



# **Epigenetic Regulation and Dietary Control of Triple Negative Breast Cancer**

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Triple negative breast cancer (TNBC) represents a highly heterogeneous group of breast cancers, lacking expression of the estrogen (ER) and progesterone (PR) receptors, and human epidermal growth factor receptor 2 (HER2). TNBC are characterized by a high level of mutation and metastasis, poor clinical outcomes and overall survival. Here, we review the epigenetic mechanisms of regulation involved in cell pathways disrupted in TNBC, with particular emphasis on dietary food components that may be exploited for the development of effective strategies for management of TNBC.

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# INTRODUCTION

Breast cancer (BC) is the most common and deadly cancer in women worldwide, with a new case being diagnosed every 18 seconds (1, 2). Four molecular BC subtypes have been characterized, based on the expression of estrogen (ER) and progesterone (PR) receptor, and the human epidermal growth factor receptor-2 (HER2). The four molecular BC subtypes are usually classified as luminal ER positive (luminal A and Luminal B), her 2 enriched, and basal like [**Table 1**; (3)]. Luminal-A (LUM-A) are ER and/or PR positive, HER2 negative and have low proliferation markers such as Ki-67; luminal-B (LUM-B) are ER positive and/or PR positive, and either HER2 positive or negative with higher levels of Ki-67; her 2 enriched is ER and PR negative and HER2 positive; triple-negative/basal-like breast cancer (TNBC) is defined as ER and PR negative, and HER2-negative; LUM-A cancers are low-grade, tend to grow slowly and have the best prognosis, whereas LUM-B cancers generally grow slightly faster than LUM-A and LUM-B tumors and can have a worse prognosis, but they are often treated with targeted therapies against HER2 with vastly improved outcomes (4). The TNBC subtype is more common in women with *BRCA1* mutations, among pre-menopausal, and African-American and Hispanic women (5).

Hope for successful treatment and prevention of BC was sparked by the identification in 1994 of the breast cancer 1 (*BRCA1*) gene (6). However, optimism was tempered by the finding that only a minor percentage (5–10%) of all BC associated with mutations in either BRCA1 or BRCA2. Nevertheless, carriers of mutated *BRCA* susceptibility genes have a higher risk of developing BC by age 70 (65% for mutated *BRCA1* and 45% for *BRCA2*), highlighting the important role of *BRCA* genes in BC development (7). Notably, TNBC are associated with mutations in the *BRCA* genes. In addition, sporadic, non-hereditary TNBC are often characterized by reduced or lost expression of BRCA1, also called BRCAness (8, 9). Increased *BRCA1* promoter methylation has been observed

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TABLE 1 | Molecular subtypes.

Molecular subtype	ER		PR		Her 2
Luminal A	Positive	And/or	Positive		Negative
Luminal B	Positive	And/or	Positive (or negative <20% and Ki67>14%)	or	Negative
Luminal B	Positive	And/or	Positive (or negative)		Positive
Her 2 enriched	Negative	and	Negative		Positive
Basal-like (TNBC)	Negative	and	Negative		Negative

in a large number of TNBC (10), underscoring the importance of epigenetic factors contributing to the TNBC subtype.

The term epigenetics refers to external modifications that do not affect the DNA, but instead turn genes on or off through several mechanisms. Epigenetic mechanisms modulating gene expression include changes in DNA CpG methylation; histone post-translational modifications (e.g., methylation and acetylation), and expression of non-coding RNA. The study of factors, endogenous and exogenous, that modulate epigenetically the expression of genes involved in TNBC phenotype, is essential for the development of therapeutic strategies targeting TNBC. In this paper, we reviewed the mechanisms of action of endogenous factors and natural food components that modulate gene expression through epigenetic modifications, mainly DNA methylation and histone modifications, and identify possible targets for strategies of TNBC prevention or intervention.

## SOURCE OF DATA

Research data published in English-language articles from the PubMed database were used for this review. Relevant studies were retrieved through the use of "triple negative breast cancer, epigenetics, dietary compounds" as keywords in searches of the database. The compounds analyzed in the second part of the review were chosen based on number of research articles found searching for "TNBC-Compound," or "Breast Cancer-Epigenetic-Compound," where "compound" was one of the following molecules: resveratrol, genistein, curcumin, (-)Epigallocatechin 3-gallate (EGCG), or folate. For each one of these compounds, between 18 and 40 research articles were found. In the "Other compounds" section we discussed bioactive molecules found searching for "TNBC-diet" and for which literature was less abundant.

#### **TRIPLE NEGATIVE BREAST CANCER**

On average, only 15-20% of breast cancers are classified as TNBC, but have the poorest short and long-term prognosis (highest risk of local/regional recurrence, distant metastases, and cancer related mortality), largely due to lack of a targeted therapy (11). However, the percentage of TNBC varies by reproductive age being more prevalent in premenopausal women; BRCA1 mutation status; and in minority populations. For example, TNBC represent  $\sim$ 39% of all BC in African American women;  $\sim$  20% in Hispanic White women; and ~16% in non-Hispanic/Caucasian White women of the same age (5). Over 80% of TNBC are invasive ductal carcinoma, with presence of lymphocytes, with a 4 to 6-fold increased risk of metastasis to the lung and the brain, rather than the bones (12). TNBC are highly heterogeneous and they have been classified in 6 distinct subtypes based on their gene expression: basal like (BL) 1 and BL 2, characterized by expression of genes involved in cell cycle and DNA damage, and high proliferative index; immunomodulatory (IM), expressing genes of the immune cell signaling pathways; mesenchymal (M) and mesenchymal stem-like (MSL) expressing genes involved in the epithelial-mesenchymal transition (EMT); and finally the LAR subtype positive for the luminal androgen receptor (AR), therefore responsive to therapy using AR antagonists such as bicalutamide (13). An alternative classification in four TNBC subgroups, combining BL1 with BL2, and M with MLS, was proposed by Burstein et al. (14) to account for tumor impurities derived by infiltrations of stromal and immune cells. Therefore, in addition to lacking target hormone receptors for targeted therapy (i.e., tamoxifen, herceptin, etc.), the heterogenicity within the TNBC subtype further complicates the design of effective neoadjuvant therapies.

Currently, a few treatment options exist for TNBC but have limited specificity. Taxanes are microtubules stabilizers that inhibit cell division. They have been shown to be more effective in the therapy of TNBC than in hormone receptor positive BC (15). Anthracyclines inhibit RNA synthesis and they have been used alone for the treatment of TNBC patients with limited success, but with better outcomes when used in combination

Abbreviations: ACCA, acetyl CoA carboxylase; AP-1, activator protein-1; AhR, aromatic hydrocarbon receptor; APC, Adenomatous polyposis coli gene; AREG, amphiregulin; ARNT, Aromatic hydrocarbon nuclear translocator; BC, Breast Cancer; BLBC, basal-like BC; BMI1, B lymphoma Mo-MLV insertion region 1 homolog; BRCA1, breast cancer protein 1; CDK4, cyclindependent kinase-4; CYP1B1, P450 cytochrome 1B1; DIM, diindolyl-methane; DMBA, 2,4-dimethoxybenzaldehyde; DNMT, DNA methyl transferase; EGCG, (-)epigallocatechin 3-gallate; EMT, epithelial mesenchymal transition; ERK, Extracellular Signal-Regulated Kinase; EZH2, enhancer of zeste homolog 2; ER, estrogen receptor; FABP5, fatty acid-binding protein 5; FASN, fatty acid synthase; FRA, folate receptor type alpha; HAT, histone acetylase transferase; HDAC, histone deacetylase; Her2, human epidermal growth factor receptor 2; HIF1, hypoxia-inducible factor 1; HMT, histone methyl transferase; KDAC, lysine deacetylase; KAT, lysine acetyltransferase; EGFR, epidermal growth factor receptor; FGFR4, fibroblast growth factor receptor 4; IGF-1, insulin growth factor-1; LUM-A/B, ER positive luminal A/B breast cancer; MAPK, mitogen activated protein kinase; MBD, methyl binding domain protein; MTA1, metastasis associated protein 1; NF-kB, nuclear factor kappa light chain enhancer of activated B cells; NuRD, nucleosome remodeling and deacetylating unit; PARP, poly(ADP-ribose) polymerase; PDL-1, programmed death-ligand 1; PI3k/Akt/mTOR, phosphatidylinositol-3-kinase/mammalian target of rapamycin; PIP3, phosphatidylinositol-3, 4, 5-triphosphate; PKM, mammalian pyruvate kinase; PR, Progesterone Receptor; PRMT5, protein arginine methyltransferase5; PRC1, protein regulator of cytokinesis 1; PTEN, protein tyrosine phosphatase and tensin homolog; RAR, Retinoic acid receptor; RASSF-1, Ras Association Domain Family Member 1; ROS, reactive oxygen species; SAM, S-adenosylmethionine; STAT3, signal transducer and activator of transcription 3; TCDD, 2,3,7,8-tetrachlorodibenzo-p-dioxin; TEB, terminal end buds; TET 1, teneleven translocation 1; TNBC, Triple negative breast cancer; XRE, xenobiotic responsive elements.

with taxanes. Platinum agents induce cell death in BRCA1 mutant cells, due to their ability to prevent replication fork and inducing double strands breaks. Therapy with carboplatin and cisplatin (platinum compounds) was found to improve overall survival (27 vs. 8 months) in TNBC patients. The best clinical outcomes were achieved when platinum compounds were used in combination with other chemotherapeutic agents (9, 16). Epigenetic drugs, i.e., HDAC inhibitors (hydroxamic acids vorinostat, belinostat, LAQ824, panobinostat; and the benzamides: entinostat, tacedinaline, and mocetinostat) have been used in chromatin modifier therapies as adjuvants to sensitize TNBC cells. However, results have been mixed, as HDAC inhibitors induce re-expression of silenced tumor suppressors genes, but also inhibit expression of pro-apoptotic genes that down-regulate cell proliferation (17). Recently, a window-of-opportunity study examined the possibility of using valproic acid, a histone deacetylase inhibitor, as a possible neoadjuvant in TNBC patients. Even though a low number of women were available for evaluation, valproic acid treatment caused a 10% decrease in proliferation of breast tumors assessed by Ki-67 expression (18).

