

Original Article

Evaluation of Antioxidant and Wound Healing Effects of Alcoholic and Aqueous Extract of *Ocimum sanctum* Linn in Rats**Somashekar Shetty¹, Saraswati Udupa² and Laxminarayana Udupa³**¹Department of Biochemistry, Melaka Manipal Medical College, ²Department of Biochemistry, KMC International Centre and ³Department of Pharmacology, Kasturba Medical College, Manipal 576104, India

In recent years, oxidative stress and free radicals have been implicated in impaired wound healing. *Ocimum sanctum* (*O. sanctum*), a plant widely used in Ayurveda, possesses anti-inflammatory and antioxidant properties. The present study was undertaken to assess the potential of alcoholic and aqueous extracts in wound healing in Wistar albino rats. The rats were divided into five groups of six animals each. Group 1 is normal wounded control and the other four groups were treated with two different doses each of alcoholic and aqueous extract of *O. sanctum*. The wound healing parameters were evaluated by using incision, excision and dead space wounds in extract-treated rats and controls. Both the doses of alcoholic and aqueous extract significantly increased wound breaking strength, hydroxyproline, hexuronic acid, hexosamines, superoxide dismutase, catalase, reduced glutathione and significantly decreased percentage of wound contraction and lipid peroxidation when compared with the control group. The results suggest that *O. sanctum* has antioxidant properties, which may be responsible and favorable for faster wound healing and this plant extract may be useful in the management of abnormal healing and hypertrophic scars.

Keywords: antioxidant property – *Ocimum sanctum* – wound healing activity

Introduction

Wound healing is the process of repair that follows injury to the skin and other soft tissues. It is fundamentally a connective tissue response. Initial stages of wound healing involve an acute inflammatory phase followed by synthesis of collagen and other extracellular matrix which are later remodeled to form scar.

Ocimum sanctum (Sanskrit:Tulasi; family: Labiaceae), is found throughout the semitropical and tropical parts of India. Different parts of the plant are traditionally used in Ayurveda and Siddha systems for the treatment of diverse ailments like infections, skin diseases, hepatic disorders and as an antidote for snake bite and scorpion sting (1). A methanol extract and an aqueous suspension

of *O. sanctum* leaves have anti-inflammatory, analgesic and immunostimulatory properties (2). Flavonoids isolated from *O. sanctum* scavenged free radicals *in vitro* and showed antilipoperoxidant activity *in vivo* at very low concentration (3). The free radical scavenging activity of plant flavonoids help in the healing of wounds (4). Low levels of antioxidants accompanied by raised levels of markers of free radical damage play a significant role in wound healing in rats (5).

Free radical scavenging activity is a major mechanism by which *O. sanctum* products protect against cellular damage (6). Based on studies in rats, *O. sanctum* may act at various levels in the immune mechanism, such as antibody production, release of mediators of hypersensitivity reactions and tissue responses to these mediators in the target organs in modulating the humoral immune responses (7). Significant inhibition of growth of pathogenic microorganisms was observed *in vitro* by traditional drugs like *O. sanctum*,

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Azardicta indica and *Annona squamosa*. Healing of the wound by indigenous ointment formulation was comparable with that of nitrofurazone and propamidine cream in mice infected by the organisms (8).

Keeping in view the tremendous pharmacological activities and a wealth of available literature, *O. sanctum* may be utilized to alleviate the symptoms of a variety of diseases as evident from pre-clinical data (9). Thus it appears that different mechanisms like free radical scavenging, metal chelation as well as immune modulation may act at different levels individually or in combination to bring about the wound healing effects of this medicinal plant.

From another viewpoint using animals for healing it is now universally acknowledged that maggot therapy can be used successfully to treat chronic, long-standing, infected wounds, which have previously failed to respond to conventional treatment. Such wounds are typically characterized by the presence of necrotic tissue, underlying infection and poor healing (10). Maggot therapy employs the use of freshly emerged, sterile larvae of the common green-bottle fly, *Phaenicia* (*Lucilia*) *sericata* and is a form of artificially induced myiasis in a controlled clinical situation. Maggot therapy has the following three core beneficial effects on a wound: debridement, disinfection and enhanced healing (11).

Methods

Plant Material



Leaves of *O. sanctum* were collected during September from the local areas of Udupi district, Karnataka, India, and were authenticated by Professor Gopalkrishna Bhat, Department of Botany, Poorna Prajna College, Udupi. A voucher specimen (No. pp531) has been deposited at the Department of Pharmacognosy, College of Pharmaceutical Sciences, Manipal, India.

