

Characterization of Natural and Affected Environments

**EXPLORING THE POTENTIAL OF A GLOBAL EMERGING CONTAMINANT
EARLY WARNING NETWORK THROUGH THE USE OF RETROSPECTIVE
SUSPECT SCREENING WITH HIGH-RESOLUTION MASS SPECTROMETRY**

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2 **THROUGH THE USE OF RETROSPECTIVE SUSPECT SCREENING WITH HIGH-RESOLUTION MASS**
3 **SPECTROMETRY**

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37

38 Abstract

39 A key challenge in the environmental and exposure sciences is to establish experimental evidence of the
40 role of chemical exposure in human and environmental systems. High resolution and accurate tandem
41 mass spectrometry (HRMS) is increasingly being used for the analysis of environmental samples. One
42 lauded benefit of HRMS is the possibility to retrospectively process data for (previously omitted)
43 compounds that has led to the archiving of HRMS data. Archived HRMS data affords the possibility of
44 exploiting historical data to rapidly and effectively establish the temporal and spatial occurrence of
45 newly identified contaminants through retrospective suspect screening. We propose to establish a
46 global emerging contaminant early warning network to rapidly assess the spatial and temporal
47 distribution of contaminants of emerging concern in environmental samples through performing
48 retrospective analysis on HRMS data. The effectiveness of such a network is demonstrated through a
49 pilot study, where eight reference laboratories with available archived HRMS data retrospectively
50 screened data acquired from aqueous environmental samples collected in 14 countries on 3 different
51 continents. The widespread spatial occurrence of several surfactants (e.g. [PEGs](#) and [C12AEO-PEGs](#)),
52 transformation products of selected drugs (e.g. gabapentin-lactam, metoprolol-acid, carbamazepine-10-
53 hydroxy, omeprazole-4-hydroxy-sulphide, 2-benzothiazole-sulfonic-acid), and industrial chemicals (3-
54 nitrobenzenesulfonate and bisphenol-S) was revealed. Obtaining identifications of increased reliability
55 through retrospective suspect screening is challenging and recommendations for dealing with issues
56 such as broad chromatographic peaks, data acquisition, and sensitivity are provided.

57

58 Introduction

59 One of the key challenges in the environmental and exposure sciences is to establish experimental
60 evidence of the role of chemical exposure in human and environmental systems.^{1,2} Our 'chemosphere' is
61 continuously changing and most chemicals that are indexed in the Chemical Abstract Service (CAS) are
62 not characterized with respect to their potential effects on human safety and environmental health.³
63 Non-target analysis employing high-resolution mass spectrometers has been established over the past
64 years as one of the key approaches for tackling this complexity. High resolution and accurate hybrid
65 tandem mass spectrometers, such as time-of-flight and Orbitrap instruments have facilitated increased
66 reliability in target analysis (using reference standards), enabled suspect screening (without reference
67 standards) and screening for unknowns.⁴⁻⁶ Substantial research effort has been placed on developing
68 tools and workflows that expedite these three approaches, with the overall outcome that the
69 contemporary analyst is able to obtain large amount of accurate mass data for a particular sample. For
70 example, in 2013 the NORMAN Network of reference laboratories, research centres and related
71 organisations for monitoring of emerging environmental substances (www.norman-network.net)
72 organized a non-target screening collaborative trial employing target, suspect, and non-target workflows
73 to identify substances in water samples.⁷ This trial revealed that non-target techniques are in general
74 substantially harmonized between practitioners and that although data processing can be time
75 consuming and remains a major bottleneck, suspect screening approaches are very popular. However it
76 recognized that *"better integration and connection of desired features into software packages, the
77 exchange of target and suspect lists, and the contribution of more spectra from standard substances into*

78 *(openly accessible) database*” are necessary for the technique to reach maturity.⁴ The archiving of HRMS
79 data also allows for data to be processed retrospectively, for example to investigate the occurrence of a
80 newly identified compound or simply one that was not considered at the time of analysis.⁸ This
81 possibility has led to researchers working in this field to digitally archive data in preparation for future
82 retrospective analysis and has even led to proposals for the establishment of data repositories, akin to
83 environmental data banks, where digital information can be safely stored for future retrospective
84 analysis.

85 Non-target HRMS full scan data allows the potential for rapid and cost-effective screening of the
86 occurrence of newly identified contaminants in previously archived HRMS data; often referred to as
87 retrospective analysis. Typically, it refers to the application of suspect screening workflows to archived
88 data as reference standard measurements are not available for the analytical settings. Whilst
89 retrospective analysis with HRMS in environmental sciences has been discussed for some time^{7,8,9,10}
90 there are few published studies that actually apply the approach^{11,12}. As far as we are aware there have
91 not been coordinated studies to investigate the spatial and temporal distribution of contaminants of
92 emerging concern in environmental samples through performing retrospective analysis on HRMS data
93 acquired using different instrumental platforms and data processing software. This has the potential to
94 be an improved and effective strategy for establishing the extent of a newly identified contaminant’s
95 occurrence rather than the traditional approach of a new contaminant(s) being reported in the scientific
96 literature and individual research groups subsequently validating targeted methods and reporting their
97 own data. In order to test this hypothesis, a pilot study was performed where eight reference
98 laboratories with available archived HRMS data were recruited with the goal of exploring the potential
99 of a contaminant of emerging concern early warning network through the use of retrospective suspect
100 screening employing HRMS. The pilot study was referred to as the NORMAN Early Warning System,
101 abbreviated to NormaNEWS.¹³

102

103 **Materials and Methods**

104 **Participants and samples**

105 The participants of the NormaNEWS exercise (8 reference laboratories; Eawag, KWR, NIVA, QAEHS,
106 RWS, UJI, UoA, and Vitens) submitted samples from 14 countries and 3 continents. In total 48 sets of
107 data from the analysis of environmental samples were evaluated. Detailed information on sample
108 matrix, sampling date, instrument type, chromatographic separation (flow, column, gradient programs,
109 and solvents), mass spectrometric method (acquisition mode and calibration method) are presented in
110 the “**Sample Information**” sheet in the supporting information (SI) excel spreadsheet. Further, a more
111 detailed description of the samples and methods used are presented in the SI spreadsheet, including
112 information on any previously published datasets.

113 A wide variety of environmental samples were included in this study. The majority of the samples were
114 wastewater (effluent and influent), surface water, and groundwater samples. More than half of the
115 samples (26 out of 48) were wastewater samples (mainly effluent wastewater samples). Wastewater
116 sample data sets were from Switzerland (various locations)¹⁴, Norway, Sweden, Finland, Denmark,

117 Iceland, Spain, Greece, Mexico and Australia. Fifteen of the 48 samples were samples from ecologically
118 important large rivers such as Danube (station JDS57 Bulgarian/Romanian borders)⁷ and Rhine¹⁵,
119 smaller rivers such as Swiss rivers (Furtbach and Doubs)¹⁶, Dutch rivers (Meuse and Vecht) and the Logan
120 river in Australia. Four groundwater samples were included from Spain and the Netherlands. One
121 primary sludge sample from the wastewater treatment plant (WWTP) in Athens (Greece)¹⁷ as well as
122 one seawater sample affected by treated wastewater¹⁸ were also evaluated. Finally, two drinking water
123 samples from Ridderkerk and Lekkerkerk in The Netherlands were included in the study. All the
124 participants were asked to provide only the absolute intensity of the identified features that were blank
125 subtracted in order to avoid the false positive identification.