# REGULATION OF TUMOR SUPPRESSOR GENES IN TNBC

Basal-like BC (BLBC) are mostly associated with BRCA1 mutations, but other tumor suppressor genes, such as TP53 and PTEN (protein tyrosine phosphatase and tensin homolog), are often lost in this BC subtype. Somatic mutations of TP53 are also found in the majority of TNBC. Reduced or lost activity of these genes is likely responsible for the high level of genomic instability observed in BLBC. PTEN is activated by phosphorylation at its K163 residue and translocates to the nucleus where it mediates DNA repair and chromosomal stability. In the cytoplasm, PTEN exerts lipid phosphatase activity by dephosphorylating phosphatidylinositol-3, 4, 5-triphosphate (PIP3) to PIP2 and inhibiting the PI3k/Akt/mTOR signaling pathway and tumor growth. Histone deacetylase inhibitors activate PTEN nuclear translocation, whereas hypermethylation of PTEN promoter and loss of its activity have been observed during BC progression. The PTEN promoter contains binding sites for p53, which induces PTEN transcription. B lymphoma Mo-MLV insertion region 1 homolog (BMI1) is a protein of the polycomb group involved in epigenetic regulation and overexpressed in many cancer types including breast (19). It has been implicated in promotion of anchorage-independent growth of tumor cells in vitro (20) and clonogenic potential by facilitating ubiquitination activity of protein regulator of cytokinesis 1 (PRC1). The PTEN protein binds in the nucleus to BMI1, and this interaction inhibits PTEN expression. Consequently, BMI1 reduces PTEN's ability to inhibit Akt activation, likely through its interaction with PTEN in the nucleus, making PTEN unavailable to dephosphorylate membrane-bound PIP3 to PIP2 (21). BMI1 also interacts directly with c-Myc, which binds to an enhancer sequence in the BMI1 promoter. Other common features observed in BLBC include mutations of the tumor suppressor Rb and the oncogene K-ras, as well as increased activity of Myc and hypoxia-inducible factor  $1-\alpha$  (HIF1 $\alpha$ )/ARNT, indicating higher levels of cell proliferation (**Figure 1**).

Cancer cells metabolism is characterized by increased glucose uptake and lactate production (Warburg effect) (22). Pyruvate kinases are rate-limiting glycolytic enzymes involved in the final step of glycolysis (23). Two types of genes encode mammalian pyruvate kinase (pklr and pkm) (24). The PKM2 protein is regulated by several post-translational modifications that lead to the suppression of pyruvate kinase activity and translocation of PKM2 into the nucleus where it acts as a kinase toward specific nuclear proteins (25). It also acts as a co-activator of HIF-1a (26) and it contributes to tumorigenesis. It has been shown that PKM2 promotes angiogenesis through the activation of NF-κB/p65 and HIF-1α in diverse types of cancer (27, 28). The nuclear factor kappa light chain enhancer of activated B cells (NF-kB) family of transcription factors regulate inflammation, immune response, cell differentiation, proliferation, and survival (29), by forming protein complexes with DNA sequences at promoter regions of responsive genes. Specifically, NF-KB is frequently activated in TNBC and inhibition of NF-KB activity blocks growth of TNBC cells (30). Ma et al. (31) reported that knockdown of PKM resulted in anticancer effects against TNBC cells by reducing NF-KB activation and suggested it as a potential therapeutic strategy against TNBC cell growth. Other potential therapies leading to inactivation of NF-kB are currently being investigated (32).

# BRCA1 FUNCTION AND REGULATION

The BRCA1 protein is a main player in DNA repair through homologous recombination and non-homologous end-joining (33-35). The loss of BRCA1 function is predominantly associated with the development of breast and ovarian cancer in women (35), and breast and prostate cancer in men (36). Our group summarized the role of nuclear receptors and other transcriptional factors in regulations of BRCA1 expression (37). In particular, activation of the ER by estrogen through association with p300, leads to BRCA1 basal transcriptional activation on the exon1b (38). In addition, binding of Sp1 to GC rich regions located upstream of an activator protein-1 (AP1) site in the BRCA1 promoter, contributes to estrogendependent transcriptional activation (39). Phosphorylation and activation of the ER and Sp1 are also induced through nongenomic pathways involving a mitogen activated protein kinase (MAPK)-cascade, further regulating the cross-talk between ER and BRCA1. The aromatic hydrocarbon receptor (AhR) also binds directly to the BRCA1 promoter and activates its expression in the presence of estrogen (40). However, in the presence of ligands, the bound AhR is recruited to xenobiotic responsive elements (XRE = 5'-GCGTG-3') harbored in the BRCA1 promoter and inhibits its estrogen-mediated transactivation, by blocking the recruitment of transcription factors and cofactors (ER, p300, SRC1) and histone modifications (i.e., AcH4, AcH3K9) that enhance transcription. Conversely, the bound AhR promotes the association of factors (DNMTs, and



methyl binding domain proteins such as MBD2) and histone modifications (H3K9me3, H3K27me3) that repress transcription (**Figure 2**). The AhR and ER pathways also interact at multiple levels in a cell and tissue specific manner. For example, binding of the xenobiotic 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD), activates AhR-mediated activity of P450 enzymes, including CYP1B1, which is involved in estrogen metabolism (41). Also, numerous exogenous and endogenous AhR ligands have been described, with different binding affinity for the AhR (42) and specificity for the two forms of the ER ( $\alpha$ and  $\beta$ ).

TNBC accumulate high levels of reactive oxygen species (ROS) because of their genetic (e.g., *BRCA1* silencing and *TP53* mutations) and metabolic alterations. Our group and others (43, 44) documented overexpression of AhR in TNBC, compared to other subtypes of BC. Specifically, Kubli et al. (44) reported that in normal and malignant mammary cells, AhR directly promoted the expression of amphiregulin (AREG), a ligand of the epidermal growth factor receptor (EGFR). The authors demonstrated that AhR-AREG signaling pathway induced tumorigenesis by controlling ROS and promoting the tumorigenic functions of the tumor microenvironment. They further showed that AhR loss of function sensitized tumor cells to Erlotinib, an EGFR inhibitor, suggesting a promising combinatorial antitumor strategy for the treatment of TNBC. Importantly, these data

implicate a causative role of the AhR in the development of TNBC.

Recently, a novel non-genomic role for BRCA1 has been described in which the BRCA1 protein in the cytoplasm interacts with acetyl CoA carboxylase (ACCA) in a phospho-dependent manner, and this interaction modulates lipogenesis (**Figure 3**). In this model, IGF-1 induces phosphorylation of ACCA, and inhibits its association with BRCA1 (45, 46). Cytoplasmic BRCA1 is rare in less aggressive ER-positive BC (47) and has been linked to metastasis in older patients (>40 years of age) (48). In ER positive cells, Koobotse et al. (46) concluded that inhibiting the association between cytoplasmic BRCA1 and ACCA, e.g., by increasing IGF-1 activity or reducing BRCA1 levels, would induce fatty acid synthesis and promote cell growth. Further studies are necessary to elucidate how the BRCA1/ACCA/IGF-1 axis contribute to tumorigenesis.

Because loss of BRCA1 function is commonly observed in sporadic BC and in subgroups of TNBC, there is great interest in food components that may increase the expression and function of the BRCA1 protein, for developing therapeutic strategies aimed at preventing or stopping BC development. Hypermethylation of the BRCA1 promoter gene has been described in 30–65% of TNBC, and some studies have suggested that *BRCA1* hypermethylation is a hallmark of this BC subtype (49, 50). Hypermethylation of *BRCA1* and *ESR1* (ER $\alpha$ ) genes are usually concurrent (51).



mediates estrogen metabolism.

# **BIOACTIVE FOOD COMPONENTS**

#### Resveratrol

Resveratrol (3,4',5-trihydroxystilbene) is a polyphenolic compound naturally occurring in the skin of dark colored fruits as grapes and berries, in peanuts and strongly pigmented vegetables. Certain plants produce resveratrol and other stilbenoids in response to stress, injury, fungal infection, or ultraviolet (UV) radiation. Resveratrol has been extensively studied in vitro and in animal models for its properties against cardiovascular and cognitive diseases, metabolic diseases as diabetes type 2, and cancer (52). Several studies have investigated the mechanisms of action of resveratrol in BC cells. Our group (53) reported that resveratrol effectively reversed several epigenetic changes associated with activation of the AhR and its binding to the BRCA1 promoter, in ER/PR positive breast cancer (MCF-7) cells. In particular, resveratrol (10 and 20 µM) antagonized the association of H3K9me3, DNMT1, and MBD2 on the BRCA1 promoter; increased transcription of BRCA1; and reduced accumulation of DNA strand breaks. Recently, Chatterjee et al. (54) reported that resveratrol increased the expression of BRCA1, p53, and p21, while decreasing protein arginine methyltransferase5 (PRMT5) and enhancer of zeste homolog 2 (EZH2) in both MCF-7 and MDA-MB-231 (TNBC)

cells. In particular, exposure to resveratrol induced loss of catalytic products of PRMT5, H4R3me2S, and H3K27me3. Moreover, resveratrol reduced lysine deacetylase (KDAC) activity and expression of KDAC1-3, whereas the expression of lysine acetyltransferase KAT2A/3B was enhanced. Overall, resveratrol increased H3K9ac and H3K27ac activity, while reducing repressive histone marks (H4R3me2s and H3K27me3) and increasing the level of activating histone marks (H3K9/27ac) in the proximal promoter region of the tumor suppressor genes BRCA1, p53, and p21. The authors observed that the effects of resveratrol were more pronounced in TNBC cells MDA-MB-231, compared to ER positive MCF7 cells. Other studies showed that resveratrol reduced the expression of metastasis associated protein 1 (MTA1), which is a component of the NuRD complex, functioning as a nucleosome remodeling and deacetylating unit. The resveratrol-induced disruption of the complex MTA1-HDAC lead to increased expression and acetylation of p53, BAX, p21, and apoptosis (55). Earlier studies had reported increased expression of PTEN induced by resveratrol in BC cells (56).

