

UNIVERSIDADE FEDERAL DO RIO GRANDE DO SUL
FACULDADE DE FARMÁCIA
TRABALHO DE CONCLUSÃO DE CURSO

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Rafael Teixeira Ribeiro

Porto Alegre, Dezembro de 2017.

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**Prof. Dr. Moacir Wajner
Orientador**

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Experimental evidence that *in vivo* intracerebral administration of L-2-hydroxyglutaric acid to neonatal rats provokes disruption of redox status and histopathological abnormalities in the brain

Rafael Teixeira Ribeiro³, Ângela Zanatta¹, Alexandre Umpierrez Amaral^{1,2}, Guilhian Leipnitz^{1,3}, Bianca Seminotti¹, Moacir Wajner^{1,3,4}

1 - Programa de Pós-Graduação em Ciências Biológicas: Bioquímica, Instituto de Ciências Básicas da Saúde, Universidade Federal do Rio Grande do Sul, Porto Alegre, RS, Brazil

2 - Departamento de Ciências Biológicas, Universidade Regional Integrada do Alto Uruguai e das Missões, Erechim, RS, Brazil

3 - Departamento de Bioquímica, Instituto de Ciências Básicas da Saúde, Universidade Federal do Rio Grande do Sul, Rua Ramiro Barcelos, 2600-Anexo, Porto Alegre, RS CEP 90035-003, Brazil

4 - Serviço de Genética Médica, Hospital de Clínicas de Porto Alegre, Porto Alegre, RS, Brazil

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***Corresponding author:** Moacir Wajner, Departamento de Bioquímica, Instituto de Ciências Básicas da Saúde, Universidade Federal de Rio Grande do Sul, Rua Ramiro Barcelos, 2600 – Anexo, CEP 90035-003, Porto Alegre, RS, Brazil.

Abstract

Tissue accumulation of L-2-hydroxyglutaric acid (L-2-HG) is the biochemical hallmark of L-2-hydroxyglutaric aciduria (L-2-HGA), a rare neurometabolic inherited disease characterized by neurological symptoms and brain white matter abnormalities whose pathogenesis is not yet well established. L-2-HG was intracerebrally administered to rat pups at postnatal day 1 (P1) to induce a rise of L-2-HG levels in the central nervous system (CNS). Thereafter, we investigated whether L-2-HG *in vivo* administration could disturb redox homeostasis and induce brain histopathological alterations in cerebral cortex and striatum of neonatal rats. L-2-HG markedly induced the generation of reactive oxygen species (increase of 2',7'-dichlorofluorescein-DCFH- oxidation), lipid peroxidation (increase of malondialdehyde concentrations) and protein oxidation (increase of carbonyl formation and decrease of sulfhydryl content), besides decreasing the antioxidant defenses (reduced glutathione-GSH) and sulfhydryl content in cerebral cortex. Alterations of the activities of various antioxidant enzymes were also observed in cerebral cortex and striatum following L-2-HG administration. Furthermore, L-2-HG-induced lipid peroxidation and GSH decrease in cerebral cortex were prevented by the antioxidant melatonin and by the classical antagonist of NMDA glutamate receptor MK-801, suggesting the involvement of reactive species and of overstimulation of NMDA receptor in these effects. Finally, L-2-HG provoked vacuolation and edema particularly in the cerebral cortex with less intense alterations in the striatum that were possibly associated with the unbalanced redox homeostasis caused by this metabolite. Taken together, it is presumed that these pathomechanisms may underlie the neurological symptoms and brain abnormalities observed in the affected patients.

Keywords: L-2-hydroxyglutaric aciduria; L-2-hydroxyglutaric acid; redox homeostasis; cerebral cortex; striatum; histopathology.

Introduction

L-2-Hydroxyglutaric aciduria (L-2-HGA; MIM#236792) is a rare autosomal recessive inborn error of metabolism caused by mutations in the *L-2-hydroxyglutarate dehydrogenase* gene (*L2HGDH*), resulting in highly elevated levels of L-2-hydroxyglutaric acid (L-2-HG) in tissues and biological fluids (urine, plasma, and cerebro-spinal fluid). The *L2HGDH* gene is expressed in multiple tissues, with the strongest expression in the brain [1]. To date, there are over three hundred documented cases of this disease in the medical literature [2-7]. The main neurological manifestations include progressive mental retardation, variable motor impairment and cerebellar ataxia. Most patients also present macrocephaly, pyramidal and extrapyramidal signs, dystonia and a typical MRI profile with striking subcortical leucoencephalopathy and basal ganglia abnormalities [5,8]. Characteristic brain MRI findings in L-2-HGA comprise alterations in the subcortical cerebral white matter, dentate nucleus, globus pallidus, putamen and caudate nucleus [5]. As the disease progresses, alterations of white matter and basal ganglia signal intensities become more diffuse, followed by cerebral white matter atrophy [3]. A strong correlation between the clinical disability and the extent of pathologic changes on MRI has been reported, but this is still under debate [5].

The mechanisms of the neurological symptoms presented by L-2-HGA patients are still poorly understood. However, increased concentrations of L-2-HG were associated with the appearance of neurological symptoms, indicating that this organic acid may be potentially neurotoxic in this disorder [9-12]. In this context, it was verified that L-2-HG inhibits creatine kinase activity in rat cerebellum homogenates [13], induces oxidative stress [14] and increases glutamate uptake in synaptosomes and synaptic vesicles [15]. Furthermore, it was demonstrated that intracerebral administration of L-2-HG to rats induces oxidative stress by decreasing the antioxidant defenses and raising reactive oxygen species in striatum and cerebellum of rats, besides causing marked brain histopathological alterations [16].

Considering that the brain is highly susceptible to free radical attack due to its low antioxidant defenses and high oxygen consumption potentially leading to a proportionally high mitochondrial production of reactive species [17], in the present study we investigated the effects of intracerebral administration of L-2-HG on a large spectrum of redox homeostasis parameters in cerebral cortex and striatum of neonatal rats. Lipid oxidative damage (malondialdehyde levels), protein oxidation (carbonyl formation and sulfhydryl content), reactive oxygen species (DCFH oxidation), and the non-enzymatic (GSH levels) and enzymatic antioxidant defenses (glutathione peroxidase, glutathione reductase, glutathione S-transferase, glucose-6-phosphate dehydrogenase, catalase and superoxide dismutase) were evaluated. We also investigated whether L-2-HG could provoke-histopathological changes in the brain of these animals.