126 Participating laboratories analyzed their samples using their own routines (i.e. sample preparation and
127 data processing) for all the analytes included in the NormaNEWS suspect list (“**NormaNEWS**
128 **compounds**” sheet in the SI, on the NORMAN [Suspect Exchange](#) and in the CompTox [Chemistry](#)
129 [Dashboard](#)). No specific method (i.e. chromatographic, ion source, and polarity) was recommended to
130 the participants. This was in order to test the applicability of this approach for the data generated via
131 different methods. For these analyses, a wide range of mass analyzers as well as chromatographic
132 conditions was employed by different participants (“**Sample Information**” sheet in the SI). All of the
133 reported results were further examined, through a quality control assessment, to produce harmonized
134 and comparable results (see section ‘Quality control criteria’). Finally, each identified peak was assigned
135 with an appropriate confidence level.¹⁹ These quality assurance steps were deemed necessary for
136 interpretation of the results.

137

138 **NormaNEWS suspect list**

139 The final chemical screening suspect list consisted of 156 analytes including: 74 surfactants i.e. [PEGs](#),
140 [C12AEO-PEGs](#), glycol ether sulfates ([GES](#)), linear alkylbenzyl sulfonates ([LAS](#)), sulfophenyl alkyl carboxylic
141 acids ([SPACs](#)), and fluorosurfactants (PFAS, from several classes); 54 pharmaceuticals and their
142 transformation products (e.g. carbamazepine, carbamazepine-10-hydroxy, diltiazem, diltiazem-
143 desacetyl, and diltiazem-N-desmethyl); 17 bisphenols; and finally 11 industrial chemicals. We considered
144 the surfactants and the industrial chemicals as two separate families of compounds, even though a lot of
145 surfactants may have industrial source. This distinction was made due to multiple sources for
146 surfactants. The suspect list compounds (name, molecular formula, CAS number, SMILES, InChI and
147 InChIKey), qualifier fragment ions and lipophilic properties (logP and log K_{ow}) are included in the SI
148 “**NormaNEWS compounds**” sheet and are available online on the NORMAN [Suspect Exchange](#) and in the
149 CompTox [Chemistry Dashboard](#). The list was formed from compounds suggested by participants and
150 typically included novel emerging substances with limited environmental occurrence as well as
151 established widely occurring environmental contaminants (e.g. carbamazepine), which was included to
152 assess the overall concept. A high number of the proposed substances were transformation products
153 (TPs) of parent drugs that were detected through suspect and non-target screening from bio-
154 transformation experiments. In these cases, parent drugs (e.g. citalopram and atenolol) were also
155 included so that detection rates of the parent drugs and their TPs could be investigated. Novel
156 surfactant compounds were also included to verify their wide-spread occurrence. In addition, the
157 inclusion of a group of bisphenols as well as 3-nitrobenzenesulfonate, specified as an industrial

158 chemical, were a result of non-target screening identifications. The purpose of the NormaNEWS suspect
159 list is to provide a dynamic list of potential environmentally relevant and novel chemicals, which is
160 enriched using expert knowledge and non-target analysis results as new data become available. The list
161 is available at the NORMAN Suspect List Exchange (<http://www.norman-network.com/?q=node/236>)
162 and on the CompTox Chemistry Dashboard
163 (https://comptox.epa.gov/dashboard/chemical_lists/normanews).

164 **Quality control criteria**

165 All participants of NormaNEWS exercise were requested to submit their results together with their raw
166 LC-HRMS chromatograms. Raw chromatograms were converted to mzML using ProteoWizard
167 (msconvert module v.3.0.10827).²⁰ For data acquired in data-independent acquisition mode, different
168 collision energy channels were separated using an in-house script (provided in the SI), while lock mass
169 scans were removed. For data-dependent acquisition mode, MS/MS spectra were exported as text files
170 (named “precursor mass retention time”) and were removed from the mzML files. Treated mzML files
171 were converted to CDF files, which are readable from various data analysis software including Bruker
172 DataAnalysis v.4.3. (Bruker Daltonics, Bremen, Germany), which was used here.

173 The performance of the following parameters was checked; mass accuracy of HRMS, stability of
174 chromatography and presence of qualifier fragments of identified compounds in higher collision energy.
175 A combination of an expert panel and literature information was used in order to set the threshold of
176 each quality control criterion.

177 The quality control step enabled us to minimize the effect of analyst expertise and the instrumentation
178 on the final results given that the evaluation of the analysts and/or the instrumentation was not within
179 the goals of this exercise. Therefore, the data points that did not meet the quality control criteria were
180 excluded from the finally reported results.

181 **RESULTS AND DISCUSSION**

182 **Quality control assessment**

183 Quality control was performed to ensure that data were generated from well-calibrated instruments
184 and that the data submitted were reliable. The first and most important step of the procedure was to
185 check that the mass difference between the experimental and theoretical mass did not exceed ± 5 mDa,
186 which was considered the maximum tolerable mass error in the provided complex environmental
187 samples.^{21, 22} This was highly relevant in assessing the confidence level assigned to each identified
188 analyte in the list.

189 The mass accuracy quality control is summarized in the SI “**QC_mass accuracy_ppm/ QC_mass**
190 **accuracy_Da**” sheet and the results presented in Figure 1. According to the submitted datasets, Orbitrap
191 mass analyzers showed better mass accuracy performance (absolute average mass error 0.55 mDa)
192 comparing to other TOF instruments (absolute average mass error 0.91 mDa), based on successfully
193 identified compounds. Mass errors are caused by the complexity of the samples, saturation of the
194 detector (see section challenges and recommendations), and the instrument itself (i.e. the age and
195 hardware). LC-HRMS data obtained using LTQ Orbitrap instruments showed lower mass accuracy
196 (absolute average mass error 1.1 mDa) when compared with the LTQ Orbitrap XL (absolute average

197 mass error 0.52 mDa), which showed lower mass accuracy in comparison with the QExactive. We further
198 investigated the effect of instrumentation used on the observed mass accuracies through a non-
199 parametric statistical test Kruskal-Wallis. ²³A Kruskal-Wallis p value > 0.01 indicated the rejection of null-
200 hypothesis and statistical significance of the observed differences in the measured averaged masses.
201 The method used to calibrate each instrument was also considered. LC-HRMS data obtained using a
202 Bruker QTOF were calibrated by injecting the calibrant substance at the beginning of the chromatogram,
203 while data from Waters QTOF (in both cases) were calibrated by lock-mass every 0.5 or 2 minutes
204 (injecting, recording and recalibrating based on calibrant peaks appearing every 0.5/2 minutes). High
205 mass accuracy is an extremely crucial parameter to achieve high quality results during the suspect
206 analysis. Especially, high accuracy measurements enable a decreased number of false positive
207 detections.

208 The chromatographic stability of the LC separation was also assessed. All participants submitted at least
209 3 datasets for evaluation. Retention time data from the same instrumental set-up (and same partner)
210 were grouped together and the normalized standard deviations (NSD) of the retention times of the
211 detected substances were calculated (retention times of the detected substances in seconds can be
212 found in the SI “**QC_observed_ret.time_Minutes**” sheet). A criterion of the maximum tolerable NSD of
213 10% was adopted for accepting the detection of a single compound across samples in data coming from
214 the same partner. The average normalized standard deviation of retention times in all samples was $< 2\%$
215 (Figure S1). The largest variability of 8.6 % was observed for analyte valsartan, whereas the lowest
216 variability ($< 0.1\%$) was observed for acesulfame in samples from Netherlands, GES-07 in samples from
217 Australia, and GES-09 and GES-06 in samples from Greece. Retention time stability was considered as
218 another extremely important parameter, which has a direct effect on the identification confidence. The
219 low deviation observed in all the submitted datasets indicated the high quality and reliability of the LC
220 separation of the participating laboratories.