Lysine acetylation within the signal transducer and activator of transcription 3 (STAT3) can interfere with the interaction between DNMT1 and STAT3 causing demethylation and reexpression of tumor suppressor genes. In the TNBC cell line



FIGURE 3 | In this model, IGF-1 induces phosphorylation of ACCA, and inhibits its association with BRCA1, which remains inactive in the cytoplasm. In addition, phosphorylated ACC would promote fatty acid synthesis inducing conversion of acetyl coA into malonyl coA. The conversion of malonyl coA into palmitate is mediated by the fatty acid synthase (FASN), which is inhibited by epigallocatechin-3-gallate (EGCG).

MDA-MB-468, treatment with resveratrol significantly reduced STAT3 acetylation as well as *ESR1* (ER $\alpha$ ) gene promoter DNA methylation. This resulted in the increased expression of ER $\alpha$  and the sensitization to ER-targeted therapy with tamoxifen. Further, growth of *in vivo* tumors in mice was significantly reduced by resveratrol combined with tamoxifen, but not by tamoxifen alone (57).

We reported that the in utero exposure to TCDD increased the number of terminal end buds (TEB) (undifferentiated, proliferative structures) and reduced BRCA-1 expression in mammary tissue of rat offspring. The treatment with TCDD induced occupancy of the BRCA-1 promoter by DNMT-1, CpG methylation of the BRCA-1 promoter, and expression of cyclin D1 and cyclin-dependent kinase-4 (CDK4). These changes were partially overridden by pre-exposure to resveratrol, which stimulated the expression of the AhR repressor AhRR, and its recruitment to the BRCA-1 gene (58). In this study, the concentration of resveratrol in the prenatal diet (7 ppm) approximated the level (10 ppm) used in previous studies to prevent mammary carcinogenesis induced by agonists of the AhR (59). Taken together, these results suggest that the anti-DNMT properties of resveratrol may be a useful tool for prevention of BRCA-1-related tumors.

In a pilot study enrolling 39 women with increased BC risk, the subjects were divided into three groups, receiving 0, 5, and 50 mg resveratrol, twice daily for 3 months. An

inverse relationship between serum resveratrol levels and RASSF-1 $\alpha$  methylation was observed, leading to higher levels of expression of this tumor suppressor gene (60). One of the practical problems presented by resveratrol is how to reach effective *in vivo* concentrations, due to its poor water solubility. To overcome the low bioavailability of resveratrol, oxidized mesoporous carbon nanoparticles (OMCNs) with size below 200 nm and high solubility in water were used to encapsulate resveratrol for delivery in TNBC cells. The results showed good biocompatibility, and cellular uptake efficiency. *In vitro* toxicity and apoptosis analyses indicated that the effects of resveratrol were mediated by poly(ADP-ribose) polymerase (PARP) and caspase-3 cleavage (61).

In a recent article, Lucas et al. (62) suggested that resveratrol and its derivative piceatannol, co-administered with anti-PDL1 (Programmed death-ligand 1) immunotherapy may result in positive response and improved clinical outcome in cancer patients expressing low levels of PDL1. The authors reported that in a panel of BC cell lines, including the TNBC cell line Cal51, the expression of PD-L1 was up-regulated through HDAC3/p300mediated NF- $\kappa$ B control.

Thus, hypothetically, even though the upregulation of PD-L1 by polyphenols in cancer could promote disease progression, agents capable of upregulating PD-L1 expression in tumor cells could sensitize cancer cells for an improved clinical response to PD-L1 immune checkpoint blockade therapy. The authors cautioned that the upregulation of PD-L1by resveratrol or piceatannol occurred at doses not achievable physiologically.

#### Genistein

Genistein and daidzein are the most abundant isoflavones found in various legumes, grains, and vegetables, with soybeans contributing to the higher amount of isoflavones in the human diet (63). Average dietary isoflavone intakes in Asian countries range from 25 to 50 mg/day (64), in contrast, the levels are on average 10-fold lower in Western countries (65).

Consumption of high levels of soybeans associated with early and lifelong exposure to isoflavones have been suggested as possible reasons for the lower rate of BC in Asian populations. Epidemiological studies have shown that higher levels of dietary soy isoflavones ( $\geq 20$  vs.  $\leq 5$  mg/day) were linked to a 29% reduced risk of BC in Asian and American Asian women (66), and early soy exposure may lower risk of BC later in life (67). Also, a meta-analysis of four prospective cohort studies indicated that high vs. low isoflavone intakes (>20 vs. <5 mg/day) might reduce risk of recurrence (RR = 0.84, 95% CI: 0.71–0.99) in BC survivors (68). An inverse association between soy isoflavone intake and recurrence was shown only among women undertaking tamoxifen treatment (69). Even if there are not sufficient data for discouraging moderate consumption of dietary isoflavones, it was recommended that women with a history of ER positive BC, should not increase their consumption of phytoestrogens, including soy isoflavones (70). Guo et al. (71) studied the effects of long-term pre-diagnosis soy food intake on the expression of 800 miRNAs and 302 pre-selected genes in tumor tissues from 272 TNBC cases from the Shanghai Breast Cancer Survival Study. Their findings indicated that soy food consumption (on an average of 10.8 grams/day) for a year before diagnosis may lead to increased expression of tumor suppressor genes and miRNAs, and decreased expression of oncogenes. In particular, higher soy food consumption was associated with higher levels of several miRNA involved in regulation of TP53 cancer-related network, and lower expression of oncogenes such as KRAS and FGFR4 (71).

Soy isoflavones have weak estrogenic activity due to their structural similarity with 17- $\beta$ -estradiol, and they can act as estrogen agonists or antagonists based on their binding to either the ER $\alpha$  or ER $\beta$ , respectively. It has been shown *in vitro* that genistein inhibits protein tyrosine kinases (e.g., EGFR, PDGFR), NF-kB (72), and DNMT (73–76). In ER $\alpha$  positive BC cells, we reported (77) that genistein reversed hypermethylation of the *BRCA1* promoter in part through antagonism of constitutively active AhR. We recently extended our studies in TNBC cell lines showing that genistein upregulated *BRCA-1* expression leading to expression of ER $\alpha$ , and these effects were linked to acquired sensitivity of TNBC cells to the growth inhibitory effects of tamoxifen (78).

Other groups reported that the treatment of MDA-MB-231 (TNBC) cells with 0, 5, 10, or 20  $\mu$ M genistein induced apoptosis and cell cycle arrest in G2/M in a dose-dependent manner. The authors suggested that these effects were mediated by inhibition of NF- $\kappa$ B activity via the Nocth-1 signaling pathway (79). Recently, Paul et al. (80) investigated the combinatorial effects of

low doses of genistein and sulforaphane (SFN) on cell viability. SFN, which is enriched in cruciferous vegetables such as broccoli sprouts and kale, has been shown to possess HDAC inhibiting properties. The doses of  $5 \,\mu\text{M}$  SFN +  $10 \,\mu\text{M}$  genistein and  $5 \,\mu\text{M}$  $SFN + 15 \mu M$  genistein acted synergistically decreasing cellular viability in both ER positive and TNBC cell lines, and the activity of HDACs and HMTs. However, the combination of genistein and SFN was not effective in synergistically downregulating DNMT activity. Previous studies have shown that genistein increased acetylated histones 3, 4, and H3/K4 at the p21 and p16 transcription start sites, and also increased the expression of HAT enzymes that function in transcriptional activation (81). Thus, the combination of genistein and SFN acting synergistically may be due to the HAT promoting activity of genistein that activates tumor suppressor genes such as p21 and p16, whereas SFN may be acting as an HDACi and suppressor of the activity of oncogenes.

Studies in rat and mouse models have produced inconsistent results regarding a protective effect of genistein on mammary tumorigenesis. In rats, mammary gland tumorigenesis was induced by exposure to 2,4-Dimethoxybenzaldehyde (DMBA), an AhR ligand, in animals receiving a wide range of genistein (from 10 to 300 ppm) at different stages of life. Data consistently showed that prepubertal exposure to genistein produced morphological changes in the mammary gland from a proliferative to a differentiated (reduced TEB) phenotype (82). Recently, Zhang et al. (83) reported that lifetime intake of genistein that mimicked Asian dietary patterns, reduced the de novo resistance to tamoxifen in rats, compared to a control group of animals that had received post-diagnosis genistein supplementation. The control group mimicked a dietary model in which genistein supplementation started in adulthood (Caucasian model).

In mice, most studies have been carried out in animals where mammary tumorigenesis was induced by an oncovirus, and genistein exposure was started in early life, pre, or post-puberty. Results of these studies were mixed and did not provide a clear answer whether genistein is protective against tumorigenesis. Possible reasons for this discrepancy include differences between species sensitivity to mammary tumors, tumor inducing agent (oncovirus in mice vs. tumor agent in rats), route, timing and dose of exposure to genistein [for a complete review Warri et al. (82)].

