences among groups in tier 2 KO, data were visualized using PCA. Functional classifications of chitinases and vitamin B₁₂ synthesis enzymes were compared between wild and laboratory-housed bighead carp using Student's t test. All statistical analysis of functional data was done using JMP, Version 10 (SAS Institute Inc., Cary, NC).

Results

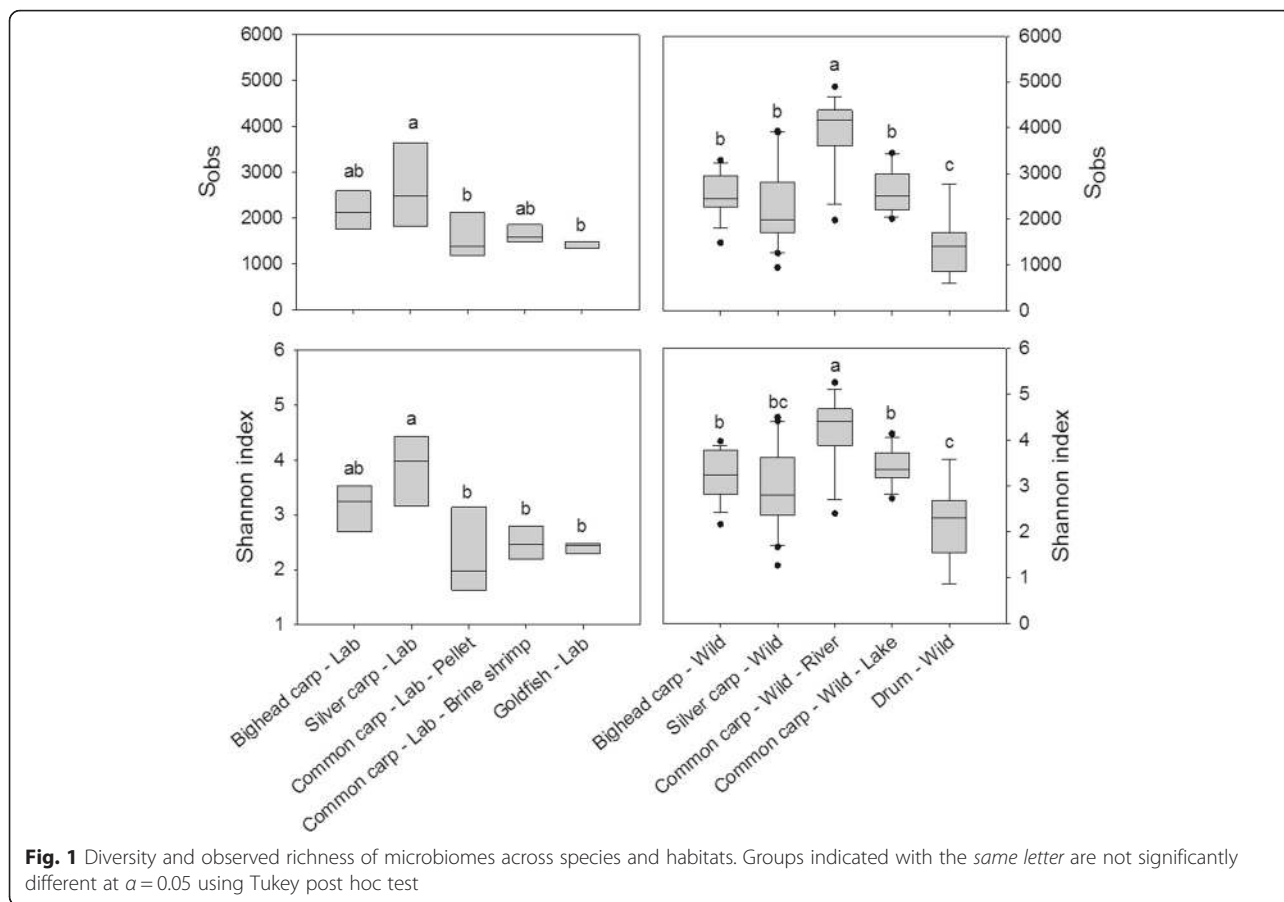
Diversity and richness

A total of 14,651 OTUs were identified across all 102 samples, with a mean coverage (estimate of total diversity that has been sampled) of $99 \% \pm 0.2 \%$ (mean \pm standard deviation) which ranged from 98 to 100 %. Observed species richness (S_{obs}) and alpha diversity, calculated using Shannon index, differed significantly among species ($p < 0.0001$, Fig. 1). Among lab fishes, silver carp had the greatest mean richness and diversity but were not significantly different than bighead carp ($p > 0.05$). Among wild fishes, common carp from the river environment had greater richness and diversity than did the other wild fishes examined ($p \leq 0.05$). Student's t test comparison between the gut microbiome of river and laboratory-housed invasive carps showed that common carp exhibited significantly higher richness ($p \leq 0.0001$) and diversity in wild fish ($p = 0.002$). Diversity was higher in captive silver carp relative to wild fish ($p = 0.04$), but richness did not differ ($p = 0.46$). Bighead carp did not show differences in either metric ($p > 0.05$).

Members of the phyla *Proteobacteria*, *Firmicutes*, and *Fusobacterium* dominated the gut microbiomes, comprising 76.9 % of total reads (Fig. 2). A portion (22.3 %) of all reads could not be classified to specific phyla, and other phyla comprised $< 1 \%$ of total reads. The most abundant orders included *Clostridiales*, *Fusobacteriales*, *Aeromonadales*, *Enterobacteriales*, *Xanthomonadales*, and *Vibrionales*. All other orders made $< 1 \%$ of total reads. The proportion of OTUs for each species at the taxonomic rank of family can be found in Additional file 1: Table S1 for families that had $> 1 \%$ of total reads.

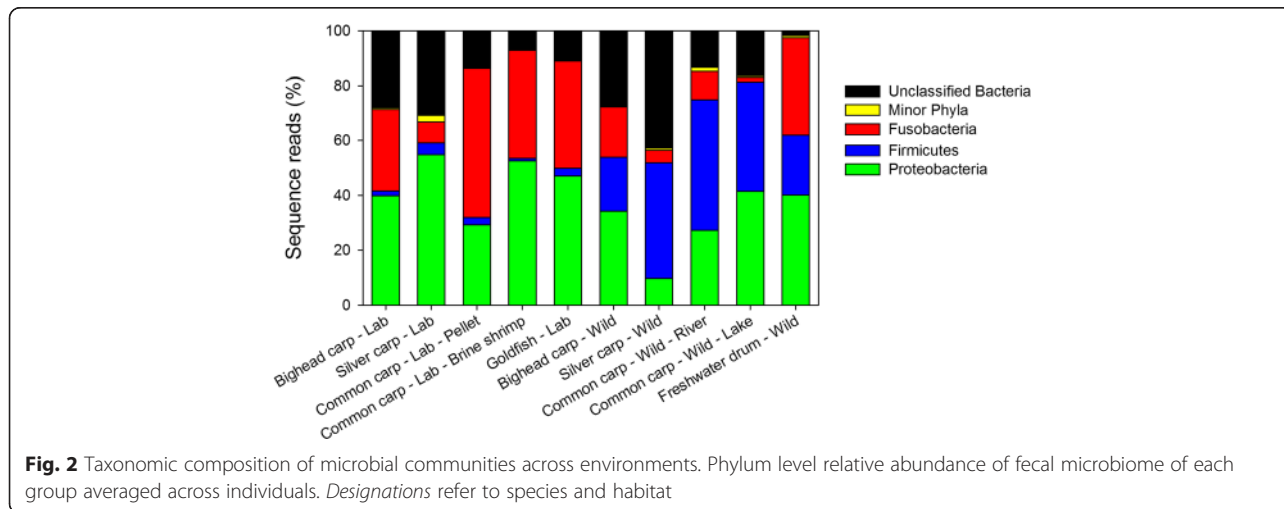
Distinct and shared OTUs

There were many OTUs that differed among fish species within and between environments. Of the 13,793 OTUs identified in wild fish, 76.6 % differed in