Material and Methods

Animals

Neonatal Wistar rats obtained from the Central Animal House of the Department of Biochemistry, ICBS, Universidade Federal do Rio Grande do Sul (UFRGS), Porto Alegre, RS, Brazil, were used. Six whole litters of 12 animals each were used for each experimental *in vivo* condition. The animals had free access to water and 20% (w/w) protein commercial chow (SUPRA, Porto Alegre, RS, Brazil) and were maintained on a 12:12-h light/dark cycle in an air-conditioned constant temperature ($22\text{ }^{\circ}\text{C} \pm 1\text{ }^{\circ}\text{C}$) colony room. The experimental protocol was approved by the Ethics Committee for animal research of the Universidade Federal do Rio Grande do Sul, Porto Alegre, Brazil, and followed the “Guide for the Care and Use of Laboratory Animals” (NIH publication 85-23, revised 2011). All efforts were made to minimize the number of animals used and their suffering.

Reagents

All chemicals were purchased from Sigma Chemical Co., St. Louis, MO, USA. L-2-HG was dissolved in phosphate-buffered saline (PBS) on the day of the experiments, and the pH was adjusted to 7.4.

L-2-HG administration and drug treatment

Seventy-two neonatal pups were used in the experiments. Each littermate was injected intracerebrally into the Cisterna magna, as previously described [18,19], between 12 and 48 hours after birth (P1) with L-2-HG (0.75 $\mu\text{mol/g}$, pH 7.4) or vehicle (phosphate buffered saline (PBS), 0.01 M, pH 7.4). After injection, animals were allowed to recover during 15–30 min and returned to their mother. In some experiments, the antioxidant melatonin (MEL; 20 mg/kg body weight) [20] or the classical NMDA receptor antagonist MK-801 (0.25 mg/kg body weight) [16] was administered intraperitoneally 60 min before L-2-HG administration.

Sample Preparation

Six hours after L-2-HG injection, animals were euthanized by decapitation, and the brain was immediately removed and kept on an ice Petri plate. The olfactory bulb, pons, and medulla were discarded and the cerebral cortex and striatum were dissected, weighed, and kept chilled until homogenization. These structures were homogenized (1:10 w/v) in 20 mM sodium phosphate, pH 7.4, containing 140 mM chloride potassium (1:10 w/v), centrifuged at 750 g for 10 min and the supernatants used for the assays.

Malondialdehyde Levels

MDA concentrations were measured according to the method described by Yagi [21] with some modifications. Briefly, one hundred microliters of cerebral cortex or striatum supernatants containing 0.3 mg of protein were treated with 200 μ L of 10 % trichloroacetic acid and 300 μ L of 0.67 % thiobarbituric acid in 7.1 % sodium sulfate and incubated for 1 h in a boiling water bath. The tubes containing the mixture were allowed to cool on running tap water for 5 min. The resulting pink-stained complex was extracted with 400 μ L of butanol. Fluorescence of the organic phase was read at 515 and 553 nm as excitation and emission wavelengths, respectively. A calibration curve was performed using 1,1,3,3-tetramethoxypropane and subjected to the same treatment as supernatants. MDA levels were calculated as nmol MDA / mg protein.

Sulfhydryl Group Content

This assay was performed according to Aksenov and Markesbery [22]. It is based on the reduction of 5,5'-dithio-bis(2-nitrobenzoic acid) DTNB by thiol groups, generating a yellow-stained compound TNB whose absorption is measured spectrophotometrically at 412 nm. Thirty microliters of 0.1 mM DTNB was added to 120 μ L of cerebral cortex or striatum supernatants containing 0.36mg of protein. This was followed by 30-min incubation at room temperature in a dark room. Absorption was measured at 412 nm. Results were calculated as nmol TNB / mg protein.

Protein carbonyl formation

Protein carbonyl formation was measured according to the method of Reznick and Packer [23]. The assay is based on the reaction of the carbonyl groups with 2,4-dinitrophenylhydrazine (DNPH) generating a corresponding dinitrophenyl hydrazone, whose absorbance is read in spectrophotometer at 370 nm. Briefly, 200 μ L of cerebral cortex or striatum supernatants were treated with 400 μ L of 10 mM DNPH dissolved in 2.5 M HCl or with 2.5 M HCl (blank) and left in the dark for 1 h. Samples were then precipitated with 600 μ L of 20 % TCA and centrifuged for 5 min at 9,000 g. The pellet was washed with 1 mL of ethanol:ethyl acetate (1:1, v/v) and suspended in 550 μ L of 6 M guanidine prepared in 2.5 N HCl. The difference between the absorbance of the samples treated with DNPH and with HCl (blank) was used to calculate the carbonyl content. The results were calculated as nmol carbonyl groups / mg of protein, using the extinction coefficient of $22,000 \times 10^6$ nmol / mL for aliphatic hydrazones.

Reduced glutathione concentrations

This parameter was measured according to Browne and Armstrong [24] with some modifications. One hundred and eighty-five microliters of 100 mM sodium phosphate buffer, pH 8.0, containing 5 mM EDTA, and 15 μ L of o-phthalaldehyde (1 mg/mL) were added to 30 μ L of sample (0.3–0.5 mg of protein) previously deproteinized with metaphosphoric acid. This mixture was incubated at room temperature in a dark room for 15 min. Fluorescence was measured using excitation and emission wavelengths of 350 and 420 nm, respectively. Calibration curve was prepared with standard GSH (0.001–1 mM), and the concentrations were calculated as nmol GSH / mg protein.

2',7'-Dichlorofluorescein oxidation

DCFH oxidation was determined according to the method of LeBel [25] with slight modifications. Homogenates (0.04–0.07 mg protein) were incubated with 2',7'-dichlorofluorescein diacetate (DCF-DA) (5 μ M) during 1 h at 37°C. DCF-DA is permeable to the cell membrane and is deacetylated by esterases to DCFH in the intracellular medium. DCFH is then converted by reactive species into the highly fluorescent product dichlorofluorescein (DCF). DCF fluorescence was measured using wavelengths of 480 nm (excitation) and 535 nm (emission). The calibration curve was performed with standard DCF (0.01–0.75 μ M). The production of reactive species was calculated as μ mol DCF / mg protein.

Nitrate and nitrite content

Reactive nitrogen species (RNS) formation was evaluated by measuring nitrate and nitrite content according to Navarro-Gonzalvez and collaborators [26], with some modifications. One hundred and fifty microliters of supernatants (containing approximately 1.2 mg of protein) was deproteinized by adding 125 mL of 75 mM ZnSO₄ solution, followed by centrifugation at 9,000 g for 2min at 25°C. The supernatant obtained was neutralized with 55mM NaOH solution and diluted in 5 vol of glycine buffer solution, pH 9.7. Copper-coated cadmium granules (600–1,000 mg) were added to the supernatants to convert all nitrates into nitrite in the biological samples. Aliquots of 200 μ L were then treated with 200 μ L of Griess reagent (2% sulfanilamide in 5% HCl and 0.1% N-1-(naphthyl)ethylenediamine in H₂O) and incubated at room temperature for 10 min. The absorbance was read at 505nm. A calibration curve was prepared with NaNO₂ at concentrations ranging from 1 to 125 mM. The final results were expressed in μ mol nitrate and nitrite / mg protein.