221 The third QC criterion related to the presence of qualifier ions (QI) in the MS/MS spectra (SI
222 “**NormaNEWS compounds**” sheet). These ions are fragments of the parent ion and are observable at
223 higher collision energy or even at low collision energy as in-source fragments. The criterion was set on
224 the presence of the QIs as either an in-source fragment or at higher collision energy. The identification
225 level of compounds that did not comply with the third QC criterion were regarded as questionable and
226 were marked accordingly.¹⁹ As these QIs proved to be a very efficient way of improving the confidence
227 of the suspect hit, Top 3 fragments have now been extracted from all mass spectra submitted to
228 [MassBank.EU](#) and also put on the [NORMAN Suspect Exchange \(direct download\)](#) and the [CompTox
229 Chemistry Dashboard Downloads \(direct link\)](#) for community use. The QC stage was used to exclude the
230 features that did not meet the previously set criteria, thus harmonization. Consequently, we have
231 reported only the features that met these mentioned criteria.

232 **Overview of the retrospective screening**

233 PolyEthylene Glycol 09 (PEG-09) was the most frequently detected compound, being present in 41 out
234 of the 48 samples (85%) analyzed. Several bisphenols, transformation products of perfluorooctane
235 sulfonate, and the pharmaceutical omeprazole were not detected in any of the samples analyzed (“**Max.
236 Absolute Intensity_counts**” sheet in the SI and Figures 2, XS, X1S, X2S). Series of surfactants, such as
237 [PEGs](#), [C12AEO-PEGs](#), and [GES](#), resulted in a higher detection frequency for compounds with masses

238 varying between 400 and 600 Da compared to both smaller and larger molecules from the same families
239 (Figure S2.A). Schymanski et al and Gago-Ferrero et al. have previously observed a similar trend for
240 these surfactants.^{14, 24} The observed trend may be explained by the efficient ionization of mid-size
241 molecules compared to other compounds and potentially the fact that they are used as technical
242 mixtures.²⁵ LAS had an average frequency of detection of around 50%. The largest measured LAS, in
243 terms of mass (i.e. C14-LAS), were detected in only 4 samples out of 48 samples. Based on the estimated
244 retention time for LAS-C14, we interpret that the chromatographic run times used by different partners
245 were not sufficiently long to successfully detect this suspect analyte in the evaluated samples. Only 3 of
246 the 5 suspect fluorinated surfactants were detected with perfluorooctane sulfonate (PFOS) having the
247 highest detection frequency of ~ 35%. For industrial chemicals and pharmaceuticals, venlafaxine was the
248 suspect analyte with the highest frequency of detection (68%), while several bisphenols were not
249 detected in any of the samples. Additionally, we observed a higher occurrence frequency of the suspect
250 analytes in the locations with higher population density such as Spain, Switzerland, and Greece
251 compared to locations such as Scandinavia and Australia with lower population density, Figures 2 and
252 S3. The observed trend was consistent across all the analyzed matrices. However, it should be noted
253 that considering the limited data set for this pilot study, further interpretation of the spatial and
254 temporal distribution of pollutants is not possible. The future implementation of this approach will
255 provide larger datasets for comprehensive spatial and temporal assessment of CEC occurrence across
256 the globe.

257 The presence of a large number of successfully detected surfactants and industrial chemicals in both
258 wastewater influents, effluents, and surface waters suggests the wide spread occurrence of these CECs
259 in the environment across the globe, Figure 2. Although modern wastewater treatment plants are to
260 some extent equipped to remove these pollutants²⁶⁻²⁹, the high production/consumption volumes of
261 these chemicals used in households and industrial applications translates into their release into the
262 environment. The environmental occurrence, fate and behavior of surfactants have been widely
263 investigated, however more reliable environmental data for these pollutants are necessary.³⁰⁻³²
264 Collective exercises such as NormaNEWS are therefore an important step forward towards producing a
265 comprehensive and reliable database on the environmental occurrence of surfactants and/or other
266 chemicals of emerging concern (CEC), which can be used for better understanding of their
267 environmental fate and behavior. Furthermore, this exercise, through the provided QC criteria,
268 metadata template (i.e. SI spreadsheet), provides all necessary information and guidelines for
269 laboratories across the globe for the reliable detection, identification, and reporting of CECs in different
270 environmental compartments.

271 **Challenges and recommendations**

272 For analysts to obtain high-confidence identifications through retrospective suspect screening they face
273 several challenges. Here, recommendations for dealing with difficulties such as broad peaks, data
274 acquisition, and sensitivity are provided in the following.

275 The presence of broad peaks in the chromatograms of complex samples is often caused by the physico-
276 chemical properties of that compound and the selected chromatographic method is unavoidable. For
277 example, the LAS surfactants that elute at the end of the gradient of a typical reverse phase
278 chromatographic run result in characteristic broad peaks (Figure 3A). Many peak picking algorithms are

279 unable to detect such broad peaks. Therefore, employing peak picking independent approaches^{33, 34},
280 prior knowledge of those analytes, and visualization tools, even though not comprehensive, may be
281 useful in dealing with broad peaks.

282 Data-dependent acquisition is often used in non-target analysis. Certain limitations with data-dependent
283 acquisition may potentially cause false identification of features due to its limitations. This acquisition
284 mode isolates and provides MS/MS spectra of some of the most abundant ions per full scan. Even
285 though this approach is the ideal acquisition mode during identification of peaks with the most
286 abundant ions, this mode is not suitable for retrospective screening, due to the limited number of
287 MS/MS spectra obtained. In case the peak of an environmentally relevant compound is not one of those
288 most abundant ions, the MS/MS spectra of this chemical would not be recorded (Figure 3B). Therefore,
289 confident identification of that peak would not be possible. As a solution, it is highly recommended that
290 samples are injected in data-independent acquisition mode which is the ideal acquisition mode for
291 retrospective screening. In data-independent acquisition, HRMS is recording full scan and MS/MS
292 spectra without prior isolation of any mass. Therefore, all fragments (and fragments of fragments in case
293 of in-source fragments) of all co-eluting compounds are recorded, resulting in complex but information-
294 rich MS/MS spectra that requires adequate data processing tools for confident identification of features.
295 However, to our knowledge this is the most effective acquisition method for the samples that are meant
296 for retrospective analysis. As different compounds have different fragmentation behavior depending on
297 the different collision energies, the use of multiple (e.g. low, medium, high) or ramped collision energies
298 should be considered during acquisition of data for retrospective screening to cover as many
299 compounds as possible. As different instruments have different settings and acquisition speeds, a
300 compromise may need to be found to provide sufficient resolution in the full scan while obtaining as
301 much fragmentation information as possible. Pilot studies such as these and the upload of
302 corresponding suspect lists and fragment information to public resources greatly help exchange
303 experience to find these ideal compromises for future investigations.

304 Another inherent concern about LC-HRMS data is sensitivity. Among other reasons, one possible case for
305 non-detection of pollutants is that current HRMS instruments operated in full scan are sensitive
306 depending on the frequency with which they acquire full scans.³⁵ This means that low abundant or
307 poorly ionized chemicals are not detected in case HRMS instrument records full scans at a high
308 frequency rate. For example, recording full-scans at low frequency (2 Hz) will enable the detection of
309 more compounds in comparison with a higher frequency rate (i.e. 20 Hz). Therefore, the analysts should
310 try to find a compromise between the sampling speed and the sensitivity required for the analyses. For
311 the samples, that are meant to be analyzed via retrospective screening a lower sampling frequency is
312 recommended given that under these conditions a higher sensitivity is achieved.