abundance across species ($p \leq 0.05$). However, OTUs that differed significantly among species made up 94 to 99 % of total reads per species. Among the laboratory fishes examined, only 20.5 % of the 7262 OTUs identified varied among species, making up 78 to 86 % of reads.

There were systematic differences in the order of OTUs that differed between wild and laboratory-housed fish of the same species (Fig. 3). Significantly different OTUs within *Clostridiales* were more abundant in wild fish of all species, whereas OTUs within *Fusobacteriales* were more abundant in captive bighead and common



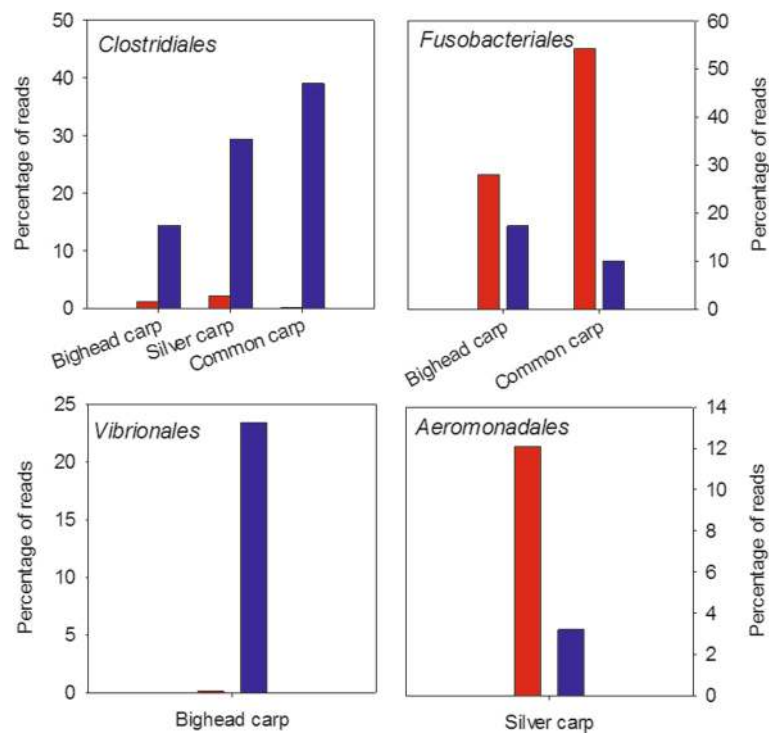


Fig. 3 Order-level classification of OTUs that varied significantly between lab (red) and wild (blue) fish by Kruskal-Wallis test at $\alpha = 0.05$. Unclassified bacteria and orders with less than 10 % of reads are not shown

carp. A large number of *Vibrionales* differed between wild and captive bighead carp, with higher abundance in wild fish. Finally, OTUs within *Aeromonadales* were higher in laboratory-housed silver carp.

Core gut microbiota must be present within a fish species across habitats [45]. We examined the OTUs that were present across wild and lab fish of the same species, and five OTUs emerged as the dominant bacteria of the core gut microbiota of invasive carps (Table 2). These bacteria were present at abundances of at least 1 % in all habitats, and they comprised, on average, approximately 40 % of the total fecal microbiome of invasive carps. There were three OTUs that were common across all invasive carps, and they were classified to the

orders *Aeromonadales*, *Xanthomonadales*, and *Fusobacteriales*. One OTU unclassified at the phylum level was common to silver and bighead carps, whereas another unclassified OTU was prevalent in bighead carp only. We used the sequence of the closest Blast_N match for both unclassified bacterial OTUs to search against the 16S ribosomal RNA database. Both sequences returned close matches to species within the *Bacteroidetes* phylum. For common carp, there was no single OTU that was prevalent, yet specific, to that species.

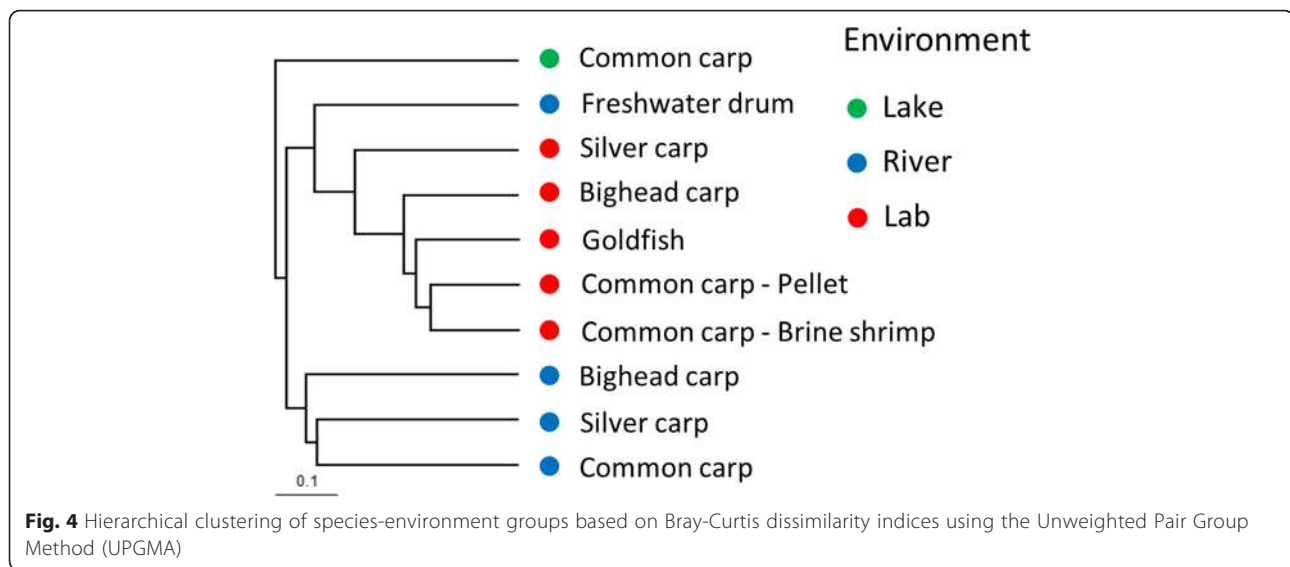
Differences in community composition

Hierarchical clustering showed that bacterial communities clustered primarily by environment (Fig. 4). Non-