Glutathione peroxidase (GPx) activity

GPx activity was measured according to Wendel [27] using tert-butylhydroperoxide as substrate. The enzyme activity was determined by monitoring the NADPH disappearance at 340 nm in a medium containing 600 μ L of buffer (100 mM potassium phosphate containing 1 mM EDTA, pH 7.0), 10 μ L of 40 mM sodium azide, 15 μ L of 100 mM GSH, 15 μ L of 10 U/mL glutathione reductase, 10 μ L of 10 mM NADPH, and 10 μ L of sample (3 μ g of protein). One GPx unit (U) is defined as 1 μ mol of NADPH consumed per minute. The specific activity was calculated as U / mg protein.

Glutathione reductase (GR) activity

GR activity was measured as previously described by Carlberg and Mannervik [28]. The enzyme activity was determined by monitoring NADPH consumption at 340 nm in a medium with 200 mM sodium phosphate buffer, pH 7.5, containing 6.3 mM ethylenediaminetetraacetic acid, 1 mM GSSG, 0.1 mM NADPH, and approximately 3 µg of protein. One GR unit (U) is defined as 1 µmol of GSSG reduced per minute. The specific activity was calculated as U / mg protein.

Glutathione-S-transferase (GST) activity

GST activity was measured according to Mannervik and Guthenberg [29], with slight modifications, by monitoring the rate of formation of dinitrophenyl-S-glutathione at 340 nm in a medium containing 50 mM potassium phosphate, pH 6.5, 1 mM GSH, 1 mM 1-chloro-2,4-dinitrobenzene (CDNB) as substrate, and tissue supernatants (approximately 0.045 mg of protein). The results were calculated as U / mg protein.

Glucose-6-phosphate dehydrogenase (G6PDH) activity

G6PDH activity was measured according to Leong and Clark [30], following NADPH formation at 25°C at 340 nm. The reaction medium contained tissue supernatants (approximately 45 µg of protein), 100 mM Tris-HCl buffer, pH 7.5, 1.0 mM MgCl₂, 0.05 mM NADP⁺ and 0.1 mM glucose-6-phosphate. The results were calculated as U / mg protein.

Catalase (CAT) activity

CAT activity was assayed according to Aebi [31] by measuring the absorbance decrease at 240 nm in a reaction medium containing 20 mM H₂O₂, 0.1 % Triton X-100, 10 mM potassium phosphate buffer, pH 7.0, and approximately 1 µg of protein. One unit (U) of the enzyme is defined as 1 µmol of H₂O₂ consumed per minute. The specific activity was calculated as U / mg protein.

Superoxide dismutase (SOD) activity

SOD activity was assayed according to Marklund [32] and is based on the capacity of pyrogallol to autoxidize, a process highly dependent on superoxide anion, which is the substrate for SOD. The inhibition of the autoxidation of pyrogallol occurs in the presence of SOD and, therefore, is proportional to the activity of the SOD present in homogenates. The reaction medium contained 50 mM Tris buffer/1 mM ethylenediaminetetraacetic acid, pH 8.2, 80 U/mL CAT, 0.38 mM pyrogallol, and approximately 1 µg of protein, and the absorbance was read at 420 nm. A calibration curve was performed with purified SOD as standard, in order to calculate the activity of SOD present in the samples. The specific activity was calculated as U / mg protein.

Histological analysis

We performed histological analysis in the brain with hematoxylin and eosin staining. Four rats of each group (PBS or L-2-HG-injected animals) were euthanized 24 hours after PBS or L-2-HG (0.75 $\mu\text{mol/g}$) administration. The whole brain was removed and postfixed in 10% formaldehyde-buffered solution, pH 7.0, for 48 hours at room temperature and processed for paraffin-embedded sectioning. Three-micrometer-thick brain slices were obtained with a microtome (MICROM HM 360). Brain sections were stained in hematoxylin solution for 5 min and then washed in running tap water. The sections were then placed in eosin solution for 3 min, dehydrated with 100 % ethanol and cleared with xylene for 2 min. Image analysis (magnification of 100x and 400x) was performed using the Q Capture Pro Software (Olympus).

Protein Determination

Protein content was measured by the method of Lowry [33], using bovine serum albumin as a standard.

Statistical Analysis

Results are presented as mean \pm standard deviation. Assays were performed in triplicate, and the mean was used for statistical calculations. Data were analyzed using the Student's *t* test for unpaired samples or one-way analysis of variance (ANOVA) followed by the post-hoc Duncan multiple range test when F was significant. Only significant values are shown in the text. Differences between groups were considered significant at $P < 0.05$. All analyses were carried out using the GraphPad Prism 5 software.

Results

In vivo L-2-HG administration provokes lipid and protein oxidative damage in neonatal rat cerebral cortex

We initially observed that L-2-HG markedly enhanced (75%) MDA levels in cerebral cortex of neonatal rats 6 h after its administration [$t_{(7)} = 4.321$; $P < 0.01$] (Figure 1A). Since MDA is an end product of membrane polyunsaturated fatty acid peroxidation, these data indicate that L-2-HG causes lipid oxidative damage. L-2-HG also increased carbonyl formation (11%) [$t_{(5)} = 3.317$; $P < 0.05$] (Figure 1B), and decreased sulfhydryl content (34%) [$t_{(5)} = 5.020$; $P < 0.01$] (Figure 1C). Since carbonyl group generation is currently used as a marker of free radical-mediated protein oxidation and two thirds of total cellular sulfhydryl groups are protein bound [34,35], our results indicate that L-2-HG induces protein oxidative damage in cerebral cortex.

L-2-HG decreases the antioxidant defenses in rat cerebral cortex

Next, we observed that L-2-HG decreased the concentrations of GSH (25%) [$t_{(8)} = 5.655$; $P < 0.001$] (Figure 2), implying an impairment of brain antioxidant defenses by this organic acid.

L-2-HG induces reactive oxygen species (ROS) generation in rat cerebral cortex

We also investigated whether L-2-HG could induce reactive species generation that could possibly lead to protein and lipid oxidative damage as well as to reduction of antioxidant defenses. Thus, we tested the effects of L-2-HG on ROS production, determined by DCFH oxidation in cerebral cortex, and on RNS generation measured by nitrate and nitrite concentrations. L-2-HG significantly increased DCFH oxidation (up to 25 %) [$t_{(5)} = 2.914$; $P < 0.05$] (Figure 3A) but, in contrast, did not change the levels of nitrates and nitrites (Figure 3B).