313 Substances at high concentration levels in extracts and/or having high ionization efficiency can often
314 result in the detector becoming saturated (Figure 3C). In this case, the peak reaches a plateau, which
315 makes peak picking and determination of exact mass and retention time very difficult. For example,
316 surfactants such as [PEGs](#) and [C12AEO-PEGs](#) were affected by detector saturation due to their high
317 concentrations in the evaluated samples. The mentioned uncertainties in the exact mass and retention
318 time are caused by the fact that saturation reduces the mass accuracy of the measurements for certain
319 instruments, which is of extreme importance when performing identification. However, increasing the

320 mass extraction window may solve these issues. On the other hand, such less strict mass accuracy
321 criterion may increase the likelihood of false positive detection.

322 Another open issue in mass spectrometry is related to structural isomers (Figure 3D). Isomers are
323 structurally similar compounds with the same molecular formula (same mass and isotopic profile) and
324 share very similar fragmentation. This happened in the case of the detection of bisphenol S in the
325 surface waters of the Netherlands. Two peaks, with different retention times, with acceptable mass
326 accuracy, isotopic fit and same qualifier ions seem to belong to two different isomers of bisphenol S. In
327 such cases, deeper knowledge of fragmentation behavior and/or retention time prediction could help to
328 identify the peak that belongs to the suspected substance. Ion ratio (ratio of the intensity of a fragment
329 to the intensity of another fragment) can be also considered. However, this information should be
330 carefully examined, because of ion suppression caused by high background signal produced by complex
331 sample's matrix. Classes of substances such as the surfactants mentioned here also contain many
332 structurally related substances that cannot be distinguished easily with mass spectrometry. These are
333 now being grouped as "related substances" in the CompTox Chemistry Dashboard (see hyperlinks for
334 the different surfactant classes throughout this manuscript) as a first step in working towards
335 computational solutions to deal with the extremely complex challenge of chemical substances of
336 Unknown or Variable Composition, Complex Reaction Products and Biological Materials (UVCBs).^{36, 37}
337 Finally, all the samples need to be analyzed both in positive and negative mode in order to cover a wider
338 chemical space compared to only single polarity.

339 **The early warning system and its potential**

340 This exercise confirmed the high occurrence frequency of several surfactants (e.g. [PEGs](#) and [C12AEO-](#)
341 [PEGs](#)), transformation products of selected drugs (e.g. gabapentin-lactam, metoprolol-acid,
342 carbamazepine-10-hydroxy, omeprazole-4-hydroxy-sulphide, 2-benzothiazole-sulfonic-acid), and
343 industrial chemicals such as 3-nitrobenzenesulfonate and bisphenol S. These chemicals are not typically
344 included in target/suspect lists used for surface water monitoring programs. Subsequently, there are
345 limited environmental occurrence data available for these pollutants.³⁸⁻⁴⁰ This clearly demonstrates that
346 an early warning network such as NormaNEWS enables the efficient and reliable detection and
347 identification of novel CECs in different environmental compartments at both a temporal and spatial
348 scale. Consequently, a reasonably large and diverse dataset on the environmental occurrence of novel
349 CECs in different matrices has been generated during this pilot project. Clearly, this study was a proof of
350 concept to test the applicability of such an approach to a diverse global dataset. Further development
351 and larger global coverage is necessary in order to generate a dataset suitable for both environmental
352 interpretation and policy making practices. Such a dataset provides an initial screen that can be used to
353 inform contaminant prioritization exercises leading to further monitoring, fate and effect studies and
354 subsequent risk assessment. Furthermore, given that the data are harmonized across a large number of
355 laboratories and the confidence level of each identification is provided, the inherent reliability of each
356 identification becomes more intuitive to non-experts. The purpose of this network activity would not be
357 to replace ongoing targeted monitoring and screening programs, but to provide a robust and
358 comprehensive complementary collaborative approach for informing the refinement of priority
359 substance lists. This also shows the great potential for screening much larger lists in the future, although
360 the manual verification of the results is still a demanding task. More computationally efficient methods
361 will be needed before this can be expanded to potentially lists of tens of thousands of substances.

362 The NormaNEWS pilot was performed using a very simple approach where all participants manually
363 submitted data on their CECs of interest in order to create a suspect screening list for the collaborative
364 exercise. This enabled researchers to easily obtain additional data on the CECs that they are particularly
365 interested in. Future lists could be generated by a number of different approaches including from open
366 resources, such as massbank.eu. As highlighted recently by Schymanski and Williams,³⁶ open resources
367 will be instrumental in defining the evolution of suspect screening. The community-wide sharing of CECs
368 through the exchange of suspect lists (e.g. the [NORMAN Suspect Exchange](#) and the [Chemistry
369 Dashboard lists](#)) as well as tentatively and unequivocally identified spectra and sharing the related
370 fragments is therefore key to the success of a global early warning network. Also key will be the
371 willingness of the scientific community to share their HRMS data in an open MS format (e.g. mzML⁴¹,
372 mzXML⁴², and netCDF⁴³). The Global Natural Products Social Molecular Networking (GNPS;
373 <http://gnps.ucsd.edu/>) provides a vision as to how global collaboration and social cooperation can be
374 used to address major scientific challenges in the sharing and community curation of MS data.⁴⁴ Taking
375 inspiration from GNPS, we propose that HRMS data are made available (through a virtual repository and
376 with necessary metadata) in order to facilitate living data along with periodic automated re-analysis of
377 data (i.e. with updates to the suspect list or the addition of new data sets). Ideally, this repository will be
378 easily accessible through a web-application and free of the aforementioned challenges. The
379 environmental and exposure sciences currently lag behind other fields, such as proteomics⁴⁵,
380 metabolomics⁴⁶ and natural product research⁴⁷ in globally collaborating and sharing data through
381 open/social platforms in order to revolutionize the way data are processed to achieve significant
382 outcomes. We acknowledge that not all the data tools are currently in place to make our proposal a
383 reality, however progress is being made in this area^{33, 34, 48, 49}. For example, within the NORMAN Network
384 (<http://www.norman-network.net/>) there is an initiative to develop a digital sample freezing platform. A
385 global emerging contaminant early warning network based on adopting the successful practices of other
386 similar networks will play a pivotal role in identifying chemicals using HRMS that has the potential to
387 possess significant outcomes in protecting human and environmental health.

388 SUPPORTING INFORMATION

389 Text, tables and figures with detailed information on experimental methods, QA/QC procedures
390 and supplemental data (xls, PDF).