Table 2 Mean relative abundance of OTUs in invasive carp species across environments

Order	Description, similarity and accession number, and source of closest NCBI Blast _N match	Abundance		
		Bighead carp	Silver carp	Common carp
Unclassified	Uncultured prokaryote, 98 % identity to KC601630, Asian seabass intestine	6.2	19.4	NA
<i>Aeromonadales</i>	<i>Psychrobacter</i> sp., 100 % identity to EU753148, marine intertidal flat	4.3	7.1	10.0
<i>Xanthomonadales</i>	Uncultured clone, 98 % identity to KM312603, earthworm gut	2.7	5.3	6.2
<i>Fusobacteriales</i>	<i>Cetobacterium</i> sp., 98 % identity to KM85610, Zebrafish intestine	21.8	5.4	23.8
Unclassified	Uncultured prokaryote, 98 % identity to KC601623, Asian seabass intestine	4.3	NA	NA
Sum		39.3	37.2	40.0

OTUs with < 1 % prevalence across all groups and species are not reported
NA not applicable



metric multidimensional scaling (NMDS) of wild fishes showed separation of most fish by species, and the lowest stress value was 0.29 with an R^2 of 0.67 (Fig. 5). Although the distribution of freshwater drum slightly overlapped with bighead and lake common carp, all groupings were significantly different (AMOVA, $p < 0.001$). The NMDS analysis was able to capture slightly more variability within the data for lab fishes (stress value = 0.23, $R^2 = 0.77$). The distribution of bighead and common carp that were fed pellets and brine shrimp overlapped (Fig. 5), and these communities did not have significantly different grouping (AMOVA, $p > 0.05$). The gut bacterial communities of silver carp and goldfish did not group with other lab fishes (Fig. 5), and both species were significantly different from other lab fish species (AMOVA, $p \leq 0.05$).

ANOSIM showed significant differences among groups in community composition and abundance (beta diversity). All wild fish species had distinct microbial communities ($p < 0.0001$), and the gut microbiota between wild and lab fish of the same species were different ($p < 0.0001$). For lab fishes, however, fewer differences were observed. Bighead carp were not significantly different from common carp fed pellet feed ($p = 0.08$) or brine shrimp ($p = 0.18$) diet. Diet did not change the bacterial community composition of common carp ($p = 0.30$). All other comparisons among lab fish were significant at $\alpha = 0.05$.

Functional analysis

Functional assignments were predicted from microbial community composition using PICRUST, which revealed differences in predicted microbial function across species and environments. The majority of predicted tier 1 KEGG

Orthology (KO) were in the functional category of metabolism (Fig. 6).

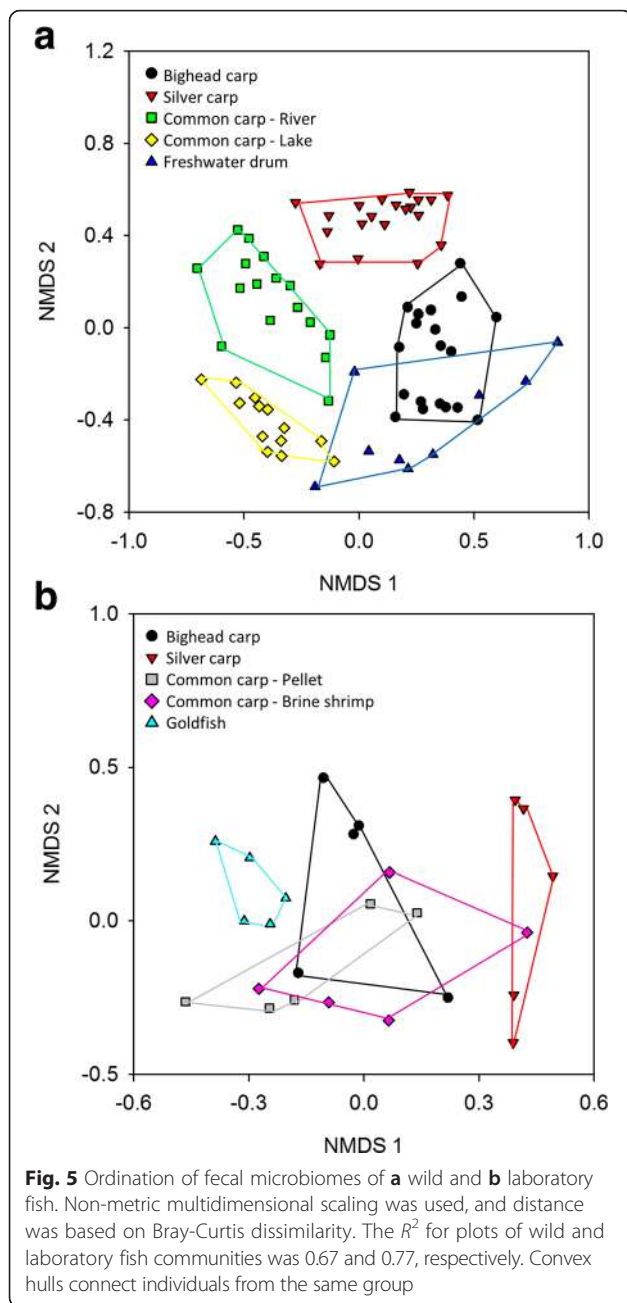
Comparisons of species within wild and lab environments were done using principal components analysis (PCA) due to numerous significant differences in tier 2 KO functions. The PCA components 1 and 2 explained a large proportion in the variation of the data (Fig. 6; wild fish = 89.9 %, lab fish = 88.4 %). Although there was overlap in the distribution of points representing the inferred function of the microbiota of wild fish, captive fish had more similarity in the predicted function of the microbiome than among wild fish. All lab fish, apart from silver carp, had overlapping distributions of the graphical representation of inferred microbiome functional data (Fig. 6). Transcription was weighted heavily in axis 1 of both wild and lab fish, indicating that this category is particularly important in the differentiation of gene functions among fish species (Table 3).

In a comparison of wild and laboratory-housed fish of the same species, there were no differences in tier 1 KO functions that were consistent across the three species. Comparisons of the proportion of proposed function for five vitamin B₁₂ biosynthesis proteins between wild and lab fishes showed some differences. For instance, there was a higher proportion of vitamin B₁₂ biosynthesis for two enzymes in wild silver carp and all five in common carp ($p \leq 0.05$). Chitinase functional classification was not different ($p = 0.20$) between wild and laboratory bighead carp.