Preparation of Extracts

Aqueous Extract

Leaves of *O. sanctum* were dried in shade and powdered. The leaf powder (100 g) was refluxed with 750 ml of double distilled water for 1 h at 75–80°C. It was then cooled and filtered. This was repeated in three trials. The extracts were pooled and evaporated using lyophilizer (12). The yield was about 20–25%.

Alcoholic Extract

75 g of leaf powder was extracted with 700 ml of 95% ethanol in a soxhlet apparatus at 60–75°C. Extract was concentrated by evaporation (12). The yield was about 10–15%. The semisolid extract was dissolved in saline by using gum acacia as a vehicle during the study.

Rats

Healthy Wistar albino rats of either sex and of approximately the same age, weighing between 150–250 g were used for the study. They were individually housed, maintained in clean polypropylene cages and fed with commercially pelleted rat chow (M/s Hindustan Lever Ltd., Mumbai) and water *ad libitum*. The experimental protocol was subjected to scrutiny of Institutional Animal Ethical Committee for experimental clearance (No. IAEC/KMC/UA/2000).

The animals were divided into five groups of six animals each and were daily administered the extract of *O. sanctum* via intragastric tube. The extracts were dissolved by using the vehicle gum acacia in normal saline during the study. Two grams of gum acacia was dissolved in 100 ml of normal saline. From this, 10 ml of solution, which contains 200 mg of gum acacia, was used for dissolving 1 g of aqueous and alcoholic extracts. So each ml of solution contains 100 mg of extracts.

- Group I: Control group with wound and treated with gum acacia in normal saline
- Group II: Test group with wound and treated with aqueous extract (400 mg/kg body weight)
- Group III: Test group with wound and treated with aqueous extract (800 mg/kg body weight)
- Group IV: Test group with wound and treated with alcoholic extract (400 mg/kg body weight)
- Group V: Test group with wound and treated with alcoholic extract (800 mg/kg body weight)

Experimental Procedure

The wounding procedure was carried out using ketamine (1 ml/kg body weight) anesthetized rats in three different wound models. The extract was given daily to the rats orally in the case of incision and dead space wound for

10 days. In the case of excision wound model, the extract was given everyday, till the day of epithelization.

Wound Models

Incision Wound

Two paravertebral incisions (6 cm long) were made through the full thickness of the skin on either side of the vertebral column (13). Wounds were closed with interrupted sutures, 1 cm apart. The sutures were removed on the 7th day. Wound breaking strength (WBS) was measured on the 10th post-wounding day (14).

Determination of Wound Breaking Strength

Rats were secured to the operation table and a line was drawn on either side of the wound 3 mm away from the wound. Two allice forceps were firmly applied on to the line facing each other. One of the forceps was fixed, while the other was connected to a freely suspended lightweight polypropylene graduated container through a string run over to a pulley. Water was allowed to flow from the reservoir slowly and steadily into the container. A gradual increase in weight was transmitted to the wound site pulling apart the wound edges. As and when the wound just opened up, the water flow was arrested and the volume of water collected in the container (approximately equal to its weight) was noted. Three readings were recorded for a given incision wound and the procedure was repeated on the wound on the contralateral side. The average reading of the group was taken as an individual value of breaking strength. Mean value gives the breaking strength for a given group.

Excision Wound

A circular piece of full thickness ($\sim 500 \text{ mm}^2$) was cut off from a predetermined area on the back of the rat. Wounds were traced on 1 mm^2 graph paper on the day of wounding and subsequently on the alternate days, until healing was complete. Changes in wound area were calculated, giving an indication of the rate of wound contraction. Number of days required for falling of eschar without any residual raw wound gave the period of epithelization (15).

Dead Space Wound

These wounds were created by implanting two polypropylene tubes ($0.5 \times 2.5 \text{ cm}^2$ each), one on either side in the lumbar region on the dorsal surface of each rat. On the 10th post-wounding day, the granulation tissue formed on the implanted tubes was carefully dissected out. The granulation tissues were collected, and dried at 60°C for 24 h. The dry tissue was used for the estimation of hydroxyproline (16), hexosamines (17) and hexuronic acid (18).

