SCIENTIFIC REPORTS

OPEN

SUBJECT AREAS: ENVIRONMENTAL CHEMISTRY MAGNETIC MATERIALS CATALYST SYNTHESIS SUSTAINABILITY

> Received 18 April 2013

Accepted 16 July 2013

Published 13 August 2013

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Facile synthesis of highly efficient and recyclable magnetic solid acid from biomass waste

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In this work, sawdust, a biomass waste, is converted into a magnetic porous carbonaceous (MPC) solid acid catalyst by an integrated fast pyrolysis–sulfonation process. The resultant magnetic solid acid has a porous structure with high surface area of 296.4 m² g⁻¹, which can be attributed to the catalytic effect of Fe. The catalytic activity and recyclability of the solid acid catalyst are evaluated during three typical acid-catalyzed reactions: esterification, dehydration, and hydrolysis. The favorable catalytic performance in all three reactions is attributed to the acid's high strength with 2.57 mmol g⁻¹ of total acid sites. Moreover, the solid acid can be reused five times without a noticeable decrease in catalytic activity, indicating the stability of the porous carbon (PC)–sulfonic acid group structure. The findings in the present work offer effective alternatives for environmentally friendly utilization of abundant biomass waste.

ith the depletion of fossil fuels, development of renewable energy has attracted an increasing amount of interest. Acidic materials are widely used in the catalytic production of a variety of renewable fuels (e.g., biodiesels) and fine chemicals (e.g., furfurals and sugars)¹⁻³. In the past, liquid mineral acids (e.g., HCl, H₂SO₄, and H₃PO₄) have been widely used to catalyze these reactions^{1,4}. However, many disadvantages, such as strong corrosiveness, costly and inefficient separation from homogeneous reaction mixtures, and creation of a large amount of waste by-products, have limited the applications of liquid mineral acids in many fields⁵. Alternatively, solid acids including solid Brønsted acids (e.g., SO₃H functionalized carbon and polymers)⁶⁻⁹ in addition to solid Lewis acids (transition-metal (W, Zr, Nb) oxides)^{10,11} are easily separated from the reaction mixture and can be reused. Thus, solid acids have been used to replace liquid mineral acids in catalytic reactions such as esterification, dehydration, and hydrolysis during biofuel production to minimize adverse effects on the environment and to reduce expenses^{5,12-16}. Porous carbonaceous (PC) solid acids are widely used catalysts in acidcatalyzed reactions. Nakajima et al.⁶ reported that these acids can be synthesized by sulfonation of partially carbonized organic matter including sugars, cellulosic materials, and synthetic polymers and can be used in many acid catalytic reacions such as hydrolysis of β -1,4 glucan and biodiesel production. Arancon et al.¹⁷ synthesized a PC solid acid from corncob residue, which demonstrated favorable catalytic performance in the esterification/transesterification of waste oils. Wu et al.¹⁸ prepared sulfonated bio-char as a solid acid for the hydrolysis of crystalline cellulose.

Compared with conventional solid acids, magnetic solid acids can be separated more easily in the presence of external magnetic fields, particularly in viscous or solid reaction mixtures^{19,20}. The most common method used for synthesis of magnetic solid acid catalysts is sulfonation of a mesoporous structure with a magnetic core, generally Fe_3O_4 . In a previous study, a magnetic solid acid with a high acid strength of 2.2–2.5 H⁺/mol was prepared through sulfonation of polystyrene-encapsulated Fe_3O_4 spheres²¹. More recently, a series of magnetic nano-sized solid acid catalysts was synthesized by sulfonation of a series of particles containing Fe_3O_4 cores with various shell materials (e.g., polystyrene, poly(glycidyl methacrylate), and silica)²². All of these catalysts exhibit high activity and excellent separability in the acid-catalyzed reactions. However, many magnetic solid acid catalysts reported thus far have been determined as unsatisfactory for acid catalyzed reactions. For example, SO₃H-functionalized polymers often exhibit low thermal stability and are costly²³. In addition, the preparation of these magnetic solid acid catalysts involves complex synthetic procedures and often requires expensive reagents, which impedes large-scale production and commercialization.