#### Curcumin

Curcumin, extracted from the rhizome of turmeric and widely used in Indian medicine, shares with resveratrol biosynthesis pathways, short half-life, low retention, rapid elimination, and low availability of parent molecules. Piperine and  $\beta$ -glucan have been used as adjuvants for curcumin and resveratrol, respectively, and resulted in a substantial increase (2,000-fold for piperine) in their availability (84). In a recent study, Pandolfi et al. (85) reported on the development of a biomimetic nanodrug consisting of a self-assembling variant (HFn) of human apoferritin loaded with curcumin (CFn). The HFn construct improved the solubility, chemical stability, and bioavailability of curcumin, when tested in two cell lines (MDA-MB-468

and MDA-MB-231) representative of two TNBC subtypes. CFn enhanced the cytotoxic effect of doxorubicin, possibly by interfering with the activity of multidrug resistance transporters. In addition, CFn halted cell cycle in both cell lines, and inhibited Akt phosphorylation, suggesting that the effect on the proliferation and cell cycle were mediated by alteration of the PI3K/Akt pathway. In a study by Lv et al. (86), curcumin induced apoptosis in human BC cell line MDA-MB-231 (TNBC, basal-like). In a similar study, it was observed that curcumin inhibited the proliferation of MDA-MB-231 cells via the EGFR pathway (87). Fatty acid-binding protein 5 (FABP5), which has been indicated as a possible marker for interference with retinoic acid (RA) via the FABP5/PPAR  $\beta/\delta$  pathway, was inhibited by curcumin and led to sensitization of the RA-resistant TNBC cells to RA-mediated growth suppression (88). Kundur et al. (89) reported that curcumin and quercetin (a flavonoid) exhibited synergetic anticancer effects in TNBC cells, possibly by promoting acetylation of the BRCA1 promoter. Similarly to resveratrol, curcumin was found to stimulate β-oxidation of fatty acids, inhibiting adipogenesis, and inflammation (90). Al-Yousef et al. (91) recently reported that curcumin treatment restored BRCA1 expression through reduction of its promoter methylation level in TNBC cell lines HCC-38 and UACC-3199. Lower levels of BRCA1 promoter methylation were attributed to upregulation of the ten-eleven translocation 1 (TET1) gene, which mediates DNA demethylation via hydroxylation of 5methylcytosine to 5-hydroxymethylcytosine. In addition, the authors suggested that TET1 may act as a target of miR-29b, an epi-miRNA which has been shown to inhibit both DNMTs and TETs (92). The miR-29b may balance DNA methylation levels, affecting the function of both methylation and demethylation enzymes (93).

## EGCG

(-)Epigallocatechin 3-gallate (EGCG), is the most abundant catechin found in green tea and displays strong antioxidant activity. Bao et al. (94) reported that exercise and consumption of green tea reduced recurrence and improved survival in TNBC patients. The antiproliferative effects of EGCG have been attributed to inhibition of fatty acid synthase (FASN), which is responsible for the *de novo* synthesis of palmitate, the most abundant fatty acid (**Figure 3**). Notably, FASN inhibition has negligible effect on non-malignant cells, which express low levels of FASN.

*In vivo*, EGCG displays low potency, poor bioavailability, and limited stability. Crous-Maso et al. (95) reported on the design and synthesis of a novel collection of polyphenolic compounds, containing two galloyl moieties (3,4,5-trihydroxybenzoyl group) linked by a variable cyclic subunit. These molecules were tested alongside EGCG for their anticancer properties, in particular in TNBC cell models.

It was reported that 92 % of tumor samples derived from 100 TNBC patients expressed FASN (96) and that doxorubicinresistant cell lines were sensitive to chemotherapeutic drugs through inhibition of FASN (97). The same authors studied the effects of EGCG and its diester derivatives on a TNBC cell line, MDA-MB-231, and two models of the same cell line resistant to chemotherapy agents (98). Their results suggested that the highly proliferative phenotype of chemo-resistant TNBC cells could be treated with FASN inhibitors such EGCG or its more stable diesters.

Braicu et al. (99) evaluated the impact of *p53* silencing and EGCG treatment on genes involved in apoptosis in the Hs578T cell culture model of TNBC. The combined therapy led to the activation of pro-apoptotic genes (i.e., *Bcl-2*) and the inhibition of pro-survival genes (such as BAG cochaperone 3, X-linked inhibitor of apoptosis, and receptor interacting serine/threonine kinase 2), while reducing cell pathways leading to autophagy, thus confirming possible benefits of EGCG regimens for the prevention of TNBC.

In a recent study, Steed et al. (100) used suberoylanilide hydroxamic acid (SAHA), a HDAC inhibitor, alone or in combination with EGCG in the TNBC cell line MDA-MB-231. The two compounds (SAHA and EGCG) decreased the expression of cellular inhibitor of apoptosis 2 (*cIAP2*) while increasing the expression of pro-apoptotic caspase 7. The authors observed also changes in histone modifications, which may mediate the reduction in expression of *cIAP2*. Overall, these changes induced apoptosis. SAHA and EGCG further inhibited TNBC cell migration through fibronectin.

#### Folate

Folates are water-soluble molecules functioning as methyl donors in one-carbon metabolism cycle, which requires vitamin B6, B12, and riboflavin. Fruits and dark leafy green vegetables are rich sources of folates (101). Folates are essential for the synthesis of amino acids and regulate the methylation of DNA and chromosomal stability by controlling the level of the methyl donor, S-adenosyl-methionine (SAM). Consequently, aberrant changes in folate metabolism may contribute to the development of cancer. As an example, RAR B, BRCA1, and Ras association domain family member 1 are frequently methylated in BC. Dietary intake of folate and cobalamin were found to be inversely associated with the methylation status of RAR B and BRCA1 in a study by Pirouzpanah et al. (102). The same authors suggested that a low intake of folate and cobalamin correlated with the age-dependent tendency of promoter regions of these genes to be hypermethylated in tumors. On the other hand, increasing concentrations of folic acid were reported to cause a dosedependent down-regulation of PTEN, APC, and RAR B2 tumor suppressor genes in both ER positive MCF-7 and triple negative MDA-MB-231 BC cell lines (103), suggesting caution with folic acid supplementation.

Folate receptor type alpha (FRA) is over-expressed by a majority of cancers including breast. Recently, two studies reported a strong association of FRA expression with ER/PR-negative and TNBC (>80%) status, poor prognosis, metastatic BC and worse overall/ disease-free survival (104, 105). Therefore, FRA represents a promising target against TNBC. In fact, some folate conjugates, such as folatefluorescein or folate-IgG showed promising anti-tumor activity in mice (106). Anti-FRA IgG antibodies such as MORAB-003 (Farletuzumab) have also been used to target ovarian cancer in patients, although with limited success (107).

Dietary compound	Targets	Epigenetic mechanism
Resveratrol	BRCA1, p53, p21, BAX, PTEN, MTA1, ERa, RASSF-1a, PDL1	Promotes deacetylation/ demethylation through inhibition of H3K9me, DNMT1, MBD2, KDAC, and induction of H3K27ac and H3K29ac
Genistein	P53, KRAS-FGFR4, Tyrosine kinases, NF-kB, BRCA1, ERa, p21, p16	Acts as estrogen agonist or antagonist through binding to ER alpha or beta, respectively. Inhibits protein tyrosine kinases, DNMT; induces HAT activity
Curcumin	FAB P5/PPAR, BRCA1, FASN, miR-29b	Induces DNA acetylation, Beta-oxidation of FASN, inhibits promoter methylation
EGCG	FASN, Bcl2, cIAP2, Caspase7	Inhibits FA synthesis, HDAC
Folate	BRCA1, RARBeta, PTEN,	Modulates methylation through FRA, inhibits Src and ERK

In a recent study, Frontera et al. (108) demonstrated that an IgA Fc-folate conjugate can bind strongly to FRA receptors on TNBC stimulating neutrophils (PMN)-mediated cell killing. PMNs are heavily present in breast tumors and exert cytotoxic action against tumor cells, representing a possible tool against TNBC (109).

Cheung et al. (110) reported that TNBCs show dysregulated expression of thymidylate synthase, folate hydrolase 1, and methylenetetrahydrofolate reductase, involved in folate metabolism. This group used RNA interference to deplete FR $\alpha$  and showed a decrease in Src and ERK signaling which lead to reduced cell growth. An anti-FR $\alpha$  antibody (MOv18-IgG1) conjugated with a Src inhibitor was able to inhibit TNBC xenograft growth. Moreover, MOv18-IgG1 triggered immune-dependent cancer cell death *in vitro* by human volunteer and BC patient immune cells, and significantly restricted orthotopic and patient-derived xenograft growth.

Target genes and mechanisms of action of the main dietary compounds discussed in the manuscript are summarized in **Table 2**.

#### **Other Phytochemicals**

Chromatin modifier therapy is currently used as an adjuvant to sensitize TNBC cells with mixed results. HDAC inhibitors promote the re-expression of tumor suppressor silenced genes, and at the same time reduce the expression of prosurvival genes in favor of pro-apoptotic genes (17). Using phytochemicals to modify the methylation patterns of cancer cells and sensitize them to conventional treatments, may lead to a better alternative to general chemotherapy in particular for TNBC patients. To this end, Szarc Vel Szic et al. (111) showed that withaferin A (WA), a steroidal lactone, commonly known as Ashwagandha, Indian ginseng or Indian winter cherry, downregulated HER2/PR/ESR-dependent gene expression interactions and repressed aggressive triple-negative MDA-MB-231 BC cells with a specific DNA hypermethylation profile of tumor oncogenes. These included a urokinase-type plasminogen activator, ADAM metallopeptidase domain 8, tumor necrosis factor (ligand) superfamily member 12, and genes related to mitochondrial metabolism (malic enzyme 3, ME3) and enzymes of cell detoxification, such as glutathione S-transferase mu 1. In another study, physiologic concentrations of dietary phytochemicals, such as curcumin, DIM, EGCG, or indole-3-carbinol (I3C), altered DNA methylation and expression of genes involved in EMT (cadherin-11), p21Cip1, invasion (urokinase-type plasminogen activator), and interleukin-6. The authors concluded that, even though different targets, all these phytochemicals induced apoptosis of MDA-MB-231 cancer cells (112).

Li et al. (113) reported that fucoidan (a complex sulfated polysaccharide extracted from brown seaweed) inhibited migration of TNBC cells through reduced expression of markers of EMT (N-cadherin and vimentin). Also, the expression of *HIF1a*, which is elevated in metastatic cancer characterized by drug-resistance and high mortality rate, was reduced by fucoidan. However, HIF1a expression does not appear to be different in TNBC compared to other BC subtypes (114).