L-2-HG-induced lipid peroxidation and decreased antioxidant defenses are prevented by melatonin (MEL) and MK-801 in neonatal rat cerebral cortex

Since L-2-HG induced ROS formation, we tested whether the reactive species scavenger MEL could prevent lipid oxidation and the decrease of the antioxidant defenses caused by this metabolite. We observed that MEL totally prevented MDA increase [$t_{(4)} = 2.788$; $P < 0.05$] (Figure 4A) and GSH decrease [$t_{(6)} = 7.476$; $P < 0.001$] (Figure 4B) caused by L-2-HG. These data support the involvement of ROS in these effects and may offer a new perspective of treatment with antioxidants. Furthermore, the effects on MDA levels [$t_{(5)} = 3.995$; $P < 0.05$] and GSH concentrations [$t_{(6)} = 4.208$; $P < 0.01$] were similarly abolished by the NMDA receptor antagonist MK-801 (Figures 4A and 4B), indicating overactivation of NMDA receptors by L-2-HG.

L-2-HG modulates the activities of antioxidant enzymes in neonatal rat cerebral cortex

The effects of L-2-HG intracerebroventricular administration on the enzymatic antioxidant defenses were further studied. We verified that L-2-HG reduced G6PDH activity (14%) [$t_{(6)} = 2.775$; $P < 0.05$] (Figure 5A), and increased GPx (18%) [$t_{(5)} = 4.326$; $P < 0.05$] (Figure 5B), GR (20%) [$t_{(6)} = 4.326$; $P < 0.01$] (Figure 5C) and GST (39%) [$t_{(6)} = 3.914$; $P < 0.01$] (Figure 5D) activities. In contrast, CAT (Figure 5E) and SOD (Figure 5F) activities were not altered by the metabolite.

L-2-HG mildly disrupts redox homeostasis in neonatal rat cerebral striatum

We also observed that L-2-HG administration decreased sulfhydryl content (50%) [$t_{(4)} = 10.13$; $P < 0.001$] (Figure 6A), but did not alter MDA levels, GSH concentrations and ROS formation (DCFH oxidation) in rat striatum. It was also verified that L-2-HG modulated the enzymatic antioxidant defenses by increasing GPx (35%) [$t_{(5)} = 3.255$; $P < 0.05$] (Figure 6B), G6PDH (20%) [$t_{(5)} = 3.354$; $P < 0.05$] (Figure 6C) and SOD (22%) [$t_{(7)} = 4.633$; $P < 0.01$] (Figure 6D) activities, with no changes in the activities of GR and GST.

L-2-HG provokes extensive vacuolation in neonatal rat cerebral cortex

The effect of L-2-HG intracerebroventricular administration on the brain histological integrity was then studied by HE staining. We observed that L-2-HG administration provoked intense edema and vacuolation in the cerebral cortex (Figure 7) with less effects in the striatum (Figure 8).

Discussion

L-2-HGA is clinically characterized by predominant neurological findings, such as psychomotor delay/mental deficiency, extrapyramidal and pyramidal symptoms, macrocephaly, seizures demyelination of white matter, cerebral and cerebellar atrophy, seizures, leukoencephalopathy, and other neurologic manifestations [4-6]. In some patients, this disorder leads to the formation of brain tumors [36-38], whose etiology is still poorly known. Accumulation of L-2-HG in tissues, predominantly in the brain, is the biochemical hallmark of this disease, and this may be because the mutated gene is highly expressed in the brain [1]. Although the disorder is predominantly neurologic, its pathogenesis is not yet established. However, it was suggested that the pathologic phenotypes of affected patients result from the toxic effects of L-2-HG on central nervous system [39]. In this context, the structural resemblance of L-2-HG to L-glutamate and α -ketoglutarate, two major metabolites of the glutamatergic system, makes it likely that the pathophysiology of the disease may involve alterations of this system or pathways of α -ketoglutarate metabolism. Furthermore, previous *in vitro* and *in vivo* studies have shown disruption of redox homeostasis provoked by L-2-HG, as well as alterations of the glutamatergic system in developing rats and affected patients with L-2-HGA [14-16,40].

It is of note that nothing has been reported on the brain toxicity of L-2-HG in neonatal brain. Therefore, in the present study we investigated whether L-2-HG *in vivo* administration could disturb redox status and induce histopathological alterations in cerebral cortex and striatum of neonatal rats. It is emphasized that these cerebral structures are mainly affected in L-2-HGA, as observed in cerebral imaging by leukoencephalopathy/spongiosis of white matter, subcortical abnormalities and caudate and putamen alterations [41,42].

We verified that L-2-HG administration markedly increased MDA levels in cerebral cortex, corroborating the *in vitro* and *in vivo* findings previously reported and showing that L-2-HG increases the levels of thiobarbituric acid-reactive substances in brain of 30-day-old rats [14,16]. Since MDA is an end product of membrane fatty acid peroxidation, these data indicate that L-2-HG induces lipid oxidative damage. L-2-HG also markedly increased carbonyl groups and decreased sulfhydryl content. Since carbonyl groups are formed by oxidation of amino acid side chain protein residues (Pro, Arg, Lys, and Thr) and two thirds of sulfhydryl groups in cells are protein bound [34,35], it is assumed that L-2-HG induces protein oxidative damage in the brain.

L-2HG also caused a significant decrease of GSH concentrations in cerebral cortex that could be tentatively attributed to its oxidation by increased reactive species generation. Since GSH represents the major naturally-occurring tissue antioxidant and plays a key role in maintaining the intracellular redox

state protecting crucial cellular sulfhydryl groups [17], our present findings suggest that L-2-HG impairs the tissue non enzymatic antioxidant defenses.

DCFH oxidation was also increased by L-2-HG, but, in contrast, this organic acid did not change nitrate and nitrite content, reflecting induction of ROS generation. It could be also hypothesized that the increase of ROS formation could lead secondarily to increased lipid and protein oxidation and decreased GSH concentrations. On the other hand, since GSH is considered an important defense against lipid oxidative damage by scavenging reactive species responsible for the initiation of this process, it is feasible that the decrease of this antioxidant could make lipids and proteins more vulnerable to the oxidative damage caused by L-2-HG through ROS formation.

Moreover, the free radical scavenger melatonin prevented L-2-HG-induced reduction of GSH levels and increase of MDA concentrations, strongly indicating that disturbance of cellular redox status was secondary to reactive species production provoked by L-2-HG. It is likely that hydroxyl and peroxy radicals were involved in these effects, since these reactive species are scavenged by melatonin [17,43,44]. On the other hand, it is feasible that NMDA receptor overstimulation was involved in the L-2-HG-induced impairment of cell redox since the classical antagonist of these receptors MK-801 prevented lipid oxidative damage and antioxidant defense decrease.