In our continuing efforts toward biomass waste disposal and resource recovery²⁴⁻²⁷, we have developed a rapid, low-cost, and readily scalable approach for synthesizing a magnetic porous carbonaceous solid acid catalyst (MPC-SO₃H) by using biomass waste as a precursor. This method involves fast pyrolysis of Fe-preloaded

biomass to produce MPC material; subsequent sulfonation of the obtained material is used to prepare a solid acid catalyst. This process also produces bio-oil, a renewable liquid that can be used for fuel and chemical production²⁸⁻³⁰.

Results

A schematic illustration of the synthetic pathway for MPC–SO₃H is shown in Fig. 1. Synthesis of MPC precursor materials is the primary step for MPC-SO₃H synthesis, which includes the following steps: The Fe preloaded on the biomass in the form of FeCl₃ is hydrolyzed to Fe hydroxides (FeO(OH)) in the drying process (Eqs. 1, 2). The Fepreloaded biomass is then subjected to a pyrolysis process to be converted into MPC material. In this process, the Fe-preloaded biomass is thermally decomposed, and the produced volatile components are swept out by the carrier gas (N₂) and condensed to produce bio-oil. The FeO(OH) is reduced to Fe₃O₄ under mesothermal conditions at 673–873 K by reducing components including H₂, CO, and amorphous carbon, which were formed during the pyrolysis process (Eqs. 3–5)^{31,32}. For comparison, PC materials without magnetism that were derived from raw biomass are also synthesized.

 $FeCl_3 + 3 H_2O \rightarrow Fe(OH)_3 + 3 HCl \uparrow$ (1)

$$Fe(OH)_3 \rightarrow FeO(OH) + H_2O$$
 (2)

$$6 \operatorname{FeO}(OH) + 4 \operatorname{H}_2 \rightarrow 2 \operatorname{Fe}_3 O_4 + 4 \operatorname{H}_2 O \uparrow \tag{3}$$

 $6 \operatorname{FeO}(OH) + 4 \operatorname{C} \rightarrow 2 \operatorname{Fe}_3 O_4 + 2 \operatorname{CO} \uparrow \tag{4}$

$$6 \operatorname{FeO}(OH) + 4 \operatorname{CO} \rightarrow 2 \operatorname{Fe}_3 O_4 + 4 \operatorname{CO}_2 \uparrow \tag{5}$$

Figure 2a shows X-ray diffraction (XRD) patterns of MPC (Pattern A) and PC (Pattern B). Both patterns show a weak but broad diffraction peak of 2θ at $\sim 23^{\circ}$, which can be attributed to the amorphous carbon composed of aromatic carbon sheets oriented in a relatively random manner³³. The XRD peaks in Pattern A of 2θ at 30.1° , 35.7° , 37.1° , 43.2° , 53.6° , 62.7° , and 74.9° indicate the formation of Fe₃O₄ crystallite in the fast pyrolysis process according to Joint Committee on Powder Diffraction Standard (JCPDS) 19-0629. Figure 2b shows

the nitrogen sorption isotherms of MPC and PC. For Pattern A, the isotherms exhibit a typical type III pattern with an H2-type hysteresis loop at high relative pressures, indicating that the pores of MPC have uniform sizes and shapes³⁴. For Pattern B, however, the isotherms exhibit a typical type II pattern without a significant hysteresis loop, suggesting that PC is nearly devoid of pores. Based on the nitrogen quantity adsorbed at different relative pressures, the surface area of the MPC is calculated by the Brunauer-Emmett-Teller (BET) method to be 391.7 m² g⁻¹, which is significantly higher than that of PC at 14.0 $m^2 g^{-1}$. This result indicates that the presence of Fe can catalyze the formation of porous structures during biomass carbonization mainly because the dehydration and decomposition of FeCl₃ can enhance the release of volatile matter such as HCl, H2O, CO, and CO₂ (Eqs. 1-5) during the biomass pyrolysis process, which generally produces an increase in surface area and pore volume. Similar phenomena have been observed in the carbonization of FeCl₃-impregnated pinewood sawdust and carbon aerogels containing Fe^{35,36}. FeCl₃ easily dehydrates carbohydrate polymers (e.g., cellulose and hemicellulose) at high temperatures and can change the decomposition pathway of lignocellulosic biomass during fast pyrolysis to inhibit the formation of heavy tars that may block pore structures, thus improving the formation of open pores in the carbon matrix. Furthermore, the formed Fe species, e.g., FeOOH or Fe₃O₄ may act as an *in situ* catalyst in the fast pyrolysis process for the generation of porous structures³⁷.