Discussion

The role of diet and environment in shaping the fish microbiome

Bacterial community structure and function in the guts of carps was strongly affected by environment. There



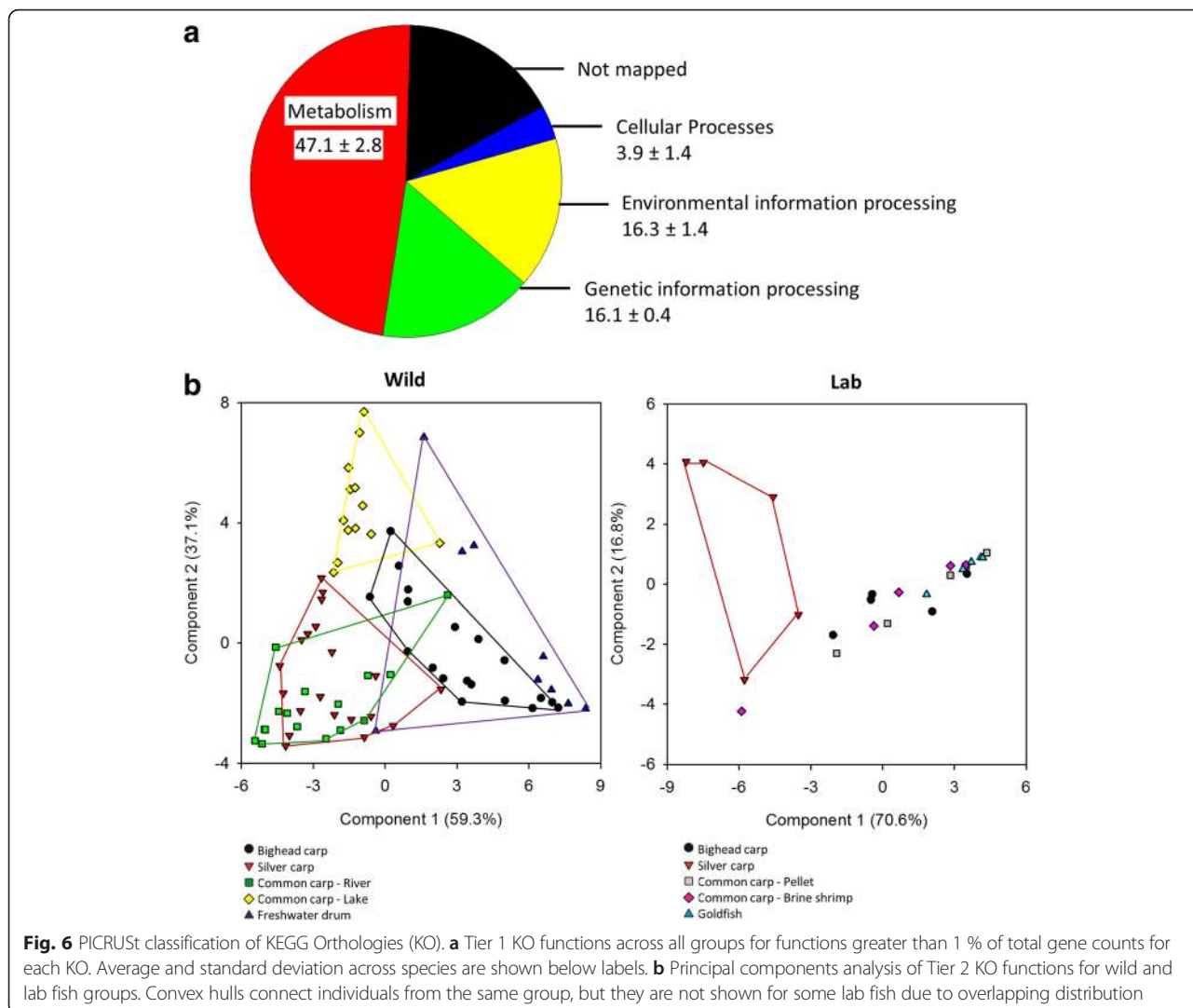
were large differences between laboratory-housed and wild fish of the same species and between common carp and lake and river environments. Many of the differences between wild and lab fishes can likely be partially explained in the context of diet, and the three taxonomic orders that varied most between wild and lab fish were *Clostridiales*, *Fusobacteriales*, *Vibrionales*, and *Aeromonadales*.

Clostridiales, which were more abundant in all wild-invasive carp species compared to those reared in the laboratory habitat, are associated with the degradation and metabolism of carbohydrates, specifically sugars [46, 47].

Clostridiaceae also aid in glucose fermentation as has been shown in earthworm guts [48] and are responsible for producing short chain fatty acids in vertebrates. Prebiotic arabinoxylan oligosaccharides have been shown to increase *Clostridium* spp. in sturgeon [23, 49]. Therefore, greater plant matter intake might increase the proportion of *Clostridiales*. Despite this supposition, however, both silver and bighead carps in the lab were fed a diet composed primarily of cyanobacteria and green algae. Moreover, feeding preference for cyanobacteria was similarly confirmed for wild silver carp [29]; thus, differences in plant matter intake cannot solely explain patterns of abundance of the *Clostridiales*.

An alternative explanation is that feed timing or availability in the wild carps resulted in changes in their gut microbiota. Transgenic common carp exhibited higher food intake and growth rate, which was associated with a greater proportion of *Clostridiales* in the gut microbiome [50]. In animals and humans, this increase is thought to be due to enhanced production of the short chain fatty acids acetate, butyrate, and propionate [50]. Although lab fish were fed to satiation, they ate only once daily during the daytime. Wild-invasive carps, in contrast, are able to consume food over a larger time frame, and they feed primarily at night [7, 51]. There is limited information on how food availability or timing affects fish gut microbiota; however, a study on Asian seabass showed starvation-induced changes in proportion of *Bacteroides* [52]. While members of the genus *Bacteroides* were not abundant in invasive carps, it is unclear what effect food limitation might have on invasive carp species, and this topic warrants further study.

Laboratory-housed carp had a much higher proportion of *Fusobacteriales* compared to wild fish. Over 95 % of *Fusobacteriales* were classified to the genus *Cetobacterium*. *Cetobacterium somerae*, a bacterium within the order *Fusobacteriales*, is a common and widely distributed species within the guts of freshwater fishes, and its prevalence is negatively correlated with dietary availability of vitamin B₁₂ (cobalamin) [53, 54]. Hence, the main role of *Cetobacterium somerae* in the fish gut is assumed to be synthesis of vitamin B₁₂ [53, 54]. The relative proportion of vitamin B₁₂ within each food type in this study is not known, so we are unable to make comparisons in the relative proportion of *Fusobacteriales* across diets. Bighead and silver carp can obtain B₁₂ from algal food sources. Cyanobacteria are capable of synthesizing vitamin B₁₂, and although many eukaryotic algae do not directly synthesize B₁₂, they have symbioses with B₁₂-producing bacteria [55, 56]. As wild common carp eat a significant amount of detritus, they may also satisfy their B₁₂ requirements through consumption of bacteria [2]. However, we found that wild common carp and silver carp had a



higher proportion of vitamin B₁₂ functional classifications than captive fish with a higher abundance of total *Fusobacteriales*.

