

## **Original Article**

# **Epigenome-wide Association Study Analysis of Calorie Restriction in Humans, CALERIE™ Trial Analysis**

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Received: June 2, 2022; Editorial Decision Date: August 8, 2022

Decision Editor: Rafael de Cabo, PhD, FGSA

## Abstract

Calorie restriction (CR) increases healthy life span and is accompanied by slowing or reversal of aging-associated DNA methylation (DNAm) changes in animal models. In the Comprehensive Assessment of Long-term Effects of Reducing Intake of Energy (CALERIE<sup>TM</sup>) human trial, we evaluated associations of CR and changes in whole-blood DNAm. CALERIE<sup>TM</sup> randomized 220 healthy, nonobese adults in a 2:1 allocation to 2 years of CR or ad libitum (AL) diet. The average CR in the treatment group through 24 months of follow-up was 12%. Whole blood (baseline, 12, and 24 months) DNAm profiles were measured. Epigenome-wide association study (EWAS) analysis tested CR-induced changes from baseline to 12 and 24 months in the n = 197 participants with available DNAm data. CR treatment was not associated with epigenomewide significant (false discovery rate [FDR] < 0.05) DNAm changes at the individual-CpG-site level. Secondary analysis of sets of CpG sites identified in published EWAS revealed that CR induced DNAm changes opposite to those associated with higher body mass index and cigarette smoking (p < .003 at 12- and 24-month follow-ups). In contrast, CR altered DNAm at chronological-age-associated CpG sites in the direction of older age (p < .003 at 12- and 24-month follow-ups). Although individual CpG site DNAm changes in response to CR were not identified, analyses of sets CpGs identified in prior EWAS revealed CR-induced changes to blood DNAm. Altered CpG sets were enriched for insulin production, glucose tolerance, inflammation, and DNA-binding and DNA-regulation pathways, several of which are known to be modified by CR. DNAm changes may contribute to CR effects on aging.

Keywords: Caloric restriction, Epigenome, Human aging

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The geroscience hypothesis proposes that interventions that slow or reverse biological processes of aging can simultaneously prevent multiple chronic diseases and extend healthy life span (1). Proof of concept for geroscience is emerging from studies with animals, in which interventions that slow or reverse the accumulation of molecular "hallmarks" of aging delay the onset of disease and functional impairment and extend healthy aging (2–4). One of the best-evidenced geroscience intervention in animals is calorie restriction (CR) (5). CR is defined as a reduction in caloric intake from a normal intake ("ad libitum" [AL]) diet while maintaining adequate nutrient intake (6). From worms to mice to monkeys, CR is associated with delayed onset of age-associated diseases, including diabetes, cancer, cardiovascular disease, osteoarthritis, and increased healthy life span (7–9).

The mechanisms by which CR slows aging and extends healthspan in animal models are several and include alterations at physiological, metabolic, and genomic levels (6,10). Studies in animals have identified slowing or reversal of epigenetic changes associated with aging in response to CR, including alterations of whole blood DNA methylation (DNAm) (11,12). However, the effects of CR on whole blood DNAm in nonobese humans are unknown.

The Comprehensive Assessment of Long-term Effects of Reducing Intake of Energy (CALERIE<sup>TM</sup>) study is the first long-term, randomized clinical trial of CR in healthy, nonobese humans (13). The goal of CALERIE<sup>TM</sup> was to identify the effects of 2 years of CR on predictors of longevity, disease risk factors, and quality of life. The intervention yielded substantial and sustained weight loss and signs of improved cardiometabolic health, reduced inflammation, and slowed biological aging, as measured by physiology-based algorithms (14,15). In ancillary studies in subsets of CALERIE<sup>TM</sup> participants, CR induced signs of metabolic slowing and reversal of markers of immune-system aging (16,17).

We conducted a genome-wide analysis of whole-blood DNAm changes over 12 and 24 months in CALERIE™. The primary analysis tested changes in methylation levels at each of 828,613 C-G dinucleotides (CpGs). Secondary analyses tested changes at sets of CpGs identified in published epigenome-wide association studies (EWAS) of the body mass index (BMI), cigarette smoking, and chronological age (18-21). BMI EWAS analysis was of interest because the CALERIE<sup>TM</sup> intervention induced substantial weight loss. Smoking- and chronological-age EWAS analyses were of interest in CALERIE<sup>TM</sup> because these are established risk factors for shortening healthy life span and have associations with DNAm differences at large numbers of CpG sites, which are not currently known to be directly affected by CR. We hypothesized that CR would offer a geroprotective effect which could be measured molecularly via DNAm, especially at regions associated with risk factors for a shorter life span.