We also observed that L-2-HG increased GPx, GR and GST activities, and inhibited G6PD activity in cerebral cortex. We presume that the increased activities of GPx and GST might have occurred due to the overexpression of these enzymes at gene level in order to compensate the increased formation of lipid peroxides, which is in line with the fact that L-2-HG increases MDA levels, a lipid peroxidation end product. In this particular, it is known that GST also detoxifies 4-hydroxynonenal and isoprostanes that are lipid peroxidation products [17]. Furthermore, increased GR activity also suggests overexpression of this enzyme protein aiming to recycle GSH once the concentrations of this antioxidant are decreased. As regards to the L-2-HG-induced decrease of G6PD activity, we speculate that it occurred due to reactive species attack causing modifications in the enzyme protein structure as previously shown for this and other antioxidant enzymes [45,46].

Since oxidative stress results from an imbalance between the total antioxidant defenses and the reactive species generated in a tissue, our present data strongly indicate that L-2-HG induces oxidative stress *in vivo* in rat cerebral cortex. It must be emphasized that ROS are capable to oxidize various molecules, including lipids, proteins, sugars, and DNA, representing a key event in the pathogenic cascade leading to necrotic and apoptotic cell death [47,48]. Furthermore, oxidative stress is a very deleterious condition especially to the brain due to its high rate of oxidative metabolism coupled to ROS production, lower activity of antioxidant enzymes, and higher peroxidation potential (high content of polyunsaturated fatty acids) [17].

It was also demonstrated in this study that L-2-HG provoked extensive edema and vacuolation in the cerebral cortex of the animals, with less intense effects in the striatum. Although not proven, we assume that these brain abnormalities could be possibly related to the disruption of redox homeostasis elicited by this organic acid [16,49-51].

It is difficult at present to determine the pathophysiological relevance of our data since brain concentrations of L-2-HG in L-2-HGA are still unknown. We cannot rule out the possibility that intracerebral L-2-HG concentrations in the present study were supraphysiological, so that our findings must be taken carefully. However, the concentrations of organic acids accumulating within neural cells in organic acidemias were reported to be usually much higher than those of plasma and cerebrospinal fluid (trapping theory) of patients with various organic acidemias [52]. Furthermore, L-2-HGA is considered a cerebral organic acidemia because L-2-HG is trapped in the CNS [53,54], where at high concentrations it potentially can exert its neurotoxic effects disturbing cellular redox homeostasis.

Conclusion

In conclusion, this is the first report showing that L-2-HG administration to neonatal rats induces *in vivo* lipid and protein oxidative damage and disrupts the antioxidant defenses mainly in the cerebral cortex. These effects were prevented by the potent antioxidant melatonin and the NMDA antagonist MK-801, implying that reactive species generation and overstimulation of these glutamate receptors may underlie the altered cellular redox status. These *in vivo* data allied to previous *in vitro* [14] and *in vivo* [16] experimental animal and humans studies [40,55] strongly indicate that imbalance of redox homeostasis may potentially contribute at least in part to the neurologic alterations found in L-2-HGA. Moreover, it is conceivable that sustained elevated levels of L-2-HG found in the patients may lead to chronic oxidative stress. Another novel and important finding of the present study is that this organic acid provoked histopathological alterations more evident in the cerebral cortex probably due to the induced disruption of redox homeostasis. It may be presumed that our present findings acting synergistically with other intrinsic (severity of mutations) and extrinsic factors may contribute to explain the pathophysiology of the neurologic symptoms and brain abnormalities observed in the affected patients. In case our present *in vivo* results can be confirmed in tissues (blood and cultivated fibroblasts) from individuals affected by L-2HGA, it is therefore tempting to speculate that antioxidants could be supplemented to the affected patients as a potential adjuvant therapy to prevent or attenuate oxidative damage.

Compliance with Ethical Standards

Ethical Approval

The experimental protocol was approved by the Ethics Committee for Animal Research of the UFRGS, Porto Alegre, Brazil, and followed the “National Institutes of Health Guide for the Care and Use of Laboratory Animals” (NIH Publications No. 85–23, revised 2011).

Conflict of Interest

The authors declare that they have no conflict of interest.

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Legend to figures

Fig. 1 Effect of intracerebral administration of L-2-hydroxyglutaric acid (L-2-HG; 0.750 $\mu\text{mol/g}$) on malondialdehyde (MDA) levels (A), carbonyl formation (B) and sulfhydryl content (C) in cerebral cortex of neonatal rats. Data are represented as mean \pm SD for four to six independent experiments (animals) performed in triplicate and are expressed as nmol / mg protein. * P <0.5, ** P <0.01, compared to control (Student's t test for unpaired samples).

Fig. 2 Effect of intracerebral administration of L-2-hydroxyglutaric acid (L-2-HG; 0.750 $\mu\text{mol/g}$) on concentrations of GSH in cerebral cortex of neonatal rats. Data are represented as mean \pm SD for four to six independent experiments (animals) performed in triplicate and are expressed as nmol / mg protein. *** P <0.001, compared to control (Student's t test for unpaired samples)

Fig. 3 Effect of intracerebral administration of L-2-hydroxyglutaric acid (L-2-HG; 0.750 $\mu\text{mol/g}$) on DCFH oxidation (A) in cerebral cortex and nitrate and nitrite concentrations (B) in forebrain of neonatal rats. Data are represented as mean \pm SD for four to six independent experiments (animals) performed in triplicate and are expressed as μmol / mg protein. * P <0.5, compared to control (Student's t test for unpaired samples)

Fig. 4 Effect of melatonin (MEL; 20 mg/Kg) and MK-801 (0,25mg/Kg) on L-2-hydroxyglutaric acid (L-2-HG 0.750 $\mu\text{mol/g}$)-induced increase of MDA (A) and decrease of GSH concentrations (B) in cerebral cortex of neonatal rats. Data are represented as mean \pm SD for four to six independent experiments (animals) performed in triplicate and are expressed as nmol / mg protein. * P <0.05, compared to control; # P <0.05, ## P <0.01, ### P <0.001, compared to L-2-HG (Student's t test for unpaired samples)

Fig. 5 Effect of intracerebral administration of L-2-hydroxyglutaric acid (L-2-HG; 0.750 $\mu\text{mol/g}$) on the activities of glucose-6-phosphate dehydrogenase (G6PDH) (A), glutathione peroxidase (GPx) (B), glutathione reductase (GR) (C), glutathione S-transferase (GST) (D), catalase (CAT) (E) and superoxide dismutase (SOD) in cerebral cortex of neonatal rats. Data are represented as mean \pm SD for four to six independent experiments (animals) performed in triplicate and are expressed as U / mg protein. * P <0.05, ** P <0.01 compared to control (Student's t test for unpaired samples)

Fig. 6 Effect of intracerebral administration of L-2-hydroxyglutaric acid (L-2-HG; 0.750 $\mu\text{mol/g}$) on sulfhydryl content (A) and the activities of glutathione peroxidase (GPx) (B), glucose-6-phosphate dehydrogenase (G6PDH) (C) and superoxide dismutase (SOD) (D) in striatum of neonatal rats. * P <0.05, ** P <0.01, *** P <0.001, compared to control (Student's t test for unpaired samples)

Fig. 7 Light microscopy images showing hematoxylin and eosin (HE) staining of cerebral cortex from PBS (control, A and C) and L-2-hydroxyglutaric acid (L-2-HG)-injected neonatal rats (B and D). Animals were sacrificed 24 hours after L-2-HG intracerebroventricular administration. Representative images were obtained from four rats per group. HE staining with magnification of 100x and 400x. Arrows in panel D indicate vacuolation.