For bighead carp, the bacterial order *Vibrionales* exhibited the greatest difference in abundance between

wild and laboratory environments. These bacteria are may be dietary in origin. *Vibrionales* are associated with exoskeletons of zooplankton [57], a major food source for bighead carps. Although silver carp consume zooplankton, such as cladocerans and copepods, these

Table 3 Axis loadings for principal components analysis of tier 2 KO functions

Wild fish		Laboratory-housed fish	
Axis 1	Axis 2	Axis 1	Axis 2
Positive	Positive	Positive	Positive
<ul style="list-style-type: none"> ○ Xenobiotics biodegradation and metabolism ○ Metabolism of terpenoids and polyketides ○ Lipid metabolism 	<ul style="list-style-type: none"> ○ Signal transduction ○ Cellular processes and signaling ○ Metabolism of other amino acids 	<ul style="list-style-type: none"> ○ Carbohydrate metabolism ○ Nucleotide metabolism ○ Replication and repair 	<ul style="list-style-type: none"> ○ Lipid metabolism ○ Metabolism of terpenoids and polyketides ○ Xenobiotics biodegradation and metabolism
Negative	Negative	Negative	Negative
<ul style="list-style-type: none"> ○ Membrane transport ○ Transcription 	<ul style="list-style-type: none"> ○ Carbohydrate metabolism ○ Translation 	<ul style="list-style-type: none"> ○ Transcription ○ Cellular processes and signaling 	<ul style="list-style-type: none"> ○ Amino acid metabolism ○ Enzyme families

Factors shown had the five largest loadings and were ≥ 1 % of inferred function

organisms make up a smaller proportion of the silver carp diet compared to bighead carp [58]. Dietary chitin has not been conclusively shown to increase the proportion of chitinase-producing bacteria in fish guts in aquaculture applications [59]; however, chitinase-producing bacteria have been isolated from many fishes, including common carp [12]. We found no difference in the proportion of chitinase function between wild and captive bighead carp, further indicating that *Vibrionales* were not contributing significantly to chitinase activity in the gut.

The fecal microbiota of common carp from lake and river habitats were different with respect to bacterial community structure, richness, and diversity. Generally, all these parameters were greater in river fish than those dwelling in lakes. Common carp exhibit similar diets in both lake and river environments. The primary component of the stomach contents of both lake and river carp is detritus, while seeds and invertebrates make up the majority of the remaining contents [1, 2, 60]. Therefore, differences are likely due to environment-specific factors rather than diet alone.

In wild fishes, patterns in relative abundance of bacterial phyla reflected feeding preferences. While silver carp and bighead carp are both filter feeders, phytoplankton makes up a larger proportion of the silver carp diet [7]. Common carp are omnivores, and their diet includes detritus, invertebrates, and plant matter [2], whereas freshwater drum are carnivorous [61, 62]

We found no consistent trends in phyla across trophic level, possibly due to the large variation in common carp between lake and river habitats.

We found a much greater proportion of unclassified bacteria than in previous studies [22, 29]. When considering the cumulative frequency of unclassified reads, most were present in high abundance (Additional file 2: Figure S1). Moreover, abundant sequences that were unclassified at the phylum level were distributed across species. For example, the three most abundant sequences are very prevalent across species (Additional file 3: Figure S2), and they are present in 88 to 99 % of individual fish sampled. While removal of sequence reads that were ≤ 10 in abundance decreased overall diversity, results of statistical analyses and beta diversity indices remained the same. Taken together, the abundance of bacteria unclassified at the phylum level and their distribution across fish species indicates that there is considerable diversity in the fish gut that is uncharacterized.

There are differences in the microbial community structure observed in the present study to previous studies on wild carps. For example, we did not observe a significant proportion of *Bacteroidetes* or *Actinobacteria* in the silver carp microbiome, as was observed by Ye et al. [29]. The differences may be due to differences in the

geographic location of the fish sampled or gut samples were taken. Generally, fish in the present study were sampled further north, and fecal material was collected by stripping rather than dissecting. Li et al. [22] found a predominance of *Proteobacteria* and comparatively few *Firmicutes* in bighead carp in their native range. However, samples were collected in the winter, when fish were under starvation conditions, which is known to alter gut microbial composition [52].

The distribution of phyla in captive carps was similar to that found in previous studies, with a few exceptions. In common carp, we found nearly 50 % of phyla classified to *Fusobacteria*, similar to van Kessel et al. [33]. The proportion of *Fusobacteria* and *Firmicutes* in bighead carps was similar to Li et al.'s [22]. However, we found a smaller proportion of *Bacteroidetes* than both studies.

Effect of captivity on the fish microbiome

Although patterns in microbiota in wild fish appeared to be linked with feeding preferences, we found that diet had little effect on fecal microbiome in a lab setting. For example, there was no difference in microbial community of common carp fed pellet or brine shrimp by any statistical measure of community composition. Previous studies have shown that diet can influence the fish gut microbiota by introduction of prey-associated microbes [26]. However, we do not have any data in this study on the microbes present in the foods ingested by fishes or the microbiome of the surrounding habitat. Intraspecific changes in the gut microbiome may also result from alteration in food metabolism, as shown in Eastern African cichlids and surgeon fishes [25, 63]. However, changes in response to diet can be slight [24] and may depend on species [64] or ecotype [28]. We also found that bighead and silver carp had different microbiota, despite being fed a similar algal feed mixture. Our results support those of Li et al. [65] who found that paddlefish and bighead carp reared in the same pond had distinct intestinal microbiota.

In lab fishes, species differences were not as pronounced as for wild fishes. For example, fewer OTUs differed in their relative abundance among lab fish species, and fewer differences among bacterial communities were observed. Temperature is a driving force in the biology and behavior of poikilotherms, such as fishes [66]. In laboratory or captive environments, fishes are not exposed to diurnal cycles in water temperature, nor is there the level of habitat complexity that is found in the natural environment. Differences in habitat use between closely related species of cichlids within a lake were associated with differences in microbiota [67]. Thus, differences in microbiota among species in the wild are likely due to a combination of the effects of diet,

habitat usage, temperature, physiology, and taxonomy. Thus, the homogeneous and homeostatic environment of the laboratory might modulate behavioral effects on microbiota, such as habitat usage.

Only common carp exhibited a difference in alpha diversity between wild and captive fish, with lower diversity in captive fish. The effect of captivity on gut microbiome diversity may be species-dependent. Captivity did not reduce the diversity of zebrafish gut microbiome [30]. However, captivity dramatically reduced in cichlid fish [25] and mummichog [68].

Our results indicate a striking effect of environment on the fecal microbiome of invasive carps and, in particular, a dramatic effect of captivity. In a previous study, few differences between the gut microbiota of wild and lab populations of zebrafish were observed [30]. But this may be due to rearing practices as the wild zebrafish were held for approximately 1 month under laboratory conditions prior to sampling, and this time frame was previously shown to alter the gut microbial community of silver carp [29]. Several studies have observed differences in microbiota of fishes in captive versus wild environments that mirror our findings. In a study of pond-reared and wild grass carp, *Fusobacteria* was more prevalent in pond fish [69]. A predominance of *Fusobacteria* was also found for grass carp, crucian carp, and bighead carp held in a rearing pond and fed a commercial feed [22]. Thus, we support the contention that environment shapes the fish gut microbiome and that a true understanding of their gut microbiota needs to come from wild-caught fish.