Scanning electron microscopy (SEM) and transmission electron microscopy (TEM) show a very rough MPC surface morphology (Fig. 2c). Energy dispersive X-ray (EDX) results indicate that the surface is composed mainly of C, O, and Fe elements (Fig. 2d) and many wire-like structures, which are also confirmed by TEM (Fig. 2e). In addition, porous structures are also observed in the TEM image. Conversely, neither wire-like nor porous structures are detected in the SEM and TEM images for PC (Figs. 2f and 2g). These results confirm those of the nitrogen sorption isotherms, which indicate that MPC has porous structures with high surface area, and PC has very low surface area and is nearly devoid of porous structures.

A sulfonation process is applied to both MPC and PC to produce $MPC-SO_3H$ and $PC-SO_3H$. Figure 3a shows the XRD pattern of



Figure 1 | Schematic illustration of the synthesis of a magnetic porous carbonaceous (MPC) solid acid catalyst from naturally abundant biomass waste (all photographs and images were produced by W. J. Liu).



Figure 2 | (a) X-ray diffraction (XRD) patterns of the magnetic porous carbonaceous (MPC) solid acid precursor (pattern A) and the precursor of the carbonaceous solid acid without magnetism (PC) (pattern B). (b) N_2 adsorption–desorption isotherms of MPC and PC. (c) Scanning electron microscopy (SEM) image of MPC. (d) Energy dispersive X-ray (EDX) analysis of MPC; the Au is a film applied before SEM analysis to improve conductivity of MPC. (e) Transmission electron microscopy (TEM) image of MPC. (f) SEM image of PC. (g) TEM image of PC.

MPC-SO₃H, which indicates that Fe₃O₄ remains after sulfonation. The density and distribution of the S and Fe elements in MPC-SO₃H are evaluated by quantitative EDX mapping. S is determined to be homogeneously distributed on the entire surface of MPC-SO₃H (Fig. 3e), suggesting effective sulfonation of the carbonaceous precursor material. As shown in Fig. 3f, Fe is also homogeneously distributed on the entire surface of MPC-SO3H after sulfonation, which is consistent with the XRD results such that the Fe₃O₄ structure is not damaged in the sulfonation reaction and thus the magnetism of the solid acid catalyst remains. The results of analysis of surface area and pore volume of MPC-SO₃H and PC-SO₃H by nitrogen adsorptiondesorption isotherms are shown in Fig. 3b, Fig. S1, and Table S1. The surface area and pore volume of MPC-SO₃H are 296.4 m² g⁻¹ and $0.14 \text{ cm}^{-3} \text{ g}^{-1}$, respectively, which are slightly lower than those of MPC. This result suggests that pore structure of MPC has been partially damaged in the sulfonation process. However, as shown in the SEM and TEM images (Figs. 3c and 3d), the wire-like and porous structures of MPC are nearly unchanged after the sulfonation process.

The surface composition of MPC–SO₃H is analyzed by X-ray photoelectron spectroscopy (XPS). The C 1s spectrum includes six peaks with differentiated binding energy values via deconvolution (Fig. 3g). These peaks could be assigned to the carbon atoms in the forms of C–S (284.1 eV), C–C (284.6 and 284.9 eV), C–O (alcoholic, phenolic hydroxyl and/or ether, 285.4 eV), C = O (carbonyl and/or

quinine, 286.7 eV), and O = C-O (carboxyl groups, 289.0 eV)^{38,39}. For comparison, the C 1s spectrum of MPC prior to sulfonation is shown in Fig. S2, in which only two carbon forms, C–C and C–O, are apparent. This result indicates that the sulfonation process not only introduces SO₃H groups on the carbon matrix through the formation of C–S bonds but also modifies the surface of the materials with additional oxygenated functional groups (e.g., C = O and O-C = O). The S 2p spectrum of MPC–SO₃H shows three different peaks (Fig. 3h), which could be attributed to the S–C (168.1 eV), S–O (168.9 eV), and S = O bonds (169.9 eV)^{6,40}. These three bonds further indicate that the S in MPC–SO₃H is mainly in the form of SO₃H groups bonded to carbon matrix⁴¹.