Inactivation of TP53 is a requirement for tumor progression in BRCA1 deficient BC, and current therapies include treatment with HDAC inhibitors. Zinc metallochaperones are being developed as anticancer drugs that target a class of zinc-binding p53 mutations by restoring wildtype p53 structure and function (115). Consumption of pomegranate has been observed to reduce beta-catenin, EMT and overall metastasis in TNBC (116). In a recent study, Rzepecka-Stojko et al. (117) evaluated the in vitro cytotoxic activity of ethanol extract of propolis (EEP) and its derivative caffeic acid phenethyl ester (CAPE) toward the TNBC cell lines, MDA-MB-231 and Hs578T. The authors reported morphological changes of these cells were observed following exposure to EEP and CAPE. In addition, propolis and CAPE inhibited the growth of both cell lines in a dosedependent and exposure time-dependent manner, with CAPE showing more cytotoxic activity than EEP. Recently, the antineoplastic effects of the electrophilic fatty acid nitroalkene derivative, 10-nitro-octadec-9-enoic acid (nitro-oleic acid, NO2-OA), were investigated in multiple preclinical models of TNBC (118). Electrophilic fatty acid nitroalkene derivatives (NO2-FA) are formed by the acidic conditions of digestion and the redox environment that is up-regulated during inflammation. Other electrophilic species present in vegetables such as broccoli, namely the isothiocyanate derivative sulforaphane, have been shown to mediate therapeutic actions in preclinical models of BC (119). The authors reported that NO2-OA reduced TNBC cell growth and viability *in vitro*, attenuated TNF $\alpha$ -induced TNBC cell migration and invasion, and inhibited the tumor growth of MDA-MB-231 TNBC cell xenografts in the mammary fat pads of female nude mice. These effects were mediated in part by inhibition of TNF $\alpha$ -induced NF-kB transcriptional activity and suppression of downstream NF-kB target gene expression, including the metastasis-related proteins intercellular adhesion molecule-1 and urokinase-type plasminogen activator.

#### CONCLUSIONS

TNBC are heterogeneous in nature, highly metastatic, and their lack of steroid hormone receptors makes ineffective strategies targeting the ER (e.g., tamoxifen). The processes responsible for their propensity to metastasize primarily in lungs and brain are largely unknown, and contribute to the poor prognosis and high mortality rate in TNBC patients (120).

In this review, we underlined how transcription factors can alter the function of tumor suppressors involved in DNA damage, cell proliferation, and differentiation. Promoter methylation and histone deacetylation emerge as central epigenetic mechanisms silencing the expression of tumor suppressors, although the importance of other mechanisms (e.g., siRNA) has been amply described, in particular regarding the mode of action of dietary anticancer molecules (121). We highlighted the complexity of the systems that modulate the expression of BRCA1, whose absence of function is a characteristic of TNBC. Interestingly, the sequestration of BRCA1 in the cytoplasm through its interaction with the ACC enzyme provides a link between dysregulation of lipid metabolism and BRCA1 function in the development of TNBC. Investigations on fatty acid synthesis and their regulation are essential for understanding TNBC development and for identification of targets of treatment connected with inflammation and metabolic pathways, including arachidonic acid and prostaglandins (122). Of particular interest is the central role of the AhR, both as a modulator of BRCA1 function and

#### REFERENCES

- Ferlay J, Colombet M, Soerjomataram I, Mathers C, Parkin DM, Pineros M, et al. Estimating the global cancer incidence and mortality in 2018: GLOBOCAN sources and methods. *Int J Cancer.* (2019) 144:1941– 53. doi: 10.1002/ijc.31937
- Torre LA, Islami F, Siegel RL, Ward EM, Jemal A. Global cancer in women: burden and trends. *Cancer Epidemiol Biomarkers Prev.* (2017) 26:444– 57. doi: 10.1158/1055-9965.EPI-16-0858
- Fragomeni SM, Sciallis A, Jeruss JS. Molecular subtypes and localregional control of breast cancer. Surg Oncol Clin N Am. (2018) 27:95– 120. doi: 10.1016/j.soc.2017.08.005
- Brown-Glaberman U, Dayao Z, Royce M. HER2-targeted therapy for earlystage breast cancer: a comprehensive review. Oncology. (2014) 28:281–9.
- Morris GJ, Naidu S, Topham AK, Guiles F, Xu Y, McCue P, et al. Differences in breast carcinoma characteristics in newly diagnosed African-American and Caucasian patients: a single-institution compilation compared with the National Cancer Institute's Surveillance, Epidemiology, and End Results database. *Cancer*. (2007) 110:876–84. doi: 10.1002/cncr.22836

as a possible target for development of therapeutic strategies against TNBC.

Numerous phytochemicals are being considered as promising allies in preventing or reversing different phases of TNBC progression *in vitro* and *in vivo*, and we described some of the more recent findings. One of the major challenges emerged from these studies is in general the poor bioavailability of these molecules, specifically their high degradability, low solubility, and high degree of metabolic transformation. Another critical issue is the lack of knowledge of how different phytochemicals interact with each other, and how their properties may be used for enhanced efficacy or to avoid negative interference.

Further studies are warranted to unravel the most promising strategies for the treatment of TNBC, through development of both novel drugs and use of available biomolecules. Implementation of dietary regimens rich in phytochemicals with anticancer properties and low in proinflammatory molecules also may prove to be a powerful tool for prevention and treatment of TNBC.

#### **AUTHOR CONTRIBUTIONS**

OS, MD, BS, LN, and DR contributed to the conception and development of the manuscript. OS and DR have primary responsibility for the writing of the manuscript. MD contributed to the writing of data related to BRCA1, AhR and genistein, and review and editing of the manuscript. BS and LN were responsible for the clinical content of the work and contributed to the writing and review of the manuscript. All authors contributed to the article and approved the submitted version.

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- Futreal PA, Liu Q, Shattuck-Eidens D, Cochran C, Harshman K, Tavtigian S, et al. BRCA1 mutations in primary breast and ovarian carcinomas. *Science*. (1994) 266:120–2. doi: 10.1126/science.7939630
- Nattinger AB, Mitchell JL. Breast cancer screening and prevention. *Ann Intern Med.* (2016) 164:81–ITC96. doi: 10.7326/AITC2016 06070
- Brianese RC, Nakamura KDM, Almeida F, Ramalho RF, Barros BDF, Ferreira ENE, et al. BRCA1 deficiency is a recurrent event in early-onset triplenegative breast cancer: a comprehensive analysis of germline mutations and somatic promoter methylation. *Breast Cancer Res Treat.* (2018) 167:803– 14. doi: 10.1007/s10549-017-4552-6
- Tian T, Shan L, Yang W, Zhou X, Shui R. Evaluation of the BRCAness phenotype and its correlations with clinicopathological features in triple-negative breast cancers. *Hum Pathol.* (2019) 84:231–8. doi: 10.1016/j.humpath.2018.10.004
- Zhang J, Fan M, Xie J, Wang Z, Wang B, Zhang S, et al. Chemotherapy of metastatic triple negative breast cancer: experience of using platinum-based chemotherapy. *Oncotarget.* (2015) 6:43135–43. doi: 10.18632/oncotarget.5654

- Yao H, He G, Yan S, Chen C, Song L, Rosol TJ, et al. Triple-negative breast cancer: is there a treatment on the horizon? *Oncotarget*. (2017) 8:1913–24. doi: 10.18632/oncotarget.12284
- Sihto H, Lundin J, Lundin M, Lehtimaki T, Ristimaki A, Holli K, et al. Breast cancer biological subtypes and protein expression predict for the preferential distant metastasis sites: a nationwide cohort study. *Breast Cancer Res.* (2011) 13:R87. doi: 10.1186/bcr2944
- Lehmann BD, Bauer JA, Chen X, Sanders ME, Chakravarthy AB, Shyr Y, et al. Identification of human triple-negative breast cancer subtypes and preclinical models for selection of targeted therapies. *J Clin Invest.* (2011) 121:2750–67. doi: 10.1172/JCI45014
- Burstein MD, Tsimelzon A, Poage GM, Covington KR, Contreras A, Fuqua SA, et al. Comprehensive genomic analysis identifies novel subtypes and targets of triple-negative breast cancer. *Clin Cancer Res.* (2015) 21:1688– 98. doi: 10.1158/1078-0432.CCR-14-0432
- Martin M, Rodriguez-Lescure A, Ruiz A, Alba E, Calvo L, Ruiz-Borrego M, et al. Molecular predictors of efficacy of adjuvant weekly paclitaxel in early breast cancer. *Breast Cancer Res Treat.* (2010) 123:149– 57. doi: 10.1007/s10549-009-0663-z
- Sirohi B, Arnedos M, Popat S, Ashley S, Nerurkar A, Walsh G, et al. Platinum-based chemotherapy in triple-negative breast cancer. *Ann Oncol.* (2008) 19:1847–52. doi: 10.1093/annonc/mdn395
- Bolden JE, Peart MJ, Johnstone RW. Anticancer activities of histone deacetylase inhibitors. *Nat Rev Drug Discov.* (2006) 5:769–84. doi: 10.1038/nrd2133
- Cohen A, Neumayer L, Boucher K, Factor R, Shrestha G, Wade M. Windowof-Opportunity Study of Valproic Acid in Breast Cancer Testing a Gene Expression Biomarker. JCO Precision Oncology (2017). Available online at: "po.ascopubs.org." (accessed June 10, 2020).
- Glinsky GV, Berezovska O, Glinskii AB. Microarray analysis identifies a death-from-cancer signature predicting therapy failure in patients with multiple types of cancer. J Clin Invest. (2005) 115:1503–21. doi: 10.1172/JCI23412
- Douglas D, Hsu JH, Hung L, Cooper A, Abdueva D, van Doorninck J, et al. BMI-1 promotes ewing sarcoma tumorigenicity independent of CDKN2A repression. *Cancer Res.* (2008) 68:6507–15. doi: 10.1158/0008-5472.CAN-07-6152
- Fan QW, Cheng C, Knight ZA, Haas-Kogan D, Stokoe D, James CD, et al. EGFR signals to mTOR through PKC and independently of Akt in glioma. *Sci Signal.* (2009) 2:ra4. doi: 10.1126/scisignal.2000014
- Fantin VR, Leder P. Mitochondriotoxic compounds for cancer therapy. Oncogene. (2006) 25:4787–97. doi: 10.1038/sj.onc.1209599
- Chaneton B, Gottlieb E. Rocking cell metabolism: revised functions of the key glycolytic regulator PKM2 in cancer. *Trends Biochem Sci.* (2012) 37:309–16. doi: 10.1016/j.tibs.2012.04.003
- Mazurek S. Pyruvate kinase type M2: a key regulator of the metabolic budget system in tumor cells. *Int J Biochem Cell Biol.* (2011) 43:969– 80. doi: 10.1016/j.biocel.2010.02.005
- Gao X, Wang H, Yang JJ, Liu X, Liu ZR. Pyruvate kinase M2 regulates gene transcription by acting as a protein kinase. *Mol Cell.* (2012) 45:598– 609. doi: 10.1016/j.molcel.2012.01.001
- Luo W, Semenza GL. Pyruvate kinase M2 regulates glucose metabolism by functioning as a coactivator for hypoxia-inducible factor 1 in cancer cells. *Oncotarget*. (2011) 2:551–6. doi: 10.18632/oncotarget.299
- Azoitei N, Becher A, Steinestel K, Rouhi A, Diepold K, Genze F, et al. PKM2 promotes tumor angiogenesis by regulating HIF-1alpha through NF-kappaB activation. *Mol Cancer*. (2016) 15:3. doi: 10.1186/s12943-015-0490-2
- Han D, Wu G, Chang C, Zhu F, Xiao Y, Li Q, et al. Disulfiram inhibits TGF-beta-induced epithelial-mesenchymal transition and stem-like features in breast cancer via ERK/NF-kappaB/Snail pathway. *Oncotarget.* (2015) 6:40907–19. doi: 10.18632/oncotarget.5723
- Ghosh S, Hayden MS. New regulators of NF-kappaB in inflammation. Nat Rev Immunol. (2008) 8:837–48. doi: 10.1038/nri2423
- Barbie TU, Alexe G, Aref AR, Li S, Zhu Z, Zhang X, et al. Targeting an IKBKE cytokine network impairs triple-negative breast cancer growth. *J Clin Invest.* (2014) 124:5411–23. doi: 10.1172/JCI75661