The core microbiome of invasive carps

Ringø and Birkbeck [45] enumerated five criteria required to be considered core gut microbiota in fishes: (1) they must be present in healthy individuals, (2) they colonize the gut at early life stages and persist throughout the lifespan of the fish, (3) they are found in both wild and cultured fish populations, (4) they are able to grow anaerobically, and (5) they are associated with the stomach, foregut, or hindgut. We identified five OTUs that comprise a large proportion of the core microbiome of silver, bighead, and common carps. These OTUs satisfied criteria 1, 3, and 5, but additional experiments would be needed to evaluate whether these OTUs satisfy all criteria.

Previous studies have shown that the proportion of bacteria that make up the core microbiome is variable [68]. In zebrafish, 21 OTUs comprise the core microbiome [30]. In rainbow trout, the core microbiota makes up over 80 % of the total community, and it is resistant to environmental factors [24]. Future work is needed to further identify and characterize the core microbiota of carps. In addition, studies should assess when these

bacteria initially colonize the gut and their degree of persistence over time. Moreover, due to the disproportionate abundance of some OTUs in bighead and silver carp, the potential for utilizing core microbiota to identify the presence of these invasive fishes in water bodies through the identification of their associated microbes be explored [70, 71].

Conclusions

Our results indicate that environment is an important factor controlling invasive carp fecal microbiota. We draw this conclusion from the difference between lab and wild fishes of the same species and the difference between lake and river habitats for common carp. Diet may partially explain some patterns in phyla abundance for wild fishes. However, in laboratory-housed fishes, diet did not exert a strong effect on fish gut microbiota, rather, fish species was the factor controlling differences among lab fishes. Future studies are needed to tease apart the multitude of factors which potentially control fish microbiota in wild populations. The mechanisms underlying differences in microbiota between lake- and river-dwelling common carp, for example, are not known. Due to the role of environment in shaping the microbiome, source-tracking markers for specific fish species should be developed from large representative samples of individuals from different geographical areas and habitat types. Studies on laboratory populations of fishes should be interpreted with caution, as lab fish have distinct microbiome structure from those of wild fishes, a pattern which is apparent, but not well characterized, in other studies. Invasive carps have a core fecal microbiome comprised primarily of five bacterial species, which make up approximately 40 % of the total fish gut microbiome. Future research is needed on the specific functional role of these organisms within the carp fecal microbiome. In addition, potential for augmenting these microbes for aquaculture applications or species detection or control should be evaluated.

Additional files

Additional file 1: Table S1. Percentage abundance of OTUs at the family level greater than 1 % of total reads. (DOCX 14 kb)

Additional file 2: Figure S1. The cumulative proportion of sequences unclassified at the phylum level with increasing frequency. (TIF 131 kb)

Additional file 3: Figure S2. Percentage of reads across samples for the three most abundant sequences that are unclassified at the phylum level. (TIF 209 kb)

Abbreviations

AMOVA, analysis of molecular variance; ANOVA, analysis of variance; IACUC, Institutional Animal Care and Use Committee; KO, KEGG Orthology; NMDS, non-metric multidimensional scaling; NSTI, nearest sequenced taxon index; OTU, operational taxonomic unit; PCA, principal components analysis; RDP, Ribosomal Database Project; UPGMA, Unweighted Pair Group Method

Acknowledgements

We thank Kevin Irons, David Wyffels, and their crew from the Illinois Department of Natural Resources for the help in obtaining fish from the Illinois River. We also thank Mary Headrick, Brett Miller, and Reid Swanson for obtaining samples from Minnesota lakes, Ratna Ghosal and Aaron Claus for providing assistance with the collection of fecal samples from fishes, James Lamer for analyzing Asian carp DNA to determine hybridization, Chanlan Chun for assistance with data analysis, and Sendréa Best for help with the preparation of samples for sequencing. The authors acknowledge the Minnesota Supercomputing Institute (MSI) at the University of Minnesota for providing resources that contributed to the research results reported within this paper.

Funding

Funding for this project was provided by the Minnesota Environment and Natural Resources Trust Fund as recommended by the Legislative-Citizen Commission on Minnesota Resources (LCCMR).

Availability of data and material

Sequencing results are available in the Sequence Read Archive (SRA) database at NCBI under BioProject ID PRJNA311219, accession number SRP071816.

Authors' contributions

PWS and MJS conceived of the study, participated in its design, and helped to draft the manuscript. MJH and CS helped to analyze the data and draft the manuscript. JJE collected and analyzed the data and drafted the manuscript. All authors read and approved the final manuscript.

Competing interests

The authors declare that they have no competing interests.

Consent for publication

Not applicable.

Ethics approval and consent to participate

All animals used in this study were treated in accordance with the approved University of Minnesota's Institutional Animal Care and Use Committee (IACUC) Protocol: 1407-31659A.

Author details

¹Department of Fisheries, Wildlife, and Conservation Biology, Minnesota Aquatic Invasive Species Research Center, University of Minnesota, Twin Cities, Saint Paul, MN 55108, USA. ²Department of Soil, Water and Climate, Biotechnology Institute, University of Minnesota, Saint Paul, MN 55108, USA.