#### Method

The CALERIE<sup>TM</sup> trial randomized 220 healthy, nonobese (BMI  $22.0 \le BMI < 28.0 \text{ kg/m}^2$ ), adults aged 21-50 years to either a 25% CR intervention condition or AL control at a 2:1 (CR:AL) ratio across 3 sites (Pennington Biomedical Research Center, Washington University, and Tufts University; Figure 1A, Table 1) (13,22). Participants were excluded from the study if they had significant medical conditions, abnormal laboratory markers, present or potential psychiatric or behavioral problems, regular use of medications (except oral contraceptives), currently smoked, were highly physically active, or were pregnant or breastfeeding. Randomization was stratified by study site, sex, and BMI. The trial duration was 24 months. As measured using



Figure 1. Study design. (A) CALERIE<sup>™</sup> trial design. Two hundred and twenty participants were randomly assigned to either 25% calorie restriction (CR) or ad libitum (AL) at a 2:1 ratio. Of the 220 participants assigned, 218 started and 188 completed the intervention. (B) Blood samples were collected from participants at baseline and 12- and 24-month follow-ups. DNA was isolated and stored. DNA methylation was assayed with Illumina methyl EPIC bead chip arrays. After quality control and normalization, epigenome-wide association study (EWAS) analysis tested CALERIE<sup>™</sup> intervention effects at 12- and 24-month follow-ups at each of 828,613 CpG sites. Finally, we conducted secondary analysis comparing results from CALERIE<sup>™</sup> EWAS with results from published EWAS of BMI, cigarette smoking, and chronological age.

Table 1. Study Participant Characteristics at Pre-treatment Baseline

Intervention	CR (n = 142)	AL $(n = 72)$
Females	97	50
Males	45	22
Hispanic	2	4
Asian	12	3
African American	15	10
White	110	55
Other	3	0
BMI (mean $\pm$ SD)	25.17 (1.8)	25.14 (1.7)
Age (mean $\pm SD$ )	38.19 (7.3)	38.16 (7.1)

Note: AL = ad libitum; BMI = body mass index; CR = calorie restriction.

doubly labeled water, the CR intervention group achieved an average of  $11.7 \pm 0.7\%$  CR ( $19.5 \pm 0.08\%$  in the first 6 months,  $9.1 \pm 0.7\%$  during the subsequent 18 months) (23).

#### **DNA Methylation**

DNAm profiling was conducted in the Kobor Lab from whole-blood DNA stored at -80°C. After quality controls and normalization, DNAm data sets were generated for n = 595 samples from 214 individuals (142 CR, 72 AL; Figure 1B, Table 1). Briefly, 750 ng of DNA was extracted from whole blood and bisulfite was converted using the EZ DNA Methylation kit (Zymo Research, Irvine, CA). Methylation was measured from 160 ng of bisulfite-converted DNA using the Illumina EPIC Beadchip (Illumina, Inc., San Diego, CA). Quality control (QC) and normalization were performed using methylumi (v. 2.32.0) (24) and the Bioconductor (v 2.46.0) (25) packages from the R statistical programming environment (v 3.6.3). Probes with detection p-values > .05 were coded as missing; probes missing in > 5% of samples were removed from the dataset (final probe n = 828,613 CpGs). Normalization to eliminate systematic dye bias in the 2-channel probes was carried out using the methylumi default method. We conducted a principal component analysis of EPIC-array control-probe beta values to compute controls for technical variability across the samples (26).

#### Statistical Analysis

The primary analysis was an epigenome-wide association study (EWAS) of CALERIE<sup>TM</sup> treatment effects in which the treatment

group was the exposure and changes in probe beta value from baseline to 12 months and baseline to 24 months were the outcome variables. Secondary analyses examined sets of CpG sites identified in published EWAS of obesity, cigarette smoking, and chronological age to test if CALERIE<sup>TM</sup> treatment specifically affected DNAm at CpG sites known to be altered by these exposures.