- Ma C, Zu X, Liu K, Bode AM, Dong Z, Liu Z, et al. Knockdown of pyruvate kinase m inhibits cell growth and migration by reducing NFkB activity in triple-negative breast cancer cells. *Mol Cells*. (2019) 42:628– 36. doi: 10.14348/molcells.2019.0038
- Kuo WY, Hwu L, Wu CY, Lee JS, Chang CW, Liu RS. STAT3/NFkappaB-regulated lentiviral TK/GCV suicide gene therapy for cisplatin-resistant triple-negative breast cancer. *Theranostics.* (2017) 7:647–63. doi: 10.7150/thno.16827
- Hu Y, Scully R, Sobhian B, Xie A, Shestakova E, Livingston DM. RAP80-directed tuning of BRCA1 homologous recombination function at ionizing radiation-induced nuclear foci. *Genes Dev.* (2011) 25:685– 700. doi: 10.1101/gad.2011011
- Saha J, Davis AJ. Unsolved mystery: the role of BRCA1 in DNA end-joining. J Radiat Res. (2016) 57(Suppl. 1):18–i24. doi: 10.1093/jrr/rrw032
- King MC, Marks JH, Mandell JB, New York Breast Cancer Study G. Breast and ovarian cancer risks due to inherited mutations in BRCA1 and BRCA2. *Science*. (2003) 302:643–6. doi: 10.1126/science.1088759
- Lecarpentier J, Silvestri V, Kuchenbaecker KB, Barrowdale D, Dennis J, McGuffog L, et al. Prediction of breast and prostate cancer risks in male BRCA1 and BRCA2 mutation carriers using polygenic risk scores. J Clin Oncol. (2017) 35:2240–50. doi: 10.1200/JCO.2016.69.4935
- Romagnolo DF, Zempleni J, Selmin OI. Nuclear receptors and epigenetic regulation: opportunities for nutritional targeting and disease prevention. *Adv Nutr.* (2014) 5:373–85. doi: 10.3945/an.114.005868
- Jeffy BD, Hockings JK, Kemp MQ, Morgan SS, Hager JA, Beliakoff J, et al. An estrogen receptor-alpha/p300 complex activates the BRCA-1 promoter at an AP-1 site that binds Jun/Fos transcription factors: repressive effects of p53 on BRCA-1 transcription. *Neoplasia*. (2005) 7:873–82. doi: 10.1593/neo.05256
- Hockings JK, Degner SC, Morgan SS, Kemp MQ, Romagnolo DF. Involvement of a specificity proteins-binding element in regulation of basal and estrogen-induced transcription activity of the BRCA1 gene. *Breast Cancer Res.* (2008) 10:R29. doi: 10.1186/bcr1987
- Hockings JK, Thorne PA, Kemp MQ, Morgan SS, Selmin O, Romagnolo DF. The ligand status of the aromatic hydrocarbon receptor modulates transcriptional activation of BRCA-1 promoter by estrogen. *Cancer Res.* (2006) 66:2224–32. doi: 10.1158/0008-5472.CAN-05-1619
- Feng S, Cao Z, Wang X. Role of aryl hydrocarbon receptor in cancer. *Biochim Biophys Acta*. (2013) 1836:197–210. doi: 10.1016/j.bbcan.2013.05.001
- Donovan MG, Selmin OI, Romagnolo DF. Aryl hydrocarbon receptor diet and breast cancer risk. Yale J Biol Med. (2018) 91:105–27.
- Romagnolo DF, Papoutsis AJ, Laukaitis C, Selmin OI. Constitutive expression of AhR and BRCA-1 promoter CpG hypermethylation as biomarkers of ERalpha-negative breast tumorigenesis. *BMC Cancer*. (2015) 15:1026. doi: 10.1186/s12885-015-2044-9
- 44. Kubli SP, Bassi C, Roux C, Wakeham A, Gobl C, Zhou W, et al. AhR controls redox homeostasis and shapes the tumor microenvironment in BRCA1-associated breast cancer. *Proc Natl Acad Sci USA*. (2019) 116:3604– 13. doi: 10.1073/pnas.1815126116
- Moreau K, Dizin E, Ray H, Luquain C, Lefai E, Foufelle F, et al. BRCA1 affects lipid synthesis through its interaction with acetyl-CoA carboxylase. *J Biol Chem.* (2006) 281:3172–81. doi: 10.1074/jbc.M504652200
- Koobotse M, Holly J, Perks C. Elucidating the novel BRCA1 function as a non-genomic metabolic restraint in ER-positive breast cancer cell lines. *Oncotarget*. (2018) 9:33562–76. doi: 10.18632/oncotarget.26093
- 47. Kao J, Salari K, Bocanegra M, Choi YL, Girard L, Gandhi J, et al. Molecular profiling of breast cancer cell lines defines relevant tumor models and provides a resource for cancer gene discovery. *PLoS ONE.* (2009) 4:e6146. doi: 10.1371/journal.pone.0006146
- Santivasi WL, Wang H, Wang T, Yang Q, Mo X, Brogi E, et al. Association between cytosolic expression of BRCA1 and metastatic risk in breast cancer. *Br J Cancer*. (2015) 113:453–9. doi: 10.1038/bjc.2015.208
- Yamashita N, Tokunaga E, Kitao H, Hitchins M, Inoue Y, Tanaka K, et al. Epigenetic inactivation of BRCA1 through promoter hypermethylation and its clinical importance in triple-negative breast cancer. *Clin Breast Cancer*. (2015) 15:498–504. doi: 10.1016/j.clbc.2015.06.009
- 50. Lips EH, Mulder L, Oonk A, van der Kolk LE, Hogervorst FB, Imholz AL, et al. Triple-negative breast cancer: BRCAness and concordance of clinical

features with BRCA1-mutation carriers. Br J Cancer. (2013) 108:2172-7. doi: 10.1038/bjc.2013.144