Received: 8 February 2016 Accepted: 2 August 2016

Published online: 12 August 2016

References

- Lubinsky KS, Van Vooren A, Farabee G, Janecek J, Jackson SD. Common carp in the Upper Mississippi River. *Hydrobiologia*. 2014;154:141–53.
- Crivelli AJ. The biology of the common carp, *Cyprinus carpio* L. in the Camargue, southern France. *J Fish Biol*. 1981;18:271–90.
- Williamson CJ, Garvey JE. Growth, fecundity, and diets of newly established silver carp in the middle Mississippi River. *Trans Am Fish Soc*. 2005;134:1423–30.
- Koehn JD. Carp (*Cyprinus carpio*) as a powerful invader in Australian waterways. *Freshw Biol*. 2004;49:882–94.
- Bajer PG, Sorensen PW. Effects of common carp on phosphorus concentrations, water clarity, and vegetation density: a whole system experiment in a thermally stratified lake. *Hydrobiologia*. 2015;746:303–11.
- Breukelaar AW, Lammens E, Breteler J, Tátrai I. Effects of benthivorous bream (*Abramis brama*) and carp (*Cyprinus carpio*) on sediment resuspension and concentrations of nutrients and chlorophyll a. *Freshw Biol*. 1994;32:113–21.
- Kolar CS, Chapman DC, Courtenay WR, Housel CM, Williams JD, Jennings DP. Asian carps of the genus *Hypophthalmichthys*: A Biological Synopsis and Environmental Risk Assessment. Wahington, DC: US Fish and Wildlife Service; 2005.
- Sass GG, Hinz C, Erickson AC, McClelland NN, McClelland MA, Epifanio JM. Invasive bighead and silver carp effects on zooplankton communities in the Illinois River, Illinois, USA. *J Great Lakes Res*. 2014;40:911–21.
- Bajer PG, Chizinski CJ, Silbernagel JJ, Sorensen PW. Variation in native micro-predator abundance explains recruitment of a mobile invasive fish, the common carp, in a naturally unstable environment. *Biol Invasions*. 2012;14:1919–29.
- Human Microbiome Project Consortium. Structure, function, and diversity of the healthy human microbiome. *Nature*. 2012;486:207–14.
- O'Hara AM, Shanahan F. The gut flora as a forgotten organ. *EMBO Rep*. 2006;7:688–93.
- Ray AK, Ghosh K, Ringø E. Enzyme-producing bacteria isolated from fish gut: a review. *Aquac Nutr*. 2012;18:465–92.
- Sampson TR, Mazmanian SK. Control of brain development, function, and behavior by the microbiome. *Cell Host Microbe*. 2015;17:565–76.
- Ghanbari M, Kneifel W, Domig KJ. A new view of the fish gut microbiome: advances from next-generation sequencing. *Aquaculture*. 2015;448:464–75.
- Nayak SK. Role of gastrointestinal microbiota in fish. *Aquac Res*. 2010;41:1553–73.
- Llewellyn MS, Boutin S, Hoseinifar SH, Derome N. Teleost microbiomes: the state of the art in their characterization, manipulation and importance in aquaculture and fisheries. *Front Microbiol*. 2014;5:1–17.
- de Vos WM, de Vos EA. Role of the intestinal microbiome in health and disease: from correlation to causation. *Nutr Rev*. 2012;70:545–56.
- Khoruts A, Dicksved J, Jansson JK, Sadowsky MJ. Changes in the composition of the human fecal microbiome after bacteriotherapy for recurrent *Clostridium difficile*-associated diarrhea. *J Clin Gastroenterol*. 2010;44:354–60.
- Hamilton MJ, Weingarden AR, Unno T, Khoruts A, Sadowsky MJ. High-throughput DNA sequence analysis reveals stable engraftment of gut microbiota following transplantation of previously frozen fecal bacteria. *Gut Microbes*. 2013;4:125–35.
- Sullam KE, Essinger SD, Lozupone CA, O'Connor MP, Rosen GL, Knight R, Kilham SS, Russell JA. Environmental and ecological factors that shape the gut bacterial communities of fish: a meta-analysis. *Mol Ecol*. 2012;21:3363–78.
- Wong SD, Rawls JF. Intestinal microbiota composition in fishes is influenced by host ecology and environment. *Mol Ecol*. 2012;21:3100–2.
- Li T, Long M, Gatesoupe FJ, Zhang Q, Li A, Gong X. Comparative analysis of the intestinal bacterial communities in different species of carp by pyrosequencing. *Microb Ecol*. 2014;69:25–36.
- Geraylou Z, Souffreau C, Rurangwa E, D'Hondt S, Callewaert L, Courtin CM, Delcour JA, Buyse J, Ollevier F. Effects of arabinosyl-oligosaccharides (AXOS) on juvenile Siberian sturgeon (*Acipenser baerii*) performance, immune responses, and gastrointestinal microbial community. *Fish Shellfish Immunol*. 2012;33:718–24.
- Wong S, Waldrop T, Summerfelt S, Davidson J, Barrows F, Kenney PB, Welch T, Wiens GD, Snekvik K, Rawls JF, Good C. Aquacultured rainbow trout (*Oncorhynchus mykiss*) possess a large core intestinal microbiota that is resistant to variation in diet and rearing density. *Appl Environ Microbiol*. 2013;79:4974–84.
- Baldo L, Riera JL, Tooming-Klunderud A, Albà MM, Salzburger W. Gut microbiota dynamics during dietary shift in Eastern African cichlid fishes. *PLoS One*. 2015;10(5), e0127462.
- Smith CR, Snowberg LK, Caporaso G, Knight R, Bolnick DI. Dietary input of microbes and host genetic variation shape among-population differences in stickleback gut microbiota. *ISME J*. 2015;9:2515–26.
- Bolnick DI, Snowberg LK, Hirsch PE, Lauber CL, Knight R, Caporaso JG, Svanbäck R. Individuals' diet diversity influences gut microbial diversity in two freshwater fish (threespine stickleback and Eurasian perch). *Ecol Lett*. 2014;17:979–87.
- Sullam KE, Rubin BER, Dalton CM, Kilham SS, Flecker AS, Russell JA. Divergence across diet, time and populations rules out parallel evolution in the gut microbiomes of Trinidadian guppies. *ISME J*. 2015;9:1508–22.
- Ye L, Amberg J, Chapman D, Gaikowski M, Liu WT. Fish gut microbiota analysis differentiates physiology and behavior of invasive Asian carp and indigenous American fish. *ISME J*. 2014;8:541–51.
- Roeselers G, Mittge EK, Stephens WZ, Parichy DM, Cavanaugh CM, Guillemin K, Rawls JF. Evidence for a core gut microbiota in the zebrafish. *ISME J*. 2011;5:1595–608.
- Rawls JF, Mahowald MA, Ley RE, Gordon JI. Reciprocal gut microbiota transplants from zebrafish and mice to germ-free recipients reveal host habitat selection. *Cell*. 2006;127:423–33.
- Clements KD, Angert ER, Montgomery L, Choat JH. Intestinal microbiota in fishes: what's known and what's not. *Mol Ecol*. 2014;23:1891–8.