Granulation tissue from the other tube was collected in phosphate-buffered saline for the estimation of anti-oxidant enzymes like superoxide dismutase (SOD) (19), catalase (20), reduced glutathione (GSH) (21) and tissue lipid peroxidation (22).

Acute Toxicity Studies

Healthy rats ($n=6$) were orally fed with increasing doses (400 mg/kg to 6 g/kg body weight) of extracts for 14 days. The doses up to 4 g/kg body weight did not produce any toxicity and mortality. Of the maximum-tolerated doses (400 and 800 mg/kg), 10 and 20% were selected for the study.

Statistical Analysis

The results were analyzed using one way analysis of variance (ANOVA) with *post hoc* Scheffe's test. *P*-values <0.05 were considered statistically significant.

Results

Physical Parameters

In the incision wound model, WBS was measured. WBS was significantly increased in the *O. sanctum*-treated group compared with control with both the extracts and with both the doses. There was not much significant difference in WBS in the dose and type of extracts (Fig. 1A).

In the excision wound model, the percent of wound contraction was measured on alternate days. The percentage wound contraction was nearly 100% for the both the doses of alcoholic extract in 16 days while for the control it was 22 days ($P<0.05$). The percentage wound contraction was nearly 100% for the both the doses of aqueous extract in 14 days in comparison with controls ($P<0.05$) (Fig. 2). In the above physiological and pharmacological parameters of wound healing, aqueous extract of *O. sanctum* was more effective than the alcoholic extract.

Biochemical Parameters

In the dead space wound model, the granulation tissue hydroxyproline level was significantly increased in alcoholic extract 400 mg/kg ($P<0.01$), and in all other types of extract-treated groups compared with control ($P<0.001$). There was no significant difference among different doses of alcoholic extract. When compared with alcoholic extract 400 mg/kg, there was significant increase in hydroxyproline level in aqueous extract 800 mg/kg ($P<0.01$) (Fig. 1B). Hexuronic acid level was significantly increased in all extract-treated groups compared with control ($P<0.01$) and it was highly significant