Because the XPS results indicate that the S in MPC–SO3H is mainly associated with SO₃H groups, the SO₃H contents of MPC– SO₃H can be estimated from the S content. The S content of MPC– SO₃H and PC–SO₃H is 5.6 wt.% and 3.1 wt.%, respectively (Table S2). Thus, the SO₃H contents of MPC–SO₃H and PC–SO₃H are calculated to be 1.75 mmol g⁻¹ and 0.97 mmol g⁻¹, respectively. The total acid sites of MPC–SO₃H and PC–SO₃H are 2.57 mmol g⁻¹ and 1.26 mmol g⁻¹, respectively, as determined through titration of a NaOH solution. The larger contents of the acid sites compared with those of SO₃H in the solid acid suggest that acidic groups other than SO₃H groups, such as COOH, are also present in the solid acid^{18,42}. This theory is confirmed through XPS and Fourier transform infrared spectroscopy (FTIR) (Fig. S3).





Figure 3 | (a) X-ray diffraction (XRD) patterns of magnetic porous carbonaceous solid acid (MPC–SO₃H). (b) N₂ adsorption–desorption isotherms of MPC–SO₃H and porous carbonaceous solid acid (PC–SO₃H). (c) Scanning electron microscopy (SEM) image of MPC–SO₃H. (d) Transmission electron microscopy (TEM) image of MPC. (e) Distribution of S in MCP–SO₃H. (f) Distribution of Fe in MCP–SO₃H. (g) C 1s X-ray photoelectron spectroscopy (XPS) spectrum of MPC–SO₃H. (h) S 2p XPS spectrum of MPC–SO₃H.

The catalytic activities of the solid acids are evaluated in three typical acid-catalyzed reactions (Fig. 4). For Reaction A, MPC-SO₃H exhibits high activity for the catalytic formation of benzyl acetate with a yield of 93%, which is significantly higher than that of PC-SO₃H at 63% and comparable to that of concentrated sulfuric acid at 89% (Fig. 5a). For Reaction B, one of the primary reactions for the production of furfural from renewable biomass, only 35% of the xylose is converted in the absence of a catalyst, and no furfural is produced (Fig. 5b). The furfural yields of 45%, 61%, and 65% with the catalysis of PC-SO₃H, MPC-SO₃H, and commercially available H₂SO₄, respectively, indicate a favorable catalytic performance of the magnetic solid acid^{43,44}. Moreover, as shown in Fig. S4, more than 96% of the xylose can be converted within 6 h with the catalysis of MPC-SO₃H. For Reaction C, no sucrose is hydrolyzed in the absence of a catalyst (Fig. 5c); however, a hydrolysis conversion of 89% with the catalysis of PC-SO₃H is noted with a glucose and fructose yield of 83%. These results are significantly lower than those of MPC-SO₃H and H₂SO₄, in which the hydrolysis conversions are both near 100% with monosaccharide yields of 94% and 97%, respectively. Furthermore, MPC-SO₃H catalyzed hydrolysis of the sucrose occurs rapidly because more than 90% of the sucrose can be hydrolyzed in the first hour (Fig. S5).

For the solid acid catalyst, one of the most important advantages over the liquid mineral acid in the acid-catalyzed reaction is that it can be separated and reused easily. In the present study, the spent solid acid catalyst is separated from the reaction mixture by using a magnet (Fig. S6a) and is reused in the same esterification reaction. After five reuses, the catalytic activity of MPC–SO₃H remains unchanged with an average benzyl acetate yield of ~90% (Fig. 5d). Moreover, the magnetism of MPC–SO₃H also remains after five reuses (Fig. S6b). SEM image (Fig. S7) of MPC–SO₃H after five repeated experiments shows no significant difference in surface morphology compared to that of the raw catalyst (Fig. 3a). The XPS results of the reused MPC–SO₃H (Table S3 and Fig. S8) show no noticeable change in surface element compositions of *C*, O, S, and Fe, which indicates high stability of MPC–SO₃H in the acid–catalytic reaction. In addition, MPC–SO₃H also exhibits high thermal stability (Fig. S9).