- Wei M, Xu J, Dignam J, Nanda R, Sveen L, Fackenthal J, et al. Estrogen receptor alpha, BRCA1, and FANCF promoter methylation occur in distinct subsets of sporadic breast cancers. *Breast Cancer Res Treat.* (2008) 111:113– 20. doi: 10.1007/s10549-007-9766-6
- Berretta M, Bignucolo A, Di Francia R, Comello F, Facchini G, Ceccarelli M, et al. Resveratrol in cancer patients: from bench to bedside. *Int J Mol Sci.* (2020) 21:2945. doi: 10.3390/ijms21082945
- Papoutsis AJ, Lamore SD, Wondrak GT, Selmin OI, Romagnolo DF. Resveratrol prevents epigenetic silencing of BRCA-1 by the aromatic hydrocarbon receptor in human breast cancer cells. *J Nutr.* (2010) 140:1607– 14. doi: 10.3945/jn.110.123422
- Chatterjee B, Ghosh K, Kanade SR. Resveratrol modulates epigenetic regulators of promoter histone methylation and acetylation that restores BRCA1, p53, p21(CIP1) in human breast cancer cell lines. *Biofactors*. (2019) 45:818–29. doi: 10.1002/biof.1544
- 55. Farhan M, Ullah MF, Faisal M, Farooqi AA, Sabitaliyevich UY, Biersack B, et al. Differential methylation and acetylation as the epigenetic basis of resveratrol's anticancer activity. *Medicines*. (2019) 6:24. doi: 10.3390/medicines6010024
- 56. Stefanska B, Salame P, Bednarek A, Fabianowska-Majewska K. Comparative effects of retinoic acid, vitamin D and resveratrol alone and in combination with adenosine analogues on methylation and expression of phosphatase and tensin homologue tumour suppressor gene in breast cancer cells. *Br J Nutr.* (2012) 107:781–90. doi: 10.1017/S0007114511003631
- Lee H, Zhang P, Herrmann A, Yang C, Xin H, Wang Z, et al. Acetylated STAT3 is crucial for methylation of tumor-suppressor gene promoters and inhibition by resveratrol results in demethylation. *Proc Natl Acad Sci USA*. (2012) 109:7765–9. doi: 10.1073/pnas.1205132109
- Papoutsis AJ, Selmin OI, Borg JL, Romagnolo DF. Gestational exposure to the AhR agonist 2,3,7,8-tetrachlorodibenzo-p-dioxin induces BRCA-1 promoter hypermethylation and reduces BRCA-1 expression in mammary tissue of rat offspring: preventive effects of resveratrol. *Mol Carcinog.* (2015) 54:261–9. doi: 10.1002/mc.22095
- Banerjee S, Bueso-Ramos C, Aggarwal BB. Suppression of 7,12dimethylbenz(a)anthracene-induced mammary carcinogenesis in rats by resveratrol: role of nuclear factor-kappaB, cyclooxygenase 2, and matrix metalloprotease 9. *Cancer Res.* (2002) 62:4945–54.
- Zhu W, Qin W, Zhang K, Rottinghaus GE, Chen YC, Kliethermes B, et al. Trans-resveratrol alters mammary promoter hypermethylation in women at increased risk for breast cancer. *Nutr Cancer.* (2012) 64:393– 400. doi: 10.1080/01635581.2012.654926
- Fan C, Kong F, Shetti D, Zhang B, Yang Y, Wei K. Resveratrol loaded oxidized mesoporous carbon nanoparticles: a promising tool to treat triple negative breast cancer. *Biochem Biophys Res Commun.* (2019) 519:378– 84. doi: 10.1016/j.bbrc.2019.09.016
- Lucas J, Hsieh TC, Halicka HD, Darzynkiewicz Z, Wu JM. Upregulation of PDL1 expression by resveratrol and piceatannol in breast and colorectal cancer cells occurs via HDAC3/p300mediated NFkappaB signaling. *Int J Oncol.* (2018) 53:1469–80. doi: 10.3892/ijo.2018.4512
- Munro IC, Harwood M, Hlywka JJ, Stephen AM, Doull J, Flamm WG, et al. Soy isoflavones: a safety review. *Nutr Rev.* (2003) 61:1– 33. doi: 10.1301/nr.2003.janr.1-33
- Messina M, Nagata C, Wu AH. Estimated Asian adult soy protein and isoflavone intakes. *Nutr Cancer.* (2006) 55:1– 12. doi: 10.1207/s15327914nc5501\_1
- 65. Zamora-Ros R, Knaze V, Lujan-Barroso L, Kuhnle GG, Mulligan AA, Touillaud M, et al. Dietary intakes and food sources of phytoestrogens in the European Prospective Investigation into Cancer and Nutrition (EPIC) 24-hour dietary recall cohort. *Eur J Clin Nutr.* (2012) 66:932– 41. doi: 10.1038/ejcn.2012.36
- Wu AH, Yu MC, Tseng CC, Pike MC. Epidemiology of soy exposures and breast cancer risk. Br J Cancer. (2008) 98:9–14. doi: 10.1038/sj.bjc. 6604145
- Thanos J, Cotterchio M, Boucher BA, Kreiger N, Thompson LU. Adolescent dietary phytoestrogen intake and breast cancer risk (Canada). *Cancer Causes Control.* (2006) 17:1253–61. doi: 10.1007/s10552-006-0062-2

- Dong JY, Qin LQ. Soy isoflavones consumption and risk of breast cancer incidence or recurrence: a meta-analysis of prospective studies. *Breast Cancer Res Treat.* (2011) 125:315–23. doi: 10.1007/s10549-010-1270-8
- 69. Nechuta SJ, Caan BJ, Chen WY, Lu W, Chen Z, Kwan ML, et al. Soy food intake after diagnosis of breast cancer and survival: an in-depth analysis of combined evidence from cohort studies of US and Chinese women. Am J Clin Nutr. (2012) 96:123–32. doi: 10.3945/ajcn.112.035972
- Duffy C, Perez K, Partridge A. Implications of phytoestrogen intake for breast cancer. CA Cancer J Clin. (2007) 57:260–77. doi: 10.3322/CA.57.5.260
- Guo X, Cai Q, Bao P, Wu J, Wen W, Ye F, et al. Long-term soy consumption and tumor tissue MicroRNA and gene expression in triple-negative breast cancer. *Cancer.* (2016) 122:2544–51. doi: 10.1002/cncr.29981
- Russo M, Russo GL, Daglia M, Kasi PD, Ravi S, Nabavi SF, et al. Understanding genistein in cancer: the "good" and the "bad" effects: a review. *Food Chem.* (2016) 196:589–600. doi: 10.1016/j.foodchem.2015.09.085
- Bosviel R, Dumollard E, Dechelotte P, Bignon YJ, Bernard-Gallon D. Can soy phytoestrogens decrease DNA methylation in BRCA1 and BRCA2 oncosuppressor genes in breast cancer? *OMICS*. (2012) 16:235– 44. doi: 10.1089/omi.2011.0105
- 74. Lubecka K, Kaufman-Szymczyk A, Cebula-Obrzut B, Smolewski P, Szemraj J, Fabianowska-Majewska K. Novel clofarabine-based combinations with polyphenols epigenetically reactivate retinoic acid receptor beta, inhibit cell growth, and induce apoptosis of breast cancer cells. *Int J Mol Sci.* (2018) 19:3970. doi: 10.3390/ijms19123970
- King-Batoon A, Leszczynska JM, Klein CB. Modulation of gene methylation by genistein or lycopene in breast cancer cells. *Environ Mol Mutagen*. (2008) 49:36–45. doi: 10.1002/em.20363
- Xie Q, Bai Q, Zou LY, Zhang QY, Zhou Y, Chang H, et al. Genistein inhibits DNA methylation and increases expression of tumor suppressor genes in human breast cancer cells. *Genes Chromosomes Cancer*. (2014) 53:422–31. doi: 10.1002/gcc.22154
- Romagnolo DF, Donovan MG, Papoutsis AJ, Doetschman TC, Selmin OI. Genistein prevents BRCA1 CpG methylation and proliferation in human breast cancer cells with activated aromatic hydrocarbon receptor. *Curr Dev Nutr.* (2017) 1:e000562. doi: 10.3945/cdn.117.000562
- Donovan MG, Selmin OI, Doetschman TC, Romagnolo DF. Epigenetic activation of BRCA1 by genistein *in vivo* and triple negative breast cancer cells linked to antagonism toward Aryl hydrocarbon receptor. *Nutrients*. (2019) 11:2559. doi: 10.3390/nu11112559
- Pan H, Zhou W, He W, Liu X, Ding Q, Ling L, et al. Genistein inhibits MDA-MB-231 triple-negative breast cancer cell growth by inhibiting NFkappaB activity via the Notch-1 pathway. *Int J Mol Med.* (2012) 30:337– 43. doi: 10.3892/ijmm.2012.990
- Paul B, Li Y, Tollefsbol TO. The effects of combinatorial genistein and sulforaphane in breast tumor inhibition: role in epigenetic regulation. *Int J Mol Sci.* (2018) 19:1754. doi: 10.3390/ijms19061754
- Majid S, Kikuno N, Nelles J, Noonan E, Tanaka Y, Kawamoto K, et al. Genistein induces the p21WAF1/CIP1 and p16INK4a tumor suppressor genes in prostate cancer cells by epigenetic mechanisms involving active chromatin modification. *Cancer Res.* (2008) 68:2736– 44. doi: 10.1158/0008-5472.CAN-07-2290
- Warri A, Saarinen NM, Makela S, Hilakivi-Clarke L. The role of early life genistein exposures in modifying breast cancer risk. *Br J Cancer.* (2008) 98:1485–93. doi: 10.1038/sj.bjc.6604321
- Zhang X, Cook KL, Warri A, Cruz IM, Rosim M, Riskin J, et al. Lifetime genistein intake increases the response of mammary tumors to tamoxifen in rats. *Clin Cancer Res.* (2017) 23:814–24. doi: 10.1158/1078-0432.CCR-16-1735
- Bezerra DP, de Castro FO, Alves AP, Pessoa C, de Moraes MO, Silveira ER, et al. *In vitro* and *in vivo* antitumor effect of 5-FU combined with piplartine and piperine. *J Appl Toxicol.* (2008) 28:156–63. doi: 10.1002/jat.1261
- Pandolfi L, Bellini M, Vanna R, Morasso C, Zago A, Carcano S, et al. Hferritin enriches the curcumin uptake and improves the therapeutic efficacy in triple negative breast cancer cells. *Biomacromolecules*. (2017) 18:3318– 30. doi: 10.1021/acs.biomac.7b00974
- Lv ZD, Liu XP, Zhao WJ, Dong Q, Li FN, Wang HB, et al. Curcumin induces apoptosis in breast cancer cells and inhibits tumor growth *in vitro* and *in vivo*. *Int J Clin Exp Pathol*. (2014) 7:2818–24.