## Epigenome-wide association study of CALERIE<sup>TM</sup> treatment effects

We tested associations of CALERIE<sup>™</sup> intervention with changes in DNAm at each QC'ed CpG site using a mixed model. The model took the form of:

$$\beta_{it} \sim a_i | + |Follow - up Time| + |CR| + |Follow - up Time| \times CR| + |X_{it}| + |e|$$

where " $\beta$ " is the level of methylation for CpG site "i" at time "t"; "a" is the model intercept, including sample-wide and person-specific components, "Follow-up Time" is a pair of indicator variables encoding the 12- and 24-month follow-ups; "CR" is an indicator of treatment group; "Follow-up Time × CR" is a series of interaction terms between follow-up time and treatment group; "X" is a matrix of covariates; and "e" is the error term comprising both sample-wide and person-specific components. The effect of the intervention is tested by the coefficients for the interaction terms, which evaluate the treatment effect at 12 and 24 months as the difference in change from baseline between the treatment (CR) and control AL groups.

Time-invariant covariates were pre-intervention-baseline chronological age, sex, BMI stratum (22–24.9, 25–27.9), study site, and the first 3 principal components estimated from genome-wide SNP data in order to correct for population stratification. Time-varying covariates were proportions of monocytes, neutro-phils, and CD4T, CD8T, Natural Killer, and B-cell lymphocytes estimated from the DNAm data using the Houseman Equation via the Minfi and FlowSorted.Blood.EPIC R packages and the first 7 principal components were estimated from EPIC-array control probes (26–28). Benjamini–Hochberg correction was applied to account for nonindependence of tests. Statistical significance was established at a false discovery rate (FDR) < 0.05. EWAS analysis was conducted using the lmerTest R package (29).

#### Secondary analyses of EWAS summary statistics

We evaluated whether DNAm changes associated with CALERIE<sup>™</sup> intervention reflected changes expected based on published EWAS. We conducted analyses of EWAS results from studies of BMI, cigarette smoking, and chronological age (18–21). Hypothesis testing was performed using a Wilcoxon Rank Sum Test to compare distributions of CALERIE<sup>™</sup> EWAS test statistics for phenotype-associated CpGs to the distribution of CALERIE<sup>™</sup> EWAS test statistics for all other CpGs. Independent tests were performed for CpG sites identified as hypermethylated and hypomethylated in association with the target phenotype. Because all target-phenotype EWAS used an earlier generation of Illumina array technology, we restricted these analyses to the 431,205 EPIC-array CpGs measured in CALERIE<sup>™</sup> that were also included on the Illumina 450k array.

#### Secondary analysis of BMI-associated CpGs

The CALERIE<sup>™</sup> intervention was associated with an average weight loss of 8 kg by 12 months of follow-up (23). We therefore evaluated whether DNAm changes associated with the CALERIE<sup>™</sup> intervention overlapped with DNAm associations with BMI. We examined 129 CpGs identified in a prior EWAS of BMI (18). Specifically, we tested if CpGs hypomethylated in individuals with higher BMI showed signs of increased DNAm in response to the CALERIE<sup>TM</sup> intervention, and if CpGs hypermethylated in individuals with higher BMI showed signs of decreased DNAm in response to CALERIE<sup>TM</sup> intervention, that is, we tested the hypothesis that DNAm changed induced by CALERIE<sup>TM</sup> intervention would be opposite to the pattern of association with higher BMI.

#### Secondary analysis of smoking-associated CpGs

We tested if DNAm changes associated with the CALERIE<sup>™</sup> intervention overlapped with DNAm associations with cigarette smoking, a potent risk factor for aging-related disease and mortality known to have pervasive effects on blood DNAm. We examined 2 622 CpGs identified in a prior EWAS of smoking (21). We tested if CpGs hypomethylated in smokers showed signs of increased DNAm in response to the CALERIE<sup>™</sup> intervention and if CpGs hypermethylated in smokers showed signs of decreased DNAm in response to the CALERIE<sup>™</sup> intervention, that is, we tested the hypothesis that DNAm changes induced by CALERIE<sup>™</sup> intervention would be opposite to the pattern of association with smoking.