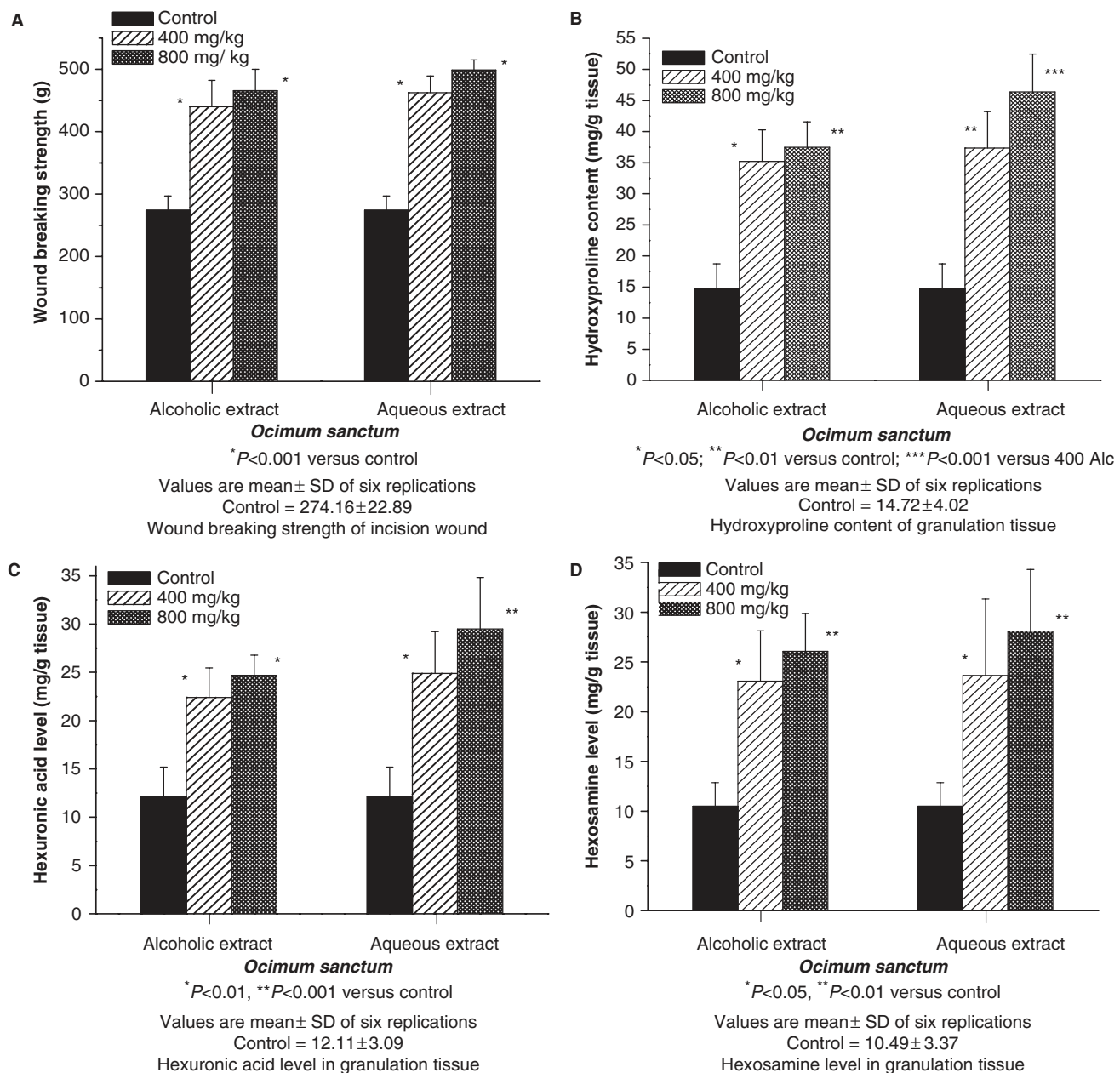


Figure 1. Effect of aqueous and alcoholic extracts of *O. sanctum* on physical (WBS) and biochemical parameters.

for aqueous 800 mg/kg treated group ($P < 0.001$) (Fig. 1C). Hexosamine level was significantly increased in aqueous and alcoholic (400 and 800 mg/kg) extract-treated groups compared with control ($P < 0.05$ and $P < 0.01$) (Fig. 1D).

Antioxidant Parameters

SOD activity in granulation tissue was significantly increased in the case of rats treated with alcoholic and aqueous extract 400 mg/kg ($P < 0.05$), alcoholic 800 mg/kg ($P < 0.05$) and aqueous extract 800 mg/kg ($P < 0.01$) when compared with control (Fig. 3A). Catalase level in

granulation tissue was significantly increased in the case of alcoholic and aqueous extract 400 mg/kg ($P < 0.05$), alcoholic 800 mg/kg ($P < 0.05$) and aqueous extract 800 mg/kg treated groups ($P < 0.01$), when compared with control (Fig. 3B). Reduced GSH concentration in granulation tissue was significantly increased in rats treated with different extracts ($P < 0.001$), when compared with control. There was significant increase in GSH level in aqueous 800 mg/kg treated group when compared with alcoholic 400 mg/kg group ($P < 0.05$) in intercomparison statistical analysis (Fig. 3C). Malondialdehyde (MDA) level (which indicates lipid peroxidation) in granulation tissue was significantly

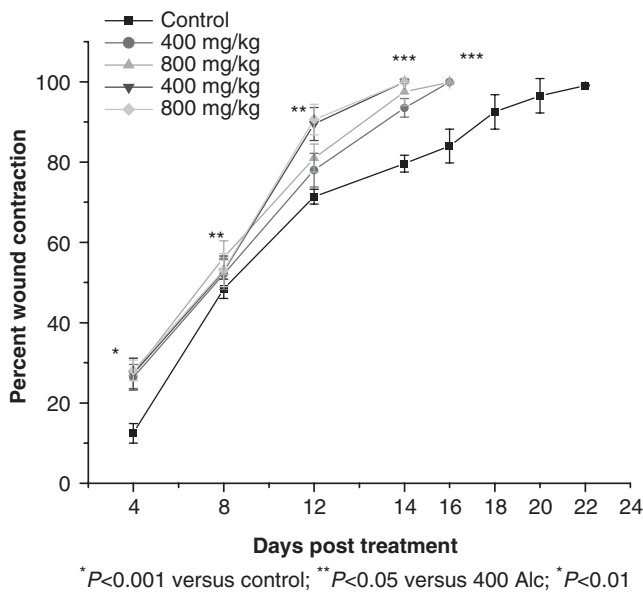


Figure 2. Effect of aqueous and alcoholic extracts of *O. sanctum* on excision wound model.

decreased in the case of all the extract-treatment groups ($P < 0.001$) compared with controls (Fig. 3D).