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402 REFERENCES

- 403 1. Kortenkamp, A.; Faust, M.; Scholze, M.; Backhaus, T., Low-level exposure to multiple chemicals:
404 reason for human health concerns? *Environ Health Perspect* **2007**, *115 Suppl 1*, 106-114.
- 405 2. Pleil, J. D., Categorizing Biomarkers of the Human Exposome and Developing Metrics for
406 Assessing Environmental Sustainability. *Journal of Toxicology and Environmental Health, Part B* **2012**, *15*,
407 (4), 264-280.
- 408 3. Muir, D. C. G.; Howard, P. H., Are There Other Persistent Organic Pollutants? A Challenge for
409 Environmental Chemists. *Environmental Science & Technology* **2006**, *40*, (23), 7157-7166.
- 410 4. Rager, J. E.; Strynar, M. J.; Liang, S.; McMahan, R. L.; Richard, A. M.; Grulke, C. M.; Wambaugh, J.
411 F.; Isaacs, K. K.; Judson, R.; Williams, A. J.; Sobus, J. R., Linking high resolution mass spectrometry data
412 with exposure and toxicity forecasts to advance high-throughput environmental monitoring. *Environ Int*
413 **2016**, *88*, 269-280.
- 414 5. Andra, S. S.; Austin, C.; Patel, D.; Dolios, G.; Awawda, M.; Arora, M., Trends in the application of
415 high-resolution mass spectrometry for human biomonitoring: An analytical primer to studying the
416 environmental chemical space of the human exposome. *Environ Int* **2017**, *100*, 32-61.
- 417 6. Leendert, V.; Van Langenhove, H.; Demeestere, K., Trends in liquid chromatography coupled to
418 high-resolution mass spectrometry for multi-residue analysis of organic micropollutants in aquatic
419 environments. *TrAC Trends in Analytical Chemistry* **2015**, *67*, 192-208.
- 420 7. Schymanski, E. L.; Singer, H. P.; Slobodnik, J.; Ipolyi, I. M.; Oswald, P.; Krauss, M.; Schulze, T.;
421 Haglund, P.; Letzel, T.; Grosse, S.; Thomaidis, N. S.; Bletsou, A.; Zwiener, C.; Ibanez, M.; Portoles, T.; de
422 Boer, R.; Reid, M. J.; Onghena, M.; Kunkel, U.; Schulz, W.; Guillon, A.; Noyon, N.; Leroy, G.; Bados, P.;
423 Bogialli, S.; Stipanicev, D.; Rostkowski, P.; Hollender, J., Non-target screening with high-resolution mass
424 spectrometry: critical review using a collaborative trial on water analysis. *Anal Bioanal Chem* **2015**, *407*,
425 (21), 6237-55.
- 426 8. Krauss, M.; Singer, H.; Hollender, J., LC-high resolution MS in environmental analysis: from
427 target screening to the identification of unknowns. *Anal Bioanal Chem* **2010**, *397*, (3), 943-51.
- 428 9. Hernandez, F.; Sancho, J. V.; Ibanez, M.; Abad, E.; Portoles, T.; Mattioli, L., Current use of high-
429 resolution mass spectrometry in the environmental sciences. *Anal Bioanal Chem* **2012**, *403*, (5), 1251-
430 64.
- 431 10. Gomez-Ramos, M. M.; Ferrer, C.; Malato, O.; Aguera, A.; Fernandez-Alba, A. R., Liquid
432 chromatography-high-resolution mass spectrometry for pesticide residue analysis in fruit and
433 vegetables: screening and quantitative studies. *J Chromatogr A* **2013**, *1287*, 24-37.
- 434 11. Polgar, L.; Garcia-Reyes, J. F.; Fodor, P.; Gyepes, A.; Dernovics, M.; Abranko, L.; Gilbert-Lopez, B.;
435 Molina-Diaz, A., Retrospective screening of relevant pesticide metabolites in food using liquid
436 chromatography high resolution mass spectrometry and accurate-mass databases of parent molecules
437 and diagnostic fragment ions. *J Chromatogr A* **2012**, *1249*, 83-91.
- 438 12. Chiaia-Hernandez, A. C.; Krauss, M.; Hollender, J., Screening of lake sediments for emerging
439 contaminants by liquid chromatography atmospheric pressure photoionization and electrospray
440 ionization coupled to high resolution mass spectrometry. *Environ Sci Technol* **2013**, *47*, (2), 976-86.
- 441 13. Gomez-Ramos Mdel, M.; Perez-Parada, A.; Garcia-Reyes, J. F.; Fernandez-Alba, A. R.; Aguera, A.,
442 Use of an accurate-mass database for the systematic identification of transformation products of
443 organic contaminants in wastewater effluents. *Journal of chromatography. A* **2011**, *1218*, (44), 8002-12.

- 444 14. Schymanski, E. L.; Singer, H. P.; Longree, P.; Loos, M.; Ruff, M.; Stravs, M. A.; Ripolles Vidal, C.;
445 Hollender, J., Strategies to characterize polar organic contamination in wastewater: exploring the
446 capability of high resolution mass spectrometry. *Environ Sci Technol* **2014**, *48*, (3), 1811-8.
- 447 15. Ruff, M.; Mueller, M. S.; Loos, M.; Singer, H. P., Quantitative target and systematic non-target
448 analysis of polar organic micro-pollutants along the river Rhine using high-resolution mass-
449 spectrometry--Identification of unknown sources and compounds. *Water Res* **2015**, *87*, 145-54.
- 450 16. Moschet, C.; Wittmer, I.; Simovic, J.; Junghans, M.; Piazzoli, A.; Singer, H.; Stamm, C.; Leu, C.;
451 Hollender, J., How a complete pesticide screening changes the assessment of surface water quality.
452 *Environ Sci Technol* **2014**, *48*, (10), 5423-32.
- 453 17. Gago-Ferrero, P.; Borova, V.; Dasenaki, M. E.; Tauhomaidis Nu, S., Simultaneous determination
454 of 148 pharmaceuticals and illicit drugs in sewage sludge based on ultrasound-assisted extraction and
455 liquid chromatography-tandem mass spectrometry. *Anal Bioanal Chem* **2015**, *407*, (15), 4287-97.
- 456 18. Alygizakis, N. A.; Gago-Ferrero, P.; Borova, V. L.; Pavlidou, A.; Hatzianestis, I.; Thomaidis, N. S.,
457 Occurrence and spatial distribution of 158 pharmaceuticals, drugs of abuse and related metabolites in
458 offshore seawater. *Sci Total Environ* **2016**, *541*, 1097-105.
- 459 19. Schymanski, E. L.; Jeon, J.; Gulde, R.; Fenner, K.; Ruff, M.; Singer, H. P.; Hollender, J., Identifying
460 small molecules via high resolution mass spectrometry: communicating confidence. *Environ Sci Technol*
461 **2014**, *48*, (4), 2097-8.
- 462 20. Chambers, M. C.; Maclean, B.; Burke, R.; Amodei, D.; Ruderman, D. L.; Neumann, S.; Gatto, L.;
463 Fischer, B.; Pratt, B.; Egertson, J.; Hoff, K.; Kessner, D.; Tasman, N.; Shulman, N.; Frewen, B.; Baker, T. A.;
464 Brusniak, M. Y.; Paulse, C.; Creasy, D.; Flashner, L.; Kani, K.; Moulding, C.; Seymour, S. L.; Nuwaysir, L. M.;
465 Lefebvre, B.; Kuhlmann, F.; Roark, J.; Rainer, P.; Detlev, S.; Hemenway, T.; Huhmer, A.; Langridge, J.;
466 Connolly, B.; Chadick, T.; Holly, K.; Eckels, J.; Deutsch, E. W.; Moritz, R. L.; Katz, J. E.; Agus, D. B.;
467 MacCoss, M.; Tabb, D. L.; Mallick, P., A cross-platform toolkit for mass spectrometry and proteomics.
468 *Nat Biotechnol* **2012**, *30*, (10), 918-20.
- 469 21. Zedda, M.; Zwiener, C., Is nontarget screening of emerging contaminants by LC-HRMS
470 successful? A plea for compound libraries and computer tools. *Anal Bioanal Chem* **2012**, *403*, (9), 2493-
471 502.
- 472 22. Kaufmann, A.; Walker, S., Evaluation of the interrelationship between mass resolving power and
473 mass error tolerances for targeted bioanalysis using liquid chromatography coupled to high-resolution
474 mass spectrometry. *Rapid Commun Mass Spectrom* **2013**, *27*, (2), 347-56.
- 475 23. Breslow, N., A generalized Kruskal-Wallis test for comparing K samples subject to unequal
476 patterns of censorship. *Biometrika* **1970**, *57*, (3), 579-594.
- 477 24. Gago-Ferrero, P.; Schymanski, E. L.; Bletsou, A. A.; Aalizadeh, R.; Hollender, J.; Thomaidis, N. S.,
478 Extended Suspect and Non-Target Strategies to Characterize Emerging Polar Organic Contaminants in
479 Raw Wastewater with LC-HRMS/MS. *Environ Sci Technol* **2015**, *49*, (20), 12333-41.
- 480 25. Mazzoni, M.; Rusconi, M.; Valsecchi, S.; Martins, C. P.; Polesello, S., An on-line solid phase
481 extraction-liquid chromatography-tandem mass spectrometry method for the determination of
482 perfluoroalkyl acids in drinking and surface waters. *J Anal Methods Chem* **2015**, *2015*, 942016.
- 483 26. Prats, D.; Ruiz, F.; Vazquez, B.; M., R.-P., Removal of anionic and nonionic surfactants in a
484 wastewater treatment plant with anaerobic digestion. A comparative study. *Water Res* **1997**, *31*, (8),
485 1925-1930.
- 486 27. Aboulhassan, M. A.; Souabi, S.; Yaacoubi, A.; Baudu, M., Removal of surfactant from industrial
487 wastewaters by coagulation flocculation process. *Int J Environ Sci Tech* **2006**, *3*, (4), 327-332.
- 488 28. Gonzalez, S.; Petrovic, M.; Barcelo, D., Removal of a broad range of surfactants from municipal
489 wastewater--comparison between membrane bioreactor and conventional activated sludge treatment.
490 *Chemosphere* **2007**, *67*, (2), 335-43.