#### Secondary analysis of chronological-age-associated CpGs

We tested if DNAm changes associated with the CALERIE<sup>TM</sup> intervention overlapped with DNAm associations with chronological age. We examined 1 000 CpGs identified in a prior EWAS of chronological age (19). We tested if CpGs hypomethylated in chronologically older individuals showed signs of increased DNAm in response to the CALERIE<sup>TM</sup> intervention and if CpGs hypermethylated in chronologically older individuals showed signs of decreased DNAm in response to CALERIE<sup>TM</sup> intervention, that is, we tested the hypothesis that DNAm changes induced by CALERIE<sup>TM</sup> intervention would be opposite to the pattern of association with older chronological age. We repeated the analysis using 875 CpGs identified in a prior EWAS of chronological age (20).

For all secondary analyses, we applied a Bonferoni-corrected threshold of p < .003 to establish statistical significance (16 tests; 0.05/16 = 0.003).

#### Enrichment analyses

To inform the interpretation of secondary analyses, we performed an enrichment analysis of sets of CpGs identified in published EWAS (18-20). We annotated each CpG to the nearest transcription start site to conduct gene enrichment analysis. We used the Reactome Database to identify enriched biological processes, pathways, and functional relationships (30). We used the GM12878 chromatin immunoprecipitation sequencing (ChIP-seq) data from the ENCODE data portal (31) to identify whether certain transcription factor binding sites were enriched amongst phenotype-associated CpGs. Briefly, BEDtools were used to identify the intersection between the Methyl 450 annotation file and the ChIP-seq bed file (32). Enrichment of transcription factors bound within 500 bp of the phenotype-associated CpGs compared to non-phenotype-associated CpGs was tested with permutation analysis. We additionally tested ontological enrichment using the gene ontology enrichment analysis and visualization tool (Gorilla) (33).

#### **Results**

#### EWAS of CALERIE<sup>™</sup>Treatment Effects

We conducted an intent-to-treat analysis of CALERIE<sup>TM</sup> treatment effects at 12- and 24-month follow-ups. Genome-wide comparison of

DNAm between CR and AL at 12 and 24 months did not identify any CpG-site-specific changes that were statistically different from zero at FDR < 0.05 (Figure 2; Supplementary Table 1). The top-ranked CpG site at 12 months was within the first exon of T-Cell Receptor T3 Delta Chain (CD3D) (cg07728874, *p*-value =  $4.05 \times 10^{-6}$ ). At 24 months, the top-ranking CR-associated site was located on chromosome 1 within Long intergenic Non-Protein Coding RNA 1344 (LNC01334) (cg12040931, *p*-value =  $2.5 \times 10^{-6}$ ).

### Secondary Analyses of CpG Sites Identified in Published EWAS of BMI, Cigarette Smoking, and Chronological Age

We conducted secondary analyses of summary statistics from the CALERIE<sup>TM</sup> EWAS using published results from EWAS of BMI, cigarette smoking, and chronological age.

Because CR induced substantial weight loss (23), we first compared CALERIE<sup>TM</sup> EWAS results for DNAm at n = 129 CpG sites identified in a published EWAS of BMI (18) with results for all other CpG sites. For CpG sites identified as hypermethylated in individuals with higher BMI (n = 50), CR tended to reduce DNAm (12 months, p = 2.06E-07; 24 months, p = 3.96E-11). For CpG sites identified as hypomethylated in individuals with higher BMI (n = 79), CR tended to increase DNAm (12 months, p = 1.04E-06; 24 months, p = 7.04E-04). Thus, for both sets of CpGs, CR reversed BMI-associated DNAm.

We next compared CALERIE<sup>TM</sup> EWAS results for DNAm at n = 2 622 CpG sites identified in EWAS of cigarette smoking (34) with results for all other CpG sites. For CpG sites identified as hypermethylated in smokers (n = 1 555), compared with AL, CR tended to reduce DNAm (12 months, p = 1.03E-05; 24 months, p = 2.63E-30). For CpG sites identified as hypomethylated in smokers (n = 1 067), compared with AL, CR tended to increase DNAm, although this finding was statistically different from the null only at 24 months of follow-up (12 months, p = .08; 24 months, p = 4.3E-04). Overall, CR showed signs of reversing smoking-associated DNAm.