Fig. 8 Light microscopy images showing hematoxylin and eosin (HE) staining of striatum from PBS (control, A and C) and L-2-hydroxyglutaric acid (L-2-HG)-injected neonatal rats (B and D). Animals

were sacrificed 24 hours after intracerebroventricular administration. Representative images were obtained from four rats per group. HE staining with magnification of 100x and 400x. Arrows in panel D indicate vacuolation.

ANEXO I – FIGURAS

FIGURE 1

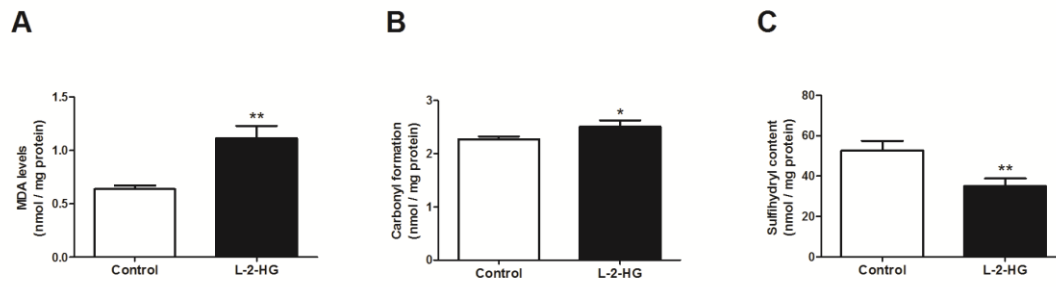