Discussion

MP solid acid was synthesized through an integrated fast pyrolysis– sulfonation process. This process minimizes the environmental impact of large-scale production because it uses naturally abundant, renewable biomass waste as feedstock. The full advantage of such a process can be achieved if the loading of Fe can be achieved by using the biomass as a sorbent for extraction of Fe in wastewater. Bio-oil, a renewable fuel resource, was also produced during the fast pyrolysis process, which may result in additional economic benefits.



Figure 4 | The three typical acid-catalyzed reactions, esterification of benzyl alcohol and acetic acid (Reaction A), dehydration of xylose to produce furfural (Reaction B), and hydrolysis of sucrose (Reaction C).

Magnetic solid acids exhibit strong acid strength and are readily separable. Therefore, they provide promising substitutions for liquid acids in some acid catalyzed reactions, which can minimize adverse effects on the environment and human health. Furthermore, because the solid acid catalyst here is easily separated and shows high catalytic activity in reuse, it offers promising applications as a low cost, environmentally friendly, and recyclable acid catalyst in the production of fine chemicals.

In summary, we have synthesized a porous magnetic solid acid catalyst through a facile integrated fast pyrolysis–sulfonation process by using naturally abundant biomass waste. During this process, the biomass waste is first used as a sorbent to adsorb Fe(III) ions from water to obtain Fe-loaded biomass, which is then used as pyrolysis feedstock for energy and resource recovery by creating bio-oil. The pyrolysis bio-char is then subjected to a sulfonation process to synthesize the MPC solid acid catalyst. It has been determined that the loaded Fe(III) on the biomass can be converted into Fe₃O₄ during the fast pyrolysis process, endowing the materials with magnetism that remains after the sulfonation process. In addition, the loaded Fe can improve formation of the porous structure on carbonaceous materials. The magnetic solid acid has a porous structure with a high surface area of 296.4 m² g⁻¹ and high acid strength with total acid sites of



Figure 5 | (a) Comparison of the catalytic performance of MPC-SO₃H with other acid catalysts in the esterification of acetic acid and benzyl alcohol in (b) the dehydration of the xylose, (c) the hydrolysis of sucrose, and (d) the reuse test of catalytic activity.

2.57 mmol g⁻¹. Moreover, this acid exhibits excellent catalytic performance, favorable separability, and high durability in typical acidcatalyzed reactions. The results presented in this work offer new opportunities for the development of facile and environmentally friendly methods of synthesizing functional materials from naturally abundant biomass waste.

Methods

Materials. All reagents and solvents were purchased from Sinopharm Chemical Reagent Co., Shanghai, China, and used without further purification. The biomass used in this work was fir sawdust, a naturally abundant lignocellulosic biomass waste obtained from a local timber treatment plant in Hefei, China. After collected, the biomass was initially washed to remove impurity particles, and then dried in an oven at 378 K overnight. The dried biomass was crushed using a high-speed rotary cutting mill, and the biomass with the particle size smaller than 120 mesh were collected. The proximate analysis and elemental composition of the biomass were determined and the results were reported in our previous work²⁴.

Synthesis of the magnetic porous carbonaceous solid acid catalyst. The precursor of solid acid catalyst was the Fe-loaded biomass, which was prepared in a biosorption process using biomass as an adsorbent at ambient temperature. Briefly, 10.0 g of biomass and 1,000 mL of FeCl₃ solution with a concentration of 10 mmol L⁻¹ were mixed in a flask and shaken in a constant temperature oscillator at 200 rpm for 300 min. Then, the water in the mixture was evaporated, and the solid residue was dried at 378 K overnight and sieved again to collect the particles with a size smaller than 120 mesh, and the Fe-loaded biomass with Fe content of approximately1.0 mmol g⁻¹ was obtained.