Finally, we compared CALERIE<sup>TM</sup> EWAS results for DNAm at 1 000 CpG sites previously associated with chronological age (19) to results for all other CpG sites. For CpG sites identified as hypermethylated in older adults (n = 980), compared with AL, CR tended to increase DNAm (12 months, p = 3.79E-41; 24 months, p = 5.73E-06). For CpG sites identified as hypomethylated in older adults (n = 20), compared with AL, CR was not associated with changes in DNAm (12 months, p = .12; 24 months, p = .29). Results were similar in repeated analyses using results from a second EWAS



**Figure 2.** Quantile-quantile (QQ) plots of *p*-value distributions from epigenome-wide association study (EWAS) analysis of CALERIE<sup>TM</sup> treatment effects at 12- and 24-month follow-ups. The figure shows QQ plots for EWAS of blood DNA methylation changes in response to CR at 12 months (Genomic Inflation–0.97; Panel **A**) and 24 months (Genomic Inflation–0.99; Panel **B**).

of chronological age (20). Thus, for sites hypermethylated in older adults, CR induced DNAm changes consistent with older age. In contrast, CR had no detectable effect on sites hypomethylated in older as compared to younger adults.

Results for analyses of BMI-, cigarette smoking-, and chronological-age-associated CpG sites are reported in Table 2. Distributions of CALERIE<sup>TM</sup> EWAS test statistics for BMI-, cigarettesmoking-, and chronological-age-associated CpGs are shown in Figure 3. Enrichment results and gene ontological process analyses are reported in Supplementary Table 2. External EWAS CpGs and test statistics are included in Supplementary Table 3.

#### Discussion

The goal of the CALERIE<sup>™</sup> Trial was to identify the effects of CR on predictors of longevity, disease risk factors and quality of life. Published analyses of CALERIE<sup>TM</sup> data establish that the intervention improved cardiometabolic health and suggest it may have slowed or reversed aging-related biological changes (14-17,23,35). In this study, we tested whether the intervention altered whole-blood DNAm. After accounting for multiple testing, EWAS analysis revealed no sites of altered CpG methylation by CR. However, secondary analyses of sets of CpG sites, identified in published EWAS of BMI, cigarette smoking, and chronological age, indicated that the CALERIE<sup>TM</sup> intervention changed blood DNAm in a manner consistent with a reversal of DNAm patterns linked with obesity and cigarette smoking, but in the direction of older chronological age. Further interrogation across BMI-, cigarette smoking-, and chronological aging-associated sites revealed enrichment of genes associated with insulin production, glucose tolerance, inflammation, and DNA binding and regulation (Supplementary Table 2).

CALERIE<sup>TM</sup>-induced DNAm changes at BMI-associated CpG sites were enriched for genes involved in insulin production, glucose tolerance, and inflammatory processes, consistent with CR-induced epigenetic changes in animal models (7-9,31,36-40). The 26 genes enriched in CpG sites hypermethylated with higher BMI include P4HB, critical for lipoprotein metabolism, insulin production, and glucose intolerance (37-39). CR-induced hypomethylation at P4HB may mediate previously reported CR-derived metabolic improvements in lipoproteins and insulin sensitivity (15). Another potential epigenetic benefit of CR on glucose tolerance may derive from hypermethylation at cg16246545 (Supplementary Table 4), located near PHGDH. Deletion of PHGDH in adipocytes of mice with dietinduced obesity improves glucose tolerance. CR-induced methylation changes at both P4HB and PHGDH likely enhance glucose tolerance. Additional CR-induced epigenetic changes at BMI-associated sites included hypomethylation at cg19750657 (Supplementary Table 4), located near UFM1, which has been identified as a mediator of the inflammatory response in diabetic mice. Taken together, these results imply that CR, especially when maintained for 24 months, may produce anti-inflammatory benefits (31,40).

CALERIE<sup>TM</sup>-induced DNAm changes at smoking-associated CpG sites were enriched for genes involved in the tumor necrosis factor receptor-2 (TNF2) noncanonical NF-kB signaling pathway (Supplementary Table 2), a key driver of systemic inflammation (41). In addition, changes at sites with less methylation in smokers versus nonsmokers included sites identified in published EWAS of C-reactive protein (CRP) (42), a well-studied biomarker of inflammation, which is elevated in smokers and was reduced with CR in CALERIE<sup>TM</sup> (43–46). Taken together, CR appears to reverse smoking-associated DNAm patterns in inflammatory pathways.