FIGURE 2

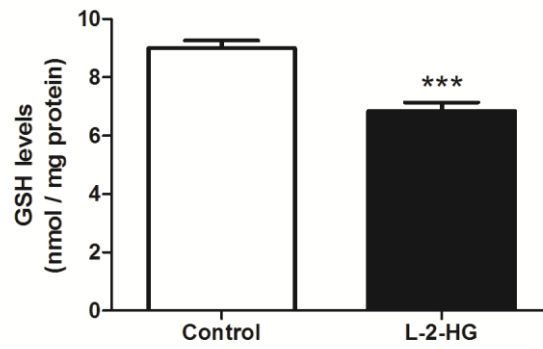


FIGURE 3

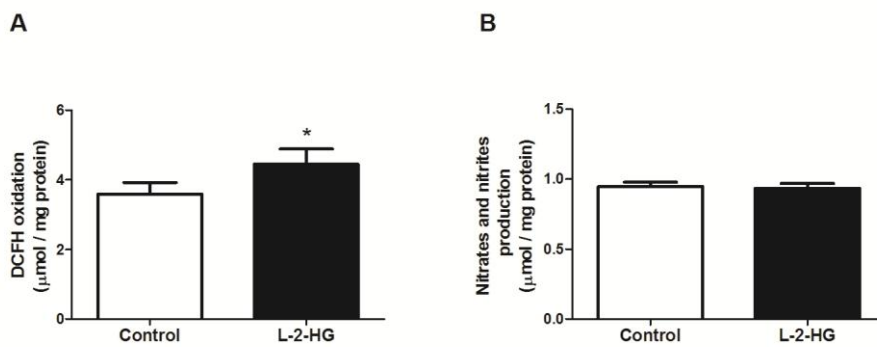


FIGURE 4

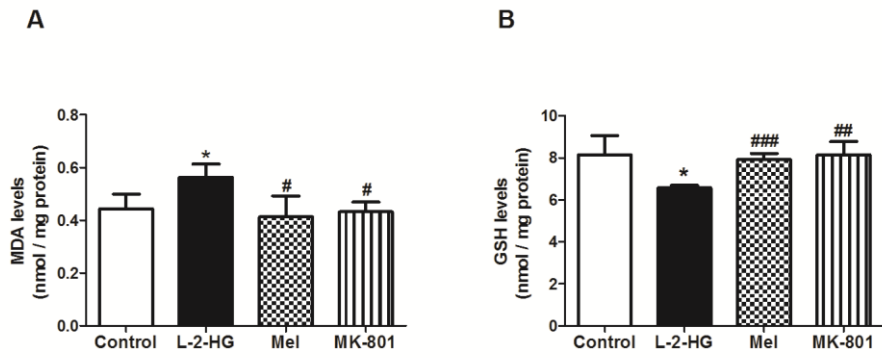


FIGURE 5

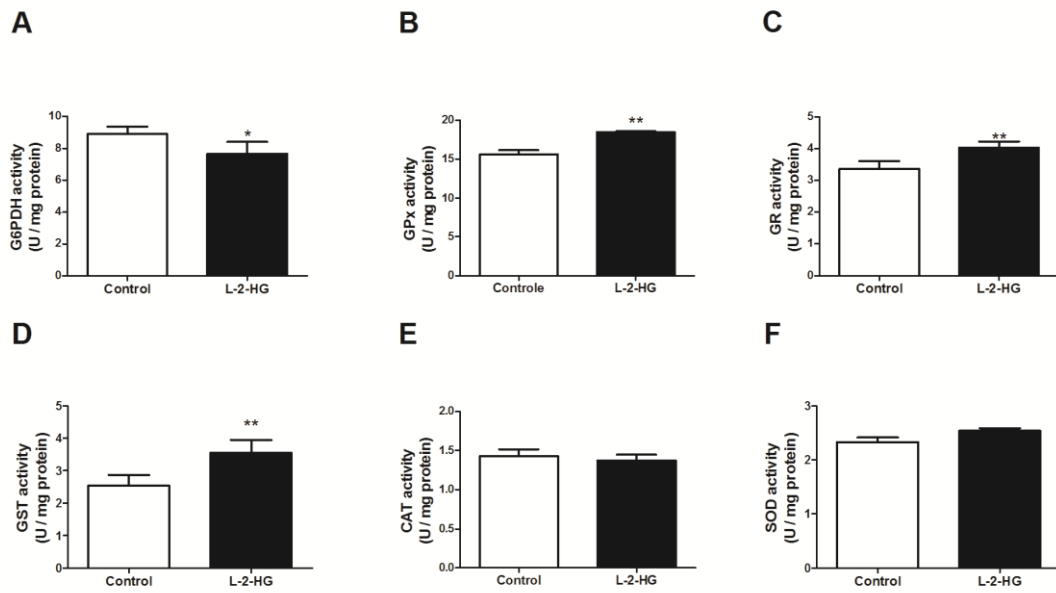


FIGURE 6

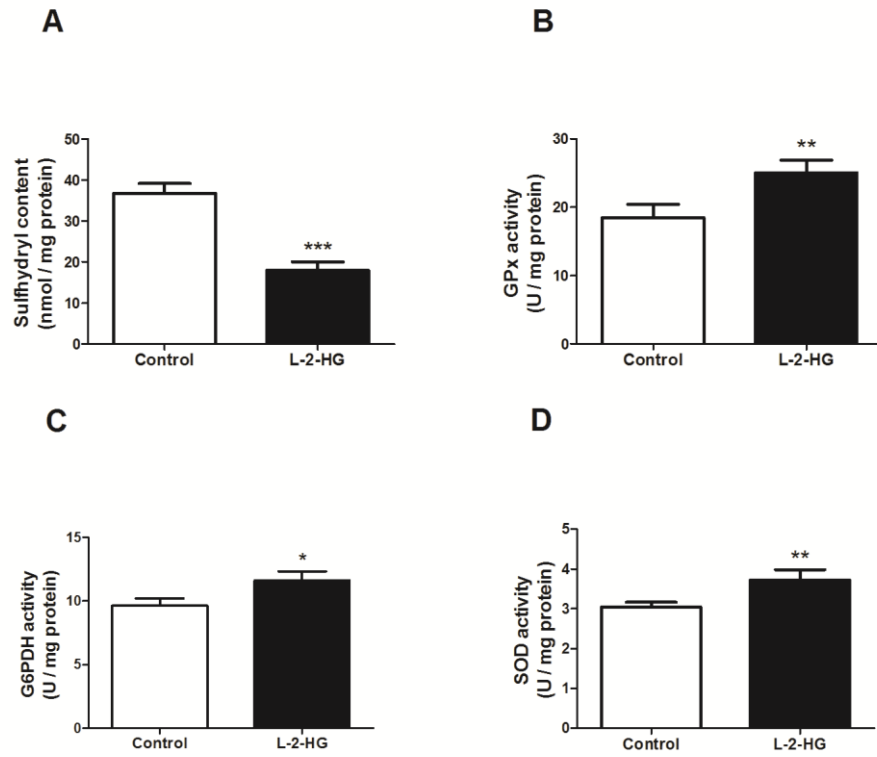


FIGURE 7

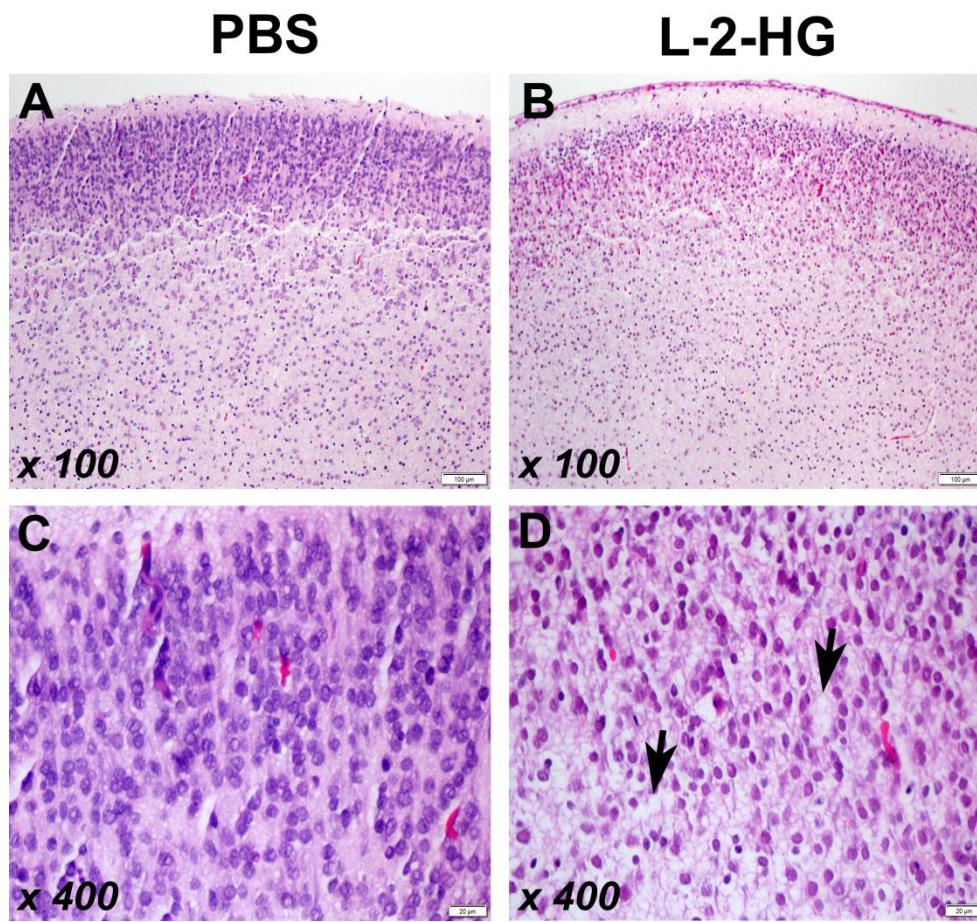
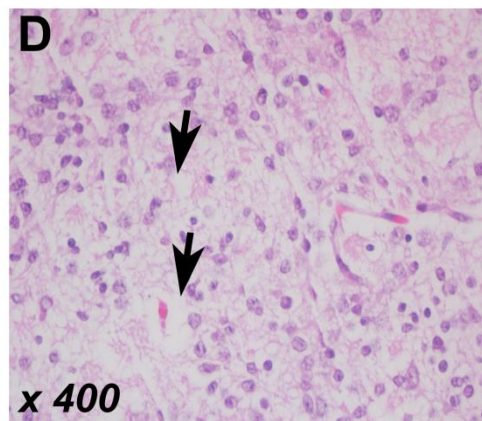
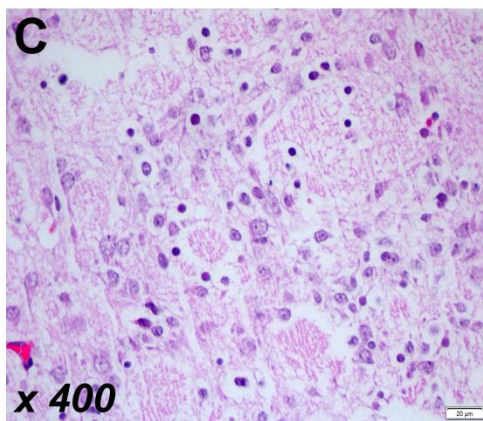
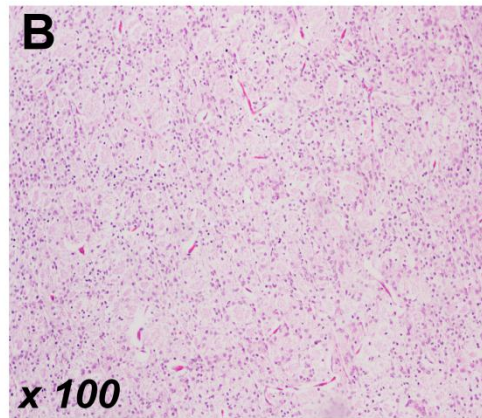
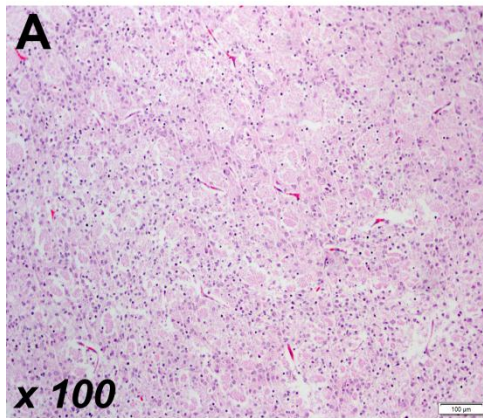