- Sun XD, Liu XE, Huang DS. Curcumin induces apoptosis of triple-negative breast cancer cells by inhibition of EGFR expression. *Mol Med Rep.* (2012) 6:1267–70. doi: 10.3892/mmr.2012.1103
- Thulasiraman P, McAndrews DJ, Mohiudddin IQ. Curcumin restores sensitivity to retinoic acid in triple negative breast cancer cells. *BMC Cancer*. (2014) 14:724. doi: 10.1186/1471-2407-14-724
- Kundur S, Prayag A, Selvakumar P, Nguyen H, McKee L, Cruz C, et al. Synergistic anticancer action of quercetin and curcumin against triple-negative breast cancer cell lines. J Cell Physiol. (2019) 234:11103– 18. doi: 10.1002/jcp.27761
- Rupasinghe HP, Sekhon-Loodu S, Mantso T, Panayiotidis MI. Phytochemicals in regulating fatty acid beta-oxidation: potential underlying mechanisms and their involvement in obesity and weight loss. *Pharmacol Ther.* (2016) 165:153–63. doi: 10.1016/j.pharmthera.2016.06.005
- Al-Yousef N, Shinwari Z, Al-Shahrani B, Al-Showimi M, Al-Moghrabi N. Curcumin induces reexpression of BRCA1 and suppression of gamma synuclein by modulating DNA promoter methylation in breast cancer cell lines. *Oncol Rep.* (2020) 43:827–38. doi: 10.3892/or.2020.7473
- Morita S, Horii T, Kimura M, Ochiya T, Tajima S, Hatada I. miR-29 represses the activities of DNA methyltransferases and DNA demethylases. *Int J Mol Sci.* (2013) 14:14647–58. doi: 10.3390/ijms140714647
- Yan B, Guo Q, Fu FJ, Wang Z, Yin Z, Wei YB, et al. The role of miR-29b in cancer: regulation, function, and signaling. *Onco Targets Ther.* (2015) 8:539–48. doi: 10.2147/OTT.S75899
- 94. Bao PP, Zhao GM, Shu XO, Peng P, Cai H, Lu W, et al. Modifiable lifestyle factors and triple-negative breast cancer survival: a population-based prospective study. *Epidemiology*. (2015) 26:909–16. doi: 10.1097/EDE.000000000000373
- Crous-Maso J, Palomeras S, Relat J, Camo C, Martinez-Garza U, Planas M, et al. (-)-Epigallocatechin 3-gallate synthetic analogues inhibit fatty acid synthase and show anticancer activity in triple negative breast cancer. *Molecules*. (2018). 23:1160. doi: 10.3390/molecules23051160
- 96. Giro-Perafita A, Sarrats A, Perez-Bueno F, Oliveras G, Buxo M, Brunet J, et al. Fatty acid synthase expression and its association with clinicohistopathological features in triple-negative breast cancer. *Oncotarget.* (2017) 8:74391–405. doi: 10.18632/oncotarget.20152
- Giro-Perafita A, Palomeras S, Lum DH, Blancafort A, Vinas G, Oliveras G, et al. Preclinical evaluation of fatty acid synthase and egfr inhibition in triple-negative breast cancer. *Clin Cancer Res.* (2016) 22:4687–97. doi: 10.1158/1078-0432.CCR-15-3133
- Giro-Perafita A, Rabionet M, Planas M, Feliu L, Ciurana J, Ruiz-Martinez S, et al. EGCG-derivative G28 shows high efficacy inhibiting the mammosphere-forming capacity of sensitive and resistant TNBC models. *Molecules*. (2019) 24:1027. doi: 10.3390/molecules24061027
- 99. Braicu C, Pileczki V, Pop L, Petric RC, Chira S, Pointiere E, et al. Dual targeted therapy with p53 siRNA and epigallocatechingallate in a triple negative breast cancer cell model. *PLoS ONE.* (2015) 10:e0120936. doi: 10.1371/journal.pone.0120936
- Steed KL, Jordan HR, Tollefsbol TO. SAHA and EGCG promote apoptosis in triple-negative breast cancer cells, possibly through the modulation of cIAP2. *Anticancer Res.* (2020) 40:9–26. doi: 10.21873/anticanres.13922
- 101. Liu JJ, Ward RL. Folate and one-carbon metabolism and its impact on aberrant DNA methylation in cancer. *Adv Genet.* (2010) 71:79– 121. doi: 10.1016/B978-0-12-380864-6.00004-3
- 102. Pirouzpanah S, Taleban FA, Mehdipour P, Atri M. Association of folate and other one-carbon related nutrients with hypermethylation status and expression of RARB, BRCA1, and RASSF1A genes in breast cancer patients. *J Mol Med.* (2015) 93:917–34. doi: 10.1007/s00109-015-1268-0
- 103. Lubecka-Pietruszewska K, Kaufman-Szymczyk A, Stefanska B, Fabianowska-Majewska K. Folic acid enforces DNA methylation-mediated transcriptional silencing of PTEN, APC and RARbeta2 tumour suppressor genes in breast cancer. *Biochem Biophys Res Commun.* (2013) 430:623– 8. doi: 10.1016/j.bbrc.2012.11.103
- 104. O'Shannessy DJ, Somers EB, Maltzman J, Smale R, Fu YS. Folate receptor alpha (FRA) expression in breast cancer: identification of a new molecular subtype and association with triple negative disease. *Springerplus*. (2012) 1:22. doi: 10.1186/2193-1801-1-22

- 105. Zhang Z, Wang J, Tacha DE, Li P, Bremer RE, Chen H, et al. Folate receptor alpha associated with triple-negative breast cancer and poor prognosis. *Arch Pathol Lab Med.* (2014) 138:890–5. doi: 10.5858/arpa.2013-0309-OA
- 106. Li H, Lu Y, Piao L, Wu J, Yang X, Kondadasula SV, et al. Folateimmunoglobulin G as an anticancer therapeutic antibody. *Bioconjug Chem.* (2010) 21:961–8. doi: 10.1021/bc900545h
- 107. Ebel W, Routhier EL, Foley B, Jacob S, McDonough JM, Patel RK, et al. Preclinical evaluation of MORAb-003, a humanized monoclonal antibody antagonizing folate receptor-alpha. *Cancer Immun.* (2007) 7:6.
- 108. Frontera ED, Khansa RM, Schalk DL, Leakan LE, Guerin-Edbauer TJ, Ratnam M, et al. IgA Fc-folate conjugate activates and recruits neutrophils to directly target triple-negative breast cancer cells. *Breast Cancer Res Treat.* (2018) 172:551–60. doi: 10.1007/s10549-018-4941-5
- 109. Gregory AD, Houghton AM. Tumor-associated neutrophils: new targets for cancer therapy. *Cancer Res.* (2011) 71:2411– 6. doi: 10.1158/0008-5472.CAN-10-2583
- 110. Cheung A, Opzoomer J, Ilieva KM, Gazinska P, Hoffmann RM, Mirza H, et al. Anti-folate receptor alpha-directed antibody therapies restrict the growth of triple-negative breast cancer. *Clin Cancer Res.* (2018) 24:5098–111. doi: 10.1158/1078-0432.CCR-18-0652
- 111. Szarc Vel Szic K, Declerck K, Crans RAJ, Diddens J, Scherf DB, Gerhauser C, et al. Epigenetic silencing of triple negative breast cancer hallmarks by Withaferin A. Oncotarget. (2017) 8:40434–53. doi: 10.18632/oncotarget.17107
- 112. Moiseeva EP, Almeida GM, Jones GD, Manson MM. Extended treatment with physiologic concentrations of dietary phytochemicals results in altered gene expression, reduced growth, and apoptosis of cancer cells. *Mol Cancer Ther.* (2007) 6:3071–9. doi: 10.1158/1535-7163.MCT-07-0117
- 113. Li W, Xue D, Xue M, Zhao J, Liang H, Liu Y, et al. Fucoidan inhibits epithelial-to-mesenchymal transition via regulation of the HIF-1alpha pathway in mammary cancer cells under hypoxia. *Oncol Lett.* (2019) 18:330– 8. doi: 10.3892/ol.2019.10283
- 114. Yehia L, Boulos F, Jabbour M, Mahfoud Z, Fakhruddin N, El-Sabban M. Expression of HIF-1alpha and markers of angiogenesis are not significantly different in triple negative breast cancer compared to other breast cancer molecular subtypes: implications for future therapy. *PLoS ONE.* (2015) 10:e0129356. doi: 10.1371/journal.pone.0129356
- 115. Na B, Yu X, Withers T, Gilleran J, Yao M, Foo TK, et al. Therapeutic targeting of BRCA1 and TP53 mutant breast cancer through mutant p53 reactivation. *NPJ Breast Cancer*. (2019) 5:14. doi: 10.1038/s41523-019-0110-1
- 116. Bagheri M, Fazli M, Saeednia S, Kor A, Ahmadiankia N. Pomegranate peel extract inhibits expression of beta-catenin, epithelial mesenchymal transition, and metastasis in triple negative breast cancer cells. *Cell Mol Biol.* (2018) 64:86–91. doi: 10.14715/cmb/2018.64.7.15
- 117. Rzepecka-Stojko A, Stojko J, Kurek-Gorecka A, Gorecki M, Kabala-Dzik A, Kubina R, et al. Polyphenols from Bee Pollen: structure, absorption, metabolism and biological activity. *Molecules*. (2015) 20:21732– 49. doi: 10.3390/molecules201219800
- Woodcock CC, Huang Y, Woodcock SR, Salvatore SR, Singh B, Golin-Bisello F, et al. Nitro-fatty acid inhibition of triplenegative breast cancer cell viability, migration, invasion, and tumor growth. J Biol Chem. (2018) 293:1120–37. doi: 10.1074/jbc.M117. 814368
- 119. So JY, Lin JJ, Wahler J, Liby KT, Sporn MB, Suh N. A synthetic triterpenoid CDDO-Im inhibits tumorsphere formation by regulating stem cell signaling pathways in triple-negative breast cancer. *PLoS ONE.* (2014) 9:e107616. doi: 10.1371/journal.pone. 0107616
- 120. Al-Mahmood S, Sapiezynski J, Garbuzenko OB, Minko T. Metastatic and triple-negative breast cancer: challenges and treatment options. *Drug Deliv Transl Res.* (2018) 8:1483–507. doi: 10.1007/s13346-018-0551-3
- 121. Ratovitski EA. natural Anticancer compounds as epigenetic modulators of gene expression. Curr Genomics. (2017)18:175-205. doi: 10.2174/13892029176661608031 65229

122. Guo M, Sinha S, Wang SM. Coupled genome-wide DNA methylation and transcription analysis identified rich biomarkers and drug targets in triple-negative breast cancer. *Cancers.* (2019) 11:1724. doi: 10.3390/cancers11111724

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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