Discussion

Granulation, collagen maturation and scar formation are some of the many phases of wound healing, which run concurrently, but independent of each other. The use of a single model is inadequate and no reference standard exists that can collectively represent the various phases of wound healing. Hence, three different models have been used in our study to assess the effect of *O. sanctum* on the various phases of wound healing.

In incision wound, the increase in tensile strength of treated wounds may be due to the increase in collagen concentration and stabilization of the fibers (23). Increase in WBS and role of antioxidants were experimentally proved by Michel and Fredrickson (24). In excision wound, both the extracts showed faster healing compared with control groups and wound contraction rate is faster with aqueous extract as compared with alcoholic extract. The faster wound contraction rate of the aqueous extract may be due to stimulation of interleukin-8, an inflammatory α -chemokine which affects the function and recruitment of various inflammatory cells, fibroblasts and keratinocytes. It may increase the gap junctional intracellular communication in cultured fibroblasts and induces a more rapid maturation of granulation tissue (25). The aqueous extract of *O. sanctum* increased cellular proliferation and collagen synthesis at the wound site as evidenced by increase in total protein and total

collagen contents reflected by hydroxyproline content of granulation tissues.

The glycosaminoglycans are a major component of the extracellular matrix of skin, joints, eyes and many other tissues and organs. In spite of its simple structure, it demonstrates remarkable viscoelastic and hygroscopic properties which are relevant for dermal tissue function. Biological activities in skin are due to its interaction with various binding proteins. Due to an influence on signaling pathways, hyaluronic acid is involved in the wound healing process and scarless fetal healing. In clinical trials, topical application of hyaluronic acid has improved the healing of wound (26). In addition, the mucopolysaccharide hyaluronic acid protects granulation tissue from oxygen free radical damage and thereby stimulates wound healing (27).

Among the glycosaminoglycans, dermatan sulfate and dermatan have also been implicated in wound repair and fibrosis. Their ability to bind and alter protein-protein interactions has identified them as important determinants of cellular responsiveness in development, homeostasis and disease (28). In our study, hexuronic acid and hexosamine concentrations which are the component of glycosaminoglycans were significantly increased with both the extracts and with both the doses when compared with control. The glycosaminoglycans are known to stabilize the collagen fibers by enhancing electrostatic and ionic interactions with it and possibly control their ultimate alignment and characteristic size. Since *O. sanctum* extracts have increased the levels of these compounds considerably, it is likely that the observed increase in tensile strength was not only due to increased collagen synthesis but also due to its proper deposition and alignment.

Molecular oxygen plays a central role in the pathogenesis and therapy of chronic wounds. Overproduction of reactive oxygen species (ROS) results in oxidative stress thereby causing cytotoxicity and delayed wound healing. Therefore, elimination of ROS could be an important strategy in healing of chronic wounds (29). Therefore, estimation of antioxidants like SOD, catalase and glutathione in granulation tissues is also relevant because these antioxidants hasten the process of wound healing by destroying the free radicals (30). The significant alteration in the antioxidant profile accompanied by the elevated levels of MDA, a marker of free radical damage, may be attributed to impaired wound healing in immunocompromised rats (31). While numerous attempts have been made to identify prognostic biomarkers of wound healing in skin, these have met with limited success. The bioactivity of flavonoids tightly correlated with their chemical structure and action mechanisms, mostly inhibitory on enzymatic systems involved in cellular activations (32). Study of 12 different plant flavonoids, mainly orientin and vicenin of *O. sanctum* has