33. van Kessel MAHJ, Dutilh BE, Neveling K, Kwint MP, Veltman JA, Flik G, Jetten MSM, Klaren PHM, Op Den Camp HJM. Pyrosequencing of 16S rRNA gene amplicons to study the microbiota in the gastrointestinal tract of carp (*Cyprinus carpio* L.). *AMB Express*. 2011;1:41.
34. Lamer JT, Ruebush BC, Arbieva ZH, McClelland MA, Epifanio JM, Sass GG. Diagnostic SNPs reveal widespread introgressive hybridization between introduced bighead and silver carp in the Mississippi River Basin. *Mol Ecol*. 2015;24:3931–43.
35. Staley C, Unno T, Gould TJ, Jarvis B, Phillips J, Cotner JB, Sadowsky MJ. Application of Illumina next-generation sequencing to characterize the bacterial community of the Upper Mississippi River. *J Appl Microbiol*. 2013; 115:1147–58.
36. Schloss PD, Westcott SL, Ryabin T, Hall JR, Hartmann M, Hollister EB, Lesniewski RA, Oakley BB, Parks DH, Robinson CJ, Sahl JW, Stres B, Thallinger GG, Van Horn DJ, Weber CF. Introducing mothur: open-source, platform-independent, community-supported software for describing and comparing microbial communities. *Appl Environ Microbiol*. 2009;75:7537–41.
37. Edgar RC, Haas BJ, Clemente JC, Quince C, Knight R. UCHIME improves sensitivity and speed of chimera detection. *Bioinformatics*. 2011;27:2194–200.
38. Pruesse E, Quast C, Knittel K, Fuchs BM, Ludwig W, Peplies J, Glockner FO. SILVA: a comprehensive online resource for quality checked and aligned ribosomal RNA sequence data compatible with ARB. *Nucleic Acids Res*. 2007;35:7188–96.
39. Wang Q, Garrity GM, Tiedje JM, Cole JR. Naive bayesian classifier for rapid assignment of rRNA sequences into the new bacterial taxonomy. *Appl Environ Microbiol*. 2007;73:5261–7.
40. Brown SP, Veach AM, Rigdon-Huss AR, Grond K, Lickteig SK, Lothamer K, Oliver AK, Jumpponen A. Scraping the bottom of the barrel: are rare high throughput sequences artifacts? *Fungal Ecology*. 2015;13:221–225.
41. Bray JR, Curtis JT. An ordination of the upland forest communities of southern Wisconsin. *Ecol Monogr*. 1957;27:325–349.
42. Kenkel N, Orloci L. Applying metric and nonmetric multidimensional scaling to ecological studies: some new results. *Ecology*. 1986;67:919–28.
43. Mizrahi-Man O, Davenport ER, Gilad Y. Taxonomic classification of bacterial 16S rRNA genes using short sequencing reads: evaluation of effective study designs. *PLoS One*. 2013;8:18–23.
44. Langille MGI, Zaneveld J, Caporaso JG, McDonald D, Knights D, Reyes JA, Clemente JC, Burkepile DE, Vega Thurber RL, Knight R, Beiko RG, Huttenhower C. Predictive functional profiling of microbial communities using 16S rRNA marker gene sequences. *Nat Biotechnol*. 2013;31:814–21.
45. Ringø E, Birkbeck TH. Intestinal microflora of fish larvae and fry. *Aquac Res*. 1999;30:73–93.
46. Clements KD, Pasch IBY, Moran D, Turner SJ. *Clostridia* dominate 16S rRNA gene libraries prepared from the hindgut of temperate marine herbivorous fishes. *Mar Biol*. 2007;150:1431–40.
47. Zhu L, Wu Q, Dai J, Zhang S, Wei F. Evidence of cellulose metabolism by the giant panda gut microbiome. *Proc Natl Acad Sci*. 2011;108:17714–9.
48. Wüst PK, Horn MA, Drake HL. *Clostridiaceae* and *Enterobacteriaceae* as active fermenters in earthworm gut content. *ISME J*. 2011;5:92–106.
49. Geraylou Z, Souffreau C, Rurangwa E, Maes GE, Spanier KI, Courtin CM, Delcour JA, Buyse J, Ollevier F. Prebiotic effects of arabinoxylan oligosaccharides on juvenile Siberian sturgeon (*Acipenser baerii*) with emphasis on the modulation of the gut microbiota using 454 pyrosequencing. *FEMS Microbiol Ecol*. 2013;86:357–71.
50. Li X, Yan Q, Xie S, Hu W, Yu Y, Hu Z. Gut microbiota contributes to the growth of fast-growing transgenic common carp (*Cyprinus carpio* L.). *PLoS One*. 2013;8, e64577.
51. Garcia LM, Adelman IR. An in situ estimate of daily food consumption and alimentary canal evacuation rates of common carp, *Cyprinus carpio* L. *J Fish Biol*. 1985;27:487–93.
52. Xia J, Lin G, Fu G, Wan Z, Lee M, Wang L, Liu X, Yue G. The intestinal microbiome of fish under starvation. *BMC Genomics*. 2014;15:266.
53. Sugita H, Miyajima C. The vitamin B₁₂-producing ability of the intestinal microflora of freshwater fish. *Aquaculture*. 1991;92:267–76.
54. Tsuchiya C, Sakata T, Sugita H. Novel ecological niche of *Cetobacterium somerae*, an anaerobic bacterium in the intestinal tracts of freshwater fish. *Lett Appl Microbiol*. 2008;46:43–8.
55. Croft MT, Lawrence AD, Raux-Deery E, Warren MJ, Smith AG. Algae acquire vitamin B₁₂ through a symbiotic relationship with bacteria. *Nature*. 2005;438:90–3.
56. Rodionov DA. Comparative genomics of the vitamin B₁₂ metabolism and regulation in prokaryotes. *J Biol Chem*. 2003;278:41148–59.
57. Huq A, Small EB, West PA, Huq MI, Rahman, Colwell RR. Ecological relationships between *Vibrio cholerae* and planktonic crustacean copepods. *Appl Environ Microbiol*. 1983;45:275–83.
58. Sampson SJ, Chick JH, Pegg MA. Diet overlap among two Asian carp and three native fishes in backwater lakes on the Illinois and Mississippi rivers. *Biol Invasions*. 2008;11:483–96.
59. Ringø E, Zhou Z, Olsen RE, Song SK. Use of chitin and krill in aquaculture - the effect on gut microbiota and the immune system: a review. *Aquac Nutr*. 2012;18:117–31.
60. Crivelli AJ. The destruction of aquatic vegetation by carp. *Hydrobiologia*. 1981;106:37–41.
61. Jacquemin SJ, Pyron M, Allen M, Etchison L. Wabash River freshwater drum *Aplodinotus grunniens* diet: effects of body size, sex, and river gradient. *J Fish Wildl Manag*. 2014;5:133–40.
62. Dettmers JM, Wahl DH, Soluk DA, Gutreuter S. Life in the fast lane: fish and foodweb structure in the main channel of large rivers. *J N Am Benthol Soc*. 2014;20:255–65.
63. Miyake S, Ngugi DK, Stingl U. Diet strongly influences the gut microbiota of surgeonfishes. *Mol Ecol*. 2015;24:656–72.
64. Givens CE. A fish tale: comparison of the gut microbiome of 15 fish species and the influence of diet and temperature on its composition, Doctoral thesis. Athens: University of Georgia; 2007.
65. Li XM, Zhu YJ, Yan QY, Ringo E, Yang DG. Do the microbiota differ between paddlefish (*Polyodon spathala*) and bighead carp (*Aristichthy nobilis*) reared in the same pond? *Appl Microbiol*. 2014;117:1245–52.
66. Beitinger TL, Bennet WA, McCauley RW. Temperature tolerances of North American freshwater fishes exposed to dynamic changes in temperature. *Environ Biol Fishes*. 2000;58:237–75.
67. Franchini P, Fruciano C, Frickey T, Jones JC, Meyer A. The gut microbial community of midas cichlid fish in repeatedly evolved limnetic-benthic species pairs. *PLoS One*. 2014;9, e95027.
68. Givens CE, Ransom B, Bano N, Hollibaugh JT. Comparison of the gut microbiomes of 12 bony fish and 3 shark species. *Marine Ecol Prog Ser*. 2015;518:209–23.
69. Ni J, Yan Q, Yu Y, Zhang T. Factors influencing the grass carp gut microbiome and its effect on metabolism. *FEMS Microbiol Ecol*. 2014;87:704–14.
70. Yan T, Sadowsky MJ. Determining sources of fecal bacteria in waterways. *Environ Monit Assess*. 2007;129:97–106.
71. Roslev P, Bukh AS. State of the art molecular markers for fecal pollution source tracking in water. *Appl Microbiol Biotechnol*. 2011;89:1341–55.
72. Hansen A, Ghosal R, Caprio J, Claus AW, Sorensen PW. Anatomical and physiological studies of bigheaded carps demonstrate that the epibranchial organ functions as a pharyngeal taste organ. *J Exp Biol*. 2014;217:3945–54.

Submit your next manuscript to BioMed Central and we will help you at every step:

- We accept pre-submission inquiries
- Our selector tool helps you to find the most relevant journal
- We provide round the clock customer support
- Convenient online submission
- Thorough peer review
- Inclusion in PubMed and all major indexing services
- Maximum visibility for your research

Submit your manuscript at
www.biomedcentral.com/submit