The magnetic porous carbonaceous solid acid catalyst was synthesized by fast pyrolysis of the Fe-loaded biomass and subsequent sulfonation of the produced biochar. The fast pyrolysis experiments were carried out in a vertical drop fixed bed reactor described in our previous work²⁷. A 7.0 g portion of the Fe-loaded biomass sample was first placed in the feed pipe under a nitrogen flow of 400 mL min⁻¹, and the nitrogen flow was maintained for 20 min to remove air from the pyrolysis system. Subsequently, when the temperature rose to the setting value of 773 K, the Fe-loaded biomass sample was inserted into the quartz tubular reactor with a piston for pyrolysis. The volatiles produced in the pyrolysis process were swept out by a nitrogen flow of 200 mL min⁻¹, and condensed by cold ethanol to obtain bio-oil. When the fast pyrolysis process was accomplished, the temperature of the heating zone was increased to 873 K, and the bio-char was further carbonized for 1 hour. Finally, the reactor was moved out of the heating zone and cooled in the nitrogen flow to room temperature, and the magnetic porous carbonaceous material was obtained.

The solid acid catalyst was synthesized by sulfonation of the magnetic porous carbonaceous material. Briefly, the magnetic porous carbonaceous material was mixed with concentrated sulfuric acid (10 mL of sulfuric acid to 1 gram of solid), sonicated for 15 min, and subsequently heated under vigorous stirring at 423 K for 10 hours to introduce sulfo-group (SO₃H) on the surface of the materials. Afterwards, the mixture was cooled to room temperature, and then slowly added to a beaker containing 500-mL deionized water. The suspension mixture was then filtered off and washed repeatedly with deionized water until there was no sulfate ions (SO₄²⁻) detected in the washing (detected by Ba²⁺ solution). The solid was further washed with absolute ethanol to remove the water, and then dried at 353 K overnight. The dried sample was the magnetic porous carbonaceous solid acid and was stored for further use (MPC–SO₃H). For comparison, another carbonaceous solid acid was synthesized in the same way using raw biomass without Fe-loading as precursor (PC–SO₃H).

Characterization. The carbon (C) and sulfur (S) contents of the solid acids were analyzed using a high-frequency infrared carbon and sulfur analyzer (CS-600, LECO, USA), while the Fe content of the MPC-SO3H was determined by an X-ray fluorescence spectrometer (XRF-1800, Shimadzu Co., Japan). The total amount of acid sites was determined by an acid-base titration method using NaOH solution (10 mmol L^{-1})⁴⁵.

FTIR spectra were recorded on a Fourier transform infrared spectroscopy (Bruker EQUINOX55, Germany) to analyze the functional groups of the solid acid. The thermal stability of the solid acid was characterized with a DTG-60H/DSC-60 thermogravimetric analyzer (TGA, Shimadzu Co., Japan). The structural features of the solid acids were analyzed by nitrogen adsorption-desorption isotherms, performing at 77 K on a Micromeritics Gemini apparatus (ASAP 2020 M+C, Micromeritics Co. USA) after degassing the sample at 393 K under vacuum for 10 hours. Specific surface areas of the solid acid were measured by tBET method, while their pore volumes were calculated by the amount of nitrogen adsorbed at a relative pressure of 0.99. The surface morphology of the solid acid was analyzed using TEM (JEOL-2100F, Japan) and SEM (Sirion 200, FEI electron optics company, USA) coupled with EDX (INCA energy, UK). XPS measurements of the solid acid were performed using a Thermo VG-Scientific spectrometer (ESCALAB250, USA) with a monochromatized Al Ka radiation (1486.92 eV). XRD analysis of the solid acids was carried out in an 18-KW rotating anode X-ray diffractometer (MXPAHF, Japanese Make Co., Japan).

Acid-catalyzed reactions. The catalytic activities of the solid acids were evaluated in three typical acid-catalyzed reactions, including esterification of benzylacohol and

acetic acid (Reaction A), dehydration of the xylose to produce furfural (Reaction B), and hydrolysis of sucrose (Reaction C).