			Hypometh	ıylated with Phenoty	ype			Hypermet	hylated with Phenot	ype		
henotype	Publication	EWAS Sample Size	Number of CpGs	12 mo CALERIE <sup>TM</sup> EWAS Median <i>T</i> -statistic	12 Months Wilcox <i>p</i> -value	24 mo CALERIE <sup>TA</sup> EWAS Median <i>T</i> -statistic	4 24 mo Wil- coxon <i>p</i> -value	Number of CpGs	12 mo CALERUE <sup>TM</sup> EWAS Median T-statistic	12 Months Wilcoxon <i>p</i> -value	24 mo CALERIE <sup>TM</sup> EWAS Median T-statistic	24 mo Wilcoxor <i>p</i> -value
3MI	Wahl et al. (18)	10,261	79	0.61	1.04E-06	0.32	7.04E-04	50	-0.93	2.06E-07	-1.08	3.96E-11
moking	Joehanes et al. 21)	15,907	1  067	-0.12	.08	-0.17	4.3E-04	1 555	-0.14	1.03E-05	-0.32	2.63E-3(
Age	McCartney et al. (19)	7 036	20	0.67	.12	0.27	.29	980	0.39	3.79E-41	0.16	5.73E-06
Age	Rönn et al. (20)	294	15	-0.08	.90	-0.47	.06	860	0.19	1.08E-09	0.05	9.04E-0∠

association between the phenotype and DNA methylation was negative). The table reports the number of ntified in pare CALERIE<sup>TM</sup>-EWAS T-statistic distributions between phenotype-associated CpGs and all other CpGs. Tests were conducted separately for CpGs hypermethylated with the phenotype (ie, for which the association between direction as the DNAm the published EWAS with *p*-value < 1E-7, with the exception of the chronological-age EWAS by McCartney et al., which reported only the top 1 000 sites (all *p*-values < 1E-7). We used a Wilcoxon Rank Sum Test to comwas in the opposite direction of DNAm association with the target phenotype. same ( the Ξ. was i intervention indicate that DNAm change in response to CALERIETM association with the target phenotype. A negative median t-statistic indicates that the DNAm change in response to CALERIE<sup>TM</sup> intervention the phenotype and DNA methylation was positive) and CpGs hypomethylated with the phenotype (ie, for which the Positive median CALERIE<sup>TM</sup> EWAS *t*-statistics and *p*-value. anlaysis of CpG sites resulting z-statistic the and in each test CpGs included Notes: The

The overwhelming majority of CpG sites identified in EWAS of chronological age exhibited greater DNAm in older as compared to younger individuals. These sites, at which we observed increased DNAm in response to CR, are enriched for multiple transcription factors and DNA binding proteins, including T-Box Transcription Factor 15 (TBX15), SRY-Box Transcription Factor 1 (SOX1), Zic Family Member 4 (ZIC4), SIM BHLH Transcription Factor 1 (SIM1), and SRY-Box Transcription Factor 17 (SOX17). Therefore, CR may induce gain of methylation parallel to aging at genomic sites serving regulatory functions. An important next step is to better understand if such gain of methylation reflects processes of aging-related decline in system integrity or, instead, genomic changes that preserve health in aging. For example, the association of CpG methylation at these sites with chronological age could reflect survivor bias, in which relatively fewer individuals with lower levels of DNAm at these sites survive to advanced ages. CR slows the accumulation of aging-related DNAm changes in mice and monkeys (11,47). Further investigation of the significance of chronological-age-associated CpG sites for phenotypes of aging is needed to clarify the interpretation of our findings. Specifically, studies are needed that establish if DNAm correlates of older chronological age are predictive of morbidity and mortality and if changes in DNAm at these loci correspond to worsening health trajectories.