FIGURE 8

PBS

L-2-HG



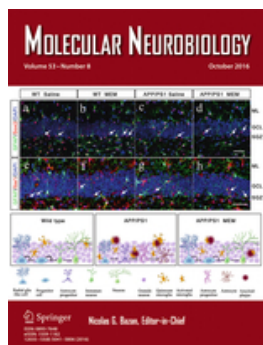
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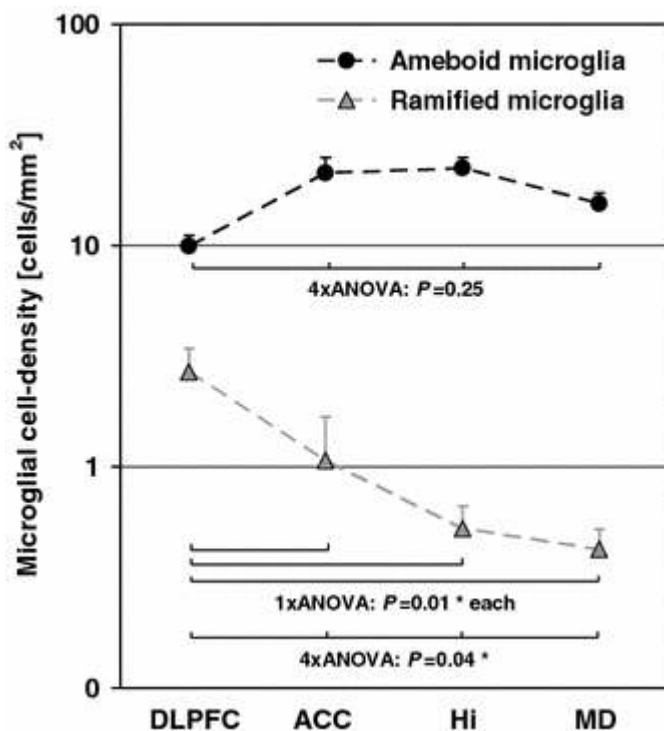
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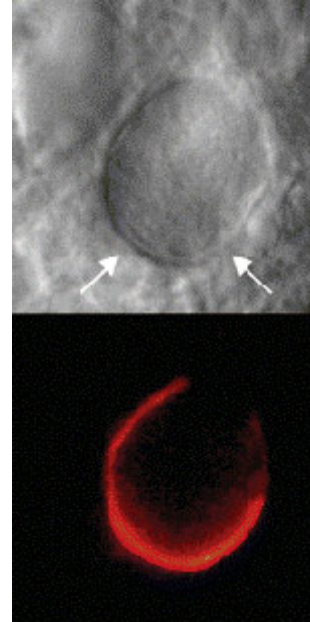
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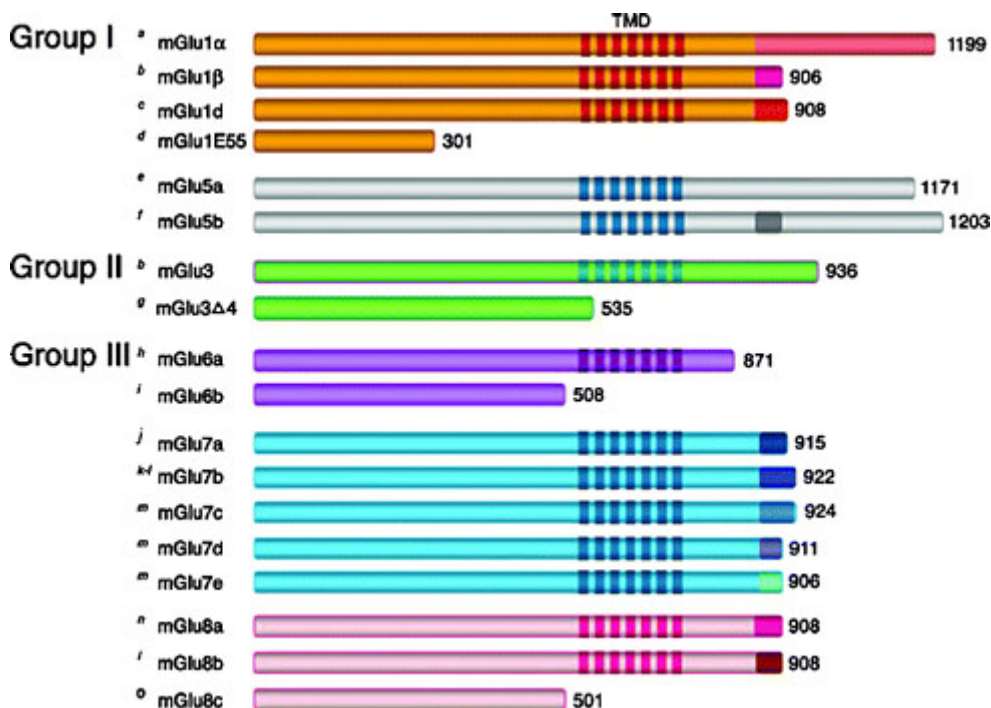
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