The esterification reaction was performed in a solvent-free system as follows: 0.100 g of catalyst was added to a 25 mL reaction tube, followed by the dose of 0.108 g (1.0 mmol) of benzyl alcohol and 0.290 g (5.0 mmol) of acetic acid. Then, the mixture was vigorously stirred with a magnetic stirrer at 363 K to conduct the reaction. The reaction was completed in a nitrogen atmosphere and monitored by a thin layer chromatograph (TLC). After the reactions were accomplished (no benzyl alcohol was detected in the TLC), the reaction mixture was washed with cyclohexane of 10 mL, and the catalyst was separated with the aid of a powerful magnet. The remaining cyclohexane solution was washed with NaHCO₃ solution of 0.01 mol L⁻¹ to remove the unreacted acetic acid, and then dried over anhydrous MgSO₄. Finally, the products (benzyl acetate) was obtained after removal of the solvent under reduced pressure and confirmed by comparing the retention time with that of the pure benzyl acetate in a gas chromatograph. After reaction, the catalyst was separated by an external magnet, then washed with ethyl acetate several times, then dried at 333 K overnight and reused for subsequent reactions.

For the dehydration reaction, $d^{-}(+)$ -xylose (0.150 g, 1 mmol), DMSO (5.000 g), and acid catalyst (0.100 g) were loaded into a 25-mL reaction tube. The mixture was vigorously stirred and heated in a constant-temperature oil bath (423 K) for 300 min. After reaction, the mixture was cooled to room temperature. The concentration of formed furfural was analyzed by gas chromatography, and the unreacted xylose content was determined by a phloroglucinol colorimetric assay using a UV-vis spectrophotometer (UV-1700, Phenix Optical Scientific Instrument Co., China)⁴⁶. The conversion of xylose and yield of furfural were calculated with Eqs. 6 and 7, respectively.

$$Xylose \ yield = \frac{initial \ xylose - unreacted \ xylose}{initial \ xylose} \times 100\%$$
(6)

$$Furfural yield = \frac{furfural \ content}{initial \ xylose} \times 100\%$$
(7)

For the hydrolysis reaction, sucrose (0.342 g, 1 mmol), water (20 mL), and acid catalyst (0.100 g) were added into a 100-mL round bottomed flask. The mixture was vigorously stirred and heated in a constant temperature oil bath (353 K) for 150 min. After reaction, the mixture was cooled to room temperature. The hydrolysis products, i.e., glucose and fructose, were determined by a 3,5-dinitrosalicylic acid colorimetric assay on a UV-vis spectrophotometer (UV-1700, Phenix Optical Scientific Instrument Co., China)⁴⁷. The unreacted sucrose was analyzed using the same method after hydrolysis in 6-mM HCl solution (373 K) for 5 min. The conversion of sucrose and yields of hydrolysis products were calculated with Eqs. 8 and 9, respectively.

$$Sucrose \ conversion = \frac{initial \ sucrose - unreacted \ sucrose}{initial \ sucrose} \times 100\%$$
(8)

$$Hrdrolysis \ products \ yield = \frac{glu \cos e \ and \ fructose \ content}{initial \ sucrose} \times 100\%$$
(9)

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Acknowledgments

The authors gratefully acknowledge financial support from National 863 Program (2012AA063608-01), National Key Technology R&D Program of the Ministry of Science and Technology (2012BAJ08B00), and the Key Special Program on the S&T for the Pollution Control and Treatment of Water Bodies (No.2012ZX07103-001).

Author contributions

W.J.L. designed and conducted the experiments, analyzed the data, and wrote the manuscript; H.J. provided guidance on the manuscript preparation, modified the manuscript, and supervised the whole work; K.T. conducted part of the experiments; H.Q.Y. modified this manuscript and provided important suggestions. All authors contributed through scientific discussions.

Additional information

Supplementary information accompanies this paper at http://www.nature.com/ scientificreports

Competing financial interests: The authors declare no competing financial interests.

How to cite this article: Liu, W.-J., Tian, K., Jiang, H. & Yu, H.-Q. Facile synthesis of highly efficient and recyclable magnetic solid acid from biomass waste. Sci. Rep. 3, 2419; DOI:10.1038/srep02419 (2013).



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