- 491 29. Luo, Y.; Guo, W.; Ngo, H. H.; Nghiem, L. D.; Hai, F. I.; Zhang, J.; Liang, S.; Wang, X. C., A review on
492 the occurrence of micropollutants in the aquatic environment and their fate and removal during
493 wastewater treatment. *Sci Total Environ* **2014**, *473-474*, 619-41.
- 494 30. Jackson, M.; Eadsforth, C.; Schowanek, D.; Delfosse, T.; Riddle, A.; Budgen, N., Comprehensive
495 review of several surfactants in marine environments: Fate and ecotoxicity. *Environ Toxicol Chem* **2016**,
496 *35*, (5), 1077-86.
- 497 31. Jarda, K.; Drogui, P.; Daghrir, R., Surfactants in aquatic and terrestrial environment: occurrence,
498 behavior, and treatment processes. *Environ Sci Pollut Res Int* **2016**, *23*, (4), 3195-216.
- 499 32. Ying, G.-G., Fate, behavior and effects of surfactants and their degradation products in the
500 environment. *Environment International* **2006**, *32*, (3), 417-431.
- 501 33. Samanipour, S.; Langford, K.; Reid, M. J.; Thomas, K. V., A two stage algorithm for target and
502 suspect analysis of produced water via gas chromatography coupled with high resolution time of flight
503 mass spectrometry. *J Chromatogr A* **2016**, *1463*, 153-61.
- 504 34. Samanipour, S.; Baz-Lomba, J. A.; Alygizakis, N. A.; Reid, M. J.; Thomaidis, N. S.; Thomas, K. V.,
505 Two stage algorithm vs commonly used approaches for the suspect screening of complex environmental
506 samples analyzed via liquid chromatography high resolution time of flight mass spectroscopy: A test
507 study. *J Chromatogr A* **2017**, *1501*, 68-78.
- 508 35. Acena, J.; Stampachiachiere, S.; Perez, S.; Barcelo, D., Advances in liquid chromatography-high-
509 resolution mass spectrometry for quantitative and qualitative environmental analysis. *Anal Bioanal*
510 *Chem* **2015**, *407*, (21), 6289-99.
- 511 36. Schymanski, E. L.; Williams, A. J., Open Science for Identifying "Known Unknown" Chemicals.
512 *Environ Sci Technol* **2017**, *51*, (10), 5357-5359.
- 513 37. Williams A.; Grulke, C. M.; McEachran A.; Richard, A.; Jolley R.; Dunne J.; Edmiston E.; J, E. Comptox
514 Chemistry Dashboard: Web-based data integration hub for environmental chemistry and toxicology
515 data.
516 [https://www.slideshare.net/AntonyWilliams?utm_campaign=profiletracking&utm_medium=sssite&utm](https://www.slideshare.net/AntonyWilliams?utm_campaign=profiletracking&utm_medium=sssite&utm_source=ssslideview)
517 [_source=ssslideview](https://www.slideshare.net/AntonyWilliams?utm_campaign=profiletracking&utm_medium=sssite&utm_source=ssslideview)
- 518 38. Beretsou, V. G.; Psoma, A. K.; Gago-Ferrero, P.; Aalizadeh, R.; Fenner, K.; Thomaidis, N. S.,
519 Identification of biotransformation products of citalopram formed in activated sludge. *Water Res* **2016**,
520 *103*, 205-14.
- 521 39. Nika, M. C.; Bletsou, A. A.; Koumaki, E.; Noutsopoulos, C.; Mamais, D.; Stasinakis, A. S.;
522 Thomaidis, N. S., Chlorination of benzothiazoles and benzotriazoles and transformation products
523 identification by LC-HR-MS/MS. *J Hazard Mater* **2017**, *323*, (Pt A), 400-413.
- 524 40. Christophoridis, C.; Nika, M. C.; Aalizadeh, R.; Thomaidis, N. S., Ozonation of ranitidine: Effect of
525 experimental parameters and identification of transformation products. *Sci Total Environ* **2016**, *557-558*,
526 170-82.
- 527 41. Martens, L.; Chambers, M. C.; Sturm, M.; Kessner, D.; Levander, D.; Shofstahl, J.; Tang, W. H.;
528 Römpf, A.; Neumann, S.; Pizarro, A. D.; Montecchi-Palazzi, L.; Tasman, N.; Coleman, M.; Reisinger, F.;
529 Souda, P.; Hermjakob, H.; Binz, P.-A.; Deutsch, E. W., mzML - a Community Standard for Mass
530 Spectrometry Data. *Mol Cell Proteomics* **2011**, *10*, (1).
- 531 42. Pedrioli, P. G.; Eng, J. K.; Hubley, R.; Vogelzang, M.; Deutsch, E. W.; Raught, B.; Pratt, B.; Nilsson,
532 E.; Angeletti, R. H.; Apweiler, R.; Cheung, K.; Costello, C. E.; Hermjakob, H.; Huang, S.; Julian, R. K.; Kapp,
533 E.; McComb, M. E.; Oliver, S. G.; Omenn, G.; Paton, N. W.; Simpson, R.; Smith, R.; Taylor, C. F.; Zhu, W.;
534 Aebersold, R., A common open representation of mass spectrometry data and its application to
535 proteomics research. *Nat Biotechnol* **2004**, *22*, (11), 1459-66.
- 536 43. Erickson, B., ANDI MS standard finalized. *Anal Chem* **2000**, *72*, (3), 103 A-103 A.
- 537 44. Wang, M.; Carver, J. J.; Phelan, V. V.; Sanchez, L. M.; Garg, N.; Peng, Y.; Nguyen, D. D.; Watrous,
538 J.; Kapon, C. A.; Luzzatto-Knaan, T.; Porto, C.; Bouslimani, A.; Melnik, A. V.; Meehan, M. J.; Liu, W. T.;