We acknowledge limitations. Foremeost, response to the CR intervention was heterogeneous, as is typical in lifestyle interventions (48). Over the 2-year intervention, the treatment group achieved on average 12% CR (23). The trial sample was relatively small for genome-wide analysis; EWAS analyses were powered to detect only medium-to-large effect-size changes in DNAm at individual CpG sites. Identification of such changes is hampered by imperfect measurement precision for individual CpG-site DNAm (48), which will bias estimates of change toward the null. Nevertheless, aggregate analyses of sets of CpGs identified in prior EWAS suggest that the CALERIE<sup>TM</sup> intervention altered the blood methylome. As EWAS consortia uncover new CpG sites associated with a broader array of aging-related diseases, this analysis can be expanded. As new methods are developed to improve the precision of DNAm measurement from Illumina array data, it may be possible to revisit analyses to identify specific regions in which DNAm may be altered by the intervention (49). Future studies testing stronger doses of CR or including larger samples may also improve the detection of DNAm changes. In that light, sex-dependent effects of weight loss interventions, particularly in CR, have been identified in rodents (50). Although this study was underpowered to identify sexdependent methylation changes in response to CR, future studies should incorporate a study population and size better suited to address these phenomena. Additionally, the majority of participants enrolled in CALERIE<sup>TM</sup> were White. A priority for future trials of lifestyle interventions, including CR, is increased representation of non-White race/ethnic groups. Last, because follow-up extended only to the end of the intervention period, we cannot know if DNAm changes associated with CR persisted after the intervention concluded.

In conclusion, while CR did not result in individual CpG-site DNAm changes that reached epigenome-wide significance, analyses of sets of CpGs identified in prior EWAS of BMI, cigarette smoking, and chronological age identified clear evidence of DNAm changes in response to CR. As expected, the BMI-associated changes were consistent with CR-induced reversal of BMI-associated patterns of DNAm. Likewise, CR reversed DNAm patterns associated with



**Figure 3.** Distributions of test statistics from epigenome-wide association study (EWAS) analysis of CALERIE<sup>TM</sup> treatment effects for CpG sites identified in published EWAS of body mass index, cigarette smoking, and chronological age. The figure shows box plots of CALERIE<sup>TM</sup> EWAS test statistics for 3 groups of CpGs sites for each target phenotype. The blue-shaded box plot shows CALERIE<sup>TM</sup> EWAS test statistics for CpG sites that exhibit lower levels of DNA methylation in association with the target phenotype in published EWAS of independent samples. The red-shaded box plot shows CALERIE<sup>TM</sup> EWAS test statistics for CpG sites that exhibit higher levels of DNA methylation in association with the target phenotype in published EWAS. The gray-shaded box plot shows test statistics for CpG sites not associated with the target phenotype in published EWAS. Box plots are drawn for CALERIE<sup>TM</sup> EWAS results from 12- and 24-month follow-ups. Stars indicate *p*-value threshold for comparisons based on a Wilcoxon Rank SumTest. (\* <.05, \*\* < .005). \*\*\* < .005). Panel **A** graphs data grouped according to EWAS of cigarette smoking by Joehanes et al. (21). The figure illustrates reversal of smoking-associated DNAm changes in response to CALERIE<sup>TM</sup> intervention. Panel **B** graphs data grouped according to EWAS of chronological age by McCartney et al. (19). The figure illustrates induction of older-chronological-age-associated DNAm changes in response to CALERIE<sup>TM</sup> intervention. Panel C graphs data grouped according to EWAS of chronological age by McCartney et al. (19). The figure illustrates induction of older-chronological-age-associated DNAm changes in response to CALERIE<sup>TM</sup> intervention. Panel C graphs data grouped according to EWAS of chronological age by McCartney et al. (19). The figure illustrates induction of older-chronological-age-associated DNAm changes in response to compared to younger people.

cigarette smoking, a known correlate of premature aging. Last, and to our surprise, CR appeared to increase methylation at sites where hypermethylation is associated with older as compared to a younger age. That these sites were enriched for regulatory mechanisms suggests a complex interplay of CR with genomic changes characteristic of older age. Whether they imply pro-aging effects of CR or reflect signatures of healthy aging remains to be determined.

## **Supplementary Material**

Supplementary data are available at *The Journals of Gerontology,* Series A: Biological Sciences and Medical Sciences online.

## Funding

This work was supported by National Institute on Aging Grant R01AG061378. W.E.K. and K.M.H. are also supported by the National Institute of Aging Grant R33AG070455. V.B.K. is supported by National Institute of Health Grants R01AG054840 and P30-AG028716. R.W. is the recipient of a McKnight Scholar Award from the McKnight Brain Research Foundation through the American Brain Foundation in collaboration with the American Academy of Neurology.

## **Conflict of Interest**

None declared.

## Acknowledgment

We thank the CALERIETM Biorepository (R33AG070455) for its support of this work.

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