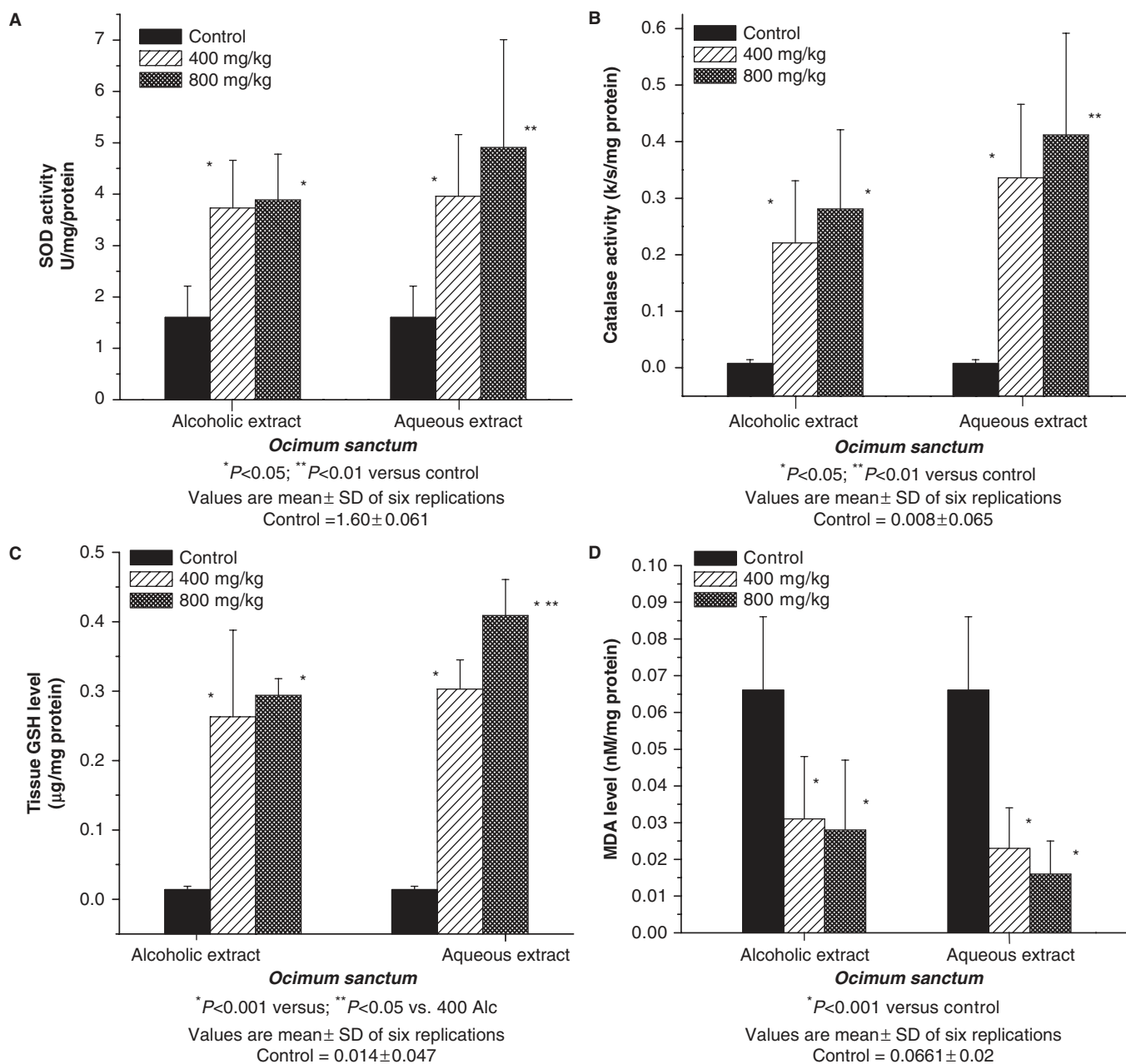


Figure 3. Effect of alcoholic and aqueous extracts of *O. sanctum* on antioxidant parameters. (A) SOD activity in granulation tissue; (B) catalase activity in granulation tissue; (C) reduced GSH level in granulation tissue; (D) lipid peroxidation-MDA level in granulation tissue.

revealed that their *in vitro* antioxidant activity works also under *in vivo* conditions (33).

In case of the antioxidant parameters, rats treated with the aqueous extracts of *O. sanctum* in the dose of 800 mg/kg body weight showed significant increase in the activity of SOD catalase and GSH with a decrease in MDA level in granulation tissue compared with controls. These enzymes are known to quench the superoxide radical and thus prevent the damage of cells caused by free radicals (34). Better collagenation, seen under the influence of this plant extract, may be because of the presence of flavonoids, which is responsible for the

free radical scavenging activity which is believed to be one of the most important components of wound healing (35).

In our study on the effect of alcoholic and aqueous extracts of *O. sanctum* on wound healing (400 and 800 mg/kg body weight), we found that aqueous extract possesses a better effect than alcoholic extract at a dose of 800 mg/kg body weight. Since *O. sanctum* is ubiquitous and abundantly grown, it could be a fairly economical therapeutic agent for wound management as a prohealer, as well as to control abnormal healing.

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Received June 15, 2006; accepted January 11, 2007



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