- 539 Crusemann, M.; Boudreau, P. D.; Esquenazi, E.; Sandoval-Calderon, M.; Kersten, R. D.; Pace, L. A.; Quinn,
540 R. A.; Duncan, K. R.; Hsu, C. C.; Floros, D. J.; Gavilan, R. G.; Kleigrew, K.; Northen, T.; Dutton, R. J.;
541 Parrot, D.; Carlson, E. E.; Aigle, B.; Michelsen, C. F.; Jelsbak, L.; Sohlenkamp, C.; Pevzner, P.; Edlund, A.;
542 McLean, J.; Piel, J.; Murphy, B. T.; Gerwick, L.; Liaw, C. C.; Yang, Y. L.; Humpf, H. U.; Maansson, M.;
543 Keyzers, R. A.; Sims, A. C.; Johnson, A. R.; Sidebottom, A. M.; Sedio, B. E.; Klitgaard, A.; Larson, C. B.; P, C.
544 A. B.; Torres-Mendoza, D.; Gonzalez, D. J.; Silva, D. B.; Marques, L. M.; Demarque, D. P.; Pociute, E.;
545 O'Neill, E. C.; Briand, E.; Helfrich, E. J. N.; Granatosky, E. A.; Glukhov, E.; Ryffel, F.; Houson, H.; Mohimani,
546 H.; Kharbush, J. J.; Zeng, Y.; Vorholt, J. A.; Kurita, K. L.; Charusanti, P.; McPhail, K. L.; Nielsen, K. F.; Vuong,
547 L.; Elfeki, M.; Traxler, M. F.; Engene, N.; Koyama, N.; Vining, O. B.; Baric, R.; Silva, R. R.; Mascuch, S. J.;
548 Tomasi, S.; Jenkins, S.; Macherla, V.; Hoffman, T.; Agarwal, V.; Williams, P. G.; Dai, J.; Neupane, R.; Gurr,
549 J.; Rodriguez, A. M. C.; Lamsa, A.; Zhang, C.; Dorrestein, K.; Duggan, B. M.; Almaliti, J.; Allard, P. M.;
550 Phapale, P.; Nothias, L. F.; Alexandrov, T.; Litaudon, M.; Wolfender, J. L.; Kyle, J. E.; Metz, T. O.; Peryea,
551 T.; Nguyen, D. T.; VanLeer, D.; Shinn, P.; Jadhav, A.; Muller, R.; Waters, K. M.; Shi, W.; Liu, X.; Zhang, L.;
552 Knight, R.; Jensen, P. R.; Palsson, B. O.; Pogliano, K.; Lington, R. G.; Gutierrez, M.; Lopes, N. P.; Gerwick,
553 W. H.; Moore, B. S.; Dorrestein, P. C.; Bandeira, N., Sharing and community curation of mass
554 spectrometry data with Global Natural Products Social Molecular Networking. *Nat Biotechnol* **2016**, *34*,
555 (8), 828-837.
- 556 45. Sturm, M.; Bertsch, A.; Gropl, C.; Hildebrandt, A.; Hussong, R.; Lange, E.; Pfeifer, N.; Schulz-
557 Trieglaff, O.; Zerck, A.; Reinert, K.; Kohlbacher, O., OpenMS - an open-source software framework for
558 mass spectrometry. *BMC Bioinformatics* **2008**, *9*, 163.
- 559 46. Uppal, K.; Walker, D. I.; Liu, K.; Shuzhao, L.; G., Y.-M.; P., J. D., Computational Metabolomics: A
560 Framework for the Million Metabolome. *Chem Res Toxicol* **2016**, *29*, (12), 1956-1975.
- 561 47. Allard, P. M.; Genta-Jouve, G.; Wolfender, J. L., Deep metabolome annotation in natural
562 products research: towards a virtuous cycle in metabolite identification. *Curr Opin Chem Biol* **2017**, *36*,
563 40-49.
- 564 48. Samanipour, S.; Reid, M. J.; Thomas, K. V., Statistical Variable Selection: An Alternative
565 Prioritization Strategy during the Nontarget Analysis of LC-HR-MS Data. *Anal Chem* **2017**, *89*, (10), 5585-
566 5591.
- 567 49. Samanipour, S.; Reid, M.; Baek, K.; Thomas, K. V., Combining a deconvolution and a universal
568 library search algorithm for the non-target analysis of data independent LC-HRMS spectra. *Environ Sci*
569 *Technol* **2018**, In Press.

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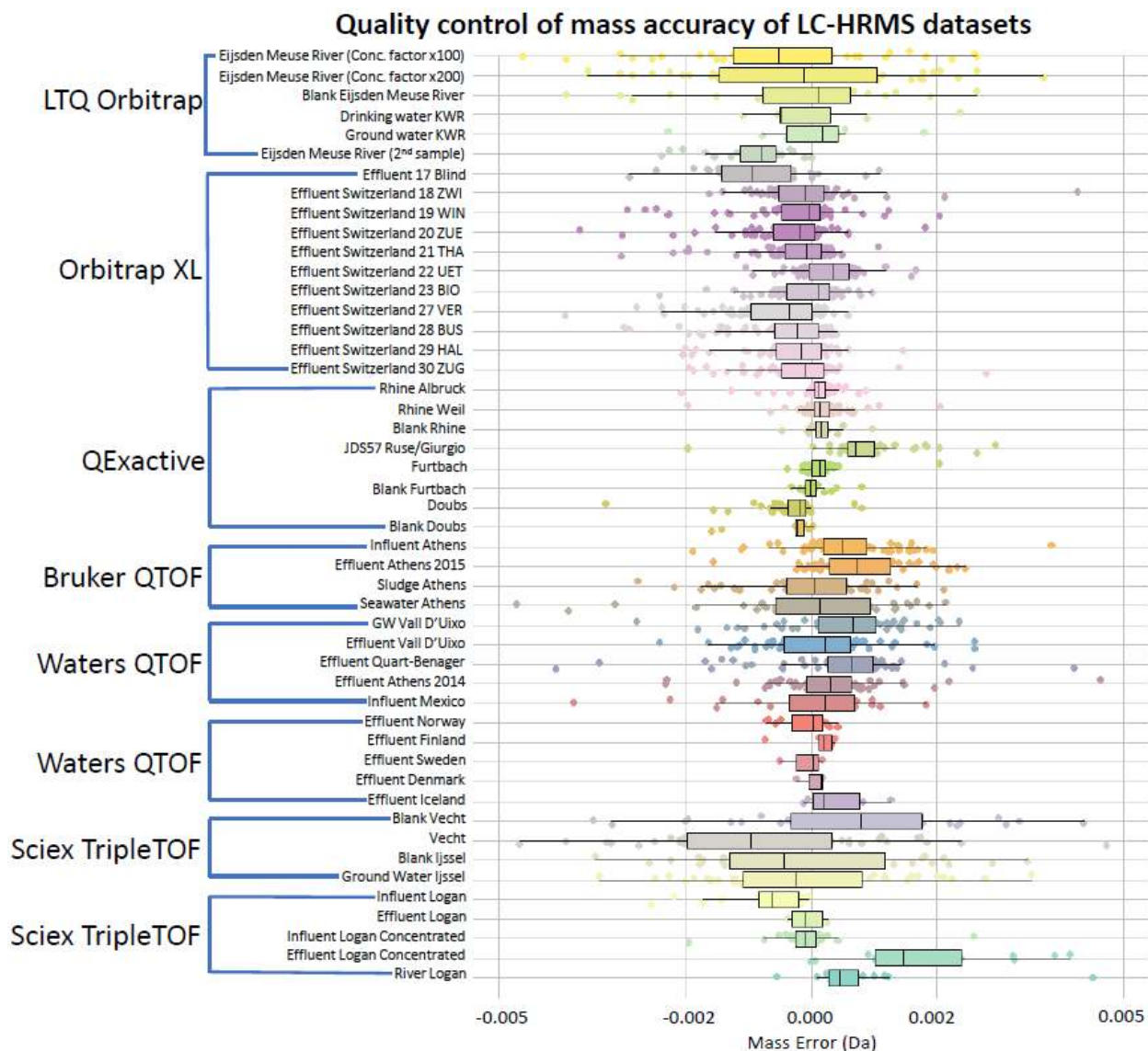


Figure 1. Quality control of mass accuracy of the submitted datasets based on the identified compounds. Type of mass analyzer, calibration type of the mass analyzer as well as other factors (age of equipment, scan sampling rate of the detector) affect the performance and the quality of the results.

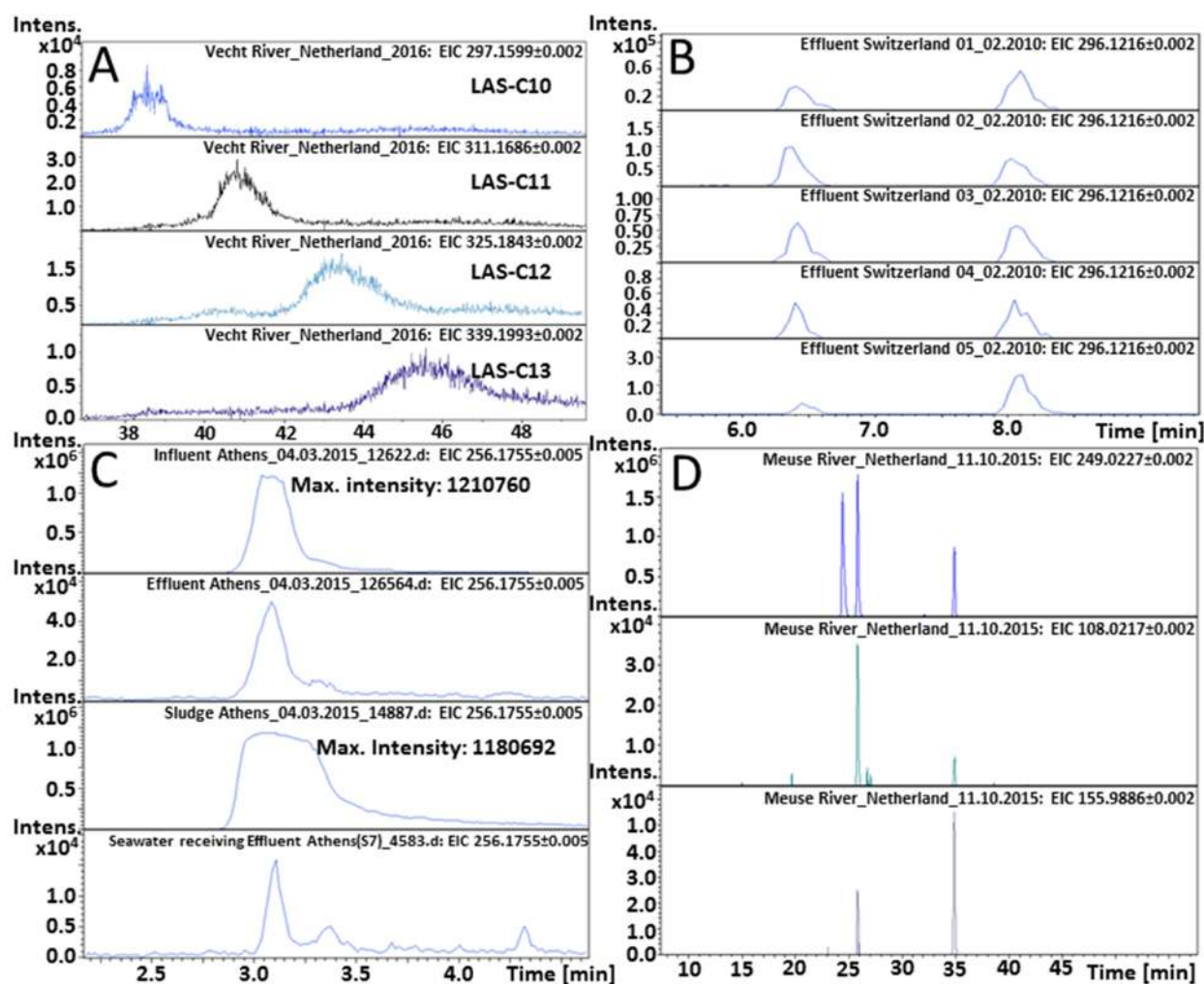
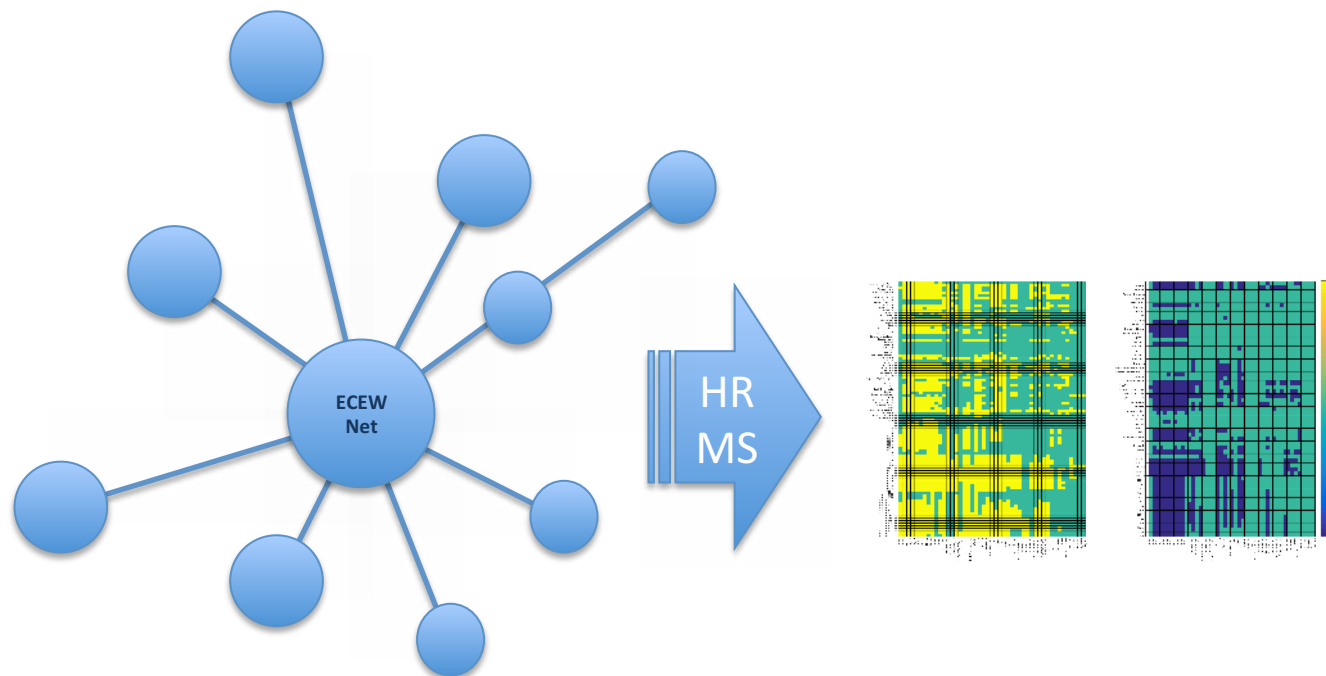


Figure 3. Challenges faced during evaluation of the results; A. Broad peaks of Linear alkylbenzene sulphonate (LAS) surfactants makes peak-picking challenging, B. Missing fragmentation information (MS/MS) decreases identification confidence because of data-dependent acquisition. Peaks are mass accuracy and isotopic profile consistent but not abundant enough so that MS/MS spectra have not been acquired (case of Quetiapine-N-desalkyl), C. Saturation of detector deteriorates mass accuracy, affects peak-picking and causes quantification mistakes when quantification is done by maximum intensity and not by peak area (case of PEG-05), D. Bisphenol S isomers cannot be distinguished, because in both cases qualifier fragment ions (m/z 108.0217 and 155.9886) are present in both peaks in the high collision energy channel.



A global emerging contaminant early warning network through the use of retrospective suspect screening with high-resolution mass spectrometry